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Diphyllobothrium sprakeri n. sp. (Cestoda: Diphylobothriidae): a hidden broad tapeworm from sea lions off North and South America

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Abstract

Background: The systematic of several marine diphylobothriid tapeworms of pinnipeds has been revised in recent years. However, 20 species of *Diphyllobothrium* from phocids and otariids are still recognized as *incertae sedis*. We describe a new species of *Diphyllobothrium* from the intestine of California sea lions *Zalophus californianus* (Lesson) (type-host) and South American sea lions *Otaria flavescens* (Shaw).

Methods: *Zalophus californianus* from the Pacific coast of the USA and *O. flavescens* from Peru and Argentina were screened for parasites. Partial fragments of the large ribosomal subunit gene (*18S*rDNA) and the cytochrome *c* oxidase subunit 1 (*cox1*) mitochondrial gene were amplified for 22 isolates. Properly fixed material from California sea lions was examined using light and scanning electron microscopy.

Results: A total of four *18S*rDNA and 21 *cox1* sequences were generated and aligned with published sequences of other diphylobothriid taxa. Based on *cox1* sequences, four diphylobothriid tapeworms from *O. flavescens* in Peru were found to be conspecific with *Adenocephalus pacificus* Nybelin, 1931. The other newly generated sequences fall into a well-supported clade with sequences of a putative new species previously identified as *Diphyllobothrium* sp. 1. from *Z. californianus* and *O. flavescens*. A new species, *Diphyllobothrium sprakeri* n. sp., is proposed for tapeworms of this clade.

Conclusions: *Diphyllobothrium sprakeri* n. sp. is the first diphylobothriid species described from *Z. californianus* from the Pacific coast of North America, but *O. flavescens* from Argentina, Chile and Peru was confirmed as an additional host. The present study molecularly confirmed the first coinfection of two diphylobothriid species in sea lions from the Southern Hemisphere.

Keywords: Parasites, Helminths, *cox1*, *18S*rDNA, Pinnipedia, Otariidae, *Zalophus californianus*, *Otaria flavescens*

Background

The systematic of the tapeworms (Cestoda) has been under detailed revision since the last decade including adding new molecular data [1]. However, current knowledge and systematics of the tapeworms of marine mammals are still superficial [2–7]. The pinnipeds are well-known hosts of several groups of tapeworms

composed of two orders, Tetrabothriidea with only seven members of the genus *Anophryocephalus* Baylis, 1922, and Diphylobothriidea with around 30 species of several genera [7, 8]. Waeschenbach et al. [4] revised the systematic of diphylobothriids based on morphological as well as molecular data and proposed systematic changes mainly in the polyphyletic genus *Diphyllobothrium* Cobbold, 1858, which now includes only seven species parasitizing cetaceans. Non-monophyletic *Diphyllobothrium* (designated as '*Diphyllobothrium*' by Waeschenbach et al. [4]) provisionally comprises 20 species mainly from pinnipeds (eight of which have been characterized

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molecularly) that do not form a monophyletic assemblage and are considered *incertae sedis* [4].

The California sea lion *Zalophus californianus* (Lesson) (CSL) is a well-known otariid species of North America whose range in the Pacific is from Baja California, Mexico, to British Columbia, Canada. Nowadays, its population is increasing and includes around 340,000 individuals [9, 10]. The first diphyllbothriid tapeworm from CSL was reported by Stunkard [11] who studied a scolex of '*Diphyllbothrium*' sp. (see fig. 2 in Stunkard [11]). Later, several authors reported diphyllbothriids from CSL as *Adenocephalus pacificus* Nybelin, 1931 (under the names *D. glaciale* [Cholodkovsky, 1915] or *D. pacificum* [Nybelin, 1931]) [12–17]. The recent revision of metazoan parasites of CSL considered 24 valid species, including two undescribed tapeworm species [18].

The South American sea lion *Otaria flavescens* (Shaw) (SASL) is distributed exclusively in South America from the Pacific coast of Peru to the Atlantic coast of southern Brazil [19]. Nowadays, its population has been estimated as at least 445,000 individuals [20]. The first diphyllbothriid tapeworm from SASL was reported by Baylis [21] from Falkland Island (Islas Malvinas) as *Diphyllbothrium scoticum* (Rennie & Reid, 1912). This material was later studied by Markowski [22] who confirmed Baylis' specimens as *D. scoticum*. However, the evaluation Baylis' [21] material by Baer et al. [23] and Hernández-Orts et al. [2] suggested that these specimens belong to *A. pacificus*. Hermosilla et al. [24] reported a '*Diphyllbothrium scoticum*-like cestode' from SASLs from the Pacific coast of Chile to be molecularly conspecific with samples from CSLs. The recent revision of metazoan parasites of SASL by Ebmer et al. [25] reported 44 metazoan taxa (only 25 were identified to the species level), including *A. pacificus* with its synonyms, *Clistobothrium delphini* (Bosc, 1802) [as *Phyllobothrium delphini* (Bosc, 1802)] and unidentified species of *Anophryocephalus* Baylis, 1922 and '*Diphyllbothrium*'.

A detailed study of newly obtained material, examination of museum specimens, as well as molecular phylogenetic analysis and extensive literature revision confirmed the presence of an undescribed species of the genus *Diphyllbothrium* which parasitizes CSLs. The main goal of this article is the description of a new species previously reported as '*Diphyllbothrium*' sp. 1 by Waeschenbach et al. [4], Hermosilla et al. [24] and Kuzmina et al. [18] on the basis of detailed morphological and molecular data.

Methods

Sample collection

A total of 39 CSLs, 9–10 months to 16 years old, were collected stranded on the Pacific coast of central California (36°57'–38°32'N, 121°95'–123°00'W), USA, between 2012 and 2018 (see [18] for details). CSLs died in the Marine Mammal Center (Sausalito, California, USA) from different causes and were necropsied using a standard procedure [26]. Two recently dead SASLs were collected from two localities in South America: (i) a subadult male from Bellavista beach (12°04'S, 77°07'W), Callao City, Callao, Peru, in October 2017; (ii) a subadult female from Playa Unión (43°19'S, 65°03'W), Chubut, Argentina, in October 2013. The intestines of fresh SASLs were excised from the carcasses, opened and washed with tap water through a series of sieves. Intestinal contents were placed in Petri dishes with saline and examined under a dissecting microscope. Tapeworms were washed in saline and killed with hot (90 °C) tap water and fixed in 70% ethanol. A few posterior proglottids of selected specimens were cut off and fixed in molecular-grade ethanol (99%) for DNA sequencing before killing the worm.

Molecular data and phylogenetic analyses

Three diphyllbothriid specimens from CSLs and 18 from SASLs (17 from Peru and one from Argentina) were selected for molecular studies. Pieces of strobila were used for DNA isolation and sequencing. The remaining parts of the worms were stained and mounted in Canada balsam and were kept as molecular vouchers (i.e. hologenophores sensu Pleijel et al. [27]). Total genomic DNA was extracted using DNeasy Blood and Tissue Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. Phylogenetic relationships of the studied diphyllbothriids were evaluated based on two molecular markers: the large subunit nuclear ribosomal RNA gene (*lsrDNA*) and the cytochrome *c* oxidase subunit 1 (*cox1*) mitochondrial gene.

Partial *lsrDNA* (D1–D3 domains, ca. 1400 bp) sequences were generated using the primers LSU5 (5'-TAG GTC GAC CCG CTG AAY TTA AGC A-3'; [28]) and 1500R (5'-GCT ATC CTG AGG GAA ACT TCG-3'; [29]). Partial (ca. 420 bp) and almost complete (ca. 1500 bp) *cox1* sequences were amplified using the primers JB3 (5'-TTT TTT GGG CAT CCT GAC GTT TAT-3'; [30]) and JB 4,5 (5'-TAA AGA AAG AAC ATA ATG AAA ATG-3' [30]) or the primers Cox1Forward (5'-TAT CAA ATT AAG TTA AGT AGA CTA-3';

[31]) and Cox1Reverse (5'-CCA AAT AGC ATG ATG CAA AAG-3'; [31]), respectively. PCR amplification reactions were performed following the procedures described by Brabec et al. [32] for the *lsrDNA* gene and Wicht et al. [31] or Gomez-Puerta et al. [33] for the *cox1* gene. All products were purified through an enzymatic treatment with Exonuclease I and FastAP alkaline phosphatase (Thermo Fisher Scientific, Waltham, MA, USA) or using a Microcon® Centrifugal Filters (Millipore, Bedford, MA, USA). Purified products were Sanger sequenced at GATC Biotech (Konstanz, Germany) or using BigDye™ Terminator Cycle Sequencing Ready Reaction Kit (Applied Biosystems, Foster City, CA, USA) and an ABI 3100 automated sequencer (Applied Biosystems).

Sequences were assembled and inspected for errors using Geneious v.11 and deposited in the GenBank database (accession numbers MW600336–MW600339 for the *lsrDNA* sequences and MW596661–MW596682 for the *cox1* sequences). Newly generated sequences were aligned in two independent datasets following the alignments from Hernández-Orts et al. [2] and Waeschenbach et al. [4]. Sequences from other diphylobothriids were retrieved from GenBank and aligned with our novel sequences using default settings of MUSCLE [34] implemented in Geneious (Additional file 1: Table S1). The extremes were trimmed resulting in an alignment with 1574 bp for the *lsrDNA* (Additional file 2: *lsrDNA_alignment*) and 1571 bp for the *cox1* (Additional file 3: *cox1_alignment*). A combined *lsrDNA* + *cox1* alignment (3145 bp; Additional file 4: *lsrDNA_cox1_alignment*) was also constructed using only taxa with sequences for both markers available from GenBank (Additional file 1: Table S1).

