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Ecological interactions disrupted by habitat alteration in the Neotropics

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Ecological interactions disrupted by habitat alteration in the Neotropics

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Abstract: Ecological interactions help determine the distribution of species across landscapes and play crucial roles in ecosystem services such as pollination, seed dispersal, and pest control¹. Human disturbances, particularly habitat alteration, have the potential to modify or erase ecological interactions^{2,3} and so jeopardise the processes they control. While examples of interactions becoming rewired under human influence have been recorded, studies of this process for speciose assemblages at regional to continental scales are uncommon⁴ and obstructed by logistical difficulties². The consequences for ecological communities and people are therefore poorly understood. Here we show that human habitat alteration is associated with a decrease in the spatial aggregation of Neotropical bat pairs and bird pairs that share similar dietary requirements. We find that groups of species pairs with similar vs. different diets have positive spatial associations on average, but pairs within dietary guilds have stronger associations than pairs with disparate diets when habitats are relatively intact. Our results suggest that species with similar resource requirements typically coexist in relatively intact natural settings. By contrast, exclusion becomes more common (though not dominant) when habitats are altered. Altered habitats thus fail to support the coexistence of diverse competitive interactions, reversing patterns observed in the wild.

Main text:

Biological interactions are essential for maintaining biodiversity because they directly affect which species can coexist. Interactions are one of the four main drivers of species distributions⁵ and key moderators of ecosystem functioning¹. Although this interpretation is still contentious^{5–7}, it is increasingly recognised that biotic interactions across assemblages can scale up to influence community assembly at much larger spatiotemporal scales than that of individual encounters^{8–10}. For example, competition and facilitation can influence the modern distribution of species at regional scales¹¹, and at continental scales over millennia¹², and drive the diversification and extinction of entire clades over millions of years^{10,13,14}.

Paradoxically, despite agreement that interactions affect species distributions on either small or very large spatiotemporal scales, the idea that the footprints of biological interactions are broadly discernible from modern occurrence data at regional or continental spatial scales is famously contentious^{11,15–21}. The debate is largely driven by the difficulty of distinguishing the effect of interactions from those of other landscape variables such as biogeographic historical effects, dispersal limitations, and habitat preferences¹¹. Furthermore, the relatively low occupancies of many species mean that there is insufficient occupancy data to provide biological information on the vast majority of possible pairwise interactions, leaving scientists to focus mainly on a relatively small number of common species.

These questions are also contentious partly because human activities may alter community assembly processes. For example, humans have been implicated in homogenising communities²² even when species richness is stable^{23,24}, and reducing species co-occurrences²⁵. Complicating matters, rapid change can rewire biological interactions²⁶ without necessarily adding or removing species²⁷. Evidence that human disturbance can decouple predator-prey interactions²⁸, increase

competition for pollinators²⁹, and simplify or break apart interaction networks^{3,30} has been gathered mainly at local scales on systems with only a few species due to logistical challenges. Patterns documenting changing interactions in entire assemblages are also rare, except for laboriously compiled interaction data spanning decades, such as plant-pollinator networks². Nonetheless, these are beset by sampling inconsistencies². Thus, there is an urgent need to fill this gap in our understanding of how interactions scale up and how human disturbance might alter such community assembly processes.

Here, we explore how one interaction type (food competition) measurably influences species cooccurrences in Neotropical birds and bats. We further demonstrate how human habitat alteration changes the effects of food competition on the spatial associations of species pairs, meaning the degree to which pairs co-occur at the same sites more or less often than expected based a noncompeting set of pairs that serves as a control. We analyse assemblages of Neotropical bats and understory birds downloaded from the Ecological Register³¹. The dataset comprises 2,396 occurrences of bats and 4,371 occurrences of birds at 132 and 91 sites, respectively. The sites in the Register are classified into two levels of degradation (altered and intact habitats: Fig. S1). We begin by running a suite of classical community analyses to illustrate the differences detectable in altered and intact faunas based on traditional approaches. Next, we introduce a statistical model that estimates an underlying association parameter which describes the spatial patterns in groups of species pairs. Our analysis demonstrates that habitat degradation influences spatial association patterns among competitors by altering – or even reversing – the average outcome of food competition at regional to continental scales.

Classical community analyses conducted using several metrics produce inconsistent results with respect to whether community patterns differ between altered and intact sites. Specifically, bird

assemblages experience significant differences in species composition without changes in richness (Extended Data Table 1) or beta diversity (Extended Data Table 2). Bats have no significant disparity in beta diversity, but possible differences in richness (Extended Data Table 1) and composition, depending on the analysis and metric used (Extended Data Table 2). A multidimensional scaling analysis demonstrates that compositional dissimilarities between altered and intact habitats are not visually striking (Fig. 1). In sum, classical analyses detect possible differences in patterns between habitat types, but they are not necessarily robust.

To characterise the effect of food competition on community assembly, we consider every pairwise combination of species within each taxon (46952 bat pairs and 1290588 bird pairs). Dietary data are easily obtained for these two groups and widely used in large-scale studies. Each pair is classified as potentially competing (henceforth: competing) or not competing (henceforth: control) based on basic dietary guild data (Fig. 2). Competing pairs share dietary guilds while control pairs do not share any dietary sources. We excluded pairs with partially overlapping diets (5.3% of bat and 28% of bird pairs) to produce the strongest possible test of our hypothesis that dietary relatedness should affect spatial association.

The intersection of diet and habitat type produces four experimental groups: competing pairs in intact sites, non-competing pairs in intact sites, competing pairs in altered sites, and noncompeting pairs in altered sites. We fit a hierarchical Bayesian model to estimate averages by group (g) for a spatial association parameter ($\bar{\theta_g}$, see Methods). A negative value of θ indicates spatial segregation or repulsion between members of species pairs, whereas a value of approximately 0 indicates independent patterns of occupancy for the pair and a positive value indicates spatial aggregation or attraction. We estimate $\bar{\theta_g}$ after first pooling the species pairs within each group into occupancy sets on the basis of shared occupancy values (e.g. all pairs with an occupancy pattern of {1,1} in one set, {1,2} in another set and so forth, see Fig. 2). This approach, which we term "occupancy-set analysis", (see Methods), confers several advantages over classical "co-occurrence analysis", which entails seperately assessing the spatial association for each species pair, or else calculating a single value for a complete matrix³². Model results for competing pairs are compared to control pairs in each habitat for each taxon using 95% highest density intervals on 1,200 posterior samples (Fig 3, Extended Data Table 3).

