



RESEARCH ARTICLE

Biochemical profile, antioxidant effect and antifungal activity of Saudi Ziziphus spina-christi L. for vaginal lotion formulation

Hallouma Bilel1*, Hani Mohamed Awad Abdelzaher2 & Shaima Mohamed Nabil Moustafa2

- ¹Department of Chemistry, College of Science, Jouf University, Sakaka 72341, Saudi Arabia
- ²Department of Biology, College of Science, Jouf University, Sakaka 72341, Saudi Arabia

*Email: hbilel@ju.edu.sa



ARTICLE HISTORY

Received: 08 January 2022 Accepted: 31 August 2022

Available online Version 1.0:05 November 2022



Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS etc.
See https://horizonepublishing.com/journals/index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an openaccess article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (https://creativecommons.org/licenses/by/4.0/)

CITE THIS ARTICLE

Bilel H, Abdelzaher H M A, Moustafa S M N. Biochemical profile, antioxidant effect and antifungal activity of Saudi *Ziziphus spina-christi* L. for vaginal lotion formulation. Plant Science Today (Early Access). https://doi.org/10.14719/pst.1659

Abstract

Ziziphus spina-christi L. extract from the northern region of Saudi Arabia, was investigated to determine its chemical composition and to evaluate its antioxidant and antifungal properties. Fresh leaves were extracted using Soxhlet apparatus and the yield was 8% w/w. Results of the qualitative study showed that this extract is rich in chemical compounds belonging to several classes (saponins, phenols, tannins). GC-MS analysis detected 38 chemical compounds with different concentrations representing 99.71 % of the total extract. However, Z. spina-christi leaves extract is mainly composed of Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethylester (18.80%). The extract has free radical scavenging activity at different concentrations and the best result was obtained with IC₅₀ of 148.33 µg/ml. C. albicans and other Candida species caused vulvovaginal candidiasis, which is a high-risk occurrence in hospitalized patients. In vitro antifungal activity was investigated by the agar well diffusion test to measure and compare diameter of zones of inhibition (in mm) against Candida albicans, Candida glabrata and Candida tropicalis. Ethanolic extract of Z. spina-christi demonstrated a substantial inhibitory impact on several Candida species, especially against C. glabrata which has the highest inhibitory effect (90%). Therefore, Saudi Z. spina-christi leaves extract is a source of natural antioxidants and it can be used as well antifungal pharmaceutical product.

Keywords

Antioxidant activity, *Candida albicans*, Pharmaceutical preservative, Fresh leaves extract, GC-MS analysis

Introduction

In hospitalized patients, vulvovaginal candidiasis is a high-risk occurrence. *Candida albicans* and other *Candida* species were known as responsible for the great majority of vulvovaginal candidiasis cases (1). Fungal infection is more likely in those with several predisposing conditions, such as diabetes, cellular urine catheters, antibiotics and corticosteroid users (2). Due to a lack of acceptable treatment choices and pathogen cross-resistance to earlier medications (fluconazole and itraconazole), researchers are looking for novel antifungal agents from a variety of sources, including medicinal plants (3, 4).

Candidiasis can be caused by a variety of yeast species in the genus *Candida* (5) They are part of the skin's, mucous membranes and gastrointestinal tract's regular biota. All humans' mucosal surfaces are colonized by *Candida* species during or shortly after birth, whereas the possibil-

ity of endogenous infection is always present. Subsequently, the most common systemic mycosis is candidiasis. Difficulties that arise during *C. albicans* chemotherapy need the development of innovative treatment techniques (6).

Plants have been used from ancient times in folk medicine. They are important sources to develop therapeutic products for health care (7). Medicinal plants are rich in therapeutically bioactive molecules such as flavonoids, alkaloids, coumarins, tannins and terpenes. These compounds are of great interest as sources of natural antioxidants and are recognized by their diverse biological activities (8).

Z. spina-christi tree (family Rhamnaceae) grows in warm-dry areas, largely cultivated in South and West Asia, North and East Africa and the Middle East (9). It is of great concern for the reason that fruits, seeds, bark, roots and leaves have been used in traditional medicine for the treatment of several diseases (10). This plant is rich in biologically active components like flavonoids, terpenoids, vitamins, polysaccharides, polyphenols and tannins and is commonly used in medicine as antimicrobial, antioxidant, anti-inflammatory, antifungal, analgesic, sedative, anticancer, hypoglycemic and reducing cholesterol agent (11, 12). Fruits of Z. spina-christi are consumed as a source of energy due to their richness in carbohydrates (13). Moreover, leaves of Z. spina-christi had biological applications and can be used for their antipyretic, anti-diarrhoeal, immunomodulator and anti-fertility activities (10). In Arabic countries, Z. spina-christi tree has historical, religious and medicinal interests. In Saudi Arabia, Z. spina-christi tree is still growing and is distributed in different regions of the Kingdom (14). In traditional Arabic medicine, it is used to treat diarrhea, ulcers and fevers (15). Oxidative stress causes an alteration of the constituents of human cells and is responsible for premature cellular aging. In general, medicinal plants can be used to protect and fight against oxidative damage in the biological system, also to maintain the anti-oxidant system balance to prevent chronic degenerative diseases (16).

In this context, the purpose of this research is to provide a comprehensive study about Saudi *Z. spina-christi* leaves grown on Al-Jouf region, to determine the chemical profile, to evaluate the antioxidant and the *in vitro* antifungal activities of *Z. spina-christi* extract for the treatment of a variety of vaginal infections and inflammatory illnesses using natural substances.

