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Global Biobank Meta-analysis Initiative: powering genetic discovery across human disease

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China Kadoorie Biobank collaborative group

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PI: Ben Neale, Claire Churchhouse

Overview: “Methodological extensions to estimate genetic heritability and shared risk factors for phenotypes of the UK Biobank”.

Website for Pan-UKBB results can be found: <https://pan.ukbb.broadinstitute.org/>

Other

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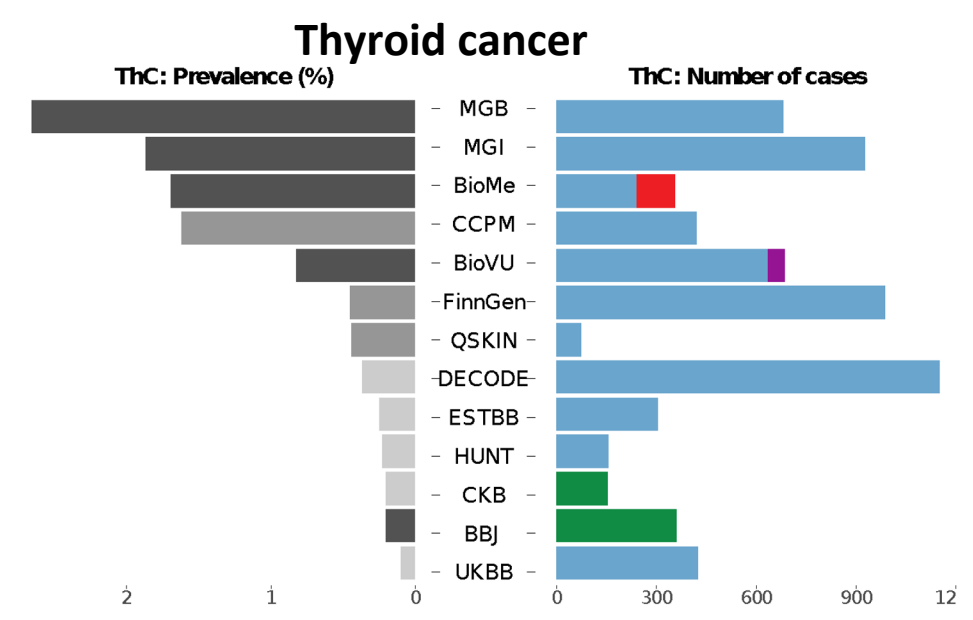
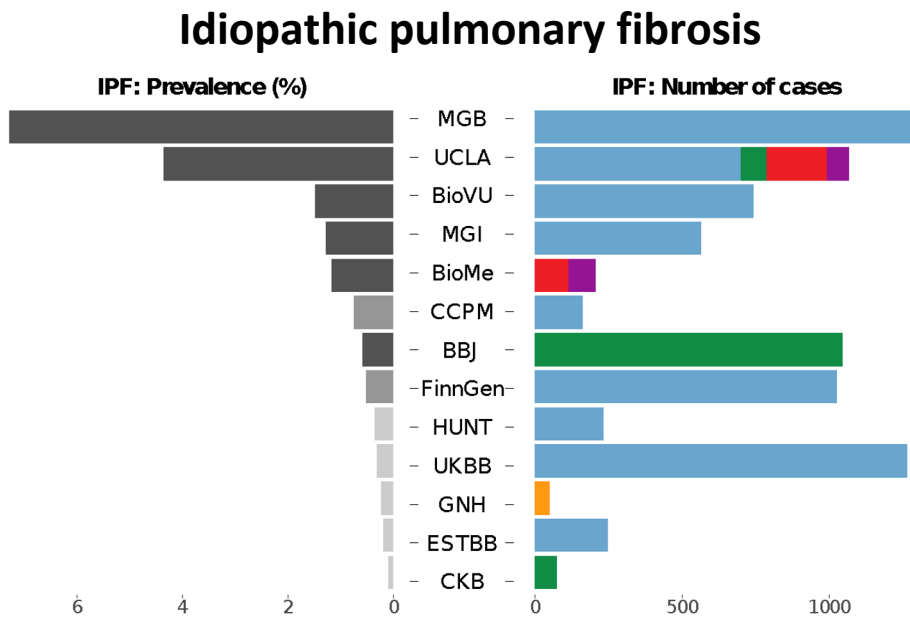
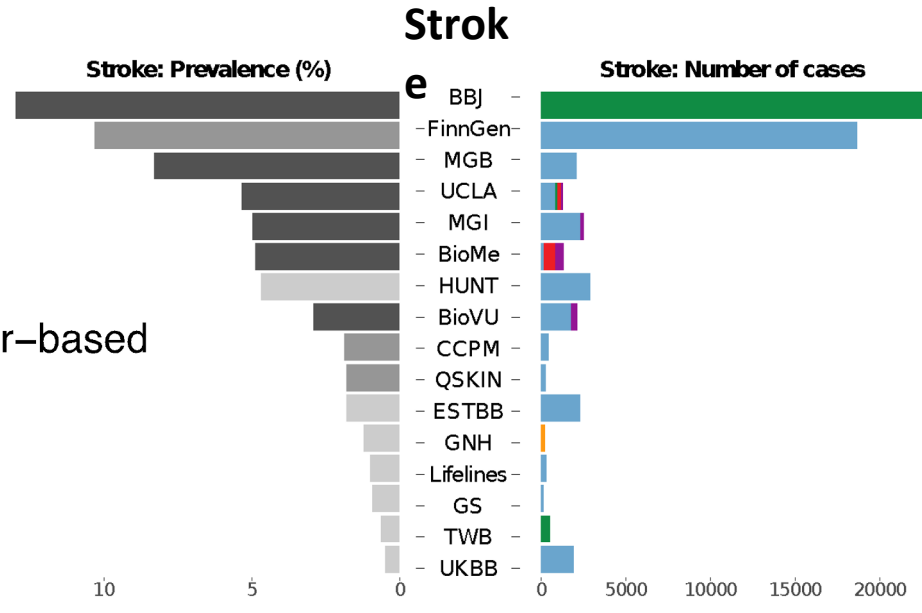
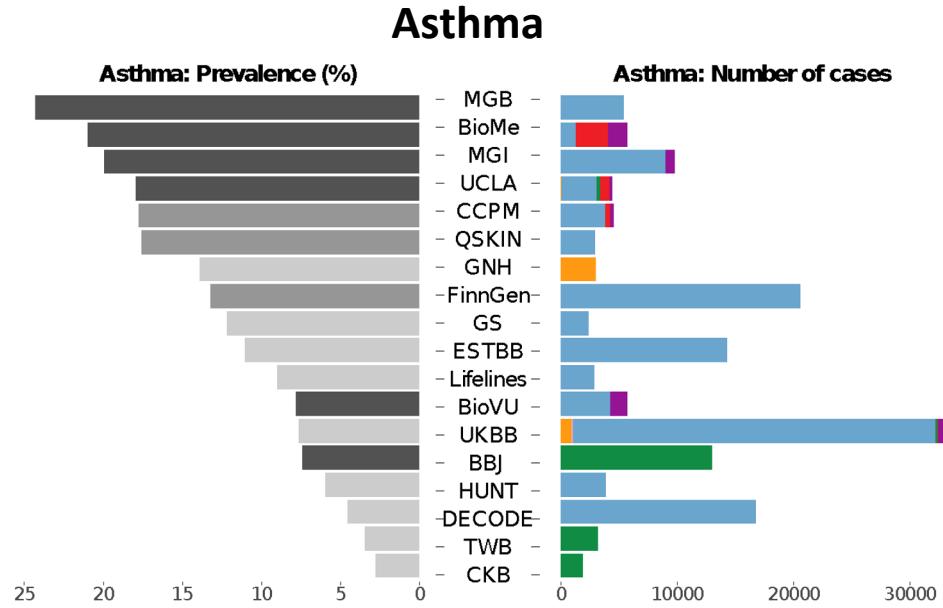
Website for ICDA can be found here: <https://www.icda.bio/>

