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Common variants at 21q22.3 locus influence *MX1* and *TMPRSS2* gene expression and susceptibility to severe COVID-19

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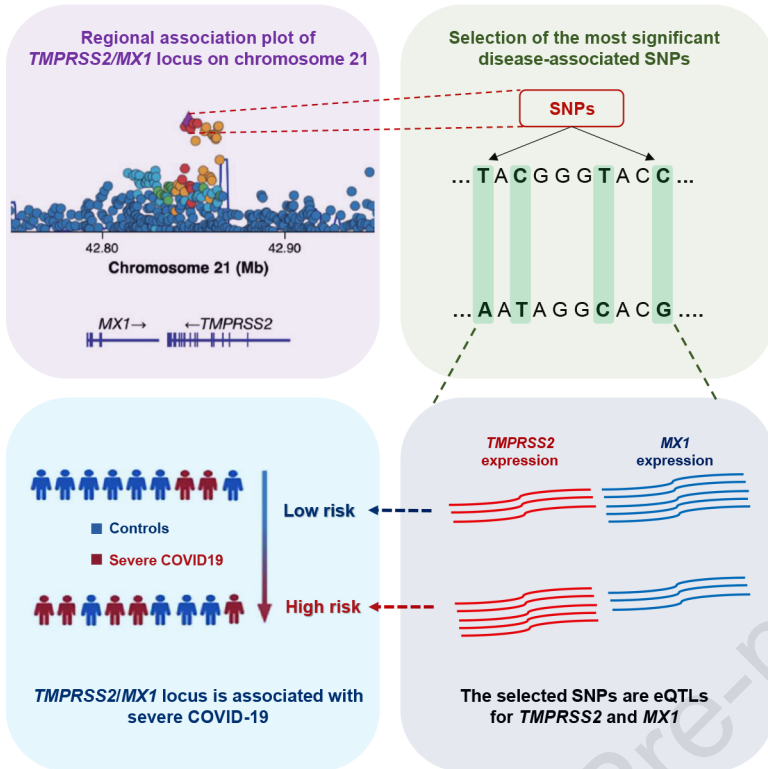
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1 **Common variants at 21q22.3 locus influence *MX1* and *TMPRSS2* gene**
2 **expression and susceptibility to severe COVID-19**

3

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29 **Keywords:** COVID-19, SARS-CoV-2, *TMPRSS2*, *MX1*, SNP genotyping.

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31

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41

42 **Summary**

43

44 The established risk factors of coronavirus disease 2019 (COVID-19) are advanced age, male
45 sex and comorbidities, but they do not fully explain the wide spectrum of disease
46 manifestations. Genetic factors implicated in the host antiviral response provide for novel
47 insights into its pathogenesis.

48 We performed an in-depth genetic analysis of chromosome 21 exploiting the genome-wide
49 association study data, including 6,406 individuals hospitalized for COVID-19 and 902,088
50 controls with European genetic ancestry from the COVID-19 Host Genetics Initiative. We
51 found that five single nucleotide polymorphisms within *TMPRSS2* and near *MX1* gene show
52 associations with severe COVID-19. The minor alleles of the five SNPs correlated with a
53 reduced risk of developing severe COVID-19 and high level of *MX1* expression in blood.

54 Our findings demonstrate that host genetic factors can influence the different clinical
55 presentations of COVID-19 and that *MX1* could be a potential therapeutic target.

56

57 **Introduction**

58

59 The recent severe acute respiratory syndrome coronavirus 2 (SARS-Cov-2) pandemic has
60 caused so far more than over 2.5 million deaths (<https://covid19.who.int/>). The coronavirus
61 disease 2019 (COVID-19), caused by the SARS-Cov-2, is associated with diverse clinical
62 presentations, ranging from asymptomatic or mildly symptomatic infections to respiratory
63 failure and death (Bellani et al., 2021; Grasselli et al., 2021; Grasselli et al., 2020; Richardson
64 et al., 2020). Advanced age is an established risk factor, as well as male sex and
65 comorbidities such as hypertension and diabetes (Zhou et al., 2020). Since these risk factors
66 do not fully explain the wide spectrum of disease manifestations, dissecting the genetics of
67 the host response to SARS-CoV-2 infection may provide novel insights into its pathogenesis
68 (Anastassopoulou et al., 2020).

69 A genome-wide association study (GWAS) (Ellinghaus et al., 2020) identified two
70 susceptibility loci of severe COVID-19: the first locus on chromosome 3 harbors multiple
71 genes (*SLC6A20*, *LZFTL1*, *CCR9*, *CXCR6*, *XCRI*, *FYCO1*) that could be functionally
72 implicated in COVID-19 pathology; the second on chromosome 9 that defines the ABO
73 blood groups (Ellinghaus et al., 2020). Other very recent papers reported the results from the
74 analysis of two large independent GWASs that validated the two previous risk loci and found
75 novel risk variants at chromosome 19p13.3, 12q24.13, and 21q22.1 associated with severe
76 COVID-19 (Pairo-Castineira et al., 2020; Shelton et al., 2020).

77 Two whole exome sequencing studies showed that inactivating rare mutations in genes
78 belonging to the type I interferon pathway predispose to life-threatening COVID-19
79 pneumonia (van der Made et al., 2020; Zhang et al., 2020). Additionally, preliminary results
80 on a small set of Italian cases suggest that coding variants in *TMPRSS2* and *PCSK3* may
81 contribute to the variability in infection susceptibility and severity. (Latini et al., 2020).