Though sharing diet guilds clearly does not guarantee that a species pair competes for food, this design ensures that — barring errors in guild assignment — all pairs that do compete for food are analysed in the same group. The inclusion of potentially many pairs that do not truly compete in this group reduces the power of the analysis and thus should produce conservative estimates of differences between groups. Because each species occurs in multiple pairs, and hence occupancy sets, the pairwise approach results in non-independence. Consequently, the significance of our results cannot be formally evaluated. Based on these two considerations, it is difficult to say whether the underlying effect is large in absolute terms. However, we report striking qualitative differences between independent habitat types that lend weight our analysis.

Results and Discussion

Overall, birds and bats both tend to aggregate ($\theta > 0$; Figs 3-4). This observation contradicts well-studied differences in the ecology of birds and bats. Bats exhibit limiting morphological similarity³³, share roosts³⁴, display varied foraging behaviours^{34,35}, and micro-partition resources, habitats, and foraging times³⁶, thus coexisting locally. By contrast, Neotropical bird species are well known for interspecific aggression³⁷, separating out along elevational gradients³⁷, and competitive exclusion³⁸. Despite these differences, the average pairwise associations among species are generally positive for both taxa at the spatial scale of our analysis. The large spatial extent of this study means that biogeography likely plays a role in this outcome. There is compositional turnover in both taxa between Mexico and Argentina^{39,40}, but nestedness of the community structure may also play a role.

Competition for food affects the spatial community structure of both taxa, as demonstrated by offsets in the posterior distributions for θ_g of competing and control groups (Fig. 3, Table S2). In intact habitats, we find no compelling evidence for assemblage-wide incidence of diet-based competitive exclusion (i.e., competing pairs co-occurring less than controls) in either taxon. The posterior distributions for θ_g have higher modes for competing than control pairs, particularly among bats (Fig. 3), suggesting that spatial aggregation attributable to shared habitat preferences is of greater importance than any processes of competitive exclusion in structuring these communities. The variance of the θ estimates is also higher for competing pairs than control pairs that mutual food sources encourage resource sharing and partitioning in the wild.

In both taxa, this pattern changes in altered habitats. Specifically, the offset in $\bar{\theta_g}$ between competing and control pairs disappears in altered habitats. For birds, there is some evidence that $\bar{\theta_g}$ is lower for competing pairs than control pairs (Fig. 3). While aggregation may still occur in many pairs, exclusion is much more common for competing pairs under altered than intact conditions in both taxa. There is also a wholesale decrease in $\bar{\theta_g}$ in altered habitats for birds, so natural coexistence is less common in altered habitats, irrespective of overlap in resource requirements.

To discover whether compositional differences between habitats are responsible for our results, we re-run our models while excluding species that are not sampled in both altered and intact habitats ("no-turnover" models). There is no qualitative change in the results for either bats or birds (Fig. 3). Taken together, these findings indicate that spatial associations among species differ between habitat types, and that this is not simply due to compositional differences between habitat types (e.g., some species occurring only in intact habitats or vice versa).

Conclusion

We find positive average associations among bat and bird species in intact habitats. However, for both taxa, there was greater coexistence for competing pairs than expected, suggesting that resource partitioning occurs more frequently than competitive exclusion when species pairs share resource requirements. This pattern changes in altered habitats. Altered habitats afford fewer opportunities for the coexistence of birds more generally. Competing pairs, however, exhibit equal or lower coexistence than control pairs under altered conditions, suggesting that competitive exclusion is more common. Thus, altered habitats are unable to consistently support varied local communities of competing species through niche and resource partitioning to the same level that intact habitats do. The no-turnover analyses indicate that these effects are largely attributable to changes in species spatial distributions with respect to their interaction partners, and not merely to differences in species composition.

Our results bring new evidence to the hotly debated effect of resource overlap on bird cooccurrence. A long history of studies argues that competitive exclusion detectable by distribution patterns alone^{15–17} is a common phenomenon in bird pairs that share dietary guilds⁴¹ or are closely related¹⁶. The opposing viewpoint holds that bird pairs have statistically weak spatial associations which cannot be used to infer the effects of competition^{18–20} or at best, associations with mundane geographical explanations, rather than biological ones²¹. However, all of these studies used classic approaches to co-occurrence analysis that consider each species pair in isolation, leaving inadequate statistical power to assess spatial associations among relatively rare taxa, which generally comprise the vast majority of species. Factors leading to positive spatial associations are seldom discussed at any length and are often chalked up to mere similarity of habitat preferences or matching geographical origins of these pairs²¹. However, Gilpin and Diamond¹⁶ mentioned that myriad similar bird species are seen to co-occur due to offsets in space utilization, specific diets, or foraging strategies. Our results supply evidence that this is more common in species with resource overlap than those without, hinting that competition may have driven the evolution of these relationships¹⁶. Moreover, our analysis shows that the effects of resource overlap can indeed be detected by using occurrence data alone when using a proper control group as a benchmark for comparison, and further suggest that similar mechanisms may operate for a functionally similar taxon (bats).

Overall, habitat alteration reduces and may even reverse the effect of competition on community structure in both bats and birds. Our results add to mounting evidence that human activity changes community structure and interactions without necessarily adding or removing species, and further corroborates that this can play out over landscape scales. Though diversity loss is known to influence ecosystem services⁴, analogous studies examining the relationships between ecological interactions and functionality of ecosystems are rare (but see e.g. Ref.⁴²). Consequences of shifting competition outcomes are therefore poorly understood, but detrimental scenarios are easy to envision. For example, population control of a destructive pest by a competing species cannot take place if the two species are unable to coexist. Our results shed light on the relationship between ecological interactions and regional community assembly, and our approach helps pave the way for future research seeking to understand how interaction outcomes change across various contexts and at large spatial scales.

Main Text References:

 Cardinale, B. J. *et al.* Biodiversity loss and its impact on humanity. *Nature 2012* 486:7401 486, 59–67 (2012).

2. Doré, M., Fontaine, C. & Thébault, E. Relative effects of anthropogenic pressures, climate, and sampling design on the structure of pollination networks at the global scale. *Global Change Biology* **27**, 1266–1280 (2021).

3. de Assis Bomfim, J., Guimarães, P. R., Peres, C. A., Carvalho, G. & Cazetta, E. Local extinctions of obligate frugivores and patch size reduction disrupt the structure of seed dispersal networks. *Ecography* **41**, 1899–1909 (2018).

