

1 **The pro- and anti-inflammatory activity of fatty acids**

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16 **Abstract**

17 Inflammation is crucial to maintain homeostasis in the body. The contribution of fatty
18 acids to the inflammatory process is exerted through a variety of mechanisms leading to cell
19 surface modifications, activation of intracellular receptors that control inflammatory signaling
20 processes, and changes in gene expression patterns. While long-chain saturated fatty acids induce
21 NFkB pathway activation through TLR-4 binding, unsaturated fatty acids, such as
22 monounsaturated, polyunsaturated and conjugated fatty acids anti-inflammatory ability is
23 mediated through PPARs or GPR120. Moreover, these unsaturated fatty acids, especially omega-
24 3 fatty acids, have immunomodulatory and cytoprotective potential, which is highly relevant for
25 diseases with a neuroinflammatory component, such as obesity, Alzheimer's disease, multiple
26 sclerosis and schizophrenia.

27 **Keywords**

28 Bioactive lipids; fatty acids; pro-inflammatory; anti-inflammatory; neuroinflammation; omega-3

29 *Abbreviations*

30 2-arachidonoylglycerol (2-AG); Alpha-linolenic acid (ALA); Alzheimer's disease (AD); Arachidonic acid
31 (AA); Arachidonylethanolamide (AEA); Blood-brain barrier (BBB); Ceramide 1-phosphate (C1P); Cyclooxygenase
32 (COX); Conjugated linoleic acid (CLA); Conjugated linolenic acid (CLNA); Damage-associated molecular patterns
33 (DAMPs); Docosahexaenoic acid (DHA); Eicosapentaenoic acid (EPA); Endocannabinoids (eCBs); G protein coupled
34 receptors (GPRs); G protein receptor 120 (GPR120); high-fat diet (HFD); Insulin receptor substrate (IRS); Linoleic
35 acid (LA); lipopolysaccharide (LPS); lysophosphatidic acid (LPA); lysophosphatidilinositol (LPI);
36 lysophosphatidylcholine (LPC); Long-chain saturated fatty acids (LC-SFAs); Monounsaturated fatty acids (MUFAs) ;
37 Multiple Sclerosis (MS); Myeloid differentiation primary response 88 (Myd88); nuclear factor kappa B (NFkB);
38 inhibitory subunit (IκB); Palmitoylethanolamide (PEA); Pathogen-associated molecular patterns (PAMPs); Peroxisome
39 proliferator activated receptors (PPARs); Polyunsaturated fatty acids (PUFAs); Prostaglandins (PGs); Polyunsaturated
40 fatty acids (PUFAs); Saturated fatty acids (SFAs); Specialized pro-resolving mediators (SPMs); sphingosine 1-
41 phosphate (S1P); Tetrahydrocannabinol (THC); TGF-β activated kinase 1 (TAK1); TGF-β activated kinase binding
42 protein 1 (TAB1); T helper cells (Th); Tumor necrosis factor (TNF); Toll-like receptors (TLR); type-1 and type-2
43 cannabinoid receptors (CB1 and CB2)

44

45 **1. Introduction: inflammation**

46 Inflammation is defined as the process through which the immune system recognizes and
47 removes harmful and foreign stimuli and begins the healing process (Fritsch & Abreu, 2019;
48 Michels da Silva et al., 2019; Xufei Zhang et al., 2019). It comprises an intricate network of
49 cellular and molecular events, where newly synthesized mediators are produced to obtain a
50 temporal and spatial response (Chiurchiù et al., 2018). Inflammation can be either acute or
51 chronic. On one hand, acute inflammation is a rapid and often severe process that occurs following
52 tissue damage, microbial invasion, or exposure to noxious compounds which symptoms may last
53 for a few days. On the other hand, chronic inflammation is a slow, long-term inflammation that
54 can last for prolonged periods (several months to years) (Pahwa et al., 2021). The self-regulation
55 of the inflammatory response relies on the activation of several negative feedback mechanisms,
56 that when lost result in increased concentration of inflammatory mediators and activated
57 inflammatory cells both at the site of tissue damage and in the systemic circulation, leading to
58 excessive and irreparable damage in host tissues (Calder, 2012a).

59 **1.1. The role of lipids as inflammatory mediators**

60 Lipids are one of the most important inflammatory mediators: they are involved in all
61 phases of inflammation and in the regulation of several important inflammatory processes. Thus,
62 due to their importance in immune regulation, inflammation and maintenance of tissue
63 homeostasis, several lipids have been classified as bioactive lipids. Bioactive lipids are generally
64 divided into four main families according to their biochemical functions: classical eicosanoids,
65 specialized pro-resolving mediators (SPMs), lysoglycerophospholipids/sphingolipids and
66 endocannabinoids (eCBs). These lipids are generated from omega-6 or omega-3 essential
67 polyunsaturated fatty acids (PUFAs) precursors, that are esterified into membrane lipids and act
68 by binding to and activating specific G protein coupled receptors (GPRs) (Chiurchiù et al., 2018).

69 In the presence of an inflammatory stimuli, such as tissue insults or infections,
70 granulocytes and monocytes/macrophages (innate immune cells) are recruited to the site of injury
71 and generate a class of lipids mediators, eicosanoids, which are responsible for acute
72 inflammation (Nathan, 2002). Classical eicosanoids lipids are highly pro-inflammatory and signal
73 via autocrine and paracrine mechanisms. During the last stage of inflammation when there is
74 resolution of inflammation, innate immune cells switch the production of eicosanoids lipids to
75 another class of bioactive lipids, the SPMs (Chiurchiù et al., 2018). SPMs terminate inflammation
76 and drive the restoration of full tissue homeostasis (Basil & Levy, 2016; Serhan, 2014). In cases
77 when the inflammation is not resolved, it turns into a chronic inflammation state (or
78 histophlogosis) resulting in aberrant tissue remodeling and organ dysfunction (Nathan & Ding,
79 2010). In such context, other bioactive lipids, such as lysoglycerophospholipids/sphingolipids and
80 eCBs, mediate several cellular processes promoting cell and tissue adaption to the inflammatory
81 milieu (Chiurchiù et al., 2015; El Alwani et al., 2006).

82 **1.1.1. Eicosanoids**

83 Eicosanoids are a class of bioactive lipids derived from 20-carbon PUFAs, most
84 frequently from the omega-6 arachidonic acid (AA), which is released from membrane
85 phospholipids. This class of molecules include the prostaglandins (PGs), thromboxanes,
86 leukotrienes and lipoxins. Such molecules usually act on short distances and time periods,
87 autocrinally, in the cells that produce them, or paracrinally in neighboring cells (Macouzet et al.,
88 2010).

89 Prostaglandins, specifically, seem to be involved in the sustained inflammation that
90 causes the transition to chronic inflammation by enhancing cytokine production (Aoki &

91 Narumiya, 2012; Narumiya & Furuyashiki, 2011). Indeed, prostaglandins induce chronic
92 inflammation through enhancement of the pro-inflammatory release cascade (Honda et al., 2006),
93 amplification of innate immunity response to pathogen- and damage-associated molecular
94 patterns (PAMPs and DAMPs) (Hirata & Narumiya, 2012), activation of specific pro-
95 inflammatory subsets of T helper (Th) cells (Q. Chen et al., 2010; Yao et al., 2009), recruitment
96 of immune cells associated with chronic inflammation, such as macrophages, T and B cells, by
97 synergistically acting with chemokines (Aoki & Narumiya, 2012) and by increasing the
98 expression of pro-inflammatory genes induced by cytokines (Chiurchiù et al., 2015).

99 **1.1.2. Lysoglycerophospholipids/sphingolipids**

100 Lysoglycerophospholipids and sphingolipids are bioactive lipids with glycerol or
101 sphingosine as backbones, respectively. They can link with other molecules, such as
102 ethanolamine, choline, inositol, serine or/and fatty acids to form many different compounds
103 distributed in plasma membranes. Importantly, lysophospholipids are derived from membrane
104 phospholipids by removal of one or both fatty acids, being lysophosphatidylcholine (LPC) and
105 lysophosphatidylinositol (LPI) as well as their byproduct lysophosphatidic acid (LPA) the most
106 biologically active. Such lipids are important signaling molecules involved in relevant aspects of
107 cellular and tissue biology, including plasma membrane shaping, cell growth and death, and
108 inflammatory cascades. LPC and LPA modulate immune responses by controlling distribution,
109 trafficking and activation of immune cells (Chiurchiù et al., 2018; D'Aquilio et al., 2007;
110 Knowlden & Georas, 2014; Piñeiro & Falasca, 2012; Sevastou et al., 2013). Moreover, their
111 sustained activation has been linked with several chronic inflammatory diseases like obesity and
112 diabetes (Heimerl et al., 2014; Moreno-Navarrete et al., 2012), cancer (Piñeiro & Falasca, 2012),
113 atherosclerosis (H. Lee et al., 2002) and rheumatoid arthritis (Fuchs et al., 2005; Nikitopoulou et
114 al., 2012).

115 Regarding sphingolipids, the main active ones are ceramide and their byproducts
116 ceramide 1-phosphate (C1P) and sphingosine 1-phosphate (S1P). Sphingolipids are important
117 bioactive lipids participating in numerous inflammatory processes including intracellular
118 trafficking and signaling, cell growth, adhesion, vascularization, survival, and apoptosis
119 (Chiurchiù et al., 2018). As reviewed by (Chiurchiù et al., 2018) the role of sphingolipids in
120 chronic inflammation has been widely assessed and most studies have associated it with
121 immunodependent and vascular-related chronic inflammatory diseases (*e.g.* diabetes and obesity,
122 neuroinflammatory disorders, chronic obstructive pulmonary disease and inflammatory bowel
123 disease). Indeed, excessive ceramide signaling is associated with the development of adipose
124 tissue inflammation and insulin resistance which ultimately leads to obesity and type 2 diabetes.
125 C1P enhances both acute and chronic inflammatory responses by promoting phospholipase A2-
126 mediated eicosanoid storm and by inducing cytokine production. However, ceramide and C1P
127 have also been shown to negatively regulate some proinflammatory cytokines. In addition,
128 ceramide and its metabolites are also involved in the physiological regulation of
129 endothelial/vascular integrity and function, and alterations in these sphingolipids are associated
130 with vascular dysfunctions and therefore with chronic inflammatory states. Regarding S1P, it is
131 an important mediator for lymphocyte trafficking between lymphoid and non-lymphoid tissues
132 allowing the egress of effector T and B cells from lymph nodes, thymus, bone marrow, and spleen
133 and blocking the ability of immature dendritic cells to migrate. This is highly relevant since T and
134 B cells are important initiators of many chronic inflammatory conditions and autoimmune
135 diseases.

136 **1.1.3. Endocannabinoids**

137 eCBs are a group of bioactive lipids that are endogenously produced by humans and
138 animals and are recognized due to their ability to bind and activate the same receptors as the Δ^9 -

139 tetrahydrocannabinol (THC), the main psychoactive component of marijuana, the type-1 and
140 type-2 cannabinoid receptors (CB1 and CB2) (Chiurchiù et al., 2018). Besides CB1 and CB2
141 cannabinoid receptors, peroxisome proliferator activated receptors (PPARs) and GPR55 are also
142 engaged by some cannabinoids (Lu & Mackie, 2016). The eCBs molecules as well as their
143 enzymes and receptors constitute the eCBs system, which serves as a homeostatic system
144 controlling several physiopathological states. Indeed, this system is a widespread
145 neuromodulatory system that plays important roles in CNS development, synaptic plasticity and
146 the response to endogenous and environmental insults (Chiurchiù et al., 2018; Lu & Mackie,
147 2016).

148 The two best studied members of eCBs family are arachidonylethanolamide (commonly
149 known as anandamide, AEA) and 2-arachidonoylglycerol (2-AG). 2-AG-ester, O-
150 arachidonylethanolamine, palmitoylethanolamide (PEA) are also members of the eCBs family.
151 Such molecules are ubiquitously produced by most tissues and immune cells, which are capable
152 of metabolize them with a specific set of enzymes (Chiurchiù et al., 2018). AEA and PEA are
153 mostly anti-inflammatory, and 2-AG acts both as pro- and anti-inflammatory.

154 The eCBs are amongst the most potent immunoregulatory compounds capable of
155 regulating the function of several cell subsets of either innate or adaptive immunity. In
156 consequence, perturbations in all members of the eCBs system occur during every chronic
157 inflammatory process, from cancer, metabolic, and gastrointestinal diseases to autoimmune and
158 neuroinflammatory disorders. Moreover, the modulation of this system has been shown to be
159 beneficial by attenuating inflammatory processes.

160 **2. The pro-inflammatory actions of fatty acids**

161 Fatty acids are carboxylic acids presenting a long aliphatic chain. Such chain can be
162 straight or branched and saturated (only single bond) or unsaturated (one or more double bond).
163 These molecules, which are naturally occurring constituents of diet, have metabolic, structural
164 and functional roles in the human body. They are usually used as energy sources: β -oxidation of
165 the fatty acids is a well-known process, mostly used by the heart and the muscular tissue to obtain
166 energy (Nagy & Tiuca, 2017a). Nevertheless, different fatty acids can present different biological
167 activities. The influence of fatty acids on inflammation has been widely studied and it is known
168 to be exerted through a variety of mechanisms. Indeed, fatty acids can act at the cell surface, as
169 intracellular receptors that control inflammatory signaling processes, and can influence gene
170 expression patterns. Besides, fatty acids are important players in cell membrane function and
171 modification in the fatty acid composition of the cell membrane can impact its fluidity, lipid raft
172 formation and cell signaling, which can ultimately result in altered gene expression and in
173 alterations in lipid and peptide's mediators production (Calder, 2012b; Nagy & Tiuca, 2017b).
174 Thus, it is widely accepted that fatty acids can affect inflammatory cell function and consequently
175 inflammatory processes; for instance, they can act directly in surface or intracellular receptors,
176 they can be incorporated in the membrane phospholipids of inflammatory cells and they can act
177 as precursors of extracellular signaling molecules such as PGs, as mentioned in the previous
178 section (Calder, 2012b).

