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Extended-spectrum beta-lactamase-producing strains among diarrheagenic *Escherichia coli* – prospective traveler study with literature review

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Abbreviations: ETEC – enterotoxigenic *Escherichia coli*; ESBL-EC – extended-spectrum betalactamase-producing *E. coli*; DEC – diarrheagenic *E. coli*; TD – travelers' diarrhea

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ABSTRACT

Background: Antibiotics are no longer the primary approach for treating all travelers' diarrhea (TD): most cases resolve without antibiotics and using them predisposes to colonization by multidrug-resistant bacteria. Data are accumulating on increasing resistance among TD pathogens, yet research into the most common agents, diarrheagenic *Escherhichia coli* (DEC), remains limited.

Methods: A total of 413 travelers to the (sub)tropics were analyzed for travel-acquired diarrheal pathogens and ESBL-PE. To identify ESBL-producing DEC, ESBL-producing *E. coli* (ESBL-EC) isolates were subjected to multiplex qPCR for various DEC pathotypes: enteroaggregative (EAEC), enteropathogenic (EPEC), enterotoxigenic (ETEC), enteroinvasive (EIEC), and enterohemorrhagic (EHEC) *E. coli*.

For a literature review, we screened studies among travelers and locals in low- and middle-income countries (LMIC) on the frequency of ESBL-producing DEC, and among travelers, also DEC with resistance to ciprofloxacin, azithromycin, and rifamycin derivatives.

Results: Our rate of ESBL-EC among all DEC findings was 2.7% (13/475); among EAEC 5.7% (10/175), EPEC 1.1% (2/180), ETEC 1.3% (1/80), and EHEC (0/35) or EIEC 0% (0/5). The literature search yielded three studies reporting ESBL-EC frequency and thirteen exploring resistance to TD antibiotics among travel-acquired DEC. For EAEC and ETEC, the ESBL-EC rates were 10–13% and 14–15%, resistance to fluoroquinolones 0–42% and 0–40%, azithromycin 0–29% and 0–61%, and rifaximin 0% and 0–20%. The highest rates were from the most recent collections. Proportions of ESBL-producing DEC also appear to be increasing among locals in LMICs and even carbapenemase-producing DEC were reported.

Conclusions: ESBL producers are no longer rare among DEC and the overall resistance to various antibiotics is increasing. The data predict decreasing efficacy of antibiotic treatment, threatening its benefits, for disadvantages still prevail when efficacy is lost.

INTRODUCTION

Uncontrolled use of antibiotics is a major driver of the ongoing antimicrobial resistance (AMR) pandemic which threatens global health¹. Increasing fastest in the tropics¹, AMR is being transported worldwide by international travelers: 20–70% of visitors to low- and middle-income countries (LMIC) carry multidrug-resistant bacteria (MDR), particularly extended-spectrum beta-lactamase-producing *Enterobacteriaceae* (ESBL-PE), to their home country²⁻⁷ and may spread them further^{2,6}. During the past decade, avoiding unnecessary antibiotic use while abroad has emerged as a means to combat travel-related global spread of AMR. In addition to the general pressure to avoid unnecessary antibioties^{1,8}, this policy is particularly encouraged by findings that antibiotic use predisposes travelers to acquisition of multidrug-resistant intestinal bacteria²⁻⁷ – and thus contributes to the global spread of AMR, colonized travelers acting as intercontinental transporters^{7,9,10}.

Special attention has been paid to treatment of travelers' diarrhea (TD) which ranks as the most common indication for travelers' antibiotic use¹¹: 5–45% of those with TD take these drugs to alleviate their symptoms^{3-6,12-17}. As described in the literature, stand-by antibiotics for TD are

prescribed at pre-travel consultations for 7–20% of European^{11,13,15} and practically all US travelers^{12,16,17}. Recently, the rates have also decreased somewhat in the US¹⁸. While antibiotics certainly retain their place in treating the most severe TD cases, their use for moderate TD has recently become topical¹⁹. Although, compared to placebo, antibiotics shorten the disease duration by 0.7–1.5 days^{20,21}, in most TD cases the drugs are not necessary, since the disease usually resolves spontaneously. Anti-diarrheals such as loperamide offer an alternative with no impact on AMR colonization²²; there are no studies that prove antibiotics to be clinically superior to loperamide in treatment of mild/moderate TD²².

In discussions concerning antibiotics for $TD^{11,23-25}$, limited attention has been given to resistance among diarrheagenic Escherichia coli (DEC), the most common TD pathogens²⁶; studies have mainly examined *Salmonella*, *Campylobacter*, and *Shigella*^{27,33}. DEC include several pathotypes: enteroaggregative (EAEC), enteropathogenic (EPEC), enterotoxigenic (ETEC), enteroinvasive (EIEC), enterohemorrhagic (EHEC) or shiga-toxin-producing (STEC) *E. coli*³⁴. The paucity of resistance studies can be explained by the challenges in detecting the various DEC: as they in culture resemble any other *E. coli*, identifying a specific DEC type requires additional screening by PCR or other methods³⁵.

Resistance has been reported among DEC in LMIC against the antibiotics currently recommended for TD treatment, but for travel-acquired DEC, the rates are only provided by a few studies. Ouyang-Latimer et al. showed already 2011 a substantial increase in MIC values for ciprofloxacin and azithromycin between 1997 and 2006–08 among both EAEC and ETEC isolates from travelers to Mexico, Guatemala and India³⁶. Moreover, travel-acquired ESBL-EAEC and ESBL–ETEC have been detected³⁷⁻³⁹. ESBL-DEC are of special interest, since for severely ill travelers hospitalized, first-line intravenous drugs include third generation cephalosporins (3GC) ineffective against these pathogens⁴⁰. Emergence of MDR strains among DEC is not unexpected – a similar development has been reported for other stool bacteria such as *Salmonellae*²⁸. Scarcity of research into travel-acquired ESBL-DEC prompted us to revisit our data on 413 Finnish travelers to investigate the frequency of ESBL producers among various DEC. Since our samples were collected ten years ago and the global AMR situation is constantly deteriorating, to get a more accurate picture, we also screened the literature for investigations into ESBL producers, resistance of travel-acquired DEC to commonly used antibiotics, and rates of ESBL-DEC among locals in LMIC. Research into the resistance of TD pathogens provides fundamental information for guidance on antibiotic treatment of TD.

MATERIALS AND METHODS

The first part of this two-faceted study explored the rates and geographic origin of ESBL-producing strains among DEC contracted by Finnish travelers to LMIC (Figure 1). The second part searched PubMed for original studies of DEC exploring proportions of ESBL producers (travelers and locals) and resistance to commonly used TD antibiotics (only travelers).

Study design, volunteers, samples, and travel destinations

We prospectively recruited 526 Finnish travelers attending pre-travel consultation at the Travel Clinic of Aava Medical Center before their journey outside the Nordic countries for more than four nights³. Of these, 413 met our inclusion criteria (provided pre- and post-travel stools, filled in pre- and post-travel questionnaires, travel destination in LMIC). The details of stool collection, questionnaires and categorization of travel destinations have been described in our previous study³.

Post-travel ESBL-producing *Enterobacteriaceae* (ESBL-PE) were considered as travel-acquired only if pre-travel samples had been negative for ESBL-PE.

The protocol was approved by the Helsinki University Hospital ethics committee. All subjects provided written informed consent.

Collection of specimens

Briefly, fecal samples were collected before departure and from the first or second stools passed after returning home. For collection we used swabs in Copan M40 Transystem tubes (Copan Diagnostics, Brescia, Italy). Once the samples arrived, total nucleic acids were extracted using the standard semiautomated protocol of easyMAG (bioMérieux, Marcy l'Etoile, France) and the stools were cultured (see below).

