

Development and validation of UV Spectrophotometric method for simultaneous estimation of Efonidipine hydrochloride ethanolate and Chlorthalidone in their synthetic mixture

Dipika M Solanki^a, Dhara V Patel^{*a}, Dhananjay B Meshram^a,

^aDepartment of Pharmaceutical Quality Assurance, Pioneer Pharmacy Degree College, Vadodara, Gujarat, India

*Corresponding author: patel.dhara.j@gmail.com

Efonidipine Hydrochloride Ethanolate and Chlorthalidone is used in management of hypertension and under clinical phase 3 study. The development of quality control method is required for accurate analysis of both drugs. Two simple, precise and economical UV spectrophotometric methods have been developed for the simultaneous estimation of Efonidipine Hydrochloride Ethanolate and Chlorthalidone in their synthetic mixture. Method I is simultaneous equation method (Vierodt's Method), which is based on measurement of absorption at 251 and 227 nm i.e. λ_{max} of Efonidipine Hydrochloride Ethanolate and Chlorthalidone, respectively. Method II is first order derivative was based on the measurement of absorbance of Efonidipine Hydrochloride Ethanolate measure at 283.2 nm (ZCP of Chlorthalidone) and absorbance of Chlorthalidone measure at 250.8 nm (ZCP of Efonidipine Hydrochloride Ethanolate). Linearity was observed in the concentration range of 6.4-38.4 $\mu\text{g.mL}^{-1}$ for Efonidipine hydrochloride ethanolate and 2-12 $\mu\text{g.mL}^{-1}$ for Chlorthalidone using methanol as a solvent. The accuracy of methods was assessed by recovery studies and was found to be within range of 98-102% for both the drugs. Precision of the methods was estimated by repeatability and intermediate precision studies. The % RSD values were found to be less than 2, proving methods were precise. Two methods were compared using F-test. The results were validated statistically as per ICH Q2 R1 guideline and were found to be satisfactory.

Keywords: Efonidipine Hydrochloride Ethanolate; Chlorthalidone; simultaneous equation method; first order derivative method; recovery study; validation; F-test.

Article received at 10/06/2022 and accepted at 04/07/2022.

<https://doi.org/10.22456/2527-2616.125170>

Introduction

Efonidipine Hydrochloride Ethanolate (EFD) is a 2-(N-benzyl anilino) ethyl 5-(5,5-dimethyl-2-oxo-1,3,2 λ^5 -dioxaphosphinan-2-yl)-2,6-dimethyl-4-(3-nitrophenyl)-1,4-dihydropyridine-3 carboxylate;ethanol;hydrochloride (Figure 1). It is 1,4-dihydropyridine derivative. Therapeutically categorized as Calcium channel blocker and used in treatment of hypertension and angina pectoris. It blocks L and T type of calcium channel, which leads to vasodilation and lower automaticity of heart. It exerts negative chronotropic effect by inhibiting T-type calcium channel activity of sinoatrial (SA) node, it prolongs the late phase-4 depolarization of sinoatrial action potential and decreasing heart rate (1,2).

Chlorthalidone (CTD) is a 2-chloro-5-(1-hydroxy-3-oxo-2H-isindol-1-yl) benzenesulfonamide (Figure 2). It is benzenesulfonamide derivative. Therapeutically categorized as Thiazide diuretic and used in treatment of premenstrual tension in fluid retention and in management of hypertension. It inhibits sodium ion transport across the renal tubular epithelium in the cortical diluting segments of the ascending limb of loop of Henle by increasing the delivery of sodium to the distal renal tubule, chlorthalidone indirectly increases potassium excretion via sodium-potassium mechanism (3,4).

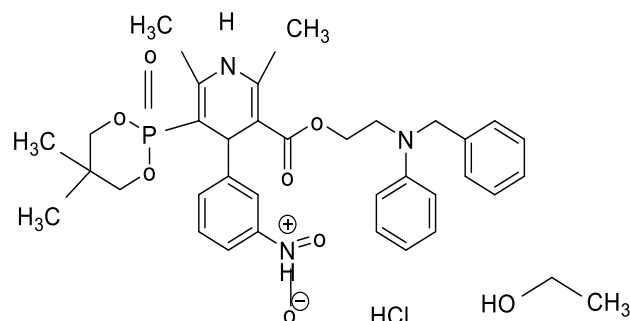


Figure 1. Chemical structure of EFD.

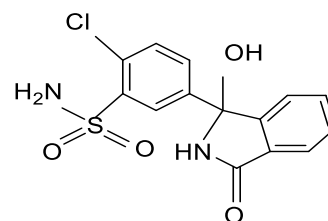


Figure 2. Chemical structure of CTD.