179 **2.1. Long-chain saturated fatty acids**

180 Long-chain saturated fatty acids (LC-SFAs) are fatty acids with 14 or more carbon atoms,
181 with no double bonds. In general, saturated fatty acids (SFAs) are important energy sources, used
182 as building blocks for structural elements, protein modification and regulation of gene
183 transcription. Adipose tissue and liver are able to *de novo* synthesize and store SFAs, namely
184 palmitic acid (C16:0) from precursors such as glucose (Ruiz-Núñez et al., 2016a). The role of
185 SFAs, namely dietary SFAs, as pro-inflammatory molecules was unraveled because obesity was

186 found to be associated with a low-grade inflammatory state, both in adipose tissue and
187 hypothalamus. Thus, the mechanisms causing this inflammation were widely studied and the role
188 of high-fat diet (HFD), specifically LC-SFAs, elucidated. It was found that the SFA excess, itself
189 a characteristic of obesity, triggers cellular stress and the activation of pro-inflammatory pathways
190 mediated by JNK and nuclear factor kappa B (NFkB) (Ellulu et al., 2017). This raises the
191 circulating levels of several acute-phase proteins and inflammatory cytokines contributing to the
192 chronic low-grade inflammation state (Gómez-Hernández et al., 2016; Longo et al., 2019).
193 Moreover, several studies have identified toll-like receptors (TLR)-4, widely known for binding
194 bacterial lipopolysaccharide (LPS), as the mediator responsible for the activation of innate
195 immune response initiated by SFA (O'Neill, 2006). TLRs are known to be expressed in several
196 cells: macrophages, dendritic cells, adipocytes, hepatocytes and muscle. In addition, TLR- 2 and
197 -4 are also recognized binding sites for LC-SFAs in the hypothalamus. Indeed, studies have shown
198 that HFD significantly increases the expression level of TLR gene expression (Nam et al., 2017).
199 It is known that when activated, TLRs are responsible for the NFkB pathway activation, by
200 facilitating the NFkB translocation to the nucleus (Kawai & Akira, 2006; Ruiz-Núñez et al.,
201 2016b). Considering this and the ability of LC-SFAs to bind TLR-4, these SFAs have been shown
202 to induce NFkB activation in several *in vitro* models: endothelial cells (Harvey et al., 2010),
203 mHypoA-POMC/GFP-2 neurons (Tse & Belsham, 2018), cocultures of 3T3-L1 adipocytes and
204 RAW264 macrophages (Takayoshi et al., 2007), skeletal muscle cells (Hommelberg et al., 2009),
205 L6 muscle cells differentiated into myotubes (Nisr et al., 2019), among others. Under normal
206 conditions NFkB is associated with its inhibitory subunit (IκB) remaining inactive in the cytosol.
207 The phosphorylation of IκB by IKKβ, results in IκB dissociation from NFkB, allowing NFkB to
208 translocate into the nucleus to initiate transcription of its target genes (Xiaoqing Zhang et al.,
209 2008). Regarding NFkB pathway activation by LC-SFAs through TLR-4, this process is mediated
210 by myeloid differentiation primary response 88 (Myd88) activation. The activation of Myd88
211 leads to TGF-β activated kinase 1 (TAK1) activation and consequent interaction with TGF-β
212 activated kinase binding protein 1 (TAB1) resulting in NFkB activation (as reviewed by (Salsinha
213 et al., 2021)). Recently, the role of Myd88 role on HFD on hypothalamic inflammation was
214 elucidated in an astrocyte specific Myd88 knockout mice model: in the knockout mice the HFD
215 or SFAs-induced inflammation was ameliorated (Jin et al., 2020).

216 Besides their direct effect on NFkB pathway activation, SFAs can be incorporated into
217 specific lipid rafts domains of the plasma membrane. Such incorporation enhances TLR-4
218 dimerization, important for its activation (Wong et al., 2009) as well as the activation of
219 downstream signaling pathways, such as JNK/AP-1 (Ruiz-Núñez et al., 2016b). Moreover, SFAs
220 are known to be involved in the *de novo* synthesis of ceramides (mentioned in the introductory
221 section): serine and palmitoyl-CoA are condensed to form 3-ketosphingamine via a synergistic
222 signaling of TLR-4 and LPS (Schilling et al., 2013). Despite strong evidences in the role of TLR-
223 4 and -2 in inducing LC-SFAs induced inflammatory responses, the direct binding of SFAs to
224 TLR-4 has been challenged (Erridge & Samani, 2009; Ruiz-Núñez et al., 2016b; Salsinha et al.,
225 2021). Pal and colleagues (Pal et al., 2012) suggested that SFAs may interact with TLR-4 through
226 fetuin A. Fetuin-A (alpha-2 Heremans-Schmid glycoprotein) is extensively expressed in the
227 kidney, brain, skin, gastrointestinal tract, and in the liver, from where is predominantly secreted
228 in adults. Importantly, both proinflammatory and anti-inflammatory roles have been attributed to
229 it, and due to its proinflammatory role this glycoprotein has been associated with several
230 inflammatory diseases, including metabolic syndrome, nonalcoholic fatty liver disease,
231 atherosclerosis, coronary artery disease, and is considered to be an important predictor of
232 cardiovascular morbidity in diabetes (as reviewed by (Mukhopadhyay et al., 2014)). Indeed,
233 fetuin A can interact with a variety of receptors including TLRs (Mukhopadhyay et al., 2014). It
234 was shown that fetuin A binds and activated TLR-4 and can also play an important role in

235 polarizing adipose tissue M2 macrophages towards the pro-inflammatory M2 phenotype
236 (Chatterjee et al., 2013).

237 The proinflammatory potential of the mentioned fatty acids is summarized in Figure 1.

238 **2.2. Omega-6 fatty acids**

239 Polyunsaturated fatty acids (PUFAs) are unsaturated fatty acids which present two or
240 more double bonds. They can be classified accordingly to the position of the first double bond
241 relative to the methyl-end group: omega-6 fatty acids have the double bond 6 carbons atoms
242 away from the terminal methyl group and omega-3 fatty acids have the double bond 3 carbons
243 away from the terminal methyl group. Omega-3 PUFAs are synthesized from the essential fatty
244 acid alpha-linolenic acid (ALA) (C18:3 *c*9,*c*12,*c*15) and omega-6 PUFAs from the linoleic acid
245 (LA) (C18:2 *c*9,*c*12). Although humans and other animals can metabolize these essential fatty
246 (ALA and LA), they are not able to synthesize them (Brenna et al., 2009; Burdge & Wootton,
247 2002; Mišurcová et al., 2011; Orsavova et al., 2015; Proust et al., 2014). Thus, they have to be
248 obtained through dietary sources (Ander et al., 2003). The balance of omega-6 to omega-3
249 obtained through diet is very important to human health since overabundance of one type of fatty
250 acids will interfere with the metabolic production of the other type and even limit it.

251 Besides LA, arachidonic acid (AA) is the other most important omega-6 fatty acid. AA
252 can be synthesized by the conversion of LA after desaturation and elongation reactions (B. Chen
253 et al., 2012). Along with omega-3 eicosapentaenoic acid (EPA), AA is a precursor of the
254 eicosanoids (mentioned in the introductory section). In general, eicosanoids derived from n-6
255 PUFA are proinflammatory while eicosanoids derived from n-3 PUFA are anti-inflammatory
256 (Patterson et al., 2012). Besides eicosanoids, ARA is a precursor of other potent pro-inflammatory
257 mediators, such as PGs and leukotrienes. Considering this, several anti-inflammatory drugs
258 targeting the ARA pathway are used to control inflammatory processes in different diseases. It
259 was reported that higher dietary intake of omega-6 fatty acids, namely ARA or LA, leads to
260 increased inflammatory processes (Patterson et al., 2012). Diets, such as the western diet, with
261 high intake of omega-6 fatty acids, should in principle exacerbate the inflammatory response.
262 This diet results in increased omega-6:omega-3 ratios, resulting in augmented production of
263 mediators and regulators of inflammation and immune responses (Patterson et al., 2012).
264 However, studies in healthy humans have showed that increased intake of ARA or LA does not
265 increase concentrations of several inflammatory markers (Innes & Calder, 2018). Indeed, the pro-
266 inflammatory profile associated with western-pattern diet has been increasingly associated with
267 the overconsumption of LC-SFAs and not with the omega-6: omega-3 ratio, as discussed below.

268 **3. The anti-inflammatory potential of bioactive lipids**

269 In contrast to what is observed for HFD, it has been noticed that some diets, such as the
270 Mediterranean diet, are associated with lower risk of developing overweight and obesity (Notario-
271 Barandiaran et al., 2020). Such observations lead to the understanding that in contrast to LC-
272 SFAs, others fatty acids may possess beneficial effects on human health. In fact, the beneficial
273 effects of the Mediterranean diet are strongly related with unsaturated fatty acids consumption,
274 namely monounsaturated fatty acids (MUFAs) olive oil-based diet. Indeed, a MUFA rich diet was
275 shown to reduce macrophage infiltration and to allow the shift to a more anti-inflammatory profile
276 with production of anti-inflammatory cytokines IL-10 and IL-4 (Montserrat-de la Paz et al.,
277 2019). Other study, aiming at addressing the impact of the omega-7 palmitoleic acid (C16:1 *c*9)
278 MUFA on atherosclerosis in mice, has shown that palmitoleic acid supplementation reduces the
279 expression of IL-1 β and tumor necrosis factor (TNF)- α genes (Z. Yang et al., 2020). It has been
280 suggested that this anti-inflammatory effect is mediated by PPAR- γ , since it was observed that
281 MUFA diet induces the expression of the corresponding gene (Montserrat-de la Paz et al., 2019).

282 Importantly, PPAR α , β/δ and γ are nuclear receptors that translate nutritional and/or
283 pharmacologic stimuli into changes in gene expression and are involved in the regulation of
284 inflammation, immunity and epithelial cell differentiation. (Bassaganya-Riera, Reynolds,
285 Martino-Catt, Cui, Hennighausen, Gonzalez, Rohrer, Benninghoff, Hontecillas, et al., 2004;
286 Cunard et al., 2002; Jones et al., 2002; Natarajan & Bright, 2002; P Tontonoz et al., 1994; Peter
287 Tontonoz et al., 1994; Y. L. Wang et al., 2002; Zakaria, 2014). While LC-SFAs induce NFkB
288 activation through TLR-4, the palmitoleic acid MUFA was shown to downregulate NFkB
289 pathway through PPAR- γ stimulation (de Souza et al., 2018). Nonetheless, *in vivo* studies have
290 shown that palmitoleic acid interacts with G protein receptor 120 (GPR120) receptor and that its
291 activation is responsible for the resolution of palmitic acid-induced inflammation (Hirasawa et
292 al., 2005; Ichimura et al., 2012). Thus, it has been suggested that MUFAs can inhibit NFkB
293 through direct binding of PPARs or GPR120, inhibiting its activation by SFAs (Ravaut et al.,
294 2021).

295 Similarly to MUFAs, PUFAs are widely known for their anti-inflammatory potential,
296 thought to be mediated through the GPR120 receptor. In fact, some PUFAs, mostly omega-3 fatty
297 acids - ALA, docosahexaenoic acid (DHA) and EPA – are proven activators of GPR120 (Oh et
298 al., 2010). It has been suggested that activation of GPR120 by these omega-3 fatty acids leads to
299 the recruitment of β -arrestin 2, and to the formation and subsequent internalization of a GPR120-
300 β -arrestin 2 complex. Such complex interacts with TAB1, inhibiting its interaction with another
301 protein, the TAK1. This inhibition is highly relevant since their interaction mediates downstream
302 inflammatory processes by activating NFkB and JNK pathways. Thus, GPR120 activation by
303 omega-3 fatty acids inhibits the activation of pro-inflammatory pathways, reverting the
304 inflammatory action of SFAs via TLR 4 receptors (Salsinha et al., 2021; Talukdar et al., 2011).
305 Importantly, GPR120 is highly expressed in adipocytes and macrophages suggesting a potential
306 anti-inflammatory action of PUFAs in several tissues and a possible therapeutical role in several
307 inflammatory diseases. In addition, DHA was able to inhibit cyclooxygenase (COX)-2 gene
308 expression in macrophages bearing constitutively active TLR-4 but not in those bearing
309 constitutively active Myd88 - used by TLR-4 to activate NFkB -, suggesting that the effects of
310 DHA on the NFkB pathway produce a direct effect on TLR-4 signaling (Calder, 2012b; J. Y. Lee
311 et al., 2001). Additionally, EPA and DHA dietary intake increase their proportion in cell
312 membranes. Such effect decreases the generation of pro-inflammatory molecules derived from
313 omega-6 PUFAs (Martínez Leo et al., 2019).

314 Other PUFAs, namely conjugated fatty acids – conjugated linoleic acid (CLA) and
315 conjugated linolenic acid (CLNA) – are also known for their anti-inflammatory potential. It was
316 observed that CLA – a group of positional and geometric isomers of LA – exerts a PPAR- γ anti-
317 inflammatory action on colitis, by repression of TNF- α expression and NFkB activation, while
318 inducing the expression of the immunoregulatory cytokine TGF- β 1 (Bassaganya-Riera,
319 Reynolds, Martino-Catt, Cui, Hennighausen, Gonzalez, Rohrer, Benninghoff, & Hontecillas,
320 2004). Regarding CLNA, the punicic acid isomer, was associated with suppression of NFkB
321 activation and TNF- α expression. Such action was also mediated by PPAR- γ , since loss of PPAR-
322 γ impaired the ability of dietary punicic acid to suppress inflammation (Hontecillas et al., 2009).

