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Fast surface disinfection with COUNTERFOG® SDR-F05A+

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Abstract COUNTERFOG® has been proposed as a rapid decontamination and disinfection technology that uses dynamic submicrometric-disinfecting fog cones. When projected onto surfaces, they create a micrometre-thick film of disinfectant minimizing the use of liquids and the impact on environment. The extremely thin film is intended to be enough to cover and kill microorganisms and simultaneously thin enough to evaporate in a few minutes—depending on the environmental conditions. In the present work, experimental tests were carried out to verify this hypothesis. These include a physical characterization of the fog in the cone, a measurement of the liquid flow projected on surfaces as well as disinfection tests with a series of microorganisms. In addition to these results, operational recommendations are derived to ensure disinfection reliability.

1 Introduction

Fomites and aerosols are two main transmission channels for many pathogens. Particularly, they are currently accepted as the main transmission mechanisms for COVID19. This virus has been proved to remain active for hours both on surfaces and in aerosols [1].

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Fig. 1 COUNTERFOG® dynamic fog cone

31 Therefore, a key for success when fighting against transmission of the pandemic is to
32 effectively disinfect both surfaces and air wherever people can be exposed to—including
33 buildings, vehicles, facilities, etc. [2].

34 COUNTERFOG® technology was recently developed¹ within the 7th Framework Pro-
35 gram for research and technological development (FP7) aiming to be a fast and universal
36 technology for decontamination of both surfaces and air from Chemical, Biological, Radio-
37 logical and Nuclear agents on a large scale [3].

38 This decontamination technology is based on dynamic fog cones composed of submicron-
39 sized liquid droplets that, when projected into the air, collapse air-borne CBRN agents posing
40 them down onto surfaces.

41 It was previously demonstrated that COUNTERFOG® cones made of biocidal liquid
42 projected on surfaces, covering them with a thin layer of biocide (a few microns thick)
43 disinfect *Bacillus Turinghiensis*. Biological agents are rapidly eliminated, saving significantly
44 in the use of biocide and in the recovery time of the operation of the place, whether the agents
45 are stuck or adhered to a surface, as if they are floating in the air or are easily resuspendable
46 [4] (Fig. 1).

47 COUNTERFOG® technology uses a specially designed nozzle fed by liquid—generally
48 water and solutions—and low-pressure compressed air. The portable device COUNTER-
49 FOG® SDR-F05A+ for rapid disinfection uses this technology to make a practical device
50 able to be operated just by a single person in a very simple way.

51 Suspended particles collapse with the fog nano-sized liquid droplets and are dragged by
52 the cone, posing on the ground or onto surfaces. This prevents the agents from continu-
53 ing to disperse, thus facilitating decontamination and generating minimal liquid waste and
54 consequent damage to the environment [5] (Fig. 2).

55 The physical principle that governs the operation of COUNTERFOG® technology is the
56 droplet and micro–nano-CBRN particles dynamics. When these substances are floating in
57 air, they are absolutely dragged by airflow. The smaller they are the fitter they behave. This
58 implies that they can only collapse or coalesce with droplets of liquid of a similar size—they

¹ Contract of the EU Commission n° 312804.

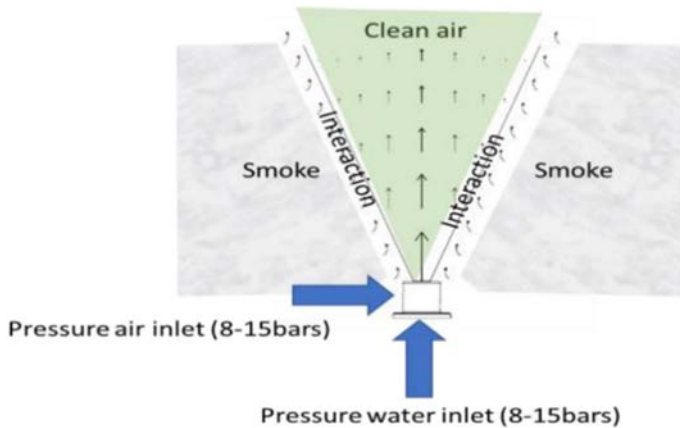


Fig. 2 Schematic representation of a COUNTERFOG® cone absorbing the air—in this case with smoke—around it

cannot collapse to droplets of a larger size, such as those produced by conventional sprinklers or sprayers [6–8].