As consequence, this combination of EFD and CTD is an essentially use to manage hypertension and effectively regulate the blood pressure via synergistic action and CTD raise the potassium excretion by Na^+/K^+ exchange mechanism which results in low blood pressure (5,6). The considerable literature survey disclose that the analytical

methods reported for estimation of EFD and CTD alone and in combination with other drug, but no analytical method was reported for simultaneous estimation of EFD and CTD (7-40). So, it was thought of interest to develop simple, accurate, precise and reproducible UV-Spectrophotometric methods for simultaneous estimation of EFD and CTD in their synthetic mixture. Methods were validated as per ICH guideline [Q2 (R1)] (41).

Material and method

Instrumentation

All absorption spectra were recorded with UV-Visible double beam spectrophotometer (Simadzu-1800 Japan) with UV probe 2.33 software, with spectral band width of 2 nm, wavelength accuracy of 0.5 nm and a pair of 10 mm matched quartz cells. in a range of 200-400 nm. The analysis was carried by using UV Probe 2.33 software. Electronic balance (Shimadzu ATX-200) was used in the experiment.

Reagents and chemicals

Pure sample of EFD and CTD were kindly supplied as gift sample by Pure Chem Pvt. Ltd., Ankleshwar, Gujarat and CTX Life Science Pvt. Ltd., Surat, Gujarat, respectively. All reagents used were of analytical grade.

Selection of solvent

The solubility of EFD and CTD was performed using distilled water, methanol, acetonitrile, 0.1 M HCl, and 0.1 M NaOH. Both the drug was soluble in methanol so methanol was selected as a solvent.

Preparation of Standard solution

Accurately weighed quantity of 25 mg of EFD and 25 mg of CTD was transferred to separate 25 mL volumetric flask, add some methanol and sonicate for 10 min and diluted up to mark with methanol to give a stock solution having the concentration strength of 1000 $\mu\text{g.mL}^{-1}$. An aliquot of 2.5 mL from the above stock solution was pipette out in to 25 mL of volumetric flask and diluted up to mark with methanol to give a stock solution having the strength of 100 $\mu\text{g.mL}^{-1}$.

Preparation of Test Solution

Take synthetic mixture equivalent to 40 mg EFD and 12.5 mg CTD in 100 mL volumetric flask (Table 1) and add methanol up to the mark to give solution strength (400, 125 $\mu\text{g.mL}^{-1}$) sonicate for 10 min. Take 1 mL from the above solution and transfer in 10 mL volumetric flask and make the volume up to mark with methanol give solution strength (40, 12.5 $\mu\text{g.mL}^{-1}$).

Table 1: Composition of synthetic mixture.

| Item | Ingredients | Quantity (mg) | Role |
|------|-----------------------------|---------------|-------------------|
| 1 | EFD | 40 | Anti-hypertensive |
| 2 | CTD | 12.5 | Anti-hypertensive |
| 3 | Micro Crystalline Cellulose | 120 | Disintegrant |
| 4 | Starch 5% | 10 | Binder |
| 5 | PVPK-30 5% | 10 | Binder |
| 6 | Magnesium Stearate | 4 | Lubricant |
| 7 | Talc | 4 | Glidant |

Procedure for determination of wavelength for measurement

2.56 ml of stock solution of EFD (100 $\mu\text{g.mL}^{-1}$) and 0.8 mL of stock solution of CTD (100 $\mu\text{g.mL}^{-1}$) were pipette out into two separate 10 mL volumetric flasks. Volume was adjusted to the mark with methanol to get 25.6 $\mu\text{g.mL}^{-1}$ of EFD and 8 $\mu\text{g.mL}^{-1}$ of CTD. Each solution was scanned between 200-400 nm against methanol as a blank reagent. The spectrum of each solution was obtained. The wavelength maximums were found to be 251 nm and 227 nm for EFD and CTD, respectively (Figure 3).

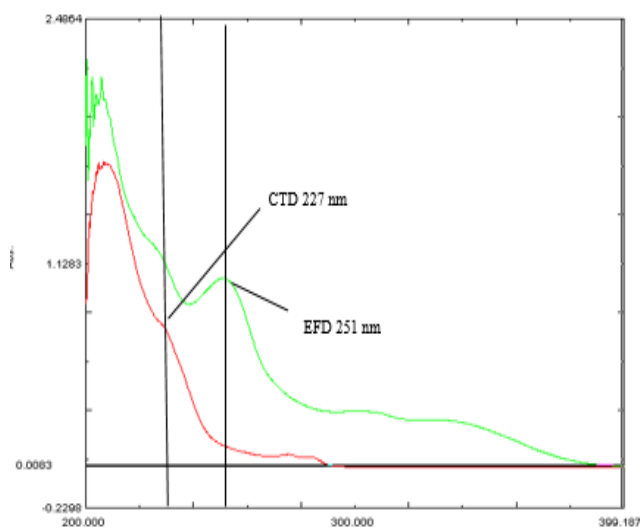


Figure 3. Overlain spectra of EFD (25.6 $\mu\text{g.mL}^{-1}$) and CTD (8 $\mu\text{g.mL}^{-1}$)

Stability of Solution

The stability of the solution in methanol was done for 48 hr at room temperature. The absorbance of the solution of EFD (25.6 $\mu\text{g.mL}^{-1}$) and CTD (8 $\mu\text{g.mL}^{-1}$) was taken at 251 nm and 227 nm, respectively, at the interval of 0, 24 and 48 hr (Table 2).

