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Oxidative state in the oestrus cycle of the buffaloes: a preliminary study

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Dear Editor and Reviewer,

thank You for the revision of our paper.

We are agree with the reviewer when he wrote that the “groups appear to be a thin sample”. We wanted to understand the trend of oxidative status in buffaloes, because we work in this field from many years. We enrolled a little sample, just to outline a first range of values, which have to be confirmed, expanding the samples. For this reason, we modified the title and added in conclusion (line 176-177) a sentence in which we specified that it is a preliminary study.

27 found in dioestrus, when they perform protective action against oxidative damage in the ovaries and
28 uterus.

29

30 Keywords: buffalo, oxidative status, oestrus, dioestrus, anoestrus.

31

32 INTRODUCTION

33 Free radicals are molecules with one or more unpaired electrons on their last orbital, a condition
34 that makes them highly reactive, so that they tend to subtract (Poston and Raijmakers, 2004) or
35 donate an electron to a non-radical molecule (Hallywell, 1991), to achieve an electronic stability.

36 Two main categories of free radicals exist: Reactive Oxygen Species (ROS) and Reactive Nitrogen
37 Species (RNS) (Agarwal et al., 2005); both derive from oxidation (loss of electrons) or reduction
38 (acceptance of electrons) reactions (Pourovà et al., 2010), occurring physiologically within the cells,
39 during normal aerobic metabolic processes.

40 Free radicals play important roles in many biologic pathways, thus they are commonly found in
41 blood. ROS are involved in respiratory burst (Saugstad, 2000; Bergendi et al, 1999), they are able to
42 amplify inflammatory responses (Poston and Raijmakers, 2004) and appear to be involved in signal
43 transduction in various biological processes (Saugstad, 2000; Fleury et al, 2002). However, when at
44 high concentrations, they exert adverse effects in the cell, leading to lipid peroxidation, oxidation of
45 proteins (with loss of their biological functions) and of nucleic acids (rupture of nucleotide
46 filaments) (Valko et al., 2006).

47 Organisms have developed antioxidant defences in order to contain the potential adverse effects of
48 ROS (Anderson and Phillips, 1999). These defences consist of exogenous components, taken with
49 the diet, vitamin compounds, and not vitamin compounds (Liebler and Stratton, 1997), besides
50 endogenous enzymatic and non-enzymatic antioxidants (Antolovich et al., 2002; Rahman, 2007;
51 Uttara et al., 2009).

52 In reproduction, the ROS/antioxidants system modulates folliculogenesis, ovulation, formation and
53 activity of the corpus luteum, luteolysis, early embryonic development, embryo implantation,
54 initiation of parturition and placentation (Rizzo et al., 2012).

55 Studies performed in cows (Rizzo et al., 2007; Rizzo et al., 2009a), bitches (Rizzo et al., 2009b) and
56 sheep (Rizzo et al., 2008) have shown that physiological ROS concentrations influence
57 reproduction.

58 When ROS generation exceed antioxidant defences oxidative stress arises, leading to many
59 diseases. In the ovary, high free radical levels inhibit oocyte development (Guerin et al., 2001),
60 causing meiosis arrest, increase the percentage of degenerated oocytes (Tatemoto et al., 2000) as
61 well as oocyte apoptosis (Liu et al., 2000). In addition, some authors have shown that oxidative
62 stress is involved in the onset of follicular cysts, repeat breeder syndrome, mastitis, metritis and
63 retained placenta in cattle (Kankofer, 2001; Miller et al., 2003; Rizzo et al., 2007; Rizzo et al.,
64 2009a).

65 Given these premises and considering that, to the best of our knowledge, no data about the
66 physiological range of blood ROS concentrations in the buffalo exist, this study aims to investigate,
67 under physiological conditions, ROS serum concentrations in the buffaloes, during the different
68 phases of the estrous cycle. Once established a physiological range, it could be helpful for
69 diagnosing reproductive or other dysfunctions, in case they should lead to altered blood ROS
70 concentrations.

