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Full-field strain distribution in multi-vertebra spine segments:

An *in vitro* application of Digital Image Correlation

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Abstract

The biomechanics of the spine is experimentally assessed in terms of range of motion and overall stiffness. Quantification of the surface strain distribution is currently limited either to the vertebrae or the discs, whereas a full-field approach to measure the strain distribution in a multi-vertebra segment is currently missing. The aim of this work was to explore the feasibility of using Digital Image Correlation (DIC) to measure the strain distribution simultaneously on the vertebral bodies and the intervertebral discs of spine segments in different loading configurations.

Three porcine spine segments were tested. A white-on-black speckle pattern was prepared which covered the hard and soft tissues. Two different loading configurations (flexion and lateral bending) were reproduced, while two sides of the spine were analyzed with DIC.

Measurements were successfully performed on the entire region of interest of all specimens, in both configurations. The DIC analysis highlighted the strain gradients present on the spine segments including tension and compression associated with bending, the direction of principal strains in the different regions, as well as bulging of the discs under compression. Strains of tens of thousands microstrain were measured in the discs, and below 2000 microstrain in the bone.

This work showed the feasibility of applying DIC on spine segments including hard and soft tissues. It also highlights the need for a full-field investigation, because of the strain inhomogeneity in the vertebrae and discs.

Keywords: Biomechanics; Spine segment; Full-field strain measurement with Digital Image Correlation; Feasibility study; Intervertebral Discs; Vertebrae.

1 **1. Introduction**

2 The spine consists in a sequence of hard (vertebrae) and soft tissue
3 (intervertebral discs), stabilized by ligaments. Spine characterization is a
4 fundamental challenge in biomechanics because it could help engineers and
5 clinicians to better understand physiological/pathological conditions, and to
6 design better implants [1, 2].

7 Spine segments have often been investigated *in vitro* [3-7]. In such experiments
8 either known motions are imposed to measure the structural stiffness [8-10], or
9 known loads are applied to measure the range of motion[11-13]. From these
10 tests, structural properties and behaviour of the spine segment as a whole is
11 assessed. Only in few cases the local strain distribution has been measured.
12 However, such experiments focused separately either on the vertebra or on the
13 intervertebral disc. Strain in the vertebra has been measured by means of strain
14 gauges [14-17]; this provided accurate measurements, but was limited to the
15 area of application of the strain gauges. The reinforcement effect of strain
16 gauges can be significant (up to 9%) [14, 18, 19]. Measuring the local strain in
17 the intervertebral disc is even more difficult, due to the intrinsic nature of the
18 disc itself (low stiffness). One of the first measurements of the strain on the
19 outer part of the disc (*annulus fibrosus*) was based on stereo-photogrammetry
20 and covered a limited portion of the disc [20]. More recently, the entire disc
21 surface was investigated (excluding the adjacent vertebrae), using digital image
22 correlation (DIC) [21]. A method to explore the strain field on spine segments
23 with large in-plane and out-of-plane deformations is missing.

24 The aim of this work was to explore the feasibility of using DIC to measure the
25 full-field strain distribution simultaneously on the vertebral bodies and
26 intervertebral discs of thoracolumbar spine segments in different loading
27 configurations. Specifically, the following research questions were addressed:

- 28 • The feasibility of applying a speckle pattern with sufficient contrast for
29 the correlation algorithm, that should not become damaged when the
30 specimen was kept hydrated;

- 31 • The robustness of the pattern to withstand the large deformations
32 expected in the soft tissue (up to 100000 microstrain) for up to 50 cycles;
- 33 • The procedure must allow measuring the small deformations expected in
34 bone (of the order of 2000 microstrain) with acceptable errors (not
35 exceeding 200 microstrain);
- 36 • The possibility of measuring strains in specimens undergoing relatively
37 large in-plane (up to 3 mm) and out-of-plane (7 mm) displacements,
38 while correlating at least 90% of the region of interest.

39 **2. Material and methods**

40 ***2.1 Specimen and pattern preparation***

41 For ethical reasons, in this methodological work porcine spine segments from
42 the slaughterhouse were preferred. The three animals were female, of the same
43 breed, approximately 9 months old and approximately 100 kg at sacrifice. The
44 segments consisted of four thoracolumbar vertebrae (T7-T10/T11-T14/L2-L5)
45 selected for resemblance in terms of ranges of motion and dimensions [22-24].
46 The specimens were kept hydrated, and stored at -28°C to avoid alteration of the
47 mechanical properties.

48 The ribs, the muscles, the anterior longitudinal ligament, and the periosteum
49 were carefully removed using surgical tools. The interspinous, supraspinous and
50 posterior longitudinal ligaments, and the capsules were left intact in order to
51 preserve the natural kinematics. The specimens were aligned so that the middle
52 disc of each segment was aligned horizontally in the frontal and lateral views
53 [25]. In this configuration, two pots of poly-methyl-methacrylate were created
54 parallel to one another, where the upper half of the most cranial vertebra and the
55 lower half of the most caudal vertebra, were embedded (Fig. 1).

56 In order to use DIC to make measurement on the spine segments (both the
57 vertebrae and the intervertebral discs), a high-contrast white-on-black speckle
58 pattern [26, 27] was prepared. The spine segments were first stained with a
59 dark background (a solution of 4 g of methylene-blue per 100 ml of water) [28,
60 29]. Two applications were required for the intervertebral discs and three for

61 the vertebrae to obtain a uniform background. Dying was preferred, rather than
62 surface painting, as a layer of paint would crumble and crack due to the large
63 deformations of the soft tissues.

64 The speckle pattern was applied with paint instead of using a powder, as was
65 done previously [26]. This allows avoiding artifacts as the dots can strain during
66 the test (unlike powder grains). Furthermore, paint dots resist to saline solution
67 without damage, allowing specimen hydration and storage. A white water-based
68 paint (Q250201 Bianco Opaco, Chrèon, Italy) was diluted at 40% with water and
69 sprayed using an airbrush-airgun (AZ3 HTE 2, nozzle 1.8 mm, Antes Iwata, Italy)
70 to prepare a white speckle pattern with the qualitative and quantitative
71 requirements described in [30]. The spraying distance (300 mm) and the
72 pressure (1000 kPa) were optimized to obtain the desired dot size, and minimize
73 the scatter of dot dimension [35]. The actual size of the speckle dots was
74 measured in the 2D digital images (acquired by DIC system) of the ROI through a
75 custom script developed for this work. The speckles had a dimension of 6 pixels
76 (average 0.18 mm with a standard deviation of 0.18 mm), larger than the
77 minimum recommended size while the facet were sufficiently large compared to
78 the speckle size [30].

79 The specimens were soaked in saline solution before the tests.

80 ***2.2 Mechanical Testing***

81 The specimens were loaded using a uniaxial servo-hydraulic testing machine
82 (8032, Instron, UK) in displacement control. In order to avoid transmission of
83 any undesired component of load, free rotation of the loading plate was allowed
84 by means of a ball joint, while free horizontal translations were granted by
85 means of two low-friction orthogonal linear bearings.

86 In order to assess the feasibility of measuring strains on such complex
87 specimens, two different loading configurations were simulated, which are
88 frequently investigated in the literature [31] (Fig. 2):

- 89 • Flexion: the vertical force had an anterior offset equal to the 20% of the
90 antero-posterior depth of the central intervertebral disc;

- 91 • Lateral bending: the vertical force had a lateral offset (both towards right
92 and left) equal to the 20% of the coronal width of the central
93 intervertebral disc.

94 Ten preconditioning cycles were applied between 0 and 1.0 mm of compression,
95 at 0.5 Hz. A compression of 3.0 mm was applied for each loading configuration in
96 0.1 mm steps, while DIC images were acquired at each step (see below). This
97 value of compression was based on preliminary tests to reach typical strains
98 associate to physiological loads [32, 33] to prevent damage of the specimens.
99 These loading configurations did not aim to mimic any specific motor task.

100 ***2.3 Digital Image Correlation***

101 A commercial 3D-DIC system (Q400, Dantec Dynamics, Denmark) with
102 directional custom designed arrays of LEDs (10000 lumen in total) was used.
103 Images were acquired by two cameras (5MegaPixels, 2440x2050 pixels, 8-bit)
104 equipped with high-quality metrology-standard 35 mm lens (Apo-Xenoplan
105 1.8/35, Schneider-Kreuznach, Germany; 135 mm equivalent) for a stereoscopic
106 view. Due to the curvature of the vertebral bodies and discs, the cameras were
107 positioned at a distance of 260 mm from the specimen and vertically to maximize
108 the area framed by both cameras (Fig. 2).

109 The field of view was set to 70 mm by 60 mm (resulting in a pixel size of 28
110 micrometers), with depth of field of 20 mm (lens aperture f/16). The field of
111 view was wide enough to frame the entire region of interest (ROI): the central
112 intervertebral disc and the two adjacent vertebrae.

113 To explore the possibility of assessing the different sides of the spine, two
114 different acquisitions were performed for each loading configuration and each
115 specimen:

- 116 • Frontal view: the cameras pointed the anterior wall of the spine segment;
117 • Lateral view: the cameras pointed the lateral side (either right or left) of
118 the spine segment.

119 During the mechanical tests, series of images were acquired starting from the
120 unloaded condition (reference step) and every 0.1 mm step of compression.

121 The correlation was performed with Istra-4D (v.4.3.1, Dantec Dynamics,
122 Denmark) using the following parameters (Fig. 1):

123 a) Facet size: 33 pixels;

124 b) Grid spacing: 19 pixels;

125 c) Contour smoothing: local regression with a kernel size of 5x5;

126 It is not possible to exactly quantify the measurement spatial resolution due to
127 the nonlinearity of the smoothing process. With the settings used, a spatial
128 resolution better than 3 mm was estimated (comparable with the typical
129 dimension of the strain gauges used in biomechanical field [14]).

130 Finally, the maximum (ε_1) and minimum (ε_2) principal strains were evaluated.

131 ***2.4 Optimization and error analysis***

132 As DIC relies on a complex preparation, acquisition and analysis, its accuracy and
133 precision cannot be taken from granted. We capitalized on past experience on
134 optimization of pattern preparation [28] [35], image acquisition and correlation
135 parameters [34] (see 2.1 and 2.3).

136 Before each session, calibration was performed using a proprietary calibration
137 target (Mod. Al4-BMB-9x9, Dantec Dynamics): 3D-residuum values from 0.031 to
138 0.173. It is a proprietary procedure and it consists in acquiring a series of images
139 of the target calibration in different orientations to define the physical
140 dimensions of the measurement volume, the correction of the distortions due to
141 lens and the compensation of the parallax effects.

142 An optimization and validation of the system was preliminarily performed [34]
143 to find the best compromise between the uncertainties and DIC parameters. A
144 pair of images of the unloaded segments were captured for each specimen and
145 analyzed with the optimal hardware and software settings in order to assess the
146 measurement uncertainties in a known configuration (zero-strain). Being in a
147 zero-strain configuration, any strain different from zero was accounted as
148 measurement error. The Kolmogorov-Smirnov test was used to check that the
149 errors followed a Gaussian distribution. The systematic and random errors [34]
150 were computed for each specimen and for each view as the average and the
151 standard deviation of ε_1 and ε_2 .

152 **3. Results**

153 Measurements were successfully performed on all specimens, over the entire
154 region of interest in both the frontal and sagittal view, for both loading
155 configurations, during the entire tests. Similar strain distributions were found in
156 the three specimens. The different tissues were easily identified observing the
157 physical specimens and the images acquired by the cameras. The evolution of
158 the strain distribution during load application (Fig. 3) is visible in the videos in
159 the Supplementary Material.

160 ***3.1 Strain error***

161 In the zero-strain test, the apparent strains followed a Gaussian distribution
162 (Kolmogorov-Smirnov). As expected, the largest component of error was the
163 random error, and it never exceeded 190 microstrain.

164 ***3.2 Flexion – Frontal view***

165 During flexion, at most 4.4% (worst specimen) of the ROI in the frontal view lost
166 correlation between the first and the last step. The discs underwent larger
167 deformations compared to the vertebral bone (Fig. 4). The strain in the discs
168 followed a strain gradient, with peaks in the central portion. In the discs, ε_1 was
169 on the order of +20000 microstrain (aligned circumferentially) and ε_2 was of the
170 order of -40000 microstrain (aligned axially).

