

The Lancet Neurology

Twelve loci provide insights into the genetic basis of lacunar stroke and small vessel disease: a meta-analysis of genome-wide association studies

--Manuscript Draft--

Manuscript Number:	THELANCETNEUROLOGY-D-20-00873R2
Article Type:	Article (Original Research)
Keywords:	STROKE; Lacunar Stroke; Small vessel disease; genome wide association study
Corresponding Author:	Markus Hugh Clinical Neurosciences Cambridge, UNITED KINGDOM
First Author:	Matthew Traylor, PhD
Order of Authors:	Matthew Traylor, PhD Elodie Persyn Liisa Tomppo Sofia Klasson Vida Abedi Mark Bakker Nuria Torres Linxin Li Steven Bell Loes Rutten-Jacobs Daniel Tozer Christoph Griessenauer Yanfei Zhang Annie Pedersen Pankaj Sharma Jordi Jimenez-Conde Tatjana Rundek Raji Grewal Arne Lindgren James Meschia Veikko Salomaa Aki Havulinna Christina Kourkoulis Katherine Crawford Sandro Marini Braxton Mitchell Steven Kittner Jonathan Rosand Martin Dichgans

	Christina Jern
	Daniel Strbian
	Israel Fernandez-Cadenas
	Ramin Zand
	Ynte Ruigrok
	Natalia Rost
	Robin Lemmens
	Peter Rothwell
	Christopher Anderson
	Joanna Wardlaw
	Cathryn Lewis
	Markus Hugh
Manuscript Region of Origin:	UNITED KINGDOM
Abstract:	<p>Background</p> <p>Despite causing up to a quarter of all strokes, the genetic basis of lacunar stroke remains poorly understood, with a single locus on 16q24 identified to date. We performed a genome-wide association study (GWAS) of lacunar stroke, expanding the sample size by recruiting robustly phenotyped cases using MRI in diagnosis, to identify novel associations and provide mechanistic insights into the disease.</p> <p>Methods</p> <p>We performed a GWAS of 7,338 cases and 225,258 controls, including newly recruited patients and reanalysis of existing studies, of which 2,987 cases (matched with 29,540 controls) were confirmed using MRI. We used multi-trait analysis of GWAS (MTAG), performing a joint analysis with a study of cerebral white matter hyperintensities (N=42,310) - an etiologically related radiological trait, to uncover additional genetic associations. We performed a transcriptome-wide association study (TWAS), to determine genes for which expression is associated with lacunar stroke, identified significantly enriched pathways using MAGMA and determined cardiovascular risk factors causally associated with the disease using Mendelian Randomization.</p> <p>Findings</p> <p>5 loci were associated with lacunar stroke in European or Transethnic meta-analysis. A further 7 loci were associated in multi-trait analysis. Two loci contain genes (COL4A2 and HTRA1) involved in monogenic lacunar stroke. Pathway analyses implicated disruption of the extracellular matrix, phosphatidylinositol 5 phosphate binding, and roundabout (ROBO) binding at FDR<0.05, while Mendelian randomization linked elevated blood pressure, history of smoking, and type 2 diabetes in the etiology of lacunar stroke.</p> <p>Interpretation</p> <p>Lacunar stroke has a substantial heritable component, with 12 loci now identified that may represent future treatment targets. These loci provide insights into lacunar stroke pathogenesis, highlighting disruption of the vascular ECM (COL4A2, LOX, SH3PXD2A, GPR126, HTRA1), pericyte differentiation (FOXF2, GPR126), TGF-beta signaling (HTRA1), and myelination (ULK4, GPR126) in disease risk.</p>

1 **Twelve loci provide insights into the genetic basis of lacunar stroke and small vessel disease:**
2 **a meta-analysis of genome-wide association studies**

3 Matthew Traylor PhD 1-2, Elodie Persyn PhD 3, Liisa Tomppo MD 4, Sofia Klasson PhD 5, Vida
4 Abedi PhD 6, Mark K Bakker MSc 7, Nuria Torres PhD 8, Linxin Li DPhil 9, Steven Bell PhD 10, Loes
5 Rutten-Jacobs PhD 11, Daniel J Tozer PhD 10, Christoph J Griessenauer MD 12-13, Yanfei Zhang
6 PhD 14, Annie Pedersen MD 5, Prof Pankaj Sharma MD 15, Jordi Jimenez-Conde MD 16, Prof
7 Tatjana Rundek MD 17, Raji P. Grewal MD 18, Prof Arne Lindgren MD 19-20, Prof James F Meschia
8 MD 21, Prof Veikko Salomaa MD 22, Aki Havulinna DSc 22-23, Christina Kourkoulis BS 24-26,
9 Katherine Crawford BS 24-25, Sandro Marini MD 24-25, Prof Braxton D. Mitchell PhD 27-28, Prof
10 Steven J. Kittner MD 28-29, Prof Jonathan Rosand MD 24-26,30, Prof Martin Dichgans MD 31-32,
11 Prof Christina Jern MD 5, Daniel Strbian MD 4-33, Israel Fernandez-Cadenas PhD 8,34, Ramin Zand
12 MD 35, Ynte Ruigrok MD 7, Prof Natalia Rost MD 36, Robin Lemmens MD 37, Prof Peter M Rothwell
13 FMed Sci 9, Christopher D Anderson MD 24-26,30, Prof Joanna Wardlaw MD 38, Prof Cathryn M
14 Lewis PhD 39,3, Prof Hugh S Markus FMed Sci 10, Helsinki Stroke Study, The Dutch Parelinoer
15 Institute-Cerebrovascular accident (CVA) Study Group, NINDS Stroke Genetics Network (SiGN), UK
16 DNA Lacunar Stroke Study Investigators, International Stroke Genetics Consortium (ISGC).

17 1, Clinical Pharmacology, William Harvey Research Institute, Queen Mary University of London,
18 London, UK

19 2, The Barts Heart Centre and NIHR Barts Biomedical Research Centre - Barts Health NHS Trust,
20 The William Harvey Research Institute, Queen Mary University London, London, UK

21 3, Department of Medical and Molecular Genetics, King's College London, London, UK

22 4, Department of Neurology, Helsinki University Hospital, Helsinki, Finland

23 5, Department of Laboratory Medicine, Institute of Biomedicine, Sahlgrenska Academy at University
24 of Gothenburg, Gothenburg, Sweden

25 6, Department of Molecular and Functional Genomics, Weis Center for Research, Geisinger Health
26 System, Danville, PA, USA

27 7, Department of Neurology and Neurosurgery, Brain Center Rudolf Magnus, University Medical
28 Center Utrecht, Utrecht, Netherlands

29 8, Stroke Pharmacogenomics and Genetics, Sant Pau Institute of Research, Hospital de la Santa
30 Creu I Sant Pau, Barcelona, Spain

31 9, Centre for the Prevention of Stroke and Dementia, Nuffield Department of Clinical Neuroscience,
32 University of Oxford, Oxford, UK

33 10, Clinical Neurosciences, University of Cambridge, Cambridge, UK

34 11, Product Development Personalized Health Care, F. Hoffmann-La Roche Ltd., Basel, Switzerland

35 12, Neuroscience Institute, Geisinger Health System, Danville, PA, USA

36 13, Institute of Neurointervention, Paracelsus Medical University, Salzburg, Austria

37 14, Genomic Medicine Institute, Geisinger Health System, Danville, PA, USA

38 15, Institute of Cardiovascular Research, Royal Holloway University of London, London, UK

39 16, Neurovascular Research Group, Department of Neurology of Hospital del Mar-IMIM (Institut
40 Hospital del Mar d'Investigacions Mediques), Universitat Autònoma de Barcelona/DCEXS-Universitat
41 Pompeu Fabra, Barcelona, Spain

42 17, Evelyn F. McKnight Brain Institute, Department of Neurology, University of Miami Miller School of
43 Medicine, Miami, USA

44 18, Neuroscience Institute, Saint Francis Medical Center, School of Health and Medical Sciences,
45 Seton Hall University, New Jersey, USA

46 19, Department of Neurology, Skane University Hospital, Lund, Sweden

47 20, Department of Clinical Sciences Lund, Neurology, Lund University, Lund, Sweden

48 21, Department of Neurology, Mayo Clinic, Jacksonville, FL, USA

49 22, Department of Public Health Solutions, Finnish Institute for Health and Welfare, Helsinki, Finland

50 23, , Institute for Molecular Medicine Finland - FIMM HiLIFE, Helsinki, Finland

51 24, Center for Genomic Medicine, Massachusetts General Hospital, Boston, USA

52 25, Program in Medical & Population Genetics, Broad Institute of Harvard and MIT, Cambridge, USA

53 26, Henry and Allison McCance Center for Brain Health, Massachusetts General Hospital, Boston,
54 USA

55 27, Division of Endocrinology, Diabetes and Nutrition, Department of Medicine, University of
56 Maryland School of Medicine, Baltimore, USA

57 28, Geriatrics Research and Education Clinical Center, Baltimore Veterans Administration Medical
58 Center, Baltimore, USA

59 29, Department of Neurology, University of Maryland School of Medicine, Baltimore, USA

60 30, Department of Neurology, Massachusetts General Hospital, Boston, USA

61 31, Institute for Stroke and Dementia Research (ISD), LMU Munich, Munich, Germany

62 32, Munich Cluster for Systems Neurology (SyNergy), Munich, Germany

63 33, Clinical Neurosciences, University of Helsinki, Helsinki, Finland

64 34, Neurovascular Research Laboratory and Neurovascular Unit, Institut de Recerca, Hospital Vall
65 d'Hebron, Universitat Autònoma de Barcelona, Barcelona, Spain

66 35, Geisinger Neuroscience Institute, Geisinger Health System, Danville, PA, USA

67 36, J. Philip Kistler Stroke Research Center, Massachusetts General Hospital, Boston, USA

68 37, KU Leuven - University of Leuven, Department of Neurosciences, Experimental Neurology; VIB
69 Center for Brain & Disease Research, University Hospitals Leuven, Department of Neurology,
70 Leuven, Belgium

71 38, Centre for Clinical Brain Sciences, UK Dementia Research Institute and Row Fogo Centre for
72 Research into the Ageing Brain, University of Edinburgh, Edinburgh, UK

73 39, Social, Genetic, and Developmental Psychiatry Centre, King's College London, London, UK

74

75 Corresponding Author:

76 Professor Hugh Markus. Stroke Research Group, Clinical Neurosciences, University of Cambridge,

77 CB2 0QQ. hsm32@medschl.cam.ac.uk

78 **Abstract**

79 *Background.* Despite causing up to a quarter of all strokes, the genetic basis of lacunar stroke remains
80 poorly understood, with a single locus on 16q24 identified to date. We performed a genome-wide
81 association study (GWAS) of lacunar stroke, expanding the sample size by recruiting robustly
82 phenotyped cases using MRI in diagnosis, to identify novel associations and provide mechanistic
83 insights into the disease.