323 **4. Bioactive lipids and brain: role of fatty acids on neuroinflammation**

324 The role of PUFAs in the brain especially during early development is widely recognized:
325 they play important biological roles, including receptor binding, neurotransmission, signal
326 transduction and eicosanoid synthesis. In fact, PUFAs represent 35% of total lipids in adult brain.
327 Specifically, AA, DHA and EPA play important roles during brain development and in the
328 maintenance of normal brain structure and function. DHA, for instance, accounts for 30 to 40%

329 of fatty acids in the gray matter of the cortex, being particularly concentrated in synaptic
330 membranes (Chew et al., 2020; Lauritzen et al., 2016; Liu et al., 2015).

331 Moreover, the bioactive fatty acids obtained from dietary intake can cross the blood-brain
332 barrier (BBB) and reach the CNS where they accumulate (Nadjar et al., 2016). Interestingly, *in*
333 *vivo* studies in mice showed that high-SFA diet altered the levels of several lipids, and increased
334 the levels of palmitic acid (LC-SFA) in brain. Later, the authors demonstrated that after entering
335 the brain, SFAs are transported into the hypothalamus and are taken up by microglia cells
336 (Valdearcos et al., 2014).

337 **4.1. Obesity and neuroinflammation: the dietary fatty acids role**

338 There is an increasing interest in obesity that despite being a preventable disease has
339 currently reached a worldwide pandemic level and plays a central role in the development of non-
340 communicable diseases. As reviewed by Salsinha and colleagues (Salsinha et al., 2021)
341 hypothalamus inflammation triggered by diet has been showing to play a relevant role in the onset
342 and progression of this disease. Several studies have reported that HFD, particularly LC-SFAs, is
343 responsible for inducing inflammatory processes in both CNS and peripheral tissues, including
344 the adipose tissue (Argente-Arizón et al., 2015; Valdearcos et al., 2014). In fact, 1 to 3 days of
345 HFD consumption is sufficient to elevate inflammatory markers in *in vivo* models (Thaler et al.,
346 2012). Importantly, the inflammatory potential of these fatty acids is associated with a persistent
347 elevation of microglia reactivity and consequent TNF- α secretion (Yi et al., 2017). The LC-SFA
348 palmitic acid induces proinflammatory activation of microglia (Duffy et al., 2015; Valdearcos et
349 al., 2014; Z. Wang et al., 2012) via TLR-4 mediated NF κ B pathway induction (Duffy et al., 2015;
350 Milanski et al., 2009; Z. Wang et al., 2012). The effects of LC-SFAs on IKK β and JNK have been
351 associated to insulin resistance (Ono, 2019). HFD-induced inflammation leads to increased
352 activation of the intracellular kinases JNK and NF κ B, through IKK β (Lawrence, 2009), which
353 induces phosphorylation of the insulin receptor substrate (IRS) on its serine residues and inhibits
354 phosphorylation on its tyrosine residues, critical to the transmission of the insulin signal to
355 downstream effectors and for biological outcomes (Rorato et al., 2017). Insulin resistance causes
356 disruption of insulin signaling processes in the CNS and suppresses the effect of insulin on food
357 intake. Moreover, besides the homeostatic imbalance caused by such processes, insulin resistance
358 is also described as the common link between obesity and the development of type 2 diabetes (W.
359 Chen et al., 2017).

360 On the other hand, omega-3 fatty acids, through dietary supplementation, have been
361 showing promising results in ameliorating the obesity effects caused by the HFD, specifically by
362 LC-SFAs. Several of the omega-3 beneficial effects have been attributed to their anti-
363 inflammatory potential (Pimentel et al., 2012; Viggiano et al., 2016) thought to be mediated
364 through the GPR120 receptor (Oh et al., 2010). It has been suggested that activation of GPR120
365 by omega-3 fatty acids abrogates NF κ B and JNK pathway activation (Talukdar et al., 2011).
366 Furthermore, Oh Da et al. (2010) reported that DHA stimulation of GPR120 inhibits both TLR
367 2/3/4 and the TNF- α proinflammatory cascade. Recently, DHA intracerebroventricular injection
368 in male C57BL/6J mice ameliorated the HFD-induced hypothalamic inflammation (Cheng et al.,
369 2020). Some *in vitro* studies have also reported relevant results in microglia cell lines: DHA was
370 able to reverse LPS inflammatory effects in N9 microglia cells (Chang et al., 2015; De Smedt-
371 Peyrusse et al., 2008).

372 Other PUFAs are also recognized for their anti-inflammatory potential, namely
373 conjugated fatty acids. For instance, CLA has been showing interesting results regarding HFD-
374 induced obesity symptoms, including normalization of serum leptin levels (Ryan et al., 2011; Y.-
375 M. Wang et al., 2005; Yanagita et al., 2005) and positive energy balance (Ryan et al., 2011). CLA
376 isomers incorporation into the brain has been detected in few cases at very low concentrations

377 (Alasnier et al., 2002; Murru et al., 2021). C18:2 *c9,t11* and C18:2 *t10,c12* were demonstrated to
378 be actively incorporated into the rat brain and in *in vitro* astrocyte cultures (Fa et al., 2005). Since
379 it is known that CLA is an activator of PPARs, specifically PPAR- α , it has been suggested that
380 CLA anti-inflammatory actions in the CNS are possibly connected to the activation of such
381 factors. Indeed, PPAR- α anti-inflammatory action is mediated through repression of many
382 activated transcription factors, such as NF κ B, among others. The same anti-inflammatory
383 potential was attributed to CLNA. A study demonstrated that pomegranate seed oil (a source of
384 punicic acid CLNA isomer) affected the morphology of activated BV-2 microglia cells. The
385 authors suggested an immunomodulatory and cytoprotective potential comparable to that of
386 omega-3 PUFAs, in neuroinflammatory disease caused by obesity (Račková et al., 2014).

387 Besides PUFAs, the possible beneficial role of MUFAs on brain, was recently assessed
388 both *in vitro* and *in vivo*, using microglia cells (BV-2 cell line) and C57BL/6J mice, respectively.
389 In both models using an olive oil source, MUFAs enhanced the microglia anti-inflammatory
390 phenotype, while SFAs polarize microglia towards a pro-inflammatory profile (Toscano et al.,
391 2020). Such results suggest a potential positive impact of olive oil MUFAs (particularly oleic
392 acid) in neuroinflammation.

393 **4.2. Alzheimer's disease**

394 Epidemiological and clinical research studies have shown that a high dietary intake of
395 PUFAs is associated with lower risk for Alzheimer's disease (AD) and related dementias.
396 Moreover, high intake of fish and unsaturated fatty acids are associated with reduced cognitive
397 decline (Gustafson et al., 2020). Importantly, DHA plays an important role in the maintenance of
398 optimal membrane functions and is known to reduce amyloid deposition. This is highly relevant
399 since amyloid deposits are present in AD patients (Gustafson et al., 2020). In addition, increased
400 ratios of omega-6: omega-3 fatty acids have been associated with a higher risk of AD
401 (MacDonald-Wicks et al., 2019). In addition, SPMs, synthesized from omega-3 fatty acids DHA
402 and EPA, are important to resolve inflammation through inhibition of polymorphonuclear
403 leukocytes and lowering vascular permeability. It has been suggested that this process is impaired
404 in AD (Chew et al., 2020; Whittington et al., 2017). Moreover, many studies have also reported
405 alterations to the eicosanoid pathway in AD (Biringer, 2019). Considering omega-3 fatty acids
406 role on SPMs and eicosanoids and their potential in resolving the inflammatory processes
407 underlying this disease, they have attracted a great deal of interest (Martínez Leo et al., 2019).

408 Importantly, in the CNS the activation of microglia and its associated cytokine production
409 are responsible for inflammatory responses. Unregulated inflammation, excessive cytokine
410 production and failure to resolve inflammatory responses, contribute to chronic
411 neuroinflammation, which is a hallmark of many neurodegenerative diseases, including AD (as
412 reviewed by (Chew et al., 2020)). Omega-3 supplementation inhibits microglial activation and
413 the subsequent inflammatory response by regulating the translocation and nuclear secretion of
414 high mobility group protein B1 (HMGB1). This factor is important since it mediates microglia
415 activation through the TLR4/ NF- κ B pathway, responsible for neuroinflammation after neuronal
416 injury (Martínez Leo et al., 2019).

417 **4.3. Multiple Sclerosis**

418 Multiple sclerosis is considered an autoimmune disorder first described in 1868 by Jean-
419 Martin Charcot. The French neurologist framed the disease as *sclérose en plaques*, which meant
420 multiple sclerosis (MS) in French. MS is characterized by myelin loss (demyelination),
421 accompanied by inflammation within the CNS, which are called the plaques or lesions (Kihara,
422 2019). The development of MS depends on both genetic and environmental factors, but several
423 studies have reported that impairment of Th cells are involved (Rezapour-Firouzi, 2017). In

424 summary, Th1 and Th2 cells are the major type of Th cells; the formers produce IL-2 and TNF-
425 α . Th2 cells produce IL-4, IL-5, IL-10 and IL-13,6,7. It is the balance between Th1 and Th2 cells
426 that is considered one of the risk factors of MS. Indeed, altered cytokine profiles in CNS tissue
427 and peripheral blood mononuclear cells have been discovered in MS patients (Imitola et al., 2005).
428 It has been reported that in MS several Th1 cytokines levels are increased, while Th2 cytokines
429 levels are decreased (Rezapour-Firouzi, 2017).

430 In 1952, Swank suggested that the variations in MS geographical distribution is related
431 with the amount and the type of fat consumption (*1952-Swank-Multiple-Sclerosis-In-Rural-*
432 *Norway*, 2010). Nowadays, several studies demonstrated that abnormalities of polyunsaturated
433 fatty acids (PUFAs) synthesis may be involved in this disease. Indeed, increased risk for MS
434 development was associated with HFD (Ghadirian et al., 1998). Moreover, when comparing the
435 values between normal and MS patients the saturated/ unsaturated fatty acids ratio is significantly
436 higher in the latter (OLIVEIRA et al., 2019). The increased concentrations of lysolecithin and its
437 SFAs is being appointed as a possible explanation for some changes in MS patients, namely in
438 the platelet behavior.

439 Moreover, a relation between dyslipidemia and early stages of MS has been found.
440 Indeed, there is a correlation between high density lipoprotein-cholesterol and inflammatory
441 cytokines in MS, confirming the existence of a relationship between lipid metabolism and
442 inflammation (Rádiková et al., 2020). Furthermore, omega-6 fatty acids have been associated
443 with the pathogenesis as well as treatment of MS. Studies evidenced that there is a disturbance in
444 omega-6 fatty acid metabolism in MS and that this disturbance has been linked with dysregulation
445 of cytokines. Indeed, the disturbance of omega-6 metabolism originates an important loss of
446 membrane omega-6 fatty acids LA and AA, which was associated with a decrease in TGF- β .
447 Moreover, the loss of these fatty acids also impact CNS structure and function and, in the long
448 term, can lead to neurological deficits associated with MS (Harbig & Sharief, 2007).

449 Omega-3 supplementation was first demonstrated in 1995, to being able to modulate
450 some immune functions altered in MS patients (Gallai et al., 1995). Recently, as reviewed by
451 (Labuschagne & Blaauw, 2018) although omega-3 fatty acids seem to have a relative low
452 influence on disease progression they reduce the frequency of relapsing episodes. Moreover, fish
453 oil supplementation was also effective in the reduction of pro-inflammatory cytokines and nitric
454 oxide levels in patients with relapsing-remitting MS (Ramirez-Ramirez et al., 2013).
455 Nevertheless, in 2017 the European Society for Clinical Nutrition and Metabolism stated that
456 there is not enough evidence regarding a positive effect of omega-3 to recommend such
457 supplementation in multiple sclerosis patients (Burgos et al., 2018).

458 **4.4.Schizophrenia**

459 Schizophrenia is a neuropsychiatric disorder with disabling symptoms and a lower life
460 expectancy (Ouyang et al., 2020).

461 The association between an abnormal lipid content in peripheral tissues and violence in
462 patients with schizophrenia has been reported: low blood levels of total cholesterol and low-
463 density lipoproteins, as well as higher blood triglycerides levels, were associated with violence
464 and a higher suicide risk in schizophrenia patients. Furthermore, disturbance of PUFA metabolism
465 was also associated with schizophrenia, where omega-3 deficits is related with psychiatric
466 symptoms in schizophrenia (as reviewed by (Ouyang et al., 2020)). In fact, low levels of EPA and
467 DHA as well as the ratio of EPA/ AA in red blood cells are associated with hostile behaviour in
468 schizophrenia patients (Watari et al., 2010). Such effects are related with the fact that omega-3,
469 mainly DHA, is important to membrane integrity. Its deficiency alters the structure and function

470 of membranes and induce dysfunctions in neurotransmission of serotonin and dopamine (Chalon,
471 2006; Healy-Stoffel & Levant, 2018; Ouyang et al., 2020)

472 Besides, neuroinflammation has been suggested to play an important role in the
473 pathogenesis of schizophrenia. Uncontrolled microglia activation increases the production of pro-
474 inflammatory factors, which ultimately result in neuronal impairments. Omega-3 fatty acids can
475 control inflammation. Indeed, omega-3 PUFAs, such as EPA and DHA, have anti-inflammatory
476 properties since they can produce resolvins and cytokines and have an inhibitory effect on
477 microglia activation.

478 **5. Conclusion**

479 Fatty acids are important cellular mediators which act on cell membrane integrity and
480 function, as intracellular signaling molecules and can influence gene expression. Thus, they are
481 important mediators in several inflammatory processes. Different fatty acids have different roles
482 in the human organism. For instance, TLR-4 was identified as the mediator responsible for the
483 activation of the innate immune response initiated by LC-SFAs, ultimately resulting in NFkB
484 activation. While LC-SFAs have a pro-inflammatory profile, the MUFAs and conjugated fatty
485 acids, CLA and CLNA, were shown to downregulate NFkB pathway through PPAR- γ
486 stimulation. Furthermore, *in vivo* studies have shown that MUFAs along with PUFAs interact
487 with the GPR120 receptor and that its activation is responsible for the inhibition of NFkB
488 activation. Thus, it has been suggested that these unsaturated fatty acids can inhibit NFkB through
489 direct binding of PPARs or GPR120, blocking its activation by LC-SFAs. Notably,
490 neuroinflammation plays an important role in the pathogenesis of schizophrenia, AD, obesity and
491 MS. For instance, uncontrolled microglia activation increases the production of pro-inflammatory
492 factors, which ultimately results in neuronal impairments. Indeed, omega-3 PUFAs, such as EPA
493 and DHA, anti-inflammatory properties have an modulatory effect on microglia, inhibiting their
494 activation and the subsequent inflammatory response.