Materials and Methods

Plant material

Fresh leaves of *Z. spina-christi* L. were collected in March 2021 from Sakaka city a region in Aljouf located the north of KSA (latitude: 29.953894, longitude: 40.197044, 29° 57' 14.0184" N and 40° 11' 49.3584" E) and were stored in plastic bags in the dark for further use. Then, the plant identified by Doctor Ben Amor botanist in the faculty of sciences, Gafsa, Tunisia.

Leaves extraction

Extract was prepared from fresh leaves of *Z. spina-christi* using Kimax * Soxhlet extractor apparatus. Fifteen gms of fresh leaves were placed in the Soxhlet extractor and then 250 ml of ethanol were added to the distillation flask. After refluxing for 8 hr, the solvent was evaporated under reduced pressure and the concentrated extract was stored at 4 °C in obscurity until the beginning of the analysis. Extraction was done in triplicate and the yield was 8% w/w (17).

Qualitative phytochemical determination

Qualitative analysis was carried out to detect the presence or absence of alkaloids, glycosides, saponins, phenols, tannins, steroids, terpenoids, anthraquinone and flavonoids according to the common methods described in the literature (18).

Quantitative chemical determination by GC-MS

Fresh leaves extract of *Z. spina-christi* was analyzed using Shimadzu GC-MS- QP2010SE single quadrupole apparatus. GC was equipped with SLB-5MS capillary column (30 m x 0.25 mm; thickness= 0.25 μ m). Injector temperature was set at 270 °C and the oven temperature increase from 40 °C to 220 °C (4 °C/min), kept for 10 min then up to 280 °C (5 °C/min). Detector temperature was 270 °C and Helium (carrier gas of 99.99% purity) was used at a flow rate 1 ml/min. A mass spectrum was recorded at energy of ionization of 70 eV. The total analysis time was 120 min and components were identified based on the comparison of their retention time and mass fragmentations patterns with those of standards data of WILEY and NIST libraries (19).

Evaluation of in vitro antioxidant activity

The anti-free radical activity of DPPH of leaves extract of *Z. spina-christi* was determined based on the standard assays with some modifications (20). Thus, at different concentrations, 1 ml of each tested extract was added to 2 ml of DPPH osolution (0.1 mM). After vigorous stirring, the mixture is incubated for 30 min in the dark and at room temperature and then the absorbance was measured at 515 nm by a UV visible spectrophotometer (JASCO-V530).

The estimated anti-free radical activity was expressed by the value of the percentage inhibition (% I) calculated using the following formula:

% $I=[(A_0 \text{ blank-}A_1 \text{ sample})/A_0 \text{ blank}] * 100$

 A_0 blank is the absorbance of the control reaction containing all reagents except the tested extract (1 ml of ethanol and 2 ml of DPPH) and A_1 sample is the absorbance of the tested sample. Trolox was used as a positive control.

Antioxidant activity was expressed as IC50 ($\mu g/ml$) which represent the extract concentration providing 50% inhibition, calculated from the graph plotting inhibition % against sample concentration. A Low IC50 value means a high antioxidant activity of the extract. Tests were carried out in triplicate.

Evaluation of in vitro antifungal activity

Collection of samples

A sterile swab was used to collect vaginal fluids from females with different ages from (25 to 50) years old. Under sterile conditions, 3 specimens were taken simultaneously, one for light microscopic investigation and the other for fungal culture. Materials were inoculated with Sabouraud's Dextrose Agar and incubated at 35 °C until colonial appearance. Fungal cultures were maintained in Sabouraud's Dextrose Agar at 5 °C followed by serial subculturing every 3 months. All fungi were kept at the Biology Department, College of Science, Jouf University with a number of *C. albicans* (JU 01032), *C. parasilopses* (JU 01033) and *C. tropicalis* (JU 10134).

Identification of isolated fungi

Candida spp. were identified using morphological and physiological methods such as growth characteristics and carbon source assimilation or fermentation, as well as the appearance on differential isolation media. According to the manufacturer's recommendations, the HiCandida identification kit was used to accurately identify *Candida* species. A plastic strip had twelve wells containing sterile media for several biochemical assays as follows: well 1, medium for urease detection, and wells 2-12, medium for carbohydrate utilization (with 11 different sugars in respective wells, including, melibiose, lactose, maltose, sucrose, galactose, cellobiose, inositol, xylose, dulcitol, raffinose and trehalose) (21, 22).

The homogeneous yeast suspension (10⁶ cells/ml) was produced and injected into kit wells, then incubated for 24-28 hrs at 22.5 2.5 °C. The color of the kit changed after the incubation period: well 1 containing urease was considered positive if the yellow color changed to pink. If the color of wells 2-12 changed from orange to yellow after 72 hrs, the result was considered positive; if the color remained orange, the result was regarded as negative. Findings were interpreted according to the manufacturer's guidelines.

Well Diffusion Assay

Inhibitory zones of *Z. spina-christi* leaves extract were tested using the well assay technique against *C. albicans*, *C. glabrata* and *C. tropicalis* strains to determine the effective concentration. Overnight inoculum of *Candida* spp. were disseminated over Sabouraud's dextrose agar media and 1 ml of different concentrations of the extracts (5, 10, 15 and 20 mg/ml) were added to each well (10 mm diam.) and incubated at 26 °C for 48 hrs (23, 24). As a positive control, miconazole was utilized (25).

Statistical Analysis

Analysis of variance as One-way (ANOVA) and Statistical Package for Social Sciences (SPSS) software version 12.0 were conducted for statistical analysis of the obtained results. Antioxidant and antifungal activities were carried out in 3 experiments. Differences among the mean values of the various treatments were determined by the least significant difference test. A probability level of P < 0.05 was used in testing the statistical significance of all experimental data (26).