Identification of ESBL-PE

As described earlier³, ESBL-PE were isolated and characterized using established methods with culture on chromID ESBL (BioMérieux, Marcy-l'étoile, France), followed by double-disk synergy (Oxoid, Thermo Fisher Scientific, Hampshire, UK) test for cefotaxime, ceftazidime, and cefpodoxime (30 µg each), alone or with clavulanic acid (10µg), and species identification by Vitek GN (BioMérieux). Susceptibility testing for ciprofloxacin, cotrimoxazole, nitrofurantoin, tobramyćin, ertapenem, imipenem, and meropenem was conducted with E-test (BioMérieux) according to criteria set by the European Committee on Antimicrobial Susceptibility Testing EUCAST 5.0 (2018; www.eucast.org). Finally, beta-lactamase genes (TEM, OXA, SHV, CTX-M) and plasmid-mediated AmpC beta-lactamase genes (DHA, CIT) were identified by multiplex PCR⁴¹. The co-resistance rates⁴², prevalence of beta-lactamase genes³, and phylogroup characterization⁴³ of the ESBL-PE strains have been reported in our previous papers.

Analysis of DEC by qPCR

To explore the proportion of ESBL producers among various DEC (Figure 1), we first explored the total rates of stool samples positive for DEC by a multiplex qPCR assay which identifies nine bacterial pathogens: *Salmonella, Yersinia, Campylobacter, Vibrio cholerae, Shigella*/EIEC, EHEC, ETEC, EAEC, and EPEC⁴⁴. Second, to identify ESBL-DEC in the same samples, we subjected the ESBL-EC isolates to the multiplex qPCR for DEC.

Search for articles in PubMed

We searched PubMed for "ESBL" or "extended-spectrum beta-lactamase" or "CTX" combined with "diarrh(o)eagenic", "enteroaggregative", "enteropathogenic", "enterotoxigenic", "enteroinvasive", "enteroh(a)emorrhagic", "shiga-toxin-producing", or "verocytoxigenic", "DEC", "ETEC", EAEC", "EPEC", "EIEC", "EHEC", "STEC", or "VTEC" and "est", "elt", "eae", "aggR", "bfpA", "ipaH"and "stx", plus selected articles in our own collections that reported ESBLproduction among the various DEC in human samples. Although *Shigella* and EIEC often cannot be distinguished by qPCR, we did not collect resistance data from studies reporting the ESBLproducing strains as *Shigella*.

RESULTS

Participants

Eight of the 13 participants with ESBL-DEC (61.5%) had traveled to South Asia, and three (23.1%) to Southeast Asia. None of the visitors to Sub-Saharan Africa or Latin America had ESBL-DEC.

ESBL producers among DEC

The rate of ESBL-EC was 2.7% (13/475) among all DEC strains; 5.7% (10/175) among EAEC, 1.1% (2/180) among EPEC, 1.3% (1/80) among ETEC, and 0% among EHEC (0/35) or *Shigella*/EIEC (0/5) strains (Table 2). EIEC and *Shigella* are indistinguishable in the qPCR assay, but as the same samples proved negative in *Shigella* culture, the isolates were considered as EIEC.

Among strains originating in South Asia, 8.3% (1/12) of ETEC and 3.3% (1/30) of EPEC produced ESBL. The highest frequencies of ESBL-EAEC were seen for South Asia (6/33; 18.2%), Southeast Asia (3/33; 9.1%), and North Africa and the Middle East (1/3; 33.3%).

Two volunteers had taken antibiotics (ciprofloxacin) for TD; both had an ESBL-DEC co-resistant to ciprofloxacin and tobramycin, whereas among those without antibiotic use, only one strain (1/11; 9.1%) was co-resistant to ciprofloxacin (Supplementary Table 1).

ESBL genes

A total of 8/13 (61.5%) of the ESBL-DEC had $bla_{CTX-M-15}$. The genes characterized for the nine ESBL-EAEC strains were $bla_{CTX-M-1}$ (5/9), $bla_{CTX-M-9}$ (3/9), bla_{TEM} (4/9), and bla_{SHV} (1/9); for the two ESBL-EPEC strains bla_{TEM} (2/2) and $bla_{CTX-M-1}$ (2/2); and the only ESBL-ETEC $bla_{CTX-M-1}$ (1/1) (Supplementary Table 2). Six of nine ESBL-DEC harbored genes of two types.

Literature on resistance among DEC, special focus on rates of ESBL-DEC

In our literature search for studies of ESBL-DEC, we omitted those not reporting total number of DEC^{43,45} or strain-specific travel data^{46,47}; these reports prove existence of ESBL-DEC, though. Instead, we selected, in accord with our initial aim, papers providing prevalence data on resistance among travel-acquired DEC or rates of ESBL-DEC among DEC originating in LMIC. Due to meager search results especially among travelers, we also reviewed our own files on TD studies.

Our search only yielded 24 original studies of ESBL-DEC rates among one or more types of DEC, three traveler studies³⁷⁻³⁹ (Table 3), and 21 looking at locals in LMIC⁴⁸⁻⁶⁸ (Table 4). As for travelers, we found four other investigations into resistance rates to 3GC^{36, 69-71}. In total, 13 traveler studies provided resistance rates to one or more TD antibiotics^{30,36-39,69-76}, all presented below by DEC pathotype.

Resistance among EAEC strains

Eight traveler studies describe resistance among EAEC strains (Table 3). Guiral et al. report for Spanish travelers with TD ESBL-EAEC rates of 10% (among 51 EAEC isolates in 2005–06) and 13% (39 EAEC in 2011–17)^{37,39}.

Among samples from language school students in Peru (2003–10), 11% of the EAEC isolates proved resistant to 3GC^{70} , for travelers to Mexico/Guatemala and India the figures were 20% and 0%, respectively (2006–08)³⁶, and for the US military in Thailand 0% (2013–17)⁷¹

Among travel-acquired EAEC, resistance rates of 0–42% have been reported to fluoroquinolones (eight articles^{30,36,39,69-71,73,76}); 0–61% to azithromycin (six articles^{30,36,39,69,70,76}), and 0% to rifaximin (three articles^{36,39,69}).

The seven LMIC investigations show rates of 11–85% for ESBL-EAEC among EAEC (Table 4)^{48,50,54,56,57,61,62}.

Resistance among ETEC strains

We found 12 resistance studies of travel-acquired $\text{ETEC}^{30,36,38,39,69-76}$ (Table 3). For ESBL-ETEC a rate of 14% was reported among 43 ETEC isolates from Spanish travelers in 2011–17³⁷ and a rate of 15% among 265 ETEC isolates (from travelers and locals) from Kathmandu, Nepal in 2001–16³⁸. Among the most recently acquired strains, the resistance rates amounted to 34–35%³⁸.

Of the three studies reporting resistance to 3GC, a rate of 0% was recorded for language school students in Peru 2003– 10^{70} and US military in Thailand⁷¹, and 5% and 6% for travelers to Mexico/Guatemala and India, respectively, in 2006– 08^{36} .

Resistance among ETEC to fluoroquinolones was explored in 12 traveler studies, showing rates of $0-33\%^{30,36,38,39,69-76}$; seven studies explored resistance to azithromycin with rates of 0-29% 30,36,39,69,70,75,76 ; and three to rifaximin with rates of 0-20% 36,39,69 .

The three investigations among locals in LMIC showed among ESBL-ETEC rates of 18% in India⁷⁰ and 75% and 100% in Iran^{57,62} (Table 4).

