IJBCP International Journal of Basic & Clinical Pharmacology

Research Article

Evaluation of wound healing activity of ethanolic extract of Azadirachta Indica leaves on incision and excision wound models in Wister albino rats

H. N. Nagesh*, P. L. Basavanna, M. S. Kishore

Department of Pharmacology, Mysore Medical College and Research Institute, Mysore, Karnataka, India

Received: 06 October 2015 Revised: 08 October 2015 Accepted: 14 November 2015

***Correspondence to:** H. N. Nagesh, Email: nagu728@gmail.com

Copyright: © the author(s), publisher and licensee Medip Academy. This is an openaccess article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT

Background: Wound healing is complex cellular and biochemical cascade that lead to restitution of integrity and function. Recently, the traditional use of plants for wound healing has received attention by the scientific community, as traditional medicine is a source of less expensive, comprehensive medical care, especially in developing countries. *Azadirachta indica* (Neem) is well-known in India, as one of the most versatile medicinal plants having a wide spectrum of biological activity. Hence, this study was undertaken to evaluate the wound healing activity of the ethanolic extract of *A. indica* leaf in the experimentally-induced wound in rats.

Methods: The healing effect produced by *A. indica* extract was assessed by the rate of wound contraction histopathology and skin breaking strength by using excision wound model and incision wound model in Wister albino rats. This was compared with control (soft white paraffin) and standard (1% w/w framycetin sulfate ointment). The results have been analyzed by calculating the mean values, standard deviation and compared by using student t-test.

Results: The ethanol extract of leaves of *A. indica* significantly promoted the wound healing activity in both excision and incision wound models.

Conclusion: The study revealed promising wound healing activity of ethanolic extract of *A. indica* and provides a scientific rationale for the traditional use in the management of wounds.

Keywords: Traditional medicine, Framycetin sulfate ointment, Soft white paraffin

INTRODUCTION

Wound is delineated as disruption of structural and physiological continuity of living tissue. They are inescapable events of life; wound may arise due to physical, chemical, or microbial agents. Wound healing is complex cellular and biochemical cascade that lead to restitution of integrity and function, accomplished by several processes which involve different phases including inflammation, granulation, fibro genesis, neo-vascularization, wound contraction and epithelization.¹ When healing takes place in a direction away from its normal course, it is common to have non-healing, under or over healing. Treatment is, therefore, aimed at either shortening the time required for healing or minimizing the undesired consequences.

The World Health Organization (WHO) has been promoting traditional medicine as a source of less expensive, comprehensive medical care, especially in developing countries. The WHO also recognized the importance of traditional medicine and has treated strategies, guidelines, and standard for botanical medicines.² Approximately, one-third of all traditional medicines in use is for the treatment of wounds and skin disorders, compared to only 1-3% of modern drugs.³

Azadirachta indica (Neem) is well-known in India for more than 2000 years, as one of the most versatile medicinal plants having a wide spectrum of biological activity. It is called as "villagers' dispensary" because of its medicinal value. Every part of the tree has been used as a household remedy against various human ailments. Various studies have been carried out and it is shown to have antipyretic, immunostimulant, antiulcer, antioxidant, hypoglycemic, hepatoprotective activity.⁴

Hence, this study was undertaken to evaluate the wound healing activity of the ethanolic extract of *A. indica* leaves in the experimentally-induced wound in albino rats.

Objectives

- 1. To evaluate the wound healing activity of ethanolic extract of *A. indica* leaves in Albino rats
- 2. To compare the wound healing activity of *A. indica* with the standard, 1% w/w framycetin sulfate cream (FSC) ointment.

METHODS

A total 36 healthy adult Wister albino rats of either sex weighing 180-250 g were randomly selected from the central animal house. Pregnant and diseased animals were excluded in this study. The study was conducted after obtaining approval from Institutional Animal Ethical Committee.

