

Article

Screen-Printed Graphite Electrode Modified with Graphene-Co₃O₄ Nanocomposite: Voltammetric Assay of Morphine in the Presence of Diclofenac in Pharmaceutical and Biological Samples

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Abstract: This work focuses on the development of a novel electrochemical sensor for the determination of morphine in the presence of diclofenac. The facile synthesis of graphene-Co₃O₄ nanocomposite was performed. The prepared material (graphene-Co₃O₄ nanocomposite) was analyzed by diverse microscopic and spectroscopic approaches for its crystallinity, composition, and morphology. Concerning the electrochemical determinations, after drop-casting the as-fabricated graphene-Co₃O₄ nanocomposite on the surface of a screen-printed graphite electrode (SPGE), their electrochemical performance was scrutinized towards the morphine detection. It was also found that an SPGE modified by a graphene-Co₃O₄ nanocomposite exhibited better electrocatalytic activity for morphine oxidation than unmodified electrode. Under optimal conditions, the differential pulse voltammetry (DPV) was employed to explore the present sensor (graphene-Co₃O₄/SPGE), the findings of which revealed a linear dynamic range as broad as 0.02–575.0 μM and a limit of detection (LOD) as narrow as 0.007 μM. The sensitivity was estimated to be 0.4 μM/(μA cm²). Furthermore, the graphene-Co₃O₄/SPGE sensor demonstrated good analytical efficiency for sensing morphine in the presence of diclofenac in well-spaced anodic peaks. According to the DPV results, this sensor displayed two distinct peaks for the oxidation of morphine and diclofenac with 350 mV potential difference. In addition, the graphene-Co₃O₄/SPGE was explored for voltammetric determination of diclofenac and morphine in pharmaceutical and biological specimens of morphine ampoule, diclofenac tablet, and urine, where recovery rates close to 100% were recorded for all of the samples.

Keywords: electrochemical sensor; morphine; diclofenac; graphene-Co₃O₄ nanocomposite; screen-printed graphite electrode



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1. Introduction

Today, pharmaceutical industries and medical sciences have realized the necessity of detecting and quantifying drugs in human biological fluids and pharmaceutical matrices. Morphine, a strong opiate analgesic psychoactive drug, is the prototypical opioid leading to central nervous system (CNS) disorders. Morphine can relieve severe pain in patients, particularly in candidates for surgery [1]. Opium poppy straw is the source for extracting morphine [2]. This substance can act as a precursor in obtaining some opioids, such as codeine, hydromorphone, heroin, dihydromorphone, and nicomorphine [3]. Heroin metabolites can cause the production of this substance, which can subsequently be an indicator of heroin use [4]. In addition, morphine is available as an illegal drug and can

be associated with side effects such as hallucinations, slow heart rate, CNS disturbance, constipation, nausea, respiratory problems, muscle stiffness, coma, and dependence [5].

Nonsteroidal anti-inflammatory drugs (NSAIDs) lead to decreased inflammation, pain, and fever throughout the body by inhibiting cyclooxygenase enzymes and reducing the production of prostaglandins [6]. One NSAID is diclofenac, which has potent analgesic, anti-inflammatory, and antipyretic performances. The use of diclofenac is due to effects such as relief of post-traumatic pain, cancer pain, and neuralgia and treatment of osteoarthritis, rheumatoid arthritis, soft tissue disorders, acute gout, and some other inflammatory responses [7,8]. Despite the significant benefits of diclofenac, its inappropriate use can be associated with disadvantages such as extensive liver metabolism, stroke, and hypertension [9]. In addition, the poor degradation of diclofenac in the environment leads to poor water quality and endangers the health of fish [10]. Researchers found that combining morphine and diclofenac in small doses is synergistic in cases of serious inflammatory pains. Actually, using the combination decreases the amount of morphine necessary for sufficient analgesia in comparison to the use of morphine itself [11]. Due to all of these issues, it is vital to detect such analytes in human biological fluids and pharmaceutical matrices.

Accordingly, there have been various analytical ways to detect diclofenac and morphine in diverse matrices, some of which are high-performance liquid chromatography [12,13], chemiluminescence [14,15], capillary electrophoresis [16,17], spectrophotometry [18,19], gas chromatography-mass spectroscopy [20,21], thin-layer chromatography [22], and some electrochemical methods [23–26]. High selectivity, high sensitivity, and minimal-interference impacts are prominent features of a potent analytical method. Despite the identification of drugs at very low doses by these methods, they require lengthy sample preparation steps, pre-analytical extraction, highly trained technical staff, and expensive instruments. Among these techniques, electrochemical techniques have emerged as important diagnostic tools in recent years, due to their high sensitivity, simplicity, accuracy, and fast detection [27–31].

Special attention in the field of electroanalytical research has recently been drawn to screen-printed electrodes (SPE) in the manufacturing of (bio)sensors [32–36]. Compared to the routine three-electrode system (working, reference, and auxiliary), SPEs have merits such as easier setup, system miniaturization, and portability. Besides the possibility of performing in situ analyses, the ease of mass production, high reproducibility, and low cost make these electrodes very interesting. Chemically modified electrodes (CMEs) are the result of the intentional fixation of a modifying agent on the surface of electrode by various physical and chemical methods [37,38]. Many studies have shown that improvements in the sensitivity and selectivity of electrochemical sensors are achieved due to modifications of the electrode surface [39,40].

Nanomaterials and their applications in various fields have become distinct and active areas of scientific and technological developments over the recent years [41–43]. Nanomaterial-supported electrochemical-sensing systems have been considered by many researchers due to their capability for carrying out the electrochemical analysis of diverse analytes resulting from enhanced detection efficacy [44–46]. Graphene, a 2D carbon-based nanomaterial, possesses carbon atoms with sp^2 hybridization connected in a hexagonal lattice architecture. It has admirable electrical conductivity, predominantly because of delocalized π bonds at the top and bottom of its base plane and its huge surface area, and it has recently become a promising candidate for the development of electrochemical sensors [47–50]. Transition-metal oxides are routine mediators in the construction of electronics and advanced catalysts because of specific traits such as impressive catalytic performance, chemically reactive facets, and great earth abundance [51]. In this regard, cobalt oxide (Co_3O_4) has shown commendable physicochemical features such as conductivity, high reversibility, and thermal stabilities [52]. Nanomaterial functionalization remarkably enhances the activity of sensors through the improvement of electrochemical capabilities leading to a greater electron transfer rate compared to a single component.

Combining the advantages of the unique features of graphene with Co_3O_4 nanoparticles, the graphene/ Co_3O_4 nanocomposite creates a sensitive and stable base for electroanalysis [53,54].

The present work describes an electrochemical sensor on the basis of SPGE modification with graphene- Co_3O_4 nanocomposite for the detection of morphine combined with diclofenac. The resulting modified electrode (graphene- Co_3O_4 nanocomposite/SPGE) had a great sensitivity towards the morphine, possessing a narrow LOD and a broad linear range. Furthermore, the co-detection of morphine and diclofenac was performed on the modified electrode surface. The designed graphene- Co_3O_4 nanocomposite/SPGE sensor had practical applicability for sensing morphine and diclofenac in real biological and pharmaceutical matrices with acceptable recoveries.

2. Materials and Methods

2.1. Equipments

An Autolab PGSTAT 320N Potentiostat/Galvanostat Analyzer (Herisau, Switzerland) with GPES (General Purpose Electrochemical System-version 4.9) software was applied for all electrochemical determinations at ambient temperature. The electrochemical sensors were prepared by DRP-110 SPEs (DropSens, Oviedo, Spain) using silver pseudo-reference electrode, graphite working electrode, and graphite auxiliary electrode. A Metrohm 713 pH meter with a glass electrode (Metrohm AG, Herisau, Switzerland) was selected to determine and adjust the solutions' pH. Direct-Q[®] 8 UV deionized water (Merck Chemicals GmbH, Darmstadt, Germany) was used to prepare fresh solutions.