Bayesian inference (BI) and maximum likelihood (ML) analyses were performed for each dataset. jModelTest 2.1.10 software [35] was used to select the best nucleotide substitution model under the Akaike information criterion. The TIM2+I+G model was chosen for the *lsrDNA* and *cox1* datasets and the TIM2+I+G & TIM1+I+G for the combined dataset. Bayesian inference analyses were constructed using MrBayes 3.2.6 [36]. The BI analyses were estimated via two independent Markov Chain Monte Carlo runs of four chains with standard settings for 10,000,000 generations with a sampling frequency of 1000th generations. Burn-in periods were set to 25% of generations. The ML analyses were run with raxmlGUI v.2.0 [37]. Bootstrap nodal support values were computed by running 1000 bootstrap resamples. The resulting trees for BL and ML were visualized in FigTree 1.4.4 [38]. Genetic distances (uncorrected *p*-distance) were calculated with MEGA 10.1.8 [39] from the total number of nucleotide

differences from the *lsrDNA* alignment and from the full *cox1* alignment excluding partially characterized sequences (i.e. < 1400 bp).

Morphological examination

For morphological evaluation, selected tapeworms were stained with Mayer's hydrochloric carmine, dehydrated through an ethanol series, cleared with eugenol and mounted in permanent slides in Canada balsam. Selected pieces of the strobila were embedded in paraffin wax, cross-sectioned (thickness 15 µm), stained with hematoxylin–eosin, and mounted in Canada balsam. Mounted specimens were examined with an Olympus BX51 microscope (Olympus Corp., Tokyo, Japan). Measurements were taken from digital images with the QuickPHOTO CAMERA 3.2 image analysis software (Promicra Ltd., Prague, Czech Republic). Measurements are expressed in micrometers unless otherwise stated and presented as the range (minimum–maximum), with the mean followed by the standard deviation (SD) and the number of measured specimens or structures in parentheses. Detailed line drawings were made using a drawing tube attached to an Olympus BX51 microscope.

Selected scoleces and proglottids were prepared for scanning electron microscopy (SEM). Specimens were dehydrated through an ethanol series, transferred to hexamethyldisilazane (Ted Pella, Inc., Redding, CA, USA) and allowed to air dry. Samples were mounted on aluminium stubs on double-sided adhesive carbon tape, gold sputter-coated and examined with a JEOL JSM 7401-F scanning electron microscope (JEOL Ltd., Tokyo, Japan) at 4 kV at the Laboratory of Electron Microscopy, Institute of Parasitology, Biology Centre, Czech Academy of Sciences. Terminology of microtriches follows Chervy [40].

Tapeworms collected in the present study were compared with voucher material of the following immature diphylobothriid species collected from *Z. californianus* identified as '*Diphylobothrium latum* (L.)' and deposited at the Natural History Museum (NHML), London, UK: two vouchers from San Diego County (NHML 1980.6.3.196–8), one from Los Angeles (NHML 1980.6.3.200), one from an unknown locality in California, USA (NHML 1994.7.21.12–13), and one from Mexican waters (NHML 1980.6.3.189).

Specimens of the type series and voucher specimens from the present study are deposited in the Helminthological Collection of the Institute of Parasitology (IPCAS), Biology Centre, Czech Academy of Sciences, České Budějovice, Czech Republic; the National Museum of Natural History of the Smithsonian Institution (NMNH-USNM), Washington, DC, USA; the Parasite Collection of the Museum of Natural History (PCMNH),

National University of San Marcos, Lima, Peru; and the Collection of the Laboratory of Epidemiology and Veterinary Economics (LEVE), School of Veterinary Medicine, National University of San Marcos, Lima, Peru.

Results

More than 150 tapeworm specimens (including immature and gravid specimens) were collected in the intestine of CSLs. In the SASLs from Argentina and Peru, 13 and 18 immature tapeworms were collected, respectively.

Phylogenetic relationships and genetic divergence

A total of four partial *lsrDNA* sequences (1450–1481 bp long; three isolates from CSLs, USA, and an isolate from SASL, Argentina) were generated. Additionally, one almost complete (1565 bp long; isolate from SASL, Argentina) and 21 partial *cox1* sequences (415–420 bp long; four isolates from CSLs, USA, and 17 isolates from SASL, Peru) were generated (see Additional file 1: Table S1).

The resulting phylograms inferred with BI and ML analyses for the *lsrDNA* dataset (34 taxa) showed generally similar topologies (Fig. 1) and congruent results with those of Waeschenbach et al. [4]. Our four newly generated sequences formed a well-supported clade with sequences of a putative new species '*Diphyllobothrium*' sp. 1 from CSL of California (KY552829) [4] and '*Diphyllobothrium*' sp. 1 from SASL of Chile (KY945917) [19] (Fig. 1). The clade composed by our novel sequences and '*Diphyllobothrium*' sp. 1 appeared to be sister to a sequence generated from a diphyllbothriid plerocercoid (reported as Diphyllbothriidae gen. sp.) from *Trematopus bernacchii* Boulenger of Antarctica (KY552830) [4] (Fig. 1). '*Diphyllobothrium*' sp. 1 and Diphyllbothriidae gen. sp. form the sister group of a clade which includes '*Diphyllobothrium*' *scoticum* from *Mirounga leonina* (L.) and *A. pacificus* isolates, but these two lineages with low support (Fig. 1).

Intraspecific sequence variability of 0.14% was only detected between the four isolates from CSLs and the two from SASLs for the *lsrDNA* gene. The *lsrDNA* sequences of isolates from CSLs and SASL diverged 0.64–0.78% from the sequences of Diphyllbothriidae gen. sp. from *T. bernacchii* from Antarctica. Genetic divergence values between our lineage and '*Diphyllobothrium*' *scoticum* were 2.48–2.55% and between *A. pacificus* ranged from 1.99–2.06%. Intergeneric distance between '*Diphyllobothrium*' *scoticum* and *A. pacificus* was 1.13%. The interspecific distances in the *lsrDNA* region of '*Diphyllobothrium*' spp. of pinnipeds range between 0.36% ('*Diphyllobothrium*' *cordatum* (Leuckart, 1863) vs '*Diphyllobothrium*' *lanceolatum* (Krabbe, 1865)) to 3.70%

('*Diphyllobothrium*' *tetrapterum* (von Siebold, 1848) vs '*Diphyllobothrium*' cf. *cameroni* Rausch, 1969).

Both ML and BI analyses of the *cox1* dataset (59 taxa) resulted in generally similar topologies (Fig. 2), but slightly differed from the recent phylogenetic study of Waeschenbach et al. [4]. Four newly generated sequences from isolates from CSLs and 14 novel sequences from SASLs (13 from Peru and 1 from Argentina) formed a strongly supported clade with three sequences identified as '*Diphyllobothrium*' sp. 1, including one sequence from an isolate from CSL (KY552890) and two from SASLs (MF893274 and KY945922) (Fig. 2). The genetic divergence between isolates from CSLs and SASLs in the *cox1* gene was 2.87–3.07%. In the phylogenetic tree, the clade formed by our novel sequences + '*Diphyllobothrium*' sp. 1. appeared also as sister to the sequence of Diphyllbothriidae gen. sp. from *T. bernacchii* (KY552888). Genetic divergence between our newly generated sequences + '*Diphyllobothrium*' sp. 1 and the isolate reported as Diphyllbothriidae gen. sp. ranged from 14.56–14.94%.

In contrast to the phylograms inferred for the *lsrDNA* dataset, our novel sequences + '*Diphyllobothrium*' sp. 1 + Diphyllbothriidae gen. sp. formed the sister group, although with low support, to an unresolved clade of diphyllbothriids (Fig. 2). The interspecific distances in the *cox1* region of '*Diphyllobothrium*' spp. of pinnipeds ranged between 1.88% ('*Diphyllobothrium*' *tetrapterum* vs '*Diphyllobothrium*' *schistochilus*) to 14.30% ('*Diphyllobothrium*' *tetrapterum* vs '*Diphyllobothrium*' cf. *hians* (Diesing, 1850)). Finally, four newly generated partial sequences from isolates from SASL from Peru formed a strongly supported monophyletic lineage with the previously published sequences of *A. pacificus* (Fig. 2).

The phylogenetic trees inferred with the combined (*lsrDNA* + *cox1*) dataset (33 taxa) were similar to the topologies inferred for the *lsrDNA* and *cox1* datasets (Additional file 5: Fig. S1). Our newly generated sequences formed a well-supported clade with sequences of '*Diphyllobothrium*' sp. 1 from CSL of California [4] and '*Diphyllobothrium*' sp. 1 from SASL of Chile [19].