4. Winfree, R. *et al.* Species turnover promotes the importance of bee diversity for crop pollination at regional scales. *Science* **359**, 791–793 (2018).

5. McGill, B. J. Matters of scale. *Science* **328**, 575–576 (2010).

Araújo, M. B. & Rozenfeld, A. The geographic scaling of biotic interactions. *Ecography* 37, 406–415 (2013).

7. Ezard, T. H. G. & Purvis, A. Environmental changes define ecological limits to species richness and reveal the mode of macroevolutionary competition. *Ecology Letters* **19**, 899–906 (2016).

8. Araújo, M. B. & Luoto, M. The importance of biotic interactions for modelling species distributions under climate change. *Global Ecology and Biogeography* **16**, 743–753 (2007).

9. Pollock, L. J. *et al.* Understanding co-occurrence by modelling species simultaneously with a Joint Species Distribution Model (JSDM). *Methods in Ecology and Evolution* **5**, 397–406 (2014).

10. Fraser, D. *et al.* Investigating Biotic Interactions in Deep Time. *Trends in Ecology & Evolution* **17**, (2020).

11. Gotelli, N. J., Graves, G. R. & Rahbek, C. Macroecological signals of species interactions in the Danish avifauna. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 5030–5035 (2010).

12. Tóth, A. B. *et al.* Reorganization of surviving mammal communities after the end-Pleistocene megafaunal extinction. *Science* **365**, 1305–1308 (2019).

13. Sepkoski, J. J. A kinetic model of Phanerozoic taxonomic diversity. III. Post-Paleozoic families and mass extinctions. *Paleobiology* **10**, 246–267 (1984).

14. Alroy, J. The shifting balance of diversity among major marine animal groups. *Science*329, 1191–1194 (2010).

 Diamond, J. M. Assembly of Species Communities. in *Ecology and evolution of communities* (eds. Cody, M. L. & Diamond, J. M.) 545 (Belknap Press of Harvard University Press, 1975).

16. Gilpin, M. E. & Diamond, J. M. Factors contributing to non-randomness in species cooccurrences on islands. *Oecologia* **52**, 75–84 (1982).

17. Sanderson, J. G., Diamond, J. M. & Pimm, S. L. Pairwise co-existence of Bismarck and Solomon landbird species. *Evolutionary Ecology Research* **11**, 771–786 (2009).

 Connor, E. F. & Simberloff, D. The Assembly of Species Communities : Chance or Competition? *Ecology* 60, 1132–1140 (1979).

19. Gotelli, N. J. & McCabe, D. Species co-occurrence: A meta-analysis of JM Diamond's assembly rules model. *Ecology* **83**, 2091–2096 (2002).

20. Sfenthourakis, S., Tzanatos, E. & Giokas, S. Species co-occurrence: the case of congeneric species and a causal approach to patterns of species association. *Global Ecology and Biogeography* **15**, 39–49 (2005).

21. Connor, E. F., Collins, M. D. & Simberloff, D. The checkered history of checkerboard distributions. *Ecology* **94**, 2403–2414 (2013).

22. Fraser, D. *et al.* Late quaternary biotic homogenization of North American mammalian faunas. *Nature Communications* **13**, 1–11 (2022).

23. Tóth, A. B., Lyons, S. K. & Behrensmeyer, A. K. A Century of Change in Kenya's Mammal Communities: Increased Richness and Decreased Uniqueness in Six Protected Areas. *PLoS ONE* **9**, (2014).

24. Dornelas, M. *et al.* Assemblage time series reveal biodiversity change but not systematic loss. *Science* **344**, 296–299 (2014).

25. Lyons, S. K. *et al.* Holocene shifts in the assembly of plant and animal communities implicate human impacts. *Nature* **529**, 80–83 (2016).

Bartley, T. J. *et al.* Food web rewiring in a changing world. *Nature Ecology & Evolution*3, 345–354 (2019).

27. Tylianakis, J. M. & Morris, R. J. Ecological Networks Across Environmental Gradients. *Annual Review of Ecology, Evolution, and Systematics* **48**, 25–48 (2017).

28. Rodewald, A. D., Kearns, L. J. & Shustack, D. P. Anthropogenic resource subsidies decouple predator-prey relationships. *Ecological Applications* **21**, 936–943 (2011).

29. Holzschuh, A., Dormann, C. F., Tscharntke, T. & Steffan-Dewenter, I. Expansion of mass-flowering crops leads to transient pollinator dilution and reduced wild plant pollination. *Proceedings of the Royal Society B: Biological Sciences* **278**, 3444–3451 (2011).

30. Haddad, N. M., Haarstad, J. & Tilman, D. The effects of long-term nitrogen loading grassland insect communities. *Oecologia* **124**, 73–84 (2000).

31. Alroy, J. The shape of terrestrial abundance distributions. *Science Advances* 1, 1–8 (2015).

32. Arita, H. T. Species co-occurrence analysis: Pairwise versus matrix-level approaches. *Global Ecology and Biogeography* **25**, 1397–1400 (2016).

33. Kingston, T., Jones, G., Zubaid, A. & Kinz, T. H. Resource partitioning in rhinolophoid bats revisited. *Oecologia* **124**, 332–342 (2000).

34. Swift, S. M. & Racey, P. A. Resource partitioning in two species of vespertilionid bats (Chiroptera) occupying the same roost. *Journal of Zoology* **200**, 249–259 (1983).

35. Denzinger, A. & Schnitzler, H.-U. Bat guilds, a concept to classify the highly diverse foraging and echolocation behaviors of microchiropteran bats. *Frontiers in Physiology* **4**, 164 (2013).

36. Aldridge, H. D. J. N. & Rautenbach, I. L. Morphology, Echolocation and Resource Partitioning in Insectivorous Bats. *Journal of Animal Ecology* **56**, 763–778 (1987).

37. Freeman, B. G., Class Freeman, A. M. & Hochachka, W. M. Asymmetric interspecific aggression in New Guinean songbirds that replace one another along an elevational gradient. *Ibis* 158, 726–737 (2016).

38. Remsen, J. V. & Graves IV, W. S. Distribution patterns of Buarremon brush-finches (Emberizinae) and interspecific competition in Andean birds. *The Auk* **112**, 225–236 (1995).

39. Alroy, J. Latitudinal gradients in the ecology of New World bats. *Global Ecology and Biogeography* **28**, 784–792 (2019).