A PANalytical X'Pert-PRO X-ray diffractometer (Almelo, The Netherlands) applying a $\text{Cu}/\text{K}\alpha$ radiation ($\lambda:1.54 \text{ \AA}$) was used for X-ray-diffraction (XRD) analysis, and a Bruker Tensor II spectrometer (Bruker, Karlsruhe, Germany) was employed to capture the Fourier transform infrared (FT-IR) spectra. A MIRA3 scanning electron microscope coupled with an energy-dispersive X-ray spectroscopy (EDS) detector (Tescan, Brno, Czech Republic) was utilized for field-emission scanning electron microscopy (FE-SEM) (Tescan, Brno, Czech Republic) images and elemental analysis.

2.2. Solvents and Chemicals

All solvents and chemicals applied in our protocol were of analytical grade obtained from Merck and Sigma-Aldrich (Darmstadt, Germany). The phosphate buffer solution (PBS) was prepared with phosphoric acid and adjusted with NaOH to the desired pH value.

2.3. Preparation of Graphene- Co_3O_4 Nanocomposite

First, graphene oxide (GO) (20 mg) was dissolved in ethanol (20 mL) under 30-min ultra-sonication. Then, $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ (0.001 mol) was dissolved in ethanol (20 mL) while stirring for 30 min at ambient temperature. Next, the two prepared solutions were blended, 3.6 mL of an ammonia ($\text{NH}_3 \cdot \text{H}_2\text{O}$ (wt. 25%)) solution was added dropwise, and the solution was subsequently transferred into a Teflon-lined stainless-steel autoclave and maintained at $180 \text{ }^\circ\text{C}$ for 24 h. After completion of the reaction, centrifugation was performed to collect the product, followed by rinsing with ethanol/deionized water. At last, the graphene- Co_3O_4 nanocomposite was oven-dried at $70 \text{ }^\circ\text{C}$ overnight.

2.4. Preparation of the Graphene- Co_3O_4 /SPGE Sensor

A drop-casting technique was followed to fabricate the graphene- Co_3O_4 /SPGE. Thus, a certain amount of as-prepared graphene- Co_3O_4 nanocomposite (1 mg) was dispersed in deionized water (1 mL) under 20-min ultra-sonication. Then, the well-dispersed suspension (4 μL) was coated on the SPGE surface dropwise and dried at the laboratory's temperature.

The electrochemically active surface area (EASA) of the unmodified SPGE and graphene- Co_3O_4 /SPGE was calculated from the cyclic voltammetry (CV) measurements in a 0.1 M KCl solution containing 1.0 mM $\text{K}_3[\text{Fe}(\text{CN})_6]$ as a redox probe at different scan rates. The value of the EASA was calculated according to the Randles-Sevcik equation [55]. The

values of the EASA for the unmodified SPGE and graphene- Co_3O_4 /SPGE are calculated as 0.05 and 0.16 cm^2 , respectively. The results reveal that the graphene- Co_3O_4 /SPGE has a larger EASA than the unmodified SPGE.

2.5. Preparation of Real Samples

Urine samples were obtained from healthy volunteers and were stored in a refrigerator immediately after collection. Ten milliliters of each sample was centrifuged for 15 min at 2000 rpm. The supernatant was filtered out using a 0.45 μm filter. Then, varying volumes of the solution were transferred into 25 mL volumetric flasks and diluted to the mark with PBS (pH 7.0). The diluted urine sample was spiked with different amounts of morphine and diclofenac. The morphine and diclofenac contents were analyzed by the proposed method using the standard addition method.

One milliliter of morphine ampoule (labeled 10 mg per mL, Alborz Darou Company, Alborz Industrial City, Iran) was diluted to 10 mL with 0.1 M PBS (pH 7.0); then, varying volumes of the diluted solution were transferred into each of a series of 25 mL volumetric flasks and diluted to the mark with PBS. The analysis of morphine and diclofenac was performed using the standard addition method.

Five diclofenac tablets (labeled 50 mg per tablet, Darou Pakhsh Company, Tehran, Iran) were ground. Then, a tablet solution was prepared by dissolving 150 mg of the powder in 25 mL water by ultrasonication. Then, a different volume of the diluted solution was transferred into a 25 mL volumetric flask and diluted to the mark with PBS (pH 7.0). The morphine and diclofenac contents were analyzed by the proposed method using the standard addition method.

3. Results and Discussion

3.1. Characterization of Graphene- Co_3O_4 Nanocomposite

FT-IR analysis was performed to confirm the formation of graphene- Co_3O_4 nanocomposite (Figure 1). The wide absorption peak at 3417 cm^{-1} was representative of functional groups of OH. The peak at 1633 cm^{-1} related to aromatic C=C graphitic carbon domain vibration. The band at 1384 cm^{-1} related to OH deformation and bending vibration of interlayer water molecules [56]. The peaks at 562 and 660 cm^{-1} related to Co-O metal-oxygen bond stretching vibrations, approving Co_3O_4 nanoparticle formation.

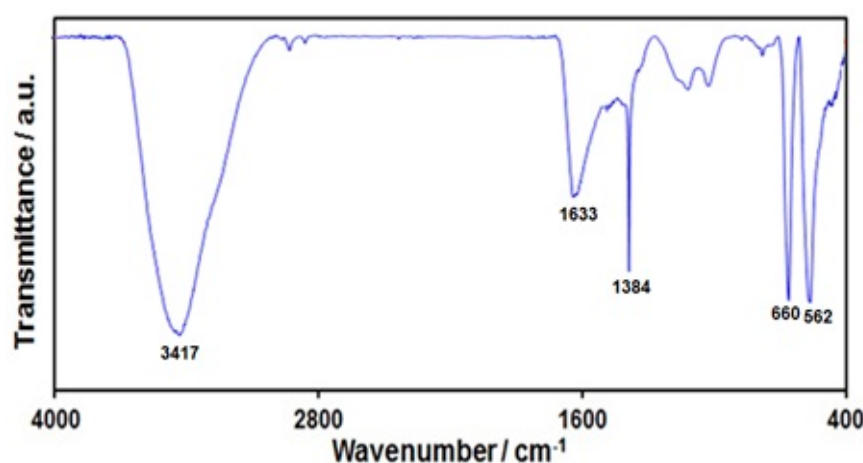


Figure 1. FT-IR spectrum of graphene- Co_3O_4 nanocomposite.

The FE-SEM image was captured to explore the morphology and structure of the graphene- Co_3O_4 nanocomposite. Figure 2 shows the FE-SEM image of the graphene- Co_3O_4 nanocomposite. It can be seen that Co_3O_4 nanoparticles are anchored and densely dispersed on the surface of graphene nanosheets.

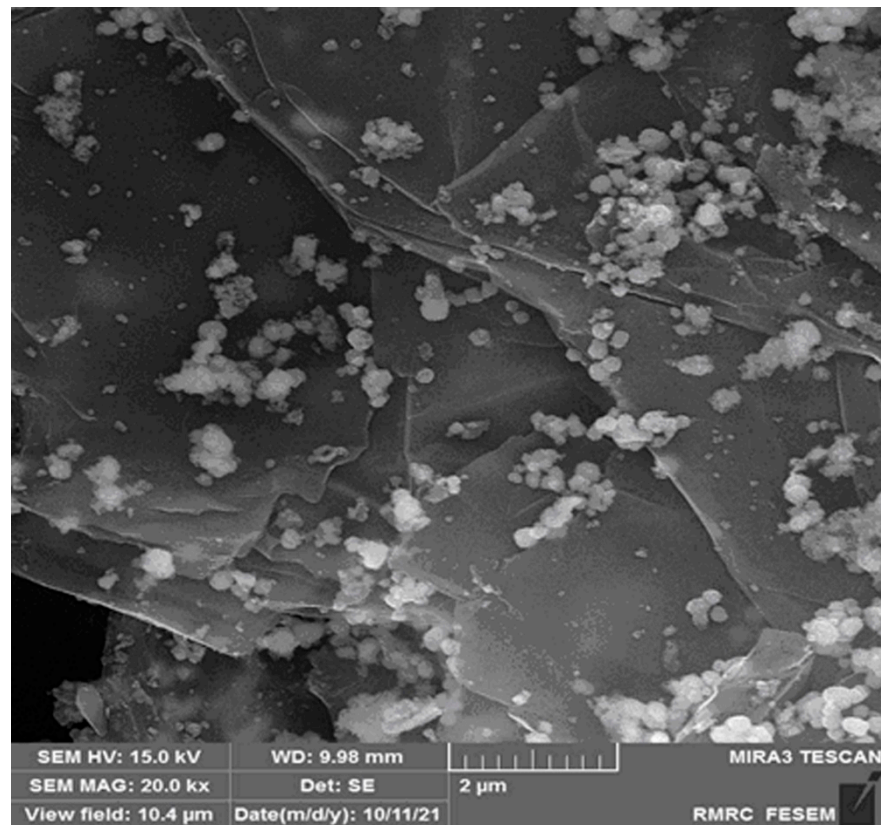


Figure 2. The FE-SEM image of graphene- Co_3O_4 nanocomposite.

