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# Raman spectroscopy of blood plasma for cancer diagnosis

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## ABSTRACT

Raman spectra of blood plasma were studied in the dynamics of the experimental glioma. We used a DXR Raman Microscope (Thermo Scientific), excitation wavelengths of 532 nm, range 80–3200  $\text{cm}^{-1}$ . Each sample of blood plasma was a droplet with a volume of 10  $\mu\text{L}$  placed on a special aluminum plate. Machine learning methods were used to identify the most informative frequencies associated with cancer molecular markers. The most significant changes in the Raman spectra are observed in the 900–1700  $\text{cm}^{-1}$  range.

**Keywords:** Raman spectra, blood plasma, U87 human glioblastoma, machine learning methods

## INTRODUCTION

A Raman spectrum represents a molecular fingerprint of the sample and provides quantitative information regarding its chemical makeup. Biochemical changes in cells and tissues that may either be caused or are the cause of a disease can lead to significant changes in the Raman spectra. Raman spectroscopy is a powerful analytical technique that can be applied in the differential diagnosis of breast, skin and other types of cancer [1]. Raman spectroscopy and machine learning techniques were used to identify glioma margins and grades [2, 3], and biochemical changes induced by IDH1 mutation in human glioma [4, 5]. The Raman spectra included molecular vibrational fingerprints of carotenoids, tryptophan, amide I/II/III, proteins, and lipids. The cross-validated sensitivity, specificity, and accuracy were found to be 100%, 96.3%, and 99.6% to distinguish glioma tissues from normal brain tissues, respectively [6]. The analysis of blood can enable powerful, minimally invasive diagnostics for many diseases, including cancer [1, 7, 8]. Raman spectroscopy combined with advanced chemometrics is allowed the discrimination between samples of blood from breast cancer patients and healthy volunteer controls [8, 9], between samples of cirrhotic patients with and without hepatocellular carcinoma [10]. Using the principal component analysis, we previously assessed the general differences in the Raman spectra of the blood serum in rats in the dynamics of development of the experimental cholangiocarcinoma [11].

The purpose of our work is to study the Raman spectra of blood plasma in the dynamics of the development of experimental glioblastoma. Glioblastoma is among the deadliest neoplasms and represents approximately 49% of all brain and other central nervous system malignant tumors [12]. A positive prognosis in treatment is observed with the early detection of a malignant tumor. Early diagnostic methods have not yet been developed for these cancer types [13].

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## METHODS

We used a DXR Raman Microscope (Thermo Scientific) with a magnification of 10x, excitation wavelengths of 532 nm, range 80–3200  $\text{cm}^{-1}$ . Each sample of blood plasma was a droplet with a volume of 10  $\mu\text{L}$  placed on a special aluminum plate [11] (Figure 1). This plate has identical holes in the form of a funnel with a diameter of 5 and a depth of 2 mm. We can assume that the distribution of samples in each well and the film thickness of the sample after drying will be the same. The measurements were carried out after complete drying of the sample, which took place for 10 min at room temperature. The Raman spectrum was recorded at three points in the center of the well and averaged. Each spectrum was averaged over 300 scans.



**Figure 1.** Special aluminum plate, top view.

The model of orthotopic xenotransplantation of the U87 human glioblastoma cells into immunodeficient mice SCID line was used [14]. The U87 cell suspension (5  $\mu\text{L}$ , 500000 cells per animal) was introduced in the subcortical brain structures through a hole in the mouse cranium. The control (healthy) mice were injected with the same volume of saline. Animals were removed from the experiment on days 7, 14, 21 and 28 after tumor inoculation. Each group consisted of 10 individuals. The dynamics of the tumor growth was estimated by means of T2-weighted images [15]. The experiments were carried out in compliance with the principles of humanity according to directive 86/609/EEC of the European Community.

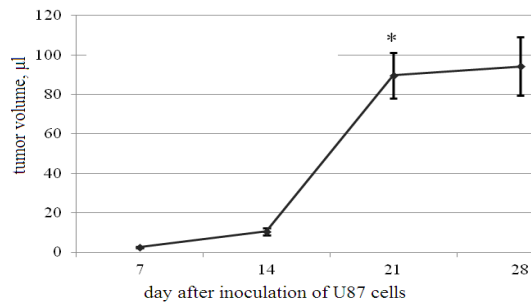
Two tasks were considered in the framework of the analysis of Raman spectra using machine learning methods. First, the difference between the control group and the comparison group was studied with the informative feature selection that has the greatest impact on group separability. Second, the dynamics of changes in Raman spectra for glioma groups with an increase in the duration of tumor development were considered. The question of the influence of the background subtraction procedure on the choice of informative features by the method of principal components analysis (PCA), and Support Vector Machines (SVM) was investigated separately. Details about PCA and SVM can be found in our previous work [16-22]. All computations were made by Python 3.8.2 with sklearn 0.22.2 (PCA and machine learning), rampy 0.4.6 (baseline correction), and scipy 1.4.1 (Savitsky-Golay filtering) libraries.