171 The vertebrae had lower strains: peak on the order of +500 microstrain for the ε_1
172 and around -1500 microstrain for ε_2 . The two regions cranial and caudal to the
173 intervertebral discs with larger strain show the large deformation of the growth
174 plates.

175 ***3.3 Flexion – lateral view***

176 During flexion, at most 5.7% (worst specimen) of the ROI in the lateral view lost
177 correlation between the first and the last step. The discs presented a strain
178 gradient: they were more deformed in tension in the posterior than in the
179 anterior side (Fig. 4). ε_1 on the intervertebral discs varied between +12000

180 microstrain (anterior side) and +87000 microstrain (posterior side). ε_2 followed
181 a similar gradient, from -87000 microstrain (anterior) to -18000 microstrain
182 (posterior). The direction of ε_1 changed from circumferential (anterior side), to
183 axial (posterior). Therefore, in flexion the discs tended to bulge in the anterior
184 part, and to stretch in the posterior side.

185 The vertebral body experienced lower strains: below +600 microstrain for ε_1 and
186 -1500 microstrain for ε_2 .

187 ***3.4 Lateral bending – frontal view***

188 During lateral bending, at most 1.5% (worst specimen) of the ROI in the frontal
189 view lost correlation between the first and the last step. Lateral bending was
190 applied both towards the right and the left sides, to all specimens: the outputs, in
191 terms of strain distribution on intervertebral discs and vertebrae were
192 symmetrical. The discs showed tension and compression respectively on the left
193 and right sides, depending on the direction of the imposed bending (Fig. 4). ε_1
194 varied between +6000 microstrain (compressed side) and +143000 microstrain
195 (stretched side). ε_2 were varied from -85000 microstrain (compressed side) to
196 +12000 microstrain (stretched side). The orientation of ε_1 changed from
197 circumferential on the compressed side, to axial on the stretched side.

198 The vertebrae had lower strains than the discs: ε_1 were lower than 500
199 microstrain and ε_2 did not exceed -1700 microstrain. In this configuration, as in
200 flexion, it was possible to observe both tension (ε_1 aligned with the vertebral
201 axis) and compression (ε_2 circumferentially) in the growth plates.

202 ***3.5 Lateral bending – lateral view***

203 During lateral bending, at most 5.2% (worst specimen) of the ROI in the lateral
204 view lost correlation between the first and the last step. The strains on the discs
205 were rather uniformly distributed compared with the other tests (Fig.4). When
206 the side in compression was observed, ε_1 in the disc were of the order of +40000
207 microstrain, and ε_2 of the order of -60000 microstrain.

208 The vertebral body had lower strains than the discs: ε_1 were lower than 700
209 microstrain and the ε_2 did not exceed -1400 microstrain.

210 **4. Discussion**

211 The main aim of this work was to develop a procedure to measure the strain
212 distribution on thoracolumbar spine segments [36] simultaneously on hard
213 tissues (the vertebrae) and soft tissues (the intervertebral discs) by means 3D-
214 DIC [30]. Two different loading configurations frequently found in the literature
215 [6, 11-13] were explored. The specimens were observed both frontally and
216 laterally.

217 The results of this work showed the technical feasibility of quantifying the full-
218 field strain distribution (small and large deformations) over multi-vertebra spine
219 segments during the mechanical tests. The measured strain distributions proved
220 to be highly inhomogeneous, confirming the importance of investigating the
221 spine using a full-field tool to complement the evaluation of the range of motion
222 and stiffness performed in the past.

223 The correlation was satisfactory throughout the tests, even if large deformations
224 and displacements were involved. The correlated surface covered the ROI, with
225 a loss of correlation lower than 5.7% of the initial correlated surface. Generally,
226 the noise originating from the camera sensors, sub-optimal speckle dots, poor
227 illumination, and bad speckle framing can contribute to loss of correlation. Here,
228 the leakage of fluids from the vertebral body under load, which is an inherent
229 phenomenon when testing fresh tissue, increased the loss of correlation.
230 Additionally, the out-of-plane movements (7 mm) of the postero-lateral elements
231 (capsule) caused some subsets to be out-of-focus in the lateral view. While DIC
232 often has some un-correlated gaps, other measurement techniques such as strain
233 gauges simply measure strains in selected points, and ignore the rest of the
234 surface.

235 The prepared white-on-black pattern confirmed its suitability both for hard and
236 soft tissues [28]. In fact, the background, prepared with methylene-blue, did not

237 crack during the tests, while the white dots remained sharp and adherent to the
238 specimen's surface.

239 A compromise was sought between reduction of measurement uncertainty and
240 spatial resolution. The hardware (camera resolution, lenses, field of view) and
241 software parameters (facet size, grid spacing, contour smoothing) provided a
242 measurement spatial resolution better than 3 mm. This is comparable with the
243 grid length of the strain gauges typically used in these applications (1-5 mm)
244 [37-40]. The strain measurement uncertainties (below 180 microstrain) were
245 acceptable for biomechanical purposes. Such uncertainties would not prevent
246 detecting failure of the bone (around 10000 microstrain [32]), as well as strain
247 associated to physiological loads (1000-2000 microstrain [41]). The same
248 considerations are confirmed for the intervertebral discs: as the failure strain is
249 around 250000 microstrain [20], and the physiological strain is below 150000
250 microstrain [33].

251 The evaluated strains confirmed the expected trends in all performed tests:
252 larger strains were measured in the intervertebral discs (in the order tens
253 thousands microstrain), lower strains in the vertebrae (below 2000 microstrain).
254 Such strain gradients are consistent with the expected biomechanics of the spine
255 [4]. DIC discriminated the portion of the discs subjected to
256 tension/compression, with the expected orientation of the principal strains. In
257 the compressed side of the discs the minimum principal strains were axial, as
258 expected; the maximum principal strains were larger in absolute value than the
259 minimum ones, and had a transversal orientation.

260 The combination of the prepared pattern, selected software parameters, and
261 hardware allowed recognizing the deformation on the growth plates of the
262 vertebrae. Because of the spine specimens came from skeletally immature
263 animals [42] the growth plates were not closed and therefore were subjected to
264 larger deformation compared to the vertebral body. This was serendipity and
265 would otherwise be impossible to observe with traditional surface measurement
266 technique such as strain gauges.

267 Usually, *in vitro* spine segments have been investigated in terms of range of
268 intersegmental motion [6, 12, 13, 43] and overall stiffness [11, 13, 44, 45]. In
269 other works, the spine was investigated measuring local deformation either in
270 the vertebrae or in the intervertebral discs. In the vertebrae, strains have been
271 evaluated using strain gauges, which provide point-wise measurements. At the
272 same time, strain gauges do cause reinforcement, especially on thin osteoporotic
273 tissues [14, 15]. Otherwise, the vertebrae have been studied using DIC but
274 without evaluating the contiguous discs [34, 46]. Conventional strain gauges
275 cannot be used on the discs due to their low elastic modulus. Strain
276 measurement is possible with optical methods [20, 21]. In the work of Stokes *et*
277 *al.* [20] the deformation on a small portion of the intervertebral disc was
278 evaluated by photogrammetry adhering small optical markers. Heuer *et al.* [47]
279 developed a method based on laser scanning device to evaluate the strain
280 distribution on intervertebral discs. In this case, the strain uncertainty (<1600
281 microstrain) would not allow measuring strain on hard tissue as vertebra.
282 Moreover, the duration of the scan (4 seconds), constrains the applicability to
283 quasi-static applications. In the work of Spera *et al.* [21] strain was measured
284 only on the discs using DIC. They used coffee powder: as the coffee grains
285 themselves are not deformable as the paint-dots, this can result in large noise,
286 apparent strain and loss of correlation.

287 A work [48] focused on the strain distribution on spine segments. High-speed
288 3D-DIC was used to evaluate the strains on the anterior side of porcine cervical
289 spine segments in axial impacts. In contrast, the present work aims to explore
290 the feasibility of the measurement under large deformation, directed out-of-
291 plane.

292 A limitation of this work could be the use of young porcine spine instead of
293 human spine. This choice was motivated by ethical considerations. The porcine
294 spines are different from the human ones but are valid models [22, 24] to
295 demonstrate the feasibility of this novel approach.

296 In this work, a quite standard but relatively simple configuration of 3D-DIC was
297 used. Array of cameras can be used to acquire 360° of the whole spine in order
298 to obtain a complete description of the strain maps.

299 The applied loading configurations were not intended to replicate any specific
300 physiological motor task, but included separate components of load which are
301 present in physiological motor tasks (flexion and lateral bending) and which are
302 typically found in spine biomechanics [31] as main causes of spine degeneration
303 [49]. This study did not cover other types of loading such as torsion.

304 The strains were evaluated only on the surface of the specimens. Digital Volume
305 Correlation (DVC) [50] has the potentiality to evaluate the strain within the
306 vertebral bone [51-53]. Nevertheless, applicability of DVC is limited due to the
307 slow tomographic image acquisition.

308 **5. Conclusion**

309 This work showed that by applying an adequate speckle pattern and selecting
310 optimal DIC parameters, the full-field strain distribution could be evaluated on a
311 complex structure (soft and hard tissues) such as a spine segment. This full-field
312 measurement would allow assessing the strain distribution, determining the
313 alignment of principal strains, and identify stress concentrators. This opens the
314 way to a new method of investigation of the spine, involving new possible
315 applications for basic biomechanics research and clinical innovations. DIC-based
316 experimental measurements could be combined with numerical models in an
317 inverse approach to measure (identify) the local tissue properties, including
318 anisotropy. Furthermore, the use of DIC will help elucidate the damage and
319 failure mechanism of the spine. DIC can also be used to investigate and improve
320 different surgical techniques and spine fixation devices.

321 **Conflict of Interest Statement**

322 None

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328 **Competing interests:**

329 None declared

330 **Funding:**

331 None

332 **Ethical approval:**

333 Tissue specimens (porcine spines) were obtained as a side-product through the
334 food chain from animals that have been bred and sacrificed for alimentation
335 purposes. No animal was specifically bred or sacrificed for this study. The
336 Biomechanics laboratory at UNIBO meets all the Italian regulations concerning
337 research, safety in the working environment, handling, storing, testing and
338 disposal of animal and human tissue samples. All the research was carried out in
339 accordance with the National and local regulations guidelines and, in accordance
340 with the EU Directive 2010/63/EU.

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Captions and figures (images here are in low resolution)

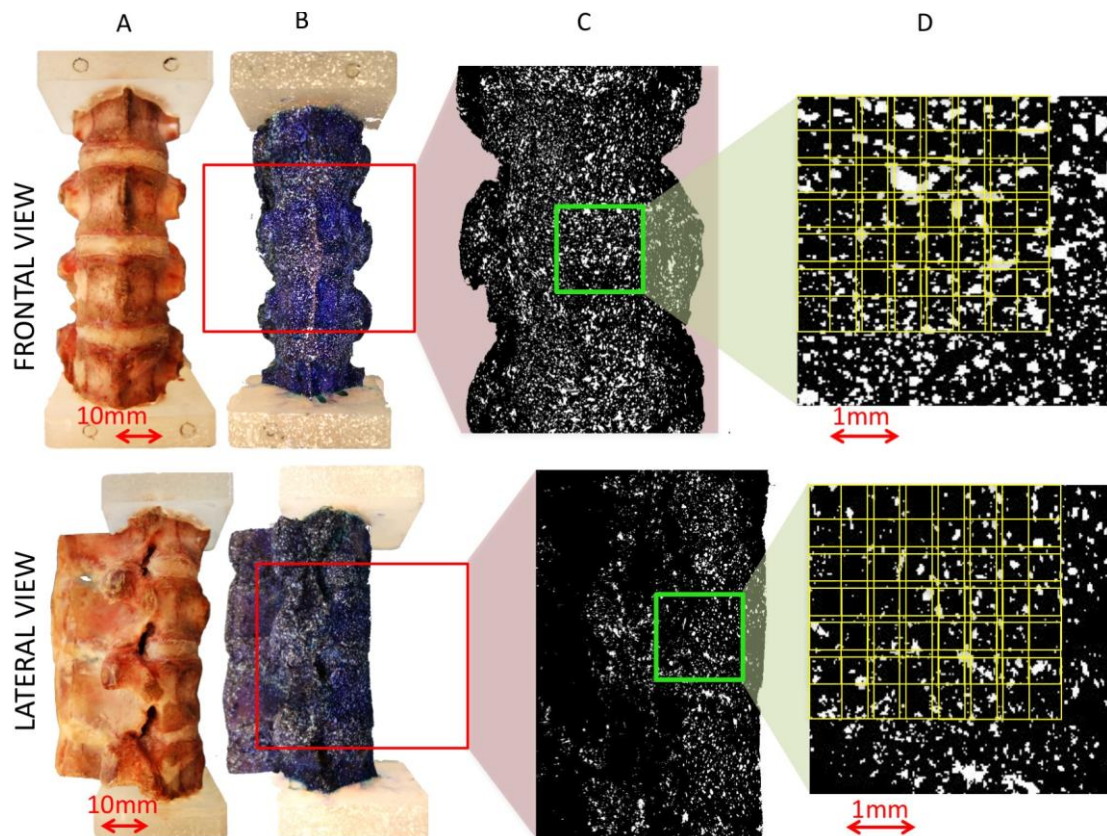


Fig. 1 - Overview at different dimensional scales of specimen preparation and analysis. The porcine spines were cleaned removing the surrounding tissues (A), and preparing the white-on-black speckle pattern (B); the red windows represent the field of view recorded by the DIC cameras. Binarized images of the entire region of interest (which covered the central disc and the two adjacent vertebrae) were generated to evaluate the dimensions of the white speckle dots (C). The facet size and grid spacing (yellow lines) can be compared to the speckle dot size in the binarized images (D).

