

Binghamton University

## The Open Repository @ Binghamton (The ORB)

---

Research Days Posters Spring 2020

Division of Research

---

2020

### Identification of genes that affect cardiac failure in diabetic *Drosophila melanogaster*

Ashley O'Toole

*Binghamton University--SUNY*

Christie Santoro

*Binghamton University--SUNY*

Follow this and additional works at: [https://orb.binghamton.edu/research\\_days\\_posters\\_spring2020](https://orb.binghamton.edu/research_days_posters_spring2020)

---

#### Recommended Citation

O'Toole, Ashley and Santoro, Christie, "Identification of genes that affect cardiac failure in diabetic *Drosophila melanogaster*" (2020). *Research Days Posters Spring 2020*. 64.

[https://orb.binghamton.edu/research\\_days\\_posters\\_spring2020/64](https://orb.binghamton.edu/research_days_posters_spring2020/64)

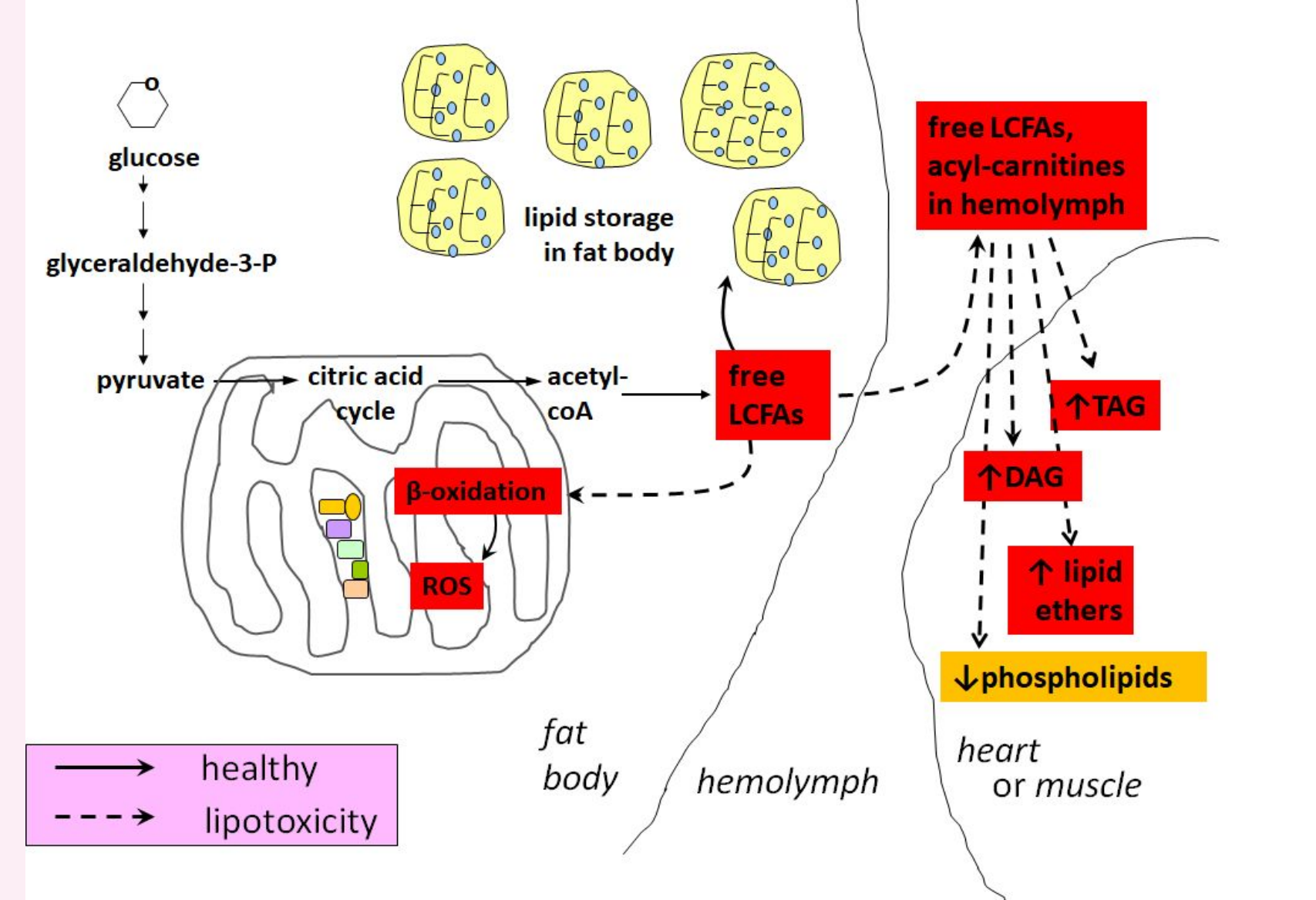
This Book is brought to you for free and open access by the Division of Research at The Open Repository @ Binghamton (The ORB). It has been accepted for inclusion in Research Days Posters Spring 2020 by an authorized administrator of The Open Repository @ Binghamton (The ORB). For more information, please contact [ORB@binghamton.edu](mailto:ORB@binghamton.edu).