In summary, both ML and BI analyses for the *lsrDNA*, *cox1* and the combined datasets revealed that most of our newly generated sequences from isolates from CSLs and SASLs belong to an as yet undescribed species reported as '*Diphyllobothrium*' sp. 1 [4]. Genetic variation was detected between sequences from isolates of the putative new species from CSLs and SASLs. However, these values were somewhat lower, especially for *cox1*, than the interspecific variation between other '*Diphyllobothrium*' species of pinnipeds included in our phylogenetic analyses. Therefore, we consider isolates of the putative new species of '*Diphyllobothrium*' from CSLs and SASLs from

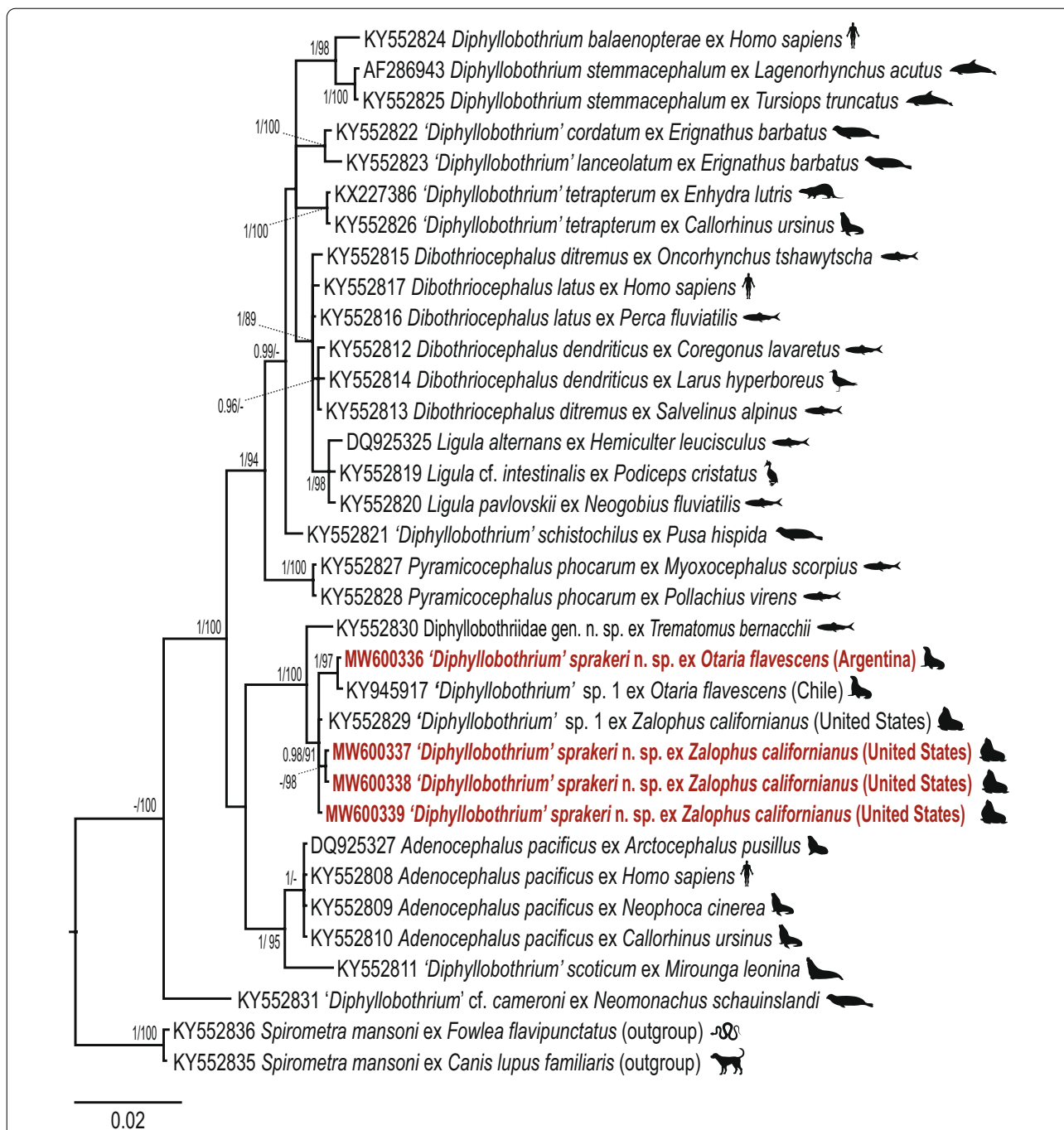


Fig. 1 Bayesian analysis for the partial *IsrDNA* dataset for selected representatives of the family Diphylobothriidae. Numbers represent posterior probabilities from BI analysis (>0.95 shown only) followed by nodal supports from ML analysis (bootstrap values > 70% shown only). The newly generated sequences are indicated in red. GenBank accession numbers are shown before the species names. The scale bar indicates the expected number of substitutions per site

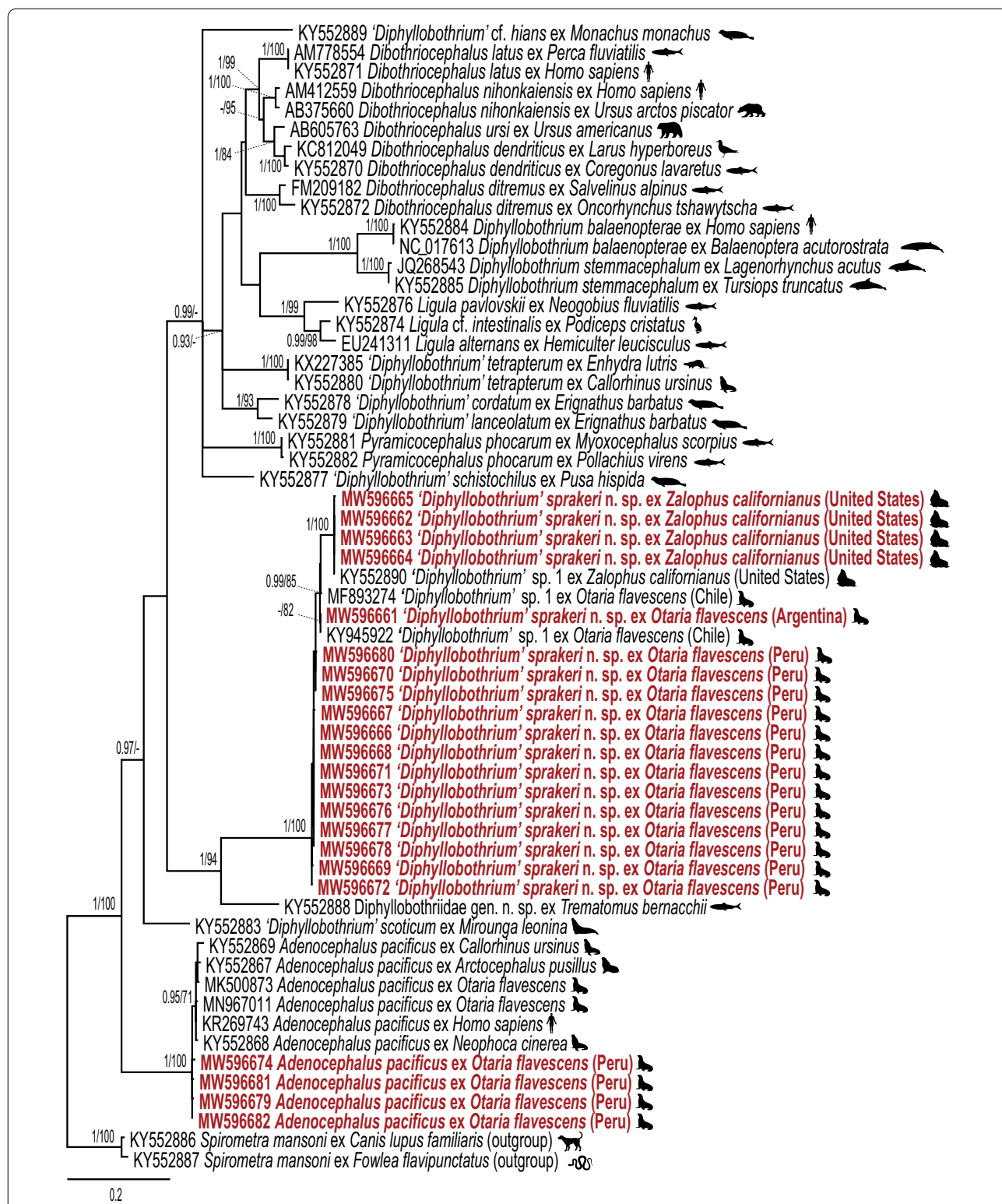


Fig. 2 Bayesian analysis of the *cox1* dataset for representatives of the family Diphylobothriidae. Numbers represent posterior probabilities from BI analysis (> 0.95 shown only) followed by nodal supports from ML analysis (bootstrap values > 70% shown only). The newly generated sequences are indicated in red. GenBank accession numbers are shown before the species names. The scale bar indicates the expected number of substitutions per site

North and South America, respectively, to be conspecific. This new tapeworm species is described below.

Family Diphylobothriidae Lühe, 1910

Genus *Diphylobothrium* Cobbold, 1858

Diphylobothrium sprakeri Hernández-Orts, Kuzmina, Gomez-Puerta & Kuchta n. sp.

Synonyms: *Diphylobothrium* sp. 1 of Waeschenbach et al. [4], Hermosilla et al. [24] and Kuzmina et al. [18]

Type-host: *Zalophus californianus* (Lesson) (Carnivora: Otariidae), California sea lion.

Type-locality: Off central California (36°57′–38°32′N, 121°95′–123°00′W), USA.

Other host: *Otaria flavescens* Shaw (Carnivora: Otariidae), South American sea lion.

