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### ARTICLE

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# The Angola Gyre is a hotspot of dinitrogen fixation in the South Atlantic Ocean

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Biological dinitrogen fixation is the major source of new nitrogen to marine systems and thus essential to the ocean's biological pump. Constraining the distribution and global rate of dinitrogen fixation has proven challenging owing largely to uncertainty surrounding the controls thereon. Existing South Atlantic dinitrogen fixation rate estimates vary five-fold, with models attributing most dinitrogen fixation to the western basin. From hydrographic properties and nitrate isotope ratios, we show that the Angola Gyre in the eastern tropical South Atlantic supports the fixation of 1.4–5.4 Tg N.a<sup>-1</sup>, 28-108% of the existing (highly uncertain) estimates for the basin. Our observations contradict model diagnoses, revealing a substantial input of newly-fixed nitrogen to the tropical eastern basin and no dinitrogen fixation west of 7.5°W. We propose that dinitrogen fixation in the South Atlantic occurs in hotspots controlled by the overlapping biogeography of excess phosphorus relative to nitrogen and bioavailable iron from margin sediments. Similar conditions may promote dinitrogen fixation in analogous ocean regions. Our analysis suggests that local iron availability causes the phosphorus-driven coupling of oceanic dinitrogen fixation to nitrogen loss to vary on a regional basis.

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he cycling of nitrogen (N) controls the fertility of the global ocean<sup>1,2</sup> and may influence the ocean's role in climate<sup>3</sup>. The ultimate source of marine N to the ocean is N<sub>2</sub> fixation, whereby specialized plankton, diazotrophs, transform inert N<sub>2</sub> gas into bioavailable ammonium. This process occurs dominantly in the warm, sunlit surface of the (sub)tropical ocean. The ultimate sink for marine N is denitrification, the bacterial reduction of oxidized N species to N<sub>2</sub> gas in oxygen-deficient waters and sediments. Considerable uncertainty exists regarding the magnitude of the global N source and sink terms, including whether these are balanced on timescales of ocean mixing<sup>4–7</sup>. An incomplete understanding of the controls on N<sub>2</sub> fixation, particularly at the regional scale, likely contributes to the apparent discrepancies in estimates of oceanic N loss and gain.

A stoichiometric phosphorus (P) excess relative to N, termed 'excess P', is a condition proposed to favour N<sub>2</sub> fixation<sup>6</sup>. Much of the global surface ocean hosts excess P<sup>8</sup>, which ultimately derives from denitrification in oxygen-deficient zones and sediments<sup>6,9</sup>. The South Atlantic receives excess P both remotely and regionally, in the latter case following seasonal N loss in the northern Benguela upwelling system (NBUS)<sup>6,10,11</sup>, yet this basin appears to support little N2 fixation. Direct and geochemical estimates of South Atlantic N<sub>2</sub> fixation rates, although notably sparse, range from 1.8 to 8.4 Tg N.a<sup>-1</sup>, with disagreement as to the regional distribution<sup>6,12–17</sup> (Table 1). These low rates have been attributed to iron limitation of diazotrophs<sup>9,14,17-19</sup> as the South Atlantic receives at least a hundred-times less aeolian iron than the North Atlantic  $(3 \pm 2 \text{ versus } 896 \pm 14 \text{ nmol iron.m}^{-2} \cdot d^{-1})^{20}$  where N<sub>2</sub> fixation is considerably higher, with estimates ranging from 24 to 47 Tg N.a $^{-1.6,9,12-16,21}$  (Table S1). The excess P that goes largely unused in the South Atlantic traverses the basin in its equatorward-flowing surface layer, ultimately fuelling N<sub>2</sub> fixation in the (sub)tropical North Atlantic following its entrainment into iron-replete, P-limited surface waters<sup>9,14,22</sup>. Nevertheless, some N<sub>2</sub> fixation likely occurs in the South Atlantic, yet its magnitude and distribution, and the controls thereon, remain poorly characterized.

The estimation of N<sub>2</sub> fixation has often relied on ocean models, most of which exploit regional variations in the nitrateto-phosphate (NO<sub>3</sub><sup>-</sup>:PO<sub>4</sub><sup>3-</sup>) ratio<sup>6,12,13</sup> to derive rates from inverse circulation fields. Variations in N:P are typically quantified by the parameters N\* and P\*<sup>6,23,24</sup>, defined here as  $[NO_3^{-}] - 15.5 \times [PO_4^{3-}]^{23}$  and  $[PO_4^{3-}] - [NO_3^{-}]/15.5^6$ , respectively, where 15.5:1 is the mean N:P ratio of nutrients supplied to the tropical South Atlantic thermocline (SI.1). The conceptual basis for the estimation is that diazotrophy pairs the production of newly-fixed N with the assimilation of P. N<sub>2</sub> fixation thus removes P but not N from surface waters and leads to a sinking organic matter (OM) flux with an N:P ratio that is higher than that of the net supply of nitrate and phosphate to the surface<sup>6,23,25</sup>. However, the N:P ratio of plankton biomass, and thus of sinking OM, varies regionally<sup>26</sup>, complicating the calculation of  $N_2$  fixation rates based on nutrient stoichiometry<sup>27</sup>. Recent modelling efforts have attempted to address this shortcoming by including variable, yet still prescribed, OM N:P ratios<sup>13</sup>.