Results and Discussion

Qualitative phytochemical determination

The qualitative analysis showed that the leaves extract of *Z. spina-christi* was rich in phytochemicals belonging to different classes as illustrated in Table 1.

Table 1. Phytochemical screening of *Z. spina-christi* leaves extract.

ı	Phytochemicals	Extract
-	Alkaloids	+
(Glycosides	+
9	Saponins	++
F	Flavonoids	-
F	Phenols	++
1	Tannins	++
9	Steroids	+
1	Terpenoids	+
A	Anthraquinone	-

⁺⁺ phytochemical detected at appreciable amount; + phytochemical detected at trace amount; - phytochemical not detected

This analysis was based on the study of the presence or absence of several phytochemicals. Alkaloids, glycosides, saponins, phenols, tannins, steroids and terpenoids were detected with different amounts, while anthraquinone and flavonoids were absent. To obtain a more detailed result on the phytochemical composition and their %, this study will be completed by an analysis using the GC-MS technique.

Quantitative chemical determination by GC-MS

The chemical composition of green leaves extract obtained from *Z. spina-christi* was identified and quantified by GC-MS technique. Analysis of the obtained result shows that the extract is rich in chemical compounds. Indeed, it contains 38 phytochemical constituents with different diverse chemical groups representing 99.71 % of the total extract (Table 2).

Table 2. Identified volatile compounds of *Z. spina-christi* leaves extract.

		·	
Peak	RT (min)	Chemical Compound	Peak Area (%)
1	11.684	N-[(E)- 3- methyl-2-butenylidene] methanamine	0.56
2	13.314	Isooctanol	0.73
3	16.428	2-tridecanol	0.64
4	16.559	Diethyl phthalate	0.45
5	19.845	11-Dodecenol	1.53
6	20.160	1-Ethynylcyclopentanol	0.52
7	20.396	Dodeca-1,6-dien-12-ol	0.62
8	21.402	n-hexadecanoic acid	0.60
9	21.465	Dibutyl phthalate	0.75
10	23.211	Phytol	2.74
11	23.520	1,2,3,4-tetrahydrostyrene	0.74
12	24.008	Hexadecanoic acid butylester	1.45

13	26.146	Octadecanoic acid butyl ester	1.32
14	27.085	2,4-Difluorophenol	0.39
15	27.252	1-Chloroheptacosane	1.31
16	27.431	Hexadecanoic acid, 2-hydroxy-1- (hydroxymethyl)ethylester	3.20
17	27.789	1-(4-bromobutyl)-2-piperidinone	0.53
18	28.217	Pentatriacontane	0.82
19	29.014	7-hydroxy-3-(1,1-dimethylprop-2-enyl) coumarin	1.12
20	29.145	Tritetracontane	2.10
21	29.286	3-chloropropionic acid octadecyl ester	1.61
22	29.428	Octadecanoic acid, 2-hydroxy-1- (hydroxymethyl)ethylester	18.80
23	30.046	14B-Pregnane	3.85
24	30.366	Squalene	5.94
25	30.943	Carbonic acid,isobutyl octadecyl ester	4.53
26	31.173	Diethylmethylborane	1.60
27	31.278	8-nitro-12-tridecanolide	1.88
28	31.750	Sulfurous acid, octadecyl 2-propyl ester	6.64
29	31.912	(cis)/trans-2-(2,2-dimethylpropanoyl)-5-methylcyclopentanone	1.40
30	32.300	Bacchotricuneatin c	4.37
31	32.463	1-bromo-11-iodoundecane	2.81
32	32.636	17-pentatriacontene	5.80
33	33.029	Vitamin E	9.70
34	33.228	(8R, 12R)-8,12-Epoxy-13,14-dihydroxy-labdane	3.31
35	34.051	HAHNFETT	1.27
36	34.392	Oxalic acid, allyl pentadecyl ester	0.67
37	35.052	Beta- sitosterol	2.35
38	36.127	1,7-dimethyl-4-(1-methylethyl)cyclodecane	1.06

Detailed chemical analysis shows that the extract was mainly composed by Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethylester (18.80%), followed by Vitamin E (9.70%), Sulfurous acid, octadecyl 2-propyl ester (6.64%), Squalene (5.94%), 17-pentatriacontene (5.80%), Carbonic acid, isobutyl octadecyl ester (4.53%), Bacchotricuneatin c (4.37%), 14B-Pregnane (3.85%), (8R, 12R)-8,12-Epoxy-13,14 -dihydroxy-labdane (3.31%), Hexadecanoic acid, 2-hydroxy -1-(hydroxymethyl) ethylester (3.2%), 1-bromo-11iodoundecane (2.81%), Phytol (2.74%), Beta- sitosterol (2.35%), Tritetracontane (2.10%), 8-nitro-12-tridecanolide (1.88%), 3-chloropropionic acid octadecyl ester (1.61%), Diethylmethylborane (1.60%), 11-Dodecenol (1.53%), Hexadecanoic acid butylester (1.45%), (cis)/trans-2-(2,2dimethylpropanoyl)-5-methylcyclopentanone (1.40%),Octadecanoic acid butyl ester (1.32%), 1-Chloro heptaco-Hahnfett (1.27%), 7-hydroxy-3-(1,1-(1.31%),dimethylprop-2-enyl) coumarin (1.12%) and 1,7-dimethyl-4-(1-methylethyl) cyclodecane (1.06%). The other compounds were detected in traces (less than 1%).