Table 2. Stability of solutions.

| Time (hr) | EFD at 251 nm | | CTD at 227 nm | |
|-----------|---------------------------|---------------|---------------------------|---------------|
| | Absorbance \pm SD (n=5) | % Degradation | Absorbance \pm SD (n=5) | % Degradation |
| 0 | 0.773 \pm 0.0007 | ----- | 0.619 \pm 0.0007 | ----- |
| 24 | 0.770 \pm 0.0008 | 0.38 | 0.617 \pm 0.001 | 0.32 |
| 48 | 0.769 \pm 0.0007 | 0.51 | 0.614 \pm 0.001 | 0.80 |

Method I: Simultaneous equation method (Vierodt's)

Simultaneous equation method is based on the absorption of drugs (X and Y) at the wavelength maximum of the other. Two wavelengths selected for the method are 251 nm and 227 nm that are λ_{max} of EFD and CTD, respectively. The absorbances were measured at the selected wavelengths and absorptivities ($A1\%_{1cm}$) for both the drugs at both wavelengths were determined as mean of six independent determinations. Concentrations in the sample were determined by using following equations:

$$C_{EFD} = (A_2 a_{y1} - A_1 a_{y2}) / (a_{x2} a_{y1} - a_{x1} a_{y2}) \quad \text{Equation 1}$$

$$C_{CTD} = (A_1 a_{x2} - A_2 a_{x1}) / (a_{x2} a_{y1} - a_{x1} a_{y2}) \quad \text{Equation 2}$$

where, A_1 and A_2 are absorbances of mixture at 251 nm and 227 nm, respectively, a_{x1} and a_{x2} are absorptivities of EFD at λ_1 (251 nm i.e. λ_{max} of EFD) and λ_2 (227 nm i.e. λ_{max} of CTD), and a_{y1} and a_{y2} are absorptivities of CTD at λ_1 and λ_2 , respectively. C_{EFD} and C_{CTD} are concentrations of EFD and CTD, respectively. Figure 3 represents the overlay spectra of both the drugs in 3.2:1 ratio and criteria for obtaining maximum precision (absorbance ratio $(A_2/A_1)/a_{x2}/a_{x1}$ and a_{y2}/a_{y1}) by this method were calculated and found to be outside the range of 0.1-2.0 which is satisfied for both the EFD and CTD.

Method II: First order derivative spectrophotometric method

For first order derivative spectrophotometric method, accurate aliquots of EFD equivalent to 6.4- 38.4 $\mu\text{g.mL}^{-1}$ were transferred from its stock solution (100 $\mu\text{g.mL}^{-1}$) into a series of 10 mL volumetric flasks and diluted to mark with methanol and mixed well. Accurate aliquots of CTD equivalent to 2-12 $\mu\text{g.mL}^{-1}$ were transferred from its working solution (100 $\mu\text{g.mL}^{-1}$) into a series of 10 mL volumetric flasks and diluted to mark with methanol and mixed well. Considering all the derivative order spectra of EFD and CTD from first to fourth derivative, the first order derivative spectra with $\delta\lambda$ 2 and scaling factor 10 found suitable. From the overlay first order derivative spectra of EFD (25.6 $\mu\text{g.mL}^{-1}$) and CTD (8 $\mu\text{g.mL}^{-1}$) the zero-crossing point (ZCP) of EFD and CTD were obtained. The wavelength selected as ZCP for EFD was 250.8 nm whereas CTD give absorbance while ZCP of CTD was 283.20 nm where EFD give absorbance. These absorbances Vs concentration were plotted in the

quantitative mode to obtain the working curves from which by extrapolating the value of absorbances of the sample solution, the concentration of the corresponding drugs was determined. Both the drugs obeyed Beer's Law.

