

RESEARCH

Open Access



Nk3R blockade has sex-divergent effects on memory in mice

Antonio Florido^{1,2} , Estefanía Moreno³ , Enric I. Canela^{3,4}  and Raúl Andero^{1,2,5,6,7*} 

Abstract

Background: Memory consolidation is a process required for the formation of long-term memories. The G-protein-coupled receptor (GPCR) neurokinin-3-receptor (Nk3R) and its interactions with sex hormones seem important for the modulation of fear memory consolidation: Nk3R antagonism in male mice impairs fear memory, but enhances it in females. However, the involvement of the Nk3R as a modulator of other memories in both sexes remains unexplored.

Methods: We use the novel object recognition paradigm to test the effect of a systemic blockade of Nk3R during memory consolidation. Further, we assess the expression of estrogen receptor α , estrogen receptor β , and androgen receptor and heterodimerization with Nk3R in the medial prefrontal cortex (mPFC) and dorsal hippocampus (DH) of mice.

Results: Nk3R systemic antagonism elicited decreased memory consolidation in males while it enhanced it in females during proestrus. Nk3R analysis in the different subregions of the mPFC and the DH showed a higher expression in males than females. Moreover, females presented upregulation of the androgen receptor in the CA1 and the estrogen receptor beta in the cingulate cortex, CA1, and dentate gyrus. Overall, males presented an upregulation of the estrogen receptor alpha. We also explored the heterodimerization of GPCR membrane sex hormone receptors with the Nk3R. We found a higher percentage of Nk3R-membrane G-protein estrogen receptors heterodimers in the prelimbic cortex of the mPFC in females, suggesting an interaction of estradiol with Nk3R in memory consolidation. However, males presented a higher percentage of Nk3R-membrane G-protein androgen receptors heterodimers compared to females, pointing to an interaction of testosterone with Nk3R in memory consolidation.

Conclusion: These data propose novel ideas on functional interactions between Nk3R, sex hormones, estrogen receptors, and androgen receptors in memory consolidation.

Highlights

- Nk3R antagonism reduces recognition memory consolidation in male mice and increases it in proestrus females.
- Androgen receptor expression is higher in the CA1 compared to DG, CA3, and the mPFC.
- Estrogen receptor α expression is higher in males than in females in the DH and mPFC.
- Estrogen receptor β expression is greater in females than in males in the DG, CA1, and CG.
- Over 60% of Nk3R in the DH and mPFC is heterodimerized with membrane estrogen receptor and androgen receptor.

*Correspondence: Raul.Andero@uab.cat

¹ Institut de Neurociències, Universitat Autònoma de Barcelona, Cerdanyola del Vallès, 08193 Barcelona, Spain
Full list of author information is available at the end of the article



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

- Nk3R–GPAR is more abundant in males than in proestrus females, whereas Nk3R–GPER is greater in proestrus females compared to males.

Keywords: Neurokinin-3 receptor, Osanetant, Sex differences, Heterodimerization, Estradiol, Testosterone

Background

Recognition memory is a type of declarative memory that allows mammals to identify an object or situation as familiar, in contrast with novel objects or situations. The organism uses this memory to compare novel situations with past personal experiences generating a congruent response toward the novel situation [1]. Although rodents do not present declarative memory itself, since they do not have the ability to explicitly expose information, analogous procedures have been developed to test this type of memory in rats and mice [2]. Declarative memory in humans has usually been related to spatial and recognition memory in mice [3, 4], and in both cases, it has been established common fundamental encoding processes in the hippocampus and its connections [5]. Memory consolidation represents a crucial point in this process when recently labile short-term memories are transformed into long-term and stable memory traces [2]. This transformation typically requires cellular and molecular adaptations, such as genetic regulation of immediate-early gene expression or increased density of dendritic spines [1]. The tachykinin 2 (Tac2) pathway is characterized by the synthesis of neurokinin B (NkB) from the Tac2 gene and its binding to the neurokinin 3 receptor (Nk3R) mediating sex differences in the consolidation of fear memories [4, 6, 7]. Moreover, Nk3R manipulations in male rodents have been shown to modulate the formation of hippocampus-dependent declarative memory [8, 9]. Interestingly, Nk3R antagonists are safe and well-tolerated drugs in humans [10] which makes them potentially attractive candidates for treating mental disorders with memory alterations. Osanetant, a potent selective Nk3R antagonist, showed safety in a clinical trial for the treatment of psychotic and affective symptoms typical of schizophrenia [11]. Although this study was discontinued, Nk3R antagonists are currently under study as a promising solution for induced vasomotor symptoms (iVS) during menopause [12]. Surprisingly, there is only one prior study that has included females involving the Tac2 pathway and memory formation [7]. The abovementioned study shows how Nk3R blockade, as well as Tac2 silencing in the amygdala, reduce fear memory consolidation in male mice while increasing it in females [7]. In contrast, the role of the amygdala in recognition memory is limited and appears to be circumscribed to the familiarization phase, but not recall [13].

On the other hand, the hippocampus and the medial prefrontal cortex (mPFC) play a major role in the consolidation of recognition memory in mice [14, 15]. Hippocampal lesions after familiarization with two objects have been shown to disrupt object recognition in male rats [16]. Moreover, the dorsal hippocampus (DH) is highly interconnected with the mPFC [17, 18]. This network has been demonstrated to mediate whether an object was previously encountered or not [19]. Moreover, lesions of the prelimbic cortex (PrL) and infralimbic cortex (IL) of the mPFC produce deficits in recognition memory [20, 21]. These findings support the role of the mPFC–DH connections in the formation of recognition memory.

The Nk3R has been shown to interact with sex hormones in both males and females in the modulation of fear memory consolidation through the modulation of the intracellular cascade comprising Akt/GSK3 β / β -Catenin [7]. In female mice, estradiol enhances recognition memory consolidation, via the activation of the PI3K/Akt pathway in the DH [22] in a similar manner. In addition, testosterone in male rats has been shown to enhance synaptic plasticity in the dentate gyrus and increase reference memory measured by the inspection of non-baited holes in a hole board [23].

In recent years, a substantial amount of evidence has shown that G-protein-coupled receptors (GPCRs) may exist as dimeric or oligomeric complexes, exhibiting biochemical properties distinct from those of the protomers in modifying cellular responses [24]. Nk3R couples to the pertussis toxin-insensitive G-protein G $_q$, whose activation results in the production of inositol triphosphate and diacylglycerol, activating intracellular signaling cascades that result in memory modulation [6, 25]. Notwithstanding, heterodimerization of Nk3R with other GPCRs remains unexplored.

Nk3R antagonists have shown sex-dependent effects on fear memory and sex hormonal fluctuations [7, 26], but sex differences in recognition memory after Nk3R antagonism have not been tested yet. Here, we found that systemic Nk3R antagonism modulated recognition memory consolidation in an opposite-sex manner, reducing consolidation in males and increasing it in females during proestrus (high levels of estradiol). Then we measured the Nk3R, estrogen receptors α (ER α) and β (ER β), and androgen receptors (AR) in the DH and mPFC

uncovering sex differences. Finally, we studied protein interactions between the Nk3R and membrane G-protein estrogen receptors (GPER) and G-protein androgen receptors (GPAR) measuring heterodimerization as a possible mechanism of the sex differences in memory formation upon Nk3R antagonism.

Methods

Animals

All experiments used male and naturally cycling female C57BL/6J adult mice (Charles River, Barcelona, Spain), 8 weeks old at the beginning of the experiments and housed in groups of 4 in a room with a 12:12 h light/dark cycle (lights on from 8 am to 8 pm). All animals were housed with a controlled temperature of 22 ± 1 °C and humidity (~40%). Behavioral procedures and pharmacological manipulations began early in the light phase of the cycle. Male and female mice were housed separately in the same room. Experiments in mice were performed under the approved ethics protocol with reference CEEAH 3603. All procedures were approved by the Ethics Committee of the Universitat Autònoma de Barcelona and the Generalitat de Catalunya. Experiments were also carried out following the European Communities Council Directive (2010-63-UE) and Spanish legislation (RD 53/2013).

Drugs

Osanetant was purchased from Merck (SML0798) and prepared for 5 and 10 mg/kg. The vehicle used for this preparation was 0.1% (v/v) Tween 20 in 0.9% (m/v) NaCl solution. Osanetant and its vehicle were administered at 10 mL/kg 30 min after the familiarization phase of the novel object recognition (NOR), to isolate the effect of the drug on the consolidation window of familiarization consolidation. Given previous data that support an attenuated effect of osanetant in memory during estrus, metestrus, and diestrus [7]; only females during the proestrus stage of the estrous cycle during familiarization were used in this experiment.

Determination of the estrous cycle

Determination of the estrous stage in females was performed by assessing vaginal smear cytology [27]. To assess the phase of the estrous cycle that females presented during the familiarization phase of the NOR, all-female mice were monitored for 3–4 consecutive cycles (approximately 10–14 days) before familiarization to test for regularity of the cycle. We performed a vaginal lavage with a 20- μ L pipette that was loaded with 10 μ L of standard NaCl 0.9% (w/v) solution, and later the tip was softly placed on the vaginal aperture. Before the collection of the sample, urine was softly removed using a tissue.