The EDS spectrum (Figure 3) also shows the existence of C, O, and Co elements in the graphene- Co_3O_4 nanocomposite.

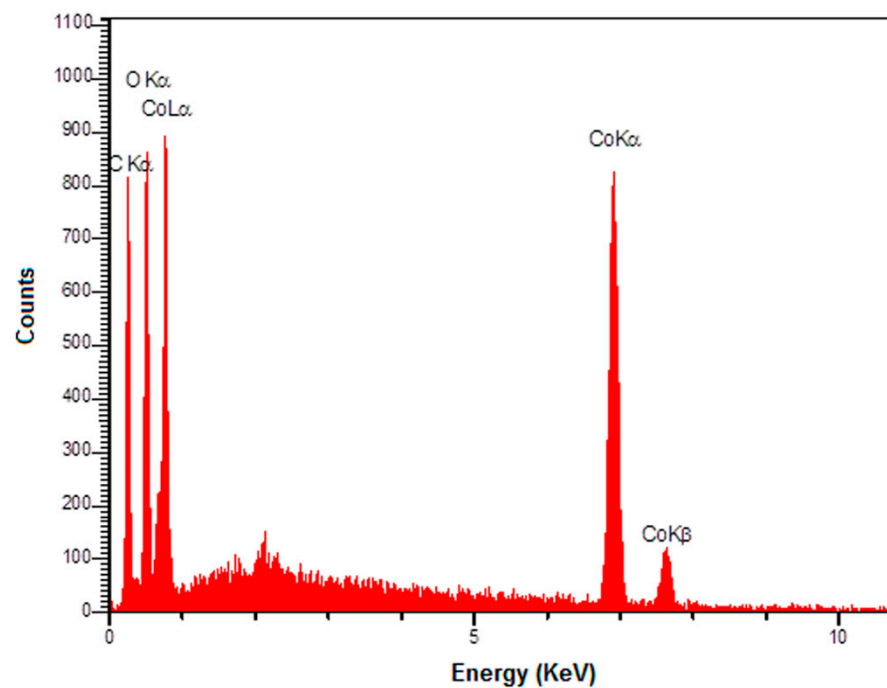


Figure 3. The EDS spectrum of graphene- Co_3O_4 nanocomposite.

The crystalline phase of the as-prepared graphene- Co_3O_4 nanocomposite was confirmed by XRD measurements (Figure 4). The obtained XRD patterns can be indexed according to the (111), (220), (311), (222), (400), (511), (440), (620), and (622) reflections of Co_3O_4 [JCPDS: 01-076-1802]. No distinct diffraction peak was observed for graphene, as the refraction of graphene is weaker than well-crystalline Co_3O_4 .

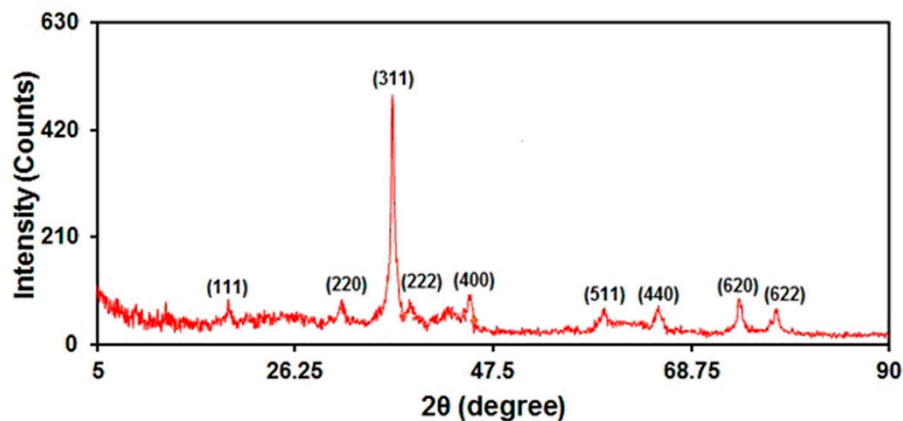


Figure 4. XRD pattern of graphene- Co_3O_4 nanocomposite.

3.2. Electrochemical Response of Morphine on Diverse Electrodes

The electrochemical response of morphine oxidation in the 0.1 M PBS adjusted to variable pH values (2.0–9.0) was explored to determine the influence of the electrolyte solution's pH. The results showed that the peak current of morphine oxidation depended on the pH value, so that it increased as the pH increased, reaching its maximum at a pH of 7.0, and then decreased with further increasing pH values (Figure 5). Hence, the pH value of 7.0 was considered to be the optimum for subsequent electrochemical determinations.

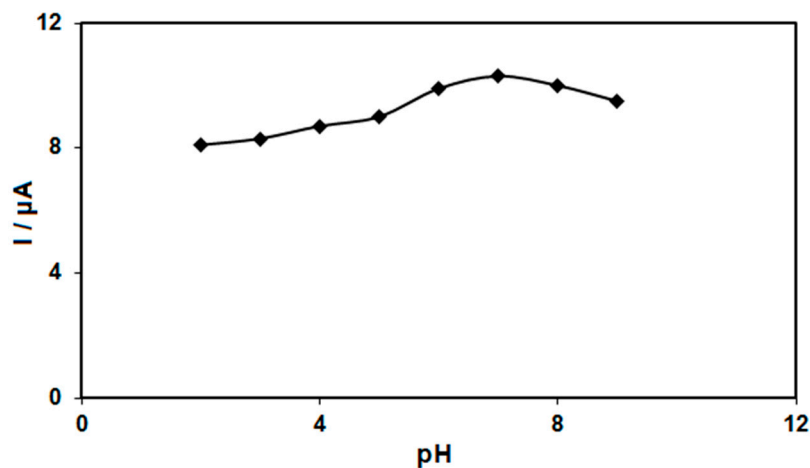


Figure 5. Plot of I_p vs. pH obtained from DPVs of graphene- Co_3O_4 /SPGE in a solution containing 200.0 μM of morphine in 0.1 PBS with different pHs (2.0, 3.0, 4.0, 5.0, 6.0, 7.0, 8.0 and 9.0).

CV was performed to clarify the electrochemical behavior of morphine on: (a) an unmodified SPGE; (b) a graphene/SPGE; (c) graphene- Co_3O_4 /SPGE surfaces. Figure 6 compares the bare SPGE and the graphene- Co_3O_4 /SPGE for 200.0 μM morphine oxidation in 0.1 M PBS at the pH value of 7.0. The morphine oxidation displayed a tiny and wide peak (3.0 μA) at the potential of 440 mV on the bare SPGE surface. Furthermore, the graphene/SPGE exhibited a shift in the peak current towards the more negative potentials (370 mV) by raising the amount of current (5.2 μA). The graphene- Co_3O_4 -modified SPGE exhibited a shift in the peak current towards the more negative potentials (250 mV) by

raising the amount of current (10.3 μA). Such an improvement can appear because of an appreciable catalytic impact of the graphene- Co_3O_4 nanocomposite for the morphine oxidation.