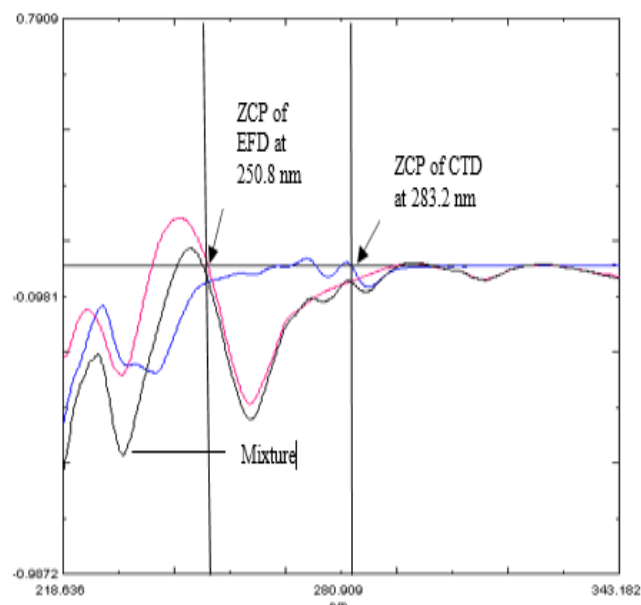


Figure 4. Overlay first order derivative spectra of EFD (25.6 $\mu\text{g.mL}^{-1}$), CTD (8 $\mu\text{g.mL}^{-1}$) and mixture (25.6 + 8 $\mu\text{g.mL}^{-1}$).

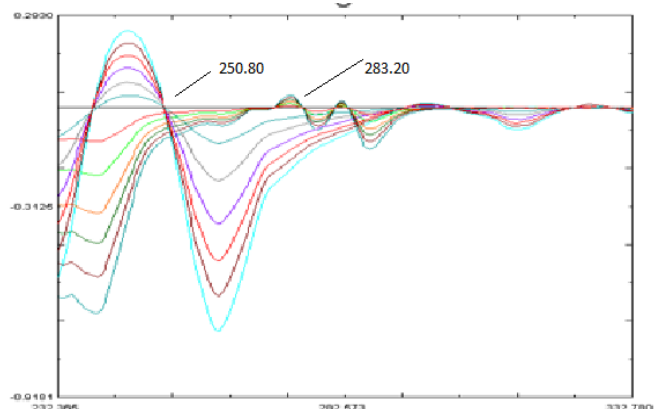


Figure 5. Overlay first order derivative spectra of EFD (6.4-38.4 $\mu\text{g.mL}^{-1}$) and CTD (2-12 $\mu\text{g.mL}^{-1}$).

Validation Parameters

Validation was carried out according to ICH guideline (ICH Q2 (R1)).

Accuracy

For studying the accuracy of the proposed methods and for checking the interference from excipients used in synthetic mixtures, experiments were carried out by the standard addition method. This study was performed by addition of known amounts of EFD and CTD to a known concentration of test solution. The amounts of standard recovered were calculated in terms of mean recovery with upper and lower limit of %RSD.

Precision

Repeatability: The precision of the instrument was checked by repeated scanning and measurement of absorbance of solutions ($n = 6$) for EFD and CTD without changing the parameter of the proposed spectrophotometry methods.

Intermediate Precision: Intra-day precision and inter-day precision for the developed methods were measured in terms of % RSD. The experiments were repeated three times a day for intra-day precision and on 3 different days for inter-day precision. The concentration values for both intra-day precision and inter-day precision were calculated three times separately and % RSD were calculated.

Limit of detection (LOD) and limit of quantitation (LOQ)

Limit of detection (LOD) and limit of quantitation (LOQ) were calculated according to the $3s/m$ and $10s/m$ criterions, respectively, where s is the standard deviation of intercept ($n = 6$) of the sample and m is the slope of the corresponding calibration curve.

Statistical Analysis

Statistical analysis was performed to assess the effect of two methods in simultaneous determination of EFD and CTD using F-test.

Results and Discussion

Method I: Simultaneous equation method (Vierodt's)

The simultaneous equation based on the absorbance of both drug EFD and CTD at their λ_{max} . Two wavelengths selected for the method are 251 nm and 227 nm that are λ_{max} of EFD and CTD, respectively. Figure 3 represent the overlain spectra of both the drugs in 3.2:1 ratio. The absorbances were measured at the selected wavelengths and specific absorptivities ($A1\%_{1cm}$) for both the drugs at both wavelengths were determined as mean of six independent determinations (Table 3). $A1\%_{1cm}$ is calculated by equation 3:

$$A = a.b.c \quad \text{Equation 3}$$

Where:

A = absorbance,

a = specific absorptivity,

b = path length 1 cm,

c = concentration of absorbing species in g.100 mL⁻¹.