The 10 μ L of the sample were unloaded and collected 5 consecutive times to collect enough cells for the assessment, and later placed on an adhesion slide (Superfrost Plus, Thermo Fisher). All vaginal smear samples were collected between 9 and 10 am. Slides were dried using a hot plate (HI1220, Leica) at 37 °C for 30 min before staining in cresyl violet acetate (C5042, Merck) 0.1% (v/v) for 5 min, washed twice for 1 min in distilled water, and read in brightfield microscopy with a 10 \times or 20 \times objective in an Eclipse 80i microscope (Zeiss, Spain).

Three different cell types may appear in the preparation: cornified epithelial cells, round nucleated epithelial cells, or leukocytes. The different stages of the estrous cycle were assessed depending on the proportion of the abovementioned cells. Proestrus is characterized by a high proportion (>80%) of nucleated epithelial cells that might present very small amounts of cornified epithelial cells or leukocytes. Estrus is typically presented with cornified epithelial cells with a lower grade of staining than leukocytes and nucleated epithelial cells. Metestrus presents a mixture of cornified epithelial cells and a considerable proportion of leukocytes. Diestrus is characterized by >90% of leukocytes that might present a very small proportion of round nucleated epithelial cells [27].

Novel object recognition

NOR is widely accepted as a test to assess prefrontal and hippocampal-dependent learning and memory processes [28, 29]. Before starting the NOR test, a different cohort of animals was used to test that animals did not present differences in the time exploring each object of the NOR test.

All animals were habituated to the testing arena for 5 min for 2 consecutive days before the familiarization phase of the test. On the 3rd day, animals were placed in the arena where they freely explored two identical objects (either a yellow plastic pin or a purple crystal jar) for 15 min. Male and female mice received 5 mg/kg, 10 mg/kg osanetant, or vehicle 30 min after the familiarization phase of the NOR test before returning to their homecage. The percentage of time exploring each object was calculated as a measure of preference for any of two identical objects.

On the test day, the animals returned to the arena where they freely explored the two objects for 15 min. The novel object was the yellow plastic pin for half of the animals and the purple crystal jar for the other half. Further, we counterbalanced the position of the novel object (right or left) to avoid an effect of place preference. All phases of the test were carried out under ~15 lux. The relative time exploring the novel object, expressed in percentage, was calculated as a measure of recognition by an

experienced observer using a stopwatch. Heatmaps were obtained using Autotype for MATLAB (Fig. 1K, L).

Immunofluorescent determination of Nk3R, ER α , ER β , and AR

Immunohistochemical assays were performed on naïve males and proestrus females whose tissue was collected early in the morning by transcardial perfusion with 50 mL of 4% (m/v) of buffered paraformaldehyde (PFA) (Casa Álvarez, Barcelona, Spain). These animals were naïve and had not undergone any treatment or behavioral procedure before tissue collection. Brains were stored in the same solution at 4 °C for postfixation overnight before being transferred to 30% (m/v) sucrose in Sorensen's PB buffer for equilibration in conic Falcon tubes. After 48 h of equilibration in the sucrose solution, brains were snap-frozen in chilled isopentane and stored at – 80 °C until sectioning.

Brains were coronally sectioned at 30 μ m/slice using a Leica cryostat (– 20 °C for the chamber, – 18 °C for the sample) and stored in cryoprotective solution before staining. For staining, brain slices containing the mPFC and DH were rinsed in KPBS 1 \times three times for 10 min each, before spending 1 h in incubation in blocking buffer (5% normal donkey serum in 0.4% Triton X solution prepared with KPBS 1 \times), to avoid nonspecific binding of the primary antibody. Then, slices were incubated overnight at 4 °C in 300 μ L of blocking buffer containing either one of the following combinations: rabbit anti-Nk3R (1:1500, donated by Phillipe Cioffi) and mouse monoclonal anti-ER α (1:50, sc-8002, Santa Cruz Biotechnology), rabbit anti-Nk3R and mouse monoclonal anti-ER β (1:500, ab288, Abcam) or rabbit anti-Nk3R and mouse monoclonal anti-AR (1:50, sc-7305, Santa Cruz Biotechnology). After primary antibody incubation, brain slices were rinsed three times, 10 min each, with KPBS 1 \times and incubated at room temperature for 2 h in blocking buffer containing donkey anti-rabbit AlexaFluor488 (1:1000, 115-546-072, Jackson ImmunoResearch) and donkey anti-mouse Rhodamine Red (1:1000, 715-295-150, Jackson

ImmunoResearch). Slices were then rinsed again and incubated for 3 min with 4',6-diamidino-2-phenylindole (DAPI) (1:10,000, 10236276001, Merck) to stain cell nuclei before mounting and covering.

Z-Stacks of the cingulate cortex (CG), PrL, IL, DG, CA1, and CA3 (50 μ m/interval) were acquired using a Leica SP3 confocal microscope with a PL APO 40 \times immersion objective, controlled by Zen 2010 software. The integrated density of the signal was then calculated using Fiji for Windows v1.52.n and used as a measure of the protein quantity of the taken pictures.

In situ proximity ligand assays (PLA)

With PLA, primary antibodies raised in different species are used to detect different proteins. Secondary antibodies specific to each primary antibody that include a short sequence of DNA are used to detect the presence of the primary antibodies. If both primary antibodies are part of the same protein complex, both DNA chains react and, after amplification of this DNA, fluorescent complementary oligonucleotides are used to detect the heterodimers. Naïve mice were deeply anesthetized and immediately perfused transcardially with buffered 4% paraformaldehyde (Casa Álvarez, Barcelona, Spain). Brains were removed and post-fixed overnight in the same solution, cryoprotected by immersion of 30% sucrose in Sorensen's PB buffer for 48 h at 4 °C, and then snap-frozen in dry ice-cooled isopentane. Serial coronal cryostat sections (30 μ m) through the whole brain were collected in a cryoprotective solution and stored at – 20 °C until PLA experiments were performed.

Immediately before the assay, mouse brain sections were mounted on glass slides, washed in PBS, permeabilized with PBS containing 0.01% Triton X-100 for 10 min, and successively washed with PBS. Interactions were detected using the Duolink in situ PLA detection Kit (Sigma-Aldrich, St. Louis, MO, USA) following the supplier's instructions. A mixture of the primary antibodies rabbit anti-Nk3R antibody (1:100; donated by Phillipe Cioffi (INSERM)) directly linked to a plus PLA probe and rabbit anti-GPER antibody (1:100; Sigma; cat

(See figure on next page.)

Fig. 1 Novel object recognition (NOR) test after the administration of osanetant or vehicle. **A** Schematic representation of the behavioral procedure. **B** Males and females' preference test between the two objects used in the test. **C, E** Familiarization phase of the NOR task in males before receiving osanetant (5 or 10 mg/kg) or vehicle. **D** NOR test after receiving 5 mg/kg or vehicle after familiarization in males. **F** NOR test after receiving 10 mg/kg or vehicle after familiarization in males. **G, I** Familiarization phase of the NOR task in females (proestrus) before receiving osanetant (5 or 10 mg/kg) or vehicle. **H** NOR test after receiving 5 mg/kg or vehicle after familiarization in females (proestrus). **J** NOR test after receiving 10 mg/kg or vehicle after familiarization in females (proestrus). The recognition index is calculated by dividing the time exploring the novel object by the total time of exploration and expressed as a percentage. The dashed line in each graph represents 50% of the relative exploration time. **K** Representative heatmaps of the NOR test showing males that received vehicle or osanetant (10 mg/kg). **L** Representative heatmaps of the NOR test showing females (proestrus) that received vehicle or osanetant (10 mg/kg). Data were analyzed using Three-way ANOVA for the familiarization phase (Sex \times Object \times Treatment) or Two-way ANOVA (Sex \times Treatment). * $p < 0.05$. Obj1 = Object 1; Obj2 = Object 2