The ethanolic extract of leaves of *A. indica* (test drug) formulation was obtained from the Himalaya drug company. 5 g of alcoholic extract of *A. indica* was mixed with 95 g of soft paraffin to prepare 5% ointment (w/w).

The selected animals were housed individually and maintained at a temperature-controlled, well-ventilated animal room for a period of 7-day prior to the experimental period to allow for acclimatization to the laboratory condition. They were kept on standard pellet diet and water *ad-libitum*. The animals were starved 12 hrs prior to wounding with water *ad-libitum*. Wounding was performed aseptically under light ether anesthesia.

Wound models

The wound healing activity was carried out in two different wound models in albino rats such as excision wound model and incision wound model.

In each model, the animals were divided into three groups, each group containing six animals.

Group-I (control) received the topical application of simple ointment base (B.P)

Group-II (standard) received the topical application of framycetin sulfate cream 1%~w/w

Group-III (test) received the topical application of 5% w/w ointment of *A. indica*.

Excision wound model

Under light ether anesthesia, the animals were secured to operation table in prone position. An impression was made on the dorsal thoracic region 1 cm away from vertebral column and 5 cm away from ear on the anesthetized rat. The particular skin area was shaved 1 day prior to the experiment. Under aseptic precautions, the skin of impressed area was excised to the full thickness to obtain a wound area of about 500 mm². Hemostasis was achieved by blotting the wound with a cotton swab soaked in normal saline.⁵ The ointments were applied topically once in a day, until the epithelization

was complete starting from the day of the experiment. The parameters studied were shown in Table 1.

The percentage of wound healing was calculated of original wound size (500 mm²) for each animal of group using the formula:⁷

	Wound area on day 0 –
Percentage of wound	wound area on respective day
closure =	wound area on day 0

Incision wound model^{8,9}

Under light ether anesthesia, the animals were secured to operation table in natural position. The skin of the back where the wound was to be made was shaved. Two paravertebral straight incisions of 6 cm each were made with the help of sharp surgical blade through full thickness of the skin on either side of the vertebral column of the rat. Care was taken to see that the incisions were at least 1 cm lateral to the vertebral column. After complete hemostasis, the wounds were closed by means of interrupted sutures with 4-0 silk thread placed at equidistant points about of 1 cm apart. Wounds were mopped with cotton swabs soaked in 70% alcohol, and animals were caged individually (Figure 1a).

The ointment containing the test formulation was applied topically once in a day. The sutures were removed on 8th post wound day, and the tensile strength (defined as the force just sufficient to disrupt the wound) of the healed wound was measured on 10th day (Figure 1b).

Statistical analysis

The data were entered in excel format and analyzed by calculating the mean, standard deviation, and the groups were compared using Student's t-test (p<0.05 was considered as significant).

RESULTS

The results of excision and incision wound models were showed in (Table 2). In excision wound model, the test group



Figure 1: Incision wound model. (a) Incision wound model showing sutured two paravertebral straight incisions. (b) Determination of tensile strength by using continuous water flow technique of Lee

Wound model	Attribute	Parameters studied	Methodology	Made on post wounding day
Excision	Physical	Percentage of wound contraction	Planimetry*	0, 4, 8, 12, 16 and subsequently on every alternate day till complete epithelisation
		Epithelization time (days)	Observed the day on which scab falls	Noted on falling of scab without raw area
		Scar area of excision wound (mm ²)	Planimetry	Studied on complete epithelization
	Histological ⁶	Cellular elements, fibroblast and collagen fibers	Microscopy	4, 8, and 12
Incision	Mechanical	Tensile strength measurement	Continuous water flow technique of Lee ⁹	10

Table 1: The parameters studied in excision and incision wound models.