Table 3. Absorptivities at 251 nm and 227 nm.

| At 251 nm | | At 227 nm | |
|-----------|-------|-----------|--------|
| a_{x1} | 398.7 | a_{x2} | 1024.6 |
| a_{y1} | 166.2 | a_{y2} | 460.6 |

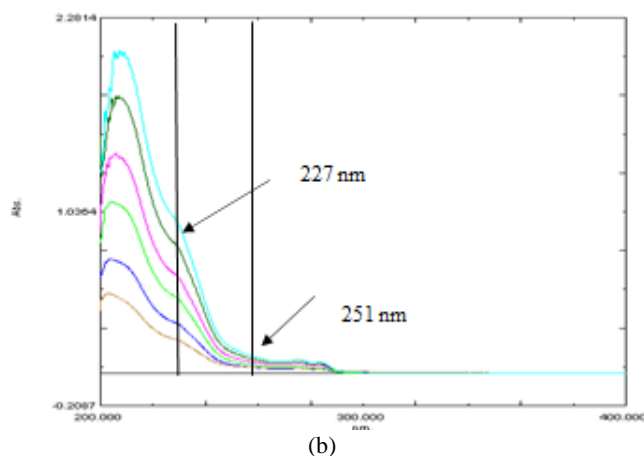
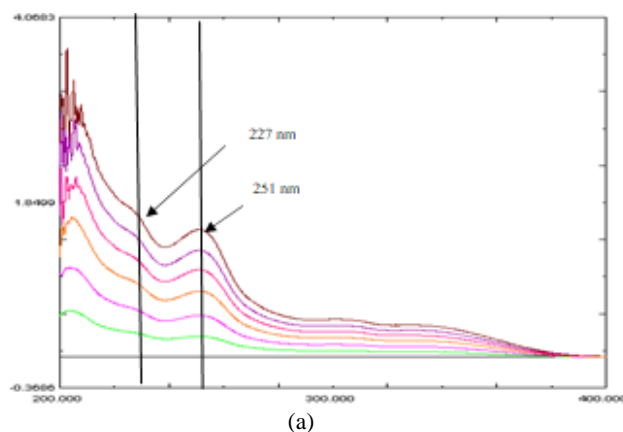


Figure 6. (a) zero-order overlain spectra of EFD (6.4 – 38.4 $\mu\text{g.mL}^{-1}$); (b) zero order overlain spectra of CTD (2 – 12 $\mu\text{g.mL}^{-1}$).

Method II: First order derivative Spectrophotometric method

In contrast to zero-order spectra, first derivative spectra show more resolution in terms of zero crossing points shown in Figure 4 and 5 which explain overlain first order derivative spectra for EFD and CTD. At 250.8 nm, EFD having zero crossing point and CTD can be determined. At 283.20 nm, CTD having zero crossing point and EFD can be determined.

Application of proposed methods for analysis of EFD and CTD in synthetic mixture

Method I: The zero-order spectrum of test solution was recorded and measured at absorbance of 251 and 227 nm to determine EFD and CTD, respectively. The concentrations of EFD and CTD in the synthetic mixture were determined using the simultaneous equation. The % assay values are given in Table 4.

Method II: The first-order spectrum of test solution was recorded and measured the absorbance at 283.20 nm and 250.80 nm for estimation of EFD and CTD. The concentrations of EFD and CTD in synthetic mixture were determined using the First order derivative method. The % assay values are given in Table 4.

Table 4, Table 5 and Table 6 exhibit results of assay, results of accuracy studies and summary of various validation parameters of all methods respectively. Statistical comparison of developed simultaneous equation method and first order derivative spectrophotometric method was done by F test (Table 7). F calculated value

was less than F critical 6.38 for both EFD and CTD, so it is indicating no significant difference observed in assay results among the two methods. Hence, it was concluded that both methods do not differ significantly.

Table 4. Assay results for tablets using the proposed methods.

| Formulation | Proposed methods | Label Claim (mg) | | Amount of drug found (mg) | | % Label Claim Assay (n=5) \pm SD | |
|-------------------|------------------|------------------|------|---------------------------|-------|------------------------------------|------------------|
| | | EFD | CTD | EFD | CTD | EFD | CTD |
| Synthetic Mixture | METHOD I | 40 | 12.5 | 39.75 | 12.45 | 99.38 \pm 0.21 | 99.60 \pm 0.35 |
| | METHOD II | 40 | 12.5 | 39.87 | 12.46 | 99.68 \pm 0.02 | 99.68 \pm 0.04 |

Table 5. Application of the standard addition technique to the analysis of EFD and CTD in synthetic mixture by the proposed methods.

| Method | Drugs | Amount present ($\mu\text{g.mL}^{-1}$) | Amount added ($\mu\text{g.mL}^{-1}$) | Total amount of drug ($\mu\text{g.mL}^{-1}$) | Amount found ($\mu\text{g.mL}^{-1}$) | %Recovery \pm SD (n = 3) | %RSD |
|-----------|-------|--|--|--|--|----------------------------|------|
| Method I | EFD | 12.8 | 6.4 | 19.2 | 19.05 | 99.22 \pm 0.39 | 0.39 |
| | | | 12.8 | 25.6 | 25.60 | 100.00 \pm 0.26 | 0.26 |
| | | | 19.2 | 32.0 | 31.98 | 99.94 \pm 0.32 | 0.32 |
| | CTD | 4 | 2 | 6 | 5.96 | 99.33 \pm 0.22 | 0.22 |
| | | | 4 | 8 | 7.96 | 99.50 \pm 0.14 | 0.14 |
| | | | 6 | 10 | 9.92 | 99.20 \pm 0.29 | 0.29 |
| Method II | EFD | 12.8 | 6.4 | 19.2 | 19.10 | 99.48 \pm 0.02 | 0.02 |
| | | | 12.8 | 25.6 | 25.57 | 99.88 \pm 0.12 | 0.12 |
| | | | 19.2 | 32.0 | 31.90 | 99.69 \pm 0.05 | 0.05 |
| | CTD | 4 | 2 | 6 | 5.99 | 99.83 \pm 0.76 | 0.76 |
| | | | 4 | 8 | 8.00 | 100.00 \pm 0.50 | 0.50 |
| | | | 6 | 10 | 9.95 | 99.50 \pm 0.19 | 0.19 |