Other localities: Los Angeles and San Diego, California, USA; Mexican waters (see below); Playa Unión (43°19′S, 65°03′W), Chubut, Argentina; beach of Bellavista (12°04′S, 77°07′W), Callao City, Callao, Peru.

Site in host: Small intestine.

Prevalence in type-host: 38% (15 out of 39 examined sea lions).

Intensity in type-host: 1–30 (average = 9.5) tapeworms per sea lion; most of the tapeworm specimens collected (> 70%) were in immature stage.

Intensity in South American sea lions: 1–17 immature tapeworms per host.

Representative DNA sequences: GenBank accession numbers: MW600337–MW600339 (*lsrDNA*) and MW596662–MW596665 (*cox1*) from *Z. californianus*: MW600336 (*lsrDNA*) and MW596661, MW596666–MW596673, MW596675–MW596678, MW596680 (*cox1*) from *O. flavescens*.

Deposition of specimens: Holotype (IPCAS C-765/1), seven slides of whole mounts and five slides of histological sections; one paratype (NMNH-USNM 1642475), 14 slides of whole mounts; one voucher (NMNH-USNM 1642476), eight slides of whole mounts; 10 vouchers (LEVE 986–990, 992–995, 997), two vouchers (PCMNH 4711, 4712) and one voucher (IPCAS C-765/2) immature worms from *O. flavescens* of Peru; three vouchers (IPCAS C-765/3), immature worms from *O. flavescens* of Argentina.

ZooBank registration: The Life Science Identifier (LSID) of the article is urn:lsid:zoobank.org:pub:68C9D942-6D3A-4CF9-8732-4766EBA494DB. The LSID for the new name *Diphylobothrium sprakeri* n. sp. is urn:lsid:zoobank.org:act: B95F5960-A589-4B34-BBE2-21E1D2EF368A.

Etymology: The species is named after Prof. Terry R. Spraker from the Colorado State University, Colorado,

USA, for his valuable contribution to the collection of this species and studies of various groups of parasites of marine mammals.

Description

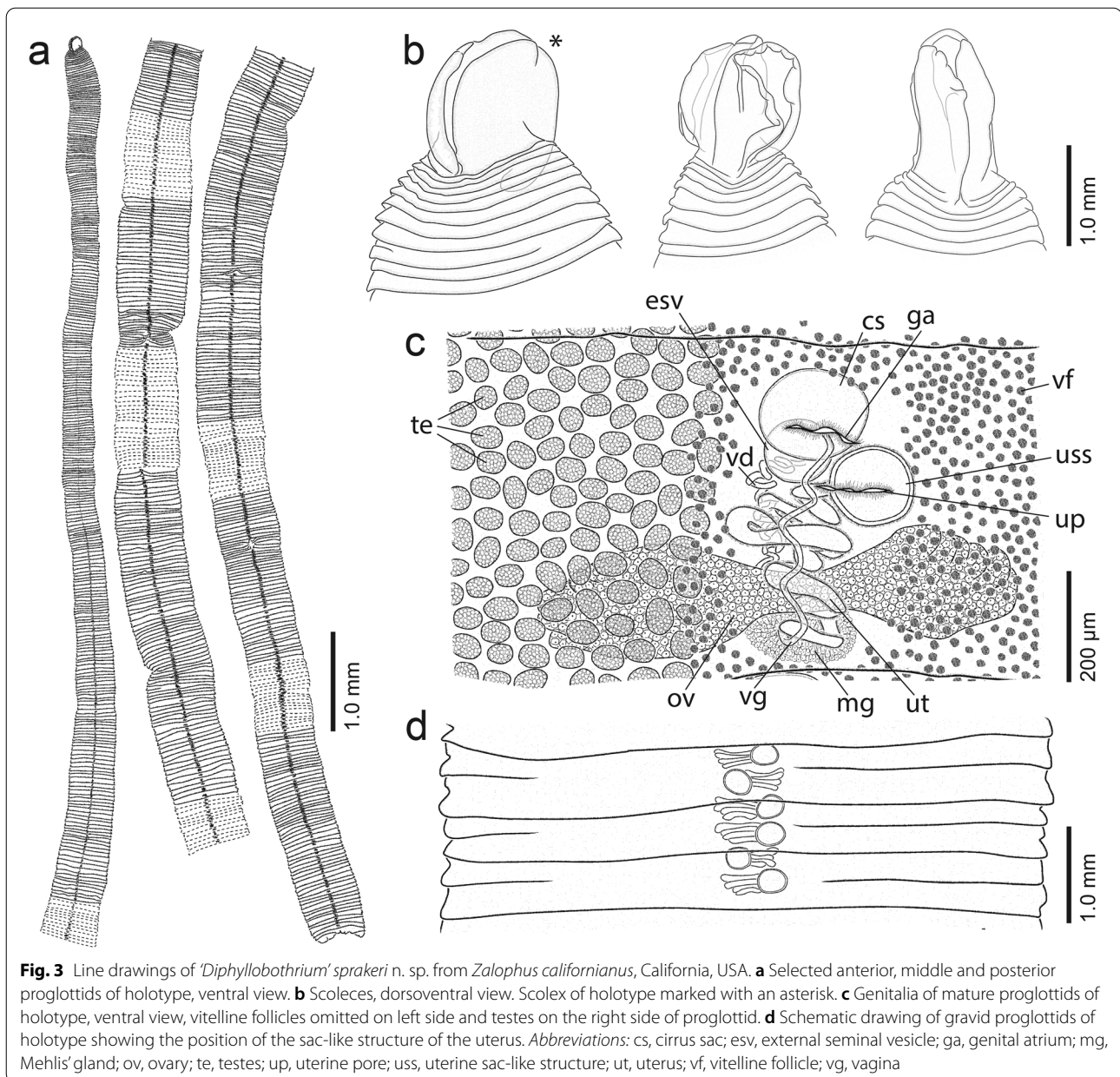
[Based on 20 specimens from CSLs. All tapeworms collected from SASL were immature specimens and were not included in the description]. Diphylobothriidea, Diphylobothriidae. Specimens anapolytic, long worms up to 2 m (holotype 56 cm; Fig. 3a) in length; maximum width ca. 8 mm. Immature worms may reach 1 m in length. Scolex surface covered with capilliform filitriches and coniform spinitriches (Fig. 4a, b). Microtriches on strobila surface not observed.

Excretory system consists of cortical and medullary longitudinal canals extending throughout strobila; canals in cortex numerous, interspersed in layer of vitelline follicles or displaced towards tegument; 2 pairs of main excretory canals in medulla alongside median axis. Longitudinal musculature formed by muscle bundles, in layer surrounding transverse musculature (Fig. 5a,c,d); longitudinal muscle layer 51–100 (71.2 ± 18 , $n = 10$) wide.

Scolex lanceolate, roundish or slightly elongated to triangular in dorsoventral view (Figs. 3b, 4c–e); 999–1484 (1207 ± 120 , $n = 15$) long by 807–1275 (935 ± 132 , $n = 15$) wide. Bothria with wide margins in lateral view, fan-like (Figs. 3b, 4c–e); bothrial margins fused anteriorly, leaving opening of varying degrees in middle and posterior parts (Figs. 3b, 4c–f). Neck absent.

Proglottids craspedote, much wider than long, first eggs appear around 10 cm posterior to scolex; first proglottids posterior to scolex short, much wider than long, 51–188 (105.9 ± 37 , $n = 30$) long by 821–1553 (1141 ± 214 , $n = 30$) wide; proglottid width/length ratio 1:0.04–0.22 (0.1, $n = 30$). Mature proglottids, i.e. with spermatozoa in vas deferens, few, 1–3 in number (Fig. 3c), 281–401 (340 ± 42 , $n = 7$) long by 3115–6202 (4451 ± 1177 , $n = 7$) wide; proglottid width/length ratio 1:0.05–0.12 (0.08, $n = 7$). Gravid proglottids numerous, larger and wider, not enlarging, 287–736 (519 ± 130 , $n = 7$) long by 2681–7890 (4712 ± 1921 , $n = 7$) wide; proglottid width/length ratio 1:0.05–0.22 (0.14, $n = 7$).

Testes medullary, subspherical to oval, 44–70 (56 ± 6 , $n = 16$) in diameter, 220–384 (325 ± 61 , $n = 7$) in number, arranged in a single dorsoventral layer (Figs. 3c, 5a), form 2 lateral fields confluent at anterior and posterior margins of proglottid, not overlapping anterior uterine loops, absent in central area of cirrus sac, uterus and ovary (Fig. 3c). Vas deferens coiled, runs dorsally to uterus in midline to posterior border of external seminal vesicle (Fig. 3c). External seminal vesicle muscular, wall up to 27 thick, posterodorsal to cirrus sac, oval to almost elliptical in sagittal section (Figs. 3c, 5b), 58–122 (103 ± 19 ,



$n=9$) long by 38–120 (82 ± 21 , $n=9$) wide in dorsoventral view; length/width ratio 1:0.7–1 (0.8, $n=9$). Cirrus sac thin-walled, wall up to 23 thick, larger than external seminal vesicle, oval in sagittal section (Fig. 5b), 142–286 (206 ± 40 , $n=9$) long by 92–159 (127 ± 18 , $n=9$) wide; length/width ratio 1:0.5–0.7 (0.6, $n=9$). Cirrus sac opening in anterior region of genital atrium (Fig. 5b). Internal sperm duct thin-walled, strongly coiled; cirrus unarmed. Genital atrium ventral, median, pre-equatorial, situated 79–242 (201 ± 32 , $n=7$) from anterior margin of proglottid, representing 29–60% (40%) of proglottid length;

surface area surrounding genital atrium covered with numerous papillae (Fig. 4g).