82 In our previous opinion article, based on the analysis of allele frequencies across different
83 populations and expression quantitative trait loci (eQTLs) data, we hypothesized that
84 common variants on chromosome 21 near *TMPRSS2* and *MXI* genes may be genetic risk
85 factors associated with the COVID-19 different clinical manifestations (Russo et al., 2020).
86 Both *TMPRSS2* and *MXI* are involved in the host response to SARS-CoV-2 infection. ACE2
87 is the main entry receptor for SARS-CoV-2 (Wang et al., 2020). Entry depends on the
88 binding of the surface unit S1 of the spike (S) protein of the virus to the receptor. SARS-
89 CoV-2 engages ACE2 as the entry receptor and employs the host cellular TMPRSS2 for S-

90 protein priming (Hoffmann et al., 2020b; Matsuyama et al., 2010). Particularly, binding of
91 SARS-CoV-2 S- protein with ACE2 receptor is then followed by host TMPRSS2-mediated
92 cleavage of the viral S-protein. This process, defined as priming, involves cleavage of the S-
93 protein at S1/S2 and S2 sites which is essential for the viral fusion with the host cell
94 membrane before entry into the cell (Hoffmann et al., 2020b; Matsuyama et al., 2020).
95 SARS-CoV-2 can use other proteases such as cathepsin B/L for S-protein in the absence of
96 TMPRSS2 receptors. However, in the lungs (the primary organ for SARS-CoV-2 infection),
97 cathepsin B/L cannot substitute for TMPRSS2 protease activity as the latter is indispensable
98 for viral entry as observed for SARS-CoV and MERS-CoV (Hoffmann et al., 2020a). *MXI* is
99 an interferon- α/β inducible gene that encodes a guanosine triphosphate metabolizing protein
100 involved in the cellular antiviral response (Ciancanelli et al., 2016).

101 In this study, to further support our hypothesis, we exploited GWAS meta-analysis data from
102 the COVID-19 Host Genetics Initiative (COVID-19 Host Genetics Initiative, 2020) and
103 performed an in-depth genetic analysis of chromosome 21 using summary statistics where
104 common variants at this chromosome were associated with severe COVID-19 at the genome-
105 wide significance level ($P \leq 5 \times 10^{-8}$). Using the cohort of 908,494 subjects with European
106 origins, we found five single nucleotide polymorphisms (SNPs) at the *TMPRSS2/MXI* locus
107 showing suggestive association with the disease. All five SNPs replicated the association in
108 two independent cohorts of Asian subjects, whereas two SNPs confirmed the association in
109 African and one SNP in the Italian cohort. Significant eQTLs signals were found for the *MXI*
110 gene in blood.

111 Results

112 *TMPRSS2/MXI* locus is associated with severe COVID-19

113 To prove that common variants at *TMPRSS2/MXI* (21q22.3) locus may affect the
 114 susceptibility to severe COVID-19 onset, we analyzed the summary statistics of a large
 115 available GWAS dataset released by the COVID-19 Host Genetics Initiative (COVID-19
 116 Host Genetics Initiative, 2020). The dataset includes 6,406 hospitalized cases and 902,088
 117 controls with European ancestry (“**Table S1**. Study groups that have contributed to GWAS
 118 meta-analyses of the COVID-19 Host Genetics Initiative, Related to Figure 1”). A region on
 119 chromosome 21 appears to be significantly associated with severe COVID-19 at the genome-
 120 wide level (<https://www.covid19hg.org/results/>) as also demonstrated in a recently published
 121 GWAS study (Pairo-Castineira et al., 2020). To investigate whether more than one
 122 association signals may exist at chromosome 21, we selected 74 SNPs showing a $P \leq 1 \times 10^{-5}$
 123 and we identified 3 independent loci among them (“**Table S2**. Summary statistics at
 124 chromosome 21 from GWAS dataset, Related to Figure 1). The most significant signal was
 125 represented by rs13050728 ($P = 2.76 \times 10^{-12}$, OR=0.83, **Figure 1a**) that maps within the
 126 *INFRA2* gene. The other two signals showed a suggestive significance level ($P \leq 1 \times 10^{-5}$) and
 127 were tagged by rs111783124 ($P = 2.39 \times 10^{-6}$, OR=1.17, **Figure 1b**) and rs3787946
 128 ($P = 2.73 \times 10^{-6}$, OR=0.87, **Figure 1c**), respectively. The rs3787946 maps in an intronic region
 129 of *TMPRSS2* and the first closest gene was *MXI* (**Figure 1c**); herein, we named this locus as
 130 “*TMPRSS2/MXI*”. An in-depth inspection of the *TMPRSS2/MXI* locus showed that 13 SNPs
 131 were in linkage disequilibrium (LD) with the lead rs3787946 ($r^2 > 0.8$, **Table 1**) and that the 5
 132 most significant SNPs (P-values ranging from 2.7×10^{-6} to 5.8×10^{-6} , **Table 1**) were in strong
 133 LD with each other ($r^2 \geq 0.90$, “**Figure S1**. Linkage disequilibrium block at *TMPRSS2/MXI*
 134 locus, Related to Figure 1”). The other 9 SNPs showed an LD with the lead SNP rs3787946
 135 ranging from 0.8 to 0.9 and P-values ranging from 6×10^{-4} to 0.04 (**Table 1**). We then sought
 136 to replicate the associations of the 14 SNPs in three independent cohorts of cases and controls
 137 of GenOMMIC GWAS (Pairo-Castineira et al., 2020) with non-European ancestry. All the 11
 138 available SNPs replicated in the east asian population (EAS) population; the top five SNPs
 139 replicated in the South Asian (SAS) ancestry population, whereas two out of five SNPs in the
 140 African (AFR) one (**Table 1**). By using the TaqMan assay, we typed the rs12329760 variant
 141 in samples from 226 hospitalized COVID-19 patients (“**Table S3**. Characteristics of Italian
 142 patients recruited by our research group, Related to Table 1”) and 1848 controls from
 143 Southern Italy collected in our Institute. An additional Italian cohort of 1915 controls and 770

144 cases, typed for rs12329760 by whole-exome sequencing, was obtained from the Network for
 145 Italian Genomes (NIG) database (Daga et al., 2021). After combining the two cohorts, we
 146 confirmed the minor allele as a protective factor against the aggressive form of the disease
 147 (**Table 2**, $OR_{\text{allele}}=0.89$, $P_{\text{allele}}=0.07$; $OR_{\text{dominant}}=0.57$, $P=0.01$; $OR_{\text{CCvsTT}}=0.57$, $P=0.01$). The
 148 results of our case-control study suggest that the protective effect against the severity of
 149 COVID-19 is mainly due to the TT genotype.