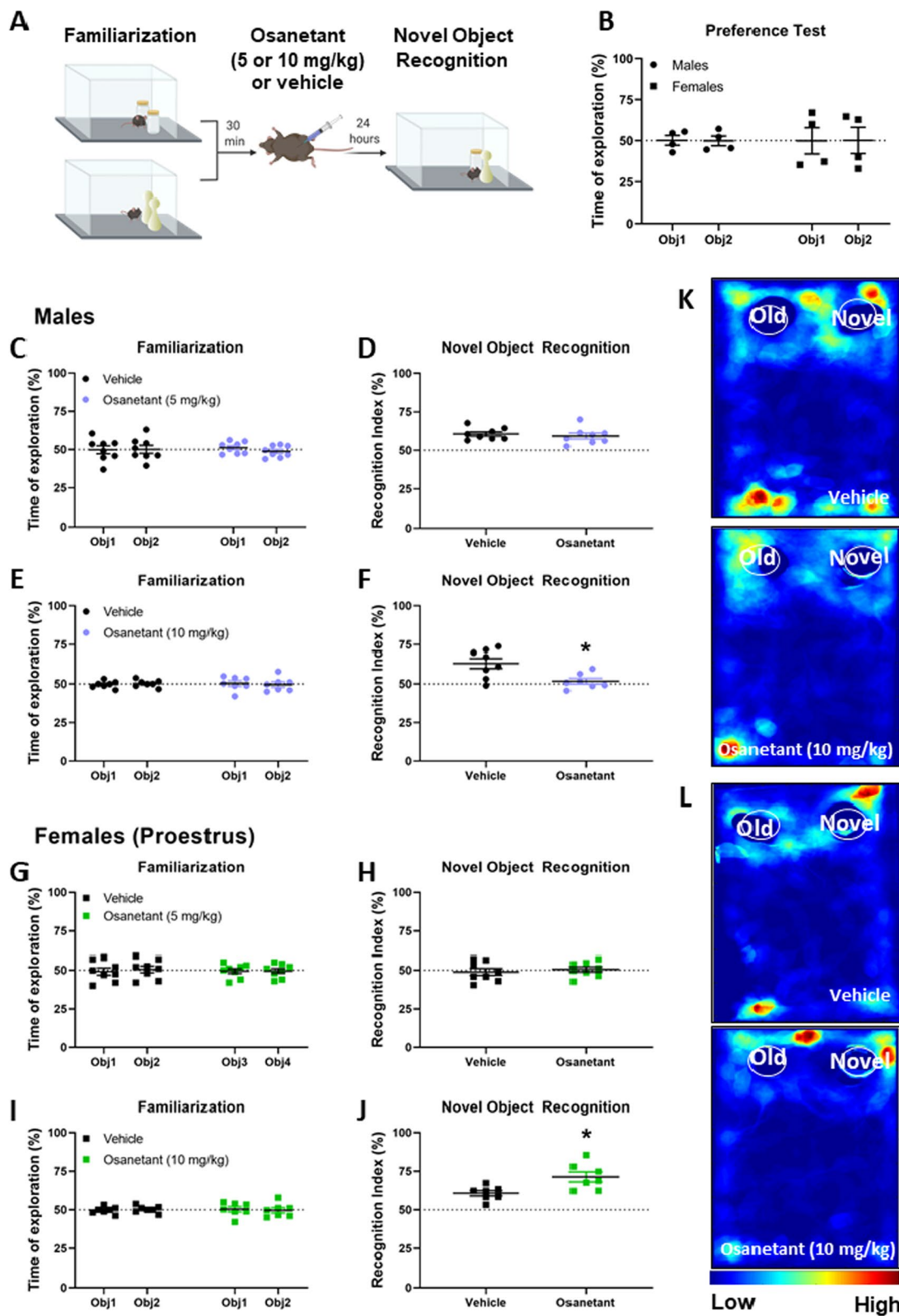


Fig. 1 (See legend on previous page.)

HPA027052) directly linked to a minus PLA probe were used to detect Nk3R–GPER interaction, or rabbit anti-GPAR antibody (1:100; Thermo Scientific, Fremont, CA, USA; Cat PA5-21074) directly linked to a minus PLA probe were used to detect Nk3R–GPAR interaction. For negative controls, one of the primary antibodies (anti-GPER or anti-GPAR) was omitted (data not shown). Then, samples were processed for ligation and amplification with a Detection Reagent Red and were mounted using a DAPI-containing mounting medium.

Samples were analyzed in a Leica SP2 confocal microscope (Leica Microsystems, Mannheim, Germany) equipped with an apochromatic 63× oil-immersion objective (1.4 numerical aperture), and 405 nm and 561 nm laser lines. For each field of view, a stack of two channels (one per staining) and 9 to 13 Z-stacks with a step size of 1 μm was acquired. Images were opened and processed with Image J software (National Institutes of Health, Bethesda, MD). Quantification of cells containing one or more red dots versus total cells (blue nuclei) was determined by using the Fiji package (<http://pacific.mpi-cbg.de/>), considering a total of 600–800 cells from 5 to 10 different fields (63×) within each region from three different animals. Nuclei and red dots were counted on the maximum projections of each image stack. After getting the projection, each channel was processed individually. The blue nuclei and red dots were segmented by filtering with a median filter, subtracting the background, enhancing the contrast with the Contrast Limited Adaptive Histogram Equalization (CLAHE) plug-in, and finally applying a threshold to obtain the binary image and the regions of interest (ROIs). Red spots were counted in each of the ROIs obtained in the nuclei images.

Experimental design and statistical analyses

The NOR test was analyzed using a multifactorial design Sex × Treatment (2 × 2) and the percentage of exploration of the new object as the dependent variable. The familiarization phase was analyzed using a multifactorial design Sex × Object × Treatment (2 × 2 × 2). Immunohistochemistry assays included main factors Sex and Area (2 × 6) to assess statistical differences or interactions of integrated density between groups on Nk3R, ERα, ERβ, and AR expression in the mPFC and the DH. Heterodimerization analysis included Area and Sex as main factors (6 × 2), and the percentage of cells including dimers among Nk3R positive cells was used as a dependent variable across the mPFC and the DH. The result of all statistical tests is summarized in Table 2.

All statistical analyses were performed using IBM SPSS v23.0. Outliers were detected using Grubb's test and removed when it was appropriate. Before proceeding with the null hypothesis testing, all datasets were

tested for normality and homoscedasticity using the Shapiro–Wilk test and Levene's test, respectively. Univariate ANOVAs were performed to assess the statistical significance of main effects and interactions, and significant interactions between main factors were analyzed using Bonferroni correction. Results are presented as mean ± SEM, and statistical significance was set at $p < 0.05$. All graphs presented in the figures were designed using Prism 7 (CA, USA).

Results

Nk3R blockade decreases recognition memory consolidation in males and increases it in females during proestrus in a dose-dependent manner

After assessing that animals presented no preference for any of the objects used in the test (Sex: [$F(1,24) = 0.057$, $p = 0.813$]; Object: [$F(1,24) = 0.072$, $p = 0.791$]; Sex × Object: [$F(1,24) = 0.072$, $p = 0.791$] (Fig. 1B), a different cohort of male and female mice underwent the familiarization phase of the object recognition. As expected, familiarization testing showed no differences in the time exploring each object either before receiving osanetant 5 mg/kg (Sex: [$F(1,56) = 0.000$, $p = 1.000$]; Object: [$F(1,56) = 0.031$, $p = 0.860$]; Treatment: [$F(1,56) = 0.000$, $p = 1.000$], Sex × Object: [$F(1,56) = 0.359$, $p = 0.552$], Sex × Treatment: [$F(1,56) = 0.000$, $p = 0.000$], Object × Treatment: [$F(1,56) = 1.128$, $p = 0.293$]; Sex × Object × Treatment: [$F(1,56) = 0.006$, $p = 0.939$] or 10 mg/kg (Sex: [$F(1,50) = 0.000$, $p = 1.000$]; Object: [$F(1,50) = 0.000$, $p = 1.000$]; Treatment: [$F(1,50) = 0.103$, $p = 0.749$]; Sex × Object: [$F(1,50) = 0.091$, $p = 0.764$]; Sex × Treatment: [$F(1,50) = 0.000$, $p = 1.000$]; Object × Treatment: [$F(1,50) = 1.638$, $p = 0.206$]; Sex × Object × Treatment: [$F(1,50) = 2.651$, $p = 0.110$] (Fig. 1C, E).

The recognition test, performed 24 h after receiving the drug, revealed no differences in recognition between males and females (no estrous cycle monitorization) that received 5 mg/kg (Sex: [$F(1,28) = 1.748$, $p = 0.197$]; Treatment: [$F(1,28) = 0.227$, $p = 0.603$]; Sex × Treatment: [$F(1,28) = 0.002$, $p = 0.965$] (Fig. 1D, H). Given that high levels of circulating estradiol have shown to be necessary for osanetant to modulate memory consolidation in a cued-fear conditioning task [7], we then tested females that were in the proestrus stage of the estrous cycle during the familiarization phase and the administration of the drug. In a different cohort of mice, males that received 10 mg/kg after the familiarization phase presented reduced memory consolidation compared to vehicle administered males, while proestrus females that received 10 mg/kg showed increased performance in the recognition test compared to the vehicle control group (Sex: [$F(1,25) = 10.595$, $p = 0.003$];

Treatment: [$F(1,25)=0.022$, $p=0.884$]; Sex \times Treatment: [$F(1,25)=17.128$, $p=0.000$]; treatment in males: [$F(1,25)=9.496$, $p=0.005$], and in proestrus females: [$F(1,25)=7.714$, $p=0.010$] (Fig. 1F, J).

This study extends our knowledge on Nk3R antagonism showing that it does not only exert an opposite-sex regulation of memory consolidation in fear memories as it has previously been demonstrated, but also in object recognition.

Nk3 and AR expression are higher in males compared to females (proestrus), while ER α and ER β expression depend on sex and area

Our previous research suggests the necessity of adult concentration of sex hormones for osanentan to modulate fear memory consolidation [7]. Following up on this data, we measured the expression of sex hormone receptors (androgen receptor -AR-, estrogen receptor alpha -ER α -, and estrogen receptor beta -ER β -) in areas related to NOR memory consolidation where the Nk3R might be exerting a regulating function.