In this way, airborne matter—or particles adhered to a surface—collapse with the COUNTERFOG® cone, making them to fall or detach. If the projected liquid is a biocide, the biological particles are neutralized as they collapse with the fog droplets. The layer of liquid formed after the application of the fog should be extremely thin, allowing its evaporation in a few minutes depending on the environmental conditions, guaranteeing the necessary time to disinfect said surfaces [9].

In this work, different tests have been carried out to physically characterize and verify the operation of the COUNTERFOG SDR-F05A+ device, as well as a set of biological tests for the neutralization of different pathogenic microorganisms with commercial disinfectants as shown below. These tests include the following: (1) COUNTERFOG® cone characterization, (2) projected micrometre film distribution and thickness, (3) liquid flow variation, (4) droplet distribution, and (5) biological tests (Fig. 3).

2 COUNTERFOG® SDR-F05A+ characterization

This section describes the tests and procedures to characterize the portable fast surface disinfection device COUNTERFOG® SDR-F05A+.

2.1 SDR-F05A+ physical characterization

The fog cone and operability of the COUNTERFOG® SDR-F05A+ device have been characterized through a series of physical tests and experimental measurements:

2.1.1 COUNTERFOG® cone geometrical and dynamic characterization

The objective of this test is measuring the absorbed airflow by the COUNTERFOG® cone; analyzing the velocity distribution of the air entering said cone; calculating the variation of the diameter and velocity of the fog cone along its horizontal axis.



Fig. 3 Operator using disinfection COUNTERFOG® SDR-F05A+ device

83 An SDR-F05A+ nozzle was horizontally fixed on a tripod. The very end of the nozzle was
84 taken as origin of coordinates. A digital anemometer *Kestrel 3000* was used to measure fog
85 velocity along its horizontal axis as well as at different peripheral points around the axis of
86 the fog cone. The criteria to define the border of the cone was based on the axial component
87 of the velocity. Points with axial velocity under 0.1 m/s are considered out of the cone.

88 The total flow of external air dragged by the cone was calculated by integration all over
89 the border of the cone.

90 *2.1.2 Projected micrometre film distribution and thickness*

91 The objective of this experimental test is to measure the aggregated liquid fog distribu-
92 tion projected on a flat surface—ml/min cm²—and its film thickness (μm) generated by a
93 COUNTERFOG® nozzle.

94 Small cups were provided onto a vertical surface in order to collect the liquid projected
95 onto 100 mm \times 100 mm patches. Total weight of liquid collected at those patches were
96 measured. This test was repeated for different distances ranging from 1 to 3 m, the projected
97 fluid flow was varied by opening the water stopcock valve in proportional angles to 45°.

98 *2.1.3 Liquid flow variation*

99 The third physical test was realized with the objective of analyzing the quantity of liquid
100 projected by the cone as a function of the opening of its corresponding liquid stopcock valve.

101 It was evaluated the time it took to empty the 12-L tank—pressurized with the 11 bar—
102 varying in each projection the opening of the water stopcock valve in proportional angles to
103 45°. As the previous tests, it was repeated a statistical number of times.

Table 1 Biological tests results

Test	Microorganism	Disinfectant	Application velocity (m/s)	Application time (min)	Film thickness (μm)	Reduction (Log_{10})	Reduction (%)
E1	<i>Escherichia coli</i>	0.6% NaClO	0.45	1	21.8	2	99.00
E2	<i>Candida albicans</i>	1% NaClO	0.36	1	27.3	3	99.90
E3	<i>Proteus hauseri</i>	1% NaClO	0.16	6	61.53	3	99.90
E4	<i>Enterococcus hirae</i>	1% NaClO	0.15	1	65.64	5	99.99
E5	<i>Pseudomonas aeruginosa</i>	1% NaClO	0.37	6	26.60	3	99.90
E6	<i>Staphylococcus aureus</i>	1% NaClO	0.21	1	46.88	0.99	93.00
E7	<i>fago phi29</i>	1% NaClO	0.57	1	17.27	3	99.90

104 2.1.4 Droplet distribution

105 The verification of the previously calculated—nanometric—size of the COUNTERFOG®
106 cone droplets, were carried out by experimental measurements using an optical emission
107 spectrometer at Instituto Nacional de Técnica Aeroespacial (INTA).