150

151 **SNPs at *TMPRSS2/MXI* locus are enriched in regulatory regions active in the thymus**

152 We tested if the 14 SNPs (**Table 1**) and their proxy SNPs ($r^2>0.8$) were significantly over-
 153 represented in active enhancers and promoters in multiple cell types and tissues by using
 154 HaploReg v4.1. These SNPs were enriched in the regulatory regions of several tissues
 155 (“**Table S4**. Results of SNP enrichment analysis in regulatory elements in different tissues
 156 and cell types, Related to Figure 2”), but the best enrichment was found in induced
 157 pluripotent stem cells and thymus (**Figure 2a**).

158

159 **Functional role of the most significant SNPs at *TMPRSS2/MXI* locus**

160 We then investigated the predicted functional role of the 14 SNPs by GWAVA and CADD
 161 tools. We found that two out of the five most significant SNPs, i.e. rs9983330 and
 162 rs12329760, showed the first (combined score=26) and second (combined score=23) most
 163 significant score (**Table 1**). The rs12329760 was classified as a coding variant (p.Val197Met)
 164 localized in the exon 6 of the *TMPRSS2* gene and was predicted to be pathogenic (PolyPhen-
 165 2=probably damaging and SIFT=deleterious).

166

167 **The most significant disease-associated SNPs are eQTLs for *MXI* in blood**

168 We verified if the top five SNPs (**Table 1**) might cause gene expression alterations
 169 interrogating the GTEx portal for all the common variants within *TMPRSS2/MXI* locus. We
 170 found that all the top five SNPs had eQTL signals for *MXI* exclusively in blood tissue.
 171 Particularly, the minor alleles of these SNPs correlated with higher expression of *MXI*
 172 compared to the major alleles (**Figure 2b**, “**Figure S2a**. Results of SNP enrichment analysis
 173 in regulatory elements in different tissues and cell types, Related to Figure 2”). Of note, all
 174 the other SNPs, except for rs2298660, did not have eQTL signals for *MXI* in the blood
 175 (“**Table S5**. Results of eQTL analysis for the common variants at *TMPRSS2/MXI* locus,
 176 Related to Figure 2). The two SNPs rs12329760 and rs2298660 were confirmed as eQTLs for
 177 *MXI* in the blood ($P=1.79\times 10^{-6}$ and 2.8×10^{-6} , minor alleles correlated with a higher

178 expression compared to the major alleles) by interrogation of another independent publicly
179 available dataset (Westra et al., 2013). *TMPRSS2* is highly expressed in lung (Russo et al.,
180 2020), so we investigated if the top five SNPs were eQTLs for *TMPRSS2* in lung tissues at a
181 nominally statistically significant level ($P \leq 0.05$). We found that the minor alleles of four out
182 of five SNPs correlated with lower expression of *TMPRSS2* compared to the major alleles
183 (**Figure 2c and “Figure S2b. Results of SNP enrichment analysis in regulatory elements in**
184 **different tissues and cell types, Related to Figure 2”**). Notably, rs12329760 is also an eQTL
185 for *TMPRSS2* in osteoblasts treated with dexamethasone (Grundberg et al., 2011).
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187 Discussion

188 Despite the substantial advances made in recent months in the field of SARS-CoV-2
189 infection, the major question remains about the identification of the factors that modulate the
190 variable clinical spectrum of COVID-19.

191 Host genetic risk factors are emerging as a potential explanation for the clinical heterogeneity
192 of COVID-19 and are also crucial to find new druggable therapeutic targets (Asselta et al.,
193 2020; Beck and Aksentijevich, 2020; Benetti et al., 2020; Pairo-Castineira et al., 2020; Singh
194 et al., 2020). The main host cell entry factors of SARS-CoV-2 are ACE2 and TMPRSS2
195 (Asselta et al., 2020; Benetti et al., 2020). The spike (S) glycoprotein of the virus binds to the
196 ACE2 making it essential for the entry of the virus into the host cell. S- protein priming by
197 the serine protease TMPRSS2 allows the fusion of viral and cellular membranes, resulting in
198 virus entry and replication in the host cells (Singh et al., 2020). TMPRSS2 is emerging as a
199 host cell factor that is critical for SARS-CoV-2 infection (Hoffmann et al., 2020b).

200 In our previous study, we hypothesized that common variants at chromosome 21, driving
201 *TMPRSS2* and *MX1* expression, might have a mild-to-moderate effect on the susceptibility to
202 SARS-CoV-2 infection. Particularly, genetic variants associated with reduced *TMPRSS2* and
203 elevated *MX1* expression might confer less individual susceptibility to SARS-CoV-2
204 infection and favor a better outcome (Russo et al., 2020). Here, to further support our
205 hypothesis, we exploited GWAS data of a cohort of 908,494 subjects with European origins
206 from the COVID-19 Host Genetics Initiative (COVID-19 Host Genetics Initiative, 2020) and
207 performed an in-depth genetic analysis of chromosome 21. We identified five common
208 variants (rs3787946, rs9983330, rs12329760, rs2298661, and rs9985159) at locus 21q22.3
209 within *TMPRSS2* and near the *MX1* gene that showed suggestive associations with severe
210 COVID-19. In particular, we found that the alleles with minor frequency were less recurrent
211 among hospitalized patients when compared to the control individuals, suggesting their
212 protective role against the progression of the disease. Interestingly, all five SNPs were
213 replicated in two cohorts of Asian origin, whereas two SNPs replicated in a case series of
214 African ancestry. Additionally, we replicated the association of the rs12329760 SNP in an
215 independent case-control cohort of Italian origin. As “proof of concept”, the rs12329760 SNP
216 was also detected in recent studies (Hou et al., 2020; Vargas-Alarcon et al., 2020). It was
217 demonstrated that the SNP, in addition to its eQTL role, decreased the stability of the protein,
218 which might impede viral entry (Vishnubhotla et al., 2020); moreover, *in silico* analysis
219 demonstrated that it created a *de novo* pocket protein (Paniri et al., 2020). These results

220 confirm 21q22.3 as a novel susceptibility locus to unfavorable outcome of COVID-19.
221 Furthermore, molecular mechanisms underlying this genetic predisposition may be common
222 among individuals with different ethnicity.