To understand a possible role for Nk3R and sex hormones in memory consolidation, we performed immunolocalization assay to assess the expression of Nk3R, ER α , ER β and AR in the DH (CA1, CA3 and DG) and the mPFC (CG, PrL and IL), areas that are typically related to the consolidation of recognition memory. This analysis revealed an overall increased expression of Nk3R in males compared to females in the DH and the mPFC (Sex: [$F(1,43)=7.770$, $p=0.008$]; Area: [$F(5,44)=1.631$, $p=0.257$]; Sex \times Area: [$F(5,44)=0.768$, $p=0.758$] (Fig. 2A, E). The expression of ER α was also enhanced in male mice compared to proestrus female mice in the DH and the mPFC (Sex: [$F(1, 47)=7.001$, $p=0.011$]; Area [$F(5,47)=1.789$, $p=0.133$]; Sex \times Area: [$F(5,47)=1.326$, $p=0.270$] (Fig. 2D, E). Contrarily, the expression of ER β was greater in proestrus females compared to males in CA1, DG and CG (Sex: [$F(1,41)=115.275$, $p=0.000$]; Area [$F(5,41)=28.398$, $p=0.000$]; Sex \times Area [$F(5,41)=26.475$, $p=0.000$], CA1 [$F(1,41)=21.410$, $p=0.000$], CA3 [$F(1,41)=0.001$, $p=0.973$], DG [$F(1,41)=98.405$, $p=0.000$], CG [$F(1,41)=105.908$, $p=0.000$], PrL [$F(1,41)=0.029$, $p=0.866$], IL [$F(1,41)=0.000$, $p=0.985$] (Fig. 2C, E). Interestingly, no differences between areas or sexes were found in the analyses AR expression in the DH and mPFC of male and proestrus females, but a sex-independent increase of AR expression (Sex: [$F(1,42)=1.388$, $p=0.245$]; Area [$F(5,42)=9.144$, $p=0.000$]; Sex \times Area [$F(5,41)=0.475$, $p=0.793$]; CG vs rest of the areas $p=0.000$) (Fig. 2B, E).

Heterodimerization of Nk3R with the membrane estrogen receptor is enhanced in females, while males present higher rates of heterodimerization of Nk3R with the membrane androgen receptor

We hypothesized a possible interaction between the Nk3R and the membrane GPER in females and with the membrane GPAR in male mice. Data showed significant interactions between Nk3R-ER α , Nk3-ER β , and Nk3R-AR in proestrus females and males using in situ proximity ligand assay (PLA) in the DH and the mPFC. Given that estradiol and testosterone appear to be necessary for modulation of fear memories upon antagonism of the Nk3R in female and male mice, respectively [7], we aimed to test whether an activational effect of sex hormones could mediate the efficacy of Nk3R in the modulation of memory consolidation. To test that hypothesis, we assessed the presence of Nk3R-GPER and Nk3R-GPAR heterodimers in male and proestrus females through PLA in the DH and the mPFC. Results suggest significant interactions of area by sex [$F(5,120)=2.461$, $p=0.037$] and dimer by sex [$F(1,120)=159.754$, $p=0.000$] in the number of positive cells, while the interactions area by dimer ([$F(5,120)=0.383$, $p=0.860$]) and area by sex by dimer ([$F(5,120)=1.250$, $p=0.290$]) did not reach statistical significance (Fig. 3A, B, Tables 1, 2).

Interaction between main effects area by sex and area by dimer describes an increased proportion of heterodimerized Nk3R with GPER in proestrus females compared to males in all areas examined ([$F(1,120)=213.664$, $p=0.000$]), with an increased proportion of heterodimerized Nk3R with GPAR in all assessed areas in male mice compared to proestrus female mice ([$F(1,120)=10.616$, $p=0.001$]). Differences in the number of heterodimers describe increased number of heterodimers in proestrus females compared to males in the DH in CA1 ([$F(1,120)=15.100$, $p=0.000$]), CA3 ([$F(1,120)=28.687$, $p=0.000$]) and the PrL ([$F(1,120)=8.884$, $p=0.003$]) and IL ([$F(1,120)=17.899$, $p=0.000$]), but not in the CG ([$F(1,120)=0.779$, $p=0.379$]) (Fig. 3A, B, Tables 1, 2), probably because of the higher rate of heterodimerization with GPER in all areas examined.

Discussion

This study shows for the first time that the systemic blockade of the Nk3R reduces novel object recognition memory consolidation in male mice and enhances this same process in female mice that present high levels of circulating sex hormones. The Nk3R antagonist osanentan is a good candidate to study memory consolidation since it presents high blood-brain barrier penetration, a half-life of ~ 2 h [30], and a high clearance [31]. Notwithstanding, since the Nk3R is a GPCR, the effect of its blockade remains in the brain until phosphatases, kinases, and other metabolite levels

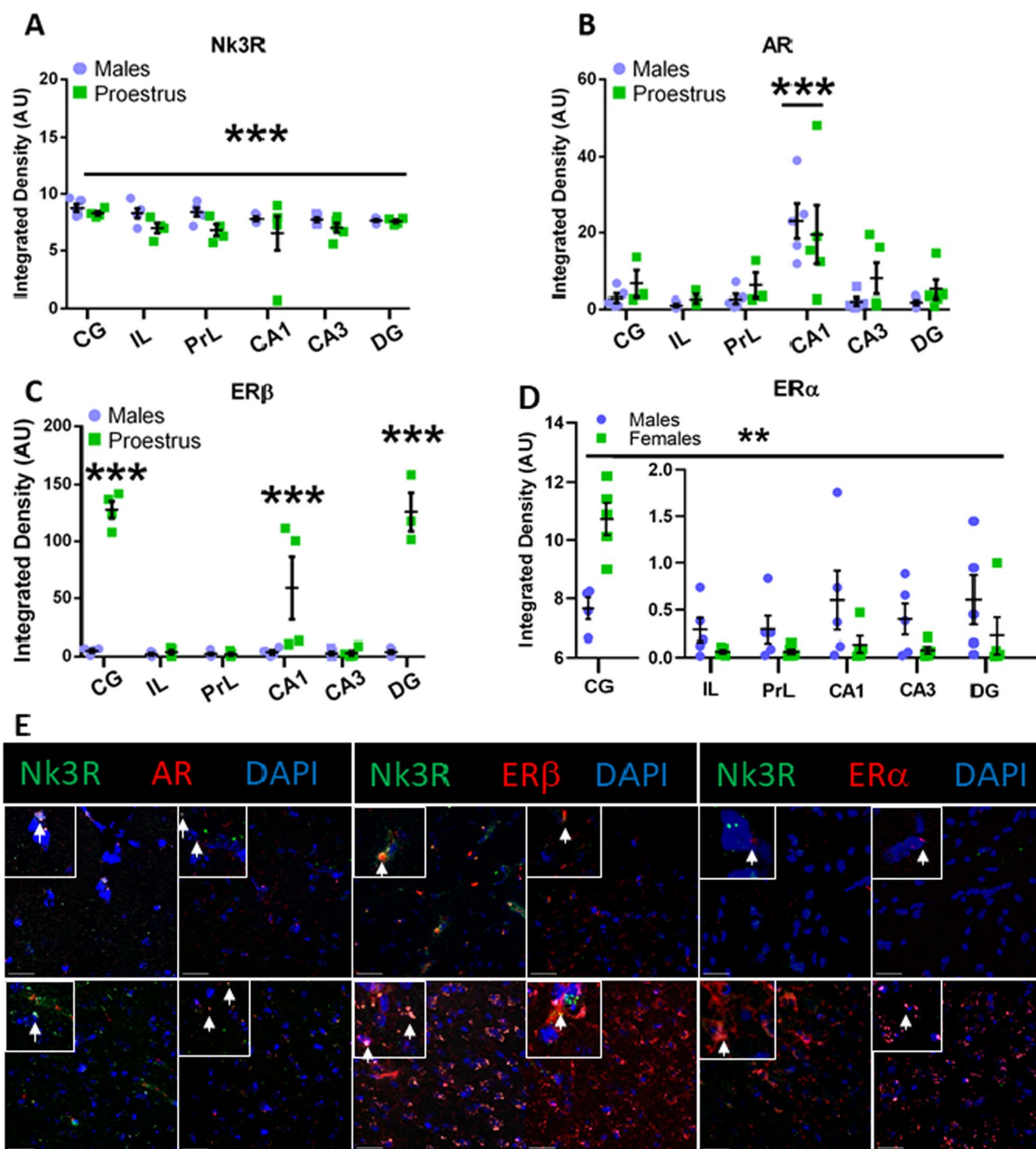


Fig. 2 Immunolocalization of neurokinin 3 receptor (Nk3R) with androgen receptor (AR), estrogen receptor α (ER α), and estrogen receptor β (ER β). **A** Immunolocalization of Nk3R in the medial prefrontal cortex (mPFC) or the dorsal hippocampus (DH). **B** Immunolocalization of AR in the mPFC or the DH. **C** Immunolocalization of ER β in the mPFC or the DH. **D** Immunolocalization of ER α in the mPFC or the DH. **D** Representative confocal images of AR, ER β , and ER α with Nk3R in the CG and CA1. CG cingulate cortex, IL infralimbic cortex, PrL prelimbic cortex, CA1 field CA1 of hippocampus, CA3 field CA3 of hippocampus, DG dentate gyrus. Data are mean \pm SEM. Yellow pixels represent colocalization between red and green. White pixels represent the colocalization of red, green, and blue. Datasets were analyzed using two-way ANOVA. ** $p < 0.01$, *** $p < 0.001$. Asterisks above a line in **A** and **D** indicate a significant main effect of sex independently of the area. Asterisks in **B** and **C** indicate significant main effect sex in discrete areas after finding a significant Sex \times Area interaction. **E** Yellow pixels indicate colocalization between Nk3R and AR, ER β , or ER α . White pixels indicate colocalization between DAPI, Nk3R, and AR, ER β , or ER α . Scale bar = 25 μ m

return to the baseline. We measured the expression of the Nk3R and sex hormone receptors in the mPFC–hippocampal circuit that underlie the formation of novel object recognition memories. We also uncovered potential new protein interaction mechanisms between the Nk3R and membrane receptors for sex hormones (GPER and GPAR), in male and female mice, that may lead to opposite-sex effects of Nk3R targeted drugs on memory consolidation (Fig. 4).