The chemical analysis of leaves extract of *Z. spina-christi* showed that fatty ester derivatives (32.78%), hydrocarbons (28.27%) and alcohols (19.22%) were the dominant compounds and represent 80% of the total bioactive constituents (Table 3).

Table 3. Classification of the volatile compounds of *Z. spina-christi* leaves extract according to the chemical classes.

S.No	Chemical classes	Percentage (%)
1	Esters	32.78
2	Hydrocarbons	28.27
3	Alcohols	19.22
4	Sulfur containing compounds	6.64
5	Pyrans	4.37
6	Ketones	3.05
7	Nitrogen containing compounds	2.44
8	Others	2.34
9	Acids	0.6
	Total	99.71

Among the identified phytochemicals, Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethylester was found to be the most dominant compound which is a fatty acid ethyl ester of glycerol derivative belonging to fatty acid esters secondary metabolites. A recent study has shown that Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethylester can be used for the treatment of C. Violaceum infections and can be evaluated for other pharmacological activities (27). Also, a theoretical predictive study showed that Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethylester has interesting biological activities such as All-trans -retinyl-palmitate hydrolase inhibitor, Lipid metabolism regulator, Eye irritation, inactive, Antieczematic, CYP2J substrate, Acylcarnitine hydrolase inhibitor, CYP2J2 substrate, Linoleate diol synthase inhibitor, Lipoprotein lipase inhibitor, GST A substrate, Macrophage colony-stimulating factor agonist, Alkenylglycerophosphocholine hydrolase inhibitor, Phosphatidylglycerophosphatase inhibitor (28).

Furthermore, some other identified phytochemicals have been reported to have interesting biological properties as antibacterial, antioxidant, analgesic, antiviral, hypoglycemic, anti-inflammatory, antifertility and more as shown in Table 4.

Table 4. Biological activities of some phytochemicals identified from *Z. spina-christi* leaves extract according to Dr. Duke's Phytochemical and Ethnobotanical databases.

S. No	Name of compounds	Biological Activity
1	Vitamin E	Analgesic, Antiaggregant, Antiagiant, Antialzheimeran, Antiatherosceloric, Antidecubitic, Antifibrostic, Antihereptic, Anti-inflammatory, antioxidant, Antitumor
2	Squalene	Antibacterial, antitumor, cancer- preventive, Immunostimilant, lipoxygenase inhibitor, perfum- ery, pesticide, sunscreen

Hexadecanoic acid. 2-hydroxy-Pesticide, hemolytic, flavor, anti-1-(hydroxymethyl)ethylester oxidant. Phytol Cancer-preventive Anorexic. Antibacterial. anticancer, antiestrogenic, antifertili-5 Beta- sitosterol ty, anti-inflammatory, antioxidant, antiviral, hypoglycemic.... Hexadecanoic acid butylester Antimicrobial, antioxidant 1,7-dimethyl-4-(1-methylethyl) 7 Cytotoxic cyclodecane Pentatriacontane Herbistat Anti-oxidant, Hypocholesterolemic. Nematicide, Anti-androgenic, n-Hexadecanoic acid Hemolytic, Pesticide, Lubricant, 5 -Alpha reductase inhibitor, antipsychotic.

The study of the bioactive components of leaves methanolic extract of *Z. spina-christi* from the campus of South Valley University, Qena, Egypt showed the presence of 13 components, while Phenol, 2,5-bis (1,1-dimethylethyl) (40.24%) and Decane, 2-methyl-(18.53%) were the most abundant components (29). Comparing to our results, the difference of chemical composition may be due to geographic conditions (humidity, temperature, altitude), the origin and the period of leaves harvest and the soil-growth conditions (30).

Evaluation of in vitro antioxidant activity

The antioxidant activity was evaluated by studying the reducing effect of different concentrations of *Z. spina-christi* leaves extract on DPPH radical compared to those of Trolox (positive control). The obtained results show that the extract has a great free radical scavenging activity with IC_{50} of 148.33 µg/ml (Fig. 1).

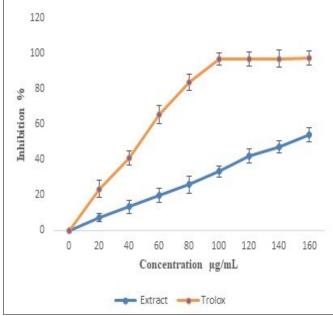


Fig. 1. Antioxidant activities of *Z. spina-christi* leaves extract at different concentrations. Trolox was used as a positive control.

A study of different *Z. spina-christi* leaves extract from five provenances in Saudi Arabia showed that our results were in consistence with those obtained from Mec-

ca Road Provenance (0.15 mg/ml), while Al-Taif, Riyadh and Jeddah provenances have a lower value of IC50 (0.06, 0.08 and 0.07 respectively) (31).

The chemical analysis shows the richness of *Z. spina -christi* leaves extract by bio-actives compounds belonging to different classes such as esters, hydrocarbons, alcohols, ketones, nitrogen and sulfur compounds and fatty acids which might be the origin of a good antioxidant capacity as Vitamin E (32), Squalene (33), 14B-Pregnane (3.85%) (34, 35), Bacchotricuneatin c (36), Phytol (37), Beta-sitosterol (38), Hexadecanoic acid butylester, (39) and Hexadecanoic acid (40).

The extract can be applied as a natural supplement in the field of the food industry due to its antioxidant activity.