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499 **7. Conflicts of interest**

500 The authors declare no conflict of interest.

501 **8. List of tables**

502 **Table 1** – Summary of some relevant studies available since 2017 regarding fatty acids' effects
503 on Alzheimer's disease (AD).

504 **Table 2** - Summary of some relevant studies available since 2017 regarding fatty acids' effects
505 on Multiple Sclerosis (MS).

506 **Table 3** - Summary of some relevant studies available since 2017 regarding fatty acids' effects
507 on Schizophrenia.

508 **9. List of Figures**

509 **Figure 1** - Pro- and anti-inflammatory action of different fatty acid.

510

511

512 **10. References**

- 513 *1952-Swank-Multiple-Sclerosis-In-Rural-Norway*. (2010).
- 514 Alasnier, C., Berdeaux, O., Chardigny, J. M., & Sebedio, J. L. (2002). Fatty acid composition and
515 conjugated linoleic acid content of different tissues in rats fed individual conjugated linoleic
516 acid isomers given as triacylglycerols small star, filled. *The Journal of Nutritional*
517 *Biochemistry*, *13*(6), 337–345.
- 518 Ander, B. P., Dupasquier, C. M., Prociuk, M. A., & Pierce, G. N. (2003). Polyunsaturated fatty
519 acids and their effects on cardiovascular disease. *Experimental and Clinical Cardiology*,
520 *8*(4), 164–172. <https://pubmed.ncbi.nlm.nih.gov/19649216>
- 521 Aoki, T., & Narumiya, S. (2012). Prostaglandins and chronic inflammation. *Trends in*
522 *Pharmacological Sciences*, *33*(6), 304–311. <https://doi.org/10.1016/j.tips.2012.02.004>
- 523 Argente-Arizón, P., Freire-Regatillo, A., Argente, J., & Chowen, J. A. (2015). Role of non-
524 neuronal cells in body weight and appetite control. *Frontiers in Endocrinology*, *6*(MAR),
525 1–15. <https://doi.org/10.3389/fendo.2015.00042>
- 526 Basil, M. C., & Levy, B. D. (2016). Specialized pro-resolving mediators: endogenous regulators
527 of infection and inflammation. *Nature Reviews. Immunology*, *16*(1), 51–67.
528 <https://doi.org/10.1038/nri.2015.4>
- 529 Bassaganya-Riera, J., Reynolds, K., Martino-Catt, S., Cui, Y., Hennighausen, L., Gonzalez, F.,
530 Rohrer, J., Benninghoff, A. U., & Hontecillas, R. (2004). Activation of PPAR γ and δ by
531 conjugated linoleic acid mediates protection from experimental inflammatory bowel
532 disease. *Gastroenterology*, *127*(3), 777–791. <https://doi.org/10.1053/j.gastro.2004.06.049>
- 533 Bassaganya-Riera, J., Reynolds, K., Martino-Catt, S., Cui, Y., Hennighausen, L., Gonzalez, F.,
534 Rohrer, J., Benninghoff, A. U., Hontecillas, R., Yeo, G., Brand, M. D., Cortright, R. N.,
535 O’Rahilly, S., Montague, C., Vidal-Puig, A. J., Podolsky, D. K., & Blumberg, R. S. (2004).
536 Activation of PPAR γ and δ by conjugated linoleic acid mediates protection from
537 experimental inflammatory bowel disease. *Gastroenterology*, *127*(3), 777–791.
538 <https://doi.org/10.1053/j.gastro.2004.06.049>
- 539 Biringer, R. G. (2019). The Role of Eicosanoids in Alzheimer’s Disease. *International Journal of*
540 *Environmental Research and Public Health*, *16*(14).
541 <https://doi.org/10.3390/ijerph16142560>
- 542 Bjørnevik, K., Chitnis, T., Ascherio, A., & Munger, K. L. (2017). Polyunsaturated fatty acids and
543 the risk of multiple sclerosis. *Multiple Sclerosis Journal*, *23*(14), 1830–1838.
544 <https://doi.org/10.1177/1352458517691150>
- 545 Brenna, J. T., Salem, N., Sinclair, A. J., & Cunnane, S. C. (2009). α -Linolenic acid
546 supplementation and conversion to n-3 long-chain polyunsaturated fatty acids in humans.
547 *Prostaglandins Leukotrienes and Essential Fatty Acids*, *80*(2–3), 85–91.
548 <https://doi.org/10.1016/j.plefa.2009.01.004>
- 549 Burdge, G. C., & Wootton, S. A. (2002). Conversion of α -linolenic acid to eicosapentaenoic,
550 docosapentaenoic and docosahexaenoic acids in young women. *British Journal of Nutrition*,
551 *88*(04), 411. <https://doi.org/10.1079/BJN2002689>
- 552 Burgos, R., Bretón, I., Cereda, E., Desport, J. C., Dziewas, R., Genton, L., Gomes, F., Jesús, P.,
553 Leischker, A., Muscaritoli, M., Pouliá, K. A., Preiser, J. C., Van der Marck, M., Wirth, R.,
554 Singer, P., & Bischoff, S. C. (2018). ESPEN guideline clinical nutrition in neurology.
555 *Clinical Nutrition*, *37*(1), 354–396. <https://doi.org/10.1016/j.clnu.2017.09.003>
- 556 Calder, P. C. (2012a). Long-chain fatty acids and inflammation. *Proceedings of the Nutrition*
557 *Society*, *71*(2), 284–289. [https://doi.org/DOI: 10.1017/S0029665112000067](https://doi.org/DOI:10.1017/S0029665112000067)

- 558 Calder, P. C. (2012b). Long-chain fatty acids and inflammation. *Proceedings of the Nutrition*
559 *Society*, 71(2), 284–289. <https://doi.org/DOI: 10.1017/S0029665112000067>
- 560 Chalon, S. (2006). Omega-3 fatty acids and monoamine neurotransmission. *Prostaglandins,*
561 *Leukotrienes, and Essential Fatty Acids*, 75(4–5), 259–269.
562 <https://doi.org/10.1016/j.plefa.2006.07.005>
- 563 Chang, P. K., Khatchadourian, A., Mckinney, R. A., & Maysinger, D. (2015). *Docosahexaenoic*
564 *acid (DHA): a modulator of microglia activity and dendritic spine morphology*. 1–15.
565 <https://doi.org/10.1186/s12974-015-0244-5>
- 566 Chatterjee, P., Seal, S., Mukherjee, S., Kundu, R., Mukherjee, S., Ray, S., Mukhopadhyay, S.,
567 Majumdar, S. S., & Bhattacharya, S. (2013). Adipocyte fetuin-A contributes to macrophage
568 migration into adipose tissue and polarization of macrophages. *The Journal of Biological*
569 *Chemistry*, 288(39), 28324–28330. <https://doi.org/10.1074/jbc.C113.495473>
- 570 Chen, B., McClements, D. J., & Decker, E. A. (2012). Design of Foods with Bioactive Lipids for
571 Improved Health. *Annual Review of Food Science and Technology*, 4(1), 35–56.
572 <https://doi.org/10.1146/annurev-food-032112-135808>
- 573 Chen, Q., Muramoto, K., Masaaki, N., Ding, Y., Yang, H., Mackey, M., Li, W., Inoue, Y.,
574 Ackermann, K., Shirota, H., Matsumoto, I., Spyvee, M., Schiller, S., Sumida, T., Gusovsky,
575 F., & Lamphier, M. (2010). A novel antagonist of the prostaglandin E(2) EP(4) receptor
576 inhibits Th1 differentiation and Th17 expansion and is orally active in arthritis models.
577 *British Journal of Pharmacology*, 160(2), 292–310. [https://doi.org/10.1111/j.1476-](https://doi.org/10.1111/j.1476-5381.2010.00647.x)
578 [5381.2010.00647.x](https://doi.org/10.1111/j.1476-5381.2010.00647.x)
- 579 Chen, W., Balland, E., & Cowley, M. A. (2017). Hypothalamic Insulin Resistance in Obesity:
580 Effects on Glucose Homeostasis. *Neuroendocrinology*, 104(4), 364–381.
581 <https://doi.org/10.1159/000455865>
- 582 Cheng, L., Hu, T., Shi, H., Chen, X., Wang, H., Zheng, K., Huang, X.-F., & Yu, Y. (2020). DHA
583 reduces hypothalamic inflammation and improves central leptin signaling in mice. *Life*
584 *Sciences*, 257(118036), 118036. <https://doi.org/https://doi.org/10.1016/j.lfs.2020.118036>
- 585 Chew, H., Solomon, V. A., Fonteh, A. N., & Martins, I. J. (2020). *Involvement of Lipids in*
586 *Alzheimer ' s Disease Pathology and Potential Therapies The Importance of Cellular Lipid*
587 *Membranes*. 11(June), 1–28. <https://doi.org/10.3389/fphys.2020.00598>
- 588 Chiurchiù, V., Battistini, L., & Maccarrone, M. (2015). Endocannabinoid signalling in innate and
589 adaptive immunity. *Immunology*, 144(3), 352–364. <https://doi.org/10.1111/imm.12441>
- 590 Chiurchiù, V., Leuti, A., & Maccarrone, M. (2018). Bioactive lipids and chronic inflammation:
591 Managing the fire within. *Frontiers in Immunology*, 9(JAN).
592 <https://doi.org/10.3389/fimmu.2018.00038>
- 593 Cunard, R., DiCampli, D., Archer, D. C., Stevenson, J. L., Ricote, M., Glass, C. K., & Kelly, C.
594 J. (2002). WY14,643, a PPAR alpha ligand, has profound effects on immune responses in
595 vivo. *Journal of Immunology (Baltimore, Md. : 1950)*, 169(12), 6806–6812.
- 596 Cutuli, D., Landolfo, E., Nobili, A., De Bartolo, P., Sacchetti, S., Chirico, D., Marini, F., Pieroni,
597 L., Ronci, M., D'Amelio, M., D'Amato, F. R., Farioli-Vecchioli, S., & Petrosini, L. (2020).
598 Behavioral, neuromorphological, and neurobiochemical effects induced by omega-3 fatty
599 acids following basal forebrain cholinergic depletion in aged mice. *Alzheimer ' s Research &*
600 *Therapy*, 12(1), 150. <https://doi.org/10.1186/s13195-020-00705-3>
- 601 D'Aquilio, F., Procaccini, M., Izzi, V., Chiurchiu', V., Giambra, V., Carotenuto, F., Di Nardo, P.,
602 & Baldini, P. M. (2007). Activatory properties of lysophosphatidic acid on human THP-1
603 cells. *Inflammation*, 30(5), 167–177. <https://doi.org/10.1007/s10753-007-9034-2>

- 604 Dal Monte, M., Cammalleri, M., Locri, F., Amato, R., Marsili, S., Rusciano, D., & Bagnoli, P.
605 (2018). Fatty Acids Dietary Supplements Exert Anti-Inflammatory Action and Limit
606 Ganglion Cell Degeneration in the Retina of the EAE Mouse Model of Multiple Sclerosis.
607 In *Nutrients* (Vol. 10, Issue 3). <https://doi.org/10.3390/nu10030325>
- 608 De Smedt-Peyrusse, V., Sargueil, F., Moranis, A., Harizi, H., Mongrand, S., & Layé, S. (2008).
609 Docosahexaenoic acid prevents lipopolysaccharide-induced cytokine production in
610 microglial cells by inhibiting lipopolysaccharide receptor presentation but not its membrane
611 subdomain localization. *Journal of Neurochemistry*, *105*(2), 296–307.
612 <https://doi.org/10.1111/j.1471-4159.2007.05129.x>
- 613 de Souza, C. O., Valenzuela, C. A., Baker, E. J., Miles, E. A., Rosa Neto, J. C., & Calder, P. C.
614 (2018). Palmitoleic Acid has Stronger Anti-Inflammatory Potential in Human Endothelial
615 Cells Compared to Oleic and Palmitic Acids. *Molecular Nutrition and Food Research*,
616 *62*(20), 1–20. <https://doi.org/10.1002/mnfr.201800322>
- 617 Duffy, C. M., Yuan, C., Wisdorf, L. E., Billington, C. J., Kotz, C. M., Nixon, J. P., & Butterick,
618 T. A. (2015). Role of orexin A signaling in dietary palmitic acid-activated microglial cells.
619 *Neuroscience Letters*, *606*, 140–144. <https://doi.org/10.1016/j.neulet.2015.08.033>
- 620 El Alwani, M., Wu, B. X., Obeid, L. M., & Hannun, Y. A. (2006). Bioactive sphingolipids in the
621 modulation of the inflammatory response. *Pharmacology & Therapeutics*, *112*(1), 171–183.
622 <https://doi.org/10.1016/j.pharmthera.2006.04.004>
- 623 Erridge, C., & Samani, N. J. (2009). Saturated Fatty Acids Do Not Directly Stimulate Toll-Like
624 Receptor Signaling. *Arterioscler Thromb Vasc Biol*, *29*(11), 1944–1949.
625 <https://doi.org/10.1161/ATVBAHA.109.194050>
- 626 Fa, M., Diana, A., Carta, G., Cordeddu, L., Melis, M. P., Murru, E., Sogos, V., & Banni, S. (2005).
627 Incorporation and metabolism of c9,t11 and t10,c12 conjugated linoleic acid (CLA) isomers
628 in rat brain. *Biochimica et Biophysica Acta - Molecular and Cell Biology of Lipids*, *1736*(1),
629 61–66. <https://doi.org/10.1016/j.bbaliip.2005.06.010>
- 630 Fritsch, J., & Abreu, M. T. (2019). The Microbiota and the Immune Response: What Is the
631 Chicken and What Is the Egg? *Gastrointestinal Endoscopy Clinics of North America*, *29*(3),
632 381–393. <https://doi.org/10.1016/j.giec.2019.02.005>
- 633 Fuchs, B., Schiller, J., Wagner, U., Häntzschel, H., & Arnold, K. (2005). The
634 phosphatidylcholine/lysophosphatidylcholine ratio in human plasma is an indicator of the
635 severity of rheumatoid arthritis: Investigations by 31P NMR and MALDI-TOF MS. *Clinical
636 Biochemistry*, *38*(10), 925–933.
637 <https://doi.org/https://doi.org/10.1016/j.clinbiochem.2005.06.006>
- 638 Gallai, V., Sarchielli, P., Trequatrini, A., Franceschini, M., Floridi, A., Firenze, C., Alberti, A.,
639 Di Benedetto, D., & Stragliotto, E. (1995). Cytokine secretion and eicosanoid production in
640 the peripheral blood mononuclear cells of MS patients undergoing dietary supplementation
641 with n-3 polyunsaturated fatty acids. *Journal of Neuroimmunology*, *56*(2), 143–153.
642 [https://doi.org/10.1016/0165-5728\(94\)00140-j](https://doi.org/10.1016/0165-5728(94)00140-j)
- 643 Ghadirian, P., Jain, M., Ducic, S., Shatenstein, B., & Morisset, R. (1998). Nutritional factors in
644 the aetiology of multiple sclerosis: a case-control study in Montreal, Canada. *International
645 Journal of Epidemiology*, *27*(5), 845–852. <https://doi.org/10.1093/ije/27.5.845>
- 646 Gustafson, D. R., Mayeux, R., Scarmeas, N., Stern, Y., Manly, J. J., & Gu, Y. (2020). *Dietary
647 fatty acids and risk of Alzheimer ' s disease and related dementias : Observations from the
648 Washington Heights-Hamilton Heights-Inwood Columbia Aging Project (WHICAP)*.
649 1638–1649. <https://doi.org/10.1002/alz.12154>
- 650 Harbige, L. S., & Sharief, M. K. (2007). Polyunsaturated fatty acids in the pathogenesis and