84 *Methods.* We performed a GWAS of 7,338 cases and 225,258 controls, including newly recruited
85 patients and reanalysis of existing studies, of which 2,987 cases (matched with 29,540 controls) were
86 confirmed using MRI. We used multi-trait analysis of GWAS (MTAG), performing a joint analysis with
87 a study of cerebral white matter hyperintensities (N=42,310) - an etiologically related radiological trait,
88 to uncover additional genetic associations. We performed a transcriptome-wide association study
89 (TWAS), to determine genes for which expression is associated with lacunar stroke, identified
90 significantly enriched pathways using MAGMA and determined cardiovascular risk factors causally
91 associated with the disease using Mendelian Randomization.

92 *Findings.* 5 loci were associated with lacunar stroke in European or Transethnic meta-analysis. A
93 further 7 loci were associated in multi-trait analysis. Two loci contain genes (*COL4A2* and *HTRA1*)
94 involved in monogenic lacunar stroke. Pathway analyses implicated disruption of the extracellular
95 matrix, phosphatidylinositol 5 phosphate binding, and roundabout (ROBO) binding at FDR<0.05, while
96 Mendelian randomization linked elevated blood pressure, history of smoking, and type 2 diabetes in
97 the etiology of lacunar stroke.

98 *Interpretation.* Lacunar stroke has a substantial heritable component, with 12 loci now identified that
99 may represent future treatment targets. These loci provide insights into lacunar stroke pathogenesis,
100 highlighting disruption of the vascular ECM (*COL4A2*, *LOX*, *SH3PXD2A*, *GPR126*, *HTRA1*), pericyte
101 differentiation (*FOXF2*, *GPR126*), TGF-beta signaling (*HTRA1*), and myelination (*ULK4*, *GPR126*) in
102 disease risk.

103 *Funding.* This work was supported by a British Heart Foundation Programme Grant.

104 **Introduction**

105 Lacunar strokes are small subcortical infarcts that arise from ischemia in the territory of the deep
106 perforating arteries of the brain.^{1,2} They comprise up to a quarter of all ischaemic strokes and are
107 usually due to cerebral small vessel disease (SVD), which is also the most common pathology
108 underlying intracerebral haemorrhage and vascular cognitive impairment. Radiologically, SVD is also
109 characterized by the presence of cerebral white matter hyperintensities (WMH), enlarged perivascular
110 spaces, microbleeds and brain atrophy.³ Despite its widespread importance, few therapeutic
111 interventions have been proven to reduce SVD. One obstacle to developing new therapeutic
112 approaches has been a lack of understanding of the underlying pathophysiology. One method which
113 has been successfully used to discover pathophysiological processes and uncover potential treatment
114 targets in other complex disease is the use of genetic data derived from genome wide association
115 studies (GWAS). Recent GWAS have identified 35 loci associated with risk of ischaemic stroke and its
116 major subtypes;⁴⁻⁶ but while many loci have been identified with the other major stroke subtypes
117 (cardioembolic and large artery stroke) only one locus has robustly associated with lacunar stroke so
118 far.⁶ This is surprising because lacunar stroke is the stroke subtype most likely to be caused by
119 monogenic disease,⁷ and sporadic lacunar stroke has been strongly associated with a family history
120 of stroke.⁸ Additionally, studies of other MRI markers of CSVD have demonstrated a substantial
121 genetic component; a recent GWAS identified 31 loci across 3 phenotypes.⁹

122 We formed a collaboration to perform GWAS of lacunar stroke to identify novel associations and
123 provide mechanistic insights into the disease. We recruited cases of lacunar stroke from hospitals
124 across the UK as part of the UK DNA lacunar stroke studies 1 and 2, and from collaborators within the
125 International Stroke Genetics Consortium (ISGC); and re-analysed data from previous studies.¹⁰⁻¹³ As
126 MRI confirmation of lacunar stroke is more reliable,^{14,15} we focused on recruiting MRI confirmed cases.
127 We first performed a GWAS of this data to identify novel genetic loci associated with lacunar stroke.
128 Secondly, we used a multi-trait approach to detect additional genetic variation associated with lacunar
129 stroke in a joint analysis with cerebral WMH from a large-scale GWAS.⁹ Thirdly, we followed up our
130 initial analyses using the transcriptome-wide association study (TWAS) approach to identify transcribed
131 genes whose expression is associated with lacunar stroke, and used Mendelian randomization to
132 assess common cardiovascular risk factors that contribute to the disease. We performed analyses

133 separately in MRI confirmed and standard phenotyping groups to assess whether MRI confirmation
134 improves power to detect genetic associations.

135

136 **Methods**

137 Study Design and Phenotype Definitions

138 The workflow and design of the study is presented in Figure 1. Lacunar stroke cases were recruited
139 from a combination of acute stroke admissions and outpatient services from Europe, United States,
140 South America, and Australia. Study inclusion criteria are detailed in the appendix, and lacunar stroke
141 classification is given below. For each contributing study, approval for inclusion in this analysis
142 complied with local ethical standards and with local institutional review board/ethics committee
143 oversight. All cases and controls provided informed consent for genetic studies.

144

145 For the purposes of this analysis, lacunar stroke samples were divided into two strata: the MRI
146 confirmed group and the standard phenotyping group.

147 In the *MRI confirmed group*, lacunar stroke was defined as a clinical lacunar syndrome,¹⁶ with an
148 anatomically compatible lesion on MRI (subcortical infarct, ≤ 15 mm in diameter) - either as a region of
149 high intensity on diffusion weighted imaging (DWI) for acute infarcts, or as a region of low intensity on
150 FLAIR or T1 imaging for non-acute infarcts,³ and absence of other non-SVD causes of stroke. MRI
151 were centrally reviewed according to a standard proforma to confirm the diagnosis of lacunar stroke
152 and identify any exclusion criteria. All patients underwent comprehensive stroke investigation, including
153 brain MRI, imaging of the carotid arteries, and electrocardiogram. Echocardiography was performed
154 where appropriate. Exclusion criteria were stenosis $>50\%$ in the extra or intracranial cerebral vessels,
155 or previous carotid endarterectomy; cardioembolic source of stroke, defined according to the TOAST
156 criteria as high or moderate probability;¹⁰ cortical infarct on MRI; subcortical infarct >15 mm in diameter,
157 as these can be caused by embolic mechanisms (striatocapsular infarcts); and any other specific cause
158 of stroke (e.g. lupus anticoagulant, cerebral vasculitis, dissection, and monogenic cause of stroke).

159 In the *standard phenotyping group*, lacunar stroke was defined according to the TOAST criteria, based
160 on a clinical lacunar syndrome, and the absence of other causes of stroke, or non-lacunar infarction
161 on computed tomography.¹⁰

162

163 Genotyping and Imputation

164 Genotyping arrays, quality control filters, and imputation reference panels are listed in the appendix
165 (pp 7-8). All studies inferred the genetic ancestry of samples by comparison with reference populations
166 using principal components analysis. European samples in this study are defined as those which
167 segregated with European ancestry reference samples. The majority of studies were imputed to the
168 Haplotype Reference Consortium build. Where this was not possible due to logistical or ethical reasons,
169 imputation to 1000 Genomes Phase 3 (All ancestry groups) panels was used.^{13,17}

170

171 Statistical Analysis

172 All studies used logistic regression to assess the association of single nucleotide polymorphisms (SNP)
173 allele dosages with lacunar stroke, including ancestry informative principal components as covariates
174 as appropriate. All studies included cases with geographically matched controls, as confirmed by
175 principal components analysis. Some studies had a combination of cases based on TOAST diagnosis
176 of lacunar stroke which were re-analysed for MRI confirmation of lacunar stroke, and cases based on
177 TOAST diagnosis of lacunar stroke only for which MRI was either not acquired or we were not able to
178 access. In these circumstances we analysed the MRI confirmed and TOAST only (standard
179 phenotyping) groups separately, and divided the study controls between the two groups to avoid any
180 sample overlap. Any cases with subsequent MRI confirmation of lacunar stroke were omitted from the
181 TOAST only group; all individuals were only analyzed once.

182 Meta-analysis was performed based on a fixed effects inverse-variance weighted procedure using
183 *METAL*.¹⁸ Meta-analysis was performed in the MRI confirmed and standard phenotyping groups
184 separately, and in all studies combined. We used the principles in Winkler et al. to scrutinize datasets
185 used in the meta-analysis.¹⁹ Per study, we filtered out SNPs with imputation INFO scores < 0.7 or

186 minor allele frequency (MAF) < 0.01. Additionally, we removed low frequency or poorly imputed SNPs
187 in smaller studies by removing variants with INFO x MAF x Number of cases < 2.⁵ Genomic control
188 correction based on genomic inflation lambda was used per study to adjust for any residual inflation.²⁰
189 LDSCORE intercept values were used to assess whether population structure had been sufficiently
190 resolved at the meta-analysis level.²¹ After meta-analysis we excluded SNPs not present in at least
191 50% of cases.

192 We defined significant loci as those containing SNPs reaching $p < 5 \times 10^{-8}$ and being in linkage equilibrium
193 ($R^2 > 0.1$) with other lead SNPs. Where multiple loci met these criteria within 1Mb we used conditional
194 and joint multiple SNP analysis (GCTA-cojo) to determine whether these SNPs remained genome-
195 wide significant in a joint modelling scenario.²² We used Nagelkerke R^2 values to calculate the
196 proportion of variance explained by genome-wide significant SNPs using the NagelkerkeR2 function in
197 the R fmsb library.²³ We subtracted the R^2 estimate for the model including only principal components
198 from the model also containing the genome-wide significant SNPs to obtain an estimate of R^2 . We used
199 a genome-based restricted maximum likelihood (GREML) approach, implemented in GCTA,^{24,25} and
200 LD Score regression,²¹ to estimate heritability of lacunar stroke (MRI confirmed and non confirmed).