108 2.2 SDR-F0A+ biological characterization

109 2.2.1 Biological tests

110 Biological tests to verify the effectiveness of the COUNTERFOG® SDR-F05A+ device
111 against different pathogenic microorganisms have been carried out. Bacteria—*Escherichia*
112 *coli*, *Proteus hauseri*, *Enterococcus hirae*, *Pseudomonas aeruginosa*, *Staphylococcus*
113 *aureus*—, fungi—*Candida albicans*—, and viruses—*fago phi29*—were disinfected.

114 The objective of the biological tests consisted in assessing the minimum quantity of
115 different commercial disinfectants necessary to apply, defining for each microorganism the
116 application velocity—sweep—of the projected cone by an operator to neutralize each type
117 of microorganism—obtaining the corresponding film thickness (μm).

118 Biological tests were realized using solutions of the different pathogenic microorganisms
119 on stainless steel coupons, in accordance with ISO-UNE standards. There were performed in
120 a closed room, projecting the cone at a distance of 2 m—from the nozzle to the impact area
121 where the coupons were located—by means of a horizontal sweep, at the velocities specified
122 in Table 1. *Biological tests results*.

123 Colony-forming units—cfu—in the control coupons without any disinfection were com-
124 pared in a laboratory with the disinfected samples, thus obtaining logarithmic and percentage
125 reductions of the cfu.

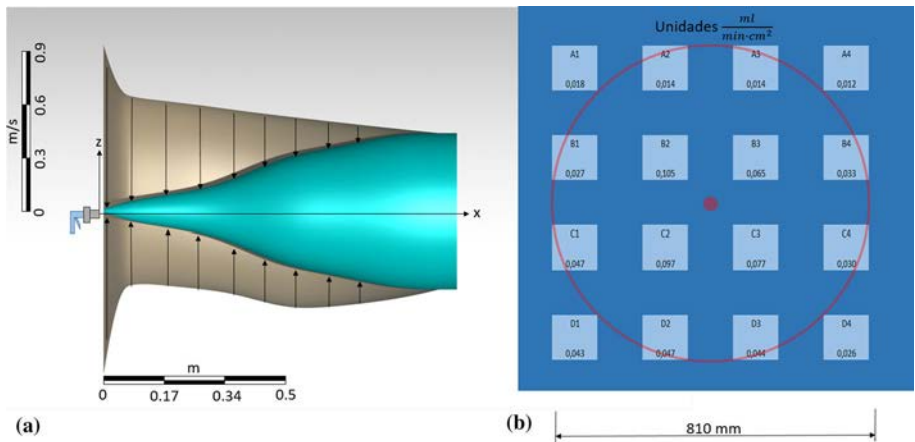


Fig. 4 **a** Distribution of the airflow drawn in by the fog cone (m/s). **b** Distribution of the projected droplets by the fog cone at 2 m (ml/(min-cm²))

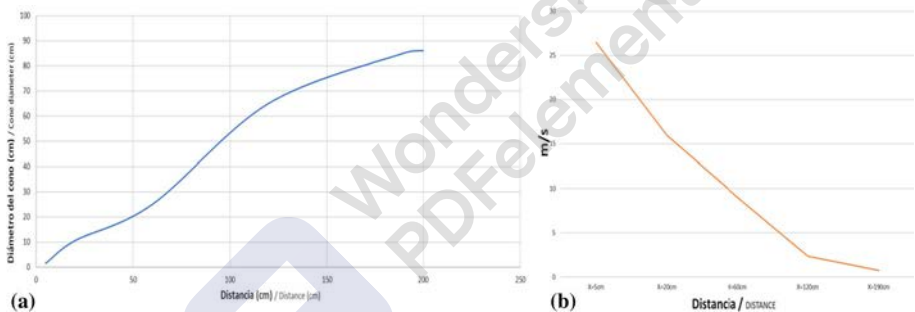


Fig. 5 **a** Cone diameter with respect to the distance from the end of the nozzle. **b** Velocity in the axis of the cone with respect to the distance from the end of the nozzle

126 3 Results

127 This section includes the results of the aforementioned tests.

128 3.1 SDR-F05A+ physical characterization results

129 3.1.1 COUNTERFOG® cone characterization results

130 The airflow COUNTERFOG® cone absorbs 0.3 m³/s air from around with an average velocity
 131 of the air drawn into the cone of 0.3 m/s and—with a maximum airflow of 0.9 m/s. Figure 4a
 132 shows the velocity distribution of the absorbed airflow in the x-z plane.