We used the following data processing pipeline. First, baseline correction was made. Second, PCA with standard scaler and loadings matrix analysis/SVM was applied. For better visualization of loadings matrix, Savitsky-Golay filter was applied with the following parameters window size 51, and polynomial of order 7 selected. SVM was used with default parameters. Results were tested by the Stratified K Fold procedure with 3 splits. Class weights were balanced to make groups of equal size.

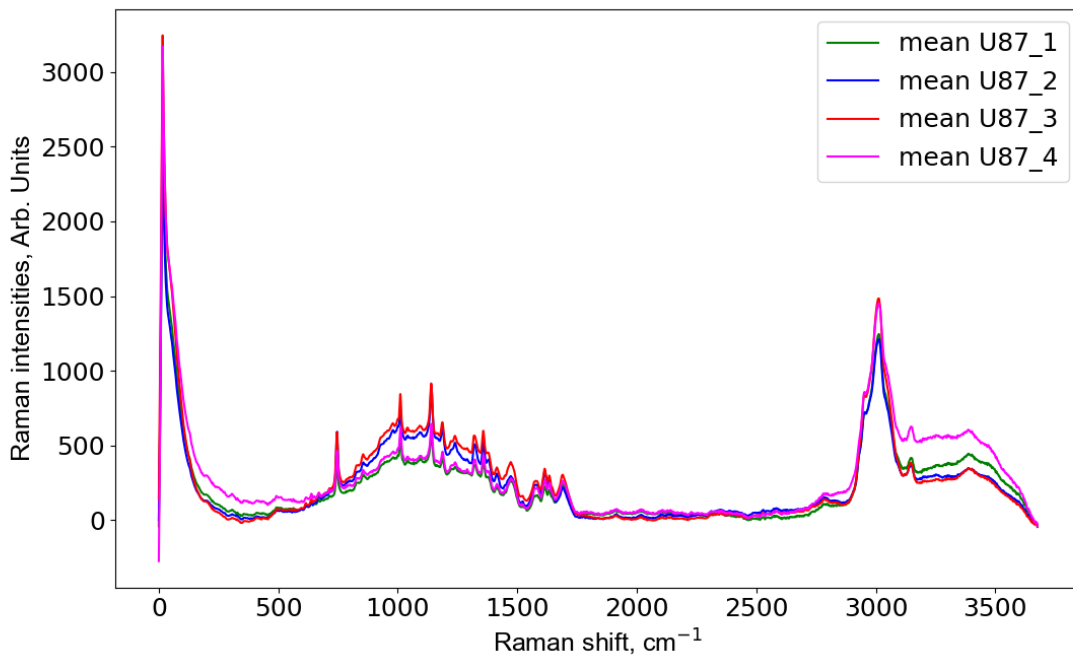
## RESULTS

Figure 2 shows the dynamics of tumor size growth. Significant tumor growth took place from 14 to 21 days after cell inoculation. The intensity of a number of bands in the Raman spectrum changes with an increase in tumor volume. Figure 3 shows the averaged Raman spectra of blood plasma for each group. The most significant changes in the Raman spectra are observed in the 900–1700  $\text{cm}^{-1}$  range. Raman spectra are high-dimensional data. PCA reduces the dimensionality of the data, reveals informative sections of the spectra, and makes it possible to visualize the results,

which makes it possible to evaluate the possibility of creating a data model for differentiating representatives of various experimental groups.