*Retracing the wound from butter paper on to mm scale graph paper and then number of squares were counted

fable 2: Th	e parameters	studied an	d results o	f excision a	and incision	wound models.
-------------	--------------	------------	-------------	--------------	--------------	---------------

Groups	Excision wound model						Incision wound model	
	Percentage of wound contraction Epithelisation Scar area							Tensile
	4th day	8th day	12th day	16th day	18th day	period (days)	(mm ²)	strength (g)
Control	$18.84{\pm}0.54$	38.54 ± 0.65	59.24±0.73	76.77±0.55	90.45±0.52	22.83±0.47	40.50 ± 1.70	205.61±6.87
Standard	29.15 ± 0.75	54.22 ± 0.65	78.50 ± 0.57	$90.54{\pm}0.78$	99.03±0.33	18.33 ± 0.49	31.66±2.01	383.05±11.23
Test	$26.94{\pm}0.36$	51.22 ± 0.64	75.86 ± 0.37	88.66±0.51	97.21±0.40	19.66±0.33	34.16±2.03	325.97±13.36

sq.mm=Square millimeter, G=Grams

showed significant increase in the percentage of wound contraction when compared to control group (Figure 2).

The mean time for complete epithelization in test group (19.66 ± 0.33) was statistically decreased (p<0.01) compared to control group (22.83\pm0.47) and statistically increased (p=0.01) compared to standard group (18.33±0.49).

The mean scar area was significantly decreased (p<0.01) in test group compared to control group; but was increased when compared to the standard group.

Histology of excision wounds in test group showed significantly increased fibrocollagenous tissue deposition anti-inflammatory activity when compared to control group (Figure 3).

In incision wound model, there was a significant increase (p<0.01) in the tensile strength of test group compared to the control group and comparable to the standard group.

DISCUSSION

Wound healing is one of most important defense mechanism of the body as proper healing is essential for restoration of disrupted anatomical continuity and disturbed functional state.

The finding of the present study clearly indicates that the test drug (*A. indica*) significantly promoted healing of excision

wound as evidenced by enhanced wound closure rate (97.21% on 18th day), decreased time duration (19 days) and mean scar area (34.16 mm²) of complete epithelization. The study also showed wound healing was promoted by abundant proliferation of connective tissues with angiogenesis.

In incision wound study, the test drug promoted healing of resutured incision wound as evidenced by significant increase (p<0.01) in the tensile strength of test group. The increased tensile strength might be due to increased proliferation and transformation of fibroblast cells into myofibroblasts.

The study of Barua et al.¹⁰ states that in the excision wound model, the mean percent of closure of wound increased significantly (p<0.01) from "0" day until 21st day which was 95.65% in case of *A. indica* treated group in comparison to the control group (75.15% on 28th day post wounding) and in the incision wound model, significant increase (p<0.05) in tensile strength was observed in the methanolic extract of *A. indica* compared to the control group.

The study results of Pandey et al.¹¹ showed wound contraction rate (83.93 ± 1.38), epithelization period (15.86 ± 0.33), tensile strength and hydroxyproline content were significantly increased for volatile oil of *A. indica* treated group as compared to control group. It also explained that terpenoids, source of *A. indica*, which play an important role in wound healing because the terpenoid strengthen the skin, increase the concentration of antioxidants in wounds,



Figure 2: Photograph of Excision wounds showing wound contraction



Figure 3: Histology of Excision wounds

and restore inflamed tissues by increasing blood supply.¹² The previous studies showed *A. indica* plant possesses Anti-oxidant activity,¹³ antibacterial activity,¹⁴ and Anti-inflammatory activity¹⁵ which may help to promote wound healing.

About 5% ointment of *A. indica* showed significant wound healing activity but when compared to the standard, 1% w/w FSC ointment, it takes more time for complete epithelization. Increased percentage concentration of *A. indica* or combined with other wound healing herbs may decrease the duration of wound healing and will be a better agent than standard as phytochemicals are not only cheap and affordable but are also safe.