Resistance among EPEC strains

Our search yielded four traveler studies of EPEC strains (Table 3). In Nepal 2001–03⁷⁵ and 2012–14⁷⁶ ESBL-EPEC were not covered, but resistance rates of 10% and 23% to fluoroquinolones, and 37% and 67% to azithromycin, were seen, respectively. Among US military in Thailand in 2002–04, resistance rates (ESBL-EPEC not covered) of 0% and 13% were recorded to fluoroquinolones and azithromycin³⁰, and in 2013–17 8% to ciprofloxacin⁷¹.

Among locals the six studies reported rates of 11–80% for ESBL-EPEC^{52,53,55,57,58,60,63} (Table 4).

Resistance among EHEC/STEC strains

None of the traveler studies reviewed provided rates of antibiotic resistance for EHEC/STEC isolates.

Amaya et al. did not find any ESBL-EC among eight EHEC strains from Nicaraguan children with diarrhea⁴⁹ (Table 4).

Resistance among EIEC strains

Our search yielded two traveler studies of resistance looking at EIEC isolates: among samples from US military in Thailand 2013–17 no resistance was detected⁷¹ but in Nepal 2012–14, 10% of the EIEC strains proved resistant to ciprofloxacin and 30% to azithromycin⁷⁶

In LMIC, studies among local children with diarrhea have found the few EIEC strains to be mostly ESBL producers^{49,57,60,68,77}.

DISCUSSION

Despite the vast discussion around antibiotic use for treating TD, paradoxically scant attention has been paid to resistance among the most common TD pathogens, DEC. The handful of reports published mostly do not focus on travelers. Apart from resistance to individual antibiotics, multidrug resistance is increasingly common among intestinal bacteria in clinical samples worldwide, ESBL-PE ranking as the most prevalent MDR type⁷⁸⁻⁸⁰. Our data together with those from a literature search for studies among travelers and locals in LMIC destinations show an emergence of ESBL producers among DEC.

Rates of ESBL producers among DEC

Our rate, 3–7% of ESBL producers among the various DEC strains collected 2009–10, appears consistent with the three other traveler studies of ESBL-DEC: among Spanish travelers, the rates of ESBL-EAEC were 10% in $2005-2006^{37}$, and 12.8% in $2011-17^{39}$. Among residents and travelers with acute diarrhea in Kathmandu an increase from 1.5% to 35% was observed between 2008 and 2016³⁸. These data suggest increasing rates of ESBL producers among DEC.

We found more investigations into the ESBL-production of DEC among locals in LMIC than among travelers, with rates of positive findings varying by pathotype, time and destination between 0% and 80% ⁴⁸⁻⁶⁸. It should be noted that none of the analyses focused on the main tourist destinations in Southeast Asia, Africa, or South and Central America, and the Caribbean. In 18 of the 21 studies the data were from local children with or without diarrhea^{48-50,52,53,55-62,64-68}, highlighting the clinical concern related to resistance. Likewise, among locals, the highest rates were recorded over the most recent years, according with the steady global increase in the rates of ESBL-producing strains among all *E. coli* in clinical samples⁷⁸⁻⁸⁰.

Our search did not focus on carbapenemase-producing DEC, but we found 16 studies from LMIC reporting resistance rates of 0-50% to carbapenems among DEC^{49-61,53,57-68}. Our samples showed no carbapenemase-producing genes³.

ESBL producers among various DEC

In our data, the ESBL-EC rates appeared higher among EAEC than EPEC and ETEC (5.7% versus 1.1% versus 1.3%). This accords with other traveler studies reporting ESBL-EAEC rates of $10\%^{37}$ and $12.8\%^{39}$ among travelers yet amounting to 85% for locals in Iran⁵⁷ and 56% in China⁵⁴.

Likewise, substantial rates (53% and 57%) of ESBL-EAEC have been reported among clinical EAEC isolates in England; yet they do not report which of the strains were travel-acquired nor their countries origin ^{46,47}.

For ESBL-ETEC, our rate, 1.3% (Table 2), was much lower than that found among Spanish travelers $(14\%)^{39}$ or in Nepal $(15\%)^{38}$. The top rates (75%) for non-travelers have been recorded among Iranian children⁵⁷.

As for EPEC, we only identified two ESBL-EPEC strains (1.1%). None of the traveler studies reviewed covered ESBL-EPEC, but among locals rates as high as 80% have been reported in Iran⁵⁸, and 59% in Pakistan⁵⁵.

We detected no ESBL-EC among EIEC and EHEC, neither did we find in the literature any other traveler studies exploring ESBL-EC of these pathotypes; only few investigations among locals report ESBL-EC for EIEC or EHEC^{49,57,60,68,77}.

We found no more than two studies looking at the rates from the other angle, describing the rates of a given pathotype *among travel-acquired ESBL-DEC*: rates of 14% in 2009– 10^{43} and 57% in 2017– 18^{45} have been shown for ESBL-DEC.

Geographic distribution of ESBL-DEC

Most of our ESBL-DEC originated in South Asia which also proved to have the highest rates of ESBL-DEC among DEC: 18.2% of EAEC strains were ESBL producers. Indeed, South Asia also has exceptionally high resistance rates among gram-negative bacteria in clinical samples^{81,82} and top ESBL-PE colonization rates among visitors^{2-6,83,84}. Our data agree with previous data showing higher resistance rates among EAEC strains from South or Southeast Asia (33.3%; 4/12) than those from Africa (6.3%; 1/16) and Latin America (0%; 0/11)³⁹.

Resistance to commonly used TD antibiotics

While our own results center around ESBL-DEC, we also reviewed the literature for data on resistance among travel-acquired DEC to commonly used TD antibiotics (fluoroquinolones, azithromycin, and rifaximin). Recent traveler studies^{39,76} present alarming data: for EAEC strains resistance rates of 15–42%, 33–61%, and 0% to fluoroquinolones, azithromycin and rifaximin, and for ETEC 23–33%, 22–29%, and 0%, respectively.

Resistance genes among DEC

Our data include thirteen ESBL-DEC isolates, with *bla*_{CTX-M-1} as the most common finding in genetic analyses, followed by *bla*_{TEM}. Only a small proportion of our strains carried the *bla*_{CTX-M-15} gene despite the worldwide spread of *E. coli* clone of sequence type 131 (ST131) carrying the CTX-M-15 ESBL both in clinical and non-clinical settings⁸⁵. In contrast, a previous traveler study³⁶ reports a total of 11 ESBL-DEC strains, all harboring either of the two genes *bla*_{CTX-M-15} or *bla*_{CTX}. M-27. Likewise, from the samples of residents and travelers in Nepal,³⁸ *bla*_{CTX-M-15} was detected in 80% of the ESBL-ETEC strains.

Clinical implications

While ESBL-EC are considered resistant to 3GC (e.g. ceftriaxone), the resistance profile as such does not cover the most commonly used TD regimens, i.e. fluoroquinolones, azithromycin, and rifaximin. Unfortunately, however, ESBL-producing strains often harbor co-resistance to other antibiotics, especially fluoroquinolones^{86,87}. Of our ESBL-DEC strains, 3/13 (23.1%) were co-resistant to fluoroquinolones, yet higher co-resistance rates have been reported among travel-acquired ESBL-PE in general, particularly for South Asia^{2,5,42,45,83} and related to fluoroquinolone intake abroad⁴². Indeed, ESBL-producing strains are of special concern, since in cases severe enough to require hospitalization empiric treatment often relies on either 3GC or fluoroquinolones⁴⁰.

Interpreting the efficacy of various antibiotics is somewhat complicated, for fecal antibiotic levels tend to exceed the minimum inhibitory concentration (MIC)²³. Furthermore, presence of antibiotics in stools, while indicating an antibiotic pressure to other intestinal bacteria, may also drive transfer of resistance genes to other *Enterobacteriaceae*, some of which are potential pathogens^{86,88}.