71

72 MATERIALS AND METHODS

73 This study was carried out between July 2017 and April 2018, in the farm "The Park", located in
74 San Giovanni Rotondo, in the province of Foggia, Apulia Region, Italy.

75 The farm has a consistency of about 500 animals, including an average of 180 lactating ones,
76 receiving a diet consisting of unifeed, deriving from corn silage, oat hay, lucerne hay, corn flour,
77 soybean meal, cotton, crushed barley, beet pulp, vitamin and trace elements supplements.

78 Female buffaloes (3-5 year old), in postpartum from at least 100 days, with a daily milk production
79 of about 10 kg and weighting 600 to 700 Kg were enrolled in this study.

80 The animals underwent a complete clinical examination, including genital tract inspection (rectal
81 palpation), necessary to rule out eventual diseases and to detect the specific phase of the oestrous
82 cycle of each buffalo. The detection of a tonic uterus, with a follicle of about 1.5 cm in diameter,
83 with tense and floating wall and a corpus luteum in regression in the ovary, let define oestrus.

84 The buffaloes with a hypotonic uterus, a corpus luteum of approximately 1.5-2 cm in diameter and
85 follicles <1 cm were referred to as being in the luteal phase (El-Shahata and Kandilb, 2012; Verma
86 et al., 2018).

87 Upon complete clinical examination, 30 healthy buffaloes, with a BCS of 3.5 to 4 on a
88 scale from 1 to 5 (Anitha et al., 2011), were chosen for this study, of which 10 were in oestrus, 10 in
89 dioestrus and 10 in anoestrus.

90 All the buffaloes underwent blood collection from the coccygeal vein using vacutainer serum-tubes
91 chilled in advance.

92 The samples were transferred on ice to the laboratory of the Obstetrics section at the Department of
93 Veterinary Medicine of Bari, where they were centrifuged at 1600xg for 10 minutes at + 4 ° C. The
94 sera obtained were stored in 1.5 mL eppendorf and frozen at -20 ° C until analytical determinations.

95 Biological antioxidant potential (BAP test) and oxidative potential (dROMs-test reactive
96 metabolites) were determined on the sera, through mono ready test photometer dedicated for use
97 Free Carpe Diem ® (Diacron International, Grosseto, Italy).

98 dROMs-test, known as d-ROMs, allows to determine the concentration of Reactive Oxygen
99 Metabolites (ROMs) in a biological sample, particularly hydroperoxides, deriving from the
100 oxidative attack of many biochemical substrates (glycids, lipids, amino acids , proteins, nucleotides,

101 etc.). The results of d-ROMs test are expressed in arbitrary units, the Carratelli Units or U.CARR,
102 where 1 U.CARR. equals 0.08 mg H₂O₂/dL.

103 As to BAP test, it is based on the ability of serum to reduce a given oxidizing substrate, suitably
104 chosen based on its redox potential. Measuring the "dynamic" or "biologically active" component of
105 the blood antioxidant barrier, provides a global measurement of many antioxidants (bilirubin, uric
106 acid, vitamins C and E).

107 The results of BAP test are expressed as micromoles (μmol) of reduced iron per liter (L) of sample.
108 All the results are shown as mean ± SD. They were compared using SPSS 19 statistical program
109 (IBM, NY). Particularly, one-way ANOVA with post-hoc LSD test was used for inter-group
110 determinations. A value of P<0.05 was set as statistically significant.

111

112 RESULTS

113 The results (mean ± SD) of ROS and BAP serum concentrations in oestrus, dioestrus and anoestrus
114 are shown in Table 1.