Evaluation of in vitro antifungal activity

From 40 vaginal swab samples that were collected from patients with vaginitis, 28 samples were positive and 12 samples were negative, as well as the frequency of isolates according to a woman's age was described in Table 5. Identification tests for all isolates reproach the following species: *C. albicans* 17 isolates, (42.5 %), *C. glabrata*, 9 isolates, (22.5%), and *C. tropicalis* 14 isolates (35%), respectively. Statistical analysis for age groups (25, 30, 35, 40 and 45) showed non-significant differences between means (p \leq 0.05) (Table 5). It was obvious that the majority of isolates belonged to the genus *C. albicans*, followed by *C. tropicalis* and *C. glabrata* respectively. The less frequent yeast species was *C. glabrata*.

Table 5. Statistical analysis of the relationship between age groups and Candida isolate.

Candida spp.	Age groups (8 samples/each group)				
	21- 25	26- 30	31- 35	36- 40	41- 45
C. albicans	5±0.3	2±0.34	3±1.54	4±0.32	3±1.54
Candida glabrata	0	1	3±0.98	0	3±.05
Candida tropicalis	3±1.43	5±0.24	2±0.54	4±0.54	2

Values are given as mean \pm SD of three replications.

Statistical significance of differences between the means of groups: No Significant at p < 0.05 according to ANOVA test.

Using an agar well diffusion test, antifungal effects of different concentrations of *Z. spina-christi* leaves extract on *Candida* spp. were evaluated in Table 6, indicating different concentration manifested inhibition zones with different diameters on Sabouraud's dextrose agar medium.

It has been shown that the concentration of 20 mg/ml of ethanolic *Z. spina-christi* leaves extract displayed the strongest antagonist effect against *C. glabrata, C. tropicalis and C. albicans* by 77, 75 and 70 mm respectively. Followed by a concentration of 15 mg/ml of extract appeared a moderate effect against *C. glabrata, C. albicans* and *C. tropicalis* (62, 60 and 58 mm respectively). Then, 5 mg/ml of extract gave the lowest effect, when compared with the control (Table 6 and Figs. 2-3).

Table 6. Mean zones of inhibition (mm) of *Z. spina-christi* leaves extract against *candida* spp.

_	Mean zone of inhibition (mm)				
Species	5 mg/ml	10 mg/ml	15 mg/ml	20 mg/ml	
Miconazol	60 ± 0.02	70 ± 0.18	80 ± 0.10	80 ± 0.32	
C. albicans	50 ±0.20	56 ± 0.16	60 ± 0.13	70 ± 1.9	
Candida glabrata	48 ± 0.12	54 ± 0.14	62 ± 0.24	77 ± 0.6	
Candida tropicalis	48 ± 0.25	56 ± 0.20	58 ± 0.5	75 ± 0.31	

Values are given as mean \pm SD of three replications. Statistical significance of differences between the means of groups: Highly Significant at p <0.05 according to ANOVA test

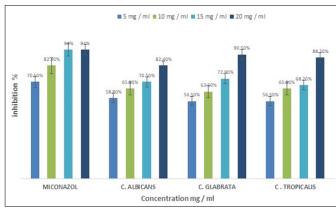


Fig. 2. Inhibition % of *Candida* spp. exposed to different concentrations of *Z. spina-christi* leaves extract on Sabouraud's dextrose agar at 35 $^{\circ}$ C for 48 hr in the dark.

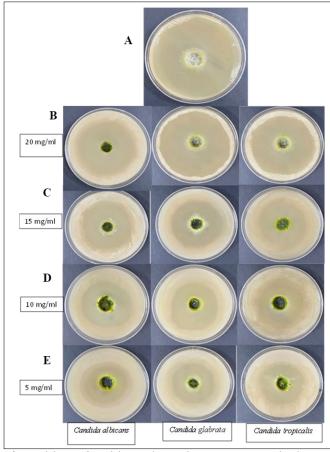


Fig. 3. Inhibition of *Candida* spp. **A)** Control containing miconazole, **B)** Treated containing *Z. spina-christi* leaves extract (20 mg/ml), **C)** Treated containing *Z. spina-christi* leaves extract (15 mg/ml), **D)** Treated containing *Z. spina-christi leaves extract* (10 mg/ml) and **E)** Treated containing *Z. spina-christi leaves extract* (5 mg/ml) on Sabouraud's dextrose agar media at 26 °C for 48 hr at the dark.

According to microscopy, *Z. spina-christi* leaves extract at 20 mg/ml caused significant changes in the shape and density of *Candida* spp. mycelia. When compared to the control, Fig. 4 demonstrates more morphological changes such as mycelium deformation, perforation, cell lysis and mycelium destruction.

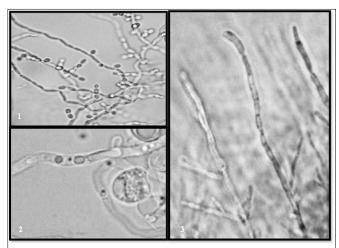


Fig. 4. Represented microscopic examination (magnification: X 10) of antagonistic effect *Z. spina-christi* leaves extract (20 mg/ml) against *C. albicans*. (1) Healthy mycelium. (2 and 3) Abnormal structure, lysis and coagulation of mycelium structure.

Several studies have highlighted the antifungal properties of the *Ziziphus* genus. It was demonstrated that the ethanolic extract of *Z. spina-christi* has antifungal efficacy against *Candida* spp. Confirmation was on the antifungal efficacy of *Z. spina-christi* extract against fungus strains in another study (41, 42). According to one report, methanolic and ethanolic extracts of *Z. Spina-christi* displayed antifungal efficacy against *Candida* spp. The microemulsion of ethanolic and methanolic extracts showed to be more fungicidal than ethanolic and methanolic extracts against *C. albicans* (43).