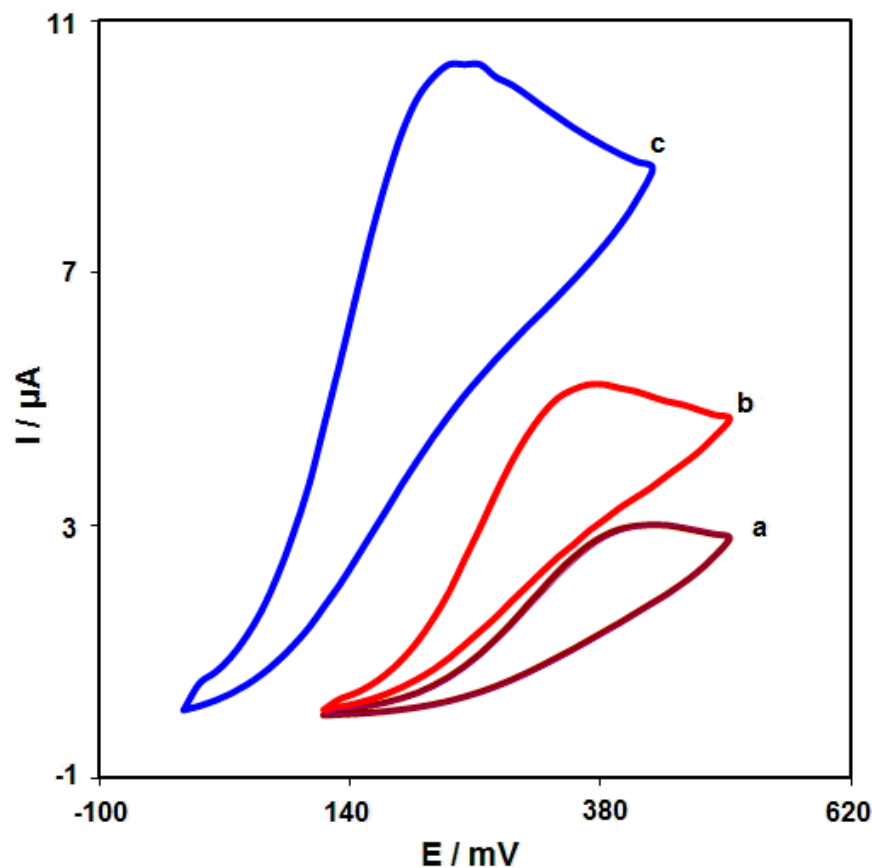


Figure 6. Cyclic voltammograms captured for oxidation of morphine (200.0 μM) in PBS (0.1 M; pH = 7.0) on: (a) unmodified SPGE; (b) graphene/SPGE; (c) graphene- Co_3O_4 /SPGE with the scan rate of 50 mV/s.

3.3. Effect of Scan Rate

The linear sweep voltammograms (LSVs) were recorded for the oxidation of morphine (100.0 μM) on the graphene- Co_3O_4 /SPGE under variable scan rates (Figure 7). There was an apparent gradual elevation in the oxidation peak by raising scan rate ranging from 10 to 300 mV/s. As seen in Figure 7 (Inset), the anodic peak current (I_{pa}) had a linear association with the scan rate square root ($U^{1/2}$). The regression equation was obtained to be $I_{pa} (\mu\text{A}) = 1.136 U^{1/2} (\text{mV s}^{-1})^{1/2} - 1.7492$ ($R^2 = 0.9993$), representing a controlled diffusion process of the morphine oxidation on the graphene- Co_3O_4 /SPGE.

3.4. Chronoamperometric Analysis

Chronoamperometry was used to explore the morphine catalytic oxidation on the graphene- Co_3O_4 /SPGE surface. Chronoamperometric analysis was performed for variable morphine contents on the graphene- Co_3O_4 /SPGE at the working electrode potential of 300 mV. The chronoamperograms captured for variable morphine contents on the graphene- Co_3O_4 /SPGE are seen in Figure 8. The Cottrell equation explains the current (I) for electrochemical reaction of an electroactive material with a D value (diffusion coefficient) under a mass transport limited condition. Figure 8 shows a linear relationship of the I value with $t^{-1/2}$ for the oxidation of variable morphine contents. The slopes from the obtained straight lines were plotted against variable morphine contents (Figure 8). The plotted slope and Cottrell equation estimated the D value to be $6.0 \times 10^{-6} \text{ cm}^2/\text{s}$ for morphine.

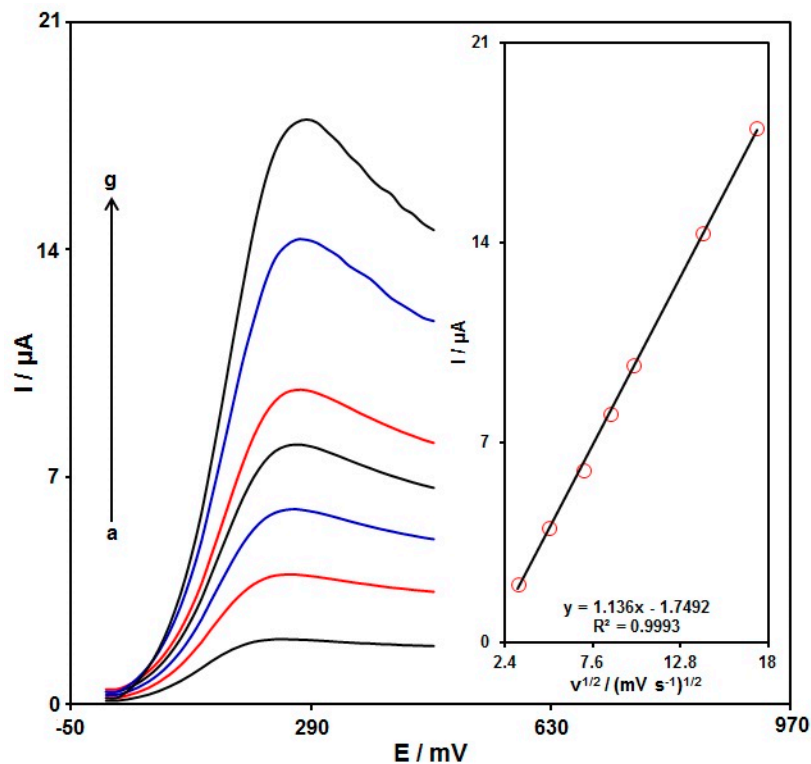


Figure 7. LSVs captured for the oxidation of morphine (100.0 μM) on the graphene-Co₃O₄/SPGE under variable scan rates (a–g: 10, 25, 50, 75, 100, 200, and 300 mV/s); Inset: the correlation of I_{pa} with $v^{1/2}$.

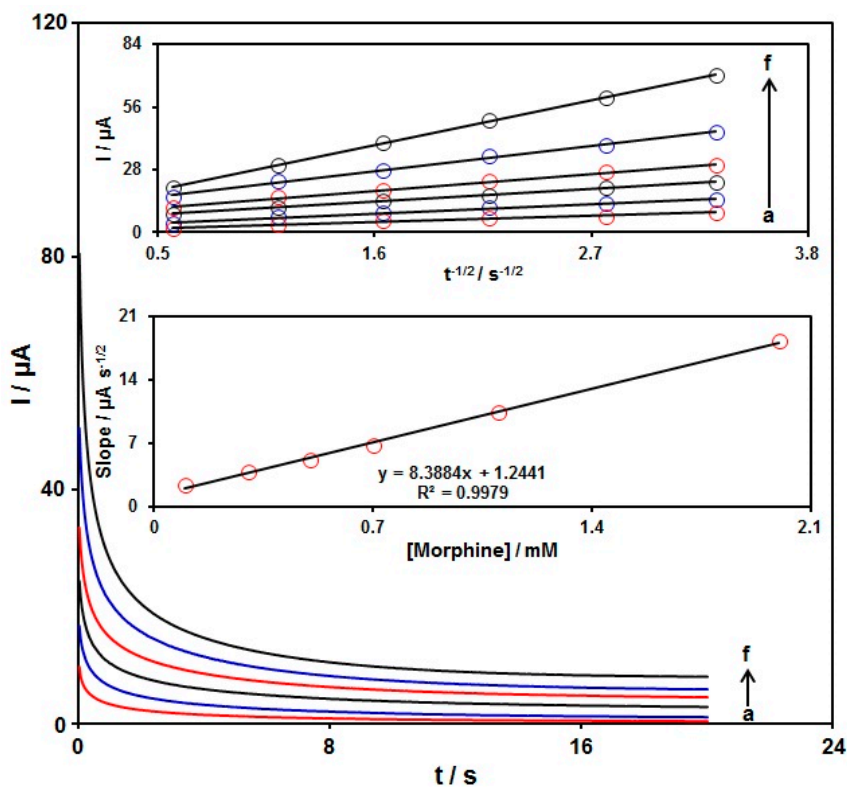


Figure 8. Chronoamperometric behavior of graphene-Co₃O₄/SPGE in PBS (0.1 M; pH = 7.00) at a potential of 300 mV for variable morphine contents (a–f: 0.1, 0.3, 0.5, 0.7, 1.1, and 2.0 mM); Insets: (top) plots of I vs. $t^{-1/2}$; (bottom) plots of the slopes from the straight lines vs. morphine level.