The Hail Team and Data Management at the Stanley Center for Psychiatric Research

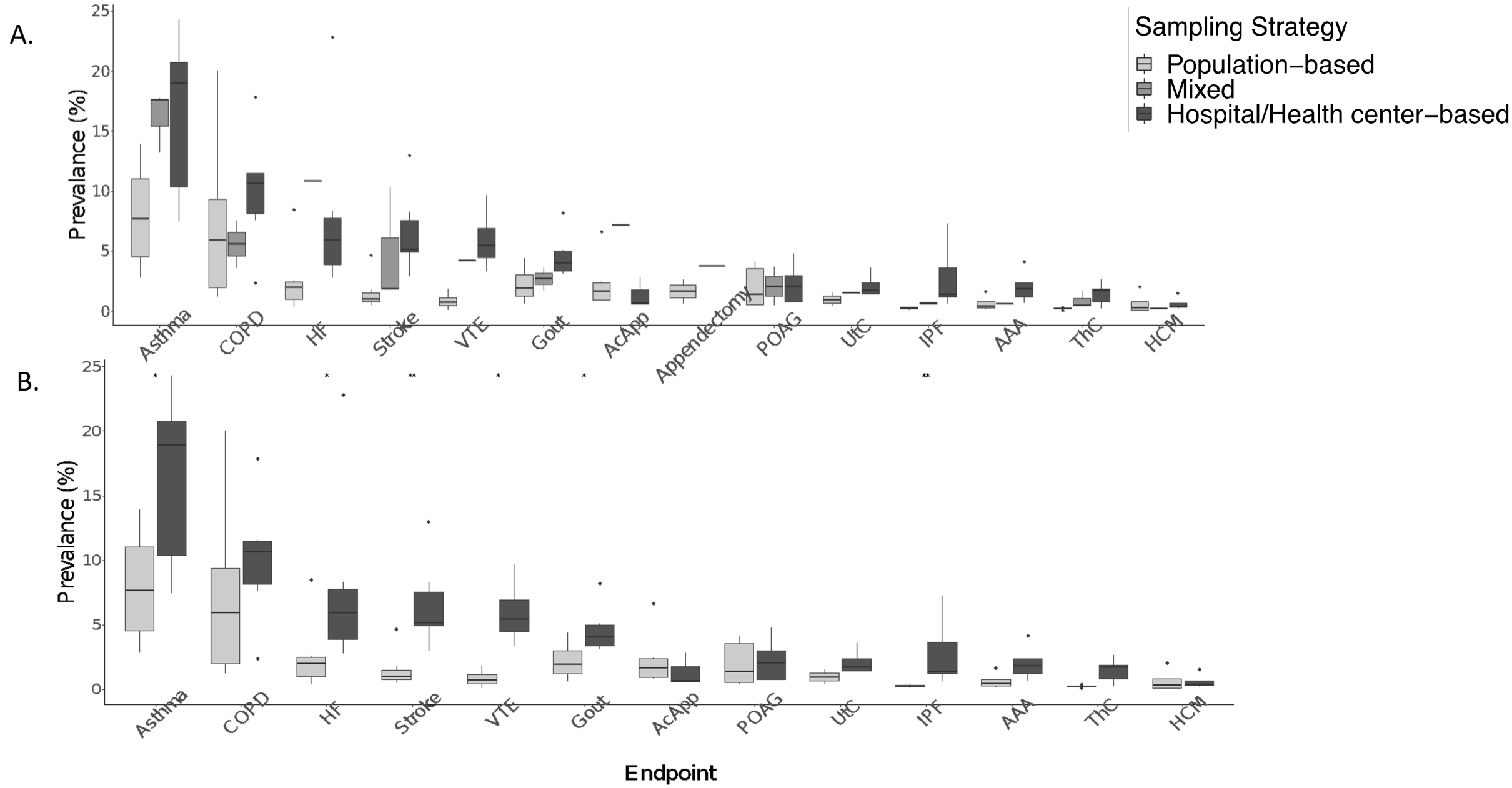
Hail is an open-source Python library that simplifies genomic data analysis in the cloud. It provides powerful, easy-to-use data science tools that can be used to interrogate biobank-scale genomic data and was used in the analysis of the data for this paper. We would especially like to thank Daniel King from the Hail team and Sam Bryant from the Stanley Center Data Management team for helping with the Google bucket set up and data sharing.

Website for Hail can be found here: <https://hail.is/>

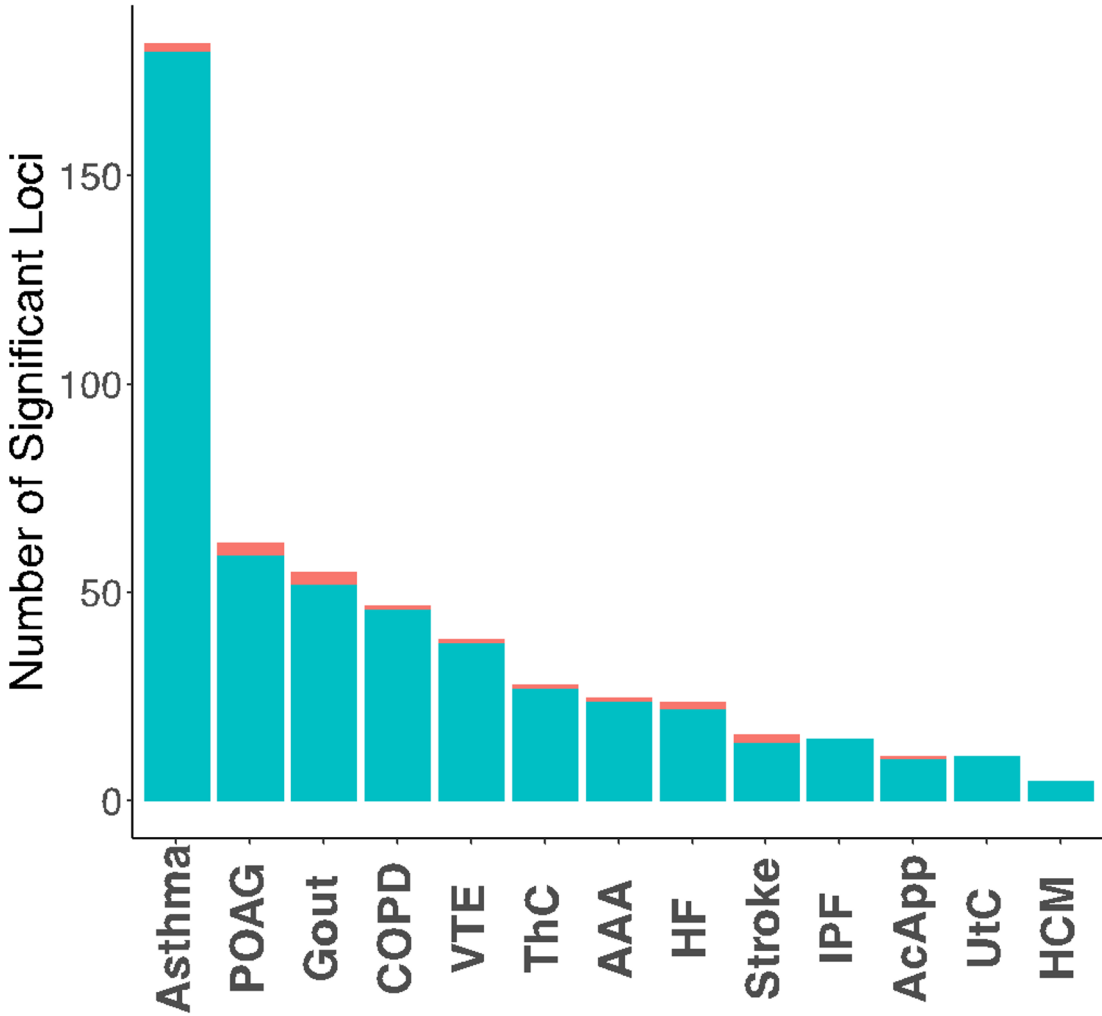
Supplementary Figure 1. Disease prevalence varies across biobanks



Supplementary Figure 2. Disease prevalence varies by difference sample recruiting strategies. A. Box plots for prevalence by three sampling strategies. B. Box plots to compare prevalence between population-based and hospital/health center-based biobanks. (**, $P < 0.01$, *, $P < 0.05$, unpaired Wilcoxon test)