An ineffective drug does not offer benefits, and yet retains its disadvantages. Although the adverse effects rate appears to be low⁸⁹, recently, for example, the US Food and Drug Administration has warned about some serious adverse effects of fluoroquinolones (e.g. tendinitis and prolonged QT interval) and azithromycin (e.g. prolonged QT interval)^{90,91}, the most popular TD antibiotics. Furthermore, data are lacking on the suggested smaller impact of one-day antibiotic treatment on acquisition of MDR bacteria abroad. The adverse effect profile would favor rifamycins such as rifaximin. However, the drug is non-absorbable and should not be used in cases with fever and invasive disease – i.e. it does not meet the most important indications for antibiotics. We only found

a few studies exploring resistance rates to rifaximin among TD pathogens; Ouyang-Latimer et al.³⁶ reported 16-25% resistance rates among ETEC already in 2011.

Limitations of our data

Firstly, collected 2009–10, our strains do not fully represent the current situation. Unfortunately, though, the same applies to the other traveler studies found in our search, only three of which provide data from a later time period^{38,39,76}. The increase in resistance recorded among locals suggests growing pressure also for travelers. Our data may thus present a slight underestimation, calling for updated surveillance.

Secondly, qPCR of stools cannot distinguish whether the samples contain one DEC strain or several of similar type. Likewise, in culturing ESBL-EC strains, those which appear phenotypically different are picked, and therefore strains may be missed that are similar or of only a slightly different phenotype, but genetically unlike. Fortunately, these sources of error may at least partly overcome one another.

Thirdly, in the various studies reviewed there are methodological differences (assessment of the various DEC, pre-analytical handling of the specimens etc.), therefore the data may not be fully comparable.

Conclusions

ESBL-producing DEC are no longer rare, particularly in Asia. Among travel-acquired DEC, their rates appear fairly low as yet, but in many regions increase is already seen among DEC isolated from locals with acute diarrhea, also portending increase among travel-acquired DEC, many strains even to be carried by travelers to their countries. While antibiotics certainly retain their place in the treatment of the most severe TD cases, data showing increasing resistance among stool pathogens further encourage cutting back on use of antibiotics for TD, and opting for non-antibiotic alternatives for mild and moderate cases. After all, an ineffective drug, while obviously useless, retains all its disadvantages.

REFERENCES

1. O'Neill J. 2016. Tackling drug-resistant infections globally: final report and recommendations.

 Paltansing S, Vlot JA, Kraakman ME, et al. Extended-Spectrum beta-Lactamase-producing *Enterobacteriaceae* among Travelers from the Netherlands. Emerg Infect Dis 2013: 19:1206-1213.

3. Kantele A, Lääveri T, Mero S, et al. Antimicrobials increase travelers' risk of colonization by extended-spectrum beta-lactamase-producing *Enterobacteriaceae*. Clin Infect Dis 2015; 60:837-846.

4. Ruppé E, Armand-Lefevre L, Estellat C, et al. High Rate of Acquisition but Short Duration of Carriage of Multidrug-Resistant *Enterobacteriaceae* After Travel to the Tropics. Clin Infect Dis 2015; 61:593-600.

5. Reuland EA, Sonder GJ, Stolte I, et al. Travel to Asia and traveller's diarrhoea with antibiotic treatment are independent risk factors for acquiring ciprofloxacin-resistant and extended spectrum beta-lactamase-producing *Enterobacteriaceae*-a prospective cohort study. Clin Microbiol Infect 2016;22:731.e1-731.e7. 10.1016/j.cmi.2016.05.003 .

6. Arcilla MS, van Hattem JM, Haverkate MR, et al. Import and spread of extended-spectrum betalactamase-producing *Enterobacteriaceae* by international travellers (COMBAT study): a prospective, multicentre cohort study. Lancet Infect Dis 2017; 17:78-85.

7. Woerther PL, Andremont A, Kantele A. Travel-acquired ESBL-producing *Enterobacteriaceae*: impact of colonization at individual and community level. J Travel Med 2017; 24: S29-S34.

8. Editorial. Prescribing antibiotics: a battle of resistance. Lancet 2014; 384:558-6736(14)61350-7.
10.1016/S0140-6736(14)61350-7.

9. Holmes AH, Moore LS, Sundsfjord A, et al. Understanding the mechanisms and drivers of antimicrobial resistance. Lancet (London, England) 2016;387:176-187.

10. Kantele A, Kuenzli E, Dunn SJ, et al. Dynamics of intestinal multidrug-resistant bacteria colonisation contracted by visitors to a high-endemic setting: a prospective, daily, real-time sampling study, Lancet Microbe, 2021. DOI: 10.1016/S2666-5247(20)30224-X

11. Vilkman K, Lääveri T, Pakkanen SH, Kantele A. Stand-by antibiotics encourage unwarranted use of antibiotics for travelers' diarrhea: A prospective study. Travel Med Infect Dis 2019; 27:64-71

12. Hill DR. Occurrence and self-treatment of diarrhea in a large cohort of Americans traveling to developing countries. Am J Trop Med Hyg 2000; 62:585-589.

13. Pitzurra R, Steffen R, Tschopp A, Mutsch M. Diarrhoea in a large prospective cohort of European travellers to resource-limited destinations. BMC Infect Dis 2010; 10: 231

14. Belderok SM, van den Hoek A, Kint JA, et al. Incidence, risk factors and treatment of diarrhoea among Dutch travellers: reasons not to routinely prescribe antibiotics. BMC Infect Dis 2011;11:295.

15. Soonawala D, Vlot JA, Visser LG. Inconvenience due to travelers' diarrhea: a prospective follow-up study. BMC Infect Dis 2011; 11:322

16. Lalani T, Maguire JD, Grant EM, et al. Epidemiology and self-treatment of travelers' diarrhea in a large, prospective cohort of department of defense beneficiaries. J Travel Med 2015; 22:152-160.

17. Stoney RJ, Han PV, Barnett ED, et al. Travelers' Diarrhea and Other Gastrointestinal Symptoms Among Boston-Area International Travelers. Am J Trop Med Hyg 2017; 96:1388-1393.

18. Gandhi AR, Rao SR, Chen LH, et al. Prescribing Patterns of Antibiotics for the Self-Treatment of Travelers' Diarrhea in Global TravEpiNet, 2009-2018. Open Forum Infect Dis 2020; 7:ofaa376.

19. Turunen K, Kantele A. Revisiting travellers' diarrhoea justifying antibiotic treatment: prospective study. J Travel Med 2020; Dec 28:taaa237. doi: 10.1093/jtm/taaa237.

20. De Bruyn G, Hahn S, Borwick A. Antibiotic treatment for travellers' diarrhoea. Cochrane Database Syst Rev 2000, (3):CD002242. doi(3):CD002242.

21. Riddle MS, Connor BA, Beeching NJ, et al. Guidelines for the prevention and treatment of travelers' diarrhea: a graded expert panel report. Journal of travel medicine 2017;24:S57-S74.

22. Lääveri T, Sterne J, Rombo L, et al. Systematic review of loperamide: No proof of antibiotics being superior to loperamide in treatment of mild/moderate travellers' diarrhoea. Travel Med Infect Dis 2016, 14(4):299-312.

23. Keystone JS, Connor BA. Antibiotic self-treatment of travelers' diarrhea: It only gets worse! Travel Med Infect Dis 2017; 16:1-2.

24. Kantele A. A call to restrict prescribing antibiotics for travellers' diarrhea--Travel medicine practitioners can play an active role in preventing the spread of antimicrobial resistance. Travel Med Infect Dis 2015; 13:213-214.