201

202 Multi-Trait Analysis

203 We applied multi-trait analysis of GWAS (MTAG),²⁶ performing a joint analysis with a large study of
204 cerebral WMH on MRI (N=42,310),⁹ which shares a common etiology with lacunar stroke through
205 cerebral SVD, to uncover additional genetic variation associated with lacunar stroke. We considered
206 associations significant if they attained a p-value < 5×10^{-8} in MTAG analysis, had a p-value < 0.05 for
207 association with WMH and lacunar stroke in univariate analysis, and showed greater significance in
208 MTAG analysis than in univariate analyses for WMH or lacunar stroke. To confirm our associations,
209 we used an alternative approach, Bayesian multivariate analysis of summary statistics (BMASS).²⁷

210

211 Transcriptome-Wide Association Study

212 We used the transcriptome-wide association study (TWAS) approach to identify genes for which
213 genetically altered expression was associated with lacunar stroke. Analyses were performed using
214 FUSION,²⁸ from gene expression models derived from the GTEx v7,²⁹ CommonMind Consortium
215 (CMC),³⁰ and Young Finns Study (YFS) datasets.³¹ The CMC gene expression tissues (labelled as
216 CMC-brain) were collected from the dorsolateral prefrontal cortex of individuals with schizophrenia or
217 controls (TWAS-N=452). In the YFS study (labelled as YFS-whole blood), peripheral blood gene
218 expression has been collected for 1,650 participants (TWAS-N=1,264). Among the available GTEx
219 tissues, we focused our TWAS analysis on the aortic artery (TWAS-N=267), coronary artery (TWAS-
220 N=152), tibial artery (TWAS-N=388) and whole blood (TWAS-N=369), based on the assumption that
221 these tissues would be the most relevant for lacunar stroke pathogenesis. Bonferroni correction for
222 multiple testing was applied taking into account the total number of tested genes across the tissues.
223 TWAS results were further investigated with colocalization analysis of expression quantitative trait loci
224 (eQTLs) and GWAS signals with the R package COLOC,³² to assess whether the observed eQTL and
225 GWAS associations were consistent with a common shared association.

226

227 Bioinformatics Analyses for Novel Associations

228 We used Phenoscanner to query whether our genome-wide significant SNPs have been associated
229 with DNA methylation,^{33,34} metabolite or protein levels from genome-wide studies at genome-wide
230 significance ($p < 5 \times 10^{-8}$) in other GWAS. We scanned DrugBank and DGIdb to assess the therapeutic
231 potential of targeting associated genes.^{35,36}

232

233 Pathway Analysis

234 To identify biological pathways significantly associated with risk of lacunar stroke, we used MAGMA.
235³⁷ We first used MAGMA to calculate significance of each gene based on association results, and then
236 used these gene-level statistics to estimate enrichment of Gene Ontology (GO) pathways from the
237 Molecular Signatures Database (MSigDB) using a gene-set enrichment analysis approach.³⁸ We

238 investigated only GO terms containing at least 4 and less than 200 genes and considered pathways
239 attaining a false discovery rate (FDR) < 0.05 as being significantly associated with lacunar stroke.

240

241 Mendelian Randomization Analysis

242 We performed Mendelian Randomization analyses to determine whether any lipid (low density
243 lipoprotein, high density lipoprotein, triglycerides),³⁹ blood pressure (systolic blood pressure, diastolic
244 blood pressure, pulse pressure),⁴⁰ metabolic (type 2 diabetes, body mass index),^{41,42} and lifestyle risk
245 factors (ever smoking) have a causal impact on lacunar stroke based on genetics.^{43,44} Instrumental
246 variables were independent (LD $r^2 < 0.01$) genome-wide significant ($p < 5 \times 10^{-8}$) variants associated with
247 each trait from previous analyses, and are listed in appendix pp 9-74. For blood pressure traits, we
248 included SNPs associated at genome-wide significance with any of the three traits in all analyses. For
249 body mass index, we used the set of independent SNPs provided by study authors.⁴¹ We calculated
250 the ratio of the SNP risk factor effect size by the corresponding effect size for lacunar stroke and
251 aggregated effects across all risk factor-associated SNPs using an inverse-variance weighted
252 procedure. As secondary analyses, we used median, weighted median and MR-Egger approaches to
253 aggregate across SNPs. We used the MR-Egger intercept to assess evidence of directional pleiotropy.
254 In all analyses we used the MendelianRandomization package in R.⁴⁵ Results are presented as odds
255 ratios per genetically predicted increase in each risk factor (original scale).

256

257 Role of the Funding Source

258 The funder had no role in the design or execution of the study, interpretation of data, writing of the
259 report, or the decision to submit the paper for publication. Study authors had full access to data included
260 in the study and accept responsibility to submit for publication.

261

262 **Results**

263 We meta-analysed studies from Europe, United States, and Australia, giving 6,030 cases and 248,929
264 controls of European ancestry, and 7,338 cases and 225,258 controls in Transethnic analysis. 2,987
265 (40.7%) cases (matched with 29,540 controls) had confirmation by MRI. Study cohorts, including
266 genomic inflation λ values, are described in the appendix (pp 5-6, 100-111). Following meta-analysis,
267 in the European analysis LDSCORE intercept values were equal to 1.046 (s.e = 0.008) and the λ_{1000}
268 value was 1.007, while in the Transethnic analysis the λ_{1000} value was 1.005, indicating no substantial
269 inflation. SNP-heritability of MRI confirmed lacunar stroke, calculated using GREML methods in a
270 European ancestry subset of 1,693 cases and 10,171 controls genotyped on the same array, was
271 $h^2=0.17-0.21$, standard error=0.02 assuming stroke prevalence of 1-3%, and that 20% of these are
272 lacunar strokes. Using LD Score regression estimates of SNP-heritability were lower than GREML
273 estimates, but were higher in the MRI confirmed ($h^2=0.065$, s.e=0.017) than in the non MRI confirmed
274 ($h^2=0.0081$, s.e=0.0025). The genetic correlation between MRI confirmed and non MRI confirmed
275 groups using LD Score regression was significant ($r_g=0.61$, s.e=0.21, $p=0.0033$).

276 Three loci were associated with lacunar stroke in European samples, while 3 were associated in
277 Transethnic analysis, giving 5 loci overall (Table 1, Figure 2). Regional association plots and forest
278 plots for these loci are provided in the appendix (pp 115-134). Four of the loci were novel, while one
279 was the previously associated 16q24 locus.⁶ One other locus (*ICA1L-WDR12-CARF-NBEAL1*) was
280 associated in gene-based analyses in MEGASTROKE, and was associated in a recent multi-trait
281 analysis of intracerebral haemorrhage and lacunar stroke.^{5,46}

282 We next applied MTAG to identify additional genetic variation underlying lacunar stroke in a joint
283 analysis with an etiologically related trait, cerebral WMH. Genetic correlation between lacunar stroke
284 and cerebral WMH, calculated using LDSCORE regression,²¹ was substantial for the MRI confirmed
285 group ($r_G(SE)=0.46(0.10)$ $p=4.6 \times 10^{-6}$) and slightly lower when including all lacunar strokes
286 ($r_G(SE)=0.37(0.09)$ $p=4.0 \times 10^{-5}$). In the joint analysis with cerebral WMH, variants in seven additional
287 loci reached genome-wide significance for lacunar stroke overall (Table 1, Figure 2). Four of these loci
288 (*SLC25A44-PMF1-BGLAP*, *LOX-ZNF474-LOC100505841*, *SH3PXD2A*, *COL4A2*) have previously
289 been associated with WMH.^{47,48} Regional association plots and Forest plots for the loci are provided
290 in the appendix (pp 120-134).

291 None of the 12 loci reaching genomewide significance showed evidence of heterogeneity ($p=0.05$ to
292 0.98 ; appendix pp 123-134). In two regions (*SH3PXD2A* and *HTRA1-ARMS2*) multiple apparently
293 independent ($LD\ r^2<0.1$) SNPs reached genome-wide significance. However, in a joint modelling
294 scenario employed using GCTA-cojo, only a single SNP at each of these regions remained genome-
295 wide significant showing that a single variant remains the most parsimonious explanation of the
296 association at this locus.²² We discarded two regions according to our protocol. One region on
297 chromosome 17q25 showed an association solely with WMH, with no association with lacunar stroke
298 (lead SNP $p=0.39$). A second region on chromosome 14 (*EVL-DEGS2*), was not as significant in MTAG
299 analysis ($p=1.2\times 10^{-9}$) as in WMH alone ($p=1.2\times 10^{-12}$) so an independent contribution of lacunar stroke
300 to the association could not be determined. Further evidence is required to determine that these regions
301 are associated with lacunar stroke, so each was discarded from this analysis. The *ZBTB14-EPB41L3*
302 locus that was associated with lacunar stroke was not associated with WMH ($p=0.33$ and effect in the
303 opposite direction). Similarly, for the *ULK4* locus associated with lacunar stroke, the lead SNP did not
304 reach significance for WMH ($p=0.12$), but was in the consistent effect direction and thus could reflect
305 a lack of study power.

306 The 12 loci showed stronger effects in the MRI confirmed group compared to the standard phenotyping
307 group (in European ancestry analysis, appendix pp 86), although not significantly so (one-tailed p -
308 value= 0.07), with a median proportional increase in odds ratio of 3.4%. The 12 loci explained 1.4% of
309 the overall heritability, and 6.5-8.1% of the lacunar stroke heritability from GWAS arrays, as calculated
310 in this study.

311 We performed a TWAS to identify genes for which expression was associated with lacunar stroke
312 (Figure 3). Genetically elevated *SLC25A44* was associated with lacunar stroke in multi-trait analysis
313 in arterial tissues, while genetically decreased *ULK4* was associated with lacunar stroke in arterial
314 tissues, whole blood, and brain. At the 2q33.2 locus, genetically elevated *CARF*, *FAM117B*, *ICA1L*,
315 and *NBEAL1* were all associated with lacunar stroke. All associations were confirmed by colocalization
316 analysis between the gene expression and lacunar stroke associations (posterior probability (pp) >0.7).
317 Five other associations were identified in the TWAS, but were not confirmed by colocalization analysis
318 ($pp<0.7$, Figure 3).

319 We used Phenoscanner to interrogate whether the 12 lead SNPs were associated with DNA
320 methylation, metabolite or protein levels from large scale studies.⁴⁹⁻⁵¹ Eleven of the 12 lead SNPs
321 showed associations with DNA methylation at genome-wide significance: more than expected by
322 chance based on randomly selected SNPs across the genome ($p < 0.01$), and 10 of which were
323 associated in multiple independent studies (appendix pp 87-97). Conversely, none of the 12 SNPs
324 were associated with metabolite or protein levels.

325 Querying databases that catalogue drug-gene relationships showed that 11 of the genes listed in Table
326 1 are categorized as 'druggable' indicating they have potential for therapeutic development (appendix
327 pp 98). However, no existing drugs target any of the genes identified in this study.