40. Kennedy, J. D. *et al.* Into and out of the tropics: The generation of the latitudinal gradient among New World passerine birds. *Journal of Biogeography* **41**, 1746–1757 (2014).

41. Diamond, J. M. & Gilpin, M. E. Examination of the "null" model of Connor and Simberloff for species co-occurrences on Islands. *Oecologia* **52**, 64–74 (1982).

42. Cardinale, B. J., Palmer, M. A. & Collins, S. L. Species diversity enhances ecosystem functioning through interspecific facilitation. *Nature* **415**, 426–429 (2002).

Figures:



Figure 1. Bat and bird and community structure. Metric multidimensional scaling analysis of altered and intact sites for (A) bats and (B) birds based on the Otsuka-Ochiai dissimilarity index⁴³.



Figure 2. Conceptual figure demonstrating the experimental design. (A) Pairwise combinations of species are depicted using an adjacency matrix, with black squares representing granivores and yellow squares representing nectarivores. White numbers along the top and left side of the adjacency matrix are the occupancies N_A and N_B (i.e., numbers of occupied sites) for species A and B, respectively. Species pairs within the adjacency matrix are assigned to one of four groups (g) on the basis of habitat (altered or intact) and dietary guild overlap (competing or control, denoted by blue and gray cells, respectively). Pairs within groups are then subdivided into *occupancy sets* on the basis of shared occupancy values (black paired numbers, calculated as $\{\min(N_A, N_B), \max(N_A, N_B)\}$). (B) Estimates of spatial association, θ_i , are calculated for each occupancy set. (C) The θ_i values are then averaged for each group, yielding posterior distributions for $\overline{\theta_g}$ for each group, $\overline{\theta_g}$. Estimates are calculated separately for altered and intact sites and then compared.



Figure 3. Posterior distributions of $\overline{\theta_g}$ for bat and bird models. Models that contained all species (full) and those containing only species that occurred in both altered and intact habitat types (noturnover) are depicted in adjacent panels.

Methods:

Data

Abundance data, site metadata (including habitat alteration status) and species dietary data for Neotropical bats and birds were downloaded from Ecological Register (http://ecoregister.org). We describe the dataset and our data preparation procedure in detail in the supplement. Our cleaned dataset had 1,876 occurrences of bats and 3,986 occurrences of birds at 106 and 84 sites, respectively. The original and final sites included in the analysis are plotted in Extended Data Fig. 1.

Species richness

We estimate richness for bats and birds in altered and intact sites using three diversity metrics: a corrected first-order jackknife (cJ1)⁴⁴, the canonical Chao1⁴⁵, and Fisher's alpha, which is robust to sample size, widely used, and well-grounded in theory⁴⁶. Each metric is applied to the raw abundance data for all sites. We compare richness estimates between altered and intact sites for both taxa using Wilcoxon rank-sum tests.

Beta diversity and species composition

We compare the beta diversity and species composition of altered and intact sites using a suite of classical analyses. We estimate beta-diversity differences between two groups of samples using a multivariate dispersion test⁴⁷, implemented using the *betadisper* function in the R package *vegan*⁴⁸. We also compare the composition of altered and intact sites for both taxa with permutational multivariate ANOVA using *adonis*⁴⁹ and canonical correspondence analysis using *cca*⁵⁰, also from the *vegan* package. Multivariate dispersion and permutational ANOVA require a distance metric, and the choice of distance metric may influence the results. We therefore run

these analyses using the relatively ubiquitous Jaccard index⁴³ and again with the Otsuka-Ochiai index⁴³. Beta diversity and composition can be compared based on abundance data or occurrence data, noting that the Otsuka-Ochiai index is equivalent to cosine similarity when run on abundance data. We run our analyses with both data types, but we focus on the results obtained with occurrence data, as these are much more common and easily obtained. However, we note cases where results from abundance data differ quantitatively.

Biotic interactions

To explore the effect of interactions on the spatial association of species, we use a combination of novel and existing methods. First, we revive the seldom-used practice of comparing spatial associations between groups of species pairs where one group acts as a control. This approach allows us to isolate the effect of interactions from other confounding variables and has been used e.g, to examine how closely related species associate compared to pairs that are not related¹⁷. Previous research, however, has almost exclusively used co-occurrence metrics that evaluate the departure of species pairs' spatial patterns from random expectations. We recognize here, however, that under a comparative approach, a metric is needed that directly quantifies the magnitude and direction of spatial association for a species pair. The fitted parameter, θ , of Fisher's noncentral hypergeometric distribution (NHD) has these attributes (see Eq. 1c below), and therefore serves as our pairwise association metric. To our knowledge, our study is the first to use the NHD to analyse co-occurrence data. Finally, we introduce a novel methodology to estimate θ , termed "occupancy-set analysis", that entails aggregating pairs into occupancy sets defined based on the numbers of sites occupied by the two species, $\{\min(N_A, N_B), \max(N_A, N_B)\}$ (see Fig. 2). We implement occupancy-set analysis using a hierarchical Bayesian model that estimates the underlying average spatial association for species-pair groups defined based on

habitat (altered or intact) and dietary guild overlap (competing or control). The details of these methods follow.

Species pair groups

We used dietary information for bats and birds from the Ecological Register to infer the effects of food competition on spatial associations. We selected all pairs with complete dietary information and classified them as competing or control. Competing pairs shared primary and secondary sources of food, but we allowed the order of these to be reversed (e.g., a frugivorenectarivore and a nectarivore-frugivore would be equated). Control pairs had no mutual food sources and presumably do not compete for food. Pairs with mutual and nonmutual food sources (e.g., a frugivore and an insectivore-frugivore) may experience a complex mix of effects (e.g. spatial or environmental variation in degree of resource overlap), and were therefore removed from the analysis for simplicity (2,478 or 5.3% of bat pairs and 358,078 or 28% of bird pairs fell into this category). Although species in the same dietary guilds do not necessarily compete, pairs with high dietary overlap have a nonzero probability of experiencing competition for food, while pairs in differing guilds have no chance of competing for food if the categorizations are accurate. Therefore, the co-occurrence patterns of competing and noncompeting groups should be different if food competition plays a role in structuring a given assemblage.