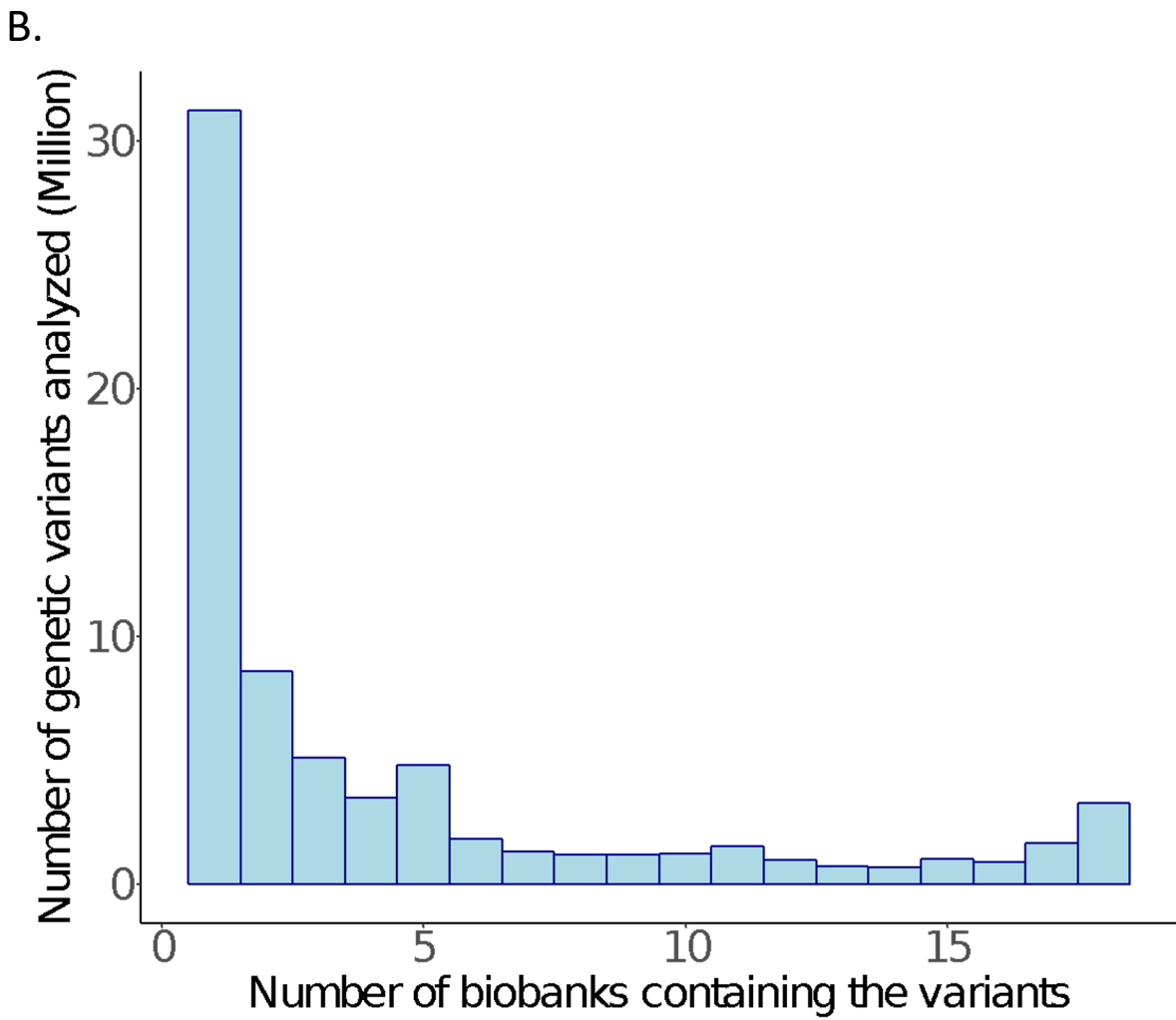
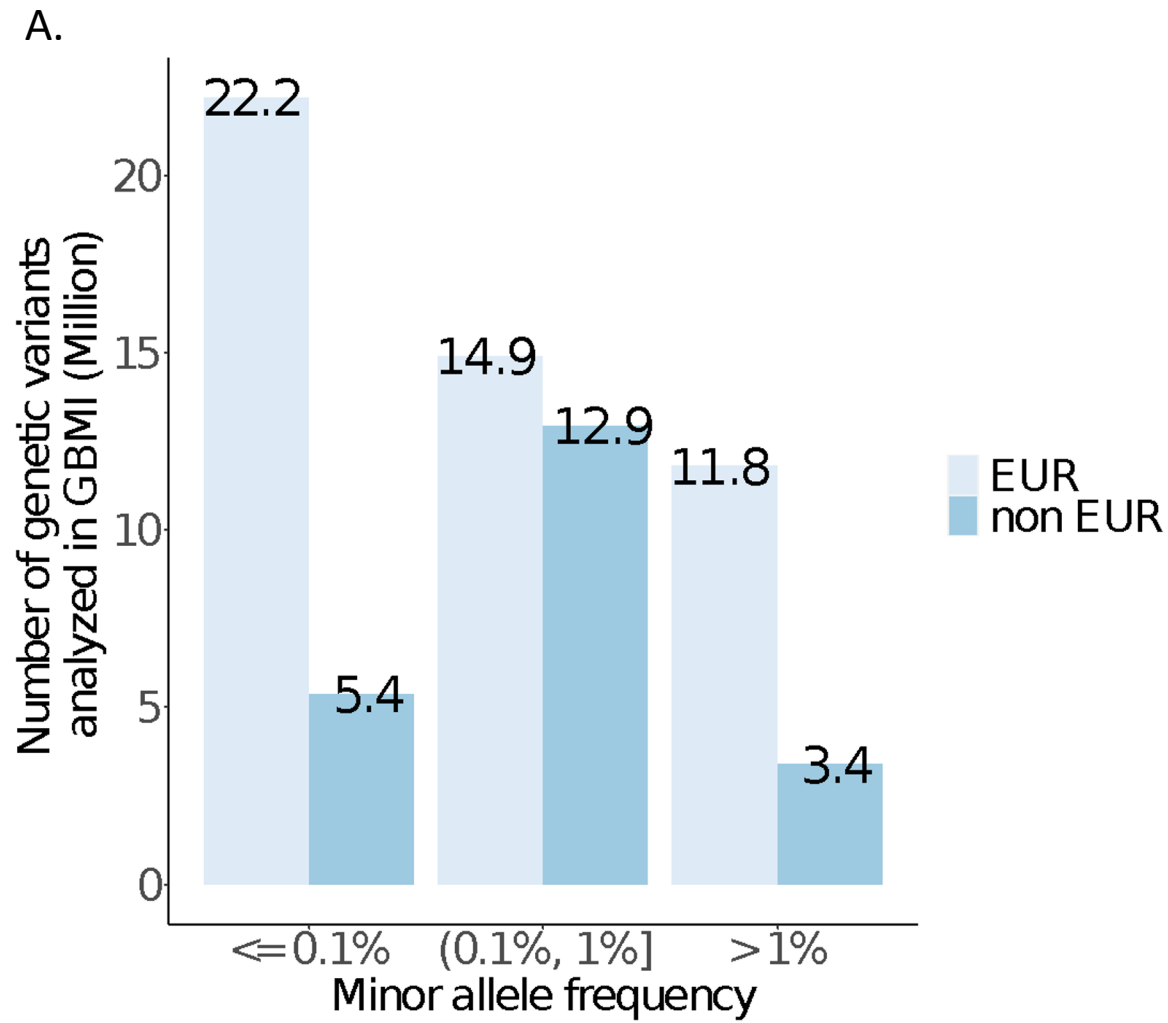


Supplementary Figure 3. Additional significant loci identified by the meta-regression approach implemented in MR-MEGA³⁰ to account for effect size heterogeneity across different data sets in meta-analyses compared to the fixed-effect meta-analyses



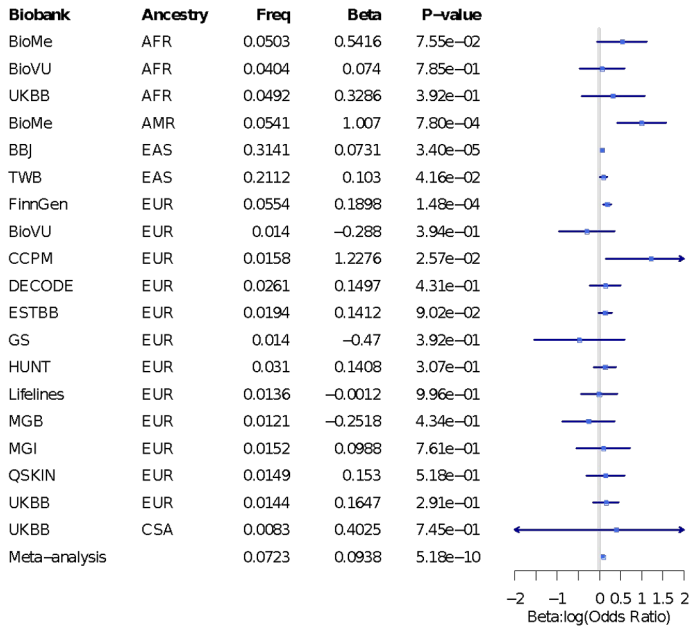
■ Additional loci by accounting for heterogeneity of variant effects in different ancestries
■ Loci identified by fixed-effect meta-analysis (assuming same variant effects in all ancestries)

Supplementary Figure 4. A. Additional genetic variants analyzed due to incorporating non-European samples. EUR: genetic variants observed in samples with European ancestry. non EUR: genetic variants only observed in samples with non-European ancestry. The highest minor allele frequency (MAF) among non EUR ancestry was used in the plot. **B.** Distribution of the number of biobanks in which the genetic variants were tested