328 A pathway analysis based on the multi-trait analysis results using MAGMA revealed five significantly
329 associated Gene Ontology gene sets: Phosphatidylinositol 5 Phosphate Binding ($p = 2.2 \times 10^{-6}$,
330 $FDR = 0.020$), Extracellular Matrix Structural Constituent ($p = 6.2 \times 10^{-6}$, $FDR = 0.027$), Extracellular Matrix
331 Constituent Conferring Elasticity ($p = 8.9 \times 10^{-6}$, $FDR = 0.027$), Middle Ear Morphogenesis ($p = 2.3 \times 10^{-5}$,
332 $FDR = 0.049$), and Roundabout (ROBO) Binding ($p = 2.7 \times 10^{-5}$, $FDR = 0.049$). No pathways were
333 significant when based solely on lacunar stroke results. Results for all pathways with $FDR < 0.5$ are
334 presented in the appendix (pp 75-76).

335 Mendelian randomization analyses using an inverse variance weighted approach found positive
336 associations with diastolic, systolic, and pulse pressure, type 2 diabetes, and ever smoking with lacunar
337 stroke (Figure 4). No significant finding showed any evidence of pleiotropy, as assessed using the MR-
338 Egger intercept. There was evidence not reaching Bonferroni corrected significance, for a protective
339 effect of increased high-density lipoprotein on risk of lacunar stroke. There was no evidence of
340 association with body mass index, low density lipoprotein or triglycerides. Secondary analysis for all
341 risk factors using median, weighted median, and MR-Egger approaches are presented in the appendix
342 (pp 135-143).

343 GWAS Summary statistics from the primary analyses are available at GWAS Catalog
344 (<https://www.ebi.ac.uk/gwas/summary-statistics>) and on the Cerebrovascular Portal
345 (<http://www.cerebrovascularportal.org>).

346

347 Discussion

348 Despite its public health importance as the cause of a quarter of all strokes, previous GWAS studies
349 have only identified one genetic locus for lacunar stroke, in contrast to the 35 identified for ischaemic
350 stroke and its major subtypes.⁵ We performed a GWAS of lacunar stroke, including the largest number
351 of cases with MRI confirmation to date, identifying 11 novel loci in addition to replicating the only
352 previously reported locus.

353 The primary analysis identified four novel loci. One association on chromosome 11, encompassing
354 *SPI1-SLC39A13-PSMC3-RAPSN* was identified in both European and Transethnic analyses. The lead
355 SNP is a synonymous variant in *SLC39A13* (Solute Carrier Family 39 Member 13), a transmembrane
356 protein with roles in zinc transport. Mutations in this gene cause a form of Ehlers-Danlos syndrome, a
357 group of connective tissues disorders which influence the vasculature and can cause stroke;⁵² vascular
358 abnormalities have been reported in *SLC39A13* knockout mice.⁵³ We additionally identified a locus for
359 which the lead SNP resides in an intron of *ULK4* (UNC-51 Like Kinase 4) on chromosome 3. The TWAS
360 analysis suggests *ULK4* is the most likely implicated gene, with genetically decreased expression of
361 *ULK4* being associated with lacunar stroke. Variants in close LD with the lead SNP have been
362 implicated in diastolic blood pressure in large scale GWAS.⁵⁴ However, the direction of effect was
363 opposite to that for lacunar stroke, suggesting this likely reflects pleiotropy rather than a causal
364 pathway. Variants in close LD have also been associated with another cardiovascular disease, Acute
365 Aortic Dissection.⁵⁵ *ULK4* belongs to the family of serine/threonine protein kinases, a group of
366 phosphorylating kinases involved in diverse processes including cell proliferation and differentiation,
367 apoptosis and embryonic development. Its deficiency leads to hypomyelination,⁵⁶ and it has been
368 associated with neuropsychiatric traits.⁵⁷ Finally, we report a novel association on chromosome 18,
369 located between *ZBTB14* (Zinc Finger and BTB Domain Containing 14), a zinc finger transcription
370 factor, and *EPB41L3* (Erythrocyte Membrane Protein Band 4.1 Like 3), a membrane protein that inhibits
371 cell proliferation and promotes apoptosis.

372 In multi-trait analysis we identified 7 further loci, all of which are reported as associated with lacunar
373 stroke at genome-wide significance for the first time. Two have not been reported as being associated
374 with any cerebrovascular disease previously. One lies in an intergenic region between the *VTA1*

375 (Vesicle Trafficking 1) and *GPR126* (G Protein-Coupled Receptor 126) genes. *GPR126* is a G-Protein
376 Coupled Receptor which is activated by type IV collagen and has an important role in myelination.⁵⁸
377 *GPR126* binds laminin-211,⁵⁹ an extracellular matrix protein produced by astrocytes and present in
378 the brain, with key roles in development and function of the blood-brain barrier,⁶⁰ in part through
379 regulation of pericyte differentiation – a mechanism previously implicated through the *FOXF2* gene.
380^{61,62} SVD-related endothelial dysfunction has also been shown to prevent oligodendrocyte precursor
381 cell maturation, contributing to impaired myelination.⁶³ One hypothesis is that the *GPR126* variant
382 might exacerbate this process, inhibiting repair from myelin damage. The second previously unreported
383 association lies in an intergenic region, the nearest gene to which is *HTRA1* (HtrA Serine Peptidase
384 1), a gene in which rare homozygous variation leads to Cerebral Autosomal Recessive Arteriopathy
385 with Subcortical Infarcts and Leucoencephalopathy (CARASIL).⁶⁴ *HTRA1*, through processing of
386 LTBP-1 (latent transforming growth factor beta binding protein 1), promotes transforming growth factor
387 beta (TGF-beta) signaling in the vascular extracellular matrix (ECM).⁶⁵ The presence of both rare and
388 common risk variants in *HTRA1* points to it being a key molecule in lacunar stroke pathogenesis, and
389 is a feature shared with another gene identified in this study, *COL4A2*, in which rare variants also cause
390 monogenic forms of cerebral small vessel disease.⁷ Candidate gene studies have previously shown
391 associations not reaching genome-wide significance in *COL4A2* with lacunar stroke and the same
392 region has also previously been associated with intracerebral haemorrhage in multi-trait analysis, and
393 coronary artery disease.^{46,66,67} Four other loci identified (*SH3PXD2A*, *LOX-ZNF474-LOC100505841*,
394 *SLC25A44-PMF1-BGLAP*, *FOXF2-FOXQ1*) were associated with broad stroke in MEGASTROKE (see
395 appendix pp 11-18 for associations of all SNPs in MEGASTROKE) or a previous meta-analysis,^{5,62}
396 although this is the first study to confirm their association specifically with lacunar stroke. At the
397 *SLC25A44-PMF1-BGLAP* locus, the TWAS results point to an association of genetically elevated
398 *SLC25A44* with lacunar stroke, which was validated in colocalization analysis. *SLC25A44* plays a key
399 role in catabolism of branched-chain amino acids in brown adipose tissue by transporting them into
400 mitochondria,⁶⁸ and thus has potential as a mediating factor in the relationship between metabolic
401 disease and lacunar stroke. However, variants in close LD have also been associated with mosaic Y
402 chromosome loss,⁶⁹ highlighting mosaicism as an alternative mechanism. Further functional studies
403 will be required to untangle these relationships with lacunar stroke. The strength of association of all

404 associated variants was moderate to large in the context of GWAS (OR ranging from 1.10 to 1.25 in
405 Europeans) and notably larger than effects previously reported for variants associated with broad
406 stroke phenotypes.⁵ This is consistent with the variants acting specifically on the lacunar stroke
407 subtype rather than on stroke as a whole.

408 We also found that 11 of the 12 lead SNPs influence DNA methylation at genome-wide significance,
409 pointing to epigenetic changes being one source through which risk of lacunar stroke is conferred.
410 Whether this genetically-altered DNA methylation influences transcription of nearby genes – and which
411 genes are affected – should be the focus of further study. A pathway analysis implicated several
412 biological processes in lacunar stroke pathophysiology. Two pathways involved the ECM, the network
413 of extracellular molecules that provide scaffolding and biochemical support to surrounding tissues.
414 Disruption of the vascular ECM has been hypothesized to be a key component in pathogenesis of
415 CSVD, particularly in monogenic forms, and several of the genes implicated in this study (*COL4A2*,
416 *LOX*, *SH3PXD2A*, *GPR126*, *HTRA1*) play a key role in the ECM.⁷⁰ This finding lends support to this
417 hypothesis and suggests ECM dysfunction also has a key role in sporadic CSVD.

418 We performed Mendelian randomization to assess whether cardiovascular risk factors showed
419 evidence of causal association with lacunar stroke. We found support for genetically predicted elevated
420 blood pressure (systolic, diastolic, and pulse pressure), type 2 diabetes and smoking being associated
421 with lacunar stroke. The results are consistent with those from observational studies, and suggest that
422 targeting these factors would reduce risk of lacunar stroke.⁷¹ There was evidence not reaching
423 Bonferroni corrected significance for a protective effect of increased high-density lipoprotein on risk of
424 lacunar stroke, and no association with low-density lipoprotein, replicating findings in previous studies.
425^{72,73} Overall these findings show that the impact of the direct effects of low-density-lipoprotein lowering
426 medications such as statins on incidence of lacunar stroke is likely to be minimal.

427 Our study emphasises the benefit of accurate phenotyping using MRI. Using this approach, the
428 heritability of lacunar stroke using GREML was substantial, and larger than previous estimates based
429 on TOAST subtyping.⁷⁴ Using LD Score, the heritability was larger in the MRI confirmed group, but
430 estimates were considerably lower than for GREML. The use of MRI subtyping also increased the
431 strength of association of the lacunar stroke associated variants although this increase was not quite

432 significant. These results suggest that further genetic risk factor studies in lacunar stroke are likely to
433 be more successful if MRI subtyping is used.

434 Our study has limitations. The analysis was performed in a predominantly European ancestry
435 population. Large studies including diverse ancestries should be performed to assess the
436 generalizability of findings to all ethnic groups. The MTAG approach relies on the relatively strong
437 assumption that associated variants act on both traits, which may not always be the case for WMH and
438 lacunar stroke, as they reflect downstream effects of a shared common ancestor, SVD. To control for
439 this, we only considered SNPs showing association with both traits and showing greater significance
440 in MTAG analysis than with WMH or lacunar stroke alone, as being significant. However independent
441 replication will remain the gold standard for confirming these and all other reported associations in this
442 article. Recent studies have suggested that a more conservative threshold of $p < 1 \times 10^{-8}$ should be
443 considered in GWAS using larger imputation panels such as here.⁷⁵ If using this threshold one locus
444 (*ZBTB14-EPB41L3*) would no longer be significant. Additional caution should therefore be applied
445 when interpreting this finding, particularly as it was not significant in MTAG analysis. To increase
446 sample size and study power, we used publicly available controls in analyses. As such it was not
447 possible to determine whether these individuals had a history of lacunar stroke. Our analyses did not
448 adjust for age and sex; there is an ongoing debate about the importance of including such covariates
449 in genetic studies.⁷⁶ In analyses with substantial differences between case and control populations it
450 is possible that this could result in subtle biases.