Ovary bilobed, situated dorsally near posteriormost margin of proglottid, 1029–1550 (1302 ± 167 , $n=10$) wide; ovarian lobes 135–233 (181 ± 30 , $n=10$) long, posteriorly enclosing Mehlis' gland (Fig. 3c). Vagina runs ventrally, 19–22 wide in cross sections (Fig. 5b). Female genital pore posterior to male genital opening in genital atrium (Fig. 5b). Vitelline follicles cortical, subspherical, numerous, 18–38 (29 ± 5 , $n=18$) in maximum diameter in dorsoventral view (Fig. 3c); area surrounding genital

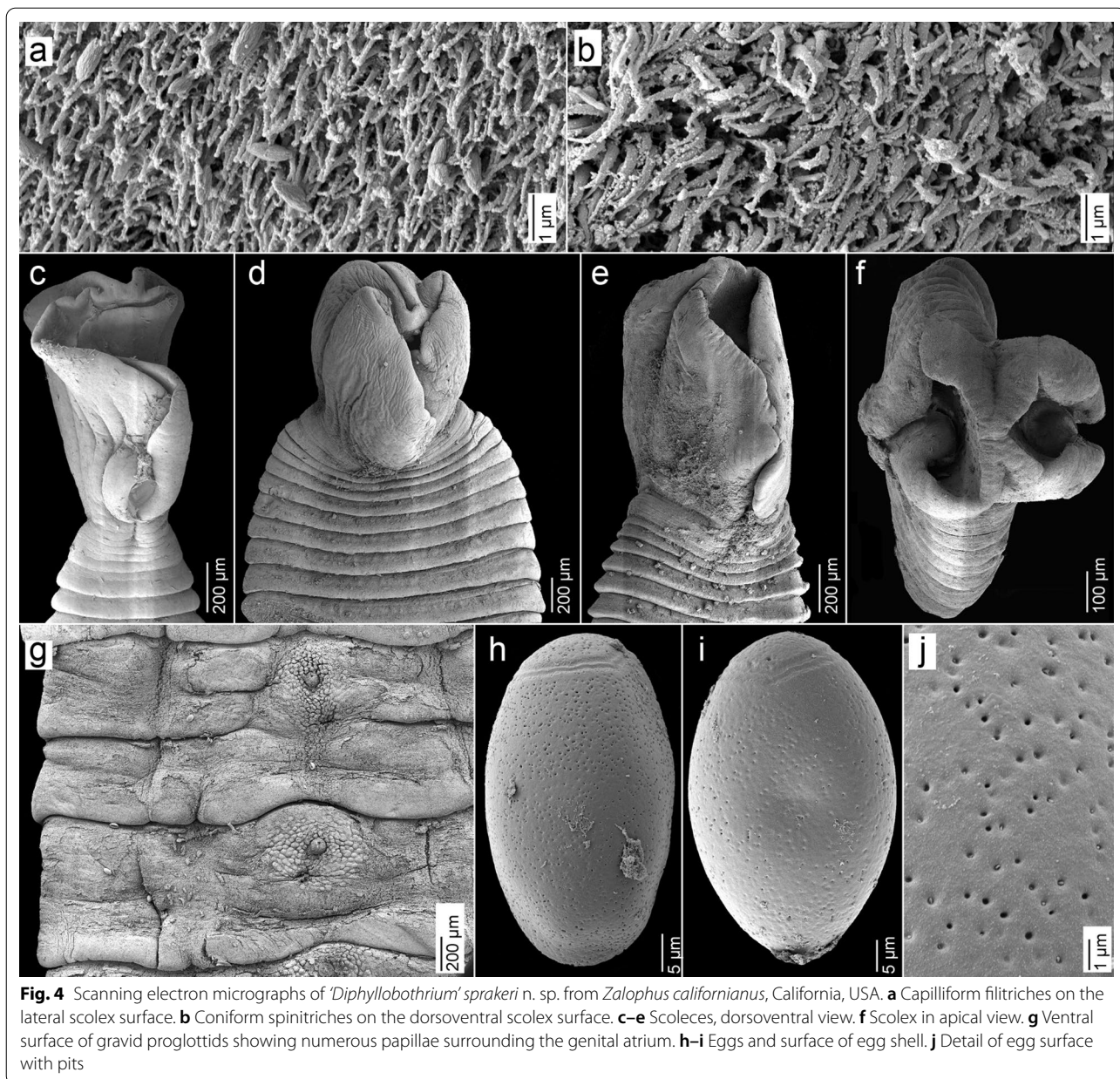


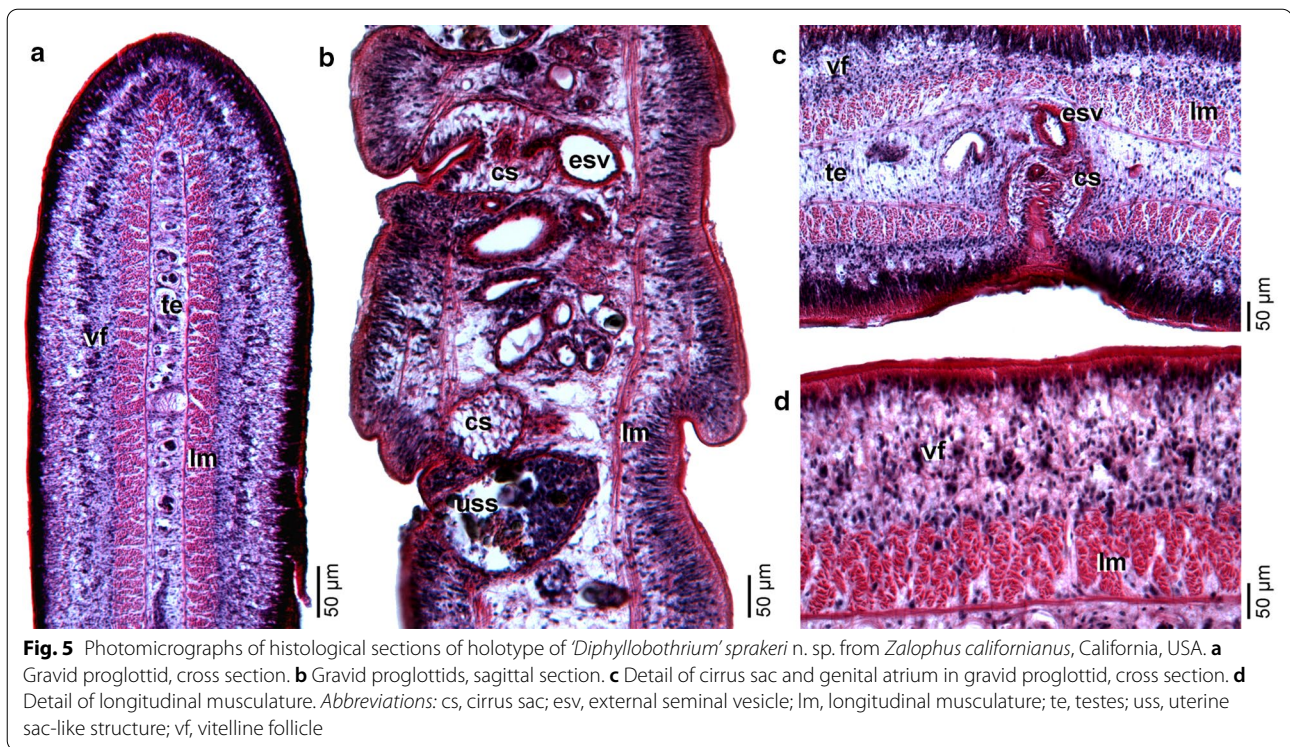
Fig. 4 Scanning electron micrographs of *Diphyllobothrium sprakeri* n. sp. from *Zalophus californianus*, California, USA. **a** Capilliform filitriches on the lateral scolex surface. **b** Coniform spinitriches on the dorsoventral scolex surface. **c–e** Scoleces, dorsoventral view. **f** Scolex in apical view. **g** Ventral surface of gravid proglottids showing numerous papillae surrounding the genital atrium. **h–i** Eggs and surface of egg shell. **j** Detail of egg surface with pits

atrium devoid of vitelline follicles (Figs. 3c, 5c). Uterus tightly coiled, containing fully developed, but unembryonated eggs, forms bilateral uterine loops; uterine loops 3–4 (3, $n=5$) in number on each side of mid-line (Fig. 3c). Terminal part of uterus enlarged, forming a thick-walled sac-like structure (Fig. 3c, d). Uterine pore situated 16–152 (75 ± 44 , $n=7$) from anterior margin of proglottid, representing 5–31% (21%) of proglottid length. Eggs oval, thick-walled, operculated (Fig. 4h, i), 59–75 (65 ± 3 ; $n=152$) long by 40–47 (43 ± 2 ; $n=152$) wide. Egg shell slightly pitted, with around 39–57 ($n=2$) pits/ $10 \mu\text{m}^2$ (Fig. 4j).

Remarks

The new species is placed in the genus *Diphyllobothrium* because of its typical shape of the scolex, composition of genital organs, absence of transverse papilla-like tegumental protuberances on the ventral surface of the proglottids and using pinnipeds as its definitive host (see [4]).