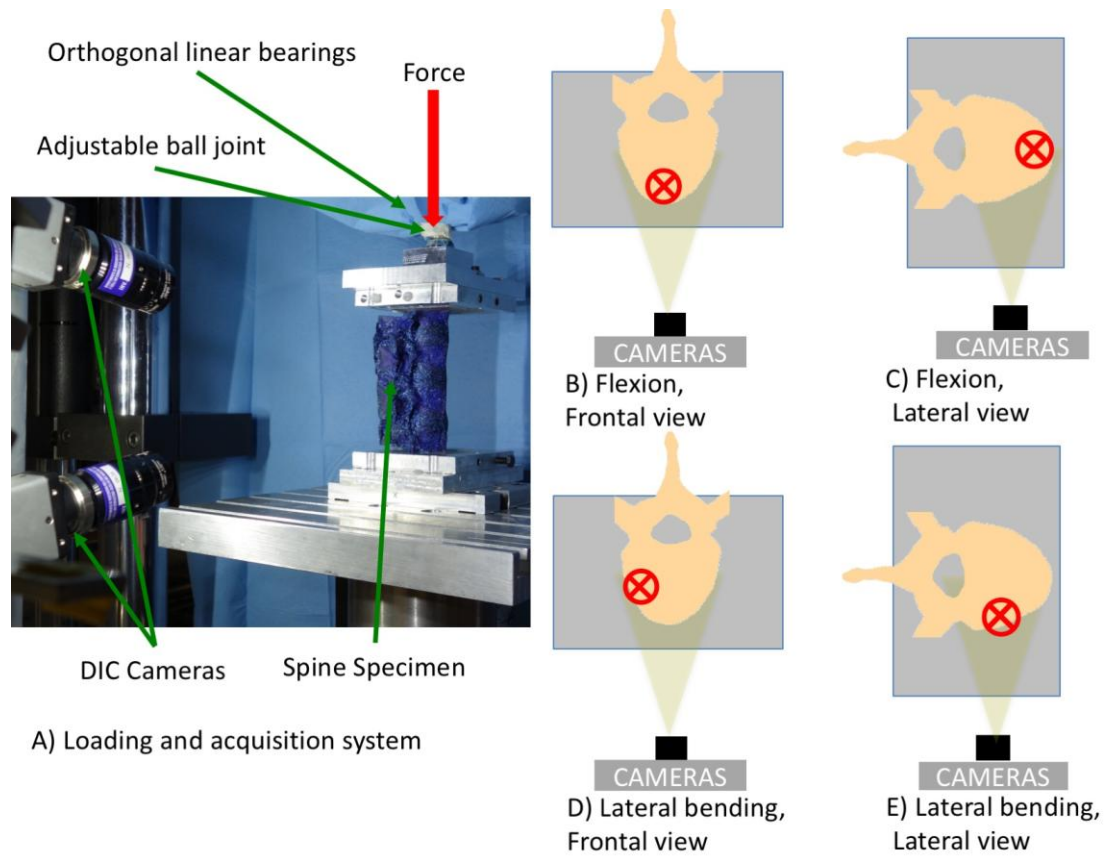


Fig. 2 - The different loading configurations were reproduced using a uniaxial testing machine and a dedicated system of low-friction linear and ball bearings to avoid transmission of undesired force components (A). The different loading configurations (flexion (B, C), lateral bending (both right and left directions were tested; the instance shown here is towards right) (D, E)) and the different views (frontal (B, D), lateral (C, E)) are sketched, viewed from top: the red cross represents the compressive force applied to the specimen. The two cameras are aligned in this top view.

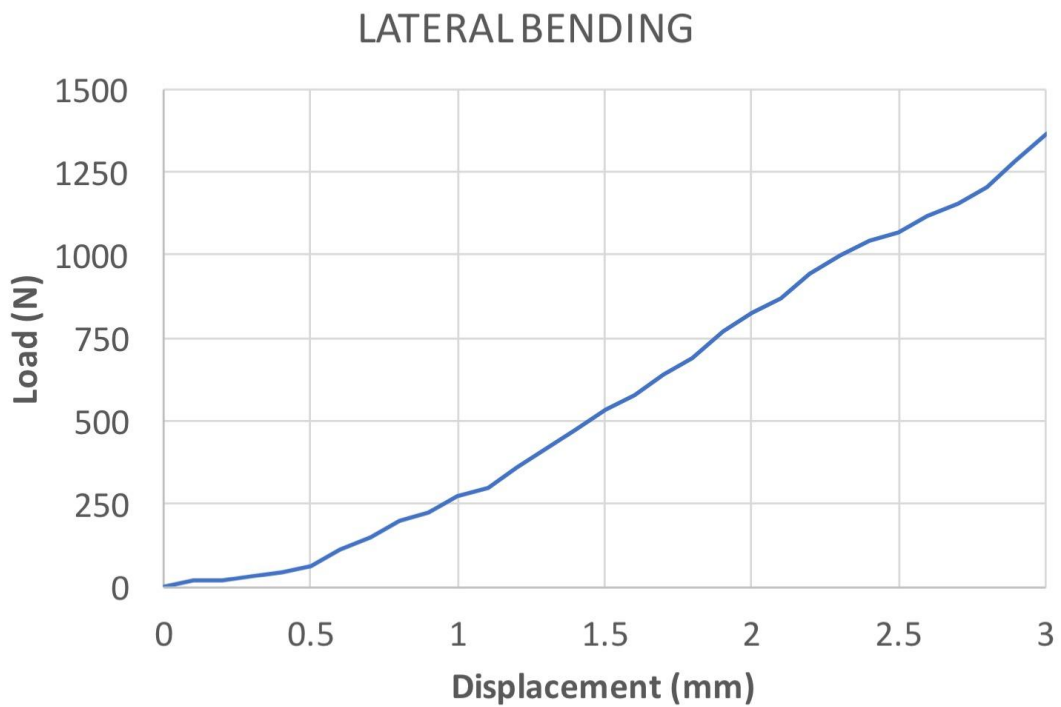
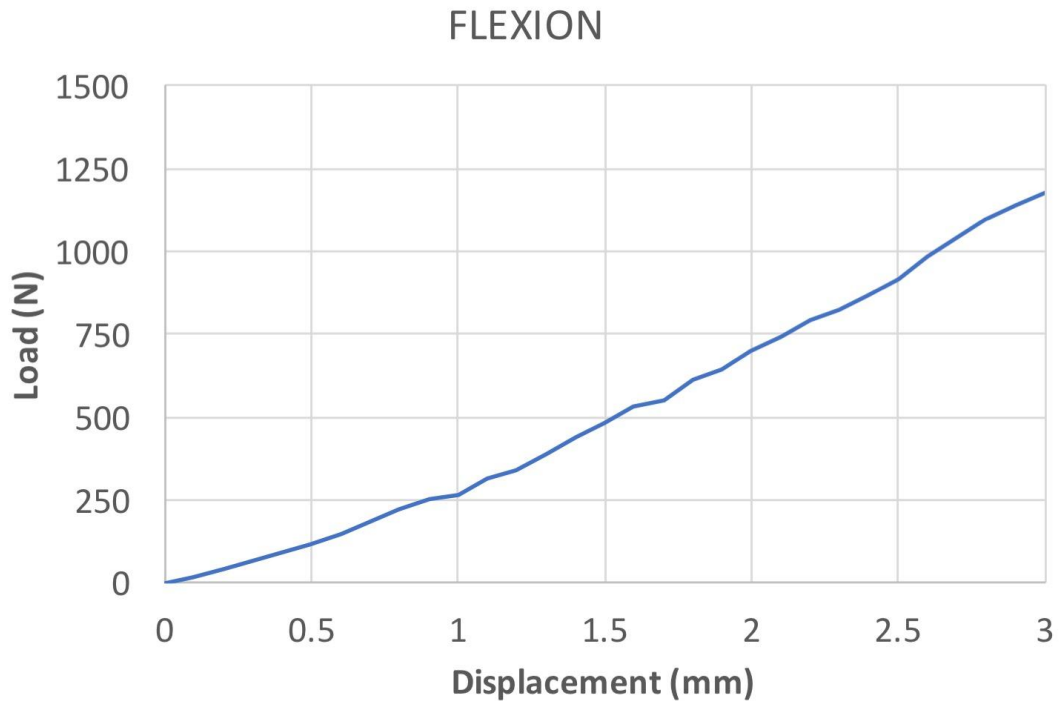


Fig. 3 – Typical Load-Displacement plot for flexion and lateral bending. The plot for one of the three specimen is represented (T11-T14): all specimens exhibited similar, rather linear, trends. The maximum load in flexion ranged between 624 N and 1956 N; for lateral bending the maximum load ranged from 566 N to 2361 N.