451 In summary, these findings represent substantial progress in identifying the genetic mechanisms
452 underlying lacunar stroke, a disease for which there remain significant deficits in our understanding of
453 the molecular causes. Our findings highlight diverse mechanisms contributing to the disease,
454 implicating disruption of the vascular ECM (*COL4A2*, *LOX*, *SH3PXD2A*, *GPR126*, *HTRA1*), pericyte
455 differentiation (*FOXF2*, *GPR126*), TGF-beta signaling (*HTRA1*), and myelination (*ULK4*, *GPR126*) in
456 disease risk. This provides novel insights into the pathogenesis of lacunar stroke, and highlights
457 multiple candidates to take forward into functional experiments to identify specific mechanisms
458 conferring risk of lacunar stroke which could be targeted therapeutically.

459

460 **Acknowledgements**

461 The UK Household Longitudinal Study is led by the Institute for Social and Economic Research at the
462 University of Essex and funded by the Economic and Social Research Council. The survey was
463 conducted by NatCen and the genome-wide scan data were analysed and deposited by the Wellcome
464 Trust Sanger Institute. Information on how to access the data can be found on the Understanding
465 Society website <https://www.understandingsociety.ac.uk/>. This research made use of the UK Biobank
466 Resource under application number 36509. Ethical approval for UK Biobank was received from the
467 research ethics committee (REC reference 11/NW/0382). We are grateful to deCODE genetics for
468 providing data for this analysis. We acknowledge the contribution of Giorgio Boncoraglio.

469

470 **Funding**

471 This work, including collection and genotyping of the UK Young Lacunar Stroke DNA Study 2 (DNA
472 Lacunar 2), was supported by a British Heart Foundation Programme Grant (RG/16/4/32218). Matthew
473 Traylor was supported by The Barts Charity and The National Institute of Health Research Barts
474 Biomedical Research Centre. The National Institute of Neurological Disorders and Stroke – Stroke
475 Genetics Network (NINDS-SIGN) study was funded by the US National Institute of Neurological
476 Disorders and Stroke, National Institutes of Health (U01 NS069208 and R01 NS100178). Collection of
477 the UK Young Lacunar Stroke DNA Study 1 (DNA Lacunar) was primarily supported by the Wellcome
478 Trust (WT072952) with additional support from the Stroke Association (TSA 2010/01). Genotyping of
479 the DNA Lacunar samples was supported by a Stroke Association Grant (TSA 2013/01). The principal
480 funding for the WTCCC2 stroke study was provided by the Wellcome Trust, as part of the Wellcome
481 Trust Case Control Consortium 2 project (085475/B/08/Z and 085475/Z/08/Z and WT084724MA).
482 Hugh Markus is supported by a National Institute for Health Research (NIHR) Senior Investigator
483 award, and his work is supported by the Cambridge Universities NIHR Comprehensive Biomedical
484 Research Centre. CML is supported by the NIHR Biomedical Research Centre at South London and
485 Maudsley NHS Foundation Trust and King's College London. Dr Anderson is supported by NIH
486 R01NS103924 and K23NS086873. Professor Rothwell and the Oxford Vascular Study are funded by
487 the NIHR Oxford Biomedical Research Centre and by the Wellcome Trust. Collection and genotyping

488 of the Sahlgrenska Academy Study on Ischemic Stroke were primarily supported by the Swedish
489 Research Council (grant #2018-02543), the Swedish Heart and Lung Foundation (20190203), and the
490 Swedish State under the agreement between the Swedish government and the county councils (the
491 ALF-agreement, ALFGBG-720081). The genetic data from Geisinger was made available through the
492 collaboration with Regeneron Genetic Centre. Dr Lemmens is a senior clinical investigator of FWO
493 Flanders. Edinburgh Mild Stroke Study (MSS2) was funded by the Wellcome Trust (WT088134/Z/09/A)

494

495 **Contributors**

496 HSM, MT, CML designed the experiment. MT performed the meta-analysis and downstream analyses.
497 HSM provided oversight. EP analysed the TWAS. YZ, NT, MKB, VA, SK, LT, SB performed statistical
498 analysis. HSM, JW, CDA, PMR, RL, NR, YR, RZ, IF-C, DS, CJ, MD, JR, SJK, BDM, SM, KC, CK, AH,
499 VS, JFM, AL, RPG, TR, JJ-C, PS, AP, LL, CJG, LRJ, DJT were responsible for recruitment and
500 phenotyping of cohorts. All authors contributed to, reviewed, and approved the final draft of the paper.

501 **Declaration of Interests**

502 CA reports grants from National Institutes of Health of the U.S., grants from American Heart
503 Association, grants from Massachusetts General Hospital, grants from Bayer AG, personal fees from
504 ApoPharma, Inc., outside the submitted work; Dr. Bell reports grants from British Heart Foundation,
505 during the conduct of the study; AH reports grants from Academy of Finland, outside the submitted
506 work; AL reports grants from Swedish Heart and Lung Foundation, grants from Region Skåne, grants
507 from Skåne University Hospital, grants from Freemasons Lodge of Instruction Eos in Lund, grants from
508 Lund University, grants from The Foundation of Färs & Frosta - one of Sparbanken Skåne's ownership
509 Foundations, grants from Swedish Research Council, during the conduct of the study; personal fees
510 from Astra Zeneca, personal fees from BMS/Pfizer, personal fees from Portola, personal fees from
511 Bayer, outside the submitted work; HSM reports grants from British Heart Foundation, during the
512 conduct of the study; personal fees from BIBA, outside the submitted work; JR reports grants from
513 National Institutes of Health, grants from OneMind, during the conduct of the study; personal fees from
514 Boehringer Ingelheim, personal fees from Pfizer, personal fees from New Beta Innovation, outside the
515 submitted work; PR reports personal fees from Bayer, personal fees from BMS, outside the submitted

516 work; VS reports other from Novo Nordisk, other from Sanofi, grants from Bayer Ltd, outside the
517 submitted work; JW reports grants from Wellcome Trust, during the conduct of the study. All other
518 authors declare no competing interesting. This information was verified by MT and HSM.

519

520 **Data Sharing**

521 GWAS Summary statistics from these analyses are available at GWAS Catalog
522 (<https://www.ebi.ac.uk/gwas/summary-statistics>) and on the Cerebrovascular Portal
523 (<http://www.cerebrovascularportal.org>). Individual level data from the NINDS-SIGN Stroke study are
524 available to researchers through dbGAP: [https://www.ncbi.nlm.nih.gov/projects/gap/cgi-](https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000615.v1.p1)
525 [bin/study.cgi?study_id=phs000615.v1.p1](https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000615.v1.p1).

526

527 **Research in context**

528 *Evidence before this study*

529 Using the terms “stroke”, “small vessel stroke”, “lacunar stroke”, “small vessel disease”, “white matter
530 hyperintensities”, “genetics”, “GWAS”, we searched PubMed (<https://pubmed.ncbi.nlm.nih.gov>) and
531 GWAS Catalog (<https://www.ebi.ac.uk/gwas/>) for relevant reports. We only considered peer-reviewed
532 reports in English. Only a single locus on chromosome 16q24 has been robustly associated specifically
533 with lacunar stroke compared to over 30 with broad stroke phenotypes.

534 *Added value of this study*

535 The present findings substantially expand the number of genetic associations with lacunar stroke, with
536 5 loci now associated directly and a further 7 associated with lacunar stroke jointly with white matter
537 hyperintensities. These loci highlight several key mechanisms in lacunar stroke pathogenesis -
538 including extracellular matrix dysfunction, myelination, and pericyte differentiation. The current findings
539 also show that individuals with increased genetic predisposition to elevated blood pressure, history of
540 smoking, and type 2 diabetes are at increased risk of lacunar stroke, pointing to the causal role of these
541 factors in disease etiology.

542 *Implications of all the available evidence*

543 There currently are no treatments that prevent lacunar stroke aside from management of vascular risk
544 factors such as blood pressure. This is due in part to lack of understanding of mechanisms underlying
545 the disease. The present findings highlight novel mechanisms underlying lacunar stroke pathogenesis,
546 and therefore point to pathways which have potential to be targeted by therapeutics. Improved
547 treatment of elevated blood pressure and type 2 diabetes, as well as prevention of smoking in the
548 population will likely reduce the burden of lacunar stroke.

Table 1. Genome-wide Significant Loci for Lacunar Stroke in (A.) Univariate or (B.) Multi-trait Analysis.