3.5. DPV Analysis of Morphine

DPV analysis was performed for variable morphine contents to explore the linear dynamic range, limit of detection (LOD), and sensitivity of the graphene- Co_3O_4 /SPGE under optimized experimental circumstances (Figure 9) (step potential = 0.01 V and pulse amplitude = 0.025 V). As expected, the elevation in the morphine level enhanced the peak current. Figure 9 (Inset) shows a linear proportion of the oxidation peak currents to variable morphine contents (0.02 μM to 575.0 μM) with the linear regression equation of $I_{pa} (\mu\text{A}) = 0.049 \text{ C-morphine} + 0.6857$ ($R^2 = 0.9994$), and the sensitivity of $0.4 \mu\text{A}/(\mu\text{M cm}^2)$. In the equation of $\text{LOD} = 3 \sigma/m$, the σ stands for the standard deviation of the response for blank solution, and the m for the slope from the standard graph. The LOD was estimated at $0.007 \mu\text{M}$ for morphine determination on the graphene- Co_3O_4 /SPGE. Table 1 compares the efficiency of the morphine sensor prepared by the graphene- Co_3O_4 -nanocomposite-modified SPGE and other reported works.

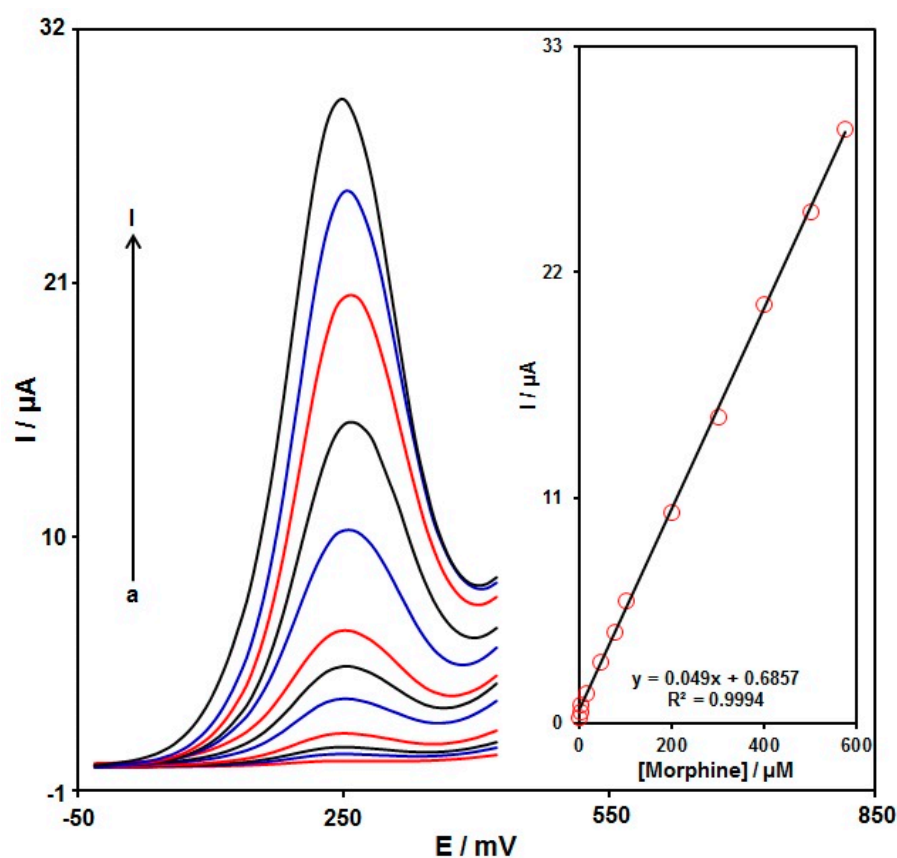


Figure 9. DPVs captured for the oxidation of variable morphine contents on the graphene- Co_3O_4 /SPGE under variable contents (a–l: 0.02, 0.25, 2.5, 15.0, 45.0, 75.0, 100.0, 200.0, 300.0, 400.0, 500.0, and 575.0 μM); Inset: Calibration curve of voltammetric response (I_{pa}) against morphine level.

3.6. DPV Analysis for Determination of Morphine in the Presence of Diclofenac

To confirm the ability of the graphene- Co_3O_4 /SPGE for co-detection of morphine with diclofenac, the electrochemical responses of these analytes were detected by simultaneously changing the concentration of both analytes in PBS at a pH of 7.0. As seen in Figure 10, with the concurrent change in their concentrations, two non-interference peaks were found on DPV curves (step potential = 0.01 V and pulse amplitude = 0.025 V). The peak currents of both morphine and diclofenac oxidation displayed a linear elevation with the respective concentrations (morphine concentration range between 2.5 μM and 550.0 μM and diclofenac concentration range between 2.5 μM and 700.0 μM) (Figure 10A,B). The intensity of the

peak current showed good linearity with the target concentration change, meaning the possibility of detecting morphine and diclofenac in the blended solution.

Table 1. Comparison of the efficiency of the graphene-Co₃O₄/SPGE sensor with other reported modified electrodes for morphine determination.

Electrochemical Sensor	Electrochemical Method	Linear Range	LOD	Ref.
ZnO-multi-walled carbon nanotubes (MWCNTs)-ionic liquid/carbon-paste electrode (CPE)	-	0.1–700.0 μ M	0.06 μ M	[1]
Polydopamine-modified MWCNTs-glassy Carbon Electrode (GCE)	DPV	0.075–75.0 μ M	0.06 μ M	[37]
Poly(cetyltrimethylammonium bromide)/graphene oxide/GCE	DPV	50–60 μ M	0.36 μ M	[38]
Au Nanoparticles/CPE	DPV	4.0×10^{-7} – 2.0×10^{-4} M	4.21 nM	[39]
Au nanoparticles and Co phthalocyanine/CPE	DPV	4.0×10^{-7} – 9.0×10^{-4} M	5.48×10^{-9} M	[40]
Ordered mesoporous carbon/GCE	CV	0.1–20 μ M	10 nM	[44]
Graphene-Co ₃ O ₄ /SPGE	DPV	0.02–575.0 μ M	0.007 μ M	This work