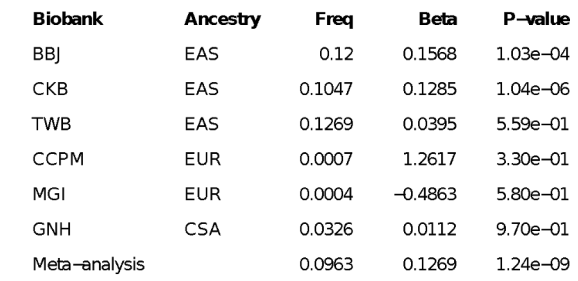


Supplementary Figure 5B. Additional significant loci identified when non-European samples were included in the meta-analysis

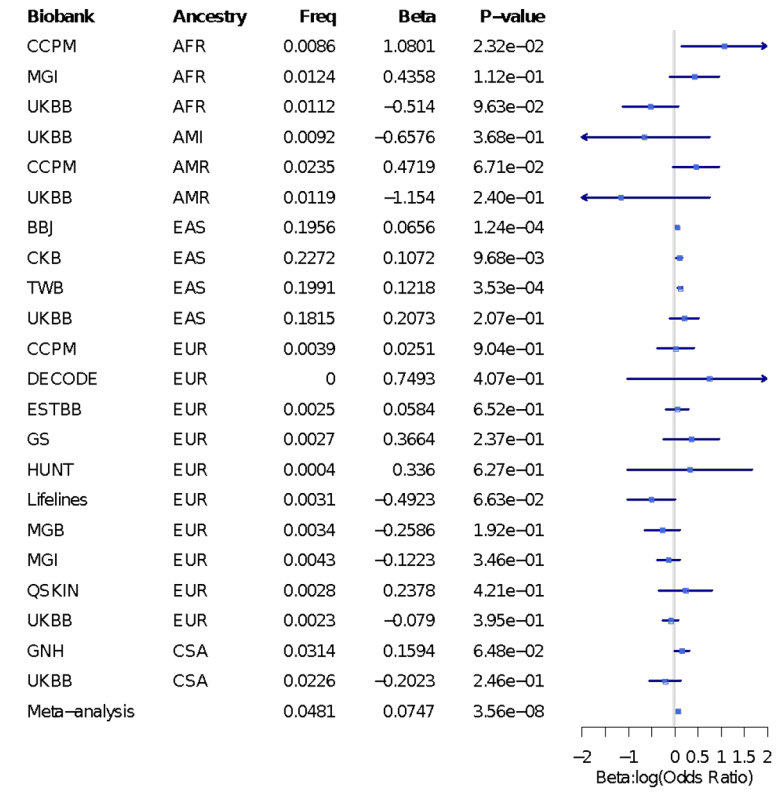
POAG
rs1355927 (intergenic)
MIR2054;INTU



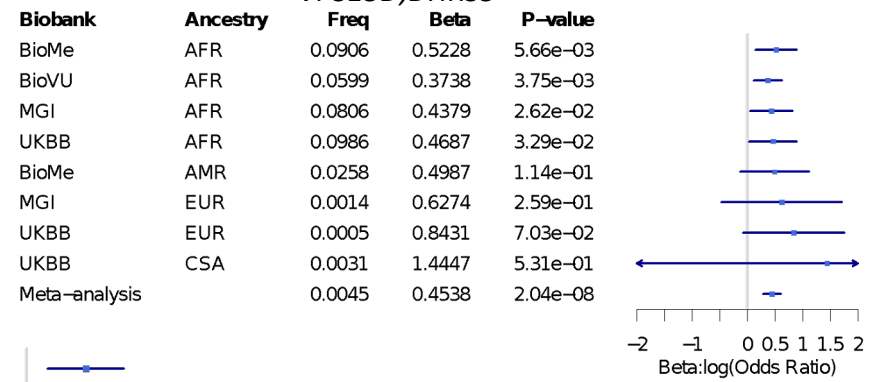
COPD
rs75550771 (intergenic)
PNPT1;EFEMP1



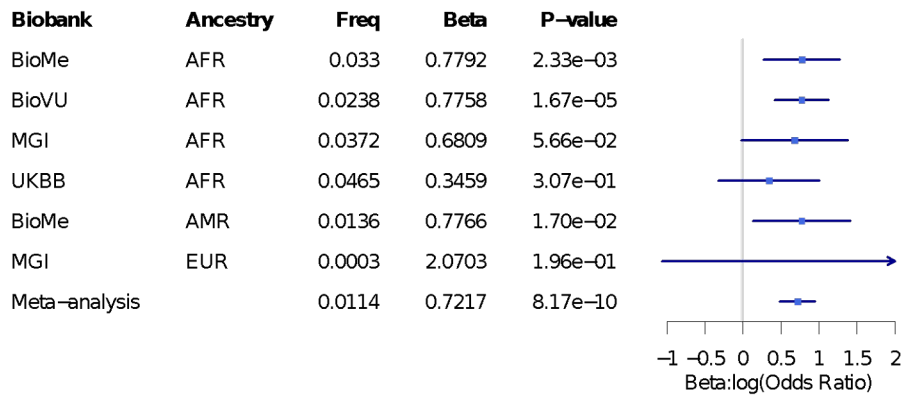
Asthma
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NAA38



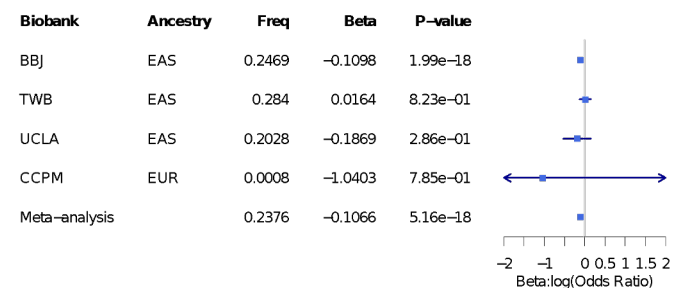
VTE
rs112106699 (intergenic)
VPS13D;DHRS3



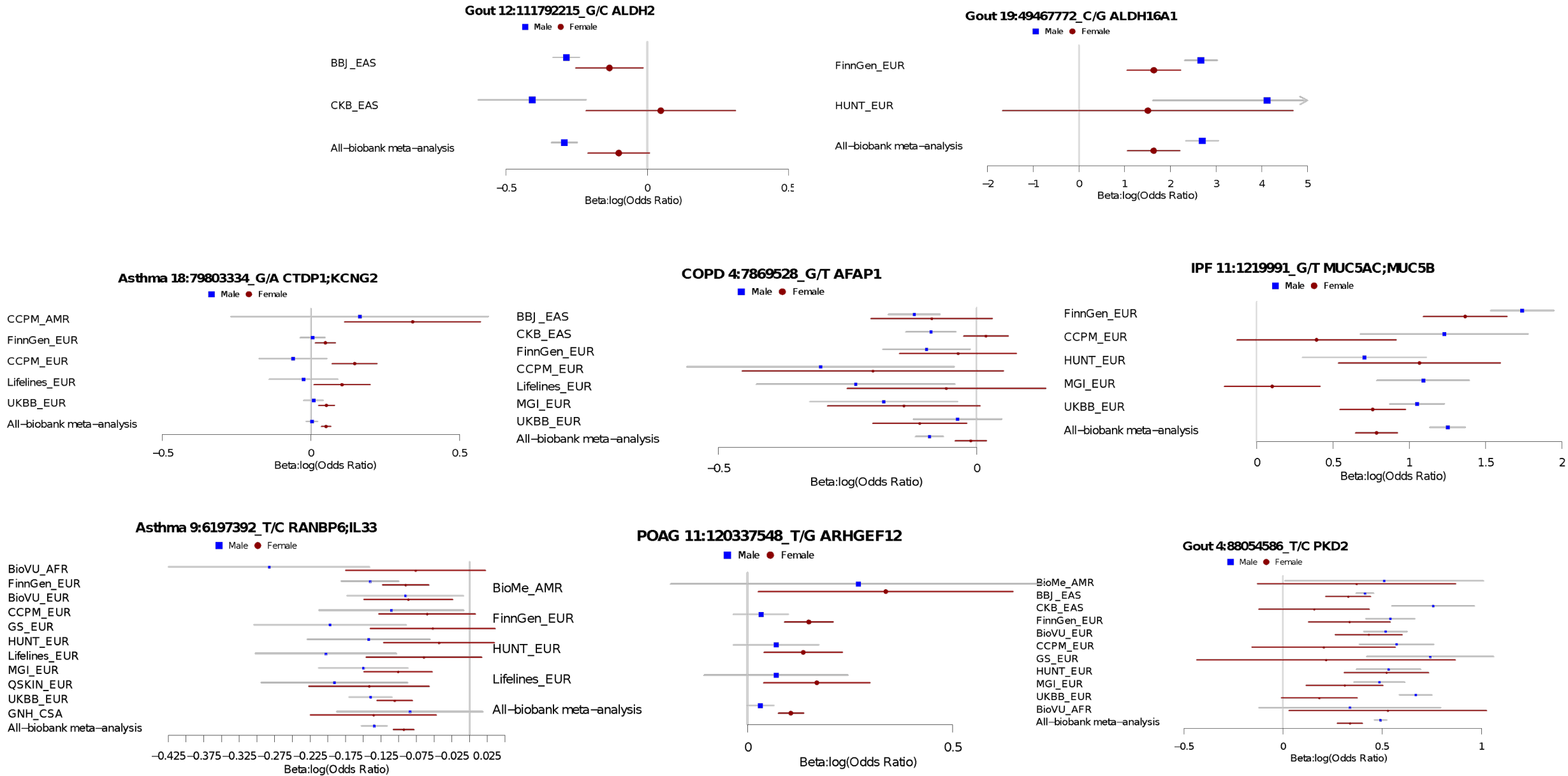
Heart failure
rs145478347 (intronic)
BCL2L12