Nearest Genes	Chr	BP	Genomic Context	rsid	RA/ OA	RAF	Lacunar Stroke (European: 6030 Cases, 219,389 Controls)			Lacunar Stroke (Transethnic: 7338 Cases, 225,258 Controls)		White Matter Hyperintensities (N=42,310)		MTAG
							OR(SE)	P value	N Studies	OR(SE)	P value	Beta(SE)	P value	P value
A.														
ICA1L-WDR12-CARF-NBEAL1	2	203,968,973	Intronic	rs72934535	T/C	0.89	1.25(0.04)	3.7x10 ⁻⁹	12	1.22(0.04)	5.2x10 ⁻⁸	0.070(0.01)	2.8x10 ⁻¹⁰	5.3x10 ⁻¹⁶
ULK4	3	41,839,370	Intronic	rs4621303	T/A	0.83	1.15(0.03)	1.7x10 ⁻⁷	14	1.16(0.03)	6.4x10 ⁻⁹	0.015(0.01)	0.12	2.2 x10 ^{-7*}
<i>SPI1-SLC39A13-PSMC3-RAPSN</i>	11	47,434,986	Exonic	rs2293576	G/A	0.67	1.14(0.02)	7.2x10 ⁻¹⁰	14	1.14(0.02)	6.0x10 ⁻¹⁰	0.030(0.01)	3.1x10 ⁻⁵	6.4x10 ⁻¹³
<i>ZCCHC14</i>	16	87,575,332	Intergenic	rs12445022	A/G	0.34	1.13(0.02)	2.5x10 ⁻⁸	13	1.12(0.02)	9.0x10 ⁻⁸	0.019(0.01)	0.0078	3.1x10 ⁻⁹
<i>ZBTB14-EPB41L3</i>	18	5,389,832	Intergenic	rs9958650	G/A	0.10	1.18(0.03)	9.9x10 ⁻⁷	12	1.19(0.03)	2.4x10 ⁻⁸	-0.011(0.01)	0.33	0.0005
B.														
SLC25A44-PMF1-BGLAP	1	156,197,380	Intronic	rs2984613	C/T	0.64	1.10(0.02)	2.5x10 ⁻⁵	13	1.09(0.02)	1.4x10 ⁻⁵	0.037(0.01)	2.3x10 ⁻⁷	8.2x10 ⁻¹⁰
<i>LOX-ZNF474-LOC100505841</i>	5	121,518,378	Downstream	rs2303655	T/C	0.81	1.14(0.03)	3.6x10 ⁻⁵	11	1.12(0.03)	0.00014	0.050(0.01)	1.4x10 ⁻⁸	1.9x10 ⁻¹⁰
<i>FOXF2-FOXQ1</i>	6	1,366,718	Intergenic	rs7766042	C/T	0.11	1.17(0.03)	3.7x10 ⁻⁶	11	1.18(0.03)	1.2x10 ⁻⁶	0.045(0.01)	7.1x10 ⁻⁵	5.2x10 ⁻⁹
<i>VTA1-GPR126</i>	6	142,562,417	Intergenic	rs225744	C/T	0.77	1.11(0.03)	3.5x10 ⁻⁵	12	1.09(0.02)	0.00050	0.037(0.01)	5.8x10 ⁻⁶	9.2x10 ⁻⁹
<i>SH3PXD2A</i>	10	105,447,838	Intronic	rs61000833	T/C	0.60	1.10(0.02)	1.7x10 ⁻⁵	12	1.07(0.02)	0.0024	0.049(0.01)	2.0x10 ⁻¹²	6.0x10 ⁻¹³
<i>HTRA1-ARMS2</i>	10	124,233,181	Intronic	rs79043147	T/C	0.07	1.21(0.04)	3.2x10 ⁻⁶	11	1.22(0.04)	1.1x10 ⁻⁶	0.057(0.01)	1.8x10 ⁻⁵	1.6x10 ⁻⁹
<i>COL4A2</i>	13	111,040,681	Intronic	rs11838776	A/G	0.29	1.11(0.02)	4.3x10 ⁻⁶	12	1.11(0.02)	1.6x10 ⁻⁶	0.050(0.01)	7.9x10 ⁻¹¹	7.9x10 ⁻¹³

A, Associations reaching genome-wide significance for Lacunar Stroke; B, Associations reaching genome-wide significance in multi-trait analysis. Chr, chromosome; BP, base position (hg19); RA, risk allele; OA, other allele; RAF, risk allele frequency; MTAG, multi-trait analysis of GWAS; logBF, log (Bayes factor); *, As A/T and C/G SNPs are removed by MTAG, results are presented for SNP in highest LD (rs9842261); Genes in bold type were associated in TWAS analysis and confirmed by colocalization.

Figure 1. Analysis pipeline

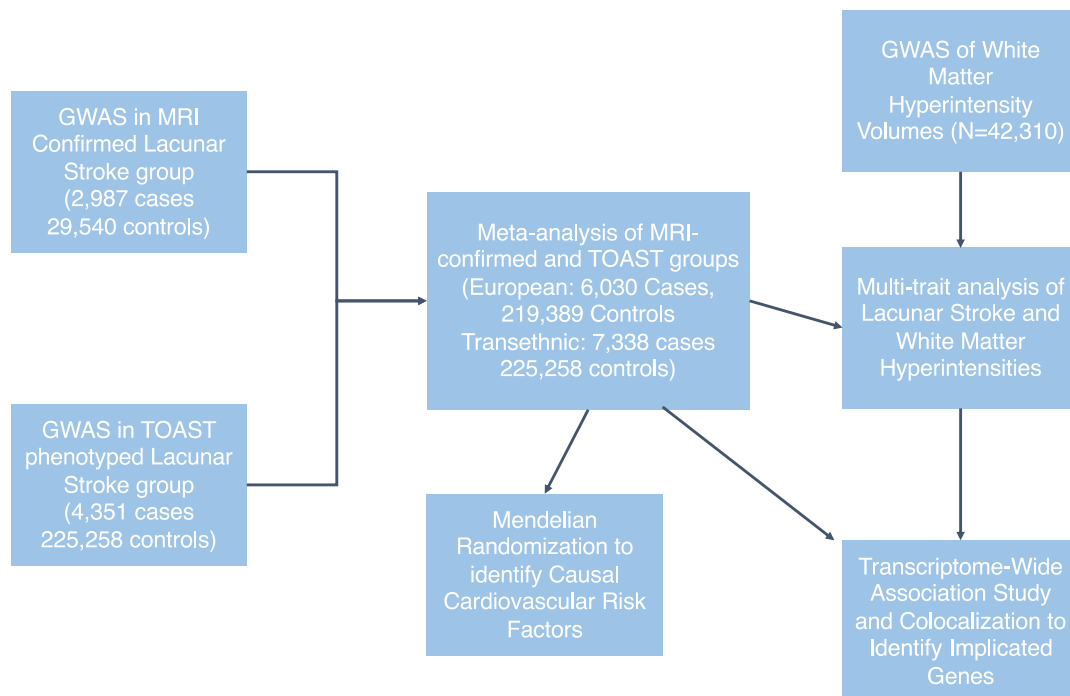


Figure 2. Manhattan plot of $-\log_{10}(\text{p-values})$ for genomewide SNP associations with A) lacunar stroke (transethnic analysis) and B) lacunar stroke multi-trait analysis, by genomic position.