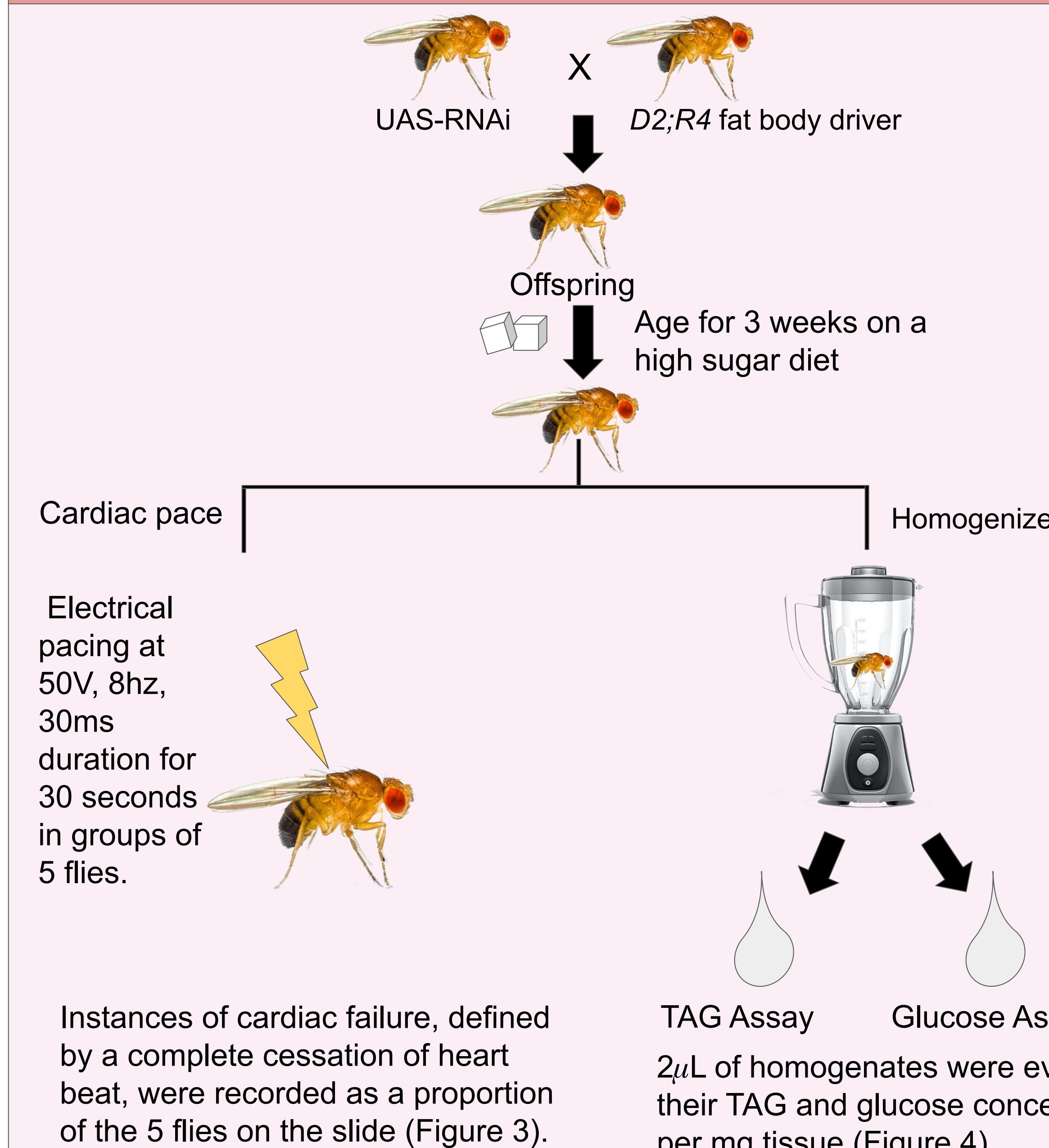
## Background

- High-sugar diets exacerbate type two diabetes and cardiovascular disease in humans and model organisms, including *Drosophila melanogaster*. Type two diabetes is characterized by hyperglycemia and increased triglyceride (TAG) levels.
- High sugar diets are linked to cardiovascular disease through lipotoxicity (1).
- Lipotoxicity is thought to be a byproduct of limited fat storage where fatty acids produced beyond that limit lead to the accumulation of free floating fatty acids in the bloodstream and, ultimately, toxicity in peripheral tissues (2,3).