The findings of the present study if extrapolated to the clinical situation, it appears that *A. indica* when used in a surgical incision or clean excision wounds could promote healing independent of their antibacterial activity.

CONCLUSION

The use of *A. indica* in Indian traditional systems of medicine for wound healing has been justified by this work. The ethanolic extract of *A. indica* leaves showed highly significant pro-healing effect almost equivalent to standard drug, which may be partly due to the anti-inflammatory activity, proliferation of fibrocollagenous tissue and angiogenesis properties. Hence, it can be used as a wound healing agent if it is confirmed by clinical trials, which would be cost effective. As animal studies cannot be directly compared with effects on humans, there is a need for clinical evaluation in humans to confirm this effect.

Funding: No funding sources Conflict of interest: None declared Ethical approval: The study was approved by the Institutional Animal Ethics Committee

REFERENCES

- Clark RA. Wound repair: overview and general considerations. In: Clark RA, Henson PM, editors. The Molecular and Cellular Biology of Wound Repair. 2nd Edition. New York: Plenum Press; 1996: 473-88.
- 2. Akerele O. Nature's medicinal bounty: don't throw it away. World Health Forum. 1993;14(4):390-5.
- Pirbalouti AG, Koohpayeh A, Karimi I. The wound healing activity of flower extracts of *Punica granatum* and *Achillea kellalensis* in Wistar rats. Acta Pol Pharm. 2010;67(1):107-10.
- Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. Biological activities and medicinal properties of neem (*Azadirachta indica*). Curr Sci. 2002;82(11):1336-45.
- Shafiuddin M, Khan A, Ali S. Wound healing activity of traditional herbal formulation. Int J Chem Sci. 2009;7(2):639-43.
- Morton JJ, Malone MH. Evaluation of vulneray activity by an open wound procedure in rats. Arch Int Pharmacodyn Ther. 1972;196(1):117-26.

- Singhal A, Gupta H, Bhati V. Wound healing activity of Argyreia nervosa leaves extract. Int J Appl Basic Med Res. 2011;1(1):36-9.
- Ehrich HP, Hunk TK. Effect of cortisone and anabolic steroids on tensile strength of healing wound. Ann Surg. 1969;170:203-6.
- Lee KH. Studies on the mechanism of action of salicylate. II. Retardation of wound healing by aspirin. J Pharm Sci. 1968;57(6):1042-3.
- Barua CC, Talukdar A, Barua AG, Chakraborty A, Sharma RK, Bora RS. Evaluation of the wound healing activity of methanolic extract of *Azadirachta indica* (Neem) and *Tinospora cordifolia* (Guduchi) in rats. Pharmacol Online. 2010;1:70-7.
- Lee G, Luna HT. Manual of Histological Staining Methods of the Armed Forces, Institute of Pathology. 3rd Edition. New York, Toronto, London, Sydney: American Registry of Pathology, The Blakiston Division; 1968: 68-9.
- 12. Pandey IP, Ahmed SF, Chhimwal S, Pandey S. Chemical

composition and wound healing activity of volatile oil of leaves of *Azadirachta indica* A. juss. Adv Pure Appl Chem. 2012;1(3):62-6.

- Amer H, Helmy WA, Taie HA. *In vitro* antitumor and antiviral activities of seeds and leaves neem (*Azadirachta indica*) extracts. Int J Acad Res. 2010;2(2):47-51.
- Mandal R, Dhaliwal PK. Antifertility effect of Melia azedarach Linn. (Dharek) seed extract in female albino rats. Indian J Exp Biol. 2007;45(10):853-60.
- Girish K, Shankara Bhat S. Neem A green treasure. Electron J Biol. 2008;4(3):102-11.

Cite this article as: Nagesh HN, Basavanna PL, Kishore MS. Evaluation of wound healing activity of ethanolic extract of *Azadirachta Indica* leaves on incision and excision wound models in Wister albino rats. Int J Basic Clin Pharmacol 2015;4:1178-82.