It was showed that aqueous extract of *Ziziphus* sp. can control the growth of *Alternaria brassicae* and *Fusarium oxysporum* (44).

Previous research has found a link between the antifungal properties of plant extracts and the solvents used; the polar extract contains saponins and glycosylated flavonoids, whereas the non-polar extract contains non-polar components such as terpenoids (45). Alkaloids, glycosides, saponins, phenols, tannins, steroids and terpenoids were detected with different amounts in *Z. spina-christi* leaves extract (Table 1) is thought to be responsible for its dominating action. Some of these chemicals, particularly terpenes were previously described to have antibacterial, fungicidal and insecticidal properties (46-49).

By entering between the fatty acyl chains, terpenes have been shown to alter the fungal cell permeability. Furthermore, terpenes impede Candida's respiratory chain, implying negative effects on mitochondria (50, 51). Extracts of *Z. spina-christi* were found to reduce *C. albicans* biomass by raising glucose levels and decreasing cell dry weight. This process might be the result of cell wall breakdown and subsequent sterilization (52). As a result, it is possible that the ethanolic extracts increased antifungal

activity and served as a catalyst for extract penetration through the fungal cell wall. These support the use of *Z. spina-christi* extract to treat yeasts that attack vagina.

Conclusion

This study demonstrated that the extract of *Z. spina-christi* L. grown in is rich in bioactive compounds and predominantly by Octadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethylester. *Z. spina-christi* leaves extract provide a good antioxidant activity, which makes it possible to use in food industry. Furthermore, this extract can be used to treat a variety of infections and inflammatory disorders caused by *C. albicans* and *C. tropicalis*. These results indicate that this raw material could be used in pharmaceutical formulation as a vaginal lotion.

Acknowledgements

The authors extend their appreciation to the Deanship of Scientific Research at Jouf University for funding this work through research grant no (DSR-2021-03-0101).

Authors contributions

SMNM performed the isolation, identification of fungal and writing the antifungal section. HB prepared the plant extract, phytochemical analysis, antioxidant activity and writing the chemical section. HMAA completed the statistical analysis and editing the final draft. All authors read and approved the final manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interests to declare.

Ethical issues: None.

References

- Kiasat N, Rezaei-Matehkolaei A, Zarei MA, Hamidavi MK, Molavi S, Khoshayand N. Prevalence of vulvovaginal candidiasis in Ahvaz, Southwest Iran: A Semi-Large Scale Study. Jundishapur J Microbiol. 2019;12(3):1-6. http://dx.doi.org/10.5812/jjm.89815
- Zarei MA, Rezaei-Matehkolaei A, Navid M, Torabizadeh M, Mazdarani S. Colonization and antifungals susceptibility patterns of Candida species isolated from hospitalized patients in ICUs and NICUs. J nephropathol. 2015; 4 (3):77-84. https://dx.doi.org/10.12860%2Fjnp.2015.15
- Sojakova M, Liptajova D, Borovsky M, Subik J. Fluconazole and itraconazole susceptibility of vaginal yeast isolates from Slovakia. Mycopathologia. 2004;157 (2):163-69. https:// doi.org/10.1023/b:myco.0000020594.35357.b0
- 4. Raesi V A, Mahdavinia M, Kalantari H, Khoshnood S, Shirani M. Antifungal effect of the effect of *Securigera securidaca* L. vaginal gel on *Candida* species. Curr Med Mycolgy. 2019;5 (3):31-35. https://doi.org/10.18502/cmm.5.3.1744
- Ciurea CN, Kosovski IB, Mare AD, Toma F, Pintea-Simon IA, Man A. Candida and Candidiasis—Opportunism Versus Pathogenicity: A Review of the Virulence Traits. Microorganisms. 2020;8(6):1-17. https://doi.org/10.3390/microorganisms8060857