115

	oestrus	dioestrus	anoestrus
ROS (U.CARR)	101.8 ± 6.03 a	88.40 ± 3.47 b	87.10 ± 11.84 b
BAP (μMmol / L)	2072.71 ± 94.50 c	2207.68 ± 111.90 d	2200.16 ± 133.11 d

116 **Table 1:** Mean ± SD concentrations of Reactive Oxygen Species (U.CARR) and Blood Antioxidant
117 Potential (μmol/L) in buffaloes in the different phases of the cycle: oestrus, dioestrus and anoestrus.
118 a, b: P <0.01; c, d: P <0.05

119

120

121

122

123 The highest ROS and BAP concentrations were observed in oestrus (101.8 ± 6.03 U. CARR) and
124 dioestrus (2207.68 ± 111.90 $\mu\text{mol/L}$), respectively.

125 Statistically significant differences between oestrus vs dioestrus and oestrus vs anoestrus were
126 found for ROS ($P < 0.01$) whereas for the BAP, the differences among the same groups showed a
127 significance of $P < 0.05$.

128

129 DISCUSSION

130 This study, even performed on a limited number of animals, is the first, to our best knowledge, to
131 show a range of reference for physiological serum Reactive Oxygen Species (ROS) and Blood
132 Antioxidant Potential (BAP), in the different phases of the estrous cycle, in buffaloes.

133 The results obtained in this study show that serum ROS levels, in buffaloes, are higher than those
134 reported in the bovine species (Rizzo et al., 2007; Rizzo et al 2009a). It is conceivable that this
135 finding may be related to inadequate feeding management (Bertoni et al., 2001). On the other hand,
136 BAP levels are similar to those found in cattle (Rizzo, 2018 unpublished thesis, University of Bari
137 Aldo Moro). This last datum is also confirmed by studies in which blood antioxidant concentrations
138 (Glutathione peroxidase and superoxide dismutase) are similar to those of cattle (Morgante et al.,
139 2001).

140 The greater rusticity of the buffalo, compared to the cow, could explain the findings of this study,
141 i.e. ROS concentrations higher in buffaloes than in cows, with a similar BAP, in healthy conditions.

142 As to the different phases of the oestrous cycle, the highest serum ROS concentrations found in
143 oestrus, could be interpreted with the knowledge that ROS play pivotal roles in this phase, in the
144 ovary. In fact, they take part to the rupture of the preovulatory follicle walls (Jozwik et al., 1999),
145 stimulating the apoptosis of granulosa cells and inducing, thereby, ovulation (Kodaman and
146 Behrman, 2001; Shkolnik et al., 2011). Many literary data prove that the ovary itself produces ROS,
147 in order to properly undergo ovulation. Ovulation was seen to be prevented if ROS generation is

148 counteracted by antioxidants (Miyazaki et al., 1991); in line with Miyazaki et al. (1991), Rizzo et al
149 (2009a) found lower ROS concentration in ovarian cystic fluid than in follicular fluid and suggested
150 that the reduced ROS concentrations had been unable to determine the rupture of the follicular wall,
151 having led to cystic formation.

152 As to the buffaloes enrolled in this study, in contrast to the high estrous ROS concentrations, a low
153 BAP was detected in the same phase. At this stage, antioxidants have the task of ensuring oocyte
154 protection from oxidative damage, until reaching the pre-ovulatory ovarian follicle stage and
155 improve the quality of the gametes (Sugino et al., 2000; Fujii et al., 2005), without, however,
156 interfere with the prime function of apoptosis induced by ROS, essential for ovulation (Sato et al.,
157 1992; Miyamoto et al., 2010).

158 During the luteal phase, however, a decline in ROS concentrations and a concurrent increase in
159 antioxidants was registered. The opposite trend observed, besides being in line with the functional
160 antagonism of ROS and BAP (Peltier and Smullan, 2006; Jones et al., 2008), might be explained
161 with the importance of antioxidants (mainly β -carotene) (Rizzo et al., 2013) in the luteal phase,
162 when they are necessary for a secretion of progesterone (Kato et al., 1997).

163 As to anoestrus, the concentrations of ROS and BAP were lower than the ones encountered in the
164 oestrus and dioestrus, a sign that, in these farming conditions, the antioxidant system is both
165 endogenous and exogenous (vitamin intake in the diet), and is able to keep under control the
166 oxidative status.