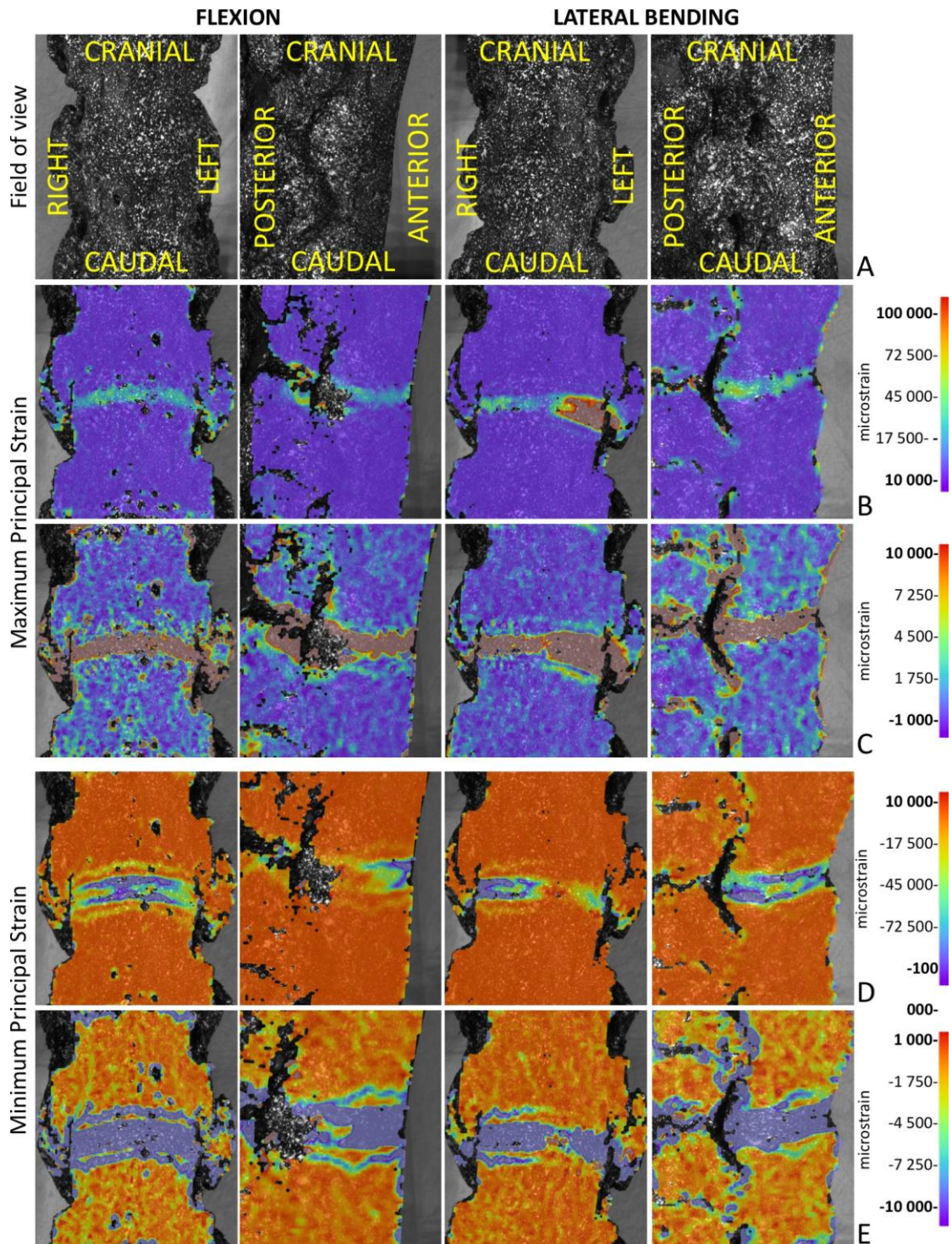


Fig. 4 - Results for a spine segment for flexion (images on the left) and for lateral bending (images on the right), both in the frontal and lateral view. The results for lateral bending with compression on the right side of the spine are shown; results for bending in the opposite direction were quite similar. The images as recorded by the DIC system are shown (A). The maps of the maximum (B, C) and minimum (D, E) principal strain are reported for the last step of the test (compression of 3 mm). As the strain in the intervertebral disc and in the vertebral bone exhibited different orders of magnitude, the strain distributions are shown twice: with a full-scale suitable for the disc (B, D) and for the vertebra (C, E). For brevity, results are shown only for one of the three specimens. The magnitude (ranges of strain observed) and distribution (localization of the strain peaks) were similar in all three specimens.

Tables

Table 1: Summary of the systematic and random errors evaluated in the anterior and lateral views for the maximum tensile (ϵ_1) and maximum compressive (ϵ_2) strains. The median and ranges over three specimens are reported.

VIEWS	STRAINS	SYSTEMATIC ERRORS Median (min-max) [microstrain]	RANDOM ERRORS Median (min-max) [microstrain]
Anterior	ϵ_1	30 (0-60)	90 (40-90)
	ϵ_2	-10 (-70-0)	90 (60-100)
Sagittal	ϵ_1	5 (0-50)	140 (90-170)
	ϵ_2	-10 (-30-10)	140 (70-180)

Full-field strain distribution in multi-vertebra spine segments:

An *in vitro* application of **Digital Image Correlation**

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Abstract

The biomechanics of the spine is experimentally assessed in terms of range of motion and overall stiffness. Quantification of the surface strain distribution is currently limited either to the vertebrae or the discs, whereas a full-field approach to measure the strain distribution in a multi-vertebra segment is currently missing. The aim of this work was to explore the feasibility of using Digital Image Correlation (DIC) to measure the strain distribution simultaneously on the vertebral bodies and the intervertebral discs of spine segments in different loading configurations.

Three porcine spine segments were tested. A white-on-black speckle pattern was prepared which covered the hard and soft tissues. Two different loading configurations (**flexion** and lateral bending) were reproduced, while two sides of the spine were analyzed with DIC.

Measurements were successfully performed on the entire region of interest of all specimens, in both configurations. The DIC analysis highlighted the strain gradients present on the spine segments including tension and compression associated with bending, the direction of principal strains in the different regions, as well as bulging of the discs under compression. Strains of tens of thousands microstrain were measured in the discs, and below 2000 microstrain in the bone.

This work showed the feasibility of applying DIC on spine segments including hard and soft tissues. It also highlights the need for a full-field investigation, because of the strain inhomogeneity in the vertebrae and discs.

Keywords: Biomechanics; Spine segment; Full-field strain measurement with Digital Image Correlation; Feasibility study; Intervertebral Discs; Vertebrae.

1 **1. Introduction**

2 The spine consists in a sequence of hard (vertebrae) and soft tissue
3 (intervertebral discs), stabilized by ligaments. Spine characterization is a
4 fundamental challenge in biomechanics because it could help engineers and
5 clinicians to better understand physiological/pathological conditions, and to
6 design better implants [1, 2].

7 Spine segments have often been investigated *in vitro* [3-7]. In such experiments
8 either known motions are imposed to measure the structural stiffness [8-10], or
9 known loads are applied to measure the range of motion[11-13]. From these
10 tests, structural properties and behaviour of the spine segment as a whole is
11 assessed. Only in few cases the local strain distribution has been measured.
12 However, such experiments focused separately either on the vertebra or on the
13 intervertebral disc. Strain in the vertebra has been measured by means of strain
14 gauges [14-17]; this provided accurate measurements, but was limited to the
15 area of application of the strain gauges. The reinforcement effect of strain
16 gauges can be significant (up to 9%) [14, 18, 19]. Measuring the local strain in
17 the intervertebral disc is even more difficult, due to the intrinsic nature of the
18 disc itself (low stiffness). One of the first measurements of the strain on the
19 outer part of the disc (*annulus fibrosus*) was based on stereo-photogrammetry
20 and covered a limited portion of the disc [20]. More recently, the entire disc
21 surface was investigated (excluding the adjacent vertebrae), using digital image
22 correlation (DIC) [21]. A method to explore the strain field on spine segments
23 with large in-plane and out-of-plane deformations is missing.

24 The aim of this work was to explore the feasibility of using DIC to measure the
25 full-field strain distribution simultaneously on the vertebral bodies and
26 intervertebral discs of thoracolumbar spine segments in different loading
27 configurations. Specifically, the following research questions were addressed:

- 28 • The feasibility of applying a speckle pattern with sufficient contrast for
29 the correlation algorithm, that should not become damaged when the
30 specimen was kept hydrated;

- 31 • The robustness of the pattern to withstand the large deformations
32 expected in the soft tissue (up to 100000 microstrain) for up to 50 cycles;
- 33 • The procedure must allow measuring the small deformations expected in
34 bone (of the order of 2000 microstrain) with acceptable errors (not
35 exceeding 200 microstrain);
- 36 • The possibility of measuring strains in specimens undergoing relatively
37 large in-plane (up to 3 mm) and out-of-plane (7 mm) displacements,
38 while correlating at least 90% of the region of interest.

39 **2. Material and methods**

40 ***2.1 Specimen and pattern preparation***

41 For ethical reasons, in this methodological work porcine spine segments from
42 the slaughterhouse were preferred. The three animals were female, of the same
43 breed, approximately 9 months old and approximately 100 kg at sacrifice. The
44 segments consisted of four thoracolumbar vertebrae (T7-T10/T11-T14/L2-L5)
45 selected for resemblance in terms of ranges of motion and dimensions [22-24].
46 The specimens were kept hydrated, and stored at -28°C to avoid alteration of the
47 mechanical properties.

48 The ribs, the muscles, the anterior longitudinal ligament, and the periosteum
49 were carefully removed using surgical tools. The interspinous, supraspinous and
50 posterior longitudinal ligaments, and the capsules were left intact in order to
51 preserve the natural kinematics. The specimens were aligned so that the middle
52 disc of each segment was aligned horizontally in the frontal and lateral views
53 [25]. In this configuration, two pots of poly-methyl-methacrylate were created
54 parallel to one another, where the upper half of the most cranial vertebra and the
55 lower half of the most caudal vertebra, were embedded (Fig. 1).

56 In order to use DIC to make measurement on the spine segments (both the
57 vertebrae and the intervertebral discs), a high-contrast white-on-black speckle
58 pattern [26, 27] was prepared. The spine segments were first stained with a
59 dark background (a solution of 4 g of methylene-blue per 100 ml of water) [28,
60 29]. Two applications were required for the intervertebral discs and three for

61 the vertebrae to obtain a uniform background. Dying was preferred, rather than
62 surface painting, as a layer of paint would crumble and crack due to the large
63 deformations of the soft tissues.

64 The speckle pattern was applied with paint instead of using a powder, as was
65 done previously [26]. This allows avoiding artifacts as the dots can strain during
66 the test (unlike powder grains). Furthermore, paint dots resist to saline solution
67 without damage, allowing specimen hydration and storage. A white water-based
68 paint (Q250201 Bianco Opaco, Chrèon, Italy) was diluted at 40% with water and
69 sprayed using an airbrush-airgun (AZ3 HTE 2, nozzle 1.8 mm, Antes Iwata, Italy)
70 to prepare a white speckle pattern with the qualitative and quantitative
71 requirements described in [30]. The spraying distance (300 mm) and the
72 pressure (1000 kPa) were optimized to obtain the desired dot size, and minimize
73 the scatter of dot dimension [35]. The actual size of the speckle dots was
74 measured in the 2D digital images (acquired by DIC system) of the ROI through a
75 custom script developed for this work. The speckles had a dimension of 6 pixels
76 (average 0.18 mm with a standard deviation of 0.18 mm), larger than the
77 minimum recommended size while the facet were sufficiently large compared to
78 the speckle size [30].

79 The specimens were soaked in saline solution before the tests.

80 ***2.2 Mechanical Testing***

81 The specimens were loaded using a uniaxial servo-hydraulic testing machine
82 (8032, Instron, UK) in displacement control. In order to avoid transmission of
83 any undesired component of load, free rotation of the loading plate was allowed
84 by means of a ball joint, while free horizontal translations were granted by
85 means of two low-friction orthogonal linear bearings.

86 In order to assess the feasibility of measuring strains on such complex
87 specimens, two different loading configurations were simulated, which are
88 frequently investigated in the literature [31] (Fig. 2):

- 89 • **Flexion:** the vertical force had an anterior offset equal to the 20% of the
90 antero-posterior depth of the central intervertebral disc;

91 • Lateral bending: the vertical force had a lateral offset (both towards right
92 and left) equal to the 20% of the coronal width of the central
93 intervertebral disc.

94 Ten preconditioning cycles were applied between 0 and 1.0 mm of compression,
95 at 0.5 Hz. A compression of 3.0 mm was applied for each loading configuration in
96 0.1 mm steps, while DIC images were acquired at each step (see below). This
97 value of compression was based on preliminary tests to reach typical strains
98 associate to physiological loads [32, 33] to prevent damage of the specimens.
99 These loading configurations did not aim to mimic any specific motor task.

100 ***2.3 Digital Image Correlation***

101 A commercial 3D-DIC system (Q400, Dantec Dynamics, Denmark) with
102 directional custom designed arrays of LEDs (10000 lumen in total) was used.
103 Images were acquired by two cameras (5MegaPixels, 2440x2050 pixels, 8-bit)
104 equipped with high-quality metrology-standard 35 mm lens (Apo-Xenoplan
105 1.8/35, Schneider-Kreuznach, Germany; 135 mm equivalent) for a stereoscopic
106 view. Due to the curvature of the vertebral bodies and discs, the cameras were
107 positioned at a distance of 260 mm from the specimen and vertically to maximize
108 the area framed by both cameras (Fig. 2).

109 The field of view was set to 70 mm by 60 mm (resulting in a pixel size of 28
110 micrometers), with depth of field of 20 mm (lens aperture f/16). The field of
111 view was wide enough to frame the entire region of interest (ROI): the central
112 intervertebral disc and the two adjacent vertebrae.

113 To explore the possibility of assessing the different sides of the spine, two
114 different acquisitions were performed for each loading configuration and each
115 specimen:

- 116 • Frontal view: the cameras pointed the anterior wall of the spine segment;
- 117 • Lateral view: the cameras pointed the lateral side (either right or left) of
118 the spine segment.

119 During the mechanical tests, series of images were acquired starting from the
120 unloaded condition (reference step) and every 0.1 mm step of compression.