- De Oliveira Santos GC, Vasconcelos CC, Lopes AJO, Cartagenes MDS, Filho AKDB, Do Nascimento FRF et al. *Candida* infections and therapeutic strategies: Mechanisms of action for traditional and alternative agents. Front Microbiol. 2018;9:1-23. https:// doi.org/10.3389/fmicb.2018.01351
- Petrovska BB. Historical Review of Medicinal Plants' Usage. Pharmacogn Rev.2012;6(11):1-5. https://dx.doi.org/10.4103% 2F0973-7847.95849
- 8. Busia K. Fundamentals of Herbal Medicine: History, Phytopharmacology and Phytotherapeutics Vol. 1. Xlibris UK. 2016.
- Mariod AA, Saeed Mirghani ME, Hussein I. Z. spina-christi (Christ's Thorn Jujube). In: unconventional oilseeds and oil sources. Elsevier; 2017. p. 243-49. https://doi.org/10.1016/B978-0-12-809435-8.00036-6
- Soni H, Jitender K M. Phyto-Pharmacological Potential of Zizyphus Jujube: A Review. Sch Int J Biochem.2021;4(1):1-5. https:// doi.org/10.36348/sijb.2021.v04i01.001
- 11. Saaty AH. Review of the nutritional values and biological activities of *Z. spina-christi* (Sidr) plant extract. American J Food Nutr. 2019; 7(4):166-72.
- El Cadi H, El Bouzidi H, Selama G, El Cadi A, Ramdan B, El Majdoub YO et al. Physico-chemical and phytochemical characterization of moroccan wild Jujube 'Zizyphus lotus (L.)' fruit crude extract and fractions. Molecules.2020;25(22):1-17. https://doi.org/10.3390/molecules25225237
- Mariod AA. Wild fruits: Composition, nutritional value and products. Springer Nature Switzerland AG. 2020. https://doi.org/10.1007/978-3-030-31885-7
- Naghmouchi S, Moodi A. Biochemical profile, antioxidant capacity and allelopathic effects from five *Ziziphyus spina-christi* (L.) provenances growing wild in Saudi Arabia. Not Bot Horti Agrobot Cluj Napoca. 2020;48(3):1600-12. https://doi.org/10.15835/nbha48312025
- 15. Hasan NM, AlSorkhy MA, Al Battah FF. *Ziziphus jujube* (Ennab) of the Middle East, food and medicine. Unique J Ayurvedic Herb Med. 2014; 2(6):7-11.
- Hassan W, Noreen H, Rehman S, Gul S, Kamal MA, Kamdem JP et al. Oxidative stress and antioxidant potential of one hundred medicinal plants. Curr Top Med Chem. 2017;17(12): 1336-70. https://doi.org/10.2174/1568026617666170102125648
- 17. Bilel H, Elsherif MA, Moustafa SMN. Seeds oil extract of *Mesembryanthemum forsskalii* from Aljouf, Saudi Arabia: Chemical composition, DPPH radical scavenging and antifungal activities. OCL..18-27:10;2020 https://doi.org/10.1051/ocl/2020005
- 18. Biqiku L, Lupidi G, Petrelli D, Vitali LA. Antimicrobial activity of single and combined extracts of medicinal plants from Cameroon. IOSR J Pharm Biol Sci. 2016;11(4):86-90. http://dx.doi.org/10.9790/3008-1104048690
- Adams RP. Identification of essential oil components by gas chromatography/mass spectromety. Carol Stream, Allured, IL. 2001.
- Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. LWT - Food Sci Technol. 1995;28(1):25-30. https://doi.org/10.1016/S0023-6438(95) 80008-5
- Maqsood I, Masood MI, Bashir S, Nawaz HM, Anjum AA et al. Preparation and in vitro evaluation of Nystatin micro emulsion based gel. Pak J Pharm Sci. 2015;28 (5):1587-93.
- Akram A, Akhtar N, Waqas MK, Rasul A, Rehman KU et al. Development, characterization and evaluation of ginger extract loaded microemulsion: *In vitro* and *Ex vivo* release studies. Pak J Pharm Sci. 2019;32(4):1873-77.
- Magaldi S, Mata-Essayag S, Hartung de Capriles C, Perez C, Colella MT et al. Well diffusion for antifungal susceptibility test-

- ing. Int J Infect Dis. 2004;8(1): 39-45. https://doi.org/10.1016/j.ijid.2003.03.002
- Sadeghi NB, Rajabi M, Zarei MA, Zarrin M. In vitro anti-candida activity of the hydroalcoholic extracts of Heracleum persicum fruit against Phatogenic candida species. Jundishapur J Microbiol. 2014;7(1):1-4. https://dx.doi.org/10.5812%2Fjjm.8703
- Kim HJ, Suh H-J, Lee CH, Kim JH, Kang SC et al. Antifungal activity of glyceollins isolated from soybean elicited with Aspergillus sojae. J Agr Food Chem. 2010;58: 9483-87. https://doi.org/10.1021/jf101694t
- Kaufmann J, Schering AG. "Analysis of Variance ANOVA". In: Wiley StatsRef: Statistics Reference Online. American Cancer Society.2014. https://doi.org/10.1002/9781118445112.stat06938
- Venkatramanan M, Ganesh PS, Senthil R, Akshay J, Ravi AV et al. Inhibition of quorum sensing and biofilm formation in *Chromo-bacterium violaceum* by fruit extracts of *Passiflora edulis*. ACS Omega. 2020;5(40):25605-616. https://dx.doi.org/10.1021% 2Facsomega.0c02483
- Rugupathi V, Stephen A, Arivoli D, Kumaresan S. Pass-assisted prediction of biological activity spectra of methanolic extract of *Gymnopilus junonius*, a wild mushroom from southern Western Ghats, India. European J Pharm Med Res. 2018;5(4):340-47.
- 29. El-Shahir AA, El-Wakil DA, Abdel Latef AAH, Youssef NH. Bioactive compounds and antifungal activity of leaves and fruits methanolic extracts of *Ziziphus spina-christi* L. Plants. 2022; 11:746-64. https://doi.org/10.3390/plants11060746
- Brito SMO, Coutinho HDM, Talvani A, Coronel C, Barbosa AGR et al. Analysis of bioactivities and chemical composition of Ziziphyus joazeiro Mart. using HPLC-DAD. Food Chem. 2015;186:185-91. https://doi.org/10.1016/j.foodchem.2014.10.031
- Naghmouchi S, Alsubeie M. Biochemical profile, antioxidant capacity and allelopathic effects from five *Ziziphyus spina-christi* (L.) provenances growing wild in Saudi Arabia. Notulae Botanicae Horti Agrobotanici Cluj-Napoca. 2020; 48(3):1600-12. https://doi.org/10.15835/nbha48312025
- Tappel AL. Vitamin E as the Biological Lipid Antioxidant. Vitam Horm. 1962;20:493-510. https://doi.org/10.1016/S0083-6729(08) 60732-3
- 33. Kohno Y, Egawa Y, Itoh S, Nagaoka S, Takahashi M, Mukai K. Kinetic study of quenching reaction of singlet oxygen and scavenging reaction of free radical by squalene in N-Butanol. Biochim et Biophys Acta. 1995;1256(1):52-56. https://doi.org/10.1016/0005-2760(95)00005-w
- 34. Prabakara R, Bibin J, Pranav P. Phyto medicinal compounds from *Urginea indica* Kunth: A synthetic drugs potential alternative. Br J of Pharm Res. 2016;11(5):1-9. https://doi.org/10.9734/BJPR/2016/25216
- 35. Gautam V, Sharma A, Arora S, Bhardwaj R. Bioactive compounds in the different extracts of flowers of *Rhododendron arboreum* Sm. J Chem Pharm Res. 2016; 8(5):439-44.
- Ashraf A, Sarfraz R A, Anwar F, Shahid SA, Alkharfy KM. Chemical composition and biological activities of leaves of *Ziziphus mauri*tiana L. native to Pakistan. Pak J Bot. 2015;47 (1):367-76.
- Santos CCMP, Salvadori MS, Mota VG, Costa LM, De Almeida AAC et al. Antinociceptive and antioxidant activities of phytol *in vivo* and *in vitro* models. Neurosci J. 2013; 1-9. https://doi.org/10.1155/2013/949452
- Yoshida Y, Etsuo N. Antioxidant effects of phytosterol and its components. J Nutr Sci Vitaminol. 2003; 49(4):277-80. https://doi.org/10.3177/jnsv.49.277