167 During dioestrus, a decrease in ROS and an increase in BAP levels were registered; this datum let
168 hypothesize that when BAP is high, it efficacely counteracts ROS generation (Peltier and Smullan,
169 2006; Jones et al., 2008), besides stimulating luteal progesterone secretion (Kato et al., 1997).

170 In this study, the high BAP levels detected in the luteal phase are in accordante with literature and
171 are related to the high luteal antioxidant content (mainly β -carotene).

172 Concluding, the results of this study, together with the cited literary data let infer that the ovary
173 might be a primary ROS source as a function of its activity and, at the same time, it is protected by

174 an adequate antioxidant system against the oxidative damage, contributing to a balance between
175 oxidant and antioxidant activity.

176 This was a preliminary investigation. Further studies are needed to confirm the obtained range and
177 to evaluate the trend of oxidative status under pathological conditions.

178

179 ETHICAL APPROVAL

180 All procedures performed in studies involving animals were in accordance with the ethical
181 standards of the institution or practice at which the studies were conducted (Ethics Committee for
182 animal experimentation, No. prot. 27/18).

183

184 CONFLICT OF INTEREST

185 The authors declare that they have no conflict of interest.

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191 REFERENCES

192

193 Agarwal, A., Gupta, S. and Sharma, RK, 2005. Role of oxidative stress in female reproduction,
194 Reproductive Biology and Endocrinology, 3, 28.

195 Anderson, D. and Phillips, BJ, 1999. Comparative in vitro and in vivo effects of antioxidants, Food
196 and Chemical Toxicology, 37, 1015-1025.

197 Anitha, A., Sarjan Rao, K., Suresh, J., Srinivasa Moorthy, P.R. and Kotilinga Reddy, Y., 2011. A
198 body condition score (BCS) system in Murrah buffaloes, Buffalo Bulletin, 30, 79-99.

199 Antolovich, M., Prenzler, PD, Patsalides, E., McDonald, S. and Robards, K., 2002. Methods for
200 testing antioxidant activity, *Analyst*, 127, 183-198.

201 Bergendi, L., Benes, L., Durackova, Z. and Ferenčík, M., 1999. Chemistry, physiology and
202 pathology of free radicals, *Life Sciences*, 65, 1865-1874.

203 Bertoni, G., Piccioli Cappelli, F. and Carli, D., 2001. The buffalo farm in Northern Italy, National
204 Congress of the breeding buffalo: reports and scientific communications, Eboli (Sa), 20-27.

205 El-Shahata, K.H and Kandilb, M., 2012. Antioxidant capacity of follicular fluid in relation to
206 follicular size and stage of estrous cycle in buffaloes, *Theriogenology*, 77, 1513-1518.

207 Fleury, C., Mignotte, B. and Vayssière, L., 2002. Mitochondrial reactive oxygen species in cell
208 death signaling, *Biochimie*, 84, 131-141.

209 Fujii, J., Iuchi, Y. and Okada, F., 2005. Fundamental roles of reactive oxygen species and protective
210 mechanisms in the female reproductive system, *Reproductive Biology and Endocrinology*, 3, 43-53.

211 Guerin, P., El Mouatassin, S. and Menezo, Y., 2001. Oxidative stress and protection against
212 reactive oxygen species in the pre-implantation embryo and its surroundings, *Human Reproduction*
213 *Update*, 7, 175-189.

214 Hallywell, B., 1991. Reactive oxygen species in living system, source, biochemistry, and role in
215 human disease, *The American Journal of Medicine*, 91, 14-24.

216 Jones, L.A., Anthony, J.P., Henriquez, F.L., Lyons, R.E., Nickdel, M.B., Carter, K.C., Alexander, J.
217 and Roberts, C.W., 2008. Toll-like receptor-4-mediated macrophage activation is differentially
218 regulated by progesterone via the glucocorticoid and progesterone receptors. *Immunology*, 125, 59-
219 69.