223 The results from our enrichment analysis for regulatory genomic regions suggested that the
224 identified SNPs and other proxy SNPs located at 21q22.3 locus can be associated with
225 different outcomes of COVID-19 by altering DNA elements that regulate the transcription of
226 *MX1* and likely of other genes relevant to the thymus functions. The thymus plays a
227 significant role in the regulation of adaptive immune responses. The effect of aging on the
228 thymus and immune senescence is well established, and the resulting inflammaging is found
229 to be implicated in the development of many chronic diseases (Gunes et al., 2020; Kellogg
230 and Equils, 2020). Both aging and diseases of inflammaging are associated with severe
231 COVID-19, and a dysfunctional thymus may be implicated in the unfavorable outcome of
232 disease (Gunes et al., 2020; Kellogg and Equils, 2020). Of note, *MX1* plays an important role
233 in the thymus as part of the innate antiviral immune response. Indeed, it is exclusively
234 expressed after engagement of the type I interferon receptor by interferon- α/β in normal fetal
235 and post-natal human thymus, but not in the periphery. The highest level of *MX1* is properly
236 found in mature thymocytes (Colantonio et al., 2011).

237 The five SNPs here identified had eQTL signals for *MX1* exclusively in blood tissue.
238 Particularly, the minor allele of these SNPs correlated with higher expression of *MX1* and
239 associated with a minor risk of developing severe COVID-19. These results support the
240 evidence that *MX1* can play a relevant role in determining less severe forms of disease and
241 are in line with a recent study that suggests *MX1* as an antiviral effector against SARS-CoV-2
242 (Bizzotto et al., 2020). Indeed, the expression of *MX1* was found to be high in SARS-CoV-2
243 positive subjects, negatively correlated with age, and independently associated with increased
244 viral load (Bizzotto et al., 2020). *MX1* is part of the antiviral response induced by type I and
245 III interferons (Zav'yalov et al., 2019). Inactivating mutations in genes belonging to type I
246 interferon pathway and the consequently decreased levels of proteins have been shown to
247 occur in patients with severe COVID-19 (Zhang et al., 2020).

248 Of note, within the region on chromosome 21, significantly associated with severe COVID-
249 19 at the genome-wide level, the most significant signal was represented by rs13050728 that
250 maps within the *INFRA2* gene. Particularly, *INFRA2* gene encodes for the type I membrane
251 protein that forms the interferon- α/β receptor, involved in the canonical host antiviral
252 signalling mediators (Duncan et al., 2015), so associated with interferon signalling like
253 *MX1*. The SNP rs13050728 was previously identified as lead variant from the meta-analysis

254 of overlapping SNPs between GenOMICC, The COVID-19 Host Genetics Initiative and
255 23andMe studies and its allele C was reported to reduce the odds of severe COVID-19 as
256 associated with an increased expression of *IFNAR2* (Pairo-Castineira et al., 2020). These
257 findings, along with ours, further strength the protective role of IFN pathway against severe
258 COVID-19.

259 We also report that the minor allele of four of the top five SNPs might reduce the expression
260 of *TMPRSS2* in lung tissues. In particular, the rs12329760 coding variant (p.Val197Met) is
261 predicted to decrease the *TMPRSS2* protein stability and ACE2 binding, thus decreasing
262 virus entry into the cells (Vishnubhotla et al., 2020). Of note, this variant was recently found
263 to be less frequent among Chinese patients with critical COVID-19 disease (Wang et al.,
264 2020). Additionally, it correlates with lower expression of *TMPRSS2* in osteoblast treated
265 with dexamethasone (Grundberg et al., 2011), a drug currently used to inhibit an excessive
266 inflammation response (Group et al., 2020). Together, these data suggest that even the
267 functions of *TMPRSS2* may be affected by the occurrence of protective variants against
268 severe COVID-19.

269 Finally, we want to point out that our findings highlight the effectiveness of investigating
270 other independent (putative) risk loci, when they do not pass genome-wide significance
271 levels. These loci, usually overlooked in extensive meta-analysis and multi-cohorts efforts,
272 might indeed contain important genetic variants associated with severe COVID-19 and map
273 genes relevant to the pathogenesis of this disease. We then encourage post-GWAS genetic
274 (re)analyses using multiple data sources to unravel novel COVID-19 risk loci and possible
275 insights on the underlying biology.

276 In conclusion, our results provide evidence that common variants, regulating the expression
277 of *MX1*, can predispose to the risk of developing severe COVID-19. Unraveling the role of
278 regulatory variants at the *TMPRSS2/MX1* locus could represent an important starting point
279 for the treatment of COVID-19.

280

281 **Limitations of the Study**

282 The data on eQTLs related to *TMPRSS2* must be interpreted with caution as these eQTL
283 signals in the lung (P=0.019) do not pass the GTEx significance threshold adjusted for
284 multiple comparisons (0.000055). Additional studies are required to further verify the role of
285 genetic variants at *TMPRSS2/MX1* locus in modulating the *TMPRSS2* expression.
286 Furthermore, the statistical approach adopted in this study did not include multivariate
287 analyses to take into account confounding factors. Although this limitation does not affect the

288 robustness of the presented genetic associations as replicated in multiple independent cohorts,
289 we believe that future studies will help to better define the effect of genetic variants at
290 *TMPRSS2/MX1* locus on the clinical subgroups of COVID-19 disease; for instance,
291 performing association analyses on patients stratified by disease aggressiveness or controlled
292 for comorbidities in larger cohorts.
293

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294 **All methods can be found in the accompanying “Transparent methods supplemental**
295 **file”.**

296

297 **Resource availability**

298 Further information and requests for resources should be directed to and will be fulfilled by
299 the Lead Contact, Prof. Mario Capasso, mario.capasso@unina.it.

300

301 **Material availability**

302 This study did not generate nor use any new or unique reagents.

303

304 **Data and code availability**

305 Manhattan plot and QQ plot of the results from the large GWAS “The COVID-19 Host
306 Genetics Initiative website” are available at the website (<https://www.covid19hg.org/results/>).

307 The 770 hospitalized COVID-19 cases and 1915 controls typed for rs12329760 by whole-
308 exome sequencing were retrieved from the web database Network for Italian Genomes (NIG)
309 available at the website (<http://nigdb.cineca.it/index.php>).