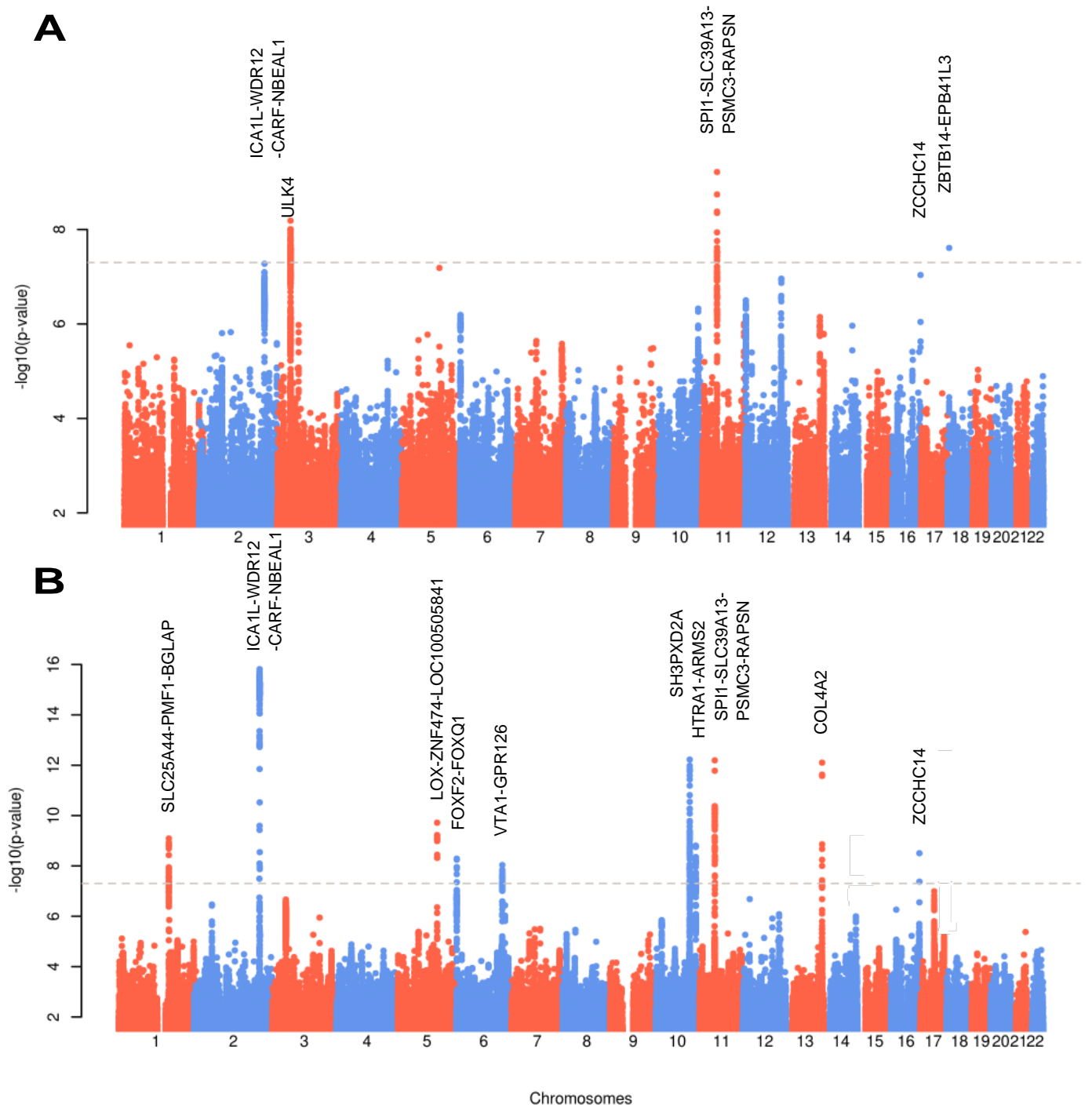
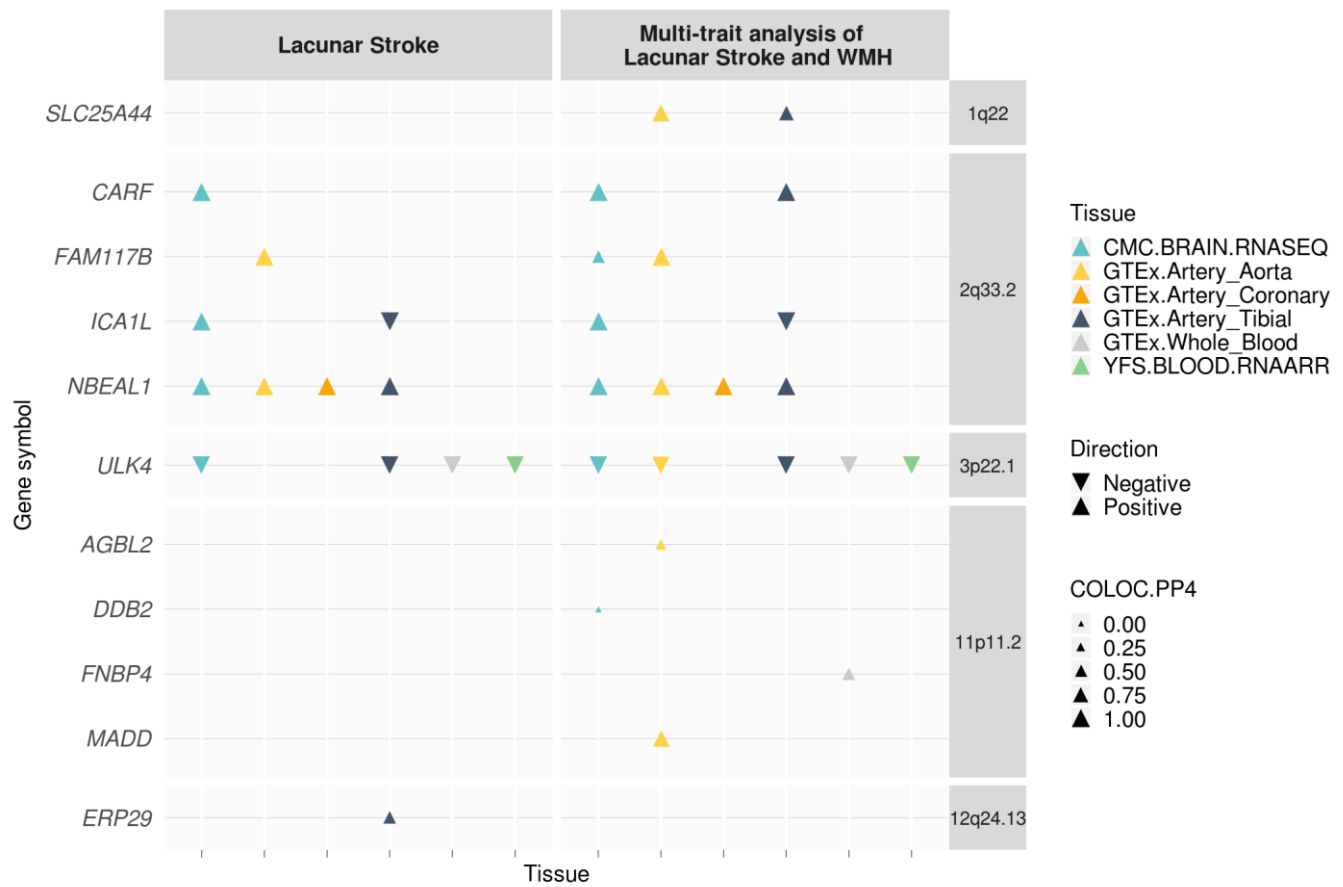
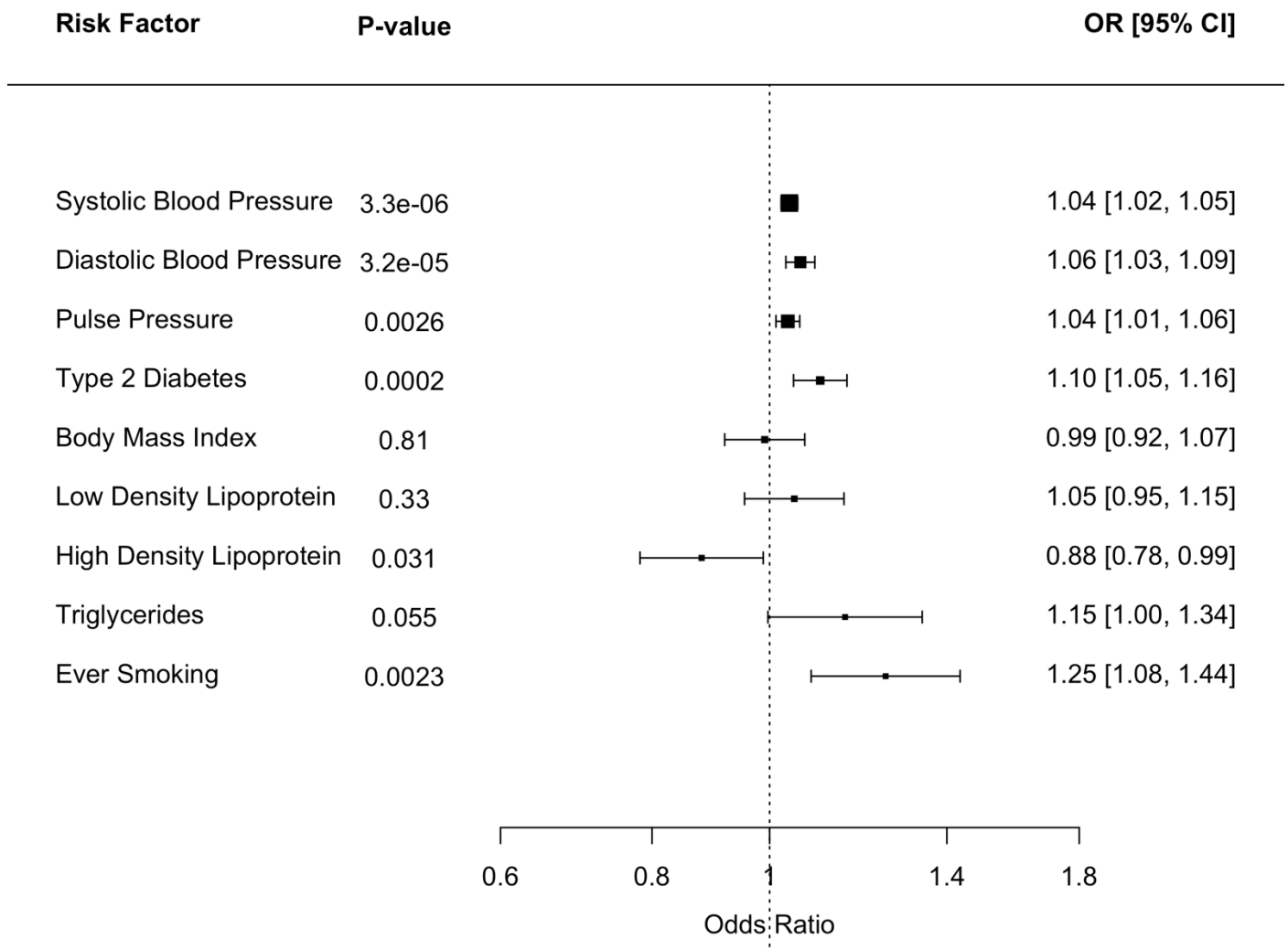


Figure 3. Genes for which expression is associated with lacunar stroke in 6 tissues from Transcriptome-Wide Association Analysis, with evidence of colocalization of gene expression and lacunar stroke signals given by triangle size



COLOC.PP4, the posterior probability of hypothesis 4 in colocalization analysis, that there is a consistent association between lacunar stroke and expression of the given gene.

Figure 4. Odds ratios for associations between genetically proxied cardiovascular risk factors and lacunar stroke from Mendelian Randomization analysis using the inverse variance weighted method



Estimates are presented as odds ratios per genetically proxied increase in each risk factor (original scale); OR, odds ratio.

References

1. Pantoni L. Cerebral small vessel disease: from pathogenesis and clinical characteristics to therapeutic challenges. *Lancet Neurol* 2010; **9**(7): 689-701.
2. Wardlaw JM, Smith C, Dichgans M. Small vessel disease: mechanisms and clinical implications. *Lancet Neurol* 2019; **18**(7): 684-96.
3. Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol* 2013; **12**(8): 822-38.
4. Malik R, Rannikmae K, Traylor M, et al. Genome-wide meta-analysis identifies 3 novel loci associated with stroke. *Ann Neurol* 2018; **84**(6): 934-9.
5. Malik R, Chauhan G, Traylor M, et al. Multiancestry genome-wide association study of 520,000 subjects identifies 32 loci associated with stroke and stroke subtypes. *Nat Genet* 2018; **50**(4): 524-37.
6. Traylor M, Malik R, Nalls MA, et al. Genetic variation at 16q24.2 is associated with small vessel stroke. *Ann Neurol* 2017; **81**(3): 383-94.
7. Tan R, Traylor M, Rutten-Jacobs L, Markus H. New insights into mechanisms of small vessel disease stroke from genetics. *Clinical science (London, England : 1979)* 2017; **131**(7): 515-31.
8. Jerrard-Dunne P, Cloud G, Hassan A, Markus HS. Evaluating the genetic component of ischemic stroke subtypes: a family history study. *Stroke* 2003; **34**(6): 1364-9.
9. Persyn E. The genetic basis of MRI markers of cerebral small vessel disease; a Genome Wide Association Study in 42,310 participants. *Under Review*.
10. Adams HP, Jr., Bendixen BH, Kappelle LJ, et al. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in Acute Stroke Treatment. *Stroke* 1993; **24**(1): 35-41.
11. Bellenguez C, Bevan S, Gschwendtner A, et al. Genome-wide association study identifies a variant in HDAC9 associated with large vessel ischemic stroke. *Nat Genet* 2012; **44**(3): 328-33.
12. Loci associated with ischaemic stroke and its subtypes (SiGN): a genome-wide association study. *Lancet Neurol* 2015.
13. McCarthy S, Das S, Kretzschmar W, et al. A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet* 2016; **48**(10): 1279-83.
14. Rajajee V, Kidwell C, Starkman S, et al. Diagnosis of lacunar infarcts within 6 hours of onset by clinical and CT criteria versus MRI. *J Neuroimaging* 2008; **18**(1): 66-72.
15. Markus HS, Khan U, Birns J, et al. Differences in stroke subtypes between black and white patients with stroke: the South London Ethnicity and Stroke Study. *Circulation* 2007; **116**(19): 2157-64.
16. Bamford J, Sandercock P, Dennis M, Burn J, Warlow C. Classification and natural history of clinically identifiable subtypes of cerebral infarction. *Lancet* 1991; **337**(8756): 1521-6.
17. Auton A, Brooks LD, Durbin RM, et al. A global reference for human genetic variation. *Nature* 2015; **526**(7571): 68-74.
18. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 2010; **26**(17): 2190-1.
19. Winkler TW, Day FR, Croteau-Chonka DC, et al. Quality control and conduct of genome-wide association meta-analyses. *Nature protocols* 2014; **9**(5): 1192-212.
20. Devlin B, Roeder K. Genomic control for association studies. *Biometrics* 1999; **55**(4): 997-1004.
21. Bulik-Sullivan BK, Loh PR, Finucane HK, et al. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet* 2015; **47**(3): 291-5.
22. Yang J, Ferreira T, Morris AP, et al. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. *Nat Genet* 2012; **44**(4): 369-75, s1-3.
23. Nagelkerke NJ. A note on a general definition of the coefficient of determination. *Biometrika* 1991; **78**(3): 691-2.
24. Lee SH, DeCandia TR, Ripke S, et al. Estimating the proportion of variation in susceptibility to schizophrenia captured by common SNPs. *Nat Genet* 2012; **44**(3): 247-50.
25. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet* 2011; **88**(1): 76-82.
26. Turley P, Walters RK, Maghziyan O, et al. Multi-trait analysis of genome-wide association summary statistics using MTAG. *Nat Genet* 2018; **50**(2): 229-37.
27. Turchin MC, Stephens M. Bayesian multivariate reanalysis of large genetic studies identifies many new associations. *PLoS Genet* 2019; **15**(10): e1008431.
28. Gusev A, Ko A, Shi H, et al. Integrative approaches for large-scale transcriptome-wide association studies. *Nat Genet* 2016; **48**(3): 245-52.