220 Jozwik, M., Wolczynski, S. and Szamatowicz, M., 1999. Oxidative stress markers in preovulatory
221 follicular fluid in humans, *Molecular Human Reproduction*, 5, 409-413.

222 Kankofer, M., 2001. Antioxidative defense mechanisms against reactive oxygen species in bovine
223 retained and not-retained placenta: activity of glutathione peroxidase, glutathione transferase,
224 catalase and superoxide dismutase, *Placenta* 22, 466-472.

225 Kato, H., Sugino, N., Takiguchi, S., Kashida, S. and Nakamura, Y., 1997. Roles of reactive oxygen
226 species in the regulation of luteal function, *Reviews of Reproduction*, 2, 81-83.

227 Kodaman, P.H. and Behrman, H.R., 2001. Endocrine-regulated and protein kinase c-dependent
228 generation of superoxide by rat preovulatory follicles, *Endocrinology*, 142, 687-693.

229 Liu, L., Trimarchi, J.R. and Keefe, D.L., 2000. Involvement of mitochondria in oxidative stress-
230 induced cell death in mouse zygotes, *Biology of Reproduction*, 62, 1745-1753.

231 Miller, JK, Brzezinska-Slebodzinska, E. and Madsen, F.C., 1993. Oxidative stress, antioxidants,
232 and animal function, *Journal of Dairy Science*, 76, 2812-2823.

233 Miyamoto, K., Sato, E.F., Kasahara, E., Jikumaru, M., Hiramoto, K., Tabata, H., Katsuragi, M.,
234 Odo, S., Utsumi, K. and Inoue, M., 2010 . Effect of oxidative stress during repeated ovulation on
235 the structure and functions of the ovary, oocytes, and their mitochondria, *Free Radical Biology and*
236 *Medicine*, 49, 674-681.

237 Miyazaki, T., Sueoka, K., Dharmarajan, A.M., Aclas, S.J., Bulkley, G.B. and Wallach, E.E., 1991.
238 Effect of inhibition of oxygen free radical on ovulation and progesterone production by the in-vitro
239 perfused rabbit ovary, *Journal of Reproduction and Fertility*, 91, 207-212.

240 Morgante, M., Beghelli D., Stelletta, C. and Ranucci, S., 2001. Glutathione peroxidase (GSHPx)
241 and superoxide dismutase (SOD) erythrocytes in adult buffaloes (*Bubalus Bubalis*), *National*
242 *Congress of the breeding buffalo: reports and scientific communications*, Eboli (Sa), 302-306.

243 Peltier, M., Tee and S. and Smullan, L., 2006. Does progesterone lower innate immunity to
244 pathogens associated with preterm birth?, *American Journal of Obstetrics and Gynecology*, 195,
245 S69.

246 Poston, L. and Raijmakers, M.T.M., 2004. Trophoblast oxidative stress, antioxidants and pregnancy
247 outcome - a review, *Placenta* 18, 72-78.

248 Pourová, J., Kottova, M., Voprsalova, M. and Pour, M., 2010. Reactive oxygen and nitrous species
249 in normal physiological processes, *Acta Physiologica*, 198, 15-35.

250 Rahman, K., 2007. Studies on free radicals, antioxidants, and cofactors, *Clinical Interventions in*
251 *Aging*, 2, 219-236.

252 Rizzo, A., Minoia, G., Trisolini, C., Manca, R. and Sciorsci, R.L., 2007. Concentrations of free
253 radicals and beta-endorphins in repeat breeder cows, *Animal Reproduction Science*, 100, 257-263.

254 Rizzo, A., Mutinati, M., Spedicato, M., Minoia, G., Trisolini, C., Jirillo, F. and Sciorsci, R.L., 2008.
255 First demonstration of an increased serum level of reactive oxygen species during the peripartal
256 period in the ewes, *Immunopharmacology and Immunotoxicology*, 30, 741-746.