310 Prediction of the functional impact of 14 SNPs at TMPRSS2/MX1 locus was assessed by
311 Genome Wide Annotation of VARIants (GWAVA) tool available at the website
312 (https://www.sanger.ac.uk/sanger/StatGen_Gwava) and by Combined Annotation Dependent
313 Depletion (CADD) tool at (<https://cadd.gs.washington.edu/>).

314 The Blood eQTL Browser is available at (<https://www.genenetwork.nl/bloodeqtlbrowser/>).

315

316

317 Author Contributions

318 IA, RR, and MC designed and conducted the study, and prepared the manuscript; MC, VAL,
319 and FB analyzed the data; BER sampled genomic DNA from COVID-19 patients; SC
320 genotyped COVID-19 patients and in-house controls; GF, AP, GMC, GS, GE, IG, CP, RV,
321 GP, PC, CB, BP cared for COVID-19 patients; MZ and AI provided a critical review of the
322 manuscript. All the authors read and approved the final manuscript.

323

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331

332

333 Declaration of interests

334 The authors declare that there are no competing interests.

335

336

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338

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457 **Figure legends**

458

459 **Figure 1. Regional association plots of the SNPs at three independent association signals**
460 **of chromosome 21.** Plots were generated using LocusZoom. Y-axes represent the
461 significance of association ($-\log_{10}$ transformed P values) and the recombination rate. SNPs
462 are color-coded based on pairwise linkage disequilibrium (r^2) with indicated lead SNPs:
463 rs13050728 (panel a), rs111783124 (panel b) and rs3787946 (panel c).

464

465 **Figure 2. Enrichment of SNPs in regulatory regions and eQTL analyses.** The statistically
466 significant fold enrichments ($P < 0.05$ after Bonferroni correction) of SNPs in regulatory DNA
467 regions active in different tissues are shown (a). eQTL violin plots between genotypes of
468 rs3787946 (b) and rs3787946 (c) with *MX1* and *TMPRSS2* expression from the from the
469 Genotype-Tissue Expression (GTEx). The significance threshold adjusted for multiple
470 comparisons is equal to 0.000055.

471

472

Table 1. Associations of SNPs at *TMPRSS2/MX1* risk locus in linkage disequilibrium with the lead rs3787946 in different populations and prioritization scores

RS number	EA	OA	MAF	r ²	OR	P_EUR	OR	P_EAS	OR	P_SAS	OR	P_AFR	*Region score	*TSS score	^Predicted Function	^Score	°Combined score
rs3787946	C	G	0.23	1.00	0.87	2.73E-06	0.63	0.026	0.71	0.02	0.74	0.07	0.16	0.29	INTRONIC	2	6
rs9983330	G	A	0.23	0.91	0.88	3.12E-06	0.54	0.004	0.73	0.04	0.79	0.16	0.31	0.64	REGULATORY	4	26
rs12329760	T	C	0.24	0.90	0.88	3.13E-06	0.64	0.029	0.76	0.08	0.78	0.14	0.32	0.41	MISSENSE	7	23
rs2298661	A	C	0.23	0.99	0.88	4.51E-06	0.63	0.030	0.67	0.01	0.60	0.01	0.18	0.35	INTRONIC	2	9
rs9985159	T	C	0.23	0.98	0.88	5.80E-06	0.61	0.018	0.75	0.06	0.98	0.89	0.16	0.46	INTRONIC	2	15
rs2298660	T	C	0.20	0.82	0.88	0.001	NA	NA	NA	NA	NA	NA	0.12	0.28	INTRONIC	2	4
rs7364088	A	G	0.26	0.84	0.91	0.002	NA	NA	NA	NA	NA	NA	0.19	0.23	INTRONIC	2	6
rs2298663	T	C	0.25	0.87	1.08	0.005	1.49	0.052	1.12	0.40	0.94	0.66	0.26	0.37	REGULATORY	4	15
rs2094881	C	T	0.25	0.87	1.08	0.005	1.47	0.058	1.10	0.47	0.93	0.60	0.29	0.26	REGULATORY	4	13
rs8131649	T	C	0.25	0.85	0.92	0.007	0.64	0.035	0.90	0.46	1.01	0.93	0.26	0.35	REGULATORY	4	12
rs8134203	T	C	0.26	0.85	1.08	0.007	1.49	0.058	1.09	0.54	0.91	0.50	0.26	0.41	REGULATORY	4	17
rs8134216	T	C	0.26	0.85	1.08	0.007	1.54	0.038	1.11	0.43	0.91	0.49	0.28	0.4	REGULATORY	4	19
rs2104810	A	G	0.26	0.85	1.08	0.008	1.54	0.040	1.10	0.47	0.90	0.48	0.23	0.35	REGULATORY	4	11
rs8131648	C	T	0.26	0.85	1.07	0.036	NA	NA	NA	NA	NA	NA	0.33	0.42	REGULATORY	4	26

*Scores from GWAVA predictor tool

^Scores from CADD predictor tool

°GWAVA and CADD scores were ranked from the smallest to largest and the obtained values were summed

In bold the SNPs that replicated in at least one cohort

EA: Effect Allele; OA: Other Allele

EUR: European; EAS: East Asian; SAS: South Asian; AFR: African; ITA: Italian

MAF: minor allele frequency

OR: Odds Ratio

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474

Table 2. Association of rs12329760 SNP with severe COVID-19 in Italian population