29. Battle A, Brown CD, Engelhardt BE, Montgomery SB. Genetic effects on gene expression across human tissues. *Nature* 2017; **550**(7675): 204-13.
30. Fromer M, Roussos P, Sieberts SK, et al. Gene expression elucidates functional impact of polygenic risk for schizophrenia. *Nat Neurosci* 2016; **19**(11): 1442-53.
31. Raitakari OT, Juonala M, Ronnema T, et al. Cohort profile: the cardiovascular risk in Young Finns Study. *Int J Epidemiol* 2008; **37**(6): 1220-6.
32. Giambartolomei C, Vukcevic D, Schadt EE, et al. Bayesian test for colocalisation between pairs of genetic association studies using summary statistics. *PLoS Genet* 2014; **10**(5): e1004383.
33. Staley JR, Blackshaw J, Kamat MA, et al. PhenoScanner: a database of human genotype-phenotype associations. *Bioinformatics* 2016; **32**(20): 3207-9.
34. Kamat MA, Blackshaw JA, Young R, et al. PhenoScanner V2: an expanded tool for searching human genotype-phenotype associations. *Bioinformatics* 2019; **35**(22): 4851-3.
35. Wishart DS, Feunang YD, Guo AC, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic Acids Res* 2018; **46**(D1): D1074-d82.
36. Cotto KC, Wagner AH, Feng YY, et al. DGIdb 3.0: a redesign and expansion of the drug-gene interaction database. *Nucleic Acids Res* 2018; **46**(D1): D1068-d73.
37. de Leeuw CA, Mooij JM, Heskes T, Posthuma D. MAGMA: generalized gene-set analysis of GWAS data. *PLoS Comput Biol* 2015; **11**(4): e1004219.
38. Subramanian A, Tamayo P, Mootha VK, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* 2005; **102**(43): 15545-50.
39. Teslovich TM, Musunuru K, Smith AV, et al. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature* 2010; **466**(7307): 707-13.
40. Hoffmann TJ, Ehret GB, Nandakumar P, et al. Genome-wide association analyses using electronic health records identify new loci influencing blood pressure variation. *Nat Genet* 2017; **49**(1): 54-64.
41. Yengo L, Sidorenko J, Kemper KE, et al. Meta-analysis of genome-wide association studies for height and body mass index in ~700,000 individuals of European ancestry. *bioRxiv* 2018: 274654.
42. Mahajan A, Taliun D, Thurner M, et al. Fine-mapping type 2 diabetes loci to single-variant resolution using high-density imputation and islet-specific epigenome maps. *Nat Genet* 2018; **50**(11): 1505-13.
43. Liu M, Jiang Y, Wedow R, et al. Association studies of up to 1.2 million individuals yield new insights into the genetic etiology of tobacco and alcohol use. *Nat Genet* 2019; **51**(2): 237-44.
44. Haycock PC, Burgess S, Wade KH, Bowden J, Relton C, Davey Smith G. Best (but oft-forgotten) practices: the design, analysis, and interpretation of Mendelian randomization studies. *Am J Clin Nutr* 2016.
45. Yavorska OO, Burgess S. MendelianRandomization: an R package for performing Mendelian randomization analyses using summarized data. *Int J Epidemiol* 2017.
46. Chung J, Marini S, Pera J, et al. Genome-wide association study of cerebral small vessel disease reveals established and novel loci. *Brain* 2019.
47. Verhaaren BF, DeBette S, Bis JC, et al. Multi-Ethnic Genome-Wide Association Study of Cerebral White Matter Hyperintensities on MRI. *Circ Cardiovasc Genet* 2015; **8**(2): 398-409.
48. Persyn E, Hanscombe KB, Howson JMM, Lewis CM, Traylor M, Markus HS. Genome-wide association study of MRI markers of cerebral small vessel disease in 42,310 participants. *Nature communications* 2020; **11**(1): 2175.
49. Bonder MJ, Luijk R, Zhernakova DV, et al. Disease variants alter transcription factor levels and methylation of their binding sites. *Nat Genet* 2017; **49**(1): 131-8.
50. Gaunt TR, Shihab HA, Hemani G, et al. Systematic identification of genetic influences on methylation across the human life course. *Genome Biol* 2016; **17**: 61.
51. Chen L, Ge B, Casale FP, et al. Genetic Drivers of Epigenetic and Transcriptional Variation in Human Immune Cells. *Cell* 2016; **167**(5): 1398-414.e24.
52. Bin BH, Hojyo S, Hosaka T, et al. Molecular pathogenesis of spondylocheirodysplastic Ehlers-Danlos syndrome caused by mutant ZIP13 proteins. *EMBO Mol Med* 2014; **6**(8): 1028-42.
53. Hirose T, Shimazaki T, Takahashi N, et al. Morphometric analysis of thoracic aorta in Slc39a13/Zip13-KO mice. *Cell Tissue Res* 2019; **376**(1): 137-41.
54. Ehret GB, Munroe PB, Rice KM, et al. Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. *Nature* 2011; **478**(7367): 103-9.
55. Guo DC, Grove ML, Prakash SK, et al. Genetic Variants in LRP1 and ULK4 Are Associated with Acute Aortic Dissections. *Am J Hum Genet* 2016; **99**(3): 762-9.
56. Liu M, Xu P, Guan Z, et al. Ulk4 deficiency leads to hypomyelination in mice. *Glia* 2018; **66**(1): 175-90.

57. Liu M, Xu P, O'Brien T, Shen S. Multiple roles of Ulk4 in neurogenesis and brain function. *Neurogenesis (Austin)* 2017; **4**(1): e1313646.
58. Paavola KJ, Sidik H, Zuchero JB, Eckart M, Talbot WS. Type IV collagen is an activating ligand for the adhesion G protein-coupled receptor GPR126. *Science signaling* 2014; **7**(338): ra76.
59. Mehta P, Piao X. Adhesion G-protein coupled receptors and extracellular matrix proteins: Roles in myelination and glial cell development. *Dev Dyn* 2017; **246**(4): 275-84.
60. Menezes MJ, McClenahan FK, Leiton CV, Aranmolate A, Shan X, Colognato H. The extracellular matrix protein laminin alpha2 regulates the maturation and function of the blood-brain barrier. *J Neurosci* 2014; **34**(46): 15260-80.
61. Yao Y, Chen ZL, Norris EH, Strickland S. Astrocytic laminin regulates pericyte differentiation and maintains blood brain barrier integrity. *Nature communications* 2014; **5**: 3413.
62. Identification of additional risk loci for stroke and small vessel disease: a meta-analysis of genome-wide association studies. *Lancet Neurol* 2016.
63. Rajani RM, Quick S, Ruigrok SR, et al. Reversal of endothelial dysfunction reduces white matter vulnerability in cerebral small vessel disease in rats. *Science translational medicine* 2018; **10**(448).
64. Hara K, Shiga A, Fukutake T, et al. Association of HTRA1 mutations and familial ischemic cerebral small-vessel disease. *N Engl J Med* 2009; **360**(17): 1729-39.
65. Beaufort N, Scharrer E, Kremmer E, et al. Cerebral small vessel disease-related protease HtrA1 processes latent TGF-beta binding protein 1 and facilitates TGF-beta signaling. *Proc Natl Acad Sci U S A* 2014; **111**(46): 16496-501.
66. Rannikmae K, Sivakumaran V, Millar H, et al. COL4A2 is associated with lacunar ischemic stroke and deep ICH: Meta-analyses among 21,500 cases and 40,600 controls. *Neurology* 2017; **89**(17): 1829-39.
67. Nikpay M, Goel A, Won HH, et al. A comprehensive 1,000 Genomes-based genome-wide association meta-analysis of coronary artery disease. *Nat Genet* 2015; **47**(10): 1121-30.
68. Yoneshiro T, Wang Q, Tajima K, et al. BCAA catabolism in brown fat controls energy homeostasis through SLC25A44. *Nature* 2019; **572**(7771): 614-9.
69. Wright DJ, Day FR, Kerrison ND, et al. Genetic variants associated with mosaic Y chromosome loss highlight cell cycle genes and overlap with cancer susceptibility. *Nat Genet* 2017; **49**(5): 674-9.
70. Joutel A, Haddad I, Ratelade J, Nelson MT. Perturbations of the cerebrovascular matrisome: A convergent mechanism in small vessel disease of the brain? *J Cereb Blood Flow Metab* 2016; **36**(1): 143-57.
71. Rutten-Jacobs LCA, Markus HS. Vascular Risk Factor Profiles Differ Between Magnetic Resonance Imaging-Defined Subtypes of Younger-Onset Lacunar Stroke. *Stroke* 2017; **48**(9): 2405-11.
72. Hindy G, Engstrom G, Larsson SC, et al. Role of Blood Lipids in the Development of Ischemic Stroke and its Subtypes: A Mendelian Randomization Study. *Stroke* 2018; **49**(4): 820-7.
73. Georgakis MK, Malik R, Anderson CD, Parhofer KG, Hopewell JC, Dichgans M. Genetic determinants of blood lipids and cerebral small vessel disease: role of high-density lipoprotein cholesterol. *Brain* 2020; **143**(2): 597-610.
74. Bevan S, Traylor M, Adib-Samii P, et al. Genetic heritability of ischemic stroke and the contribution of previously reported candidate gene and genomewide associations. *Stroke* 2012; **43**(12): 3161-7.
75. Wu Y, Zheng Z, Visscher PM, Yang J. Quantifying the mapping precision of genome-wide association studies using whole-genome sequencing data. *Genome Biol* 2017; **18**(1): 86.
76. Pirinen M, Donnelly P, Spencer CC. Including known covariates can reduce power to detect genetic effects in case-control studies. *Nat Genet* 2012; **44**(8): 848-51.