Sex differences in object recognition

Sex differences in recognition memory performance have been previously studied, although it is still controversial. While some authors report better performance for males in the NOR test [32], others report an outperformance of female mice [33]. This better performance of females has been hypothetically related to better object discrimination compared to male mice, as shown in other work [34]. In contrast, male mice have been shown to present a better discrimination index when the novel object resembles the object that they have been previously familiarized with [34]. Our study does not show a sex-dependent performance on the NOR test, nor in the preference test performed before the recognition test, suggesting that both objects used as novel and old were sufficiently different to produce a similar discrimination rate between sexes.

Nk3R opposite-sex regulation of memory consolidation

The Nk3R has been previously linked to the performance of male subjects in the object recognition test. The administration of the Nk3R agonist Senktide enhances performance in the NOR test in male Wistar rats [9]. At a molecular level, our previous studies in fear memory show that the Tac2 pathway modulations in the central amygdala (CeA) reduce the activity of the Akt/GSK3 β / β -Catenin in males and increases the activity of this molecular route in females leading to opposite-sex effects in memory [6, 7]. Our present study suggests that Nk3R antagonism may modulate in a similar manner recognition memory in both sexes with potential similarities in the mechanisms that mediate both fear and recognition memory. Increased activity of GSK3 β prevents the phosphorylation of β -Catenin and, therefore, its role in the consolidation of memories. In the same line, increased activity of GSK3 β has been shown to impair novel object recognition, as well as mood regulation and hippocampal cell proliferation

[35]. Further, activation of the Akt/GSK3 β pathway in the presence of estradiol has been shown to increase object recognition [22]. Taking all these data into account, we hypothesize a possible confluence in the mechanisms of memory consolidation after Nk3R antagonism, with the GSK3 β downstream target β -Catenin modulating the formation of long-lasting memories.

In previous works [6, 7], we demonstrated that a single dose of 5 mg/kg reduces fear memory consolidation in males and increases it in females. In contrast, this dose was not effective in modulating recognition memory consolidation, but a 10 mg/kg dose reproduced this effect in the NOR task. As commented before, the neural networks supporting consolidation for fear memories and recognition memories remain similar between humans and rodents and differ consistently. Consolidation of cued-fear memories typically requires molecular and systemic changes in the amygdala and extra-amygdalar complexes [36, 37]. In contrast, consolidation of recognition memory has been widely associated with the hippocampus and its connections with the mPFC [2, 22, 38]. Each of these brain regions presents a unique organization and information flow, which may vary its sensitivity to osanetant modulation of synaptic and systemic consolidation processes.

Previous work has demonstrated that senktide, a potent Nk3R agonist, increases memory consolidation in male rats [9]. Also, senktide reverted the scopolamine-induced memory impairment, suggesting that the muscarinic cholinergic receptors mediate the memory enhancement of senktide [39]. Senktide has also been shown to increase acetylcholine release in the mPFC and the DH [40, 41]. A tentative explanation of the observed behavioral outcome is a decrease of ACh release in males and an increase in females, after osanetant administration. Notwithstanding, NkB also interacts with the dopaminergic system regulating dopamine release in the striatum in healthy and pathological individuals [42]. We suggest that further research should evaluate ACh release in the mentioned areas as well as the behavioral outcome of site-directed manipulations of Nk3R receptors.

Sex hormones interaction with Nk3R on recognition memory consolidation

Estradiol has been shown in female mice to increase recognition memory consolidation by enhancing the activity

(See figure on next page.)

Fig. 3 Heterodimerization of neurokinin 3 receptor with membrane sex hormones receptors. **A** Quantification of heterodimerization Nk3R with G-protein androgen receptor (GPAR) and G-protein estrogen receptor (GPER) in male and female mice during proestrus. *CG* cingulate cortex, *IL* infralimbic cortex, *PrL* prelimbic cortex, *CA1* field CA1 of hippocampus, *CA3* field CA3 of hippocampus, *DG* dentate gyrus. Data are mean \pm SEM. Datasets were analyzed using two-way ANOVA. ** $p < 0.01$, *** $p < 0.001$. **B** Representative confocal images of the previously reported quantification. Scale bar = 20 μ m

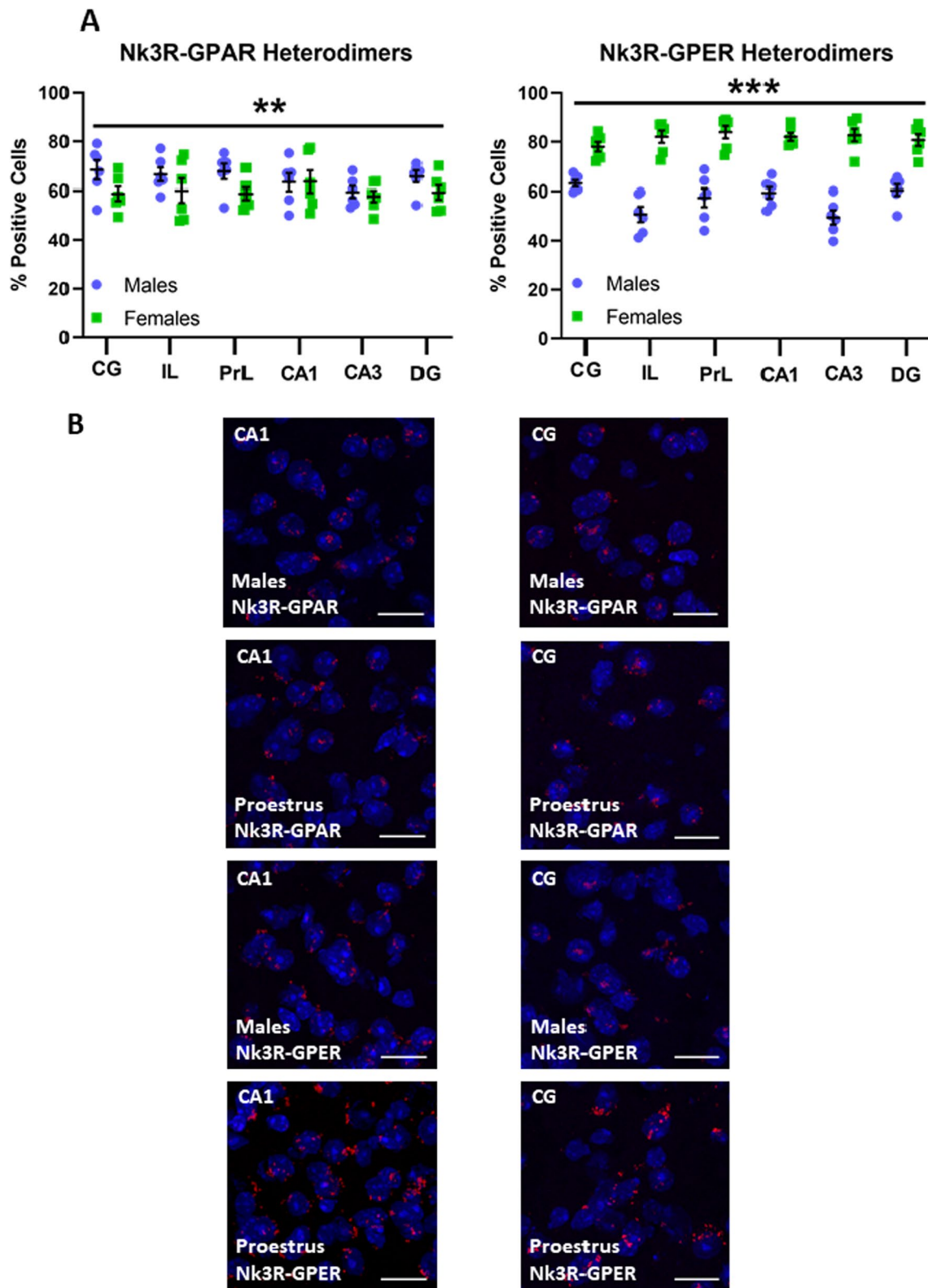
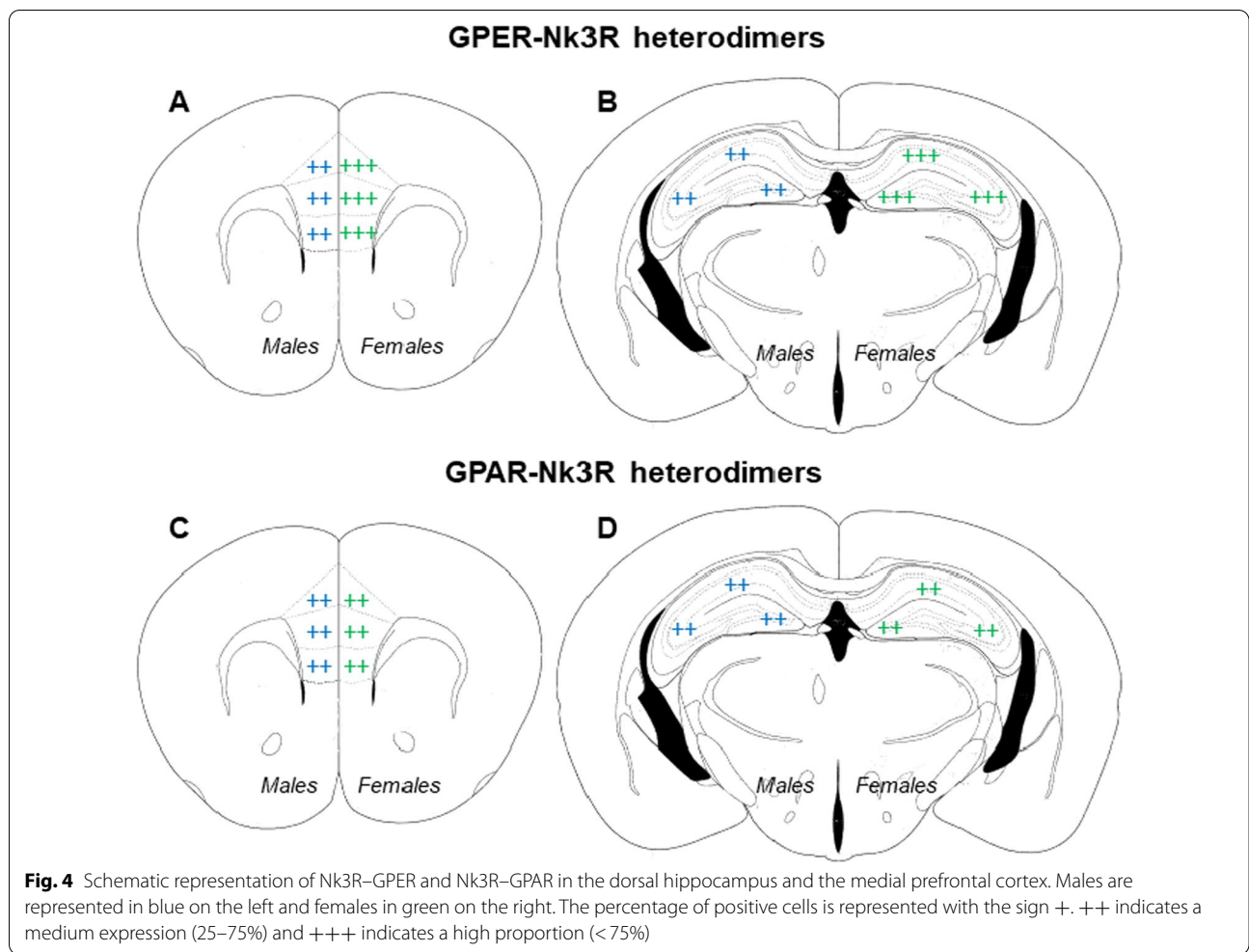


