

A toolbox of biophysical and analytical assays helps to confirm the activity of novel *hTS* dimer disrupters (Ddis) with anticancer properties

Lorenzo Tagliacruzchi ^a, Alberto Venturelli ^a, Giulia Malpezzi ^a, Maria Gaetana Moschella ^b, Daniele Aiello ^a, Ludovica Lopresti ^c, Cecilia Pozzi ^c, Gaetano Marverti ^b, Domenico d'Arca ^b, Glauco Ponterini ^a, Maria Paola Costi ^a

^a Department of Life Sciences, Università di Modena e Reggio Emilia, Via Campi 103, 41125-Modena, Italy

^b Department of Biomedical, Metabolic and Neural Sciences, Università di Modena e Reggio Emilia, Via Campi 287, 41125-Modena, Italy

^c Department of Biotechnology, Chemistry and Pharmacy, Università di Siena, Via Aldo Moro 2, 53100-Siena, Italy

Background and Aim

Thymidylate synthase (*hTS*) is an homodimeric protein existing in a dimer-monomer equilibrium. Drugs that target the *hTS* are widely used in anticancer therapy. However, treatment with classical substrate site-directed TS inhibitors such as 5-FU binds to dimer and induce *hTS* overexpression.

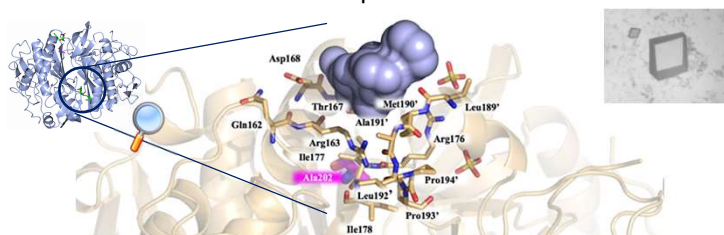


Figure 1. particular of a *hTS* monomer interface binding a Ddis

We have then synthesized and tested a large library of dissociative inhibitors (Ddis) of *hTS* that bind to the monomer interface, resulting in enzyme inhibition without overexpression. A toolbox of biophysical assays was developed to screen their activity.

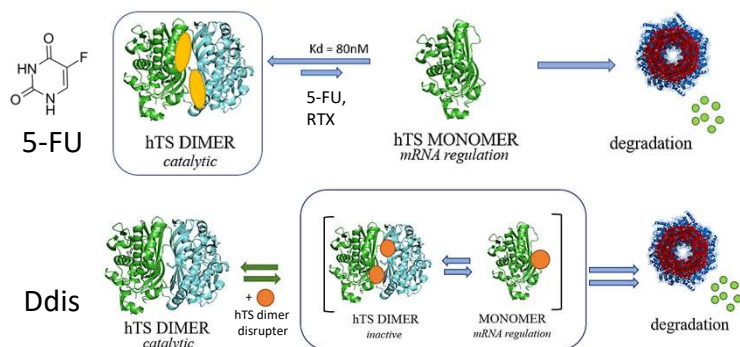


Figure 2. (a) *hTS* Monomer dimer-equilibrium when a 'traditional' inhibitor (i.e. 5-FU, RXT) is administered. (b) Novel dimer disrupters move *hTS* equilibrium towards the monomer form, causing catalytic inhibition without enzyme overexpression.

1. The direct kinetic studies on the target revealed a mixed mode inhibition model, with Ddis binding preferably to the monomers. The equilibrium is described by a Michaelis Menten mathematical model in Figure 3.

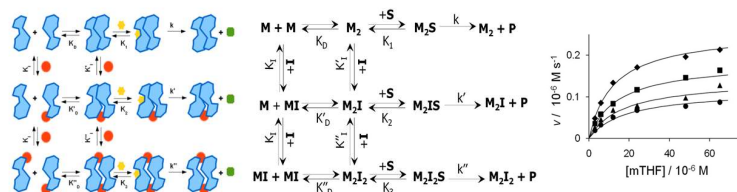


Figure 3. (left) kinetic panel describing *hTS* equilibrium when perturbed by Ddis (Mixed Mode). (right) MM graph at increasing cofactor concentration

2. We labelled each *hTS* monomer with fluorescent probes to test the actual percentage of dimers getting disrupted by the Ddis with a FRET assay.