Recent studies repeatedly point out that nonrandom co-occurrence should not be construed as evidence for interaction between species pairs^{51–53}. In our framework, we have independent evidence of potential interactions (diet) and examine the relationship between putative interactions and co-occurrence, using non-competing pairs as a control. Controls are rarely used in co-occurrence research, with most studies preferring to discard pairs with associations that are

not seen as significant (e.g., Refs.^{25,54}). Research using using a control set of pairs to factor out confounding variables can help us to understand under what circumstances, and to what extent, interactions lead to nonrandom spatial associations and how they change under shifting external conditions.

Fisher's noncentral hypergeometric distribution

We assume that the number of sites of co-occurrence, N_{AB} , for a given pair of species, A and B, adheres to Fisher's noncentral hypergeometric distribution (NHD)⁵⁵,

$$f(N_{AB}|\theta) = \frac{\binom{N_A}{N_{AB}}\binom{N_{(A)}}{N_B - N_{AB}}\exp(N_{AB}\theta)}{P_0(\theta)}$$
Eq. 1a

where N_A and $N_{(A)}$ are the respective numbers of sites with species A present and absent, $N_A + N_{(A)}$ is the total number of sites, N_B is the number of sites with species B present, and $\binom{n}{k} = n!/k!/(n-k)!$ is a binomial coefficient. The numerator of this probability mass function represents the number of ways that two species can achieve N_{AB} co-occurrences given fixed values for the two species' occupancies (N_A and N_B) and the total number of sites ($N_A + N_{(A)}$). The denominator, $P_0(\theta)$, is the total number of possible spatial configurations for the species pair given N_A , $N_{(A)}$, and N_B . It is therefore calculated by summing the quantity calculated in the numerator over all possible N_{AB} values⁵⁶, from max $(0, N_B - N_{(A)})$ to min (N_A, N_B) . We characterise this quantity using a more general expression,

$$P_k(\theta) = \sum_{n=max(0,N_B-N_{(A)})}^{min(N_A,N_B)} {N_A \choose n} {N_A \choose N_B-n} \exp(n\theta)n^k$$
Eq. 1b

with k=0, because $P_k(\theta)$ yields compact expressions for subsequent steps of the derivation presented in the supplement. The NHD has one fitted parameter,

$$\theta = \log(\psi) = \log\left(\frac{\pi_{B|A}}{\pi_{B|(A)}}\right) = \log\left(\frac{p_{B|A}(1 - p_{B|(A)})}{(1 - p_{B|A})p_{B|(A)}}\right)$$
Eq. 1c

which is the logarithm of an odds ratio $\psi = \pi_{B|A}/\pi_{B|(A)}$, where $\pi_{B|A}$ and $p_{B|A}$ are the respective odds and probability that species B occupies a site given that species A is present, and $\pi_{B|(A)}$ and $p_{B|(A)}$ are the respective odds and probability that species B occupies a site given that species A is absent. The value of θ , and the probability of having N_{AB} co-occurrences given θ , $f(N_{AB}|\theta)$, are both independent of which member of the species pair is arbitrarily chosen as species A. The NHD is typically parameterised using ψ rather than θ , but ψ has a lower bound of 0, making it less convenient for model fitting and biological interpretation.

The NHD parameter θ directly quantifies spatial patterns of co-occurrence for a species pair. If $\theta < 0$, species B is less likely to occur at a site if species A is present, as we would expect if there was competitive exclusion or differences in habitat preferences between the two species. If $\theta > 0$, species B is more likely to occur at a site if species A is present, as we would expect if the species shared habitat preferences and did not competitively exclude each other. Finally, if $\theta = 0$, the two taxa are independently distributed. When $\theta = 0$, the NHD reduces to the standard (i.e., centered) hypergeometric distribution, which has frequently been used as a null model to evaluate whether patterns of co-occurrence differ from random³². To our knowledge, our study is the first to use the noncentral form of the hypergeometric distribution for co-occurrence analyses. In light of the rich array of methodologies already available for quantifying species associations, the use of the NHD parameter θ as a co-occurrence metric must be justified. First, while θ is

mathematically related to existing co-occurrence metrics (e.g., the mid-P variant of Fisher's Exact Test⁵⁷), it fundamentally differs in that it is not based on a null model. Instead, θ directly quantifies the magnitude of the effect of one species' presence on the occurrence probability of another (analogous to the distinction between a correlation measure of effect size, *r*, and a p-value calculated under the null hypothesis that *r* = 0). Thus, θ is biologically interpretable (Eq. 1c), it is directly comparable between pairs, and it is easily programmable. Finally, θ is not sensitive to the number of sites in the study assemblage, making formal comparisons and meta-analyses possible down the track. Because existing co-occurrence methods are not commensurate with our approach, we do not attempt to draw any direct comparisons. However, we perform a series of analyses to demonstrate the behaviour of the NHD parameter and evaluate its ability to quantify spatial associations between species (see supplement).

Occupancy-set analysis

Our analysis on the performance of θ (supplement) highlights three general issues that arise when fitting the NHD to occurrence data:

- (i) A non-overlapping pair (i.e., $N_{AB} = 0$) will be observed with higher probability for a species pair with lower occurrence values (N_A and N_B), irrespective of θ .
- (ii) A *single* observation of a non-overlapping pair will not by *itself* be biologically informative about the value of θ (i.e., the maximum likelihood estimate $\hat{\theta} = -\infty$).
- (iii) The amount of information about θ that can be inferred from a *single* observation is generally lower for a species pair with lower occurrence values.

In other words, the parameter θ suffers from similar constraints as most existing metrics of species association, which attempt to leverage biological information from pairwise occurrence

data. These constraints are seldom considered explicitly in published co-occurrence analyses, aside from throwing out pairs that do not yield biological information by themselves.

To address these three issues, we present a novel approach, which we term "occupancy-set analysis". Rather than separately estimate θ for each species pair, *j*, based on the number of cooccurrences for that pair, N_{AB}^{j} , we instead *combine* observations for pairs grouped into occupancy sets, S_i , defined for pairs $j \in S_i$ on the basis of shared occupancy values, {min(N_A , N_B), max(N_A , N_B)} (see Fig. 2). We then calculate the likelihood for each occupancy set based on a single θ value, θ_i , under the explicit assumption that every pair in the set has the same θ value. This likelihood,

$$\mathcal{L}(\theta_i \mid \mathbf{N}_{AB}^{i}) = \prod_{j \in \mathcal{S}_i} f\left(N_{AB}^{j} \mid \theta_i\right)$$
 Eq. 2

is simply the product of the likelihoods for the co-occurrence observations in the occupancy set, $N_{AB}^{i} = \{N_{AB}^{j} | j \in S_{i}\}$. This approach partly addresses issues (*i*) and (*ii*) because $N_{AB} = 0$ is less likely to be observed for an entire set of observations than a single observation, and the maximum likelihood estimate for θ_{i} will be finite provided that at least one observation in the occupancy set is not an edge case (i.e., $N_{AB} > \max(0, N_B - N_{(A)})$ and $N_{AB} < \min(N_A, N_B)$). It also addresses issue (*iii*) because the Fisher Information is additive⁵⁸, meaning that it increases linearly with the number of observations. As an added benefit, performing likelihood calculations on occupancy sets reduces the time required for model fitting by orders of magnitude as compared to separately estimating θ for each pair.