121 The correlation was performed with Istra-4D (v.4.3.1, Dantec Dynamics,
122 Denmark) using the following parameters (Fig. 1):

123 a) Facet size: 33 pixels;

124 b) Grid spacing: 19 pixels;

125 c) Contour smoothing: local regression with a kernel size of 5x5;

126 It is not possible to exactly quantify the measurement spatial resolution due to
127 the nonlinearity of the smoothing process. With the settings used, a spatial
128 resolution better than 3 mm was estimated (comparable with the typical
129 dimension of the strain gauges used in biomechanical field [14]).

130 Finally, the maximum (ε_1) and minimum (ε_2) principal strains were evaluated.

131 ***2.4 Optimization and error analysis***

132 As DIC relies on a complex preparation, acquisition and analysis, its accuracy and
133 precision cannot be taken from granted. We capitalized on past experience on
134 optimization of pattern preparation [28] [35], image acquisition and correlation
135 parameters [34] (see 2.1 and 2.3).

136 Before each session, calibration was performed using a proprietary calibration
137 target (Mod. A14-BMB-9x9, Dantec Dynamics): 3D-residuum values from 0.031 to
138 0.173. It is a proprietary procedure and it consists in acquiring a series of images
139 of the target calibration in different orientations to define the physical
140 dimensions of the measurement volume, the correction of the distortions due to
141 lens and the compensation of the parallax effects.

142 An optimization and validation of the system was preliminarily performed [34]
143 to find the best compromise between the uncertainties and DIC parameters. A
144 pair of images of the unloaded segments were captured for each specimen and
145 analyzed with the optimal hardware and software settings in order to assess the
146 measurement uncertainties in a known configuration (zero-strain). Being in a
147 zero-strain configuration, any strain different from zero was accounted as
148 measurement error. The Kolmogorov-Smirnov test was used to check that the
149 errors followed a Gaussian distribution. The systematic and random errors [34]
150 were computed for each specimen and for each view as the average and the
151 standard deviation of ε_1 and ε_2 .

152 **3. Results**

153 Measurements were successfully performed on all specimens, over the entire
154 region of interest in both the frontal and sagittal view, for both loading
155 configurations, during the entire tests. Similar strain distributions were found in
156 the three specimens. **The different tissues were easily identified observing the**
157 **physical specimens and the images acquired by the cameras.** The evolution of
158 the strain distribution during load application (Fig. 3) is visible in the videos in
159 the Supplementary Material.

160 **3.1 Strain error**

161 In the zero-strain test, the apparent strains followed a Gaussian distribution
162 (Kolmogorov-Smirnov). As expected, the largest component of error was the
163 random error, and it never exceeded 190 microstrain.

164 **3.2 Flexion – Frontal view**

165 **During flexion, at most 4.4% (worst specimen) of the ROI in the frontal view lost**
166 **correlation between the first and the last step.** The discs underwent larger
167 deformations compared to the vertebral bone (Fig. 4). The strain in the discs
168 followed a strain gradient, with peaks in the central portion. In the discs, ϵ_1 was
169 **on the order of +20000 microstrain (aligned circumferentially) and ϵ_2 was of the**
170 **order of -40000 microstrain (aligned axially).**

171 The vertebrae had lower strains: peak **on the order of +500 microstrain for the ϵ_1**
172 **and around -1500 microstrain for ϵ_2 .** The two regions cranial and caudal to the
173 intervertebral discs with larger strain show the large deformation of the growth
174 **plates.**

175 **3.3 Flexion – lateral view**

176 **During flexion, at most 5.7% (worst specimen) of the ROI in the lateral view lost**
177 **correlation between the first and the last step.** The discs presented a strain
178 gradient: they were more deformed in tension in the posterior than in the
179 anterior side (Fig. 4). ϵ_1 on the intervertebral discs varied between +12000

180 microstrain (anterior side) and +87000 microstrain (posterior side). ϵ_2 followed
181 a similar gradient, from -87000 microstrain (anterior) to -18000 microstrain
182 (posterior). The direction of ϵ_1 changed from circumferential (anterior side), to
183 axial (posterior). Therefore, in **flexion** the discs tended to bulge in the anterior
184 part, and to stretch in the posterior side.

185 The vertebral **body** experienced lower strains: below +600 microstrain for ϵ_1 and
186 -1500 microstrain for ϵ_2 .

187 ***3.4 Lateral bending – frontal view***

188 During lateral bending, **at most 1.5% (worst specimen)** of the ROI in the frontal
189 view lost correlation between the first and the last step. Lateral bending was
190 applied both towards the right and the left sides, to all specimens: the outputs, in
191 terms of strain distribution on intervertebral discs and vertebrae were
192 symmetrical. The discs showed tension and compression respectively on the left
193 and right sides, depending on the direction of the imposed bending (Fig. 4). ϵ_1
194 varied between +6000 microstrain (compressed side) and +143000 microstrain
195 (stretched side). ϵ_2 were varied from -85000 microstrain (compressed side) to
196 +12000 microstrain (stretched side). The orientation of ϵ_1 changed from
197 circumferential on the compressed side, to axial on the stretched side.

198 The vertebrae had lower strains than the discs: ϵ_1 were lower than 500
199 microstrain and ϵ_2 did not exceed -1700 microstrain. In this configuration, as in
200 **flexion**, it was possible to observe both tension (ϵ_1 aligned with the vertebral
201 **axis**) and compression (ϵ_2 circumferentially) in the growth **plates**.

202 ***3.5 Lateral bending – lateral view***

203 During lateral bending, **at most 5.2% (worst specimen)** of the ROI in the lateral
204 view lost correlation between the first and the last step. The strains on the discs
205 were rather uniformly distributed **compared with the other tests** (Fig.4). When
206 the side in compression was observed, ϵ_1 in the disc were of the order of +40000
207 microstrain, and ϵ_2 of the order of -60000 microstrain.

208 The vertebral **body** had lower strains than the discs: ϵ_1 were lower than 700
209 microstrain and the ϵ_2 did not exceed -1400 microstrain.

210 **4. Discussion**

211 The main aim of this work was to develop a procedure to measure the strain
212 distribution on **thoracolumbar** spine segments [36] simultaneously on hard
213 tissues (the vertebrae) and soft tissues (the intervertebral discs) by means 3D-
214 DIC [30]. **Two different loading configurations frequently found in the literature**
215 **[6, 11-13] were explored. The specimens were observed both frontally and**
216 **laterally.**

217 The results of this work showed the technical feasibility of quantifying the full-
218 field strain distribution (**small and large deformations**) over multi-vertebra spine
219 segments during the mechanical tests. The measured strain **distributions proved**
220 **to be** highly inhomogeneous, confirming the importance of investigating the
221 spine using a full-field tool to complement the evaluation of the range of motion
222 and stiffness performed in the past.

223 **The correlation was satisfactory throughout the tests, even if large deformations**
224 **and displacements were involved. The correlated surface covered the ROI, with**
225 **a loss of correlation lower than 5.7% of the initial correlated surface. Generally,**
226 **the noise originating from the camera sensors, sub-optimal speckle dots, poor**
227 **illumination, and bad speckle framing can contribute to loss of correlation. Here,**
228 **the leakage of fluids from the vertebral body under load, which is an inherent**
229 **phenomenon when testing fresh tissue, increased the loss of correlation.**
230 Additionally, the out-of-plane movements (**7 mm**) of the postero-lateral elements
231 (capsule) caused some **subsets to be** out-of-focus in the lateral view. **While DIC**
232 **often has some un-correlated gaps, other measurement techniques such as strain**
233 **gauges simply measure strains in selected points, and ignore the rest of the**
234 **surface.**

235 The prepared white-on-black **pattern** confirmed its suitability both for hard and
236 soft tissues [28]. In fact, the background, prepared with methylene-blue, did not

237 **crack** during the tests, while the white dots remained sharp and adherent to the
238 specimen's surface.

239 A compromise was sought between reduction of measurement uncertainty and
240 spatial resolution. **The hardware (camera resolution, lenses, field of view) and**
241 **software parameters (facet size, grid spacing, contour smoothing) provided a**
242 **measurement spatial resolution better than 3 mm.** This is comparable with the
243 grid length of the strain gauges typically used in these applications (1-5 mm)
244 [37-40]. The strain measurement uncertainties (below 180 microstrain) were
245 acceptable for biomechanical purposes. Such uncertainties would not prevent
246 detecting failure of the bone (around 10000 microstrain [32]), as well as strain
247 associated to physiological loads (1000-2000 microstrain [41]). The same
248 considerations are confirmed for the intervertebral discs: as the failure strain is
249 around 250000 microstrain [20], and the physiological strain is below 150000
250 microstrain [33].

251 The evaluated strains confirmed the expected trends in all performed tests:
252 larger strains were measured in the intervertebral discs (in the order tens
253 thousands microstrain), lower strains in the vertebrae (below 2000 microstrain).
254 Such strain gradients are consistent with the expected biomechanics of the spine
255 [4]. DIC discriminated the portion of the discs subjected to
256 tension/compression, with the expected orientation of the principal strains. **In**
257 **the compressed side of the discs the minimum principal strains were axial, as**
258 **expected; the maximum principal strains were larger in absolute value than the**
259 **minimum ones, and had a transversal orientation.**

260 The combination of the **prepared pattern, selected software parameters, and**
261 **hardware** allowed recognizing the deformation on the growth **plates** of the
262 vertebrae. **Because of the spine specimens came from skeletally immature**
263 **animals** [42] the growth **plates** were not closed and **therefore** were subjected to
264 larger deformation compared to the vertebral body. **This was serendipity and**
265 **would otherwise be impossible to observe with traditional surface measurement**
266 **technique such as strain gauges.**

267 Usually, *in vitro* spine segments have been investigated in terms of range of
268 **intersegmental** motion [6, 12, 13, 43] and overall stiffness [11, 13, 44, 45]. In
269 other works, the spine was investigated **measuring** local deformation either in
270 the vertebrae or in the intervertebral discs. In the vertebrae, strains **have been**
271 **evaluated using strain gauges, which provide point-wise measurements.** At the
272 same time, strain gauges **do** cause reinforcement, especially on thin osteoporotic
273 tissues [14, 15]. Otherwise, the vertebrae have been studied using DIC but
274 without evaluating the contiguous discs [34, 46]. Conventional strain gauges
275 cannot be used on the discs due to their low elastic modulus. Strain
276 measurement is possible with optical methods [20, 21]. **In the work of Stokes *et***
277 ***al.* [20] the deformation on a small portion of the intervertebral disc was**
278 **evaluated by photogrammetry adhering small optical markers. Heuer *et al.* [47]**
279 **developed a method based on laser scanning device to evaluate the strain**
280 **distribution on intervertebral discs. In this case, the strain uncertainty (<1600**
281 **microstrain) would not allow measuring strain on hard tissue as vertebra.**
282 **Moreover, the duration of the scan (4 seconds), constrains the applicability to**
283 **quasi-static applications. In the work of Spera *et al.* [21] strain was measured**
284 **only on the discs using DIC. They used coffee powder: as the coffee grains**
285 **themselves are not deformable as the paint-dots, this can result in large noise,**
286 **apparent strain and loss of correlation.**

287 A work [48] focused on the strain distribution on spine segments. High-speed
288 3D-DIC was used to evaluate the strains on the anterior side of porcine cervical
289 spine segments in axial impacts. In contrast, the present work aims to explore
290 the feasibility of the measurement under large deformation, directed out-of-
291 plane.

292 A limitation of this work could be the use of **young** porcine spine instead of
293 human spine. **This choice was motivated by ethical considerations.** The porcine
294 spines are different from the human ones but are valid models [22, 24] to
295 demonstrate the feasibility of this novel approach.