- 651 treatment of multiple sclerosis. *British Journal of Nutrition*, 98(S1), S46–S53.
652 <https://doi.org/DOI: 10.1017/S0007114507833010>
- 653 Harvey, K. A., Walker, C. L., Pavlina, T. M., Xu, Z., Zaloga, G. P., & Siddiqui, R. A. (2010).
654 Long-chain saturated fatty acids induce pro-inflammatory responses and impact endothelial
655 cell growth. *Clinical Nutrition (Edinburgh, Scotland)*, 29(4), 492–500.
656 <https://doi.org/10.1016/j.clnu.2009.10.008>
- 657 Healy-Stoffel, M., & Levant, B. (2018). N-3 (Omega-3) Fatty Acids: Effects on Brain Dopamine
658 Systems and Potential Role in the Etiology and Treatment of Neuropsychiatric Disorders.
659 *CNS & Neurological Disorders Drug Targets*, 17(3), 216–232.
660 <https://doi.org/10.2174/1871527317666180412153612>
- 661 Heimerl, S., Fischer, M., Baessler, A., Liebisch, G., Sigrüener, A., Wallner, S., & Schmitz, G.
662 (2014). Alterations of plasma lysophosphatidylcholine species in obesity and weight loss.
663 *PloS One*, 9(10), e111348. <https://doi.org/10.1371/journal.pone.0111348>
- 664 Hirasawa, A., Tsumaya, K., Awaji, T., Katsuma, S., Adachi, T., Yamada, M., Sugimoto, Y.,
665 Miyazaki, S., & Tsujimoto, G. (2005). Free fatty acids regulate gut incretin glucagon-like
666 peptide-1 secretion through GPR120. *Nature Medicine*, 11(1), 90–94.
667 <https://doi.org/10.1038/nm1168>
- 668 Hirata, T., & Narumiya, S. (2012). *Chapter Five - Prostanoids as Regulators of Innate and*
669 *Adaptive Immunity* (F. W. B. T.-A. in I. Alt (Ed.); Vol. 116, pp. 143–174). Academic Press.
670 <https://doi.org/https://doi.org/10.1016/B978-0-12-394300-2.00005-3>
- 671 Hommelberg, P. P. H., Plat, J., Langen, R. C. J., Schols, A. M. W. J., & Mensink, R. P. (2009).
672 Fatty acid-induced NF-kappaB activation and insulin resistance in skeletal muscle are chain
673 length dependent. *American Journal of Physiology. Endocrinology and Metabolism*, 296(1),
674 E114-20. <https://doi.org/10.1152/ajpendo.00436.2007>
- 675 Honda, T., Segi-Nishida, E., Miyachi, Y., & Narumiya, S. (2006). Prostacyclin-IP signaling and
676 prostaglandin E2-EP2/EP4 signaling both mediate joint inflammation in mouse collagen-
677 induced arthritis. *The Journal of Experimental Medicine*, 203(2), 325–335.
678 <https://doi.org/10.1084/jem.20051310>
- 679 Hontecillas, R., Shea, M. O., Einerhand, A., Ba, D., & Dvm, J. B. (2009). Activation of PPAR γ
680 and α by Punicic Acid Ameliorates Glucose Tolerance and Suppresses Obesity-Related
681 Inflammation. *Journal of the American College of Nutrition*, 28(2), 184–195.
682 <https://doi.org/10.1080/07315724.2009.10719770>
- 683 Ichimura, A., Hirasawa, A., Poulain-Godefroy, O., Bonnefond, A., Hara, T., Yengo, L., Kimura,
684 I., Leloire, A., Liu, N., Iida, K., Choquet, H., Besnard, P., Lecoœur, C., Vivequin, S.,
685 Ayukawa, K., Takeuchi, M., Ozawa, K., Tauber, M., Maffeis, C., ... Froguel, P. (2012).
686 Dysfunction of lipid sensor GPR120 leads to obesity in both mouse and human. *Nature*,
687 483(7389), 350–354. <https://doi.org/10.1038/nature10798>
- 688 Imitola, J., Chitnis, T., & Khoury, S. J. (2005). Cytokines in multiple sclerosis: from bench to
689 bedside. *Pharmacology & Therapeutics*, 106(2), 163–177.
690 <https://doi.org/10.1016/j.pharmthera.2004.11.007>
- 691 Innes, J. K., & Calder, P. C. (2018). Omega-6 fatty acids and inflammation. *Prostaglandins,*
692 *Leukotrienes, and Essential Fatty Acids*, 132, 41–48.
693 <https://doi.org/10.1016/j.plefa.2018.03.004>
- 694 Jin, S., Kim, K. K., Park, B. S., Kim, D. H., Jeong, B., Kang, D., Lee, T. H., Park, J. W., Kim, J.
695 G., & Lee, B. J. (2020). Function of astrocyte MyD88 in high-fat- diet-induced
696 hypothalamic inflammation. *Journal of Neuroinflammation*, 17(195), 1–13.
- 697 Jones, D. C., Ding, X., & Daynes, R. A. (2002). Nuclear Receptor Peroxisome Proliferator-

- 698 activated Receptor alpha (PPAR alpha) Is Expressed in Resting Murine Lymphocytes. *The*
699 *Journal of Biological Chemistry*, 277(9), 6838–6845.
700 <https://doi.org/10.1074/jbc.M106908200>
- 701 Kawai, T., & Akira, S. (2006). TLR signaling. *Cell Death and Differentiation*, 13(5), 816–825.
702 <https://doi.org/10.1038/sj.cdd.4401850>
- 703 Kihara, Y. (2019). Systematic Understanding of Bioactive Lipids in Neuro-Immune Interactions:
704 Lessons from an Animal Model of Multiple Sclerosis. *Advances in Experimental Medicine*
705 *and Biology*, 1161, 133–148. https://doi.org/10.1007/978-3-030-21735-8_13
- 706 Knowlden, S., & Georas, S. N. (2014). The Autotaxin–LPA Axis Emerges as a Novel Regulator
707 of Lymphocyte Homing and Inflammation. *The Journal of Immunology*, 192(3), 851 LP –
708 857. <https://doi.org/10.4049/jimmunol.1302831>
- 709 Labuschagne, L., & Blaauw, R. (2018). An anti-inflammatory approach to the dietary
710 management of multiple sclerosis: A condensed review. *South African Journal of Clinical*
711 *Nutrition*, 31(3), 67–73. <https://doi.org/10.1080/16070658.2018.1465652>
- 712 Lauritzen, L., Brambilla, P., Mazzocchi, A., Harsløf, L. B. S., Ciappolino, V., & Agostoni, C.
713 (2016). DHA Effects in Brain Development and Function. *Nutrients*, 8(1), 6.
714 <https://doi.org/10.3390/nu8010006>
- 715 Lawrence, T. (2009). The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring*
716 *Harbor Perspectives in Biology*, 1(6), a001651–a001651.
717 <https://doi.org/10.1101/cshperspect.a001651>
- 718 Lee, H., Liao, J.-J., Graeler, M., Huang, M.-C., & Goetzl, E. J. (2002). Lysophospholipid
719 regulation of mononuclear phagocytes. *Biochimica et Biophysica Acta (BBA) - Molecular*
720 *and Cell Biology of Lipids*, 1582(1), 175–177.
721 [https://doi.org/https://doi.org/10.1016/S1388-1981\(02\)00153-1](https://doi.org/https://doi.org/10.1016/S1388-1981(02)00153-1)
- 722 Lee, J. Y., Sohn, K. H., Rhee, S. H., & Hwang, D. (2001). Saturated fatty acids, but not
723 unsaturated fatty acids, induce the expression of cyclooxygenase-2 mediated through Toll-
724 like receptor 4. *The Journal of Biological Chemistry*, 276(20), 16683–16689.
725 <https://doi.org/10.1074/jbc.M011695200>
- 726 Liu, J. J., Green, P., John Mann, J., Rapoport, S. I., & Sublette, M. E. (2015). Pathways of
727 polyunsaturated fatty acid utilization: implications for brain function in neuropsychiatric
728 health and disease. *Brain Research*, 1597, 220–246.
729 <https://doi.org/10.1016/j.brainres.2014.11.059>
- 730 Lu, H.-C., & Mackie, K. (2016). An Introduction to the Endogenous Cannabinoid System.
731 *Biological Psychiatry*, 79(7), 516–525. <https://doi.org/10.1016/j.biopsych.2015.07.028>
- 732 MacDonald-Wicks, L., McEvoy, M., Magennis, E., Schofield, P. W., Patterson, A. J., & Zacharia,
733 K. (2019). Dietary Long-Chain Fatty Acids and Cognitive Performance in Older Australian
734 Adults. *Nutrients*, 11(4), 711. <https://doi.org/10.3390/nu11040711>
- 735 Macouzet, M., Robert, N., & Lee, B. H. (2010). Genetic and functional aspects of linoleate
736 isomerase in *Lactobacillus acidophilus*. *Applied Microbiology and Biotechnology*, 87,
737 1737–1742. <https://doi.org/10.1007/s00253-010-2634-z>
- 738 Martínez Leo, E. E., Rojas Herrera, R. A., & Segura Campos, M. R. (2019). *Protective Effect of*
739 *Omega 3 Fatty Acids EPA and DHA in the Neurodegenerative Disease BT - Bioactive*
740 *Molecules in Food* (J.-M. Mérillon & K. G. Ramawat (Eds.); pp. 605–621). Springer
741 International Publishing. https://doi.org/10.1007/978-3-319-78030-6_90
- 742 Michels da Silva, D., Langer, H., & Graf, T. (2019). Inflammatory and Molecular Pathways in
743 Heart Failure-Ischemia, HFpEF and Transthyretin Cardiac Amyloidosis. *International*