Specifying the model

To further address issues (*i*) and (*ii*), we adopt a hierarchical Bayesian modeling approach⁵⁹ to estimate averages for θ_i that have been subdivided into four groups, *g*, based on habitat type (altered, intact) and diet (competing, control). At the lower (i.e., occupancy-set) level, we assume that the θ_i values are all normally distributed with means and standard deviations that can vary among groups

$$\theta_i \sim \mathcal{N}(\overline{\theta}_{g(i)}, \sigma_{g(i)})$$
 Eq. 3

where g(i) is a function that indexes the group assignment of each set. At the upper (i.e., community) level, we assign the group means, $\overline{\theta}_g$, a normal prior distribution with a mean of 0 and a standard deviation of 10,

$$\overline{\theta}_g \sim \mathcal{N}(0,10)$$
 Eq. 4

and we assign the group standard deviations, σ_g , a half-normal prior distribution with location and scale parameters of 0 and 10, respectively, on the interval $[0, \infty]$,

$$\sigma_g \sim \mathcal{H}(0,10)$$
 Eq. 5

These prior distributions were chosen to allow adequate exploration of the parameter space using the Markov chain Monte Carlo (MCMC) algorithm while ensuring that the algorithm never halted by getting stuck in regions of parameter space with negligible likelihood. These prior distributions should be viewed as very weakly informative in the parlance of Bayesian analysis⁶⁰. Our implementation of occupancy-set analysis within a hierarchical Bayesian modelling framework confers three major advantages over previous approaches when calculating spatial associations for assemblages. First, by treating the occupancy set as the experimental unit, it can extract more information about pairwise spatial associations from the co-occurrence data than analyses that treat the pair as the experimental unit, yielding more robust inferences. Second, the algorithm is formulated in such a way that adding species does not appreciably increase the computing time of the model, meaning hyper-diverse datasets for which co-occurrence analysis is not feasible can be analysed using occupancy-set analysis. Finally, the hierarchical approach ensures that sets of pairs yielding weaker evidence (as indexed by smaller likelihoods) exhibit greater shrinkage toward the mean (see supplement), and sets yielding stronger evidence are assigned greater weight in calculating the group-level means and standard deviations.

Fitting the model

We estimate the posterior distributions using the R package RStan, which provides an interface from R to the Stan probabilistic programming language⁶¹. To approximate the posterior distributions, we generate 4 MCMC chains of 2000 steps, including a warm-up period of 1000 steps. We then inspect MCMC plots for the 4 chains and calculated Gelman Rubin statistics, to ensure convergence⁵⁹. We run a series of robustness checks that ensure our model is not subject to several common biases (see supplement).

Model interpretation

In this framework, a change in the posterior distributions of $\overline{\theta_g}$ can point to various biological processes (Extended Data Fig. 2). Competing pairs can display lower $\overline{\theta_g}$ than non-competing pairs, which means that diffuse competition tends to lead to exclusion at some sites with respect to opportunity for co-occurrence (Extended Data Fig. 2A). This is the case even if all $\overline{\theta_g}$ are positive, as exclusion does not necessarily present as a negative association. Specifically,

because competing and non-competing pairs are comprised of the same presence-absence data in different pairwise arrangements, the $\overline{\theta_g}$ of non-competing pairs represents the opportunity for coexistence in the absence of food competition and controls for the effects of all confounding variables (e.g biogeography, other types of interactions, etc.). A deviation toward lower $\overline{\theta_g}$ from this baseline is evidence that competing pairs are not coexisting as often as expected–a systematic pattern of exclusion at the group level. Conversely, a higher $\overline{\theta_g}$ in competing pairs confirms that similarity in resource use leads to coexistence, suggesting that competing pairs have adapted to partition resources (Extended Data Fig. 2B). If the $\overline{\theta_g}$ of competing and noncompeting pairs do not differ significantly, but the σ_g (variance) of competing pairs is higher, this points to stronger spatial patterns (both coexistence and exclusion) in pairs that compete for food, and vice versa (Extended Data Fig. 2D). If there is no difference in $\overline{\theta_g}$ or σ_g between the two groups, then there is insufficient evidence to support the hypothesis that diffuse competition is influencing the structure of the assemblage at the landscape scale (Extended Data Fig. 2C).

The $\overline{\theta_g}$ and σ_g of competing and non-competing pairs might also differ between altered and intact sites in our example. If the relationship between competing and non-competing pairs is different, this means that alteration has changed the outcome of biotic interactions, e.g., by promoting partitioning over exclusion or vice versa. More subtly, the extent of disparity between the two groups could differ without qualitatively changing the relationship, which indicates that naturally existing mechanisms have been enhanced or dampened. On the other hand, $\overline{\theta_g}$ or σ_g could simply be higher or lower across the board, suggesting that alteration leads to changes in community structure that do not necessarily involve the rewiring of competitive interactions. In all cases, posteriors represent assemblage-level patterns and do not imply that all pairs in the group exhibit the same pattern as does their group, merely that there is a group tendency toward one process over another.

The experimental approach presented here represents two distinct innovations that allow us to deal with detection issues and context-dependence of interactions. First, we use a priori evidence for the possibility of interaction rather than direct interaction data. The resulting set of pairs includes both interacting and non-interacting pairs but encompasses all of the former. A priori evidence for many types of interactions could be derived from various functional traits. For example, fruit size and gape size of frugivorous birds can indicate which birds are physically able to disperse which fruits. Diet category and body size can similarly be used to estimate mammal predator-prey relationships⁶². This setup frees us from the necessity of manually observing all interactions and identifying the contexts under which they are realised while yielding broad insights about the impact of interactions on community assembly.