296 **In this work, a quite standard but relatively simple configuration of 3D-DIC was**
297 **used. Array of cameras can be used to acquire 360° of the whole spine in order**
298 **to obtain a complete description of the strain maps.**

299 The applied loading configurations were not intended to replicate any specific
300 physiological motor task, but included separate components of load which are
301 present in physiological motor tasks (**flexion** and lateral bending) and which are
302 typically found in spine biomechanics [31] **as main causes of spine degeneration**
303 **[49]. This study did not cover other types of loading such as torsion.**

304 **The strains were evaluated only on the surface of the specimens. Digital Volume**
305 **Correlation (DVC) [50] has the potentiality to evaluate the strain within the**
306 **vertebral bone [51-53]. Nevertheless, applicability of DVC is limited due to the**
307 **slow tomographic image acquisition.**

308 **5. Conclusion**

309 **This work showed that by applying an adequate speckle pattern and selecting**
310 **optimal DIC parameters, the full-field strain distribution could be evaluated on a**
311 **complex structure (soft and hard tissues) such as a spine segment. This full-field**
312 **measurement would allow assessing the strain distribution, determining the**
313 **alignment of principal strains, and identify stress concentrators. This opens the**
314 **way to a new method of investigation of the spine, involving new possible**
315 **applications for basic biomechanics research and clinical innovations. DIC-based**
316 **experimental measurements could be combined with numerical models in an**
317 **inverse approach to measure (identify) the local tissue properties, including**
318 **anisotropy. Furthermore, the use of DIC will help elucidate the damage and**
319 **failure mechanism of the spine. DIC can also be used to investigate and improve**
320 **different surgical techniques and spine fixation devices.**

321 **Conflict of Interest Statement**

322 None

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328 **Competing interests:**

329 None declared

330 **Funding:**

331 None

332 **Ethical approval:**

333 Tissue specimens (porcine spines) were obtained as a side-product through the
334 food chain from animals that have been bred and sacrificed for alimentation
335 purposes. No animal was specifically bred or sacrificed for this study. The
336 Biomechanics laboratory at UNIBO meets all the Italian regulations concerning
337 research, safety in the working environment, handling, storing, testing and
338 disposal of animal and human tissue samples. All the research was carried out in
339 accordance with the National and local regulations guidelines and, in accordance
340 with the EU Directive 2010/63/EU.

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Captions and figures (images here are in low resolution)

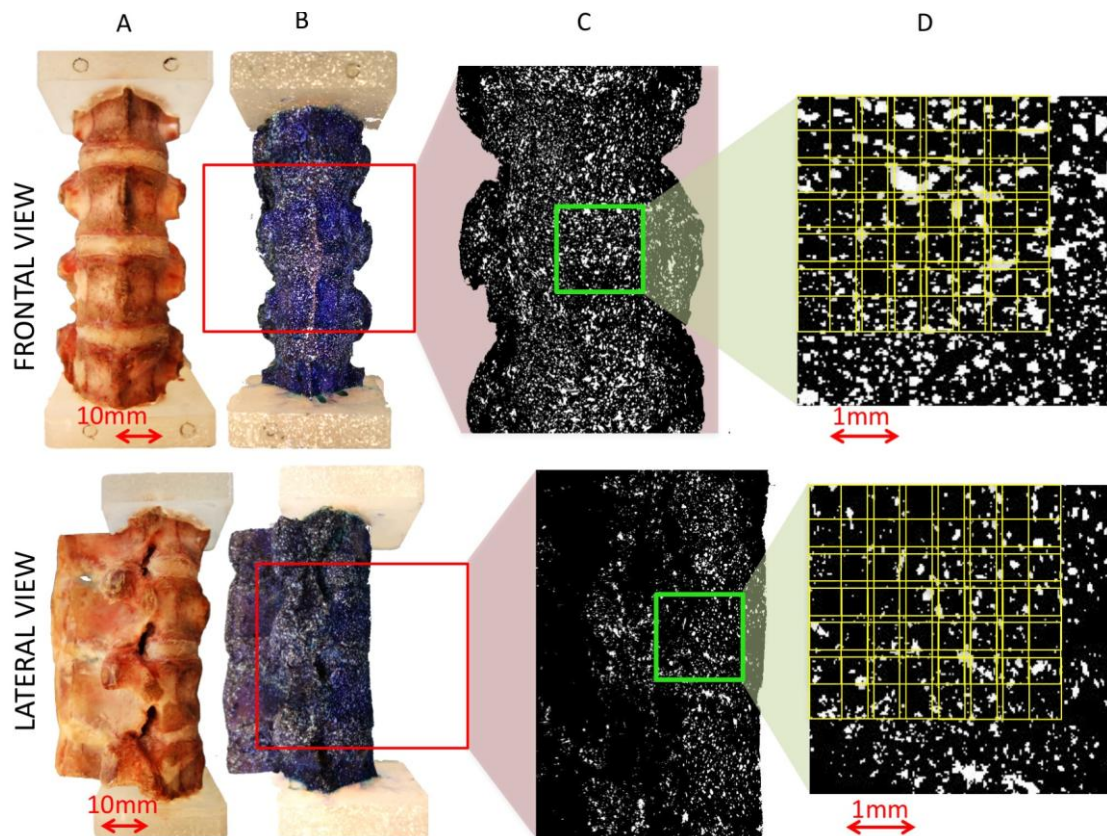


Fig. 1 - Overview at different dimensional scales of specimen preparation and analysis. The porcine spines were cleaned removing the surrounding tissues (A), and preparing the white-on-black speckle pattern (B); the red windows represent the field of view recorded by the DIC cameras. Binarized images of the entire region of interest (which covered the central disc and the two adjacent vertebrae) were generated to evaluate the dimensions of the white speckle dots (C). The facet size and grid spacing (yellow lines) can be compared to the speckle dot size in the binarized images (D).

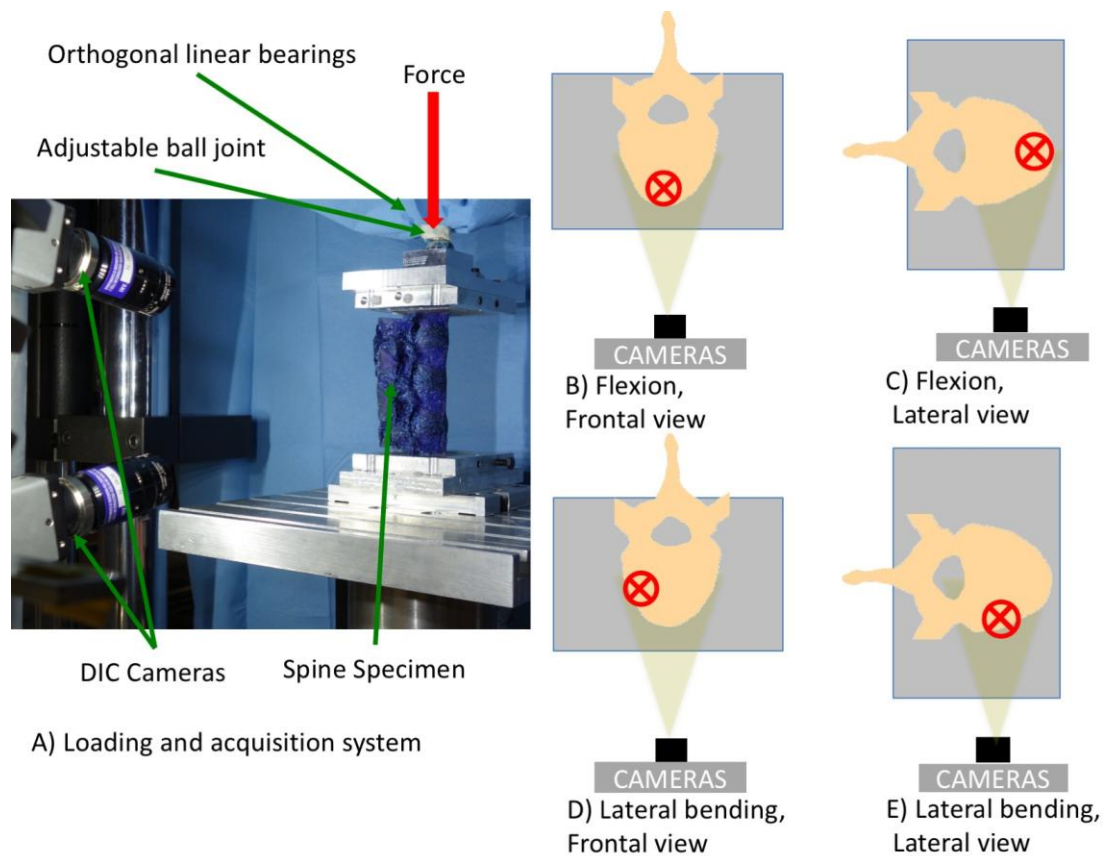


Fig. 2 - The different loading configurations were reproduced using a uniaxial testing machine and a dedicated system of low-friction linear and ball bearings to avoid transmission of undesired force components (A). The different loading configurations (*flexion* (B, C), lateral bending (*both right and left directions were tested; the instance shown here is towards right*) (D, E)) and the different views (frontal (B, D), lateral (C, E)) are sketched, viewed from top: the red cross represents the compressive force applied to the specimen. The two cameras are aligned in this top view.

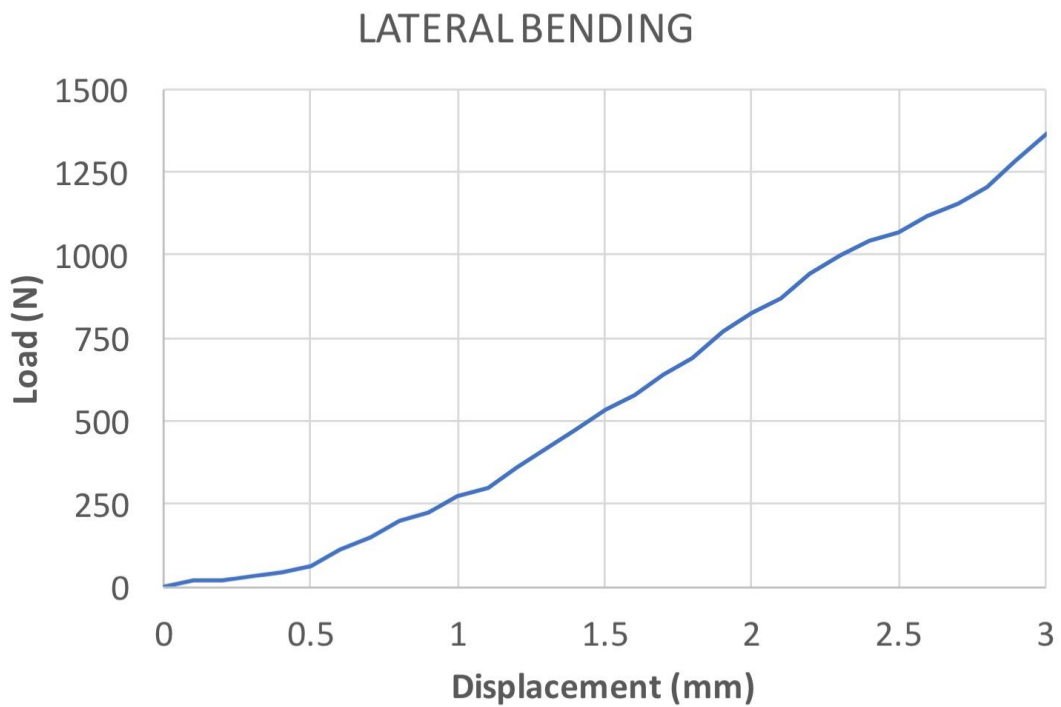
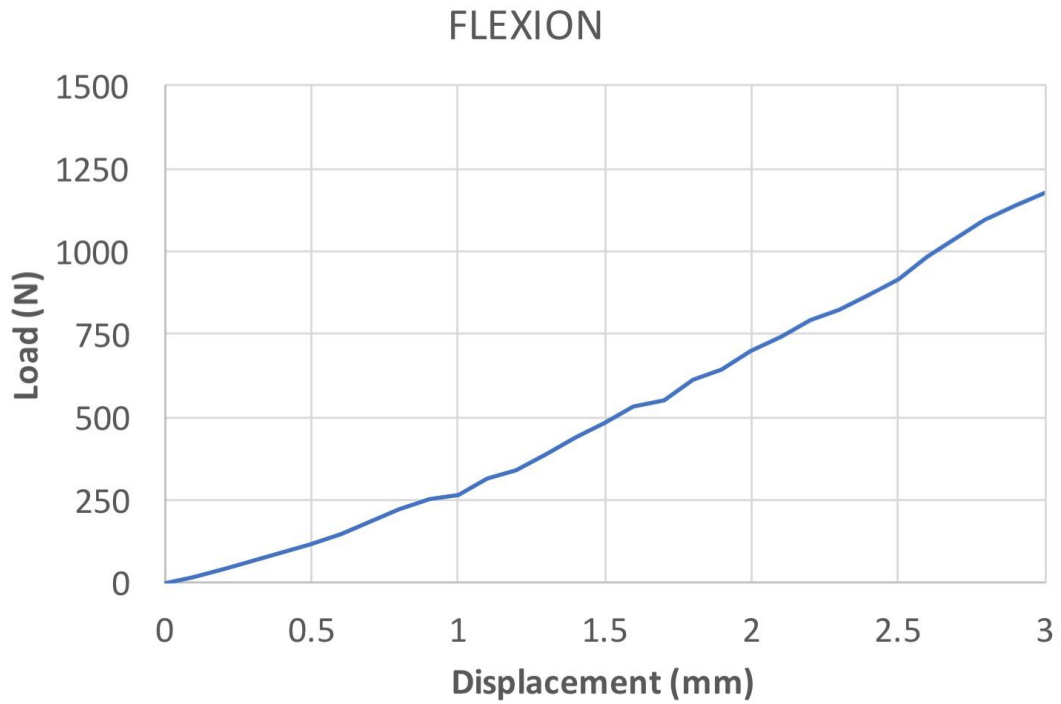


Fig. 3 – Typical Load-Displacement plot for flexion and lateral bending. The plot for one of the three specimen is represented (T11-T14): all specimens exhibited similar, rather linear, trends. The maximum load in flexion ranged between 624 N and 1956 N; for lateral bending the maximum load ranged from 566 N to 2361 N.