- 744 *Journal of Molecular Sciences*, 20(9). <https://doi.org/10.3390/ijms20092322>
- 745 Milanski, M., Degasperi, G., Coope, A., Morari, J., Denis, R., Cintra, D. E., Tsukumo, D. M. L.,
746 Anhe, G., Amaral, M. E., Takahashi, H. K., Curi, R., Oliveira, H. C., Carnevali, J. B. C.,
747 Bordin, S., Saad, M. J., & Velloso, L. A. (2009). Saturated Fatty Acids Produce an
748 Inflammatory Response Predominantly through the Activation of TLR4 Signaling in
749 Hypothalamus: Implications for the Pathogenesis of Obesity. *The Journal of Neuroscience*,
750 29(2), 359–370. <https://doi.org/10.1523/JNEUROSCI.2760-08.2009>
- 751 Mišurcová, L., Ambrožová, J., & Samek, D. (2011). Seaweed lipids as nutraceuticals. *Advances*
752 *in Food and Nutrition Research*, 64, 339–355. [https://doi.org/10.1016/B978-0-12-387669-](https://doi.org/10.1016/B978-0-12-387669-0.00027-2)
753 0.00027-2
- 754 Montserrat-de la Paz, S., Naranjo, M. C., Millan-Linares, M. C., Lopez, S., Abia, R., Biessen, E.
755 A. L., Muriana, F. J. G., & Bermudez, B. (2019). Monounsaturated Fatty Acids in a High-
756 Fat Diet and Niacin Protect from White Fat Dysfunction in the Metabolic Syndrome.
757 *Molecular Nutrition and Food Research*, 63(19), 1–17.
758 <https://doi.org/10.1002/mnfr.201900425>
- 759 Moreno-Navarrete, J. M., Catalán, V., Whyte, L., Díaz-Arteaga, A., Vázquez-Martínez, R.,
760 Rotellar, F., Guzmán, R., Gómez-Ambrosi, J., Pulido, M. R., Russell, W. R., Imbernón, M.,
761 Ross, R. A., Malagón, M. M., Dieguez, C., Fernández-Real, J. M., Frühbeck, G., &
762 Nogueiras, R. (2012). The L- α -lysophosphatidylinositol/GPR55 system and its potential
763 role in human obesity. *Diabetes*, 61(2), 281–291. <https://doi.org/10.2337/db11-0649>
- 764 Mukhopadhyay, S., Mondal, S. A., Kumar, M., & Dutta, D. (2014). Proinflammatory and
765 antiinflammatory attributes of fetuin-a: a novel hepatokine modulating cardiovascular and
766 glycemic outcomes in metabolic syndrome. *Endocrine Practice: Official Journal of the*
767 *American College of Endocrinology and the American Association of Clinical*
768 *Endocrinologists*, 20(12), 1345–1351. <https://doi.org/10.4158/EP14421.RA>
- 769 Murru, E., Carta, G., Manca, C., Sogos, V., Pistis, M., Melis, M., & Banni, S. (2021). Conjugated
770 Linoleic Acid and Brain Metabolism: A Possible Anti-Neuroinflammatory Role Mediated
771 by PPAR α Activation. *Frontiers in Pharmacology*, 11(January), 1–12.
772 <https://doi.org/10.3389/fphar.2020.587140>
- 773 Nadjar, A., Leyrolle, Q., Joffre, C., & Laye, S. (2016). Bioactive lipids as new class of microglial
774 modulators: When nutrition meets neuroimmunology. *Progress in*
775 *Neuropsychopharmacology & Biological Psychiatry*, 79(Pt A), 19–26.
776 <https://doi.org/10.1016/j.pnpbp.2016.07.004>
- 777 Nagy, K., & Tiuca, I.-D. (2017a). Importance of Fatty Acids in Physiopathology of Human Body.
778 *IntechOpen*. <https://doi.org/10.5772/67407>
- 779 Nagy, K., & Tiuca, I.-D. (2017b). Importance of Fatty Acids in Physiopathology of Human Body.
780 *IntechOpen*. <https://doi.org/10.5772/67407>
- 781 Nam, K. N., Mounier, A., Wolfe, C. M., Fitz, N. F., Carter, A. Y., Castranio, E. L., Kamboh, H.
782 I., Reeves, V. L., Wang, J., Han, X., Schug, J., Lefterov, I., & Koldamova, R. (2017). Effect
783 of high fat diet on phenotype, brain transcriptome and lipidome in Alzheimer’s model mice.
784 *Scientific Reports*, 7(1), 1–13. <https://doi.org/10.1038/s41598-017-04412-2>
- 785 Narumiya, S., & Furuyashiki, T. (2011). Fever, inflammation, pain and beyond: prostanoid
786 receptor research during these 25 years. *FASEB Journal: Official Publication of the*
787 *Federation of American Societies for Experimental Biology*, 25(3), 813–818.
788 <https://doi.org/10.1096/fj.11-0302ufm>
- 789 Natarajan, C., & Bright, J. J. (2002). Peroxisome proliferator-activated receptor-gamma agonists
790 inhibit experimental allergic encephalomyelitis by blocking IL-12 production, IL-12

- 791 signaling and Th1 differentiation. *Genes and Immunity*, 3(2), 59–70.
792 <https://doi.org/10.1038/sj.gene.6363832>
- 793 Nathan, C. (2002). Points of control in inflammation. *Nature*, 420(6917), 846–852.
794 <https://doi.org/10.1038/nature01320>
- 795 Nathan, C., & Ding, A. (2010). Nonresolving inflammation. *Cell*, 140(6), 871–882.
796 <https://doi.org/10.1016/j.cell.2010.02.029>
- 797 Nikitopoulou, I., Oikonomou, N., Karouzakis, E., Sevastou, I., Nikolaidou-Katsaridou, N., Zhao,
798 Z., Mersinias, V., Armaka, M., Xu, Y., Masu, M., Mills, G. B., Gay, S., Kollias, G., &
799 Aidinis, V. (2012). Autotaxin expression from synovial fibroblasts is essential for the
800 pathogenesis of modeled arthritis. *The Journal of Experimental Medicine*, 209(5), 925–933.
801 <https://doi.org/10.1084/jem.20112012>
- 802 Nisr, R. B., Shah, D. S., Ganley, I. G., & Hundal, H. S. (2019). Proinflammatory NFkB signalling
803 promotes mitochondrial dysfunction in skeletal muscle in response to cellular fuel
804 overloading. *Cellular and Molecular Life Sciences*, 76(24), 4887–4904.
805 <https://doi.org/10.1007/s00018-019-03148-8>
- 806 Nogueras, L., Gonzalo, H., Jové, M., Sol, J., Gil-Sanchez, A., Hervás, J. V., Valcheva, P.,
807 Gonzalez-Mingot, C., Solana, M. J., Peralta, S., Pamplona, R., & Brieva, L. (2019). Lipid
808 profile of cerebrospinal fluid in multiple sclerosis patients: a potential tool for diagnosis.
809 *Scientific Reports*, 9(1), 11313. <https://doi.org/10.1038/s41598-019-47906-x>
- 810 Nolan, J. M., Mulcahy, R., Power, R., Moran, R., & Howard, A. N. (2018). Nutritional
811 Intervention to Prevent Alzheimer’s Disease: Potential Benefits of Xanthophyll Carotenoids
812 and Omega-3 Fatty Acids Combined. *Journal of Alzheimer’s Disease*, 64, 367–378.
813 <https://doi.org/10.3233/JAD-180160>
- 814 Notario-Barandiaran, L., Valera-Gran, D., Gonzalez-Palacios, S., Garcia-de-la-Hera, M.,
815 Fernández-Barrés, S., Pereda-Pereda, E., Fernández-Somoano, A., Guxens, M., Iñiguez, C.,
816 Romaguera, D., Vrijheid, M., Tardón, A., Santa-Marina, L., Vioque, J., Navarrete-Muñoz,
817 E. M., & Project, on behalf of the I. (2020). High adherence to a mediterranean diet at age
818 4 reduces overweight, obesity and abdominal obesity incidence in children at the age of 8.
819 *International Journal of Obesity*, 44(9), 1906–1917. [https://doi.org/10.1038/s41366-020-](https://doi.org/10.1038/s41366-020-0557-z)
820 [0557-z](https://doi.org/10.1038/s41366-020-0557-z)
- 821 O’Neill, L. A. J. (2006). How Toll-like receptors signal: what we know and what we don’t know.
822 *Current Opinion in Immunology*, 18(1), 3–9.
823 <https://doi.org/https://doi.org/10.1016/j.coi.2005.11.012>
- 824 Oh, D. Y., Talukdar, S., Bae, E. J., Imamura, T., Morinaga, H., Fan, W. Q., Li, P., Lu, W. J.,
825 Watkins, S. M., & Olefsky, J. M. (2010). GPR120 Is an Omega-3 Fatty Acid Receptor
826 Mediating Potent Anti-inflammatory and Insulin-Sensitizing Effects. *Cell*, 142(5), 687–698.
827 <https://doi.org/10.1016/j.cell.2010.07.041>
- 828 OLIVEIRA, E. M. L. de, MONTANI, D. A., OLIVEIRA-SILVA, D., RODRIGUES-OLIVEIRA,
829 A. F., MATAS, S. L. de A., FERNANDES, G. B. P., SILVA, I. D. C. G. da, & LO TURCO,
830 E. G. (2019). Multiple sclerosis has a distinct lipid signature in plasma and cerebrospinal
831 fluid. *Arquivos de Neuro-Psiquiatria*, 77, 696–704.
832 [http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0004-](http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0004-282X2019001000696&nrm=iso)
833 [282X2019001000696&nrm=iso](http://www.scielo.br/scielo.php?script=sci_arttext&pid=S0004-282X2019001000696&nrm=iso)
- 834 Ono, H. (2019). Molecular Mechanisms of Hypothalamic Insulin Resistance. *International*
835 *Journal of Molecular Sciences*, 20(6), 1317. <https://doi.org/10.3390/ijms20061317>
- 836 Orsavova, J., Misurcova, L., Vavra Ambrozova, J., Vicha, R., & Mlcek, J. (2015). Fatty acids
837 composition of vegetable oils and its contribution to dietary energy intake and dependence

- 838 of cardiovascular mortality on dietary intake of fatty acids. *International Journal of*
839 *Molecular Sciences*, 16(6), 12871–12890. <https://doi.org/10.3390/ijms160612871>
- 840 Ouyang, W. C., Sun, G. C., & Hsu, M. C. (2020). Omega-3 fatty acids in cause, prevention and
841 management of violence in schizophrenia: Conceptualization and application. *Aggression*
842 *and Violent Behavior*, 50(8), 101347. <https://doi.org/10.1016/j.avb.2019.101347>
- 843 Pahwa, R., Goyal, A., Bansal, P., & Jialal, I. (2021). *Chronic Inflammation*. StatPearls.
844 <https://www.ncbi.nlm.nih.gov/books/NBK493173/#>
- 845 Pal, D., Dasgupta, S., Kundu, R., Maitra, S., Das, G., Mukhopadhyay, S., Ray, S., Majumdar, S.
846 S., & Bhattacharya, S. (2012). Fetuin-A acts as an endogenous ligand of TLR4 to promote
847 lipid-induced insulin resistance. *Nature Medicine*, 18, 1279.
848 <https://doi.org/10.1038/nm.2851>
- 849 Patterson, E., Wall, R., Fitzgerald, G. F., Ross, R. P., & Stanton, C. (2012). Health implications
850 of high dietary omega-6 polyunsaturated Fatty acids. *Journal of Nutrition and Metabolism*,
851 2012, 539426. <https://doi.org/10.1155/2012/539426>
- 852 Pimentel, G. D., Dornellas, A. P. S., Rosa, J. C., Lira, F. S., Cunha, C. A., Boldarine, V. T., Souza,
853 G. I. H. De, Hirata, A. E., Nascimento, C. M. O., Oyama, L. M., Watanabe, R. L. H., &
854 Ribeiro, E. B. (2012). High-fat diets rich in soy or fish oil distinctly alter hypothalamic
855 insulin signaling in rats. *The Journal of Nutritional Biochemistry*, 23(7), 822–828.
856 <https://doi.org/10.1016/j.jnutbio.2011.04.006>
- 857 Piñeiro, R., & Falasca, M. (2012). Lysophosphatidylinositol signalling: new wine from an old
858 bottle. *Biochimica et Biophysica Acta*, 1821(4), 694–705.
859 <https://doi.org/10.1016/j.bbali.2012.01.009>
- 860 Pompura, S. L., Wagner, A., Kitz, A., LaPerche, J., Yosef, N., Dominguez-Villar, M., & Hafler,
861 D. A. (2021). Oleic acid restores suppressive defects in tissue-resident FOXP3 Tregs from
862 patients with multiple sclerosis. *The Journal of Clinical Investigation*, 131(2).
863 <https://doi.org/10.1172/JCI138519>
- 864 Proust, F., Lucas, M., & Dewailly, É. (2014). Fatty acid profiles among the Inuit of Nunavik:
865 Current status and temporal change. *Prostaglandins Leukotrienes and Essential Fatty Acids*,
866 90(5), 159–167. <https://doi.org/10.1016/j.plefa.2014.02.001>
- 867 Račková, L., Ergin, V., Burcu Bali, E., Kuniaková, M., & Karasu, Ç. (2014). Pomegranate Seed
868 Oil Modulates Functions and Survival of BV-2 Microglial Cells in vitro. *International*
869 *Journal for Vitamin and Nutrition Research. Internationale Zeitschrift Fur Vitamin- Und*
870 *Ernahrungsforschung. Journal International de Vitaminologie et de Nutrition*, 84(5–6),
871 295–309. <https://doi.org/10.1024/0300-9831/a000216>
- 872 Rádiková, Ž., Penesová, A., Vlček, M., Havranová, A., Siváková, M., Šiarnik, P., Žitňanová, I.,
873 Imrich, R., Turčáni, P., & Kollár, B. (2020). Lipoprotein profiling in early multiple sclerosis
874 patients: effect of chronic inflammation? *Lipids in Health and Disease*, 19(1), 49.
875 <https://doi.org/10.1186/s12944-020-01221-x>
- 876 Ramirez-Ramirez, V., Macias-Islas, M. A., Ortiz, G. G., Pacheco-Moises, F., Torres-Sanchez, E.
877 D., Sorto-Gomez, T. E., Cruz-Ramos, J. A., Orozco-Aviña, G., & Celis de la Rosa, A. J.
878 (2013). Efficacy of fish oil on serum of TNF α , IL-1 β , and IL-6 oxidative stress markers
879 in multiple sclerosis treated with interferon beta-1b. *Oxidative Medicine and Cellular*
880 *Longevity*, 2013, 709493. <https://doi.org/10.1155/2013/709493>
- 881 Ravaut, G., Légiot, A., Bergeron, K. F., & Mounier, C. (2021). Monounsaturated fatty acids in
882 obesity-related inflammation. *International Journal of Molecular Sciences*, 22(1), 1–22.
883 <https://doi.org/10.3390/ijms22010330>
- 884 Rezapour-Firouzi, S. (2017). Herbal Oil Supplement with Hot-Nature Diet for Multiple Sclerosis.