Fig. 3 (See legend on previous page.)



of the Akt/GSK3β pathway in the DH [22]. Interestingly, wild-type mice, but not ERβ-KO, increased their performance on the object recognition task when treated with estradiol [43], suggesting a role for ERβ in the modulation of the enhancement effect of estradiol in object recognition memory consolidation. Our study shows that ERβ is more expressed in the CA1 and DG of the DH and the CG of the mPFC in females compared to males, suggesting possible target areas of estradiol in the enhancement of recognition memory consolidation, especially in those that represent the direct output of the DH. Notwithstanding, ERα is also present in the DH and mPFC of female mice although its expression is lower expressed than in males. Data in female Swiss mice have also highlighted the role of hippocampal ERα in enhancing memory consolidation [44].

Our study demonstrates a high degree of heterodimerization of the Nk3R with GPER in the DH and the mPFC, finding that around 80% of Nk3R found in these areas present heterodimerization with GPER in female mice (Fig. 4A, B). With these data, we hypothesize that estradiol exerts an

Table 1 Mean values and standard deviation (mean (SD)) for Nk3R–GPAR and Nk3R–GPER heterodimers in the mPFC and the DH

| Area | GPAR | | GPER | |
|------|--------------|---------------|--------------|--------------|
| | Males | Females | Males | Females |
| CA1 | 63.43 (8.92) | 63.65 (11.09) | 59.36 (5.56) | 82.00 (4.01) |
| CA3 | 59.47 (5.80) | 57.72 (5.79) | 49.47 (7.04) | 82.73 (6.38) |
| DG | 65.73 (6.02) | 59.26 (7.09) | 60.48 (5.66) | 80.70 (5.88) |
| CG | 68.42 (9.60) | 58.77 (7.08) | 63.13 (3.21) | 77.98 (4.78) |
| PrL | 67.78 (7.72) | 58.75 (6.14) | 57.37 (9.17) | 83.94 (6.31) |
| IL | 66.57 (6.85) | 60.07 (12.02) | 50.69 (7.67) | 82.07 (6.25) |

The mean value was calculated based on the quantification of cells containing one or more heteromers versus total cells

GPAR G-protein androgen receptor, GPER G-protein estrogen receptor, CG cingulate cortex, IL infralimbic cortex, PrL prelimbic cortex, CA1 field CA1 of hippocampus, CA3 field CA3 of hippocampus, DG dentate gyrus

activating function over the GPER that allows the inhibition of the Nk3R to increase intracellular cascades responsible for an increase in memory consolidation. Although

Table 2 Statistical tests for independent sample analyses

| Between-subjects variables | Snedecor's <i>F</i> | | <i>p</i> -value |
|----------------------------|---------------------|---------|-----------------|
| | <i>df</i> | Value | |
| Preference test | | | |
| Sex | 1,24 | 0.057 | 0.813 |
| Object | 1,24 | 0.072 | 0.791 |
| Sex × Object | 1,24 | 0.072 | 0.791 |
| Familiarization (5 mg/kg) | | | |
| Sex | 1,56 | 0.000 | 1.000 |
| Object | 1,56 | 0.031 | 0.860 |
| Treatment | 1,56 | 0.000 | 1.000 |
| Sex × Object | 1,56 | 0.359 | 0.552 |
| Sex × Treatment | 1,56 | 0.000 | 1.000 |
| Object × Treatment | 1,56 | 1.128 | 0.293 |
| Sex × Object × Treatment | 1,56 | 0.006 | 0.939 |
| NOR test (5 mg/kg) | | | |
| Sex | 1,28 | 1.748 | 0.197 |
| Treatment | 1,28 | 0.227 | 0.603 |
| Sex × Treatment | 1,28 | 0.002 | 0.965 |
| Familiarization (10 mg/kg) | | | |
| Sex | 1,5 | 0.000 | 1.000 |
| Object | 1,5 | 0.000 | 1.000 |
| Treatment | 1,5 | 0.103 | 0.749 |
| Sex × Object | 1,5 | 0.091 | 0.764 |
| Sex × Treatment | 1,5 | 0.000 | 1.000 |
| Object × Treatment | 1,5 | 1.638 | 0.206 |
| Sex × Object × Treatment | 1,5 | 2.651 | 0.110 |
| NOR test (10 mg/kg) | | | |
| Sex | 1,25 | 10.595 | <i>0.003</i> |
| Treatment | 1,25 | 0.022 | 0.884 |
| Sex × Treatment | 1,25 | 17.128 | <i>0.000</i> |
| Treatment in males | 1,25 | 9.496 | <i>0.005</i> |
| Treatment in females | 1,25 | 7.714 | <i>0.010</i> |
| Nk3R expression | | | |
| Sex | 1,43 | 7.770 | 0.008 |
| Area | 5,44 | 1.631 | 0.257 |
| Sex × Area | 5,44 | 0.768 | 0.758 |
| ERα expression | | | |
| Sex | 1,47 | 7.001 | 0.011 |
| Area | 5,47 | 1.789 | 0.133 |
| Sex × Area | 5,47 | 1.326 | 0.270 |
| ERβ expression | | | |
| Sex | 1,41 | 115.275 | <i>0.000</i> |
| Area | 5,41 | 28.389 | <i>0.000</i> |
| Sex × Area | 5,41 | 26.475 | <i>0.000</i> |
| Sex in CA1 | 1,41 | 21.410 | <i>0.000</i> |
| Sex in CA3 | 1,41 | 0.001 | 0.973 |
| Sex in DG | 1,41 | 98.405 | <i>0.000</i> |
| Sex in CG | 1,41 | 105.908 | <i>0.000</i> |
| Sex in IL | 1,41 | 0.000 | 0.985 |
| Sex in PrL | 1,41 | 0.029 | 0.866 |