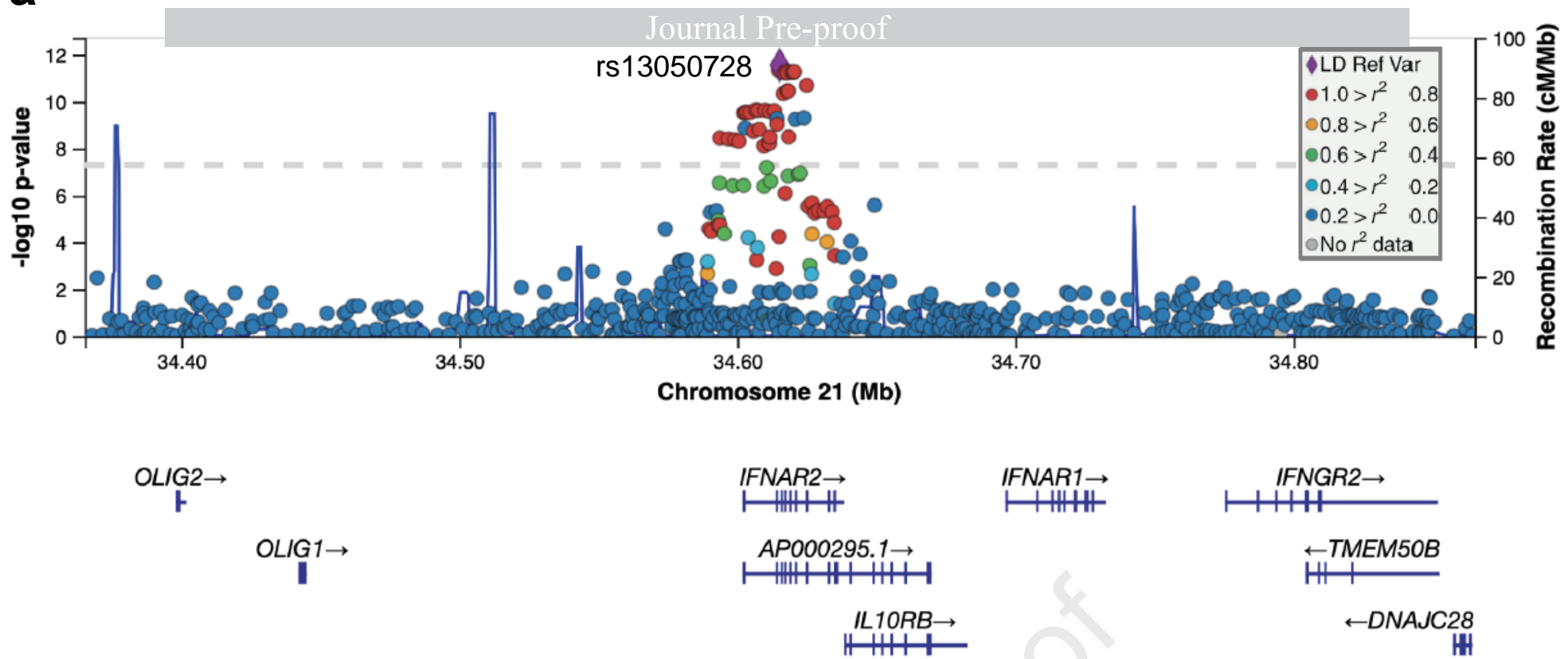
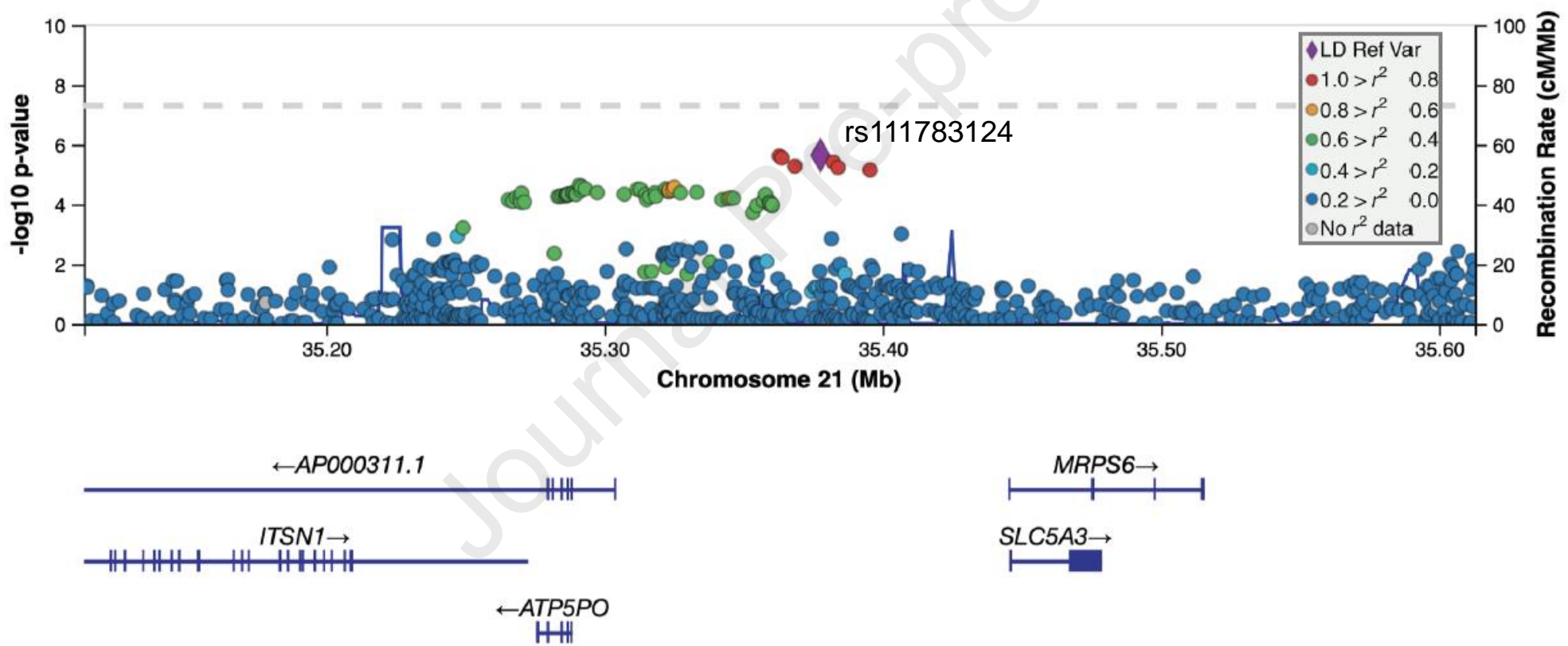
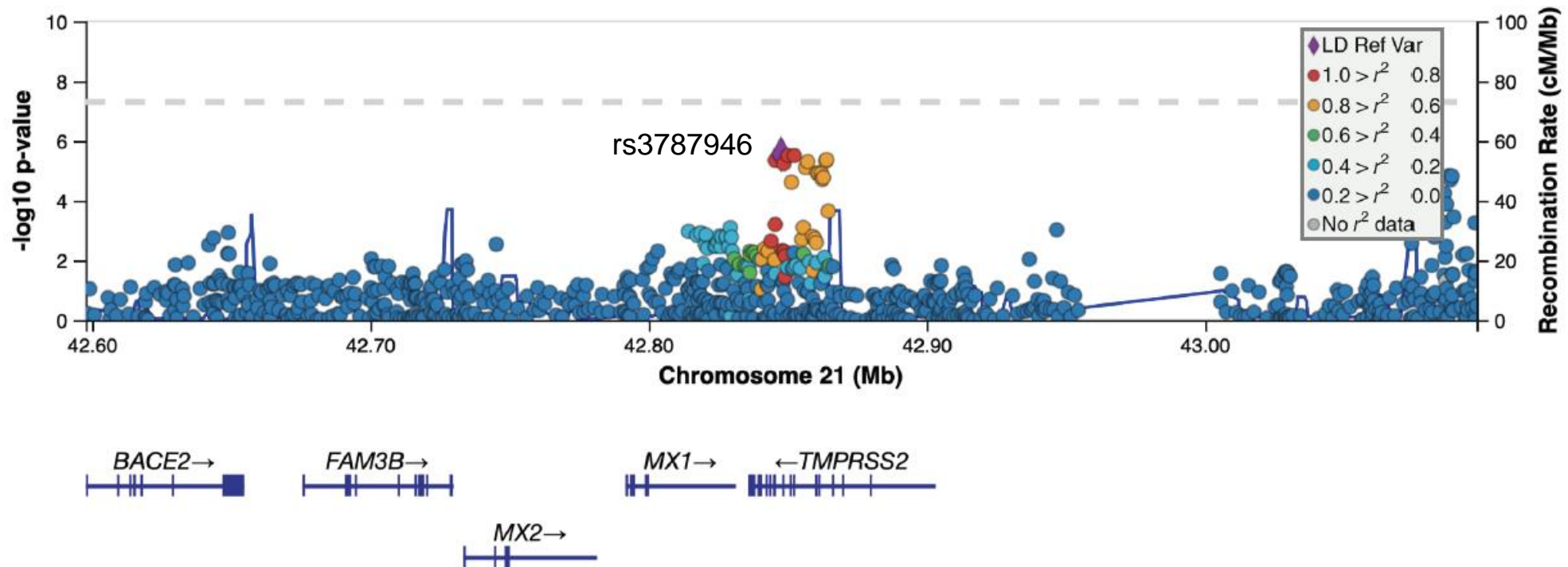
	SI cases n=226		SI controls n=1848		NIG cases n=770		NIG controls n=1915		All cases n=996		All controls n=3763		P _{SI}	OR (CI: 95%)	P _{NIG}	OR (CI: 95%)	P _{All}	OR (CI: 95%)
	n	%	n	%	n	%	n	%	n	%	n	%						
Genotype																		
CC	164	72.6	1274	68.9	532	69.1	1289	67.3	696	69.9	2563	68.1	-	-	-	-	-	-
CT	57	25.2	497	26.9	220	28.6	554	28.9	277	27.8	1051	27.9	0.47	0.89 (0.64-1.22)	0.68	0.96 (0.79-1.15)	0.71	0.97 (0.83-1.13)
TT	5	2.2	77	4.2	18	2.3	72	3.8	23	2.3	149	4.0	0.14	0.50 (0.20-1.26)	0.06	0.60 (0.35-1.02)	0.01	0.57 (0.36-0.89)
Allele																		
C	385	85.2	3045	82.4	1284	83.4	3132	81.8	1669	83.8	6177	82.1	-	-	-	-	-	-
T	67	14.8	651	17.6	256	16.6	698	18.2	323	16.2	1349	17.9	0.14	0.81 (0.62-1.07)	0.16	0.89 (0.76-1.04)	0.07	0.89 (0.78-1.01)
Dominant																		
CC/CT	221	97.8	1771	95.8	752	97.7	1843	96.2	973	97.7	3614	96.0	-	-	-	-	-	-
TT	5	2.2	77	4.2	18	2.3	72	3.8	23	2.3	149	4.0	0.15	0.52 (0.20-1.30)	0.06	0.61 (0.36-1.03)	0.01	0.57 (0.37-0.89)
Recessive																		
CC	159	70.4	1274	68.9	532	69.1	1289	67.3	691	69.4	2563	68.1	-	-	-	-	-	-
CT/TT	62	27.4	574	31.1	238	30.9	626	32.7	300	30.1	1200	31.9	0.26	0.84 (0.61-1.14)	0.37	0.92 (0.76-1.10)	0.28	0.92 (0.79-1.07)