- While the lipids and mechanisms underlying lipotoxicity are not well understood, increased abundance of some lipids are strongly associated with impaired cardiovascular function, such as plasmalogens (4).
- Our goal is to identify lipids or proteins that ameliorate or exacerbate responses to cardiac stress-induced heart failure through tissue-specific genetic loss of function.

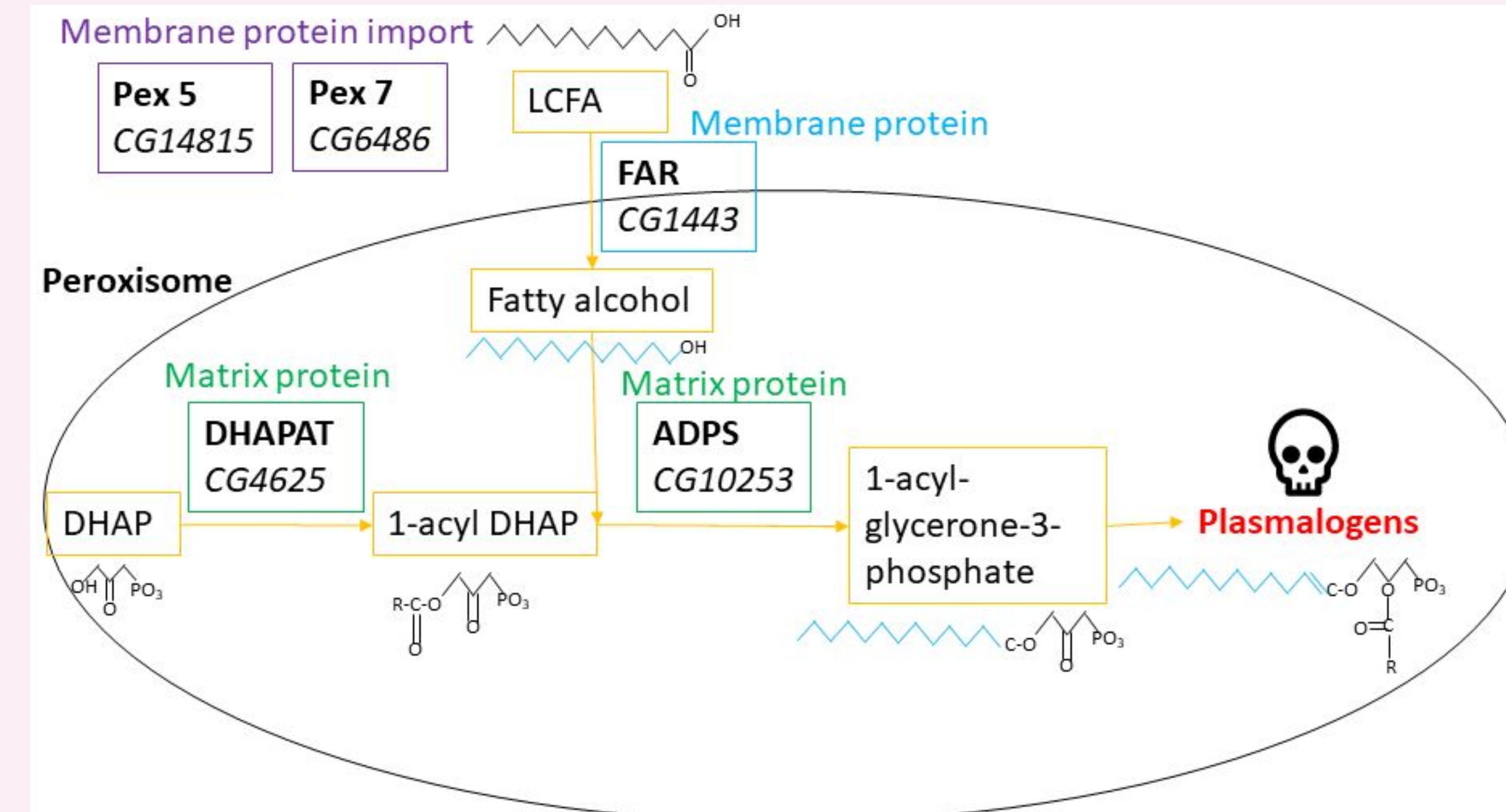
**Figure 1. Model: High sugar feeding leads to lipotoxicity via the maximum expandability model.** High sugar-induced lipogenesis makes free long chain fatty acids (LCFAs), which accumulate in the bloodstream and cause peripheral lipotoxicity.