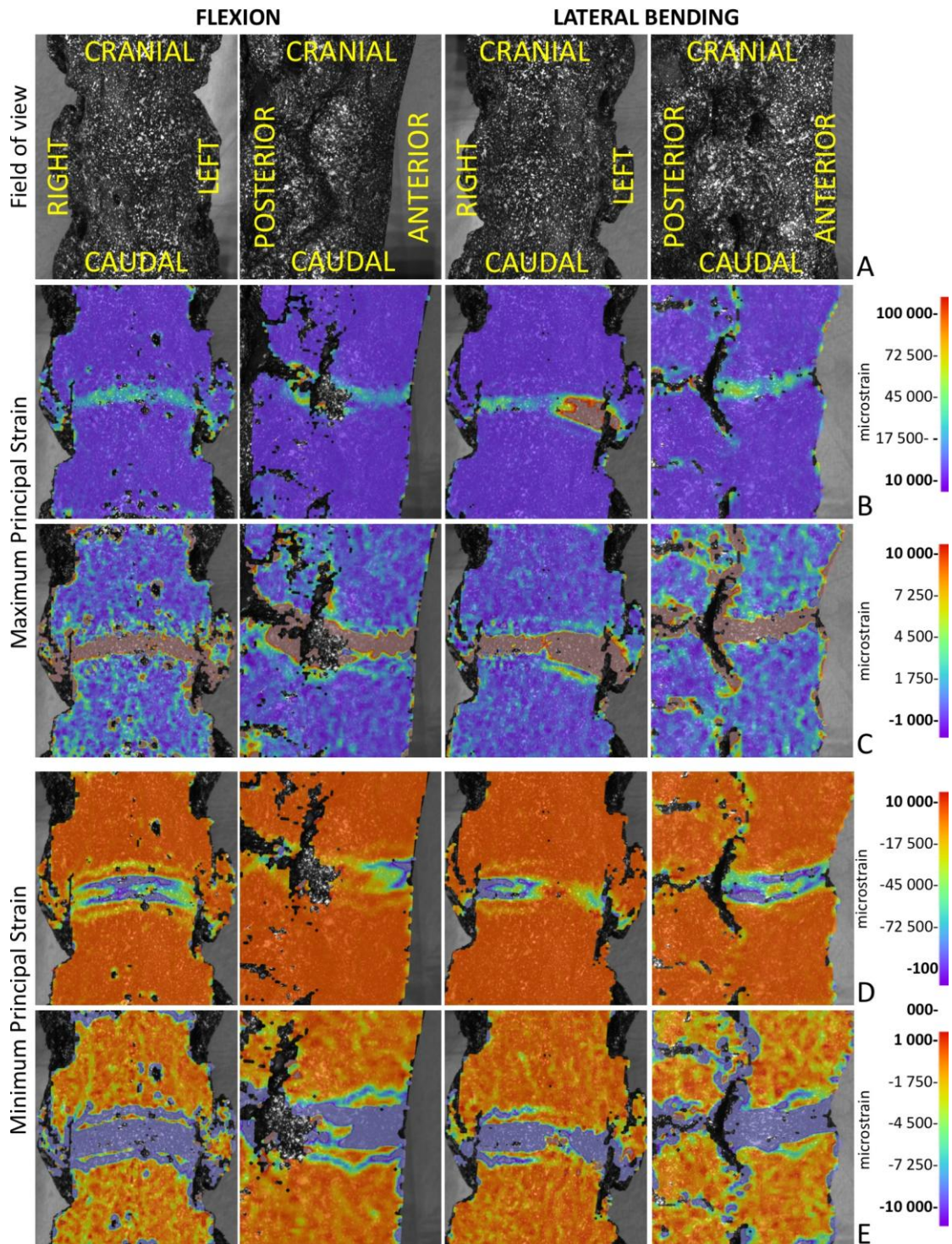


Fig. 4 - Results for a spine segment for **flexion** (images on the left) and for lateral bending (images on the right), both in the frontal and lateral view. The results for lateral bending with compression on the right side of the spine are shown; results for bending in the opposite direction were quite similar. The images as recorded by the DIC system are shown (A). The maps of the maximum (B, C) and minimum (D, E) principal strain are reported for the last step of the test (compression of 3 mm). As the strain in the intervertebral disc and in the vertebral bone exhibited different orders of magnitude, the strain distributions are shown twice: with a full-scale suitable for the disc (B, D) and for the vertebra (C, E). For brevity, results are shown only for one of the three specimens. **The magnitude (ranges of strain observed) and distribution (localization of the strain peaks) were similar in all three specimens.**

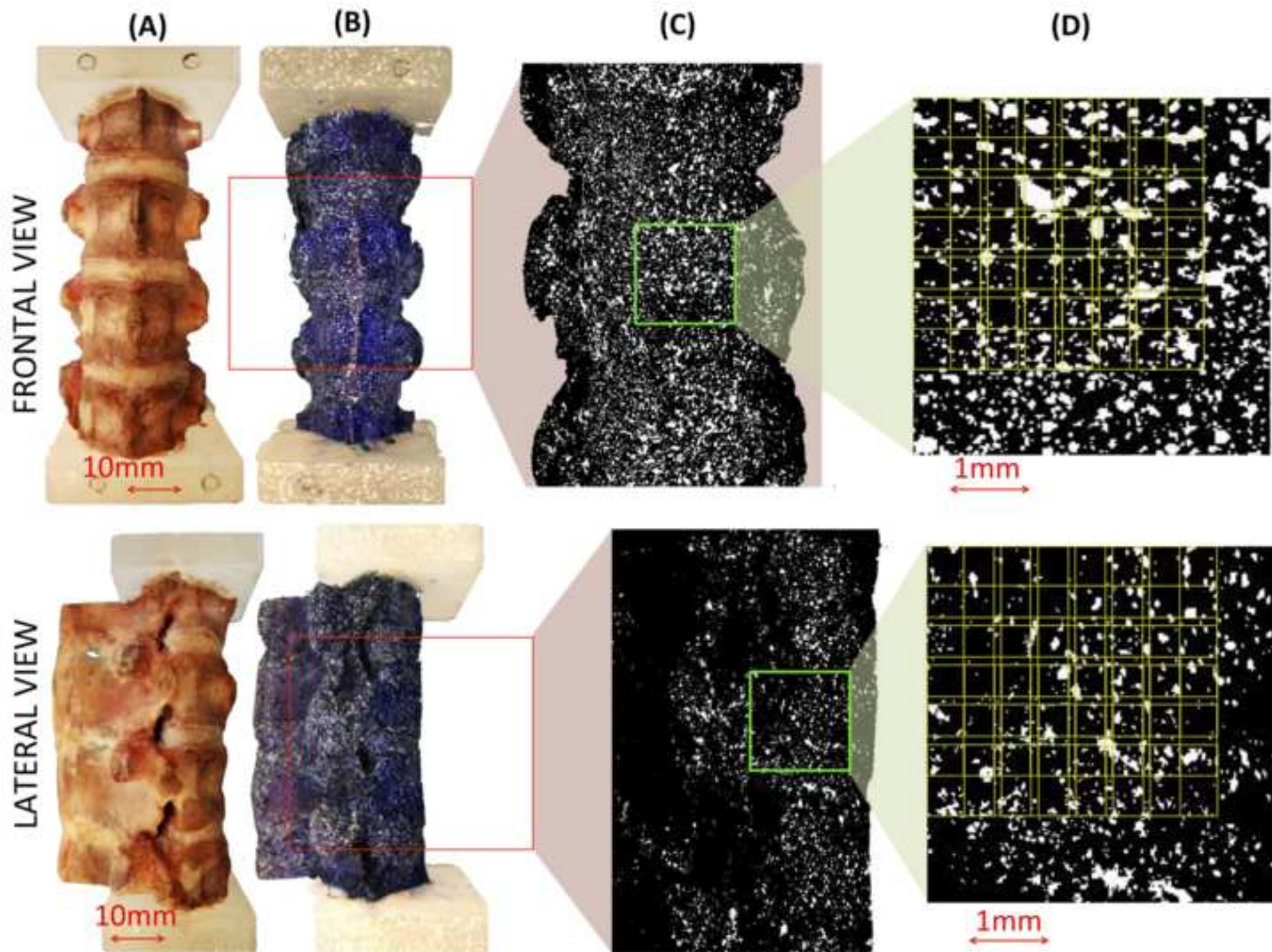
Tables

Table 1: Summary of the systematic and random errors evaluated in the anterior and lateral views for the maximum tensile (ϵ_1) and maximum compressive (ϵ_2) strains. The median and ranges over three specimens are reported.

VIEWS	STRAINS	SYSTEMATIC ERRORS Median (min-max) [microstrain]	RANDOM ERRORS Median (min-max) [microstrain]
Anterior	ϵ_1	30 (0-60)	90 (40-90)
	ϵ_2	-10 (-70-0)	90 (60-100)
Sagittal	ϵ_1	5 (0-50)	140 (90-170)
	ϵ_2	-10 (-30-10)	140 (70-180)