- 885 In *Nutrition and Lifestyle in Neurological Autoimmune Diseases: Multiple Sclerosis*.
 886 Elsevier Inc. <https://doi.org/10.1016/B978-0-12-805298-3.00024-4>
- 887 Rog, J., Błażewicz, A., Juchnowicz, D., Ludwiczuk, A., Stelmach, E., Koziół, M., Karakula, M.,
 888 Niziński, P., & Karakula-Juchnowicz, H. (2020). The Role of GPR120 Receptor in Essential
 889 Fatty Acids Metabolism in Schizophrenia. In *Biomedicines* (Vol. 8, Issue 8).
 890 <https://doi.org/10.3390/biomedicines8080243>
- 891 Rorato, R., Borges, B. D. C., & Uchoa, E. T. (2017). LPS-Induced Low-Grade Inflammation
 892 Increases Hypothalamic JNK Expression and Causes Central Insulin Resistance Irrespective
 893 of Body Weight Changes. *International Journal of Molecular Sciences*, *18*(1431), 1–14.
 894 <https://doi.org/10.3390/ijms18071431>
- 895 Ruiz-Núñez, B., Dijck-Brouwer, D. A. J., & Muskiet, F. A. J. (2016a). The relation of saturated
 896 fatty acids with low-grade inflammation and cardiovascular disease. *The Journal of*
 897 *Nutritional Biochemistry*, *36*, 1–20.
 898 <https://doi.org/https://doi.org/10.1016/j.jnutbio.2015.12.007>
- 899 Ruiz-Núñez, B., Dijck-Brouwer, D. A. J., & Muskiet, F. A. J. (2016b). The relation of saturated
 900 fatty acids with low-grade inflammation and cardiovascular disease. *The Journal of*
 901 *Nutritional Biochemistry*, *36*, 1–20.
 902 <https://doi.org/https://doi.org/10.1016/j.jnutbio.2015.12.007>
- 903 Ryan, K. K., Li, B., Grayson, B. E., Matter, E. K., Woods, S. C., & Seeley, R. J. (2011). A role
 904 for central nervous system PPAR- γ in the regulation of energy balance. *Nature Medicine*,
 905 *17*(5), 623–627. <https://doi.org/10.1038/nm.2349>
- 906 Salsinha, A. S., Rodríguez-Alcalá, L. M., Relvas, J. B., & Pintado, M. E. (2021). Fatty acids role
 907 on obesity induced hypothalamus inflammation: From problem to solution – A review.
 908 *Trends in Food Science & Technology*, *112*(September 2020), 592–607.
 909 <https://doi.org/10.1016/j.tifs.2021.03.042>
- 910 Satogami, K., Takahashi, S., Yamada, S., Ukai, S., & Shinosaki, K. (2017). Omega-3 fatty acids
 911 related to cognitive impairment in patients with schizophrenia. *Schizophrenia Research:*
 912 *Cognition*, *9*, 8–12. <https://doi.org/https://doi.org/10.1016/j.scog.2017.05.001>
- 913 Schilling, J. D., Machkovech, H. M., He, L., Sidhu, R., Fujiwara, H., Weber, K., Ory, D. S., &
 914 Schaffer, J. E. (2013). Palmitate and lipopolysaccharide trigger synergistic ceramide
 915 production in primary macrophages. *The Journal of Biological Chemistry*, *288*(5), 2923–
 916 2932. <https://doi.org/10.1074/jbc.M112.419978>
- 917 Serhan, C. N. (2014). Pro-resolving lipid mediators are leads for resolution physiology. *Nature*,
 918 *510*(7503), 92–101. <https://doi.org/10.1038/nature13479>
- 919 Sevastou, I., Kaffe, E., Mouratis, M.-A., & Aidinis, V. (2013). Lysoglycerophospholipids in
 920 chronic inflammatory disorders: The PLA2/LPC and ATX/LPA axes. *Biochimica et*
 921 *Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, *1831*(1), 42–60.
 922 <https://doi.org/https://doi.org/10.1016/j.bbalip.2012.07.019>
- 923 Snowden, S. G., Ebshiana, A. A., Hye, A., An, Y., Pletnikova, O., O'Brien, R., Troncoso, J.,
 924 Legido-Quigley, C., & Thambisetty, M. (2017). Association between fatty acid metabolism
 925 in the brain and Alzheimer disease neuropathology and cognitive performance: A
 926 nontargeted metabolomic study. *PLoS Medicine*, *14*(3), e1002266–e1002266.
 927 <https://doi.org/10.1371/journal.pmed.1002266>
- 928 Solberg, D. K., Refsum, H., Andreassen, O. A., & Bentsen, H. (2019). A five-year follow-up
 929 study of antioxidants, oxidative stress and polyunsaturated fatty acids in schizophrenia. *Acta*
 930 *Neuropsychiatrica*, *31*(4), 202–212. <https://doi.org/DOI: 10.1017/neu.2019.14>
- 931 Takayoshi, S., Kanami, T.-K., Junko, N., Michiko, I., Xunmei, Y., Shinji, M., Hidehito, K., Shoji,

- 932 Y., Kensuke, M., Seiichiro, A., Yasutomi, K., & Yoshihiro, O. (2007). Role of the Toll-like
933 Receptor 4/NF- κ B Pathway in Saturated Fatty Acid-Induced Inflammatory Changes in the
934 Interaction Between Adipocytes and Macrophages. *Arteriosclerosis, Thrombosis, and*
935 *Vascular Biology*, 27(1), 84–91. <https://doi.org/10.1161/01.ATV.0000251608.09329.9a>
- 936 Talukdar, S., Olefsky, J. M., & Osborn, O. (2011). Targeting GPR120 and other fatty acid sensing
937 GPCRs ameliorates insulin resistance and inflammatory diseases. *Trends Pharmacol Sci*,
938 32(9), 543–550. <https://doi.org/10.1016/j.tips.2011.04.004>.
- 939 Tang, W., Wang, Y., Xu, F., Fan, W., Zhang, Y., Fan, K., Wang, W., Zhang, Y., & Zhang, C.
940 (2020). Omega-3 fatty acids ameliorate cognitive dysfunction in schizophrenia patients with
941 metabolic syndrome. *Brain, Behavior, and Immunity*, 88, 529–534.
942 <https://doi.org/https://doi.org/10.1016/j.bbi.2020.04.034>
- 943 Thaler, J. P., Yi, C., Schur, E. A., Guyenet, S. J., Hwang, B. H., Dietrich, M. O., Zhao, X., Sarruf,
944 D. A., Izgur, V., Maravilla, K. R., Nguyen, H. T., Fischer, J. D., Matsen, M. E., Wisse, B.
945 E., Morton, G. J., Horvath, T. L., Baskin, D. G., Tschöp, M. H., & Schwartz, M. W. (2012).
946 Obesity is associated with hypothalamic injury in rodents and humans. *The Journal of*
947 *Clinical Investigation*, 122(1), 153–162. <https://doi.org/10.1172/JCI59660.adjacent>
- 948 Tontonoz, P, Hu, E., & Spiegelman, B. M. (1994). Stimulation of adipogenesis in fibroblasts by
949 PPAR gamma 2, a lipid-activated transcription factor. *Cell*, 79(7), 1147–1156.
950 [https://doi.org/10.1016/0092-8674\(94\)90006-x](https://doi.org/10.1016/0092-8674(94)90006-x)
- 951 Tontonoz, Peter, Hu, E., Graves, R. A., Budavari, A. I., & Spiegelman, B. M. (1994). mPPAR
952 gamma 2: tissue-specific regulator of an adipocyte enhancer. *Genes & Development*, 4,
953 1224–1234. <https://doi.org/10.1101/gad.8.10.1224>
- 954 Toscano, R., Millan-Linares, M. C., Lemus-Conejo, A., Claro, C., Sanchez-Margalet, V., &
955 Montserrat-de la Paz, S. (2020). Postprandial triglyceride-rich lipoproteins promote M1/M2
956 microglia polarization in a fatty-acid-dependent manner. *Journal of Nutritional*
957 *Biochemistry*, 75, 108248. <https://doi.org/10.1016/j.jnutbio.2019.108248>
- 958 Tse, E. K., & Belsham, D. D. (2018). Palmitate induces neuroinflammation, ER stress, and Pomc
959 mRNA expression in hypothalamic mHypA-POMC/GFP neurons through novel
960 mechanisms that are prevented by oleate. *Molecular and Cellular Endocrinology*, 472, 40–
961 49. <https://doi.org/https://doi.org/10.1016/j.mce.2017.11.017>
- 962 Valdearcos, M., Robblee, M. M., Xu, A. W., Koliwad, S. K., Valdearcos, M., Robblee, M. M.,
963 Benjamin, D. I., Nomura, D. K., & Xu, A. W. (2014). Microglia Dictate the Impact of
964 Saturated Fat Consumption on Hypothalamic Inflammation and Neuronal Function Article
965 Microglia Dictate the Impact of Saturated Fat Consumption on Hypothalamic Inflammation
966 and Neuronal Function. *CellReports*, 9(6), 2124–2138.
967 <https://doi.org/10.1016/j.celrep.2014.11.018>
- 968 Viggiano, E., Mollica, M. P., Lionetti, L., Cavaliere, G., Trinchese, G., De Filippo, C., Chieffi,
969 S., Gaita, M., Barletta, A., De Luca, B., Crispino, M., & Monda, M. (2016). Effects of an
970 High-Fat Diet Enriched in Lard or in Fish Oil on the Hypothalamic Amp-Activated Protein
971 Kinase and Inflammatory Mediators. *Frontiers in Cellular Neuroscience*, 10, 150.
972 <https://doi.org/10.3389/fncel.2016.00150>
- 973 Wang, Y.-M., Nagao, K., Ujino, Y., Sakata, K., Higa, K., Inoue, N., & Yanagita, T. (2005). Short-
974 term feeding of conjugated linoleic acid does not induce hepatic steatosis in C57BL/6J mice.
975 *Journal of Nutritional Science and Vitaminology*, 51(6), 440–444.
976 <https://doi.org/10.3177/jnsv.51.440>
- 977 Wang, Y. L., Frauwirth, K. A., Rangwala, S. M., Lazar, M. A., & Thompson, C. B. (2002).
978 Thiazolidinedione activation of peroxisome proliferator-activated receptor gamma can
979 enhance mitochondrial potential and promote cell survival. *The Journal of Biological*

- 980 *Chemistry*, 277(35), 31781–31788. <https://doi.org/10.1074/jbc.M204279200>
- 981 Wang, Z., Liu, D., Wang, F., Liu, S., Zhao, S., Ling, E. A., & Hao, A. (2012). Saturated fatty
982 acids activate microglia via Toll-like receptor 4/NF- κ B signalling. *British Journal of*
983 *Nutrition*, 107(2), 229–241. <https://doi.org/10.1017/S0007114511002868>
- 984 Watari, M., Hamazaki, K., Hirata, T., Hamazaki, T., & Okubo, Y. (2010). Hostility of drug-free
985 patients with schizophrenia and n-3 polyunsaturated fatty acid levels in red blood cells.
986 *Psychiatry Research*, 177(1–2), 22–26. <https://doi.org/10.1016/j.psychres.2010.02.016>
- 987 Whittington, R. A., Planel, E., & Terrando, N. (2017). Impaired Resolution of Inflammation in
988 Alzheimer’s Disease: A Review. *Frontiers in Immunology*, 8, 1464.
989 <https://doi.org/10.3389/fimmu.2017.01464>
- 990 Wong, S. W., Kwon, M.-J., Choi, A. M. K., Kim, H.-P., Nakahira, K., & Hwang, D. H. (2009).
991 Fatty acids modulate Toll-like receptor 4 activation through regulation of receptor
992 dimerization and recruitment into lipid rafts in a reactive oxygen species-dependent manner.
993 *The Journal of Biological Chemistry*, 284(40), 27384–27392.
994 <https://doi.org/10.1074/jbc.M109.044065>
- 995 Yan, L., Xie, Y., Satyanarayanan, S. K., Zeng, H., Liu, Q., Huang, M., Ma, Y., Wan, J.-B., Yao,
996 X., Su, K.-P., & Su, H. (2020). Omega-3 polyunsaturated fatty acids promote brain-to-blood
997 clearance of β -Amyloid in a mouse model with Alzheimer’s disease. *Brain, Behavior, and*
998 *Immunity*, 85, 35–45. <https://doi.org/https://doi.org/10.1016/j.bbi.2019.05.033>
- 999 Yanagita, T., Wang, Y.-M., Nagao, K., Ujino, Y., & Inoue, N. (2005). Conjugated linoleic acid-
1000 induced fatty liver can be attenuated by combination with docosahexaenoic acid in
1001 C57BL/6N mice. *Journal of Agricultural and Food Chemistry*, 53(24), 9629–9633.
1002 <https://doi.org/10.1021/jf052203i>
- 1003 Yang, X., Sun, L., Zhao, A., Hu, X., Qing, Y., Jiang, J., Yang, C., Xu, T., Wang, P., Liu, J., Zhang,
1004 J., He, L., Jia, W., & Wan, C. (2017). Serum fatty acid patterns in patients with
1005 schizophrenia: a targeted metabolomics study. *Translational Psychiatry*, 7(7), e1176–
1006 e1176. <https://doi.org/10.1038/tp.2017.152>
- 1007 Yang, Z., Pryor, M., Noguchi, A., Sampson, M., Johnson, B., Pryor, M., Donkor, K., Amar, M.,
1008 Remaley, A. T., Section, M., Branch, C., Heart, N., & Core, P. (2020). *HHS Public Access*.
1009 63(12), 1–22. <https://doi.org/10.1002/mnfr.201900120>. Dietary
- 1010 Yao, C., Sakata, D., Esaki, Y., Li, Y., Matsuoka, T., Kuroiwa, K., Sugimoto, Y., & Narumiya, S.
1011 (2009). Prostaglandin E2-EP4 signaling promotes immune inflammation through Th1 cell
1012 differentiation and Th17 cell expansion. *Nature Medicine*, 15(6), 633–640.
1013 <https://doi.org/10.1038/nm.1968>
- 1014 Yi, C.-X., Walter, M., Gao, Y., Pitra, S., Legutko, B., Kálin, S., Layritz, C., García-Cáceres, C.,
1015 Bielohuby, M., Bidlingmaier, M., Woods, S. C., Ghanem, A., Conzelmann, K.-K., Stern, J.
1016 E., Jastroch, M., & Tschöp, M. H. (2017). TNF α drives mitochondrial stress in POMC
1017 neurons in obesity. *Nature Communications*, 8(1), 15143.
1018 <https://doi.org/10.1038/ncomms15143>
- 1019 Zakaria, N. (2014). 4 - Body shape analysis and identification of key dimensions for apparel sizing
1020 systems. In D. Gupta & N. B. T.-A. Zakaria Apparel Sizing and Design (Eds.), *Woodhead*
1021 *Publishing Series in Textiles* (pp. 95–119). Woodhead Publishing.
1022 <https://doi.org/https://doi.org/10.1533/9780857096890.1.95>
- 1023 Zhang, Xiaoqing, Zhang, G., Zhang, H., Karin, M., Bai, H., Cai, D., & Dongsheng, C. (2008).
1024 Hypothalamic IKK β /NF- κ B and ER Stress Link Overnutrition to Energy Imbalance and
1025 Obesity. *Cell*, 135(1), 61–73. <https://doi.org/10.1016/j.cell.2008.07.043>. Hypothalamic
- 1026 Zhang, Xufei, Wu, X., Hu, Q., Wu, J., Wang, G., Hong, Z., & Ren, J. (2019). Mitochondrial DNA

1027 in liver inflammation and oxidative stress. *Life Sciences*, 236, 116464.
1028 <https://doi.org/10.1016/j.lfs.2019.05.020>
1029
1030

Table 1 – Summary of some relevant studies available since 2017 regarding fatty acids’ effects on Alzheimer’s disease (AD).