25. Ericsson CD, Riddle MS. Should travel medicine practitioners prescribe antibiotics for self-treatment of travelers' diarrhea? J Travel Med 2018; 25

26. Tribble DR. Resistant pathogens as causes of traveller's diarrhea globally and impact(s) on treatment failure and recommendations. J Travel Med 2017; 24:S6-S12.

27. Barlow RS, Debess EE, Winthrop KL, et al. Travel-associated antimicrobial drug-resistant nontyphoidal *Salmonellae*, 2004-2009. Emerg Infect Dis 2014; 20:603-611.

28. Gunell M, Aulu L, Jalava J et al. Cefotaxime-resistant Salmonella enterica in travelers returning from Thailand to Finland. 2014; Emerg Infect Dis 20:1214-1217.

29. Brown JD, Willcox SJ, Franklin N, et al. *Shigella* species epidemiology and antimicrobial susceptibility: the implications of emerging azithromycin resistance for guiding treatment, guidelines and breakpoints. J Antimicrob Chemother 2017; 72: 3181-3186.

30. Mason CJ, Sornsakrin S, Seidman JC, et al. Antibiotic resistance in *Campylobacter* and other diarrheal pathogens isolated from US military personnel deployed to Thailand in 2002-2004: a case-control study. Trop Dis Travel Med Vaccines 2017; 3:13

31. Post A, Martiny D, van Waterschoot N, et al. Antibiotic susceptibility profiles among*Campylobacter* isolates obtained from international travelers between 2007 and 2014. Eur J ClinMicrobiol Infect Dis 2017; 36:2101-2107.

32. Williamson DA, Lane CR, Easton M, et al. Increasing Antimicrobial Resistance in Nontyphoidal *Salmonella* Isolates in Australia from 1979 to 2015. Antimicrob Agents Chemother
2018; 62

33. Brown AC, Chen JC, Watkins LKF, et al. CTX-M-65 Extended-Spectrum beta-Lactamase-Producing *Salmonella* enterica Serotype Infantis, United States(1). Emerg Infect Dis 2018; 24:2284-2291.

34. Nataro JP, Kaper JB. Diarrheagenic Escherichia coli. Clin Microbiol Rev 1998; 11:142-201.

35. Vila J. New molecular diagnostic tools in traveller's diarrhea. J Travel Med 2017; 24:S23-S28.10.1093/jtm/taw071 .

36. Ouyang-Latimer J, Jafri S, VanTassel A, et al. In vitro antimicrobial susceptibility of bacterial enteropathogens isolated from international travelers to Mexico, Guatemala, and India from 2006 to 2008. Antimicrob Agents Chemother 2011; 55:874-878.

37. Guiral E, Mendez-Arancibia E, Soto SM, et al. CTX-M-15-producing enteroaggregative *Escherichia coli* as cause of travelers' diarrhea. Emerg Infect Dis 2011; 17:1950-1953.

38. Margulieux KR, Srijan A, Ruekit S, et al. Extended-spectrum beta-lactamase prevalence and virulence factor characterization of enterotoxigenic *Escherichia coli* responsible for acute diarrhea in Nepal from 2001 to 2016. Antimicrob Resist Infect Control 2018; 7:87

39. Guiral E, Goncalves Quiles M, Munoz L, et al. Emergence of Resistance to Quinolones and beta-Lactam Antibiotics in Enteroaggregative and Enterotoxigenic *Escherichia coli* Causing Traveler's Diarrhea. Antimicrob Agents Chemother 2019; 63: e01745-18.

40. Ryan ET. Treatment and prevention of enteric (typhoid and paratyphoid) fever. In: Uptodate. 2020; https://www.uptodate.com/contents/treatment-and-prevention-of-enteric-typhoid-and-paratyphoid-fever

41. Dallenne C, Da Costa A, Decre D, et al. Development of a set of multiplex PCR assays for the detection of genes encoding important beta-lactamases in *Enterobacteriaceae*. J Antimicrob Chemother 2010; 65:490-495.

42. Kantele A, Mero S, Kirveskari J, Lääveri T. Fluoroquinolone antibiotic users select fluoroquinolone-resistant ESBL-producing *Enterobacteriaceae* (ESBL-PE) - Data of a prospective traveller study. Travel Med Infect Dis 2017; 16:23-30.

43. Kantele A, Lääveri T, Mero S, et al. Despite Predominance of Uropathogenic/Extraintestinal
Pathotypes Among Travel-acquired Extended-spectrum beta-lactamase-producing *Escherichia coli*,
the Most Commonly Associated Clinical Manifestation Is Travelers' Diarrhea. Clin Infect Dis 2019;
70: 210-218

44. Antikainen J, Kantele A, Pakkanen SH, et al. A quantitative polymerase chain reaction assay for rapid detection of 9 pathogens directly from stools of travelers with diarrhea. Clin Gastroenterol Hepatol 2013; 11:1300-1307.

45. Ljungquist O, Camporeale A, Nematzadeh S, et al. ESBL-producing Enterobacterales in patients with traveller's diarrhoea - a cross sectional cohort study. Antimicrob Agents Chemother 2020; AAC.01585-20.

46. Do Nascimento V, Day MR, Doumith M, et al. Comparison of phenotypic and WGS-derived antimicrobial resistance profiles of enteroaggregative *Escherichia coli* isolated from cases of diarrhoeal disease in England, 2015-16. J Antimicrob Chemother 2017; 72:3288-3297.

47. Boxall MD, Day MR, Greig DR, Jenkins C. Antimicrobial resistance profiles of diarrhoeagenic *Escherichia coli* isolated from travellers returning to the UK, 2015-2017. J Med Microbiol 2020; 69:932-943.

48. Sonnevend A, Al Dhaheri K, Mag T, et al. CTX-M-15-producing multidrug-resistant enteroaggregative *Escherichia coli* in the United Arab Emirates. Clin Microbiol Infect 2006; 12:582-585.

49. Amaya E, Reyes D, Vilchez S, et al. Antibiotic resistance patterns of intestinal *Escherichia coli* isolates from Nicaraguan children. J Med Microbiol 2011; 60:216-222.

50. Khoshvaght H, Haghi F, Zeighami H. Extended spectrum betalactamase producing
Enteroaggregative *Escherichia coli* from young children in Iran. Gastroenterol Hepatol Bed Bench
2014; 7:131-136.

51. Ghorbani-Dalini S, Kargar M, Doosti A, et al. Molecular Epidemiology of ESBL Genes and Multi-Drug Resistance in Diarrheagenic *Escherichia coli* Strains Isolated from Adults in Iran. Iran J Pharm Res 2015; 14:1257-1262.

52. Memariani M, Najar Peerayeh S, Zahraei Salehi T, Shokouhi Mostafavi SK. Occurrence of SHV, TEM and CTX-M beta-Lactamase Genes Among Enteropathogenic *Escherichia coli* Strains Isolated From Children With Diarrhea. Jundishapur J Microbiol 2015; 8:e15620.

53. Malvi S, Appannanavar S, Mohan B, et al. Comparative analysis of virulence determinants, antibiotic susceptibility patterns and serogrouping of atypical enteropathogenic *Escherichia coli* versus typical enteropathogenic *E. coli* in India. J Med Microbiol 2015; 64:1208-1215.

54. Wang Y, Wu J, Cao Y. The extended spectrum beta-lactamases (ESBL) and virulence genes of intestinal enteroaggregative *Escherichia coli* (EAEC) in healthy elderly individuals. Int J Clin Exp Med 2015; 8:20953-20958.

55. Younas M, Siddiqui F, Noreen Z, et al.. Characterization of enteropathogenic *Escherichia coli* of clinical origin from the pediatric population in Pakistan. Trans R Soc Trop Med Hyg 2016; 110:414-420.