Stroke
12:111803962 (nonsynonymous)
ALDH2

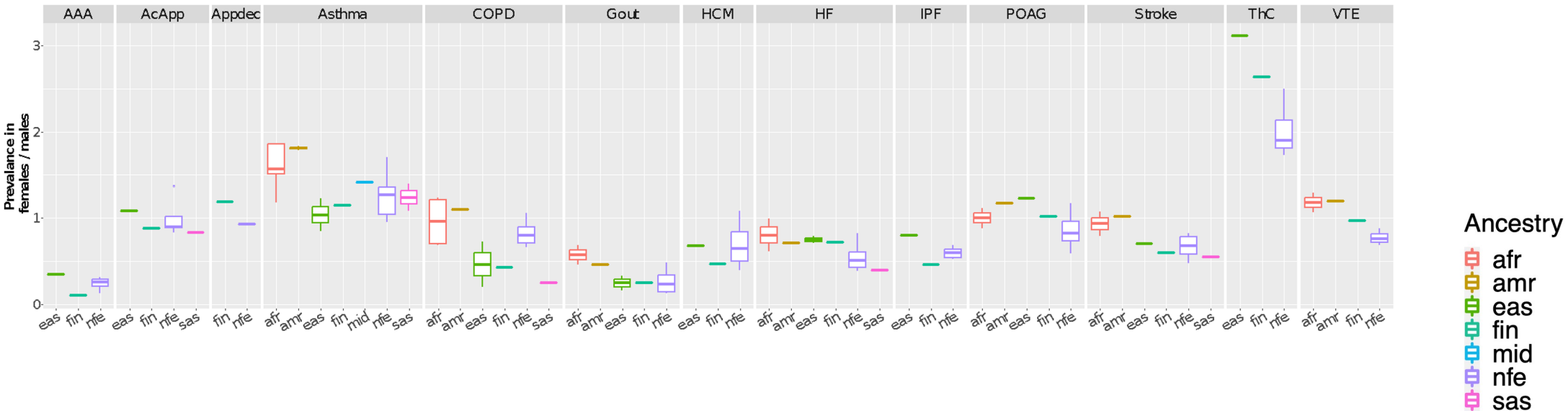


Supplementary Figure 6. LocusZoom plots of region showing differential association between sexes. All-biobank meta-analysis results stratified by sex were filtered to identify regions with different effect sizes in men and women (Phet for Cochran's Q test < 0.002).

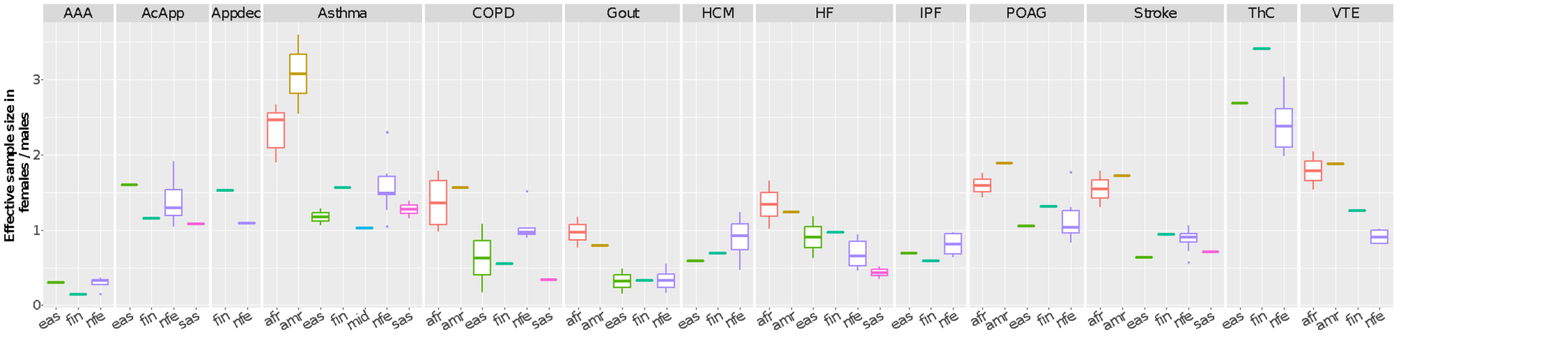


Supplementary Figure 7. Plots of A. prevalence ratios and B. effective sample sizes ratios in females and in males by ancestry across endpoints. The prevalence is calculated as number of cases / (number of cases + number of controls). The effective sample size is calculated as $4 / (1 / \text{number of cases} + 1 / \text{number of controls})$.

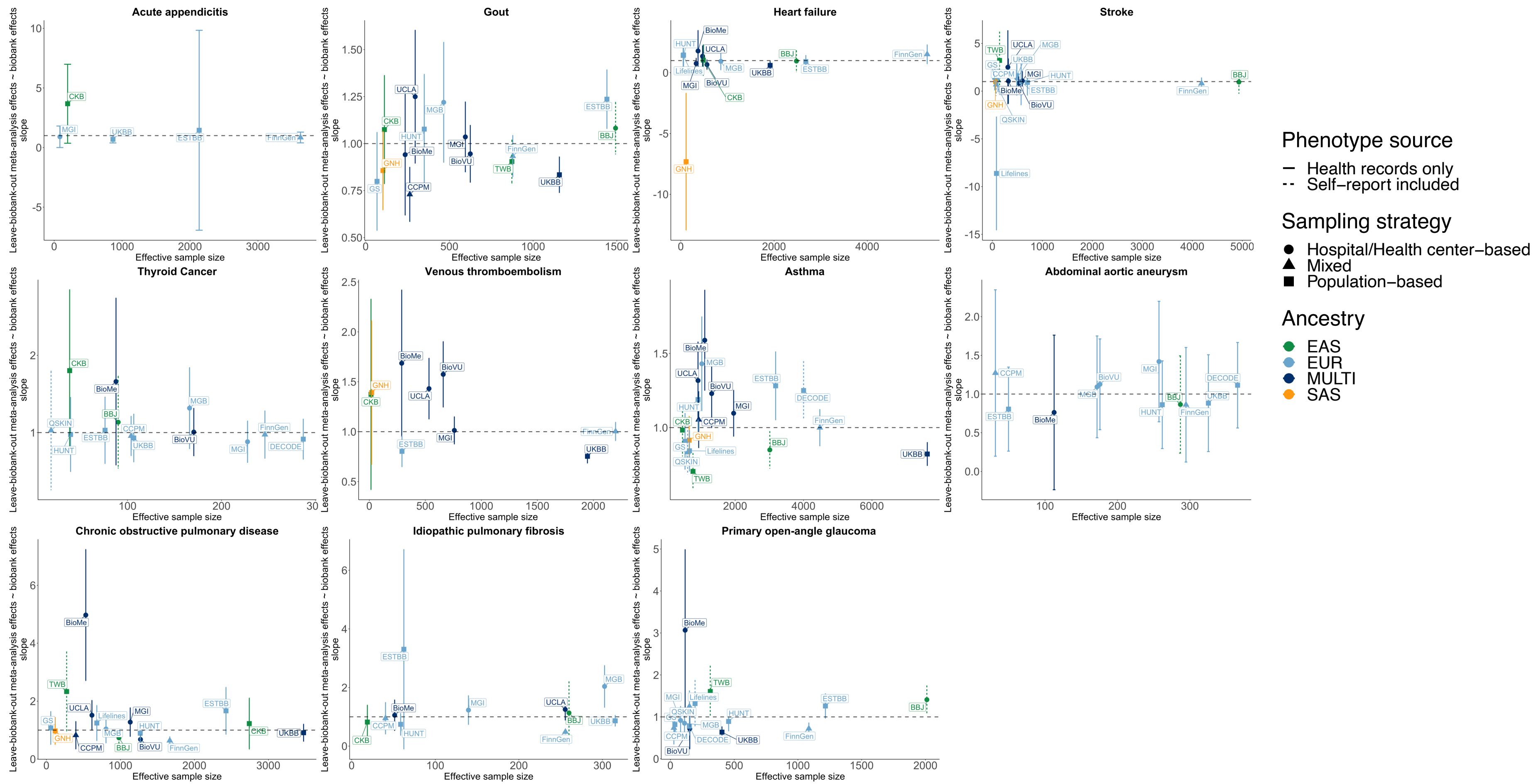
A.