Fatty acids studied	Study’s objectives	Experimental model	Main results	Reference
Dietary short-, medium-, and long-chain fatty acids.	Determine the associations between AD risk and dietary fatty acid intakes.	Multiethnic, longitudinal, population study from the Washington Heights-Hamilton Heights-Inwood Columbia Aging Project (WHICAP). Dietary intake was measured using a food frequency questionnaire.	Higher intake of DHA and EPA are protective for AD.	(Gustafson et al., 2020)
Long-chain fatty acids particularly omega-6 fatty acids.	Evaluate whether a range of dietary long-chain fatty acids are associated with cognitive performance in older men and women.	Data from the Hunter Community Study (HCS) (older men and women of Newcastle Australia); comparing dietary data from a validated Food Frequency Questionnaire with validated cognitive performance measures.	A higher intake of total omega-6, but not other types of fatty acids, was associated with better cognitive performance.	(MacDonald-Wicks et al., 2019)
Fish oil (rich in omega-3 polyunsaturated fatty acids).	Determined the dynamic alterations of blood-brain barrier transport function in the early disease stage of AD.	APPswe/PS1dE9 C57BL/6J (APP/PS1) transgenic mice	Supplementation with fish oil significantly enhanced the expression level of LRP-1 - a main efflux transporter of blood-brain barrier -, promoted Amyloid- β ($A\beta$) plaques clearance from the brain to circulation, inhibited NF κ B activation, reduced the expression of interleukin-1 β and tumor necrosis factor- α and suppressed the glial activation in mice.	(Yan et al., 2020)
Unsaturated fatty acids: linoleic acid, linolenic acid, DHA, EPA, oleic acid and arachidonic acid.	Metabolic profiling of brain tissue samples from AD individuals (57 to 95 y old) to assess how the abundance of omega-3 and omega-6 fatty acids species is affected by differing levels of disease pathology.	Autopsy sample of the Baltimore Longitudinal Study of Aging (BLSA) forming three groups: AD, controls and “asymptomatic Alzheimer’s disease”, <i>i.e.</i> , individuals with significant AD neuropathology at death but without evidence for cognitive impairment during life.	Unsaturated fatty acid metabolism is significantly dysregulated in the brains of patients with varying degrees of Alzheimer pathology.	(Snowden et al., 2017)
Fish oil supplementation	Determine the impact of supplemental xanthophyll carotenoids plus omega-3 fatty acids on disease progression in patients with AD.	Three trials: supplementation of AD patients with xanthophyll carotenoids; combined supplementation of xanthophyll and fish oil; subjects free of AD supplemented with xanthophyll.	Preliminary results suggest positive outcomes for patients with AD who consumed a combination of xanthophyll carotenoids plus fish oil.	(Nolan et al., 2018)
Omega-3 or olive oil supplementation	Determine the effect of omega-3 PUFA supplementation on behavioral performances and hippocampal neurogenesis, volume, and astrogliosis.	Aged mice subjected to a selective depletion of basal forebrain cholinergic neurons- valuable model to mimic one of the most reliable hallmarks of early AD neuropathology.	Omega-3 polyunsaturated fatty acids can counteract behavioral deficits and hippocampal neurodegeneration in the experimental model.	(Cutuli et al., 2020)

Table 2 - Summary of some relevant studies available since 2017 regarding fatty acids' effects on Multiple Sclerosis (MS).

Study's objectives	Experimental model	Main results	Reference
Evaluate the lipid profile characteristic of MS present in cerebrospinal fluid and plasma.	Preliminary untargeted qualitative lipidomics analysis comparing cerebrospinal fluid and plasma samples from patients with MS, other inflammatory neurological diseases and idiopathic intracranial hypertension.	Cerebrospinal fluid and plasma from patients with MS present a unique lipid signature that can be useful as a diagnostic biomarker.	(OLIVEIRA et al., 2019)
Determine the association between lipoprotein subfractions and inflammatory status in early stages of MS.	Lipoprotein profile analysis in 19 newly diagnosed MS patients	Male MS patients might be at higher risk of atherosclerosis development. Presence of subtle dyslipidemia in early stages of the disease. Observation of mutual links between systemic inflammation and lipid metabolism in early MS with low inflammatory activity.	(Rádiková et al., 2020)
Investigate the role of environmental lipids in shaping the tissue resident Treg (regulatory T cells) phenotype.	<i>Ex vivo</i> computational analyses and <i>in vitro</i> experimental assays with human Tregs isolated from peripheral blood and adipose tissue.	Oleic acid concentrations were reduced in patients with MS. Exposure of MS Tregs to oleic acid restored defects in their suppressive function.	(Pompura et al., 2021)
Investigate the association between dietary intake of polyunsaturated fatty acids and MS risk.	Assessment of 80,920 women from Nurses' Health Study (1984–2004) and 94,511 women from Nurses' Health Study II (1991–2009) dietary intake from a validated food frequency questionnaire every 4 years. Identification of 479 incident MS cases during follow-up.	Higher intake of total polyunsaturated fatty acids at baseline was associated with a lower risk of MS. Low dietary polyunsaturated fatty acids intake may be another modifiable risk factor for MS.	(Bjørnevik et al., 2017)
Analyze possible changes in the lipidome (namely fatty acids) of MS patients.	Lipidomic approach to analyze the cerebrospinal fluid of MS patients diagnosed with relapsing-remitting MS.	Unique cerebrospinal fluid lipidomic signature in MS patients at the time of diagnosis that might be considered as a potential diagnostic tool.	(Nogueras et al., 2019)
Optic neuritis is an acute inflammatory demyelinating disorder of the optic nerve and is an initial symptom of MS. Address a possible role of fatty acids diet supplementation in counteracting optical neuritis, namely retinal ganglion cell degeneration.	Experimental autoimmune encephalomyelitis mouse model of MS, a well-established model for optic neuritis.	Dietary supplementation with Neuro-FAG (nFAG®), a balanced mixture of fatty acids (FAs), counteracted inflammatory and gliotic processes in the retina.	(Dal Monte et al., 2018)

Table 3 - Summary of some relevant studies available since 2017 regarding fatty acids' effects on Schizophrenia.

Study's objectives	Experimental Model	Main results	Reference
Scan the free fatty acids pattern and elucidate the characteristics of lipid metabolic abnormality in schizophrenia patients.	110 patients with schizophrenia and 109 healthy controls.	Monounsaturated fatty acids and omega-6 polyunsaturated fatty acids were significantly increased in schizophrenia. Desaturation from saturated fatty acids to monounsaturated fatty acids and β -oxidation were enhanced. These results suggest that lipolysis and β -oxidation are upregulated in this disease, presumably resulting from insufficient brain energy supply.	(X. Yang et al., 2017)
Determine if omega-3 fatty acids may be of value in enhancing BDNF levels and improving cognitive function in patients with schizophrenia and metabolic syndrome.	12-week randomized placebo-controlled trial: 80 patients with both schizophrenia and Metabolic syndrome.	Omega-3 treatment improves cognitive function in schizophrenia patients with metabolic syndrome. Omega-3 enhanced BDNF levels after 12 weeks of treatment through suppressing TNF- α levels.	(Tang et al., 2020)
Investigate if the levels of oxidative markers are increased in schizophrenia compared with healthy controls and if they are associated with the levels of membrane polyunsaturated fatty acids and symptom severity.	51 control patients and 55 patients with schizophrenia spectrum disorders, assessed during an acute phase and 5 years later during a stable phase.	Redox regulation is dynamic and changes during different phases of the disorder. Abnormal levels of antioxidants (α -tocopherol) in the stable phase indicate persisting redox dysregulation. Changes in α -tocopherol were associated with polyunsaturated fatty acids levels in the acute phase.	(Solberg et al., 2019)
Examine the relationship between omega-3 fatty acids with cognitive function, social function, and psychiatric symptoms in patients with schizophrenia.	30 patients with schizophrenia or schizoaffective disorder. Psychiatric symptoms, cognitive function, and social function were assessed using the Positive and Negative Syndrome Scale, the Brief Assessment of Cognition in Schizophrenia (BACS), and the Social Functioning Scale (SFS), respectively.	Reduced blood omega-3 fatty acids are associated with cognitive impairment, which then impacts social functioning outcomes in schizophrenia.	(Satogami et al., 2017)
Determine the differences in polyunsaturated fatty acids nutritional status and metabolism between patients with schizophrenia and healthy individuals.	80 participants (40 in each group).	Different metabolisms of fatty acids in schizophrenia. The diminished anti-inflammatory response could be a component connecting GPR120 insensitivity with schizophrenia.	(Rog et al., 2020)

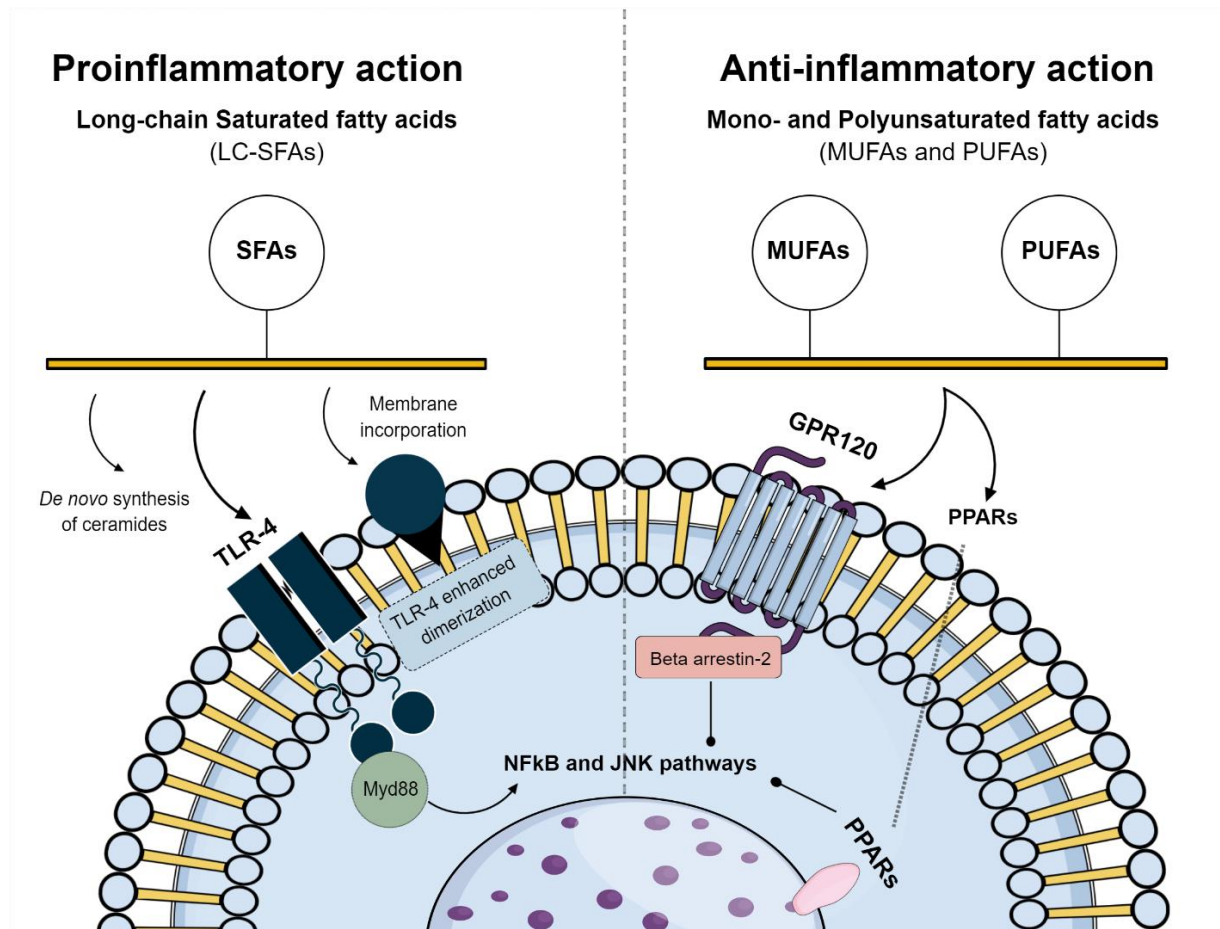


Figure 1 - Pro- and anti-inflammatory action of different fatty acids: Long-chain saturated fatty acids present a proinflammatory potential mediated by TLR-4 and Myd88 activation, ultimately leading to the activation of NFκB and JNK pathways. These saturated fatty acids can be incorporated in specific lipid rafts domains of the plasma membrane where they enhance TLR-4 dimerization, important for its activation. Besides, they are known to be involved in the *de novo* synthesis of ceramides. On the other hand, monounsaturated and polyunsaturated fatty acids through GPR120 and/or PPARs (like PPAR-γ) can repress the activation of NFκB and JNK by saturated fatty acids. Created with Mind the Graph (mindthegraph.com).