56. Khalil U, Younus M, Asghar N, et al. Phenotypic and genotypic characterization of enteroaggregative *Escherichia coli* isolates from pediatric population in Pakistan. APMIS 2016; 124:872-880.

57. Aminshahidi M, Arastehfar A, Pouladfar G, et al. Diarrheagenic *Escherichia coli* and *Shigella* with High Rate of Extended-Spectrum Beta-Lactamase Production: Two Predominant Etiological Agents of Acute Diarrhea in Shiraz, Iran. Microb Drug Resist 2017; 23:1037-1044.

58. Karami P, Bazmamoun H, Sedighi I, et al. Antibacterial resistance patterns of extended spectrum beta-lactamase -producing enteropathogenic *Escherichia coli* strains isolated from children. Arab J Gastroenterol 2017; 18:206-209

59. Konate A, Dembele R, Guessennd NK, et al. Epidemiology and Antibiotic Resistance Phenotypes of Diarrheagenic *Escherichia coli* Responsible for Infantile Gastroenteritis in Ouagadougou, Burkina Faso. Eur J Microbiol Immunol (Bp) 2017; 7:168-175.

60. Mandal A, Sengupta A, Kumar A, et al. Molecular Epidemiology of Extended-Spectrum beta-Lactamase-Producing *Escherichia coli* Pathotypes in Diarrheal Children from Low Socioeconomic Status Communities in Bihar, India: Emergence of the CTX-M Type. Infect Dis (Auckl) 2017; 10:1178633617739018.

61. Amin M, Sirous M, Javaherizadeh H, et al. Antibiotic resistance pattern and molecular characterization of extended-spectrum beta-lactamase producing enteroaggregative *Escherichia coli* isolates in children from southwest Iran. Infect Drug Resist 2018; 11:1097-1104.

62. Mahdavi Broujerdi S, Roayaei Ardakani M, Rezatofighi SE. Characterization of diarrheagenic *Escherichia coli* strains associated with diarrhea in children, Khouzestan, Iran. J Infect Dev Ctries 2018; 12:649-656.

63. Xu Y, Sun H, Bai X, et al.. Occurrence of multidrug-resistant and ESBL-producing atypical enteropathogenic *Escherichia coli* in China. Gut Pathog 2018; 10:8

64. Zhou Y, Zhu X, Hou H, et al. Characteristics of diarrheagenic *Escherichia coli* among children under 5 years of age with acute diarrhea: a hospital based study. BMC Infect Dis 2018; 18:63-017-2936-1.

65. Moharana SS, Panda RK, Dash M, et al. Etiology of childhood diarrhoea among under five children and molecular analysis of antibiotic resistance in isolated enteric bacterial pathogens from a tertiary care hospital, Eastern Odisha, India. BMC Infect Dis 2019; 19:1018-019-4501-6.

66. Taghadosi R, Shakibaie MR, Hosseini-Nave H. Antibiotic resistance, ESBL genes, integrons, phylogenetic groups and MLVA profiles of *Escherichia coli* pathotypes isolated from patients with diarrhea and farm animals in south-east of Iran. Comp Immunol Microbiol Infect Dis 2019; 63:117-126.

67. Eltai NO, Al Thani AA, Al Hadidi SH, et al. 2020. Antibiotic resistance and virulence patterns of pathogenic *Escherichia coli* strains associated with acute gastroenteritis among children in Qatar. BMC Microbiol 20:54-020-01732-8.

68. Farajzadeh-Sheikh A, Savari M, Hosseini Nave H, et al. Frequency and molecular epidemiology of class A ESBLs producing Enteroinvasive *Escherichia coli* (EIEC) isolates among patients with diarrhea. Gastroenterol Hepatol Bed Bench 2020; 13:77-85.

69. Gomi H, Jiang ZD, Adachi JA, et al. In vitro antimicrobial susceptibility testing of bacterial enteropathogens causing traveler's diarrhea in four geographic regions. Antimicrob Agents Chemother 2001; 45:212-216.

70. Jennings MC, Tilley DH, Ballard SB, et al. Case-Case Analysis Using 7 Years of Travelers' Diarrhea Surveillance Data: Preventive and Travel Medicine Applications in Cusco, Peru. Am J Trop Med Hyg 2017; 96:1097-1106. 71. Lurchachaiwong W, Serichantalergs O, Lertsethtakarn P, et al. Enteric etiological surveillance in acute diarrhea stool of United States Military Personnel on deployment in Thailand, 2013-2017. Gut Pathog 2020; 12:17

72. Vila J, Vargas M, Ruiz J et al. Quinolone resistance in enterotoxigenic *Escherichia coli* causing diarrhea in travelers to India in comparison with other geographical areas. Antimicrob Agents Chemother 2000; 44:1731-1733.

73. Mendez Arancibia E, Pitart C, Ruiz J, et al. Evolution of antimicrobial resistance in enteroaggregative *Escherichia coli* and enterotoxigenic *Escherichia coli* causing traveller's diarrhoea. J Antimicrob Chemother 2009; 64:343-347.

74. Porter CK, Riddle MS, Tribble DR, et al. The epidemiology of travelers' diarrhea in Incirlik, Turkey: a region with a predominance of heat-stabile toxin producing enterotoxigenic *Escherichia coli*. Diagn Microbiol Infect Dis 2010; 66:241-247.

75. Pandey P, Bodhidatta L, Lewis M, et al.. Travelers' diarrhea in Nepal: an update on the pathogens and antibiotic resistance. J Travel Med 2011; 18:102-108.

76. Murphy H, Bodhidatta L, Sornsakrin S, et al. Traveler's Diarrhea in Nepal - Changes in Etiology and Antimicrobial Resistance. J Travel Med 2019; 26: taz054.

77. Mahdavi Broujerdi S, Roayaei Ardakani M, Rezatofighi SE. Characterization of diarrheagenic *Escherichia coli* strains associated with diarrhea in children, Khouzestan, Iran. J Infect Dev Ctries 2018; 12:649-656.

78. Ruppe E, Woerther PL, Barbier F. Mechanisms of antimicrobial resistance in Gram-negative bacilli. Ann Intensive Care 2015; 5:61

79. European Centre for Disease Prevention and Control Surveillance of antimicrobial resistance in Europe 2018. Stockholm 2019.

80. CDC Antibiotic Resistance Threats in the United States, Atlanta, GA: U.S. Department of Health and Human Services, CDC 2019.

81. Akova M. Epidemiology of antimicrobial resistance in bloodstream infections. Virulence 2016;7:252-266.

82. Chong Y, Shimoda S, Shimono N. Current epidemiology, genetic evolution and clinical impact of extended-spectrum beta-lactamase-producing *Escherichia coli* and Klebsiella pneumoniae. Infect Genet Evol 2018; 61:185-188.

83. Kuenzli E, Jaeger VK, Frei R, et al. High colonization rates of extended-spectrum betalactamase (ESBL)-producing *Escherichia coli* in Swiss travellers to South Asia- a prospective observational multicentre cohort study looking at epidemiology, microbiology and risk factors. BMC Infect Dis 2014; 14:528-2334-14-528.

84. Lubbert C, Straube L, Stein C, et al. Colonization with extended-spectrum beta-lactamaseproducing and carbapenemase-producing *Enterobacteriaceae* in international travelers returning to Germany. Int J Med Microbiol 2015; 305:148-156. 85. Nicolas-Chanoine MH, Bertrand X, Madec JY. *Escherichia coli* ST131, an intriguing clonal group. Clin Microbiol Rev 2014; 27:543-574.

86. Carattoli A. Plasmids in Gram negatives: molecular typing of resistance plasmids. Int J Med Microbiol 2011; 301:654-658.