Second, we use non-interacting pairs of species in the assemblage to contextualise association patterns for interacting species pairs. The inclusion of an experimental control group is a central tenet of science, but non-interacting pairs or those having associations not significantly different from chance associations are routinely discarded or ignored in co-occurrence research and interaction research more generally, while raw pairwise co-occurrence scores are often interpreted at face value (e.g., any negative score equals exclusion). Our use of non-interacting pairs as a control can be applied to a broad variety of interactions. A plant-pollinator network could for instance be decomposed into groups of pairs that facilitate one another (plant-pollinator pairs), do not interact, and even compete (plant-plant pairs that compete for pollinators or pollinator-pollinator pairs that compete for nectar). In each case, the effect of the competing group(s) can be evaluated against all pairs that do not exhibit the interaction(s) in question.

The effect of turnover

Overall changes observed in association patterns could result from pairs changing their spatial relationships or their competition intensity in altered vs. intact habitats. Conversely, a shift in species composition (e.g., the removal or introduction of interaction partners), and therefore the presence of a different suite of pairs, could also cause disparities between the two habitat types. To investigate these possibilities, we ran the models with all species and then repeated them with only species occurring at least once in both habitat types ("no-turnover" models). If the results of the two model sets are very similar, we can conclude that the results are driven by changes in the spatial relationships of species occurring in both site types. If the two model sets are different but both show differences between groups, likely both turnover and changes in preserved interactions are occurring. Furthermore, the differences between the two outputs can be used to determine which changes are due to which mechanism. Model effects are due to turnover alone if the model set using only shared species has no differences while the model set using all species does.

Data and code availability

All R workflows and data that support the findings of this study are freely available at https://github.com/anikobtoth/HabitatAlteration.

Method References:

43. Hubalek, Z. Coefficients of association and similarity, based on binary (presenceabsence) data: an evaluation. *Biological Reviews of the Cambridge Philosophical Society* **57**, 669–689 (1982).

44. Alroy, J. On four measures of taxonomic richness. *Paleobiology* **46**, 158–175 (2020).

45. Chao, A. Nonparametric estimation of the number of classes in a population. *Scandinavian Journal of Statistics* **11**, 265–270 (1984).

46. Schulte, R. P. O., Lantinga, E. A. & Hawkins, M. J. A new family of Fisher-curves estimates Fisher's alpha more accurately. *Journal of Theoretical Biology* **232**, 305–313 (2005).

47. Anderson, M. J., Ellingsen, K. E. & McArdle, B. H. Multivariate dispersion as a measure of beta diversity. *Ecology Letters* **9**, 683–693 (2006).

48. Oksanen, J. *et al.* The vegan package. (2008).

49. Anderson, M. J. A new method for non-parametric multivariate analysis of variance. *Austral Ecology* **26**, 32–46 (2001).

50. Legendre, P. & Legendre, L. Numerical Ecology. 625–629 (Elsevier, 2012).

51. Freilich, M. A., Wieters, E., Broitman, B. R., Marquet, P. A. & Navarrete, S. A. Species co-occurrence networks: Can they reveal trophic and non-trophic interactions in ecological communities? *Ecology* **99**, 690–699 (2018).

52. Thurman, L. L., Barner, A. K., Garcia, T. S. & Chestnut, T. Testing the link between species interactions and species co-occurrence in a trophic network. *Ecography* **42**, 1658–1670 (2019).

53. Blanchet, F. G., Cazelles, K. & Gravel, D. Co-occurrence is not evidence of ecological interactions. *Ecology Letters* **23**, 1050–1063 (2020).

54. Araújo, M. B., Rozenfeld, A., Rahbek, C. & Marquet, P. A. Using species co-occurrence networks to assess the impacts of climate change. *Ecography* **34**, 897–908 (2011).

55. Fog, A. Calculation methods for Wallenius' noncentral hypergeometric distribution. *Communications in Statistics – Simulation and Computation* **37**, 258–273 (2008).

McCullagh, P. & Nelder, J. A. *Generalized Linear Models, 2nd Edn.* (Chapman; Hall, 1952). doi:10.2307/2347392.

57. Keil, P., Wiegand, T., Tóth, A. B., McGlinn, D. J. & Chase, J. M. Measurement and analysis of interspecific spatial associations as a facet of biodiversity. *Ecological Monographs*91, 1–22 (2021).

58. Ly, A., Marsman, M., Verhagen, J., Grasman, R. P. P. P. & Wagenmakers, E. J. A Tutorial on Fisher information. *Journal of Mathematical Psychology* **80**, 40–55 (2017).

59. Gelman, A. et al. Bayesian Data Analysis. (CRC press, 2013).

60. Lemoine, N. P. Moving beyond noninformative priors: why and how to choose weakly informative priors in Bayesian analyses. *Oikos* **128**, 912–928 (2019).

61. Stan Development Team. RStan: the R interface to Stan. (2022).

62. Pires, M. M. *et al.* Pleistocene megafaunal interaction networks became more vulnerable after human arrival. *Proceedings of the Royal Society B* **282**, (2015).

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Author contributions: ABT and JA designed the study, JA curated the data, JA and APA provided statistical expertise, ABT analysed data, produced the figures, and wrote the paper.

ABT and APA designed, coded, and tested the model. SKL helped design supporting analyses. All authors edited the manuscript.

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Additional Information:

Supplementary Information is available for this paper. Correspondence and requests for materials should be addressed to Dr. Anikó B. Tóth at aniko.toth@unsw.edu.au. Reprints and permissions information is available at www.nature.com/reprints.

Extended Data:



Extended Data Fig. 1 Maps of Neotropical bat and bird sites before (top) and after (bottom) biogeographic matching procedure, with altered sites represented in beige and intact sites represented in teal.



Extended Data Fig. 2 Interpretations of average $\overline{\theta_g}$ posterior density distributions. The dark curve indicates competing pairs and the lighter curve indicates control pairs. In (A) competing pairs co-occur less on average than control pairs, so higher spatial segregation than expected suggesting a pattern of competitive exclusion. In (B) competing pairs co-occur more than control pairs on average, suggesting that they micro-partition resources at the same sites more than expected. In (C) there is little or no separation of the posterior distributions, meaning that food competition does not aggregate or segregate pairs outside of expectations; however, other interaction types could still play an important role. (D) Both strong aggregation and segregation are observed in competing pairs, so variance is higher than expected.