Table 6: Summary of validation parameter by developed method.

| Parameters | Method-I | | Method-II | | | |
|---|-----------------------------|------------------|-------------------|------------------|------------------|-------------------|
| | EFD | CTD | EFD | CTD | | |
| Working wavelength (nm) | 251 nm | 227 nm | 283.2 | 250.8 | | |
| Concentration range ($\mu\text{g.mL}^{-1}$) | 6.4-38.4 | 2-12 | 6.4-38.4 | 2-12 | | |
| Slope | 0.031 | 0.078 | -0.0022 | -0.0524 | | |
| Intercept | 0.022 | 0.003 | 0.0008 | -0.0023 | | |
| Determination coefficient (r^2) | 0.999 | 0.998 | 0.999 | 0.999 | | |
| LOD ($\mu\text{g.mL}^{-1}$) | 0.148 | 0.012 | 0.15 | 0.04 | | |
| LOQ ($\mu\text{g.mL}^{-1}$) | 0.449 | 0.036 | 0.47 | 0.14 | | |
| Precision | Repeatability (n=6) %RSD | | 0.17 | 0.64 | 1.00 | 0.12 |
| | Intraday (n=3) %RSD | | 0.12-0.18 | 0.09-0.13 | 0.87-1.41 | 0.23-0.48 |
| | Interday (n=3) %RSD | | 0.10-0.27 | 0.12-0.64 | 0.87-1.41 | 0.13-0.66 |
| | 50% | | 99.22 \pm 0.39 | 99.33 \pm 0.22 | 99.48 \pm 0.02 | 99.83 \pm 0.76 |
| Accuracy (%) | 100% | | 100.00 \pm 0.26 | 99.50 \pm 0.14 | 99.88 \pm 0.12 | 100.00 \pm 0.50 |
| | 150% | | 99.94 \pm 0.32 | 99.20 \pm 0.29 | 99.69 \pm 0.05 | 99.50 \pm 0.19 |
| % Label claim Assay \pm SD (n=5) | | 99.38 \pm 0.21 | 99.67 \pm 0.35 | 99.60 \pm 0.02 | 99.68 \pm 0.04 | |

Table 7: F-test for EFD and CTD.

| EFD | Variable 1 | Variable 2 | CTD | Variable 1 | Variable 2 |
|---------------------|------------|------------|---------------------|------------|------------|
| Mean | 99.38 | 99.60 | Mean | 99.67 | 99.68 |
| Variance | 0.00027 | 0.00013 | Variance | 0.00028 | 0.00013 |
| Observations | 5 | 5 | Observations | 5 | 5 |
| df | 4 | 4 | df | 4 | 4 |
| F | 2.076923 | | F | 2.153846 | |
| P(F<=f) one-tail | 0.248219 | | P(F<=f) one-tail | 0.237852 | |
| F Critical one-tail | 6.388233 | | F Critical one-tail | 6.388233 | |

Abbreviations

EFD: Efonidipine hydrochloride ethanolate

CTD: Chlorthalidone

%RSD: % Relative standard deviation

SD: Standard deviation

Conclusions

Two spectrophotometric methods (simultaneous equation method and first order derivative method) were developed for simultaneous estimation of EFD and CTD in their synthetic mixture. Methods were found to be precise and accurate as can be reflected from validation data. Developed methods were successfully applied for estimation of EFD and CTD in synthetic mixture. The F-test results show that there is no significant difference between assay results obtained from these two methods. So, the proposed methods can be used in routine analysis of EFD and CTD with relatively less expensive and simple to operate instrumentation.

Acknowledgments

The authors express their sincere thanks to, Pure Chem Pvt. Ltd., Ankleshwar, Gujarat and CTX Life Science Pvt. Ltd., Surat, Gujarat, for supplying the gift samples of Efonidipine hydrochloride ethanolate and Chlorthalidone, respectively. Authors also extend thanks to the management, Pioneer Pharmacy Degree College, Vadodara for providing the facilities to carry out the present work.

Conflict of interest

The authors declare no conflicts of interest.