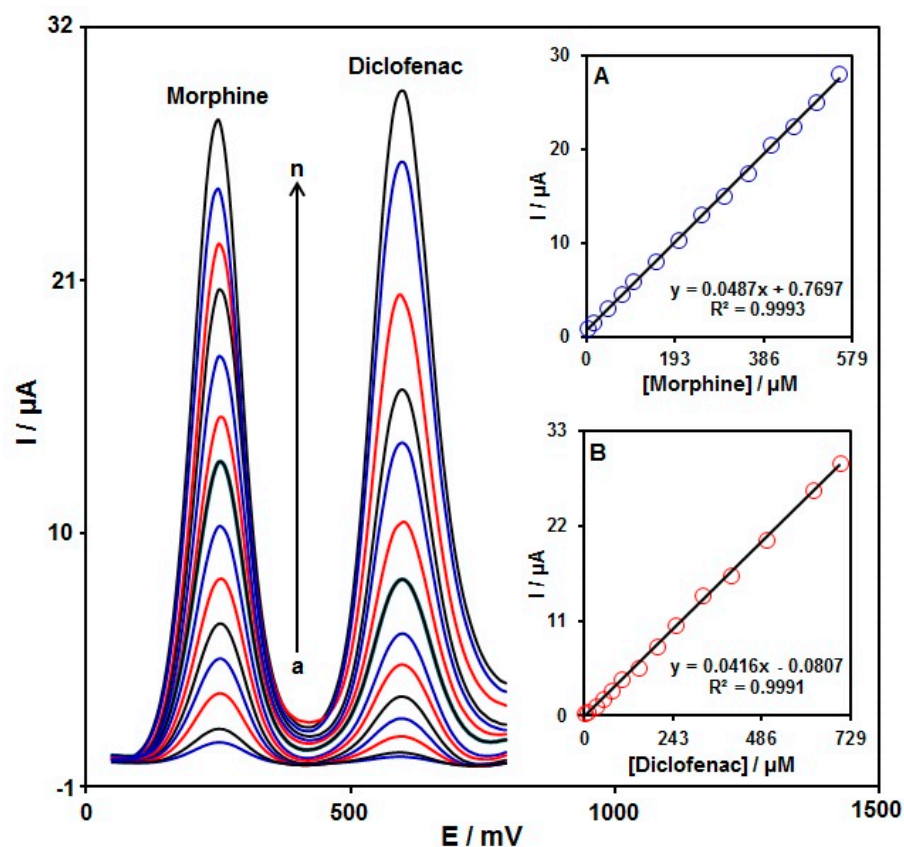


Figure 10. DPVs of graphene-Co₃O₄/SPGE in PBS (0.1 M, pH = 7.0) with variable morphine concentrations (a–n: 2.5, 15.0, 45.0, 75.0, 100.0, 150.0, 200.0, 250.0, 300.0, 350.0, 400.0, 450.0, 500.0, and 550.0 μ M) and diclofenac (a–n: 2.5, 10.0, 30.0, 50.0, 75.0, 100.0, 150.0, 200.0, 250.0, 325.0, 400.0, 500.0, 625.0, and 700.0 μ M). Inserts: (A) plot of peak current versus morphine concentration; (B) plot of peak current versus diclofenac concentration.

3.7. Reproducibility, and Stability

To test the reproducibility, seven graphene-Co₃O₄/SPGEs produced by the same procedures were applied to measure 50.0 μM morphine under identical circumstances; the obtained relative standard deviation (RSD) of 6.1% suggested commendable reproducibility.

To test the graphene-Co₃O₄/SPGE's stability, the current responses of 50.0 μM morphine were measured following 20-day storage of the sensor at ambient temperature. The decrease in the peak current of morphine to 97.1% of its original response demonstrated appreciable stability.

3.8. Interference Study

The selectivity studies of the graphene-Co₃O₄/SPGE for the determination of 50.0 μM morphine were performed in the presence of different interfering substances. The tolerance limit was defined as the maximum concentration of the interfering substance that caused an approximately ±5% relative error in the determination. The results revealed that 480-fold of Na⁺, Mg²⁺, Cl⁻, Br⁻, NH₄⁺, fructose, glucose, and lactose; 300-fold of histidine, phenyl alanine, alanine, methionine, glycine, methanol, ethanol, and tryptophan; 230-fold of ascorbic acid (after removal with ascorbic oxidase), thiourea, tyrosine, cysteine, and acetaminophen; and 10-fold of dopamine, uric acid, and methyldopa did not show interference in determination. Those results confirm the suitable selectivity of the proposed sensor for the determination of morphine.

3.9. Analysis of Real Specimens

The practical applicability of the graphene-Co₃O₄/SPGE was tested by sensing morphine and diclofenac in morphine ampoule, diclofenac tablet, and urine specimens using the DPV procedure and the standard addition method, the results of which can be seen in Table 2. The recovery rate was between 96.5% and 104.0%, and all RSD values were ≤3.5%. According to the experimental results, the graphene-Co₃O₄/SPGE sensor possessed a high potential for practical applicability.

Table 2. Voltammetric sensing of morphine and diclofenac in real specimens using graphene-Co₃O₄/SPGE. All concentrations are in μA (n = 5).

Sample	Spiked (μM)		Found (μM)		Recovery (%)		R.S.D. (%)	
	Morphine	Diclofenac	Morphine	Diclofenac	Morphine	Diclofenac	Morphine	Diclofenac
Morphine ampoule	0	0	3.0	-	-	-	3.2	-
	1.0	5.0	3.9	5.1	97.5	102.0	2.7	3.5
	2.0	7.0	5.1	6.9	102.0	98.6	1.9	2.1
	3.0	9.0	6.2	8.8	103.3	97.8	2.4	2.6
	4.0	11.0	6.9	11.1	98.6	100.9	3.0	1.8
Diclofenac tablet	0	0	-	4.0	-	-	-	2.9
	5.0	1.0	4.9	5.1	98.0	102.0	3.0	2.4
	7.5	3.0	7.6	6.8	101.3	97.1	2.0	3.3
	10.0	5.0	9.9	9.1	99.0	101.1	2.9	1.7
	12.5	7.0	13.0	10.9	104.0	99.0	2.3	2.3
Urine	0	0	-	-	-	-	-	-
	4.0	5.5	3.9	5.6	97.5	101.8	3.1	1.8
	6.0	7.5	6.2	7.4	103.3	98.7	2.8	2.9
	8.0	9.5	8.1	9.8	101.25	103.2	1.9	3.5
	10.0	11.5	9.9	11.1	99.0	96.5	2.2	2.4

4. Conclusions

The present work examined a simple and sensitive electrochemical morphine sensor using a graphene-Co₃O₄-nanocomposite-modified SPGE. The graphene-Co₃O₄/SPGE sensor showed excellent activity towards oxidation of morphine at a low overpotential

~250 mV with a high current response of around 10.3 μA . The graphene- Co_3O_4 /SPGE had a wider linear response in concentration ranging from 0.02 to 575.0 μM with an LOD as narrow as 0.007 μM . Furthermore, by separating the peaks of morphine and diclofenac oxidation with a potential difference of 350 mV, it was possible to measure morphine in the combination with diclofenac using the modified sensor. Finally, the proposed electrode was successfully employed for the determination of diclofenac and morphine in real specimens such as diclofenac tablet, morphine ampoule, and urine applying standard addition method with acceptable recoveries. The novelty of this work concerns the observed synergistic effect of graphene nanosheets and a Co_3O_4 nanoparticles-modified electrode for enhanced electrochemical sensing of morphine in the presence of diclofenac.