Kuchta and Scholz [3] reported 20 species of '*Diphyllobothrium*' (*incertae sedis*) from the intestine of pinnipeds. Of these, *D. sprakeri* n. sp. can be distinguished from *D. archeri* (Leiper & Atkinson, 1914), *D. cameroni* Rausch, 1969, *D. cordatum* (Leuckart, 1863), *D. elegans* (Krabbe,



1865), *D. fayi* Rausch, 2005, *D. hians* (Diesing, 1850), *D. lanceolatum* (Krabbe, 1865), *D. lashleyi* (Leiper & Atkinson, 1914), *D. minutus* Andersen, 1987, *D. mobile* (Rennie & Reid, 1912), *D. phocarum* Delyamure, Kurochkin & Skryabin, 1964, *D. pseudowilsoni* Wojciechowska & Zdzitowiecki, 1995, *D. quadratum* (von Linstow, 1892), *D. rauschi* Andersen, 1987, *D. roemeri* (Zschokke, 1903), *D. schistochilos* (Germanos, 1895), *D. tetrapterum* (von Siebold, 1848) and *D. wilsoni* (Shiple, 1907) by having an enlarged thick-walled sac-like structure in the terminal part of the uterus in mature and gravid proglottids [41].

Recently, Hermosilla et al. [24] collected two headless tapeworms from fecal samples of SASLs in Chile and considered them a '*Diphyllobothrium scoticum*-like cestode.' The morphological description of these tapeworms by Hermosilla et al. [24] is incomplete; however, they are similar to *D. sprakeri* n. sp. in having a thick-walled sac-like structure in the terminal part of the uterus, longitudinal muscle layer wide (51–100 vs 100) and egg size (54–61 × 38–44 vs 59–75 × 40–47). The new species slightly differs from the the '*Diphyllobothrium scoticum*-like cestode' by having smaller gravid proglottids (287–736 × 2681–7890 vs 825–1385 × 7077–7418) and number of uterine loops (3–4 vs 5–6). However, the size of diphyllobothriids and the number of uterine loops have a limited taxonomic value in distinguishing species (see [2, 5] and references therein). Our phylogenetic analyses confirmed that specimens of *D. sprakeri* n. sp. recovered

from CSLs are conspecific with those from SASLs from Chile (Figs. 1, 2).

Two species of '*Diphyllobothrium*' described from Antarctic phocids, i.e. *D. scoticum* and *D. lobodoni* Yurakhno & Maltsev, 1994, are similar to *D. sprakeri* n. sp. in having a thick-walled sac-like structure in the terminal part of the uterus [22, 42–46]. The new species can be distinguished from *D. scoticum* and *D. lobodoni* by its smaller scolex (<1484 vs >1800 and >1900, respectively), absence of a neck, lower number of uterine loops (3–6 vs 5–17 and 7–22, respectively) and somewhat smaller eggs (59–75 × 39–47 vs 60–100 × 42–53 and 71–74 × 51–53, respectively) (minimum and maximum range for biometrical data for *D. scoticum* estimated from all available descriptions; see Table 1 for details). '*Diphyllobothrium sprakeri*' n. sp. can be further differentiated from *D. scoticum* and *D. lobodoni* in the natural definitive hosts and the disparate geographical distribution (otariids from temperate waters of North and South America vs Antarctic phocids) (see below).

Our molecular analyses reported immature specimens of *A. pacificus* and *D. sprakeri* n. sp. in the intestine of a SASL from Peru. Immature specimens of both species are morphologically indistinguishable. Adult specimens of *A. pacificus* can be distinguished from the new species by the presence of the papilla-like protuberances anterior to the male gonopore [2] and the absence of a thick-walled sac-like structure in the terminal part of

the uterus. To our knowledge, this is the first confirmed report of coinfection of two species of diphylobothriid tapeworms in a single otariid from the Southern Hemisphere. Voucher specimens of *A. pacificus* collected in this study are deposited in the Laboratory of Epidemiology and Veterinary Economics (LEVE 910, 913, 917, 923, 925), Lima, Peru.

The voucher material identified as '*Diphylobothrium latum*' from CSLs off Mexico and the USA deposited in London (NHML) was substantially decomposed. These specimens were immature without developed genital organs crucial for identification. However, *D. sprakeri* n. sp. is similar to these specimens in the shape of the scolex and the absence of a neck. '*Diphylobothrium sprakeri* n. sp. has been the only species of '*Diphylobothrium*' reported from the intestine of CSLs ([18], present study). Based on this evidence, the voucher material deposited in London is tentatively conspecific with the new species. One voucher specimen deposited in London (NHML 1980.6.3.189) was collected from a CSL stranded on the Mexican coast. In Mexico, CSLs are distributed along the east and west coasts of the Baja California Peninsula [47]. Therefore, this peninsula may represent an additional locality for *D. sprakeri* n. sp.

Discussion

The taxonomy of diphylobothriids is insufficiently resolved. Identification of individual species is complicated because of their uniform strobilar morphology, the high amount of intraspecific and intraindividual variation for most morphological characters and incomplete original descriptions [3, 8].

Diphylobothriids of pinnipeds have been revised by the present authors for more than 10 years based on detailed morphological examination of well-fixed material combined with molecular data (see [2, 4–6, 8, 40]). Our previous studies suggested that otariids are only definitive hosts of three diphylobothriid species: *A. pacificus*, widely distributed in both hemispheres, and *D. tetrapterum* and *Pyramicocephalus phocarum* (Fabricius, 1780), limited to the Northern Hemisphere [2, 5, 8]. However, a revision of the metazoan parasites of CSLs recognized a new undescribed diphylobothriid tapeworm which is different from these three species [4, 5, 18]. This species showed a uterine sac-like structure in mature and gravid proglottids, which is an uncommon character in diphylobothriids. '*Diphylobothrium sprakeri* n. sp. is the first diphylobothriid species described from CSLs and, with *A. pacificus*, the second valid species of otariids from the Southern Hemisphere.

A uterine sac-like structure was described in *D. scoticum* from leopard seals *Hydrurga leptonyx* (Blainville) from several localities in Antarctica [22, 42–46]. This tapeworm species has also been reported in Weddell seals *Leptonychotes weddellii* (Lesson) in Antarctica and recently confirmed using molecular markers from southern elephant seals *Mirounga leonina* (L.) from Macquarie Island (Southwestern Pacific Ocean) [8, 46, 48].

Yurahkno and Maltsev [45] described *D. lobodoni* from the intestine of crabeater seals *Lobodon carcinophagus* (Hombron & Jacquinot) from Antarctica. This diphylobothriid species differs from *D. scoticum* by the size of the strobila, scolex, neck and uterine sac-like structure, the shape of proglottids, thickness of the tegument and muscle layer, number of testes and the position of the cirrus sac and external seminal vesicle. However, the size of the strobila and scolex and number of testes and other structures may depend on the fixation methods, host species, its size, physiological state or intensity of infection (see [5], and references therein) and are not suitable characters in species delimitation [2, 3]. Moreover, most of the used discriminant characteristics of *D. lobodoni* overlap with those of *D. scoticum* reported from leopard seals (type-host) by other authors (Table 1). Further studies of the type material and molecular data from the type-host of *D. lobodoni* are necessary to confirm the validity and systematic position of this species.

Our study suggests that *D. sprakeri* n. sp. has a wide geographical distribution in both hemispheres, including the Pacific and Atlantic Oceans, and infects at least two otariid species. The distribution of *D. sprakeri* n. sp. in the Northern Hemisphere is limited to the Pacific coast of California, USA, and Baja California, Mexico. Interestingly, our new species has not been recorded in the North Pacific coast, where the diversity of diphylobothriid tapeworms from otariids has been comprehensively evaluated in recent years [2, 5, 49, 50]. In the Southern Hemisphere, *D. sprakeri* n. sp. is more widely distributed, occurring in temperate waters of the Pacific coast of South America (Peru and Chile) and the Southwest Atlantic along the Patagonian coast of Argentina.

The life cycle of *D. sprakeri* n. sp. probably includes marine fishes as the second intermediate hosts. Recently, Mondragón Martínez [51] reported plerocercoids of *A. pacificus* and an unidentified species of *Diphylobothrium* in marine fishes from Peru based on partial *cox1* sequences. According to the phylogenetic analysis of Mondragón Martínez [51], unidentified diphylobothriid plerocercoids, collected from anchoveta *Engraulis ringens* Jenyns and Pacific jack mackerel *Trachurus symmetricus* (Ayres), formed a clade sister to *A. pacificus*

Table 1 Comparison of selected biometrical data among '*Diphyllobothrium sprakeri* n. sp.', '*Diphyllobothrium scoticum*' and '*Diphyllobothrium lobodoni*'. The incomplete description of *D. scoticum* from leopard seals from Macquarie Island, Antarctica, by Johnston [44] is not included. Measurements in micrometers, unless otherwise stated