Nitrate isotope ratio measurements provide an important alternative source of information for diagnosing N<sub>2</sub> fixation<sup>14,28,29</sup>. N<sub>2</sub> fixation introduces nitrate to the thermocline with a  $\delta^{15}$ N of -2 to  $0\%^{28,30-32}$ , distinct from that of deep-ocean nitrate ( $\geq 5\%^{33}$ ;  $\delta^{15}$ N =  $[(^{15}N)^{14}N)_{sample}/(^{15}N)^{14}N)_{ref} - 1] \times 10^3$ , in % versus N<sub>2</sub> in the air). Negative excursions in source-to-thermocline nitrate  $\delta^{15}$ N can thus be used to diagnose and quantify N<sub>2</sub> fixation<sup>28</sup>. The N isotope ratios have some major advantages over N:P ratios in this regard. In particular, the  $\delta^{15}$ N of newly-fixed N is relatively well-constrained<sup>30,31</sup>, whereas the N:P ratios of both diazotrophs and non-N<sub>2</sub>-fixing plankton are uncertain and potentially dynamic<sup>25–27</sup>.

Here we analyse nitrate isotope ratios and nutrient stoichiometries for the tropical South Atlantic Ocean based on new and existing measurements<sup>14,34–36</sup> (see 'Methods'). These data reveal substantial N<sub>2</sub> fixation in the Angola Gyre to the east of the basin and none in the west, in contrast to current notions that N<sub>2</sub> fixation occurs predominantly in the western tropical South Atlantic<sup>6,13</sup> (Fig. 1). We hypothesize that the confluence of remote and regional sources of excess P and iron supplied proximately from the ocean margin determines the incidence of N<sub>2</sub> fixation in the Angola Gyre. Our findings have implications for regional variations in N<sub>2</sub> fixation and its coupling to N loss throughout the global ocean<sup>37,38</sup>.

#### **Results and discussion**

Properties of the Angola Gyre. The Angola Gyre, a permanent upwelling feature embedded within the larger-scale cyclonic South Atlantic Tropical Gyre<sup>39-41</sup>, was encountered east of 7.5°W along a zonal transect of the Atlantic at 6-15°S (Fig. 1; CoFeMUG<sup>14,34</sup> and MET131 cruises; see 'Methods'). The gyre is characterized by a shoaling of the thermocline, defined as the waters between potential density anomalies ( $\sigma_{\theta}$ ; in kg.m<sup>-3</sup>) 26.2 and 27.0, overlying an oxygen minimum zone where apparent utilization  $(AOU = O_{2saturated} - O_{2observed})$ oxygen is  $>240 \,\mu M^{40-42}$  (Fig. 2a, b, f, g). High rates of primary productivity fuelled by persistent nutrient upwelling drive elevated OM export from the Angola Gyre mixed layer, which is remineralized in the aphotic zone below the surface<sup>43,44</sup>. A subsurface iron plume emanating from the nearby low-oxygen marginal sediments penetrates the Angola Gyre (dFe ~0.8-2 nM; Fig. 2c). Intense remineralization and repeated cycles of supply and export retain this bioavailable iron within Angola Gyre waters<sup>34</sup>. Local remineralization also yields a

Table 1 Existing areal N <sub>2</sub> fixation rate estimates (Tg N.a <sup>-1</sup> ) for the South Atlantic.			
Domain	Method	Rate [Tg N.a <sup>-1</sup> ]	Reference
0°S-25°S	N* plus forward circulation model	8.4	12
11°S-35°S	N:P plus inverse circulation model	8.3	13
0°S-40°S	P* plus inverse circulation model	7.5 <sup>a</sup>	6
11°S-30°S	Nitrate isotopes plus volume flux	3 ± 0.5	14
10°S-45°S	Direct rate measurements	5 ± 2 <sup>b</sup>	15
0°S-40°S	Direct rate measurements	1.8 ± 0.6 <sup>b</sup>	16
6°S-15°S	Nitrate isotopes plus residence time	2.0-4.1	This study

The arithmetic mean (±1 SD) for 5°S to 35°S is  $5.0 \pm 3.3$  Tg N.a<sup>-1</sup>. This average excludes our N<sub>2</sub> fixation estimate, which is relevant to the tropical South Atlantic domain only. For a detailed list of the available areal N<sub>2</sub> fixation rate estimates, see Table 51. <sup>a</sup>Estimated from a homogenized rate for the South Atlantic (area = 27 × 10<sup>12</sup> m<sup>2</sup> for 0-40°S<sup>16</sup>).

<sup>b</sup>Compilation of direct measurements made by others.