87. Canton R, Gonzalez-Alba JM, Galan JC. CTX-M Enzymes: Origin and Diffusion. Front Microbiol 2012; 3:110.

88. Martinez JL, Baquero F. Interactions among strategies associated with bacterial infection: pathogenicity, epidemicity, and antibiotic resistance. Clin Microbiol Rev 2002; 15:647-679.

89. DuPont HL, Steffen R. Use of antimicrobial agents for treatment and prevention of travellers' diarrhoea in the face of enhanced risk of transient fecal carriage of multi-drug resistant *Enterobacteriaceae*: setting the stage for consensus recommendations. J Travel Med 2017; 24:S57-S62.

90. US Food and Drug Administration. Fluoroquinolone Antimicrobial Drugs Information. 2018.

91. US Food and Drug Administration. Azithromycin (marketed as Zithromax or Zmax) Information. 2018.

FIGURES AND TABLES

Figure 1. Flow chart of prospective study of ESBL-EC (extended-spectrum beta-lactamase-producing *Escherichia coli*) rates among DEC (diarrheagenic *E. coli*) of various pathotypes.

Abbreviations: EAEC – enteroaggregative *E. coli*, EPEC – enteropathogenic *E. coli*, ETEC – enterotoxigenic E. coli, EIEC – enteroinvasive *E. coli*, EHEC enterohaemorrhagic *E. coli* or STEC – shiga-toxin-producing (STEC) *E. coli*.



Table 1. Demographics of 13 prospectively recruited travelers who contracted extended-spectrum beta-lactamase-producing diarrheagenic *Escherichia coli* (ESBL-DEC) during visits to low- and middle-income countries (LMIC). Data are provided for concomitant other ESBL-producing *Enterobacteriaceae* (ESBL-PE), antibiotic (AB) use, travelers' diarrhea (TD), destination, length of travel and non-ESBL-PE co-pathogens.

Age (yrs)	Gender	Type of ESBL- DEC	Concomitant other ESBL-PE	AB use	TD	Travel destination(s)	Length of travel (days)	Non-ESBL co-pathogens
23	male	EAEC		no	yes	Laos, Cambodia, Vietnam	22	none
31	female	EPEC		no	yes	India	11	EAEC, Campylobacter
61	female	EPEC		FQ	yes	China	12	ETEC
56	female	EAEC		no	yes	India	7	EPEC ETEC
67	male	EAEC		no	no	Egypt, Jordan	7	none
24	female	EAEC		no	yes	Thailand, Cambodia, Vietnam	110	EPEC, Campylobacter
46	female	EAEC	non-DEC E. coli	no	yes	Cambodia	19	EPEC
47	male	EAEC		no	yes	India	16	EHEC
22	female	ETEC		no	yes	India	14	EPEC EAEC
20	male	EAEC	Klebsiella pneumoniae	FQ	yes	India	16	EPEC, Salmonella, Campylobacter
31	male	EAEC		no	yes	India	27	EPEC Campylobacter
25	male	EAEC	E. hermannii	no	yes	India	32	EPEC
59	male	EAEC		no	yes	India	13	EPEC

Table 2. Proportions of ESBL-producing *Escherichia coli* (ESBL-EC) among all DEC in samples from 413 travelers visiting LMIC. DEC were determined by multiplex qPCR directly from stools; positive result was interpreted as one strain. ESBL-DEC were identified by qPCR analysis of isolates initially obtained by culture. Table shows prevalences of various ESBL-DEC among all DEC strains (total= 475) of same type plus geographic origin as judged from stools of travelers visiting each region.

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Latin	ttps://
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ESBL-DEC°	p.col
n (%)	m/jtm
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0 (0.0)	le/dc
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	n / all 90 ESBL- EC	ESBL-DEC /all respective DEC ^a	TD ^b	South Asia	South East Asia	East Asia	North Africa and Middle East	Sub- Saharan Africa	Latin America
				ESBL-DEC ^c	ESBL-DEC ^c	ESBL-DEC ^c	ESBL-DEC ^c	ESBL-DEC ^c	ESBL-DEC
	n (%)	n (%)	n (%)	n (%)	n (%)	n (%)	n (%)	n (%)	n (%)
EAEC	10 (11.1)	10/175 (5.7)	9 (90.0)	6/33 (18.2)	3/33 (9.1)	0/1 (0,0)	1/3 (33.3)	0/90 (0.0)	0/15 (0.0)
EPEC	2 (2.2)	2/180 (1.1)	2 (100.0)	1/30 (3.3)	0/44 (0.0)	1/2 (50.0)	0/4 (0.0)	0/83 (0.0)	0/17 (0.0)
ETEC	1 (1.1)	1/80 (1.3)	1 (100.0)	1/12 (8.3)	0/19 (0.0)	0/0 (0.0)	0/0 (0.0)	0/45 (0.0)	0/4 (0.0)
Fotal	13 (14.4)	13/475 (2.7)	12 (92.3)	8 (62.0)	3 (23.1)	1 (7.7)	1 (7.7)	0 (0.0)	0 (0.0)

^aESBL producers (n) among all EAEC/ EPEC/ ETEC /DEC of 413 travelers (%)

^bamong 13 travelers with ESBL-DEC

cESBL producers (n) among all EAEC/ EPEC/ ETEC /DEC in samples of travelers to region (%)

Table 3. Results of literature search for traveler studies exploring antibiotic resistance among various DEC. Some studies were conducted among both travelers and locals in LMIC. Table combines results from analyses of ESBL-DEC and resistance to TD antibiotics, fluoroquinolones, azithromycin, and rifaximin. Three studies only report resistance rates to 3rd generation cephalosporins but not ESBL-DEC. (NT=not tested)

Year(s) of	Population,	ESBL-EC	Cipro-	Azithro-	Rifaximin
stool	number		floxacin	mycin	resistance
sampling	of isolates		resistance	resistance	
2013-17	US military, Thailand	only resistance	ETEC 0%	NT	NT
	ETEC 3	to ceftriaxone	EAEC 0%		
	EAEC 3	tested 0%	EPEC 8%		
	EPEC 13		EIEC 0%		
$\langle \rangle^{\gamma}$					2
	Year(s) of stool sampling 2013-17	Year(s) of stoolPopulation, number of isolates2013-17US military, Thailand ETEC 3 EAEC 3 EPEC 13	Year(s) of stoolPopulation, number of isolatesESBL-EC2013-17US military, Thailand ETEC 3 EAEC 3 EPEC 13only resistance to ceftriaxone tested 0%	Year(s) of stoolPopulation, numberESBL-ECCipro- floxacin resistance2013-17US military, Thailand ETEC 3only resistanceETEC 0%EAEC 3to ceftriaxoneEAEC 0%EPEC 13EIEC 0%EIEC 0%	Year(s) of stoolPopulation, numberESBL-ECCipro- floxacinAzithro- mycinamplingof isolatesresistanceresistance2013-17US military, Thailand ETEC 3only resistanceETEC 0%NTEAEC 3tested 0%EPEC 8%EPEC 13EIEC 0%