Species	<i>D. sprakeri</i> n. sp.	<i>D. sprakeri</i> n. sp. ^{1a}	<i>D. scoticum</i> ^c	<i>D. scoticum</i> ^d
Host	<i>Zalophus californianus</i> (Lesson)	<i>Otaria flavescens</i> Shaw	<i>Hydrurga leptonyx</i> (Blainville)	<i>H. leptonyx</i>
Locality	California, USA	Los Lagos Region, Chile	Antarctica	Antarctica
GenBank accession no	MW600337–MW600339 (<i>IsrDNA</i>); MW596661, MW596666–MW596673, MW596675–MW596678, MW596680 (<i>cox1</i>)	KY945917 (<i>IsrDNA</i>); KY945922, MF893274 (<i>cox1</i>)	–	–
Reference	Present study	Hermosilla et al. [24, 25]	Rennie and Reid [42]	Fuhrmann [43] ^d
Strobila length (cm)	< 200 × 0.8	> 50	13.3–29	13.3
Scolex	999–1484 × 807–1275	–	1800–2500 × 1500	1800–3000 × 700–2500
Neck	Absent	–	Short	Absent
Gravid proglottid (cm)	0.03–0.07 × 0.27–0.79	0.08–0.14 × 0.71–0.74	0.09 × 0.15	0.12 × 0.55
Testes diameter or size	44–70	150 ^b	69–87	100–160
Ovary width	1029–1550	–	–	–
Number of uterine loops	3–4	5–6	> 4	5–7
Uterine sac-like structure	Present	Present	Present	Present
Egg size	59–75 × 40–47	54–61 × 39–44	70–100 × 43–51	64–80 × 44–48
Species	<i>D. scoticum</i>	<i>D. scoticum</i>	<i>D. lobodoni</i>	<i>D. scoticum</i>
Host	<i>H. leptonyx</i>	<i>H. leptonyx</i>	<i>Lobodon carcinophagus</i> (Hombron & Jacquinot)	<i>H. leptonyx</i>
Locality	Debenham Islands, Antarctica	Balleny Islands, D'Urville Sea, Antarctica	Balleny Islands, D'Urville Sea, Antarctica	King George Island, South Shetland Islands
GenBank accession no	–	–	–	–
Reference	Markowski [22]	Yurakhno and Maltsev [45]	Yurakhno and Maltsev [45]	Wojciechowska and Zdzitowiecki [46]
Strobila length (cm)	52–130	5.6–42	44.2–240	16–150
Scolex	3500 × 2000	1900–3500 (length)	1900–2870 (length)	2200–4600 × 1100–2300
Neck length	495	370–1500	910–1290	Present
Gravid proglottid (cm)	0.50–0.80 × 1.50–1.80	–	0.35 × 1.35	0.16–0.8 × 0.13–0.6
Testes diameter or size	150–210 × 150	69–193	100–220	67–150 × 47–1400
Ovary width	–	–	4900 × 5100	–
Number of uterine loops	5–12	5–17	7–22	–
Uterine sac-like structure	Present	Present	Present	Present
Egg size	76–79 × 56	68–76 × 50–53	71–74 × 51–53	60–85 × 42–56

^a Specimens without scoleces referred to as '*Diphyllobothrium scoticum*-like cestode'

^b Testes measured in transverse section

^c Metrical data on *Dibothriocephalus pygoscelis* Rennie & Reid, 1912, which is considered a junior synonym of *D. scoticum* (see Johnston [44], Markowski [22]), is included

^d Fuhrmann [43] examined the holotype of *D. scoticum*

and *Diphyllobothrium* spp. Unfortunately, partial *cox1* sequences of these diphylobothriid plerocercoids are not available in the GenBank dataset. These plerocercoids probably belong to *D. sprakeri* n. sp.; however, sequences generated from these plerocercoids need to be analyzed in a more robust phylogenetic context for reliable species identification.

The Pacific broad tapeworm *Adenocephalus pacificus* is considered the most important causative agent of diphylobothriosis among humans in South America [52]. Diphylobothriosis caused by this species has been reported predominantly in Peru, where human infections are associated with the habits of consuming raw or undercooked marine fishes [23, 52]. Our new species

may be also causative agent of human fish-borne disease in the Pacific coast of South America, but not yet recognized and certainly misidentified as *A. pacificus*. Molecular-based diagnoses represent the most reliable tool to identify clinical samples of diphyllbothriid tapeworms [53], especially because clinical samples or immature specimens of *A. pacificus* and *D. sprakeri* n. sp. could be morphologically indistinguishable.

Abbreviations

BI: Bayesian inference; *cox1*: Cytochrome c oxidase subunit 1 gene; *cs*: Cirrus sac; CSL: California sea lion; *esv*: External seminal vesicle; *ga*: Genital atrium; IPCAS: Helminthological Collection of the Institute of Parasitology; LEVE: Parasite Collection of the Laboratory of Epidemiology and Veterinary Economics; *IsrDNA*: Large ribosomal subunit nuclear ribosomal RNA gene; *mg*: Mehlis' gland; ML: Maximum likelihood; NMNH-USNM: National Museum of Natural History of the Smithsonian Institution; *ov*: Ovary; PCMNH: Parasite Collection of the Museum of Natural History; SASL: South American sea lion; *te*: Testes; *up*: Uterine pore; *uss*: Uterine sac-like structure; *ut*: Uterus; *vf*: Vitelline follicle; *vg*: Vagina.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13071-021-04661-1>.

Additional file 1: Table S1. List of taxa used in the phylogenetic analyses.

Additional file 2: *IsrDNA*_alignment. Trimmed *IsrDNA* alignment (34 taxa; 1574 bp).

Additional file 3: *cox1*_alignment. Trimmed *cox1* alignment (59 taxa; 1571 bp).

Additional file 4: *IsrDNA_cox1*_alignment. Combined *IsrDNA* + *cox1* alignment (33 taxa; 3145 bp).

Additional file 5: Figure S1. Bayesian analysis for the combined (*IsrDNA* + *cox1*) alignment. Numbers represent posterior probabilities from BI analysis (> 0.95 shown only) followed by nodal supports from ML analysis (bootstrap values > 70% shown only). The newly generated sequences are indicated in red. The scale bar indicates the expected number of substitutions per site.

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Authors' contributions

JSH-O, TK and LG-P obtained the samples. LG-P contributed to the major part of the sequencing. JSH-O and LG-P undertook phylogenetic analyses. JSH-O, TK and RK performed the identification and morphological characterisation of the tapeworms. All authors discussed the results, helped draft the MS, read and approved the final manuscript.

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Availability of data and materials

The type- and voucher material is deposited in the Helminthological Collection of the Institute of Parasitology, České Budějovice, Czech Republic, the National Museum of Natural History of the Smithsonian Institution, Washington DC, USA, the Parasite Collection of the Museum of Natural History, National University of San Marcos, Lima, Peru, and the Parasite Collection of the Laboratory of Epidemiology and Veterinary Economics, National University of San Marcos, Lima, Peru (see Results for accession numbers). DNA sequences generated in this study were deposited in the GenBank database (see Additional file 1: Table S1 for details). Alignments used in this study are included as additional files.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Caira JN, Jensen K. Planetary biodiversity inventory (2008–2017): Tapeworms from Vertebrate Bowels of the Earth. Lawrence: University of Kansas, Natural History Museum, Special Publication No. 25; 2017.
- Hernández-Orts JS, Scholz T, Brabec J, Kuzmina T, Kuchta R. High morphological plasticity and global geographical distribution of the Pacific broad tapeworm *Adenocephalus pacificus* (syn. *Diphyllbothrium pacificum*): molecular and morphological survey. *Acta Trop.* 2015;149:168–78.
- Kuchta R, Scholz T. Diphyllbothriidea. In: Caira JN, Jensen K, editors. Planetary Biodiversity Inventory (2008–2017): Tapeworms from Vertebrate Bowels of the Earth. Lawrence: University of Kansas, Natural History Museum, Special Publication No. 25; 2017. p. 167–89.
- Waeschenbach A, Brabec J, Scholz T, Littlewood DTJ, Kuchta R. The catholic taste of broad tapeworms—multiple routes to human infection. *Int J Parasitol.* 2017;47:831–43.
- Hernández-Orts JS, Scholz T, Brabec J, Kuzmina T, Kuchta R. Does the number of genital organs matter? Case of the seal tapeworm *Diphyllbothrium* (syn. *Diplogonoporos*) *tetrapterum* (Cestoda: Diphyllbothriidea). *Can J Zool.* 2018;96:193–204.
- Schaeffner BC, Ditrich O, Kuchta R. A century of taxonomic uncertainty: re-description of two species of tapeworms (Diphyllbothriidea) from Arctic seals. *Polar Biol.* 2018;41:2543–59.