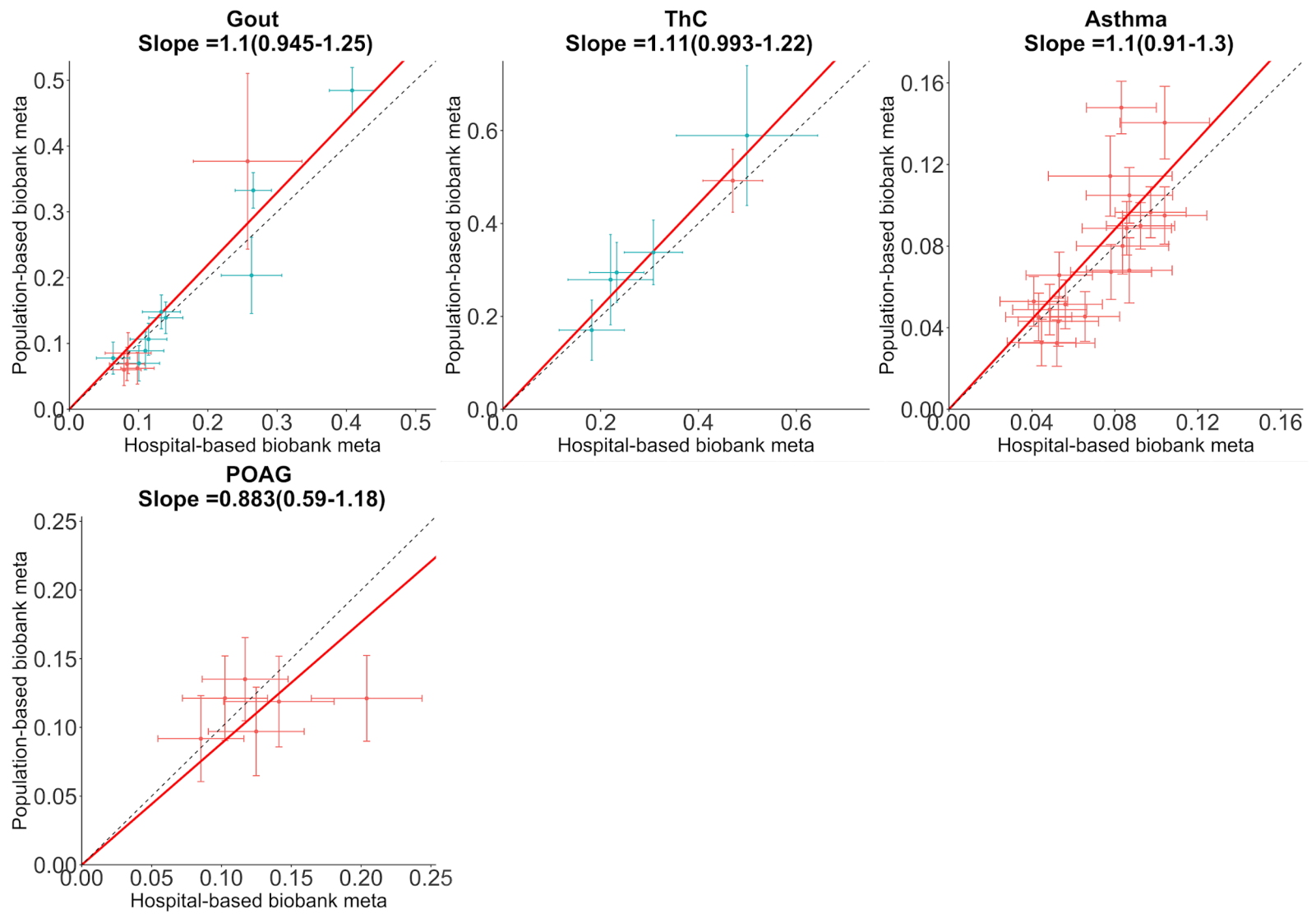
B.



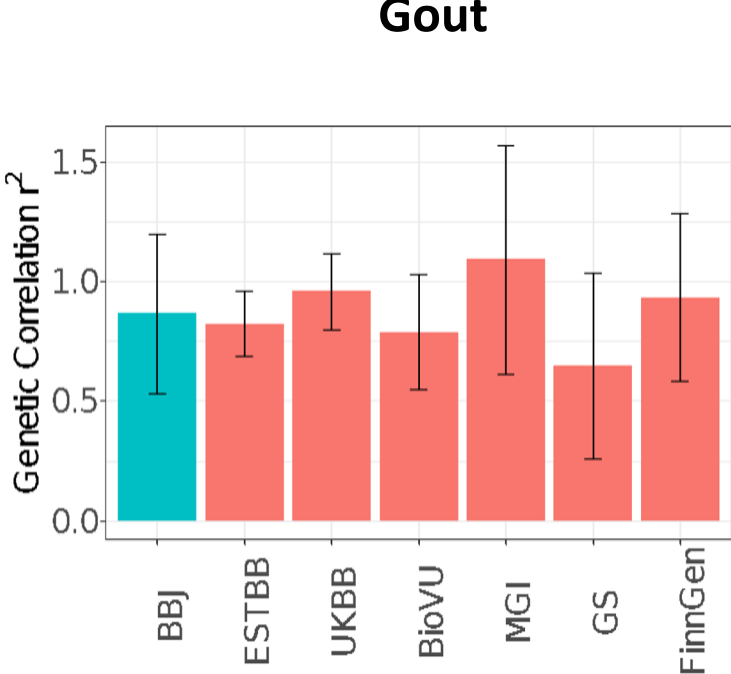
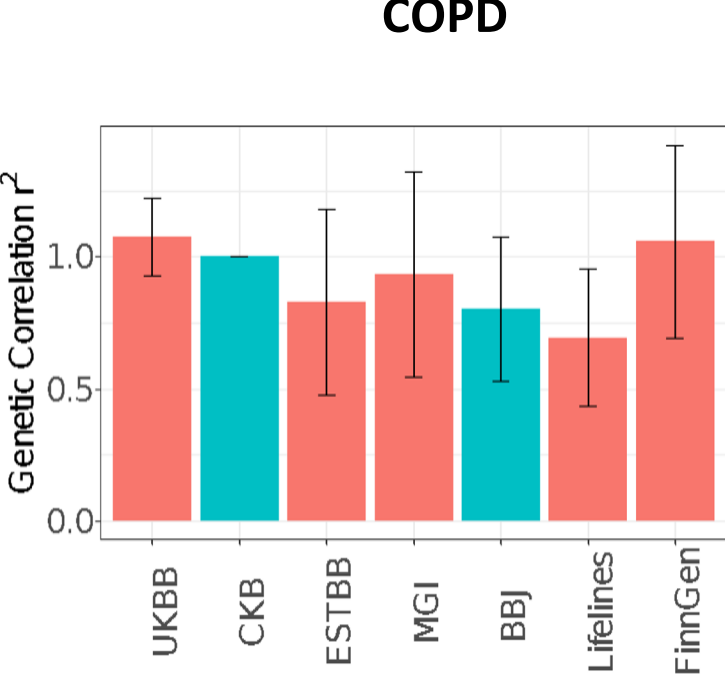
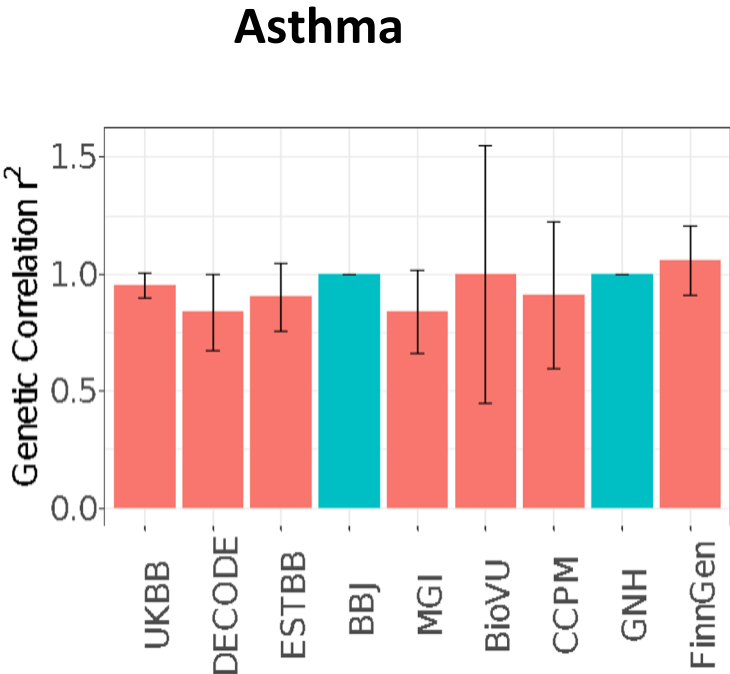
Supplementary Figure 8. The slopes of Deming regression for effect sizes for index variants in each biobank and leave-one-biobank-out meta-analysis (LOBO) pair are plotted against the effective sample sizes. Index variants with association p-values $< 1 \times 10^{-10}$ in the all-biobank meta-analysis were used for the regression. Biobanks, in which at least three index variants passed the cutoff, are plotted. Biobanks are annotated by phenotype source, sampling strategy and sample ancestry. The dotted line indicates $y=1$. A positive slope indicates that effect size estimates of the top hits are higher in the leave-one-biobank-out (LOBO) meta-analysis than in the individual biobank and a negative slope suggests lower effect size estimates in LOBO meta-analysis than in the individual biobank. The effective sample sizes is calculated as $4/(1/\text{case number} + 1/\text{control number})$.