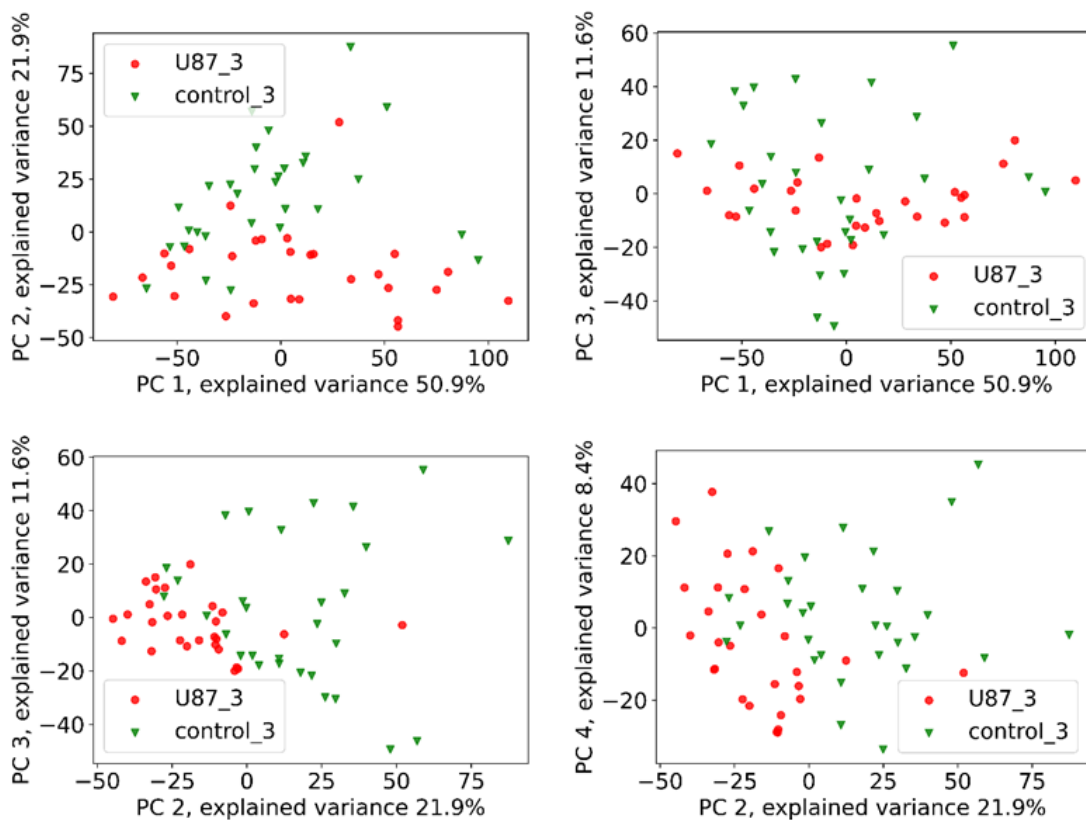


**Figure 2.** U87 glioblastoma volume versus days after inoculation. \*  $p < 0.001$ , one-way ANOVA, LSD-test.

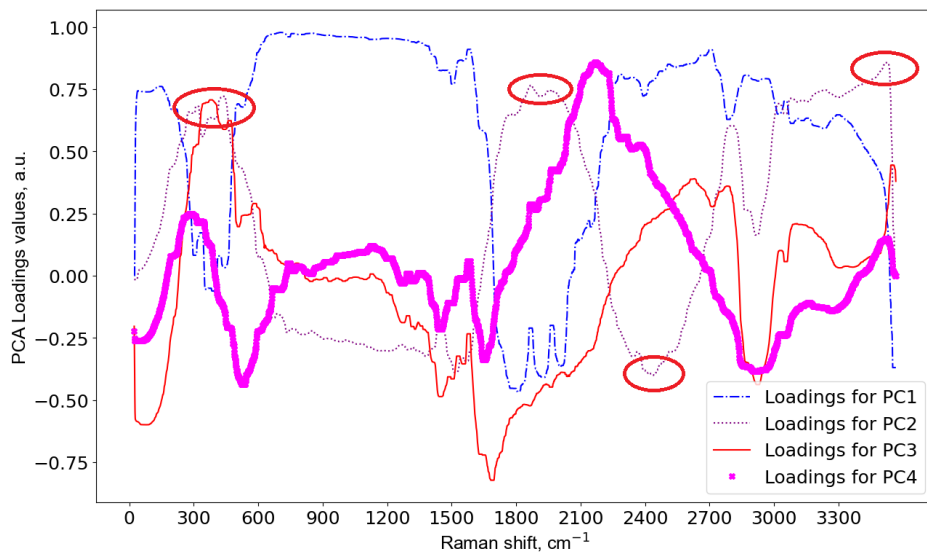


**Figure 3.** Mean Raman spectra of U87 glioblastoma experimental groups.

A significant increase in tumor size is observed on day 21 (3 weeks) after the injection of tumor cells. We will analyze the Raman spectra of this group in more detail. The first four components, which make up 97.8% of the variance, are the most informative and significant for this group. After clarifying the number of principal components containing the largest share of information, possible combinations of the projections of the four principal components onto the plane were constructed (see Figure 4.). As can be seen from Figure 4, the class separability is carried out according to the second principal component. Therefore the main contribution to the group separability will be made by the frequencies corresponding to the second principal component shows that the ranges of 450-500, 1850-1950, 2400-2500  $\text{cm}^{-1}$  will be informative (see Figure 5).