7. Mariaux J, Kuchta R, Hoberg EP. 2017. Tetrabothriidea Baer, 1954. In: Caira JN, Jensen K, editors. Planetary Biodiversity Inventory (2008–2017): Tapeworms from Vertebrate Bowels of the Earth. Lawrence: University of Kansas, Natural History Museum, Special Publication No. 25;2017: p. 357–70.
8. Scholz T, Kuchta R, Brabec J. Broad tapeworms (Diphyllobothriidae), parasites of wildlife and humans: recent progress and future challenges. *Int J Parasitol Parasites Wildl.* 2019;9:359–69.
9. Carretta JV, Forney KA, Lowry MS, Barlow J, Baker J, Hanson B, Muto MM. U.S. Pacific Marine Mammal Stock Assessments: 2007. Washington: US Department of Commerce NOAA Technical Memorandum, NMFS–SWFSC–414; 2007.
10. McClatchie S, Field J, Thompson AR, Gerrodette T, Lowry M, Fiedler PC, et al. Food limitation of sea lion pups and the decline of forage off central and southern California. *R Soc Open Sci.* 2016;3:150628.
11. Stunkard HW. Variation and criteria for generic and specific determination of diphyllobothriid cestodes. *J Helminthol.* 1965;39:281–96.
12. Dailey MD, Hill BL. A survey of metazoan parasites infesting the California (*Zalophus californianus*) and Steller (*Eumetopias jubatus*) sea lion. *Bull South Calif Acad Sci.* 1970;69:126–32.
13. Dailey MD, Brownell RL Jr. A checklist of marine mammal parasites. In: Ridgway SH, editor. Mammals of the sea, biology and medicine. Springfield: Charles C. Thomas; 1972. p. 528–89.
14. Margolis L, Dailey MD. Revised annotated list of parasites from sea mammals caught off the west coast of North America. NOAA technical report NMFS, SSRF-647. Seattle: National Oceanic and Atmospheric Administration; 1972.
15. Dailey MD. The distribution and intraspecific variation of helminth parasites in pinnipeds. *Rapp P-V Réun Cons Int Explor Mer.* 1975;169:338–52.
16. Dailey MD. Baseline data on parasites from marine mammals in the Southern California Bight area. Marine mammal and seabird survey of the Southern California Bight area. Vol. III. Principal investigator's reports. Book 1. Pinnipedia, Cetacea and Parasitology. Santa Cruz: California University; 1978. p. 442–72.
17. Felix JR. Reported incidences of parasitic infections in marine mammals from 1892 to 1978. Lincoln: Zea E-Books; 2013.
18. Kuzmina TA, Spraker TR, Kudlai O, Lisitsyna O, Zabludovskaja SO, Karbowiak G, et al. Metazoan parasites of California sea lions (*Zalophus californianus*): a new data and review. *Int J Parasitol Parasites Wildl.* 2018;7:326–34.
19. Crespo EA, Oliva D, Dans S, Sepúlveda M. Current status of the South American sea lion along the distribution range. Valparaíso: Editorial Universidad de Valparaíso; 2012.
20. Cárdenas-Alayza S, Crespo EA, Oliveira LR. *Otaria byronia*. The IUCN Red List of Threatened Species; 2016. <https://doi.org/10.2305/IUCN.UK.2016-1.RLTS.T41665A61948292.en>
21. Baylis HA, in (sic) Hamilton JE. The Southern sea lion, *Otaria byronia* (De Blainville). *Discovery Rep.* 1934;8:306.
22. Markowski S. The cestodes of seals from the Antarctic. *Brit Bull Nat Hist.* 1952;1:125–50.
23. Baer JG, Miranda H, Fernandez W, Medina TJ. Human diphyllobothriasis in Peru. *Z Parasitenkd.* 1967;28:227–89.
24. Hermosilla C, Hirtzmann J, Silva LMR, Scheufen S, Prenger-Berninghoff E, Ewers C, et al. Gastrointestinal parasites and bacteria in free-living South American sea lions (*Otaria flavescens*) in Chilean Comau Fjord and new host record of a *Diphyllobothrium scoticum*-like cestode. *Front Mar Sci.* 2018;5:459.
25. Ebmer D, Navarrete MJ, Muñoz P, Flores LM, Gärtner U, Brabec J, et al. Anthropozoonotic parasites circulating in synanthropic and Pacific colonies of South American sea lions (*Otaria flavescens*): Non-invasive techniques data and a review of the literature. *Front Mar Sci.* 2020;7:543829.
26. Spraker TR. Basic Necropsy Procedures. In: McCurnin DM, editor. Clinical textbook for the animal health technician. London: W.B. Saunders Company; 1985. p. 4702–816.
27. Pleijel F, Jondelius U, Norlinder E, Nygren A, Oxelman B, Schander C, et al. Phylogenies without roots? A plea for the use of vouchers in molecular phylogenetic studies. *Mol Phylogenet Evol.* 2008;48:369–71.
28. Littlewood DTJ, Curini-Galletti M, Herniou EA. The interrelationships of Proseriata (Platyhelminthes: Seriata) flatworms tested with molecules and morphology. *Mol Phylogenet Evol.* 2000;16:449–66.
29. Olson PD, Cribb TH, Tkach VV, Bray RA, Littlewood DTJ. Phylogeny and classification of the Digenea (Platyhelminthes: Trematoda). *Int J Parasitol.* 2003;33:733–55.
30. Bowles J, McManus DP. Genetic characterization of the Asian *Taenia*, a newly described taeniid cestode of humans. *Am J Trop Med Hyg.* 1994;50:33–44.
31. Wicht B, Yanagida T, Scholz T, Ito A, Jiménez JA, Brabec J. Multiplex PCR for differential identification of broad tapeworms (Cestoda: *Diphyllobothrium*) infecting humans. *J Clin Microbiol.* 2010;48:3111–6.
32. Brabec J, Scholz T, Králová-Hromadová I, Bazsalovicsová E, Olson PD. Substitution saturation and nuclear paralogs of commonly employed phylogenetic markers in the Caryophyllida, an unusual group of non-segmented tapeworms (Platyhelminthes). *Int J Parasitol.* 2012;42:259–67.
33. Gomez-Puerta LA, Alarcon V, Pacheco J, Franco F, Lopez-Urbina MT, Gonzalez AE. Molecular and morphological evidence of *Taenia omisssa* in pumas (*Puma concolor*) in the Peruvian Highlands. *Rev Bras Parasitol Vet.* 2016;25:368–73.
34. Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 2004;32:1792–2179.
35. Darriba D, Taboada GL, Doallo R, Posada D. jModelTest 2: more models, new heuristics and parallel computing. *Nat Methods.* 2012;9:772.
36. Huelsenbeck JP, Ronquist F. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics.* 2001;17:754–5.
37. Edler D, Klein J, Antonelli A, Silvestro D. raxmlGUI 2.0; 2019. <https://antonellilab.github.io/raxmlGUI/>
38. Rambaut A. FigTree v. 2.0; 2018. <http://tree.bio.ed.ac.uk/software/>
39. Kumar S, Stecher G, Li M, Niyaz C, Tamura K. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol.* 2018;35:1547–9.
40. Chervy L. Unified terminology for cestode microtriches: a proposal from the International Workshops on Cestode Systematics in 2002–2008. *Folia Parasitol.* 2009;56:199–230.
41. Delyamure SL, Skryabin AS, Serdiukov AM. Diphyllobothriata—Flatworm parasites of man, mammals and birds. In: Sudarikov VE, editor. Principles of Cestodology. Vol. 11. Moscow: Izdatel'stvo Nauka; 1985, p. 200. **(in Russian)**
42. Rennie J, Reid A. The Cestoda of the Scottish Antarctic Expedition (Scotia). *Trans R Soc Edinburgh.* 1912;48:441–53.
43. Fuhrmann O. Die Cestoden der Deutschen Südpolar-Expedition 1901–1903. *Deutsche Südpol-Exped.* 1920;1901–1903(16):469–524.
44. Johnston TH. The Cestoda of the Australasian Antarctic Expedition. *Sci Rep Aust Antarctic Exped.* 1937;10:1–74.
45. Yurakhno MV, Maltsev VN. *Diphyllobothrium lobodoni* sp. n. (Cestoda: Diphyllobothriidae)—The parasite of the crabeater seal. *Parazitologiya.* 1994;28:270–5. **(in Russian)**
46. Wojciechowska A, Zdzitowiecki K. Cestodes of Antarctic seals. *Acta Parasitol.* 1995;40:125–31.
47. Sztteren D, Auriolos D, Gerber LR. Population status and trends of the California sea lion (*Zalophus californianus californianus*) in the Gulf of California, Mexico. In: Trites AW, Atkinson SK, DeMaster DP, Fritz LW, Gelatt TS, Rea LD, Wynne KM, editors. Sea Lions of the World. Fairbanks: Alaska Sea Grant College Program; 2006. p. 369–84.
48. Yurakhno MV, Maltsev VN. An infection of seals from Antarctica with cestodes. *Parazitologiya.* 1997;31:81–9 **(in Russian)**.
49. Kuzmina TA, Hernández-Orts JS, Lyons ET, Spraker TR, Kornyushyn VV, Kuchta R. The cestode community in northern fur seals (*Callorhinus ursinus*) on St. Paul Island, Alaska. *Int J Parasitol Parasites Wildl.* 2015;4:256–63.
50. Kuzmina TA, Kuzmin YI, Dzeverin I, Lisitsyna OI, Spraker TR, Korol EM, Kuchta R. Review of metazoan parasites of the northern fur seal

(*Callorhinus ursinus*) and the analysis of the gastrointestinal helminth community of the population on St. Paul Island, Alaska. *Parasitol Res.* 2021;120:117–32.

51. Mondragón Martínez AM. Identificación molecular de los estadios de plerocercarioide y adulto de *Diphyllobothrium* sp. obtenidos de humanos, lobos marinos y peces. Bachelor Thesis. Lima: Universidad Ricardo Palma; 2017. p. 82. <http://repositorio.urp.edu.pe/handle/urp/1007>.
52. Kuchta R, Serrano-Martínez ME, Scholz T. Pacific broad tapeworm *Adenocephalus pacificus* as a causative agent of globally reemerging diphyllobothriosis. *Emerg Infect Dis.* 2015;21:1697–703.
53. Scholz T, Garcia HH, Kuchta R, Wicht B. Update on the human broad tapeworm (genus *Diphyllobothrium*), including clinical relevance. *Clin Microbiol Rev.* 2009;22:146–60.

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