NIG: Network for Italian Genomes

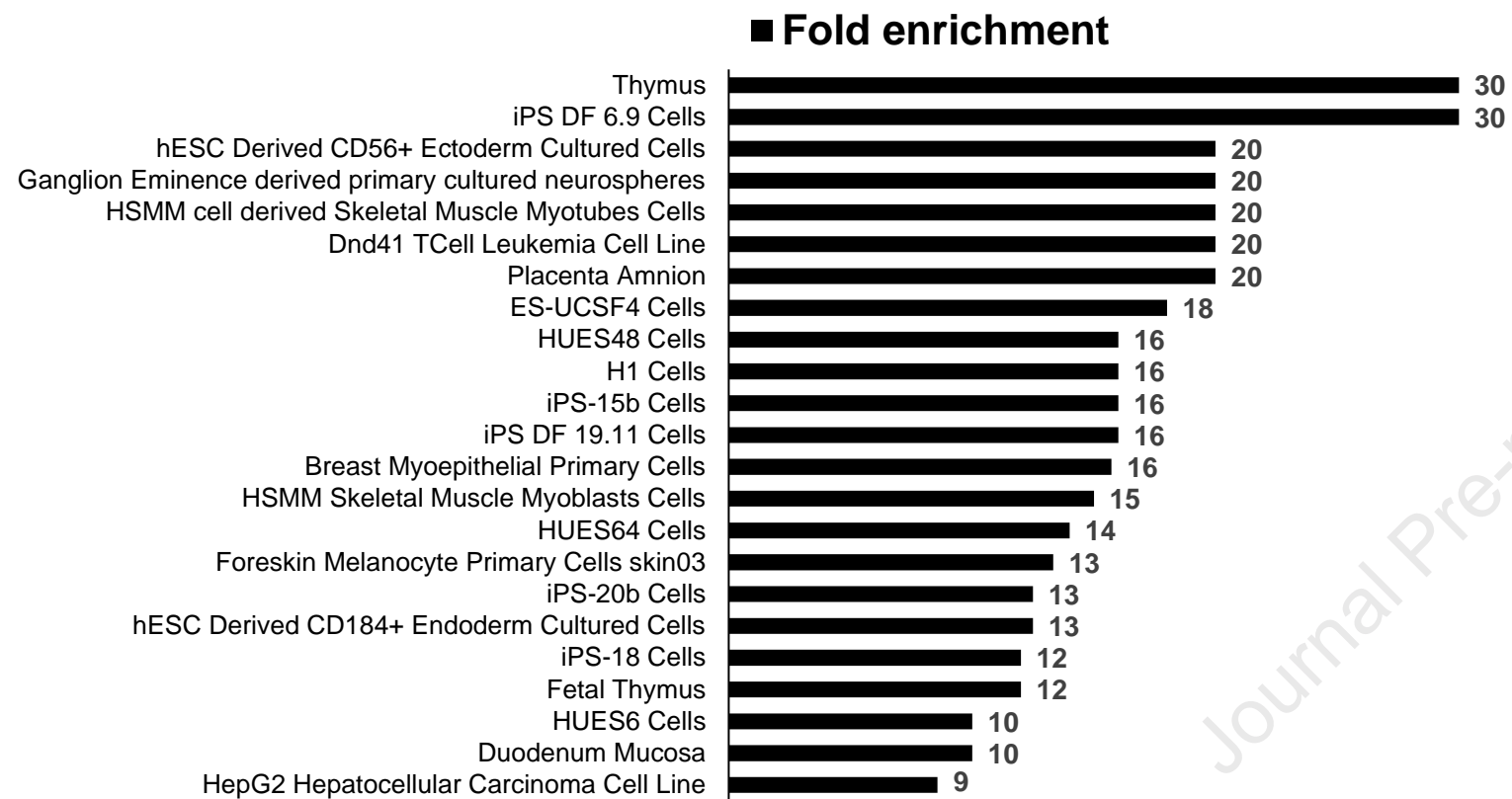
OR: odds ratio; CI: Confidence Interval

SI: Southern Italy

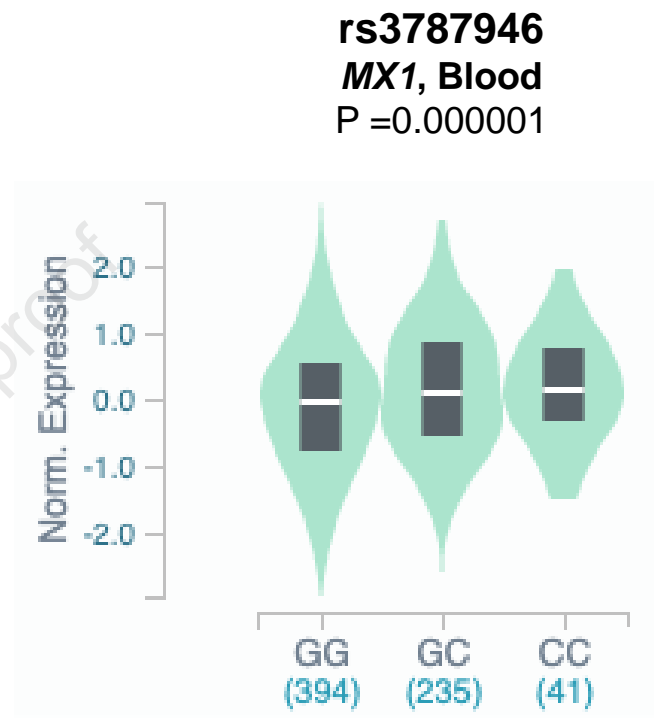
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a**b****c****Figure 1**

a



b



c

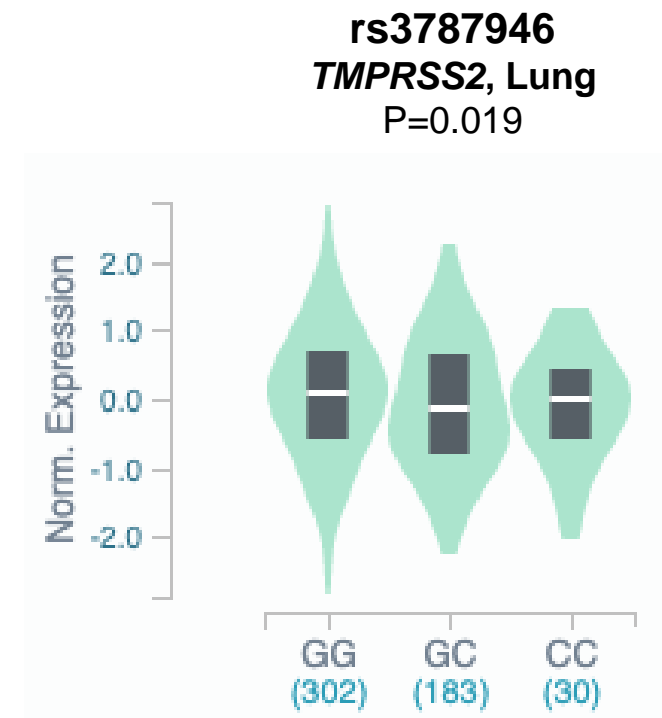


Figure 2

Common variants at 21q22.3 locus influence *MXI* and *TMPRSS2* gene expression and susceptibility to severe COVID-19

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Keywords: COVID-19, SARS-CoV-2, *TMPRSS2*, *MXI*, SNP genotyping.

Running title: Analysis of *TMPRSS2/MXI* locus in severe COVID-19.

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Highlights

- Genetic analysis was performed on 7,970 individuals hospitalized for COVID-19.
- Five SNPs within *TMPRSS2/MX1* locus (chr.21) are associated with severe COVID-19.
- The minor alleles of the five SNPs correlated with high level of *MX1* expression in blood.
- *MX1* could be a potential therapeutic target in patients with COVID-19.

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