257 Rizzo, A., Minoia, G., Trisolini, C., Mutinati, M., Mailloux, M., Jirillo, F. and Sciorsci, R.L.,
258 2009a. Reactive Oxygen Species (ROS); involvement in bovine follicular cysts etiopathogenesis,
259 *Immunopharmacology and Immunotoxicology*, 31, 631-635.

260 Rizzo, A., Roscino, M.T., Minoia, G., Trisolini, C., Spedicato, M., Mutinati, M., Pantaleo, M.,
261 Jirillo, F. and Sciorsci, R.L., 2009b. Serum levels of reactive oxygen species (ROS) in the bitch.
262 *Immunopharmacology and Immunotoxicology*, 31, 310-313.

263 Rizzo, A., Roscino, M.T., Binetti, F. and Sciorsci, R.L., 2012. Roles of reactive oxygen species in
264 female reproduction, *Reproduction in Domestic Animals*, 47, 344-352.

265 Rizzo, A., Ceci E, Pantaleo M, Mutinati M, Spedicato M, Minoia G and Sciorsci, R.L., 2013.
266 Evaluation of blood and milk oxidative status during early postpartum of dairy cows, *Animal*, 7,
267 118-123.

268 Rizzo, A., 2018. Oxidative status in oestrus cycle of buffaloes, (unpublished thesis, University of
269 Bari Aldo Moro).

270 Sato, E.F., Kobuchi, H., Edashige, K., Takahashi, M., Yoshioka, T., Utsumi, K. and Inoue, M.,
271 1992. Dynamic aspects of ovarian superoxide dismutase isozymes during the ovulatory process in
272 the rat, *FEBS Letters*, 303, 121-125.

273 Saugstad, O.D., 2000. Bronchopulmonary dysplasia-oxidative stress and antioxidants. *Seminars in*
274 *Neonatology*, 8, 39-49.

275 Shkolnik, K., Tadmor, A., Ben-Dor, S., Nevo, N., Galiani, D. and Dekel, N., 2011. Reactive oxygen
276 species are indispensable in ovulation. *Proceedings of the National Academy of Sciences*, 108,
277 1462-1467.

278 Liebler, D.C., and Stratton, S.P., 1997. Determination of singlet oxygen-specific versus radical
279 mediated lipid peroxidation in the photosensitized oxidation of lipidic bilayer: effect of beta-
280 carotene and alpha-tocopherol. *Biochemistry*, 36, 12911-12920.

281 Sugino, N., Takiguchi, S., Kashida, S., Karube, A., Nakamura, Y. and Kato, H., 2000. Superoxide
282 dismutase expression in the human corpus luteum during the menstrual cycle and pregnancy
283 in early, *Molecular Human Reproduction*, 6, 19-25.

284 Tatemoto, H., Sakurai, N. and Mute, N., 2000. Protection of porcine oocytes against apoptotic cell
285 death Caused by oxidative stress during in vitro maturation: role of cumulus cells, *Biology of*
286 *Reproduction*, 63, 805-810.

287 Uttara, B., Singh, A.V., Zamboni, P., Mahajan, R.T., 2009. Oxidative stress and neurodegenerative
288 36 diseases: a review of upstream and downstream antioxidant therapeutic options, *Current*
289 *Neuropharmacology*, 7, 65-74.

290 Valko, M., Rhodes, C.J., Moncol, J., Izakovic, M. and Mazur, M., 2006. Free radicals, metals and
291 antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, 160, 1-40.

292 Verma, A.D., Panigrahi, M., Bhushan, B., Baba, N.A., Sulabh, S., Sadam, A., Parida, S., Sonwane,
293 A. and Narayanan K., 2018. Relative expression of oxytocin receptor gene in buffalo endometrium
294 in late luteal phase and pregnancy stages, *Journal of Applied Animal Research*, 46, 146-149.