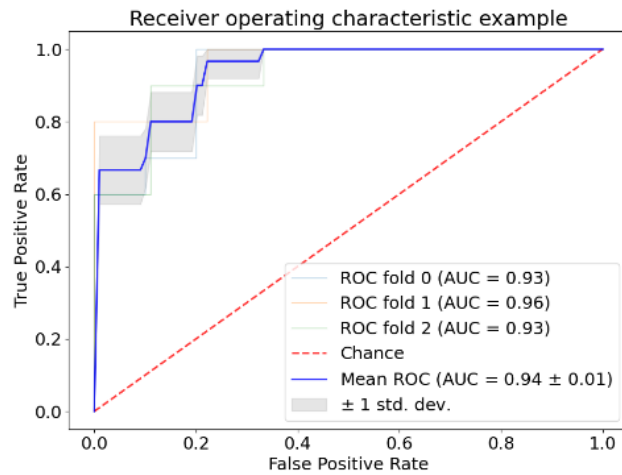
**Figure 4.** The projections of the various principal components on a plane for the third week of the experiment



**Figure 5.** Analysis of the load matrix for the third week of the experiment.

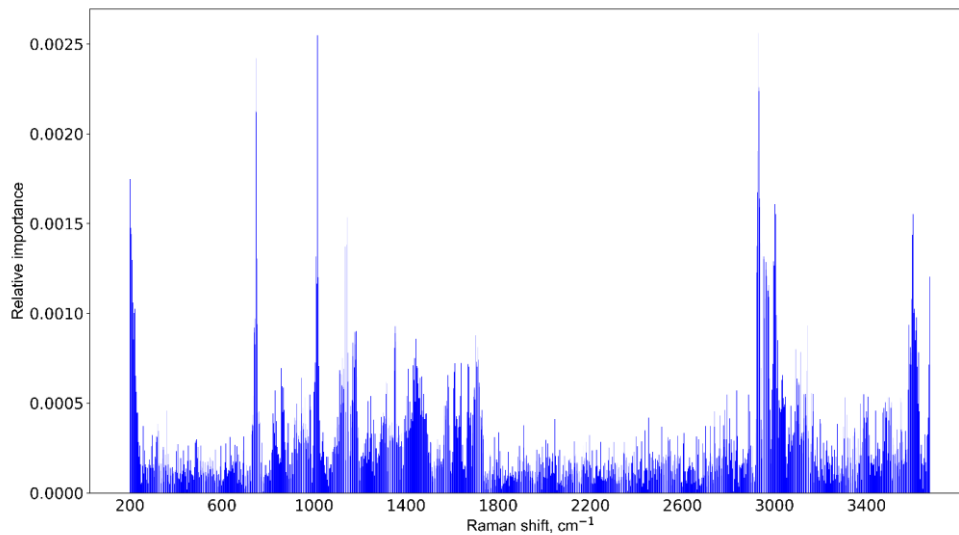
At the next stage, we used Supervised Learning methods to estimate the linear separability of the data, for example, by

using linear kernel SVM [17]. To validate estimation of the data separability, we used a standard approach for Supervised Learning: initial data were randomly divided into 3 splits with the same distribution of the data among classes as in the original dataset. Linear SVM was trained on each split with class weight balancing. Obtained quality metrics such as sensitivity, specificity, accuracy, and precision were averaged. The averaged quality metrics: sensitivity was equal to  $0.86 \pm 0.04$ ; specificity was equal to  $0.83 \pm 0.05$ ; accuracy was equal to  $0.84 \pm 0.04$ . Also, Receiver Operating Characteristic (ROC) and Area Under the Curve (AUC) analyses were performed for each data split and the averaged ones. The mean ROC curve and AUC analysis for the linear SVMs is shown in Figure 6. The results demonstrate the potential of developed models for the separation of control and experimental groups.



**Figure 6.** Mean ROC curve for linear SVMs for the 3 splits.

Also, linear SVM allows obtaining the most informative features based on the proximity of the feature vector to the separating hyperplane. An averaged relative features' importance was estimated for the three train/test splits and linear SVMs. Figure 7 illustrates the informative frequencies for this classifier. These are 700, 1000, 1180, 1500, 1770, 3000  $\text{cm}^{-1}$ .



**Figure 7.** Mean the informative frequencies for the third week of the experiment.

## CONCLUSION

We studied the blood plasma of mice in the dynamics of the development of U87 glioblastoma. Machine learning methods made it possible to conduct an analysis to separate the experimental and control groups in terms of sensitivity, specificity, and accuracy. The best models were obtained for a period of 21 days after inoculation of tumor cells when there is the greatest difference in the Raman spectra of control and experimental animals. The analysis of load matrices was carried out, and the most informative frequencies associated with specific metabolites, the concentration of which changes in the blood in the dynamics of tumor growth, were determined.

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