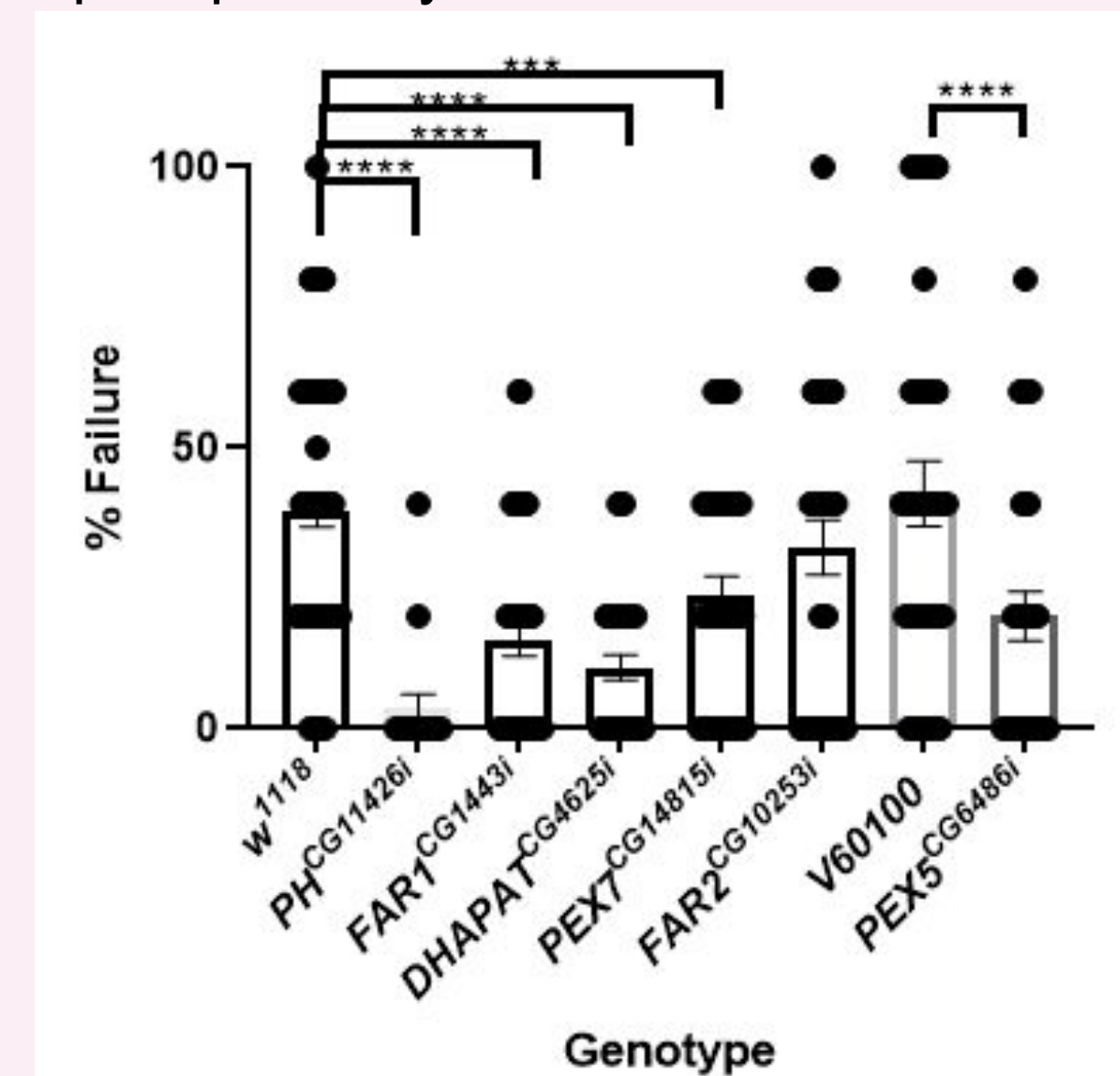
## Methods



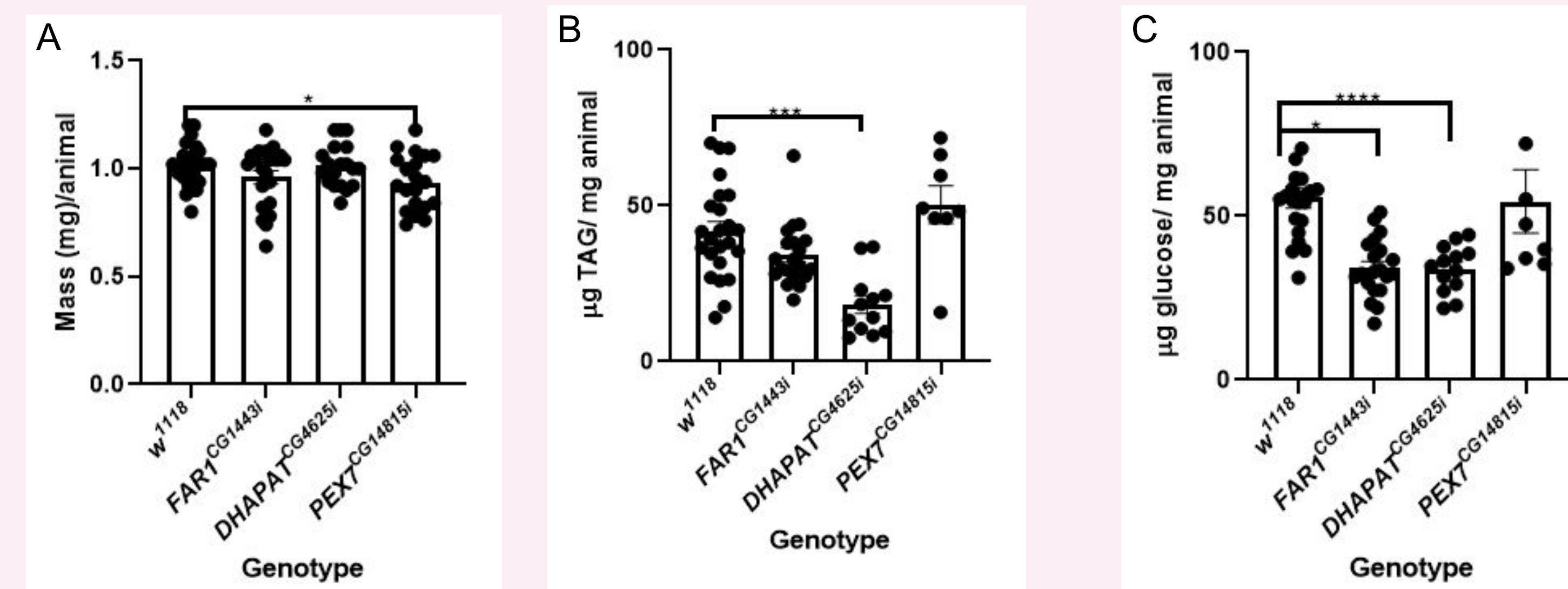
## Results



**Figure 2: Plasmalogen and peroxisomal genes function in plasmalogen formation.** Genes in the peroxisome were selected as genes of interest because they are responsible for the production of plasmalogen lipid species, which are seen at a higher concentration under high sugar feeding conditions. *CG14815* and *CG6486* encode a peroxisomal transport proteins Pex 5 and Pex 7, *CG1443* encodes for a fatty acyl reductase, *CG4625* encodes for dihydroxyacetone phosphate acyl transferase, and *CG10253* encodes for alkylidihydroxyacetonephosphate synthase.