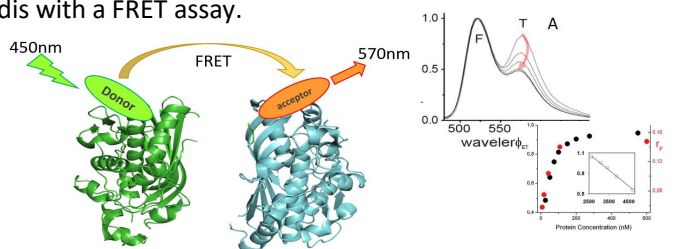


Figure 4. Representation of a FRET assay on *hTS*. Each monomer is biolabelled with fluorescein as donor, and rhodamine-5-maleimide as FRET acceptor.

Results and discussion

3a. The co-elution of *hTS* with Ddis was checked by Anionic Exchange Chromatography (AEX) on FPLC. Increasing Ddis concentration in *hTS* solution causes monomer peak to rise.

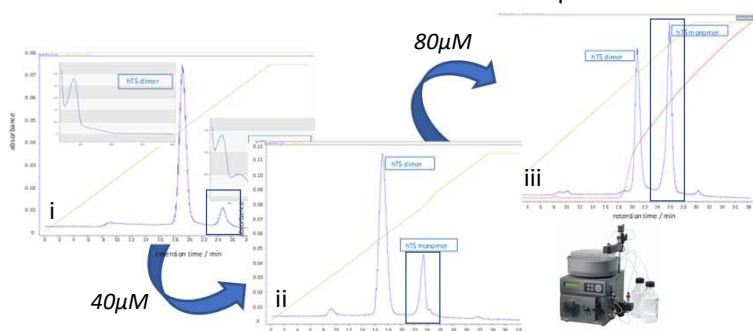


Figure 5. AEX chromatograms of *hTS* with 4 (i), 40 (ii) and 80 (iii) μM Ddis.

3b. We have employed LC-MS to analyse the actual amount of Ddis that binds *hTS* monomer and dimer fractions.

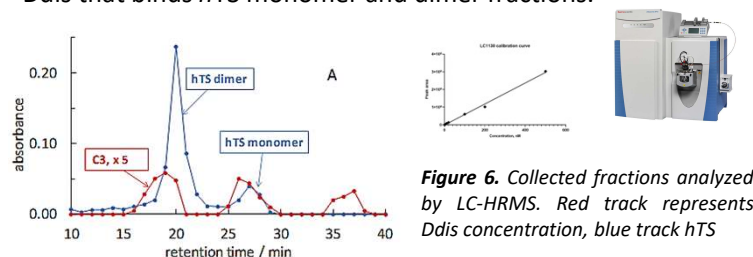


Figure 6. Collected fractions analyzed by LC-HRMS. Red track represents Ddis concentration, blue track *hTS*

Conclusions

Biophysics + bioanalytical techniques → target engagement and *Kd*'s
Elution profile with AEX → *Ddis* : *hTS* co-elution confirmation
Future investigations: Photoaffinity labelling (PAL) to validate binding

References:

- Genovese F, Ferrari S et al. Dimer-monomer equilibrium of human thymidylate synthase monitored by fluorescence resonance energy transfer. *Protein Sci.* 2010 May;19(5):1023-30.
- Costantino L, Ferrari S et al. Disrupters of the thymidylate synthase homodimer accelerate its proteasomal degradation and inhibit cancer growth, 2021 ChemRxiv

This research was funded by Associazione Italiana per la Ricerca sul Cancro, IG number 16977, to MP Costi