Table 2 (continued)

| Between-subjects variables | Snedecor's <i>F</i> | | <i>p</i> -value |
|----------------------------|-----------------------|---------|-----------------|
| | <i>df</i> | Value | |
| AR expression | | | |
| Sex | 1,42 | 1.388 | 0.245 |
| Area | 5,42 | 9.144 | <i>0.000</i> |
| Sex × Area | 5,42 | 0.475 | 0.793 |
| CG vs rest of areas | Bonferroni correction | | <i>0.000</i> |
| Nk3R–GPAR positive cells | | | |
| Sex | 1,60 | 8.402 | <i>0.005</i> |
| Area | 5,60 | 0.688 | 0.635 |
| Sex × Area | 5,60 | 0.717 | 0.613 |
| Nk3R–GPER positive cells | | | |
| Sex | 1,60 | 290.103 | <i>0.000</i> |
| Area | 5,60 | 1.609 | 0.171 |
| Sex × Area | 5,60 | 3.810 | <i>0.005</i> |
| Sex in CA1 | 1,10 | 0.001 | 0.972 |
| Sex in CA3 | 1,10 | 0.274 | 0.612 |
| Sex in DG | 1,10 | 2.906 | 0.119 |
| Sex in CG | 1,10 | 3.929 | 0.076 |
| Sex in IL | 1,10 | 3.929 | 0.076 |
| Sex in PrL | 1,10 | 5.033 | <i>0.049</i> |

Two or three-way ANOVAs were performed to assess statistical significance for independent sample analyses. Significant main effects and interactions between main effects are indicated in *italic*. Bonferroni post hoc analysis was used to assess significance between categories of a significant multiple-category main effect

not at a similar level, approximately 60% of Nk3R in the DH and the mPFC presents heterodimerization with membrane GPAR (Fig. 4C, D). Previous experiments have demonstrated that the memory consolidation effects observed after the administration of osanentan are dependent on the sexual maturity of the mouse [7], either male or female. Interestingly, given the involvement of sex hormones in the regulation of Nk3R activity on memory formation [7], estrogen GPCR has recently been shown to modulate hippocampal-dependent memory and long-term potentiation [45]. Although the Nk3R presents a significant number of heterodimers with AR in males and proestrus females, the Nk3R–GPER heterodimer appears quite more abundant than the rest, pointing to a possible explanation for the increased efficacy of Nk3R during high estradiol levels. A tentative hypothesis for this phenomenon is that the high degree of heterodimerization of Nk3R with sex hormones receptors, with new and unique pharmacological and functional properties different from those of its components [24, 46, 47], would produce a sex-specific activation of the heterodimer in the presence of sex hormones, allowing the antagonism to modulate memory consolidation in a sex-opposite way. It would be interesting for future studies to follow up our PLA analyses with co-immunoprecipitation

to further understand protein–protein interactions of the Nk3R, ER α , ER β , and AR. Additionally, an important question that could be further addressed is whether this results in heterodimerization of receptors that might interact with drug treatments, experience, or during the normal hormonal cycling or life-span in male and female mice. Further, we would like to highlight that our treatments take place 30 min after the familiarization phase of the NOR task, therefore, avoiding a possible effect of the drug in the test phase. Notwithstanding, we believe that the consolidation effect observed is affecting the storage of memories and therefore, retrieval. Further, we would also highlight that during the retrieval phase osanetant is no longer present in the animal's body, so we could isolate the effect of the drug in the consolidation phase. Notwithstanding, it will be quite interesting to manipulate Tac2/Nk3R before or during retrieval to have a more complete knowledge of the phenomenon explained here.

Perspectives and significance

This study demonstrates for the first time a sex-opposite effect of an Nk3R antagonist on recognition memory consolidation, together with a high degree of heterodimerization between Nk3R and membrane sex hormone receptors. Osanetant is a safe and well-tolerated drug in humans [10], an important finding that may lead to a rapid translation of these results into the clinic. Osanetant has not been approved yet for the treatment of mental disorders in humans, but there are clinical trials currently studying this possibility.

Previous research has shown that Nk3R is expressed in Tac2 neurons in the centromedial amygdala (CeM) [4]. As previously demonstrated [7], these neurons in the CeM are GABAergic neurons, concordantly with the anatomical data available on the distribution of GABA and glutamate in the amygdala. Notwithstanding, few articles address the colocalization of Tac2 with other neuropeptidergic systems. One example of this colocalization is the Tac2-Kisspeptin neurons in the hypothalamus [48]. Following this, Kisspeptin is not one of the representative neuropeptides found in the CeM, suggesting that the colocalization with other neuropeptidergic systems may vary depending on the area. The specific expression of Tac2 and its colocalization in the medial prefrontal cortex and the dorsal hippocampus has not been addressed in this article and could be of interest for future research.

Interestingly, Nk3R pharmacological manipulations have aroused as a promising new tool for memory deficits through its interaction with the cholinergic system [40, 41]. Moreover, interactions of the Tac2 pathway and GABA in the extended amygdala modulate the activity of the lateral hypothalamic area [47]. All these findings highlight exciting possible novel pharmacological targets for

the treatment of anterograde amnesic affection in women, suggesting a path for future research on personalized medicine that tests the possibilities of Nk3R antagonists in the treatment of memory loss in women. Moreover, the impaired memory consolidation of osanetant in males in the consolidation of both fear memory and episodic memory may be a tool for the treatment of aversive events with an emotional and episodic memory component.

Acknowledgements

Procedural images were created under license with BioRender.com.

Author contributions

This study was planned and conceptualized by RA and AF. AF performed behavioral and immunolocalization experiments, vaginal cytologies, and datasets analyses. EM and EIC performed In Situ Proximal Ligand Assays. The paper was written by RA and AF and commented on and discussed by all authors. All authors read and approved the final manuscript.

Funding

R.A. was supported by a NARSAD Young Investigator Grant #22434, Ramón y Cajal program (Agencia Estatal de Investigación) RYC2014-15784, RETOS-MICINN SAF2016-76565-R and PID2020-112705RB-I00 FEDER funds, Fundación Alicia Koplowitz, ERANET Neuron JTC 2019 (Ministerio de Ciencia, Innovación y Universidades) ISCIII AC19/00077, and CIBER—Consorcio Centro de Investigación Biomédica en Red—(CB19/09/00029); and SAF2017-87629-R to E.I.C. A.F. was the recipient of Generalitat de Catalunya (Agència de Gestió d'Ajuts Universitaris i de Recerca) 2018 FI_B00030 predoctoral fellowship.

Availability of data and materials

All data and materials are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

As indicated in "Methods" section, ethics protocols were approved for the experiments in mice with ref. CEEAH 3603. All procedures were approved by the Ethics Committee of the Universitat Autònoma de Barcelona and the Generalitat de Catalunya. Experiments were also carried out following the European Communities Council Directive (2010-63-UE) and Spanish legislation (RD 53/2013).

Consent for publication

Not applicable.

Competing interests

R.A. declares a potential conflict of interest with the patent PCT/US2015/037629. The other authors declare no competing interests.

Author details

¹Institut de Neurociències, Universitat Autònoma de Barcelona, Cerdanyola del Vallès, 08193 Barcelona, Spain. ²Departament de Psicobiologia i Metodologia de les Ciències de la Salut, Universitat Autònoma de Barcelona, Cerdanyola del Vallès, 08193 Barcelona, Spain. ³Departament de Bioquímica i Biomedicina Molecular, Facultat de Biologia, Universitat de Barcelona i Institut de Biomedicina de la Universitat de Barcelona (IBUB), 08028 Barcelona, Spain. ⁴Instituto de Salud Carlos III, 28029 Madrid, Spain. ⁵Centro de Investigación Biomédica en Red en Salud Mental (CIBERSAM), Instituto de Salud Carlos III, 28029 Madrid, Spain. ⁶Unitat de Neurociència Translacional, Parc Taulí Hospital Universitari, Institut d'Investigació i Innovació Parc Taulí (I3PT), Universitat Autònoma de Barcelona, Cerdanyola del Vallès, 08193 Barcelona, Spain. ⁷ICREA, Pg. Lluís Companys 23, 08010 Barcelona, Spain.