		EIEC 1			R	
Murphy 2019 ⁷⁶	2012-14	Travelers in Nepal ETEC 60 EAEC 208 EPEC 65 EIEC 10	NT	ETEC 23% EAEC 15% EPEC 23% EIEC 10%	ETEC 22% EAEC 61% EPEC 67% EIEC 30%	NT
Guiral 2019 ³⁹	2011- 17	TD Spain ETEC 43 EAEC 39	ETEC 14% EAEC 13%	ETÉC 33% EAEC 42%	ETEC 29% EAEC 33%	ETEC 0% EAEC 0%
Margulieux 2018 ³⁸	2001-16	Locals and travelers, Kathmandu, Nepal ETEC 265	ETEC 15%	ETEC 6%	NT	NT
Mason 2017 ³⁰	2002-04	US military, Thailand ETEC 29 EAEC 5 EPEC 16	NT	ETEC: 0% EAEC 0% EPEC 0%	ETEC: 0% EAEC 40% EPEC 13%	NT
Jennings 2017 ⁷¹	2003-10	Language school travelers, Cuzco, Peru ETEC 27 EAEC 9	ETEC 0% EAEC 11% nonsusceptible to ceftriaxone	ETEC: 0% EAEC: 7%	ETEC: 22% EAEC 33%	NT
Pandey 2011 ⁷⁵	2001-03	Travelers and expatriates, Nepal ETEC 50 EPEC 38	NT	ETEC 0% EPEC10%	ETEC 16% EPEC 37%	NT
Guiral 2011 ³⁷	2005–06	Spanish travelers to India with TD EAEC 51	EAEC 10%	not reported	not reported	not reported
Ouyang-Latimer 2011 ³⁶	2006-08	TD among travelers to Mexico, Guatemala, India	resistance to ceftriaxone India	India ETEC 28% EAEC 0%	India ETEC 25% EAEC 0%	India ETEC 20% EAEC 0% 3

		ETEC 365	ETEC 6%			
		EAEC 26	EAEC 0%	Mexico,	Mexico,	Mexico,
		India		Guatemala	Guatemala	Guatemala
		ETEC 98	Mexico,	ETEC 18%	ETEC 16%	ETEC 16%
		EAEC 3	Guatemala	EAEC 35%	EAEC 40%	EAEC 0%
		Mexico, Guatemala	ETEC 5%			
		ETEC 270	EAEC 20%			
		EAEC 20				
Porter	2002	US military, Turkey	NT	ETEC 5%	not reported	NT
2010^{74}		ETEC 82				
Mendez	1994-97	Spanish travelers	NT	1994-97	NT	NT
2009 ⁷³	and	1994-97		ETEC 1%		
	2001-04	ETEC 82		EAEC 2%		
		EAEC 50				
		2001-04		2001-04		
		ETEC 108		ETEC 8%		
		EAEC 54		EAEC4%		
Gomi	1997	travelers to India,	a	India	a	a
200169		Mexico, Jamaica, Kenya	a	ETEC 3/61 (4.9%)	
		ETEC 97		EAEC 4/44		
		EAEC 75		(9.1%)		
Vila	1994-97	Spanish travelers	▼ NT	ETEC 1%	NT	NT
2000^{72}		ETEC 82	,			

^aresistance rates for ETEC and EAEC only provided together; cases with both reported as "highly sensitive"

Table 4. Results of literature search for studies exploring rates of ESBL producers among various DEC isolated from stools of locals in various

regions in LMIC. From the same papers, resistance rates are given also for carbapenems, fluoroquinolones, azithromycin, and rifaximin, if tested

(NT=not tested).

 $\mathbf{\lambda}$

						\sim	
First author year	Year(s) of stool sampling	Population, number of isolates	ESBL-EC	Carbapenem resistance	Ciprofloxacin resistance	Azithromycin resistance	Rifaximin resistance
South Asia							
Moharana 2019 ⁶⁵	2012–17	Indian children with diarrhea DEC 77	4%	3%	74%	NT	NT
Mandal 2017 ⁶⁰	not reported ("during two consecutive years")	Indian children with diarrhea DEC 191	all DEC 38% ETEC 18% EAEC 7% EPEC 11% EIEC 100% EHEC 0%	0%	DEC 50% resistant to levofloxacin	NT	NT
Khalil 2016 ⁵⁶	2010–11	Pakistani children with diarrhea EAEC 35	34%	ŇŢ	69%	NT	NT
Younas 2016 ⁵⁵	2010–12	Pakistani children EPEC 46	59%	NT	39%	NT	NT
Malvi 2015 ⁵³	2012–13	Indian children with /without diarrhea EPEC 59	25%	30%	25%	14%	NT
Southeast Asia							
our search yielded	l no studies cond	ucted in Southeast As	ia				
East Asia							
Xu 2018 ⁶³	2006–15	Chinese patients with diarrhea aEPEC 151	25%	0%	5%	NT	NT
Zhou 2018 ⁶⁴	2015–16	Chinese children with diarrhea	52%	6%	50%	NT	NT

Wang 2015 ⁵⁴	2015	DEC 54 Chinese healthy elderly (>65yrs) EAEC 96	56%	NT	NT	NT	NT
North Africa an	d Middle East					/	
Farajzadeh- Sheikh 2020 ⁶⁸	2016–17	Iranian children EIEC 13 (5.1% of all DEC strains) ; other DEC not specified	EIEC 69%	EIEC 15%	0%	NT	NT
Eltai 2020 ⁶⁷	2017–18	Quatarian children EAEC 20 EPEC 56	EAEC; 20% EPEC: 23%	EAEC; 10% EPEC: 7%	0%	NT	NT
Taghadosi 2019 ⁶⁶	2014–15	Iranian children ETEC 13 EPEC 26	ETEC 54% EPEC 62%	0%	ETEC 46% EPEC 19%	NT	NT
Mahdavi 2018 ⁷⁷	2015–16	Iranian children with diarrhea ETEC 6 EAEC 35 EPEC 10 EIEC 6	ETEC 100% EAEC 74% EPEC 90% EIEC 83%	(imipenem) ETEC 50% EAEC 14% EPEC 40% EIEC 0%	ETEC 17% EAEC 20% EPEC 40% EIEC 0%	NT	NT
Amin 2018 ⁶¹	2015–16	Iranian children with diarrhea EAEC 32	28%	9% resistant to meropenem; 0% to imipenem	19%	78%	NT
Aminshahidi 2017 ⁵⁷	2014-15	Iranian children DEC 48	DEC 67% ETEC 75%	0%	DEC 31% ETEC 25%	NT	NT

			EAEC 85%		EAEC 27%		
			EPEC 33%		EPEC 33%		
			EIEC 50%		EIEC 50%		
Karami	not reported	Iranian children	80%	0%	21%	NT	NT
2017^{58}		with /without					
		diarrhea					
		EPEC 192					
Memariani	2011-13	Iranian children	21%	NT	17%	NT	NT
2015^{52}		with diarrhea		\sim			
		EPEC 42					
Ghorbani-Dalini	2010	Iranian adults	13%	6% resistant to	8%	NT	NT
201551		with diarrhea		imipenem			
		DEC 54; DEC					
		types not					
		specified					
Khoshvaght	2011-12	Iranian children	53%	4% resistant to	16%	NT	NT
2014 ⁵⁰		with diarrhea		imipenem			
		EAEC 36					
Sonnevend	2003–04	children and	11%	NT	NT	NT	NT
200648		adults with and					
		without diarrhea,					
		United Arab					
		Emirates	\sim				
		EAEC 44					
Sub-Saharan Afri	ica		, ,				
Konate	2013–15	children with	68%	16% resistant to	0%	NT	NT
2017 ⁵⁹		diarrhea, Burkina		imipenem			
		Faso					
	(DEC 31					
South and Centra	ll America and t	he Caribbean					
Amaya	2005-06	Nicaraguan		0.51			
-		-	diarrhea:	0%	1%	NT	NT
	\sim						7

2011 ⁴⁹	children DEC 332	ETEC 5/64 (8%) EAEC:23/134 (17%) EPEC: 3/34 (9%) EHEC: 0/8 (0%) EIEC 0/1 (0%)	R
		no diarrhea: ETEC 1/9 (11%) EAEC: 13/69 (19%) EPEC:0/13 (0%) EHEC 0/0 (0%)	
			8