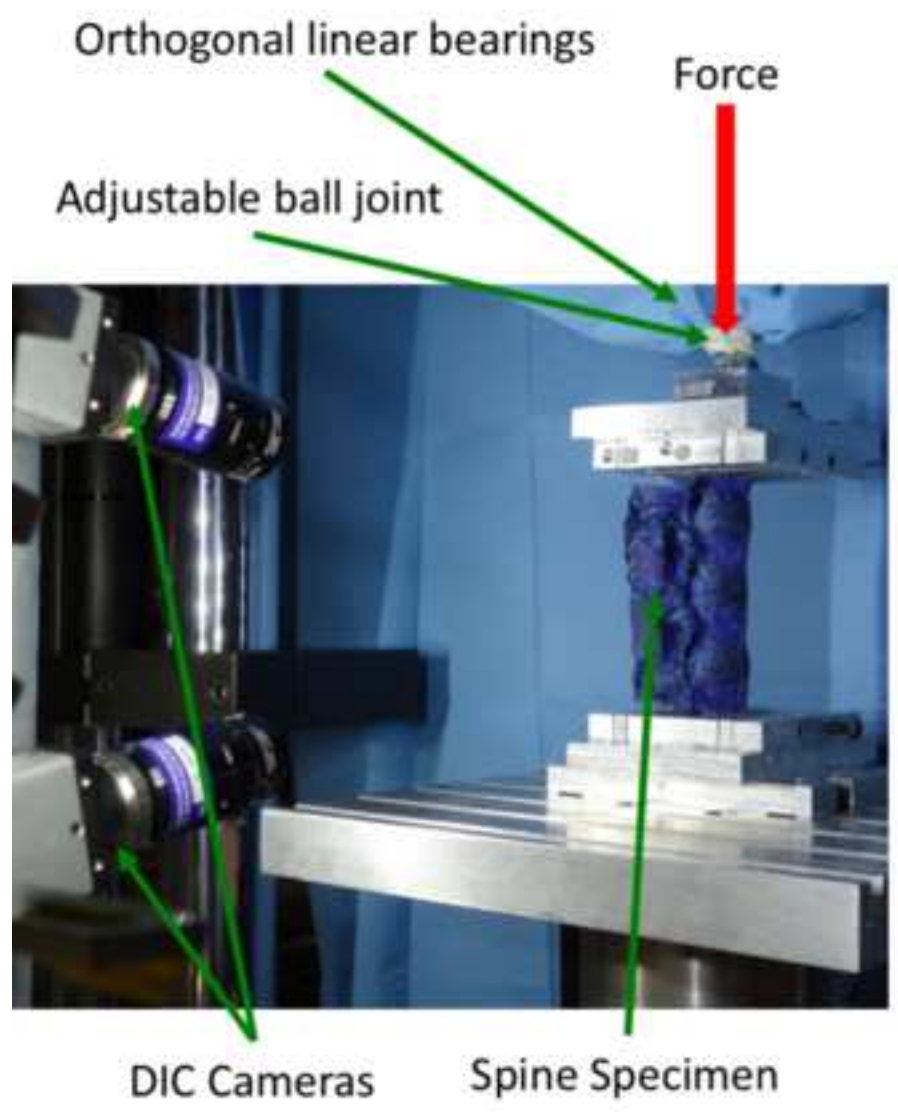
Fig_1

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Fig_2

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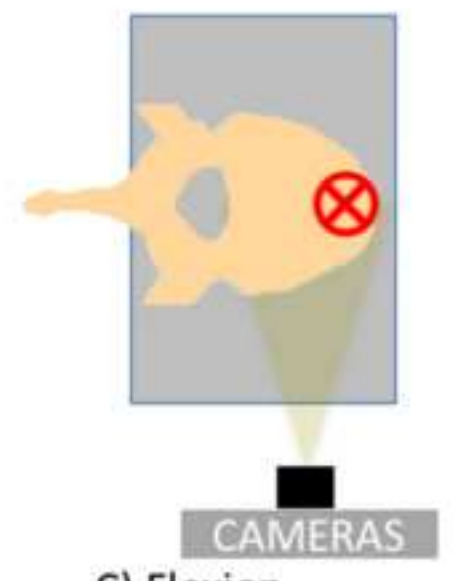
A) Loading and acquisition system



B) Flexion, Frontal view



D) Lateral bending, Frontal view



C) Flexion, Lateral view

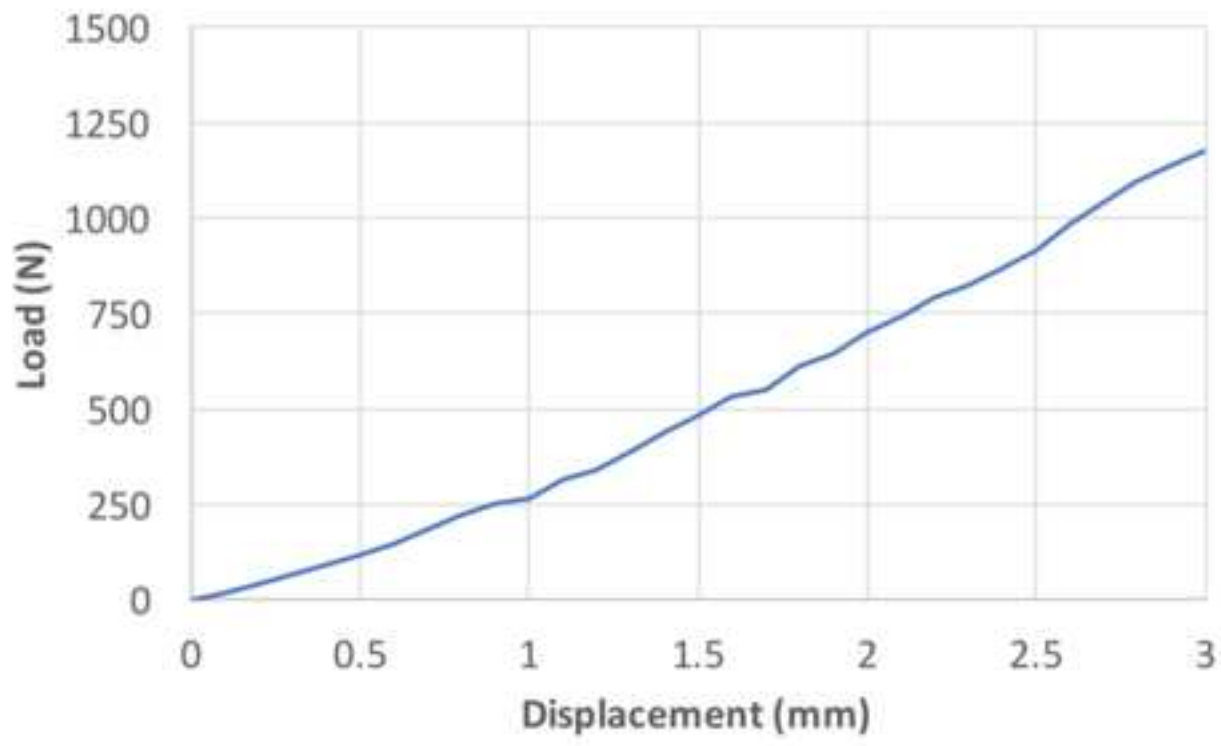


E) Lateral bending, Lateral view

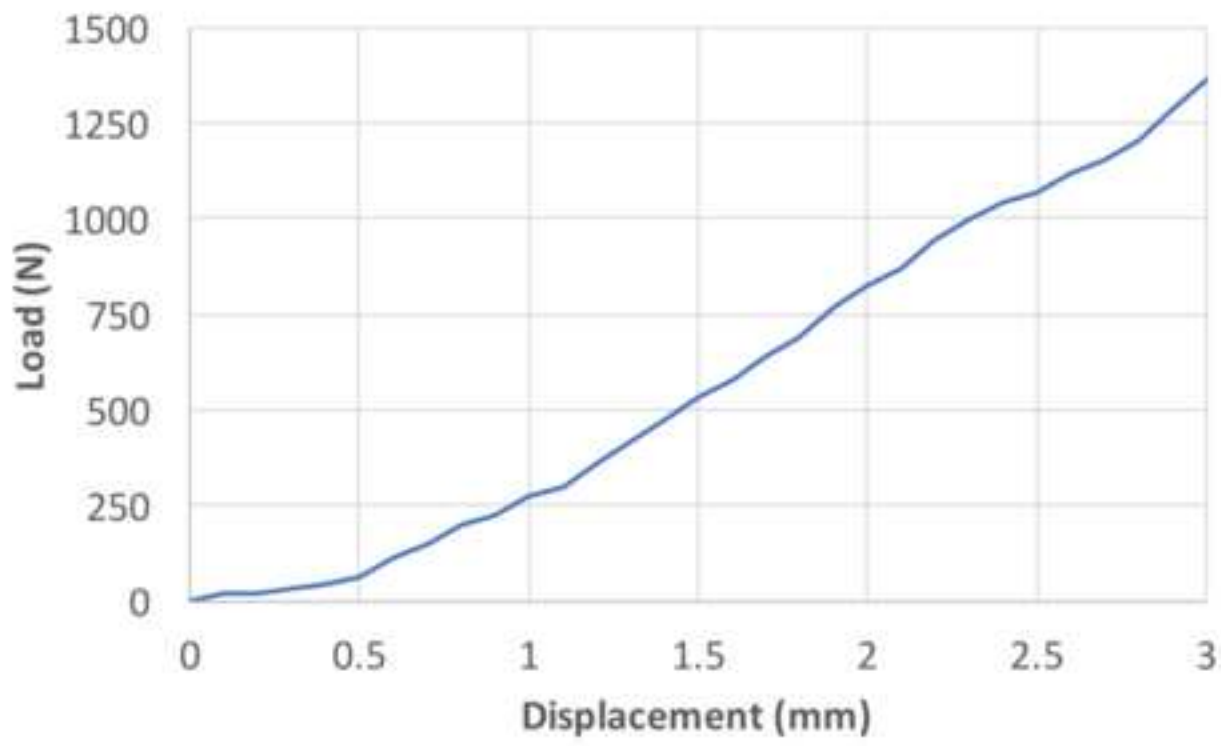
Fig_3

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FLEXION

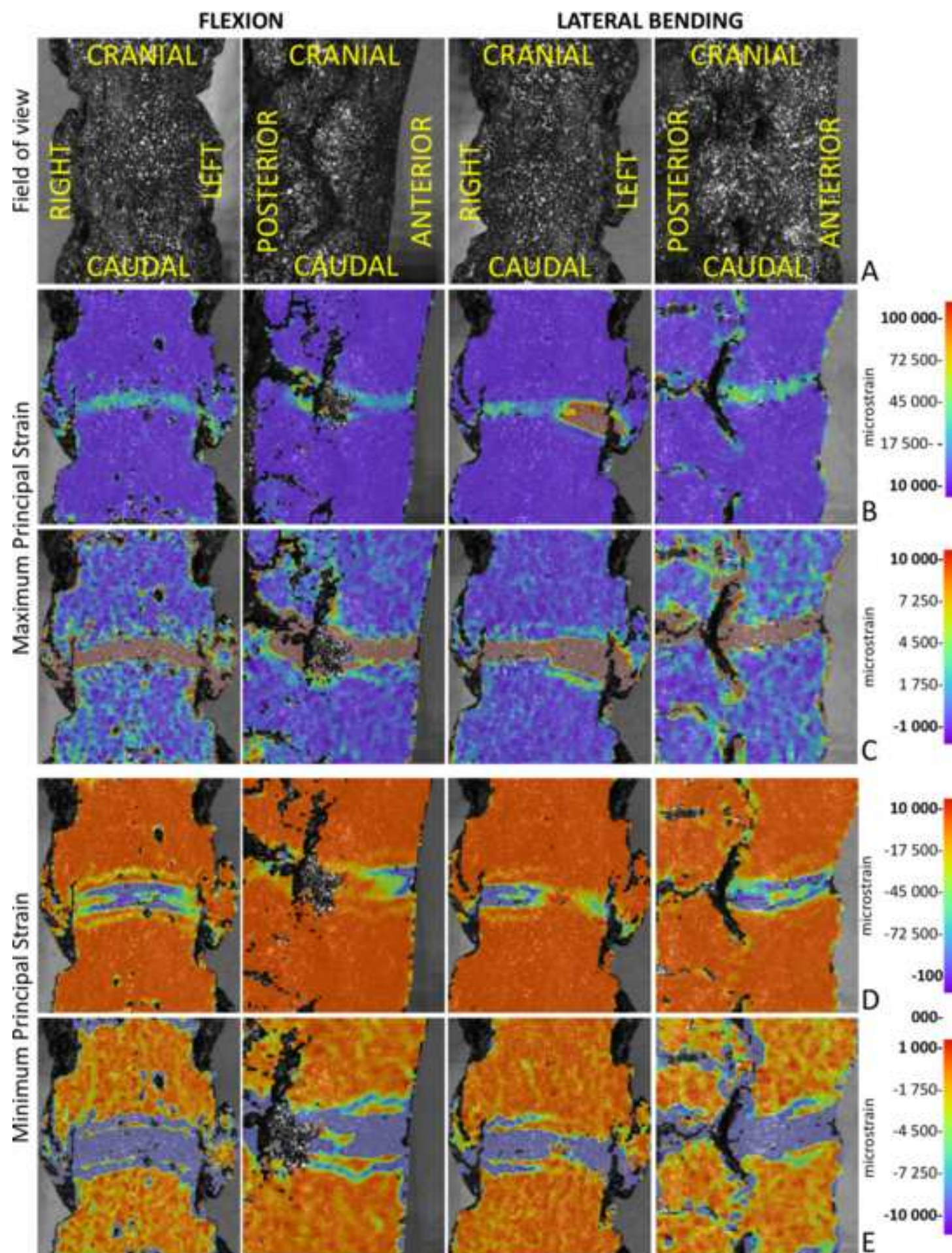


LATERAL BENDING



Fig_4

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