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References

1. Barthwal, S.; Singh, B.; Singh, N.B. A novel electrochemical sensor fabricated by embedding ZnO nano particles on MWCNT for morphine detection. *Mater. Today: Proc.* **2018**, *5*, 9061–9066. [[CrossRef](#)]
2. Maccaferri, G.; Terzi, F.; Xia, Z.; Vulcano, F.; Liscio, A.; Palermo, V.; Zanardi, C. Highly sensitive amperometric sensor for morphine detection based on electrochemically exfoliated graphene oxide. Application in screening tests of urine samples. *Sens. Actuators B Chem.* **2019**, *281*, 739–745. [[CrossRef](#)]
3. Dehdashtian, S.; Gholivand, M.B.; Shamsipur, M. Construction of a sensitive and selective sensor for morphine using chitosan coated Fe_3O_4 magnetic nanoparticle as a modifier. *Mater. Sci. Eng. C* **2016**, *58*, 53–59. [[CrossRef](#)]
4. Eissa, S.; Zourob, M. Competitive voltammetric morphine immunosensor using a gold nanoparticle decorated graphene electrode. *Microchim. Acta* **2017**, *184*, 2281–2289. [[CrossRef](#)]
5. Schug, S.A.; Sidebotham, D.A.; McGuinness, M.; Thomas, J.; Fox, L. Acetaminophen as an adjunct to morphine by patient-controlled analgesia in the management of acute postoperative pain. *Anesth. Analg.* **1998**, *87*, 368–372. [[CrossRef](#)]
6. Goyal, R.N.; Chatterjee, S.; Agrawal, B. Electrochemical investigations of diclofenac at edge plane pyrolytic graphite electrode and its determination in human urine. *Sens. Actuators B Chem.* **2010**, *145*, 743–748. [[CrossRef](#)]
7. Eteya, M.M.; Rounaghi, G.H.; Deiminiat, B. Fabrication of a new electrochemical sensor based on AuPt bimetallic nanoparticles decorated multi-walled carbon nanotubes for determination of diclofenac. *Microchem. J.* **2019**, *144*, 254–260. [[CrossRef](#)]
8. Arvand, M.; Gholizadeh, T.M.; Zanjanchi, M.A. MWCNTs/ $\text{Cu}(\text{OH})_2$ nanoparticles/IL nanocomposite modified glassy carbon electrode as a voltammetric sensor for determination of the non-steroidal anti-inflammatory drug diclofenac. *Mater. Sci. Eng. C* **2012**, *32*, 1682–1689. [[CrossRef](#)]
9. Izhar, M.; Folker, A.; Gandhi, A.; Alausa, T.; Alam, A.; Bakris, G.L. P-228: A comparative study of celecoxib versus diclofenac on blood pressure control and renal function in hypertensive African Americans and Hispanics: A randomized crossover study. *Am. J. Hypertens.* **2002**, *15*, 112A. [[CrossRef](#)]
10. Fatta-Kassinos, D.; Hapeshi, E.; Achilleos, A.; Meric, S.; Gros, M.; Petrovic, M.; Barcelo, D. Existence of pharmaceutical compounds in tertiary treated urban wastewater that is utilized for reuse applications. *Water Resour. Manag.* **2011**, *25*, 1183–1193. [[CrossRef](#)]
11. Miranda, H.F.; Silva, E.; Pinardi, G. Synergy between the antinociceptive effects of morphine and NSAIDs. *Can. J. Physiol. Pharmacol.* **2004**, *82*, 331–338. [[CrossRef](#)] [[PubMed](#)]
12. Abdolmohammad-Zadeh, H.; Zamani, A.; Shamsi, Z. Preconcentration of morphine and codeine using a magnetite/reduced graphene oxide/silver nano-composite and their determination by high-performance liquid chromatography. *J. Chromatogr. A* **2019**, *1590*, 2–9. [[CrossRef](#)]
13. Nakhaei, J.M.; Jamali, M.R.; Sohrabnezhad, S.; Rahnama, R. Solvent-Assisted dispersive solid phase extraction of diclofenac from human serum and pharmaceutical tablets quantified by high-performance liquid chromatography. *Microchem. J.* **2020**, *152*, 104260. [[CrossRef](#)]
14. Abbott, R.W.; Townshend, A.; Gill, R. Determination of morphine by flow injection analysis with chemiluminescence detection. *Analyst* **1986**, *111*, 635–640. [[CrossRef](#)]

15. Song, J.; Sun, P.; Ji, Z.; Li, J. Flow injection determination of diclofenac sodium based on its sensitizing effect on the chemiluminescent reaction of acidic potassium permanganate–formaldehyde. *Luminescence* **2015**, *30*, 32–37. [[CrossRef](#)]
16. Sebaiy, M.M.; El-Shanawany, A.A.; Baraka, M.M.; Abdel-Aziz, L.M.; Isbell, T.A.; Colyer, C.L. Determination of morphine and its metabolites in human urine by capillary electrophoresis with laser induced fluorescence detection employing on-column labeling with a new boronic acid functionalized squarylium cyanine dye. *Separations* **2016**, *3*, 1. [[CrossRef](#)]
17. Cunha, R.R.; Gimenes, D.T.; Munoz, R.A.; do Lago, C.L.; Richter, E.M. Simultaneous determination of diclofenac and its common counter-ions in less than 1 minute using capillary electrophoresis with contactless conductivity detection. *Electrophoresis* **2013**, *34*, 1423–1428. [[CrossRef](#)]
18. Rondina, R.V.; Bandoni, A.L.; Coussio, J.D. Quantitative determination of morphine in poppy capsules by differential spectrophotometry. *J. Pharm. Sci.* **1973**, *62*, 502–504. [[CrossRef](#)]
19. Gabhane, K.B.; Kasture, A.V.; Shrikhande, V.N.; Barde, L.N.; Wankhade, V.P. Simultaneous spectrophotometric determination of metaxalone and diclofenac potassium in combined tablet dosage form. *Int. J. Chem. Sci.* **2009**, *7*, 539–545.
20. Tang, S.H.; Chuang, C.H.; Chen, S.F.; Chuang, W.C.; Shang, H.S. Optimized procedures for simultaneous quantitation of low concentration levels of morphine and codeine in urine using gas chromatography–mass spectrometry. *J. Med. Sci.* **2020**, *40*, 68.
21. Česen, M.; Heath, E. Disk-based solid phase extraction for the determination of diclofenac and steroidal estrogens E1, E2 and EE2 listed in the WFD watch list by GC–MS. *Sci. Total Environ.* **2017**, *590*, 832–837. [[CrossRef](#)] [[PubMed](#)]
22. Kaale, E.; Nyamweru, B.C.; Manyanga, V.; Chambuso, M.; Layloff, T. The development and validation of a Thin Layer Chromatography densitometry method for the analysis of diclofenac sodium tablets. *Int. J. Chem. Anal. Sci.* **2013**, *4*, 73–79. [[CrossRef](#)]
23. Yang, J.; He, D.; Zhang, N.; Hu, C. Disposable carbon nanotube-based antifouling electrochemical sensors for detection of morphine in unprocessed coffee and milk. *J. Electroanal. Chem.* **2022**, *905*, 115997. [[CrossRef](#)]
24. Kumary, V.A.; Abraham, P.; Renjini, S.; Swamy, B.K.; Nancy, T.M.; Sreevalsan, A. A novel heterogeneous catalyst based on reduced graphene oxide supported copper coordinated amino acid–A platform for morphine sensing. *J. Electroanal. Chem.* **2019**, *850*, 113367. [[CrossRef](#)]
25. Mekassa, B.; Baker, P.G.; Chandravanshi, B.S.; Tessema, M. Synthesis, characterization, and preparation of nickel nanoparticles decorated electrochemically reduced graphene oxide modified electrode for electrochemical sensing of diclofenac. *J. Solid State Electrochem.* **2018**, *22*, 3607–3619. [[CrossRef](#)]
26. Kimuam, K.; Rodthongkum, N.; Ngamrojanavanich, N.; Chailapakul, O.; Ruecha, N. Single step preparation of platinum nanoflowers/reduced graphene oxide electrode as a novel platform for diclofenac sensor. *Microchem. J.* **2020**, *155*, 104744. [[CrossRef](#)]
27. Akbarian, Y.; Shabani-Nooshabadi, M.; Karimi-Maleh, H. Fabrication of a new electrocatalytic sensor for determination of diclofenac, morphine and mefenamic acid using synergic effect of NiO-SWCNT and 2,4-dimethyl-N/-[1-(2, 3-dihydroxy phenyl) methylidene] aniline. *Sens. Actuators B: Chem.* **2018**, *273*, 228–233. [[CrossRef](#)]
28. Mohammadzadeh Jahani, P.; Mohammadi, S.Z.; Khodabakhshzadeh, A.; Cha, J.H.; Shahedi Asl, M.; Shokouhimehr, M.; Peng, W.X. Simultaneous detection of morphine and diclofenac using graphene nanoribbon modified screen-printed electrode. *Int. J. Electrochem. Sci.* **2020**, *15*, 9037–9048. [[CrossRef](#)]
29. Baezzat, M.R.; Tavakkoli, N.; Zamani, H. Construction of a new electrochemical sensor based on MoS₂ nanosheets modified graphite screen printed electrode for simultaneous determination of diclofenac and morphine. *Anal. Bioanal. Chem. Res.* **2022**, *9*, 153–162.
30. Razmi, E.D.; Beitollahi, H.; Mahani, M.T.; Anjomshoa, M. TiO₂/Fe₃O₄/multiwalled carbon nanotubes nanocomposite as sensing platform for simultaneous determination of morphine and diclofenac at a carbon paste electrode. *Russ. J. Electrochem.* **2018**, *54*, 1132–1140. [[CrossRef](#)]
31. Mokhtari, A.; Karimi-Maleh, H.; Ensafi, A.A.; Beitollahi, H. Application of modified multiwall carbon nanotubes paste electrode for simultaneous voltammetric determination of morphine and diclofenac in biological and pharmaceutical samples. *Sens. Actuators B: Chem.* **2012**, *169*, 96–105. [[CrossRef](#)]
32. Antuña-Jiménez, D.; González-García, M.B.; Hernández-Santos, D.; Fanjul-Bolado, P. Screen-printed electrodes modified with metal nanoparticles for small molecule sensing. *Biosensors* **2020**, *10*, 9. [[CrossRef](#)] [[PubMed](#)]
33. Beitollahi, H.; Garkani Nejad, F.; Dourandish, Z.; Tajik, S. A novel voltammetric amaranth sensor based on screen printed electrode modified with polypyrrole nanotubes. *Environ. Res.* **2022**, *214*, 113725. [[CrossRef](#)]
34. Yan, L.; Zhang, C.; Xi, F. Disposable amperometric label-free immunosensor on chitosan–graphene-modified patterned ITO electrodes for prostate specific antigen. *Molecules* **2022**, *27*, 5895. [[CrossRef](#)] [[PubMed](#)]
35. Ma, K.; Yang, L.; Liu, J.; Liu, J. Electrochemical sensor nanoarchitectonics for sensitive detection of uric acid in human whole blood based on screen-printed carbon electrode equipped with vertically-ordered mesoporous silica-nanochannel film. *Nanomaterials* **2022**, *12*, 1157. [[CrossRef](#)] [[PubMed](#)]
36. Wang, K.; Yang, L.; Huang, H.; Lv, N.; Liu, J.; Liu, Y. Nanochannel array on electrochemically polarized screen printed carbon electrode for rapid and sensitive electrochemical determination of clozapine in human whole blood. *Molecules* **2022**, *27*, 2739. [[CrossRef](#)] [[PubMed](#)]
37. Sabeti, M.; Ensafi, A.A.; Rezaei, B. Polydopamine-modified MWCNTs-glassy Carbon Electrode, a Selective Electrochemical Morphine Sensor. *Electroanalysis* **2021**, *33*, 2286–2295. [[CrossRef](#)]