References

- Goyal RK. Elements of Pharmacology; 7th Edn; B. S. Shah Prakashan, Ahmedabad, 2007, pp 372-376.
- “Efonidipine Hydrochloride Ethanolate” accessed on May 2022, <https://pubchem.ncbi.nlm.nih.gov/compound/Efonidipine-hydrochloride-ethanolate>
- “Chlorthalidone”, accessed on May 2022, <https://go.drugbank.com/drugs/DB00310>
- Beevers G, Lip G and O' Brien E. The ABC of Hypertension: The Pathophysiology of hypertension. BMJ. 2001; 322: 912-16.
- Tamargo J and Ruilop LM. Investigational calcium channel blockers for the treatment of hypertension. Expert Opin Investig Drugs. 2016; 25: 1295-1309.
- “Thiazide Diuretic concise Medical knowledge”, accessed on May 2022, <https://www.lectorio.com/concepts/thiazide-diuretics/>
- Adeshra SD, Patel GH and Meshram DB. RP-HPLC method development and validation for simultaneous estimation of Efonidipine Ethanolate and Chlorthalidone in their synthetic mixture. Int. J. Pharma. Drug. Ana. 2021; 9: 190-195.
- Chaudhary SR and Shirkhedkar AA. Application of Plackett – Burman and Central composite designs for screening and optimization of factor influencing the chromatographic condition of HPTLC method for quantification of Efonidipine hydrochloride Ethanolate. J Anal Sci Technol. 2020; 48: 1-13.
- Pandya CP and Rajput SJ. Forced degradation study of Efonidipine HCl Ethanolate characterization of degradation products by LC- Q – TOF – MS and NMR. J Appl Pharm Sci 2020; 10:075-099.
- Rajput AS, Jha DK, Gurram S, Shah DS and Amin D. RP-HPLC method development and validation for quantification of Efonidipine Hydrochloride Ethanolate in HME processed solid dispersion. Futur J Pharm Sci. 2020, 70, 2-9.
- Kumar A, Shoni SK, Dahiya M, Kumar R, Yadav AK, Kumar V and Chaudhary H. Development and validation of liquid Chromatography (RP- HPLC) methodology for estimation of Efonidipine HCl Ethanolate. Pharm Anal Acta. 2017; 8: 1-6.
- Liu M, Deng M, Zhang D, Wang X, Ma J, Zhao H, and Zhang L et al. A Chiral LC – MS/MS method for the Spectro specific determination of Efonidipine in human plasma. J Pharm Biomed Anal. 2016; 122: 32-41.
- Otsuka M, Maeno Y, Fukami T, Inoue M, Tagami M and Ozeki T. Development considerations for Ethanolates with regard to stability and physiological characterization of Efonidipine HCl Ethanolate. CrystEngComm. 2015; 17:1-7.