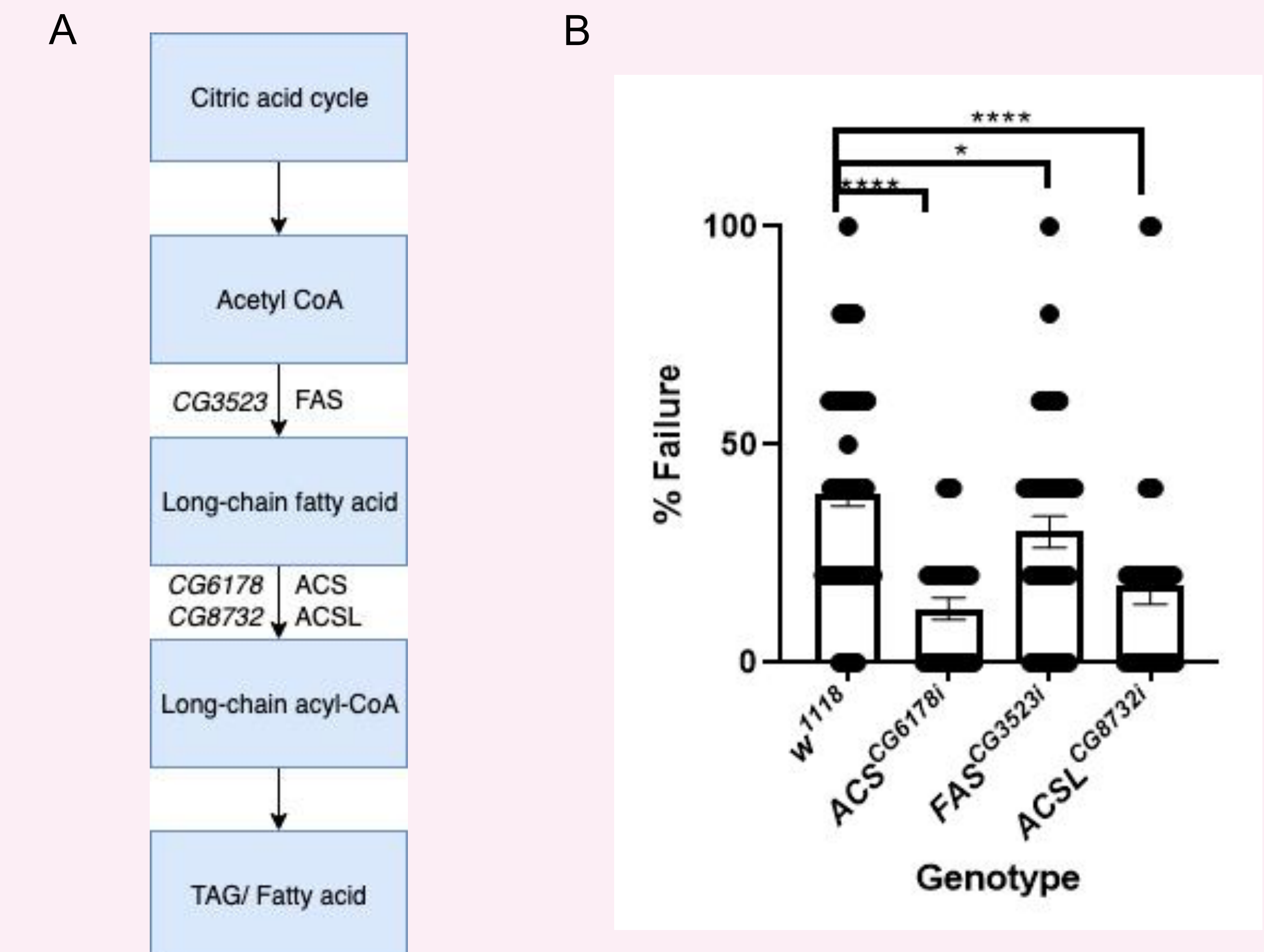


**Figure 3: Plasmalogen and peroxisomal genetic knockdown show an increased resistance to cardiac pacing.** Cardiac pacing phenotypes were evaluated for fat body specific (*r4-GAL4*) RNAi knockdown genotypes after three weeks of high sugar feeding. Control *w<sup>1118</sup>* (n=345) was compared with *CG11426* (n=85, p<0.0001), *CG4625* (n=165, p<0.0001), and *CG14815* (n=205, p=0.0003). *V60100* (n=160) was compared with *CG6486* (n=150, p<0.0001). Significance was determined by a 2X2 chi squared test.