Extended Data Fig. 3 Performance of θ . Effects of species occurrence (indexed by N_A and N_B) and spatial aggregation ($\theta > 0$) and segregation ($\theta < 0$) on the probability distribution of co-occurrences. Probabilities were calculated using Eq. 1 based on a total of $N_A + N_{(A)} = 50$ sites.



Extended Data Fig. 4 Fisher information of varying occupancy pairs. Effects of occupancy (N_A and N_B) and spatial aggregation (θ) on the probability distribution of co-occurrences. Probabilities were calculated using Eq. S2, assuming a total of $N_A + N_{(A)} = 50$ sites.



Extended Data Fig. 5 Model shrinkage. Shrinkage behavior of theta estimates for the full models for (A-B) bats and (C-D) birds. Y axes represent deviations of maximum likelihood estimates of spatial association, $\hat{\theta}_i$, which are calculated separately for each occupancy set (see Fig. 2), from Bayesian estimates, θ_i , which involve data pooling (Eq. 3). Maximum likelihood estimates will be $\pm \infty$ whenever all of the co-occurrence observations in the set take the minimum values (i.e. $N_{AB} = \max(0, N_B - N_{(A)})$, 2.9% and 0.26% of occupancy sets for bats and birds, respectively) or maximum values (i.e. $N_{AB} = \min(N_A, N_B)$, 2.1% and 0.26% of sets for bats and birds, respectively).



Extended Data Fig. 6 Distribution of θ_i values. There is one θ value for each occupancy set for each experimental group in the bat model (A) and the bird model (B).



Extended Data Fig. 7 Bayesian estimates of spatial association, θ_i , for bats plotted against: (A) the occupancy of the rarer species in the occupancy set, and (B), the number of pairs in the occupancy set. Each point represents an estimate for an occupancy set (see Fig. 2). Lowess curves (blue lines) are included for visualization only.



Extended Data Fig. 8 Bayesian estimates of spatial association, θ_i , for birds plotted against: (A) the occupancy of the rarer species in the occupancy set, and (B), the number of pairs in the occupancy set. Each point represents an estimate for an occupancy set (see Fig. 2). Lowess curves (blue lines) are included for visualization only.

Extended Data Table 1. Results of richness analyses comparing estimated richness in altered and intact habitats using three metrics. Table includes median metric output across sites and results of two-tailed Wilcoxon rank-sum tests comparing intact and altered richnesses. Sample sizes after biogeographic correction were 53 each of altered and intact bat sites and 42 each altered and intact bird sites. Key: chao= Chao 1, cJ1 = second-order jaccknife, fa = Fisher's Alpha, p = p value of Wilcoxon Rank Sum test, W = test statistic of Wilcoxon Rank Sum test.

metric	taxon	Altered	Intact	р	W
chao	bat	21.0	20.0	0.918	1,045.5
chao	bird	51.1	68.6	0.590	801.0
cJ1	bat	19.4	18.8	0.613	1,324.0
cJ1	bird	47.7	55.8	0.660	832.0
fa	bat	3.6	4.7	0.007	975.0
fa	bird	13.9	16.1	0.820	856.0

Extended Data Table 2. Results of beta diversity and composition analyses comparing community patterns in altered and intact habitats, such that significant p-values signal a difference in composition between the two site types. F-statistic and *p*-values are displayed for bats and birds.

analysis	data	metric	p.bat	p.bird	F.bat	F.bird
adonis	binary	Jaccard	0.5997	0.0360	0.9073	1.2910
adonis	binary	Ochiai	0.7402	0.0315	0.7247	1.5038
adonis	abundance	Jaccard	0.2706	0.0112	1.0941	1.2490
adonis	abundance	Cosine similarity	0.4962	0.0099	0.8959	1.6130
beta dispersion	binary	Jaccard	0.2845	0.1898	1.1573	1.7481
beta dispersion	binary	Ochiai	0.1833	0.1952	1.7944	1.7058
beta dispersion	abundance	Jaccard	0.5157	0.3984	0.4254	0.7205
beta dispersion	abundance	Cosine similarity	0.5665	0.7218	0.3306	0.1277
сса	binary	NA	0.1700	0.0370	1.1135	1.1517
сса	binary	NA	0.1590	0.0290	1.1135	1.1517
cca	abundance	NA	0.0020	0.0010	1.7831	1.6590
сса	abundance	NA	0.0020	0.0010	1.7831	1.6590

Extended Data Table 3. Median values and 95% high density posterior intervals for $\overline{\theta_g}$ and σ_g of all bat and bird models. Column "n" is the sample size, i.e., the number of occupancy sets in each group.

taxon	model	status	Pair type	n	Median θ	Lower θ	Upper θ	Median σ	Lower σ	Upper σ
bat	full	Intact	control	269	0.3359	0.2717	0.3995	0.3743	0.3111	0.4387
bat	full	Altered	control	252	0.3389	0.2610	0.4187	0.4987	0.4171	0.5844
bat	full	Intact	competing	224	0.5218	0.4232	0.6165	0.5622	0.4691	0.6724
bat	full	Altered	competing	234	0.3814	0.2401	0.5128	0.8971	0.7647	1.0377
bat	noTurnover	Intact	control	269	0.3454	0.2785	0.4104	0.3861	0.3193	0.4585
bat	noTurnover	Altered	control	252	0.3368	0.2571	0.4194	0.4962	0.4143	0.5843
bat	noTurnover	Intact	competing	224	0.5536	0.4556	0.6550	0.5813	0.4805	0.6842
bat	noTurnover	Altered	competing	234	0.3954	0.2692	0.5336	0.8647	0.7363	1.0028
bird	full	Intact	control	78	0.4882	0.4094	0.5620	0.3063	0.2445	0.3764
bird	full	Altered	control	117	0.4036	0.3428	0.4596	0.2695	0.2183	0.3212
bird	full	Intact	competing	77	0.5551	0.4784	0.6307	0.2991	0.2411	0.3700
bird	full	Altered	competing	113	0.3858	0.3232	0.4432	0.2431	0.1907	0.2959
bird	noTurnover	Intact	control	78	0.5116	0.4457	0.5805	0.2722	0.2172	0.3450
bird	noTurnover	Altered	control	117	0.3918	0.3237	0.4574	0.3122	0.2598	0.3776
bird	noTurnover	Intact	competing	77	0.5795	0.4997	0.6498	0.2829	0.2239	0.3481
bird	noTurnover	Altered	competing	113	0.3896	0.3228	0.4675	0.3081	0.2473	0.3738

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