14. Liu M, Deng M, Zhang D, Wang X, Ma J, Zhao H, and Zhang L et al. Determination of Efonidipine by LC- MS/MS for pharmacokinetic application. *J Pharm Biomed Anal.* 2015; 103: 1-6.
15. N Wang N, Ye L, Zhao BQ and Yu JX. Spectroscopic studies on the interaction of Efonidipine with bovin serum albumin. *Braz J Mad Bio Res.* 2008; 41: 589-595.
16. Indian Pharmacopoeia. Government of India, ministry of health and family welfare. Indian Pharmacopoeia Commission, 2014.
17. United States pharmacopoeia. USP 34 NF 29. Rockville: The United States Pharmacopoeial Convention; 2011.
18. Chaudhari SR and Shrikhedkar AA. Exploration of 1,2 – Naphthoquinone-4-Sulfonate Derivatizing Reagent for Determination of Chlorthalidone in Tablets: A Spectrophotometric Invention. *Futur J Pharm Sci.* 2021; 7:1-9.
19. Prajapati P, Patel M and Shah S. A robust high-performance thin-layer chromatography method for the simultaneous estimation of chlorthalidone and metoprolol succinate using quality risk assessment and design of experiments-based enhanced analytical quality by design approach. *JPC-J Planar Chromat.* 2021; 34: 229-242.
20. Kharat C, Shirsat VA, Kodgule YM and Kodgule M. Validated RP-HPLC Method for the Estimation of Chlorthalidone and its Process-Related impurities in an API and Tablet Formulation. *Int. J. Anal. Chem.* 2020; 2020; 1-11.
21. Prajapati P, Patel S and Mishra A. Simultaneous estimation of Azilsartan Medoxomil and Chlorthalidone by Chromatography method using design of experiment and quality risk management-based quality by design approach. *JPC-J Planar Chromat.* 2020; 33:631-646.
22. Vyas AJ, Gol DA, Usdad RG, Patel AI, Patel AB and Patel NK et al. A HPTLC – Densitometric Method for simultaneous estimation of Olmesartan Medoxomil and Chlorthalidone in Tablet dosage form. *Anal Chem Lett.* 2020; 10 :498-506.
23. Muthyala NI, Naresh B. Stability Indicating RP-HPLC Method Development and Validation for the Quantitative Estimation of Chlorthalidone in API and Tablet Dosage form. *Int. j. adv. res. med. pharm. sci.* 2019; 4:32-41.
24. Gandhi SV and Sanklecha AR. HPLC Method Development and Validation of Chlorthalidone in Tablet dosage form. *J. Drug. Deliv. Ther.* 2019; 9 :53-56.
25. Kumar GS, Ramya V, Mondal S and Kumar SP. Development and Validation of RP-HPLC methods for simultaneous estimation of Atenolol and Chlorthalidone from pharmaceutical formulation. *Int. Res. Pharma.* 2019;3: 215-219.
26. Akkala M, Satyavati D, Kumar A and Latha M. Validated RP-HPLC Method for Simultaneous Estimation of Chlorthalidone and Cilnidipine in API and Tablet Dosage form. *J. Sci. Res. Pharm.* 2018; 7 (11):124-129.
27. Rathod RH, Patil AS and Shirkhedkar AA. Novel NP and RP-HPLC in Praxis for Simultaneous Estimation of Chlorthalidone and Cilnidipine in Bulk and Pharmaceutical Formulation. *Ana. Chem. Lett.* 2018; 8: 862-871.
28. Chaudhari B and Dave JB. Development and Validation of Stability Indicating Gradient RP-HPLC Method for Simultaneous Estimation of Telmisartan and Chlorthalidone in bulk API and fixed dose combination. *World. J. Pharma. Res.* 2017;6: 1015-1029.
29. Abdullah N, Hassan MA and Hassan RO, Spectrophotometric Determination of Chlorthalidone in Pharmaceutical Formulation using Different Order Derivative Method. *Arab. J. Chem.* 2014; 10: 3426-3433.
30. Sonawane S, Jadhav S, Rahade P, Chhajed S, Kshirsagar S. Development and Validation of Stability Indicating method for estimation of Chlorthalidone in Bulk and Tablets with use of Experimental design in Forced degradation Experiments. *Scientifica (Cairo).* 2016.
31. Aneesh T, Radhakrishnan R Aravind PM, Sashidharan A, Choyal M. RP-HPLC method for simultaneous determination of Losartan and Chlorthalidone in pharmaceutical dosage form. *Int. Res. J. Pharma.* 2015; 6 :453-457.
32. Dangre P, Sawale V, Meshram S and Gunde M. Development and Validation of RP-HPLC method for the simultaneous Estimation of Eprosartan Mesylate and Chlorthalidone in Tablet Dosage form. *Int. Pharma. Tech. Res.* 2015; 8 :163-168.
33. Puttrevu S, Rachumallu R and Bhateria M, Bala V, Sharma VL, Bhatta RB. Simultaneous Determination of Azilsartan and Chlorthalidone in Rat and Human Plasma by Liquid Chromatography-Electrospray Tandem Mass Spectrometry. *J. Chromatogr. B.* 2015; 990:185-197.
34. Sadasivam R, Sravani P and Duganath N. Method Development and validation for the Simultaneous Estimation of Azilsartan and Chlorthalidone by RP-HPLC in pharmaceutical Dosage form. *Int. J. Pharma. Sci.* 2014; 4:725-729.
35. Kalaiselvi P and Lalitha KG. HPTLC method development and validation for simultaneous estimation of Chlorthalidone and Irbesartan in combined tablet dosage form. *Indian drugs.* 2014; 3:46-52.
36. Ingle SU and Patil PA. Development and Validation of UV Spectrophotometric method for chlorthalidone in bulk and pharmaceutical dosage form. *World. J. Pharm. Res.* 2014; 3: 958-963.
37. Parmar KE, Mehta RS and Patel ND. Development and validation of HPTLC method for simultaneous determination of telmisartan and chlorthalidone in bulk and pharmaceutical dosage form. *Int. J. Pharm. Sci.* 2013, 5, 420-425.

38. Rasha R and Hadir M. Validated stability-indicating methods for the simultaneous determination of amiloride hydrochloride, atenolol and chlorthalidone using HPTLC and HPLC with photodiode array detector. *J. AOAC. Int.* 2013; 96:313-23.
39. Abdelwaheb NS. Determination of atenolol, chlorthalidone and their degradation product by TLC – densitometric and chemometric methods with application of model updating. *Anal. Methods.* 2010; 2:1994-2001.
40. Salem H. HPTLC method for determination of certain antihypertensive mixtures. *Sci. Pharm.* 2004; 72 :157-174.
41. International Conference on Harmonization (ICH) of Technical Requirements for the Registration of Pharmaceutical for Human Use. Validation of analytical procedures: text and methodology Q2(R1). Geneva; 2005.