**Figure 4: Diabetic phenotypes are ameliorated through plasmalogen synthesis and peroxisomal gene knockdowns.** Phenotypes were evaluated for fat body specific (*r4-GAL4*) RNAi knockdown genotypes after three weeks of high sugar feeding. Genetic knockdowns in the fatty acid biosynthesis pathway revealed some statistically significant differences in weight: *w<sup>1118</sup>* (n=20), *CG1443* (n=20, p=0.1406), *CG4625* (n=8, p=0.0412), and *CG14815* (n=20, p<0.01) (A). TAG assays revealed statistically significant differences in TAG: Control *w<sup>1118</sup>* (n=20), *CG1443* (n=20, p=0.1939), *CG4625* (n=20, p=0.0002), and *CG14815* (n=20, p=0.1939) (B). Genetic knockdowns revealed a decreased level in whole animal glucose compared to genetic control: *w<sup>1118</sup>* (n=20), *CG1443* (n=20, p=0.0153), *CG4625* (n=20, p<0.0001), and *CG14815* (n=20, p=0.8859) (C). Significance was determined by a Student's T Test.

## Results



**Figure 5: Fatty acid biosynthesis gene knockdowns increase resistance to cardiac stress.** Cardiac pacing phenotypes were evaluated for fat body specific (*r4-GAL4*) RNAi knockdown genotypes after three weeks of high sugar feeding. Genes were selected based on their role in fatty acid biosynthesis as either a fatty acid synthase (*CG3523*), acyl-CoA ligase (*CG6178*), or an acyl-CoA synthetase long chain (*CG8732*) (A). Control *w<sup>1118</sup>* (n=345) was compared with *CG3523* (n=220, p=0.0306), *CG6178* (n=155, p<0.0001), and *CG8732* (n=135, p<0.0001) (B). Significance was determined by a 2X2 chi squared test.

## Conclusion and Future Direction

- Genetic knockdowns on the interface of the maximum expandability can ameliorate cardiac function after high sugar feeding.
- Knockdowns also display an improvement on other type two diabetes like phenotypes such as high glucose levels and high TAG levels.
- Future direction for this work is to identify and mechanistically define how these genes are contributing to cardiac lipotoxicity.

## Acknowledgement

We thank Dr. Carol Miles for the use of her square wave stimulator and Bryon F. Tuthill II for help with method development. We also thank BU and the American Heart Association for funding.

## References

- Kenny, H. C., & Abel, E. D. (2019). Heart Failure in Type 2 Diabetes Mellitus. *Circulation Research*, 124(1), 121–141. doi:10.1161/circresaha.118.311371
- Unger RH, Scherer PE. Gluttony, sloth and the metabolic syndrome: a roadmap to lipotoxicity. *Trends Endocrinol Metab*. 2010 Jun;21(6):345–52.
- Virtue S, Vidal-Puig A. Adipose tissue expandability, lipotoxicity and the Metabolic Syndrome—an allostatic perspective. *Biochim Biophys Acta*. 2010 Mar;1801(3):338–49.
- Diop, S. B., & Bodmer, R. (2012). *Drosophila* as a model to study the genetic mechanisms of obesity-associated heart dysfunction. *Journal of cellular and molecular medicine*, 16(5), 966–71.
- Na J, Musselman LP, Pendse J, Baranski TJ, Bodmer R, Ocorr K, et al. A *Drosophila* model of high sugar diet-induced cardiomyopathy. *PLoS Genet*. 2013 Jan;9(1):e1003175.
- Musselman LP, Fink JL, Narzinski K, Ramachandran PV, Hathiraman SS, Cagan RL, et al. A high-sugar diet produces obesity and insulin resistance in wild-type *Drosophila*. *Model Mech*. 2011 Nov;4(6):842–9.
- Palanker Musselman L, Fink JL, Baranski TJ. CoA protects against the deleterious effects of caloric overload in *Drosophila*. *J Lipid Res*. 2016 Mar;57(3):380–7.
- Musselman LP, Fink JL, Ramachandran PV, Patterson BW, Okunade AL, Maier E, et al. Role of fat body lipogenesis in protection against the effects of caloric overload in *Drosophila*. *J Biol Chem*. 2013 Mar 22;288(12):8028–42.