Supplementary Figure 9. Scatter plots of the effect size estimates in population-based biobanks and hospital-/healthcare-based biobanks with the Deming regression lines and the slope estimates (intercepts were fixed to 0). Loci that were genome-wide significant in all-biobank meta-analyses and have p-value $< 1 \times 10^{-6}$ in both meta-analyses of population-based biobanks and hospital-/healthcare-based biobanks, respectively, were included in the analyses. Endpoints that have more than 5 loci included in the analyses were plotted.

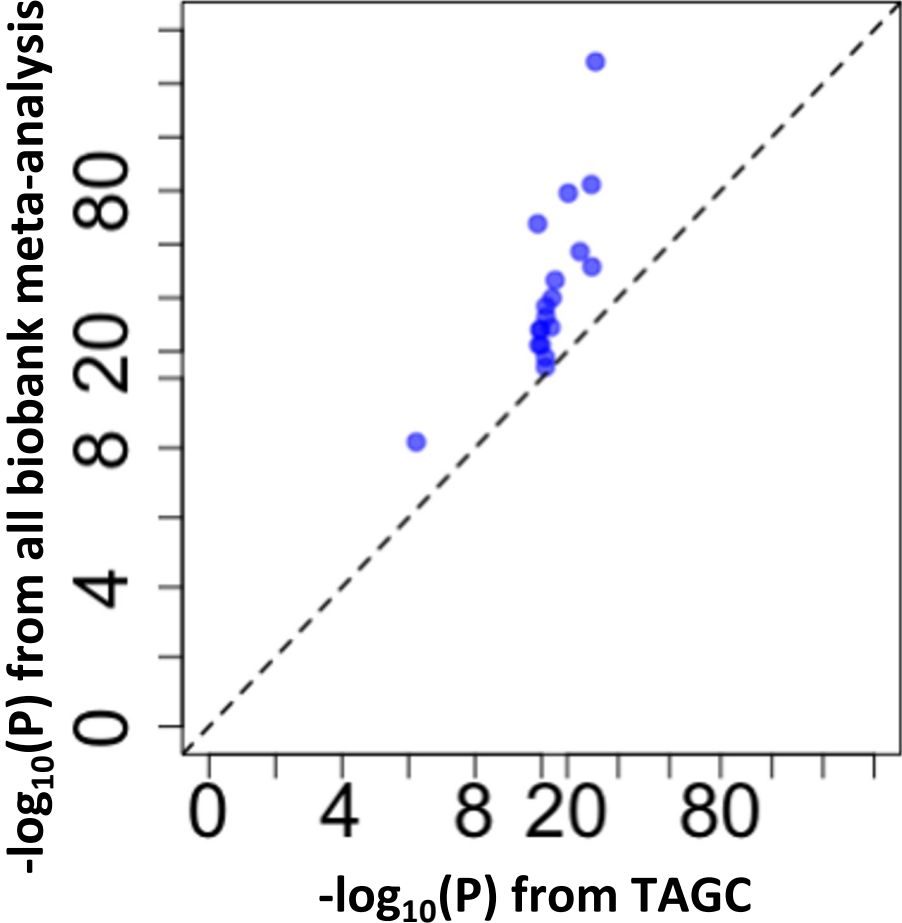


Supplementary Figure 10. Genetic correlation between each biobank and leave-on-biobank meta-analysis in GBMI

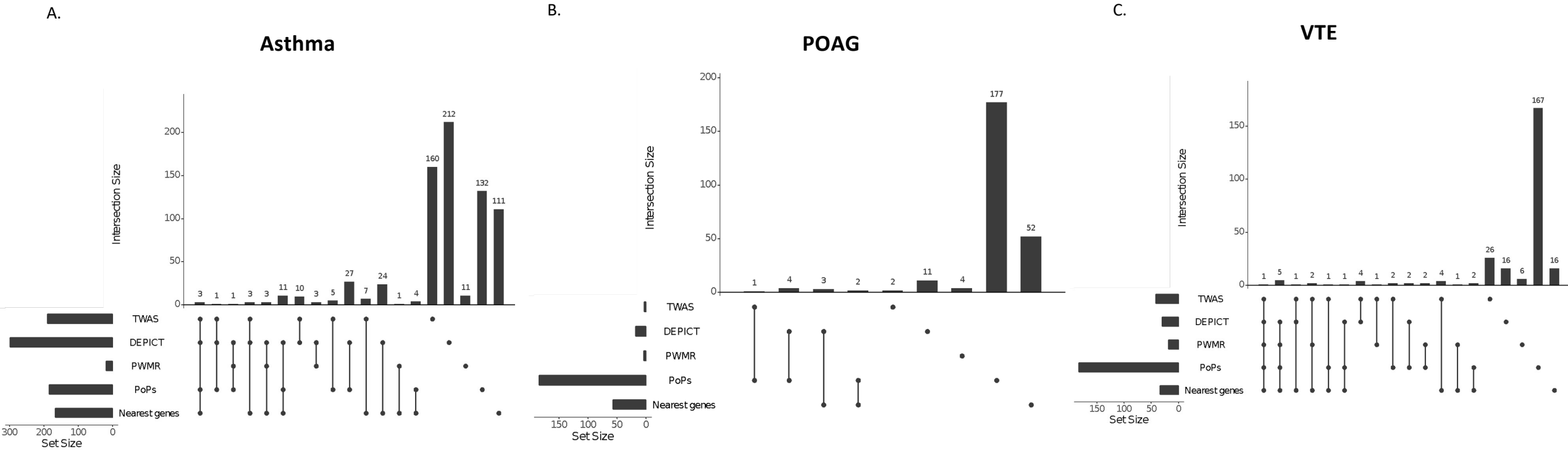


Method
ldscore
popcorn

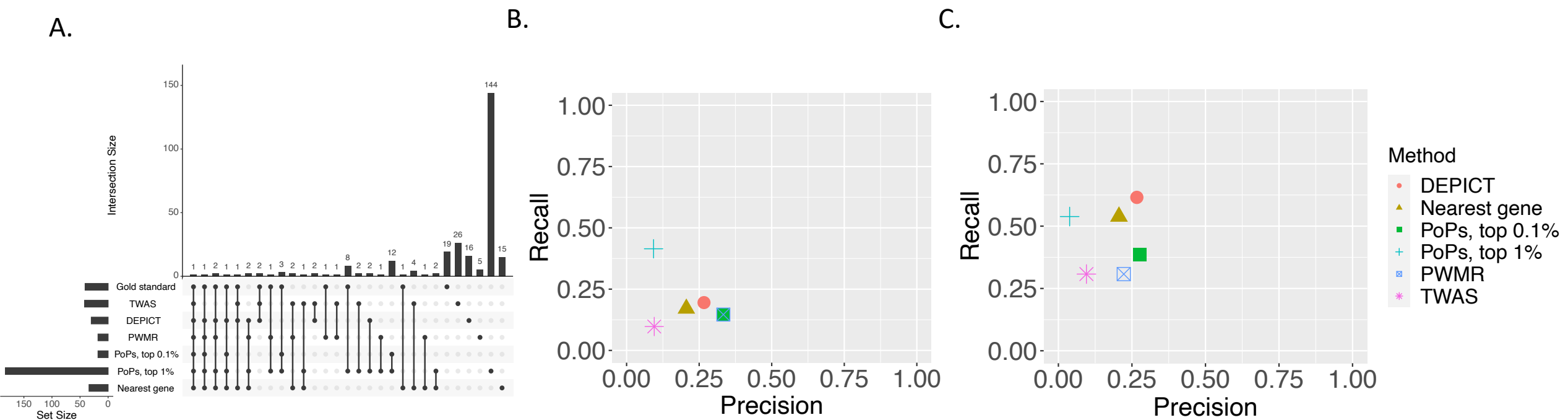
Supplementary Figure 11. All 18 loci identified by previous GWAS for Asthma (Demenais et al., 2018) have more significant p-values in all-biobank meta-analysis (**Supplementary Table 12**)