- Prakash O, Manjul G, Pant AK. Essential oils composition and antioxidant activity of water extract from seeds and fruit pulp of Skimmia anquetilia N.P. Taylor & Airy Shaw. Indian J Nat Prod Resour. 2011;2(4):435-41.
- Hidajati N, Tukiran T, Setiabudi DA, Wardana AP. Antioxidant activity of palmitic acid and pinostrobin from methanol extract of Syzygium litoralle (Myrtaceae). In: Proceedings of the International Conference on Science and Technology (ICST 2018); 2018; Bali, Indonesia: Atlantis Press. https://doi.org/10.2991/icst-18.2018.39
- 41. Mohammed GT, Abdulrahman FI, Khan IZ, Hussaini MI et al. Antimicrobial efficacies of ethanolic extract and active column fractions of the stem-bark of *Zizyphus spina-christi* L. (Desf). Int J Pharm. 2013;5:455-60.
- 42. Mardani M, Badiee P, Gharibnavaz M, Jassebi A et al. Comparison of anti-Candida activities of the ancient plants *Lawsonia inermis* and *Ziziphus spina-christi* with antifungal drugs in *Candida* species isolated from oral cavity. J Conserv Dent. 2018; 21 (4):359-62. https://dx.doi.org/10.4103%2FJCD.JCD_291_17
- 43. Salimi A, Shirani M. Antifungal activity of topical microemulsion containing *Z. spina-christi* L. for the treatment of fungal vaginitis. Iran J Pharm Sci. 2021;17 (1):43-50.
- 44. Alotibi FO, Ashour EH, Al-Basher G. Evaluation of the antifungal activity of *Rumex vesicarius* L. and *Ziziphus spina-christi* (L.) Desf. aqueous extracts and assessment of the morphological changes induced to certain myco-phytopathogens. Saudi J Biol Sci. 2020; 27(10):2818-28. https://doi.org/10.1016/j.sjbs.2020.06.051
- 45. Beatriz PM, Ezequiel VV, Azucena OC, Pilar CR. Antifungal activity of *Psidium guajava* organic extracts against dermatophytic fungi. J Med Plants Res. 2021;6 (41):5435-38. http://dx.doi.org/10.5897/JMPR12.240
- El-Din HMG, Glombitza KW, Mirhom YW, Hartmann R, Michel CG. Novel Saponins from *Zizyphus spina-christi* growing in Egypt. Planta Med. 1996;62 (2):163-65. https://doi.org/10.1055/s-2006-957842
- 47. Nazif N M. Phytoconstituents of *Zizyphus spina-christi* L. fruits and their antimicrobial activity. Food Chem. 2002;76 (1):77-81. https://doi.org/10.1016/S0308-8146(01)00243-6
- 48. Kaleem WA, Muhammad N, Khan H, Rauf. Pharmacological and phytochemical studies of genus *Zizyphus*. Middle East J Sci Res. 2014: 21 (8):1243-63. http://dx.doi.org/10.5829/idosi.mejsr.2014.21.08.21099
- 49. Almeer RS, El-Khadragy MF, Abdelhabib S, Abdel Moneim AE. *Z. spina-christi* leaf extract ameliorates schistosomiasis liver granuloma, fibrosis and oxidative stress through downregulation of fibrinogenic signaling in mice. Plos One. 2018;13(10):1-23. https://doi.org/10.1371/journal.pone.0204923
- Braga PC, Culici M, Alfieri M, Dal Sasso M. Thymol inhibits *C. albicans* biofilm formation and mature biofilm. Int J Antimicrob Agents. 2008;31(5):472-77. https://doi.org/10.1016/j.ijantimicag.2007.12.013
- Dalleau S, Cateau E, Bergès T, Berjeaud JM, Imbert C. *In vitro* activity of terpenes against Candida biofilms. Int J Antimicrob Agents. 2008; 31(6):572-76. https://doi.org/10.1016/j.ijantimicag.2008.01.028
- 52. Al-Ali S, Al-Judaibi A. Biochemical and molecular effects of *Phoenix dactylifera* and *Z. spina-christi* extracts on *C. albicans*. J Biosci Med. 2019;7:29-43. https://doi.org/10.4236/jbm.2019.73004