Received: 20 January 2022 Accepted: 24 May 2022

Published online: 11 June 2022

References

- Davis S, Renaudineau S, Poirier R, Poucet B, Save E, Laroche S. The formation and stability of recognition memory: what happens upon recall? *Front Behav Neurosci*. 2010;4:177.
- Squire LR, Genzel L, Wixted JT, Morris RG. Memory consolidation. *Cold Spring Harbor Perspect Biol*. 2015;7:a021766.
- Andero R, Dias BG, Ressler KJ. A role for Tac2, NkB, and Nk3 receptor in normal and dysregulated fear memory consolidation. *Neuron*. 2014;83:444–54.
- Andero R, Daniel S, Guo JD, Bruner RC, Seth S, Marvar PJ, et al. Amygdala-dependent molecular mechanisms of the Tac2 pathway in fear learning. *Neuropsychopharmacology*. 2016;41:2714–22.
- Eichenbaum H. The hippocampus and mechanisms of declarative memory. *Behav Brain Res*. 1999;103:123–33.
- Andero R, Dias BG, Ressler KJ. A role for Tac2, NkB, and Nk3 receptor in normal and dysregulated fear memory consolidation. *Neuron Cell Press*. 2014;83:444–54.
- Florido A, Velasco ER, Soto-Faguás CM, Gomez-Gomez Á, Perez-Caballero L, Molina P, et al. Sex differences in fear memory consolidation via Tac2 signaling in mice. *Nat Commun*. 2021;12:2496.
- Zlomuzica A, Dere E, Huston JP, de Souza Silva MA. NK(3) receptor agonism promotes episodic-like memory in mice. *Neurobiol Learn Mem*. 2008;90:420–5.
- Chao OY, Nikolaus S, Huston JP, de Souza Silva MA. The neurokinin-3 receptor agonist senktide facilitates the integration of memories for object, place and temporal order into episodic memory. *Neurobiol Learn Mem*. 2014;114:178–85.
- Malherbe P, Ballard TM, Ratni H. Tachykinin neurokinin 3 receptor antagonists: a patent review (2005–2010). *Expert Opin Ther Pat*. 2011;21:637–55.
- Meltzer HY, Arvanitis L, Bauer D, Rein W. Placebo-controlled evaluation of four novel compounds for the treatment of schizophrenia and schizoaffective disorder. *Am J Psychia*. 2004;161(6):975–84. <https://doi.org/10.1176/appi.ajp.161.6.975>.
- Modi M, Dhillon WS. Neurokinin 3 receptor antagonism: a novel treatment for menopausal hot flashes. *Neuroendocrinology*. 2019;109:242–8.
- Farovik A, Place RJ, Miller DR, Eichenbaum H. Amygdala lesions selectively impair familiarity in recognition memory. *Nat Neurosci*. 2011;14:1416–7.
- Aggleton JP, Brown MW. Interleaving brain systems for episodic and recognition memory. *Trends Cogn Sci*. 2006;10:455–63.
- Morici JF, Bekinschtein P, Weisstaub NV. Medial prefrontal cortex role in recognition memory in rodents. *Behav Brain Res*. 2015;1(292):241–51.
- Clark RE, Zola SM, Squire LR, Janssen WG, Hayashi S, Milner TA, et al. Impaired recognition memory in rats after damage to the hippocampus. *J Neurosci*. 2010;20:8853–60.
- Lee I, Kesner RP. Time-dependent relationship between the dorsal hippocampus and the prefrontal cortex in spatial memory. *J Neurosci*. 2003;23:1517.
- Jin J, Maren S. Prefrontal-hippocampal interactions in memory and emotion. *Front Syst Neurosci*. 2015;9:170.
- Warburton EC, Brown MW. Findings from animals concerning when interactions between perirhinal cortex, hippocampus and medial prefrontal cortex are necessary for recognition memory. *Neuropsychologia*. 2010;48:2262–72.
- Kesner RP, Hunt ME, Williams JM, Long JM. Prefrontal cortex and working memory for spatial response, spatial location, and visual object information in the rat. *Cereb Cortex*. 1996;6:311–8.
- Ragozzino ME, Detrick S, Kesner RP. The effects of prelimbic and infralimbic lesions on working memory for visual objects in rats. *Neurobiol Learn Mem*. 2002;77:29–43.
- Fortress AM, Lu F, Orr PT, Zhao Z, Frick KM. Estradiol-induced object recognition memory consolidation is dependent on activation of mTOR signaling in the dorsal hippocampus. *Learn Mem*. 2013;20:147–55.
- Schulz K, Korz V. Hippocampal testosterone relates to reference memory performance and synaptic plasticity in male rats. *Front Behav Neurosci*. 2010;4:187.
- Ferré S, Casadó V, Devi LA, Filizola M, Jockers R, Lohse MJ, Milligan G, Pin JP, Guitart X. G protein-coupled receptor oligomerization revisited: functional and pharmacological perspectives. *Pharmacol Rev*. 2014;66:413–34.
- Khawaja AM, Rogers DF. Tachykinins: receptor to effector. *Int J Biochem Cell Biol*. 1996;28:721–38.
- Fraser GL, Ramael S, Hoveyda HR, Gheyle L, Combalbert J. The NK3 receptor antagonist ESN364 suppresses sex hormones in men and women. *J Clin Endocrinol Metab*. 2016;101:417–26.
- Florido A, Perez-Caballero L, Velasco ER, Molina P, Marin-Blasco I, Andero R. Direct and indirect measurements of sex hormones in rodents during fear conditioning. *Curr Protoc*. 2021;1:e102.
- Rossato JI, Radiske A, Kohler CA, Gonzalez C, Bevilacqua LR, Medina JH, et al. Rapid Communication Consolidation of object recognition memory requires simultaneous activation of dopamine D1/D5 receptors in the amygdala and medial prefrontal cortex but not in the hippocampus. *Neurobiol Learn Mem*. 2013;106:66–70.
- DeVito LM, Eichenbaum H. Distinct contributions of the hippocampus and medial prefrontal cortex to the “what-where-when” components of episodic-like memory in mice. *Behav Brain Res*. 2010;215:318.
- Chen MH, Chung FZ, Roth BD, Kuo BS, Atherton J, Lee HT. Syntheses and biological activities of chiral piperidines-tachykinin NK3 antagonists. *Zhongguo Yao Li Xue Bao*. 1999;20(3):283–8.
- Quartara L, Altamura M, Evangelista S, Maggi CA. Tachykinin receptor antagonists in clinical trials. *Expert Opin Investig Drugs*. 2009;18:1843–64.
- Frick KM, Gresack JM. Sex differences in the behavioral response to spatial and object novelty in adult C57BL/6 mice. *Behav Neurosci*. 2003;117:1283–91.
- Benice TS, Rizk A, Kohama S, Pfankuch T, Raber J. Sex-differences in age-related cognitive decline in C57BL/6J mice associated with increased brain microtubule-associated protein 2 and synaptophysin immunoreactivity. *Neuroscience*. 2006;137:413–23.
- Bettis T, Jacobs LF. Sex differences in object recognition are modulated by object similarity. *Behav Brain Res*. 2012;233:288–92.
- Pardo M, Abrial E, Jope RS, Beurel E. GSK3 β isoform-selective regulation of depression, memory and hippocampal cell proliferation. *Genes Brain Behav*. 2016;15:348–55.
- Maguschak KA, Ressler KJ. β -catenin is required for memory consolidation. *Nat Neurosci*. 2008;11:1319–26.
- Schafe GE, Atkins CM, Swank MW, Bauer EP, Sweatt JD, Ledoux JE. Activation of ERK/MAP kinase in the amygdala is required for memory consolidation of pavlovian fear conditioning. *J Neurosci*. 2000;20:8177–87.
- Sutherland RJ, O'Brien J, Lehmann H. Absence of systems consolidation of fear memories after dorsal, ventral, or complete hippocampal damage. *Hippocampus*. 2008;18:710–8.
- Schäble S, Huston JP, Barros M, Tomaz C, de Souza Silva MA. The NK3 receptor agonist senktide ameliorates scopolamine-induced deficits in memory for object, place and temporal order. *Neurobiol Learn Mem*. 2012;97:235–40.
- de Souza Silva MA, Lenz B, Rotter A, Biermann T, Peters O, Ramirez A, et al. Neurokinin3 receptor as a target to predict and improve learning and memory in the aged organism. *Proc Natl Acad Sci USA*. 2013;110:15097–102.
- Schäble S, Topic B, Buddenberg T, Petri D, Huston JP, de Souza Silva MA. Neurokinin3-R agonism in aged rats has anxiolytic-, antidepressant-, and promnesic-like effects and stimulates ACh release in frontal cortex, amygdala and hippocampus. *Eur Neuropsychopharmacol*. 2011;21:484–94.
- Zhang X, Andren PE, Chergui K, Svenningsson P. Neurokinin B/NK3 receptors exert feedback inhibition on L-DOPA actions in the 6-OHDA lesion rat model of Parkinson's disease. *Neuropharmacology*. 2008;54:1143–52.
- Walf AA, Koonce C, Frye CA. Estradiol or diarylpropionitrile administration to wildtype, but not estrogen receptor beta knockout, mice enhances performance in the object recognition and object placement tasks. *Neurobiol Learn Mem*. 2008;89:513.
- Pereira LM, Bastos CP, de Souza JM, Ribeiro FM, Pereira GS. Estradiol enhances object recognition memory in Swiss female mice by activating hippocampal estrogen receptor α . *Neurobiol Learn Mem*. 2014;114:1–9.
- Kumar A, Foster TC. G protein-coupled estrogen receptor: rapid effects on hippocampal-dependent spatial memory and synaptic plasticity. *Front Endocrinol*. 2020;11:385.
- Ferré S, Baler R, Bouvier M, Caron MG, Devi LA, Durrour T, et al. Building a new conceptual framework for receptor heteromers. *Nat Chem Biol*. 2009;5:131–4.
- Gomes I, Ayoub MA, Fujita W, Jaeger WC, Pflieger KDG, Devi LA. G protein-coupled receptor heteromers. *Annu Rev*. 2016;56:403–25.

48. Goodman RL, Lehman MN, Smith JT, Coolen LM, de Oliveira CVR, Jafarzadehshirazi MR, et al. Kisspeptin neurons in the arcuate nucleus of the ewe express both dynorphin A and neurokinin B. *Endocrinology*. 2007;148:5752–60.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.