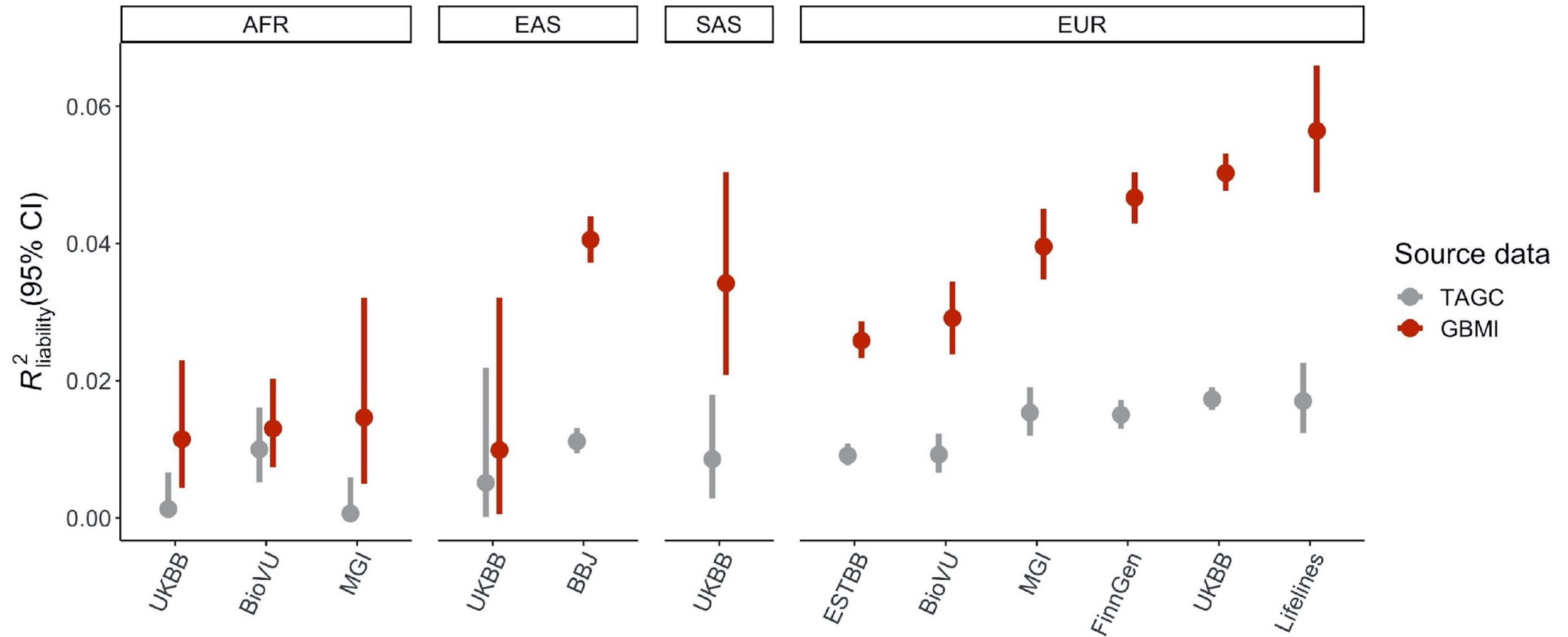
Supplementary Figure 12. Number of genes prioritized by different methods: PoPs (top 1%), DEPICT (FDR < 0.05), TWAS (P < 2.5 x 10⁻⁶), PWMR (P < 0.001, Colocalization probability > 0.7), nearest genes around the top hits (Nearest gene, for intergenic variants, the nearest gene on each side will be included if both are located within 50kb from the top hit).



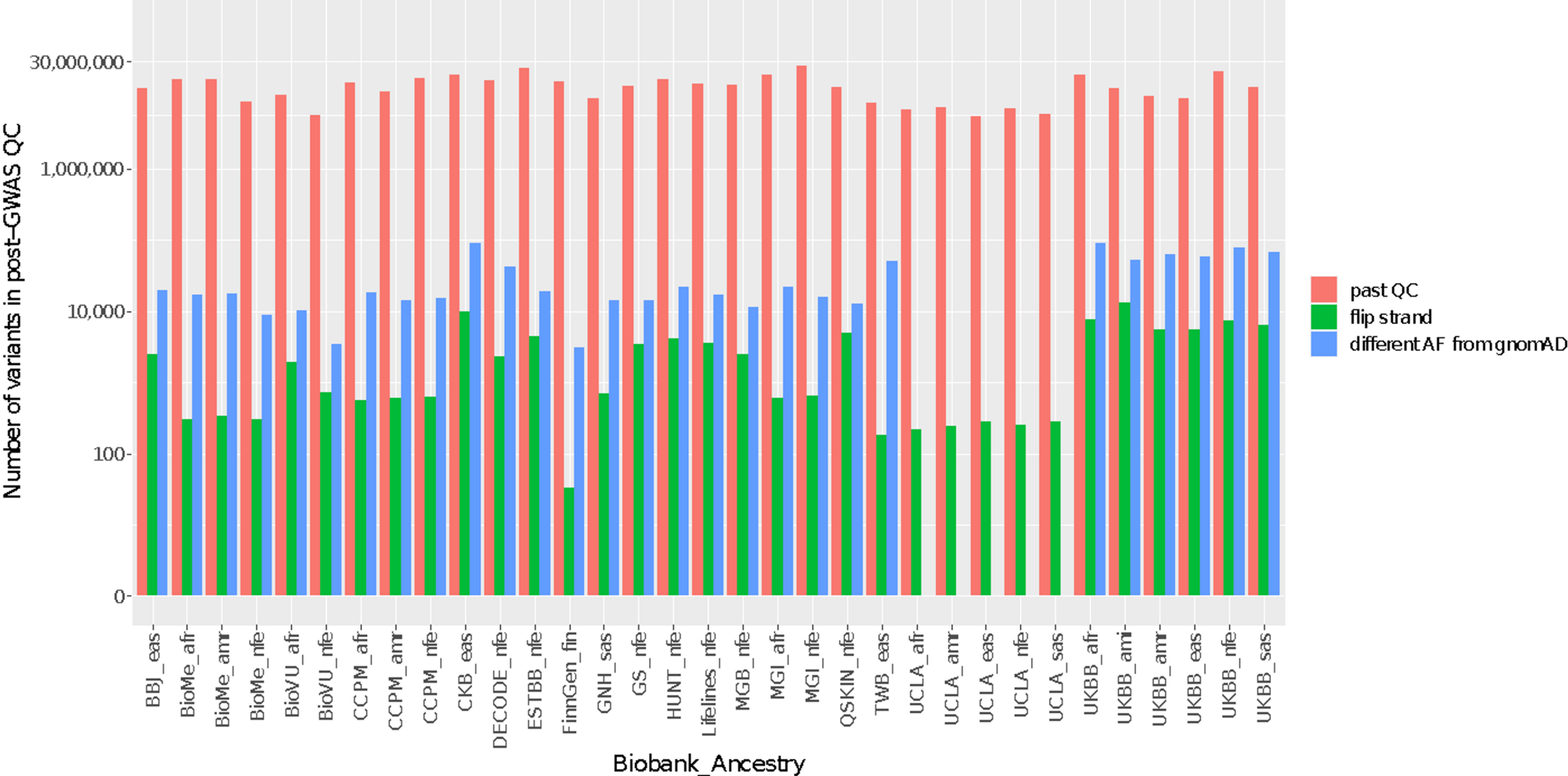
Supplementary Figure 13. For VTE, A. number of genes prioritized by different methods: PoPs (top 1% and top 0.1%), DEPICT (FDR < 0.05), TWAS (P < 2.5 x 10⁻⁶), PWMR (P < 0.001, Colocalization probability > 0.7), nearest genes around the top hits (Nearest gene, for intergenic variants, the nearest gene on each side will be included if both are located within 50kb from the top hit). B. precision and recall of the different gene prioritization methods based on a gold standard set of 41 VTE genes that was curated prior to the meta-analysis by medical and molecular genetics experts in VTE (Wolford et al. 2021). C. precision and recall of the different gene prioritization methods based on 13 genes (ADAMTS13, F10, F2, F5, F7, FGA, FGB, FGG, PROC, PROS1, PROZ, THBD, VWF) in the gold standard sets that fall within 1Mb around VTE top hits in GBMI meta-analysis. The prioritization gene lists are presented in **Supplementary Table 24**.



Supplementary Figure 14. Improved polygenic risk scores (PRS) prediction accuracy using GBMI meta-analysis results compared to TAGC summary statistics.



Supplementary Figure 15. Palindromic SNPs with potential strand flip and genetic variants with different allele frequencies compared to gnomAD were flagged when included in the meta-analyses



Supplementary Figure 16. The distribution of number of biobanks that contain the genetic variants with different allele frequencies compared to gnomAD.