38. Abraham, P.; Renjini, S.; Nancy, T.E.; Kumary, V.A. Electrochemical synthesis of thin-layered graphene oxide-poly (CTAB) composite for detection of morphine. *J. Appl. Electrochem.* **2020**, *50*, 41–50. [[CrossRef](#)]
39. Atta, N.F.; Galal, A.; Azab, S.M. Electrochemical morphine sensing using gold nanoparticles modified carbon paste electrode. *Int. J. Electrochem. Sci.* **2011**, *6*, 5066–5081.
40. Atta, N.F.; Galal, A.; Abdel-Gawad, F.M.; Mohamed, E.F. Electrochemical morphine sensor based on gold nanoparticles metalphthalocyanine modified carbon paste electrode. *Electroanalysis* **2015**, *27*, 415–428. [[CrossRef](#)]
41. Qian, L.; Durairaj, S.; Prins, S.; Chen, A. Nanomaterial-based electrochemical sensors and biosensors for the detection of pharmaceutical compounds. *Biosens. Bioelectron.* **2021**, *175*, 112836. [[CrossRef](#)] [[PubMed](#)]
42. Cheng, Z.; Li, M.; Dey, R.; Chen, Y. Nanomaterials for cancer therapy: Current progress and perspectives. *J. Hematol. Oncol.* **2021**, *14*, 1–27. [[CrossRef](#)] [[PubMed](#)]
43. Yun, Q.; Li, L.; Hu, Z.; Lu, Q.; Chen, B.; Zhang, H. Layered transition metal dichalcogenide-based nanomaterials for electrochemical energy storage. *Adv. Mater.* **2020**, *32*, 1903826. [[CrossRef](#)]
44. Li, F.; Song, J.; Shan, C.; Gao, D.; Xu, X.; Niu, L. Electrochemical determination of morphine at ordered mesoporous carbon modified glassy carbon electrode. *Biosens. Bioelectron.* **2010**, *25*, 1408–1413. [[CrossRef](#)]
45. Nazari, Z.; Es' hagh, Z. A new electrochemical sensor for the simultaneous detection of morphine and methadone based on thioglycolic acid decorated CdSe doped graphene oxide multilayers. *Anal. Bioanal. Electrochem.* **2022**, *14*, 228–245.
46. Shalauddin, M.; Akhter, S.; Basirun, W.J.; Bagheri, S.; Anuar, N.S.; Johan, M.R. Hybrid nanocellulose/f-MWCNTs nanocomposite for the electrochemical sensing of diclofenac sodium in pharmaceutical drugs and biological fluids. *Electrochim. Acta* **2019**, *304*, 323–333. [[CrossRef](#)]
47. Istrate, O.M.; Rotariu, L.; Bala, C. Electrochemical determination of NADH using screen printed carbon electrodes modified with reduced graphene oxide and poly (allylamine hydrochloride). *Microchim. Acta* **2016**, *183*, 57–65. [[CrossRef](#)]
48. Gong, J.; Tang, H.; Wang, M.; Lin, X.; Wang, K.; Liu, J. Novel three-dimensional graphene nanomesh prepared by facile electro-etching for improved electroanalytical performance for small biomolecules. *Mater. Des.* **2022**, *215*, 110506. [[CrossRef](#)]
49. Zhou, H.; Dong, G.; Sailjoi, A.; Liu, J. Facile pretreatment of three-dimensional graphene through electrochemical polarization for improved electrocatalytic performance and simultaneous electrochemical detection of catechol and hydroquinone. *Nanomaterials* **2021**, *12*, 65. [[CrossRef](#)]
50. Zhou, H.; Ma, X.; Sailjoi, A.; Zou, Y.; Lin, X.; Yan, F.; Liu, J. Vertical silica nanochannels supported by nanocarbon composite for simultaneous detection of serotonin and melatonin in biological fluids. *Sens. Actuators B Chem.* **2022**, *353*, 131101. [[CrossRef](#)]
51. Chen, S.; Zhu, J.; Wu, X.; Han, Q.; Wang, X. Graphene oxide-MnO₂ nanocomposites for supercapacitors. *ACS Nano* **2010**, *4*, 2822–2830. [[CrossRef](#)] [[PubMed](#)]
52. Ganesamurthi, J.; Keerthi, M.; Chen, S.M.; Shanmugam, R. Electrochemical detection of thiamethoxam in food samples based on Co₃O₄ Nanoparticle@Graphitic carbon nitride composite. *Ecotoxicol. Environmen. Saf.* **2020**, *189*, 110035. [[CrossRef](#)] [[PubMed](#)]
53. Wang, X.; Dong, X.; Wen, Y.; Li, C.; Xiong, Q.; Chen, P. A graphene-cobalt oxide based needle electrode for non-enzymatic glucose detection in micro-droplets. *Chem. Commun.* **2012**, *48*, 6490–6492. [[CrossRef](#)] [[PubMed](#)]
54. Liang, K.; Fu, X.; Wu, L.; Qin, Y.; Song, Y. A novel tyrosinase biosensor based on graphene and Co₃O₄ nanocomposite materials for rapid determining catechol. *Int. J. Electrochem. Sci.* **2016**, *11*, 250–258.
55. Zhang, J.; Yang, L.; Pei, J.; Tian, Y.; Liu, J. A reagentless electrochemical immunosensor for sensitive detection of carcinoembryonic antigen based on the interface with redox probe-modified electron transfer wires and effectively immobilized antibody. *Front. Chem.* **2022**, *10*, 939736. [[CrossRef](#)]
56. Jana, M.; Saha, S.; Khanra, P.; Murmu, N.C.; Srivastava, S.K.; Kuila, T. Bioreduction of graphene oxide using drained water from soaked mung beans (*Phaseolus aureus* L.) and its application as energy storage electrode material. *Mater. Sci. Eng. B* **2014**, *186*, 33–40. [[CrossRef](#)]