133 Figure 5a shows the evolution of the cone diameter—cm—with respect to the distance
 134 from the end of the nozzle—as the origin of coordinates. Figure 5b the velocity along the
 135 horizontal axis of the cone with respect to the previously mentioned distance—m/s.

D_{10} particles (μm)	D_{90} particles (μm)	% emission
0,02	0,03	47,86
0,03	0,04	85,57
0,05	0,07	76,46
0,10	0,15	47,53
0,22	0,26	23,50
0,32	0,43	12,77
0,59	0,73	4,91
0,91	1,17	1,37
1,52	1,86	0,23
2,28	2,95	0,07
3,81	4,92	0,03
6,37	7,76	0,02

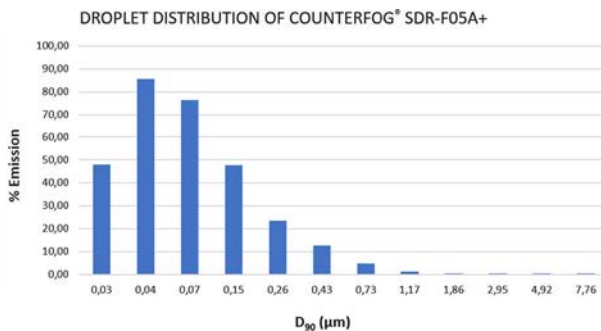


Fig. 6 Droplet distribution of COUNTERFOG® SDR-F05A+

136 3.1.2 Projected micrometre film distribution and thickness results

137 The projected fog distribution— $\text{ml}/\text{min cm}^2$ —in a flat surface has been assessed by a series
 138 of fluid projections, varying the opening of the water stopcock valve and the distance from
 139 the impact surface to the nozzle. Figure 4b shows the results of a projection carried at 2.0 m
 140 from the end of the COUNTERFOG® nozzle to the impact area, with the water stopcock
 141 valve opened 3/8 turn.

142 The flow ranges from 0.1 to 0.01 $\text{ml}/(\text{min cm}^2)$. If for instance the user moves the nozzle
 143 horizontally at 0.45 m/s, an average of 21.8 μm can be easily calculated.

144 3.1.3 Liquid flow variation results

145 The liquid mass flow projected by the cone as a function of the opening of its corresponding
 146 liquid stopcock valve has been evaluated.

147 It was observed a constant maximum projected liquid flow of 0.65 l/min when there was
 148 a valve opening greater than or equal to 3/8 turn.

149 3.1.4 Droplet distribution results

150 Droplet size distribution of COUNTERFOG® SDR-F05A+ nozzle was analyzed by experi-
 151 mental measurements in Instituto Nacional de Técnica Aeroespacial (INTA). The results are
 152 shown in Fig. 6, thus verifying the previously calculated nanometric size.

153 3.2 SDR-F0A+ biological characterization results

154 3.2.1 Biological tests results

155 Biological disinfection results show an average reduction of three orders of magnitude for
 156 the different pathogenic microorganisms assessed.

157 The following Table 1 shows the disinfectant applied for killing each pathogenic
 158 microorganism with the correspondent solution, application velocity, and time, film
 159 thickness— μm —of the projected fluid layer and the total reduction of the biological microor-
 160 ganisms—logarithmic and percentage.

161 4 Conclusions

162 COUNTERFOG® SDR-F05A+ portable device has been demonstrated to provide a fog
163 cone made of nanometric to micrometric droplets. This cone absorbs 0.3 m³/s of air from the
164 environment and projects a few micron thick liquid film over a 1 m² surface in a typical time
165 of 1–3 s. Last, with these rates of application the equipment has shown 2–3 log reduction
166 disinfection of a number of microorganisms including fungi, bacteria and virus.

167 **Data Availability Statement** Experimental test and previous researches.

168 **Funding** COUNTERFOG EBT DE LA UAH S.L.

169 Declarations

170 **Conflict of interest** The authors declare that they have no conflict of interest.

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