**Fig. 1 Surface P\* across the South Atlantic Ocean.** Station locations (coloured symbols) overlaid on an annual climatology of the South Atlantic surface P\* concentration (= $[PO_4^{3-}]-[NO_3^{-}]/15.5$ ; Sl.1). As indicated in the legend, circles represent stations sampled during the CoFeMUG cruise in November/ December 2007<sup>14,34</sup>, triangles show stations sampled during the MET131 cruise in October 2016, and diamonds indicate stations sampled during the CLIVAR/GO-SHIP A16N cruise in September/October 2013<sup>14,49</sup>. Red symbols show Angola Gyre stations and blue symbols represent western basin stations. The red cell indicates the approximate position of the Angola Gyre, and the white arrows show the circulation of thermocline waters in the tropical South Atlantic Ocean. The black box encloses P\* values calculated from the CoFeMUG data set, while the other P\* data are from WOA18<sup>8</sup> (Sl.1). The dotted light-grey contours show the distribution of recent modelled water column-integrated N<sub>2</sub> fixation rates for the South Atlantic(1, 25, 50 and 100 mmol N.m<sup>-2</sup>.a<sup>-1</sup>)<sup>13</sup>. The South Atlantic Tropical Gyre is defined by the sSEC, SEUC and AC, where sSEC is south South Equatorial Current, SEUC is South Equatorial Current, and AC is Angola Current. Additionally, nSEC is north South Equatorial Current, EUC is Equatorial Undercurrent, eSEC is central South Equatorial Current, SECC is South Equatorial Countercurrent, GCUC is Gabon-Congo Undercurrent, ABFZ is Angola-Benguela Frontal Zone (indicated by the parallel wavy lines), NBUS is northern Benguela upwelling system, BC is Benguela Current, BrC is Brazil Current, and STsG is Subtropical subgyre.

nitrate concentration maximum (>40  $\mu$ M) beneath the thermocline ( $\sigma_{\theta}$  > 27.0), contrasting with the lower concentration of sub-thermocline nitrate (<35  $\mu$ M) and AOU (<180  $\mu$ M) in the tropical western basin (west of 7.5°W; Figs. 2b, g and 3a).

Nitrate  $\delta^{15}$ N across the transect is consistently high in Tropical Surface Water ( $\sigma_{\theta} < 26.2$ ) due to isotopic fractionation during nitrate assimilation by phytoplankton, which raises nitrate  $\delta^{15}$ N (and  $\delta^{18}O^{45,46}$ ; which, in ‰ versus VSMOW = [( $^{18}O$ ) $_{ample}/$ ( $^{18}O$ ) $_{ref} - 1$ ] × 10<sup>3</sup>; SI.3) to >12‰. The mean nitrate  $\delta^{15}$ N in the underlying thermocline is 5.7 ± 0.7‰ in the Angola Gyre, reaching as low as 4.6‰, a salient decrease from the western tropical basin where the average  $\delta^{15}$ N of thermocline nitrate is 6.3 ± 0.1‰ (Figs. 2d, h and 3b). Subantarctic Mode Water (SAMW;  $\sigma_{\theta} = 27.0-27.25$ ), recently formed in the Southern Ocean, underlies the thermocline of the tropical South Atlantic and is the ultimate source of nutrients to its surface waters<sup>47</sup>. In the western tropical basin, SAMW nitrate  $\delta^{15}$ N is 6.2 ± 0.1‰, indistinguishable from the  $\delta^{15}$ N of SAMW nitrate beneath the Angola Gyre (6.4 ± 0.1‰; t(150) = -12.05, p = 0.01) (Figs. 2d, h and 3b).

Mean SAMW N\* is  $0.0 \pm 0.1 \,\mu$ M across the basin, increasing in the Angola Gyre thermocline to  $0.7 \pm 0.9 \,\mu$ M, coincident with the source-to-thermocline decline in nitrate  $\delta^{15}$ N. N\* decreases into the western tropical basin thermocline, to  $-0.4 \pm 0.5 \,\mu$ M (Fig. 3c and SI.1 and 4).

Zonal trends in the tropical South Atlantic: evidence for an exogenous source of N to the Angola Gyre. Thermocline nitrate  $\delta^{15}$ N in the western tropical South Atlantic basin is similar to that of the underlying source (6.3 ± 0.1‰ versus 6.2 ± 0.1‰) while the  $\delta^{15}$ N of nitrate in the Angola Gyre thermocline is distinctly lower, by 0.7–1.8‰ (Figs. 2d, h and 3b). Since surface-water nitrate is generally completely consumed, and given that nitrification in the ocean interior typically competes with no other process—thus

negating the impact of isotopic fractionation during remineralization in the subsurface—remineralized OM produced from Tropical Surface Water will return nitrate to the thermocline with a  $\delta^{15}$ N that is indistinguishable from the underlying SAMW source<sup>29,48,49</sup>. The fact that the  $\delta^{15}$ N of thermocline nitrate in the Angola Gyre is lower than SAMW nitrate  $\delta^{15}$ N signals an exogenous source of N to surface waters, the consumption and subsequent remineralization of which yields low- $\delta^{15}$ N nitrate in the Angola Gyre thermocline<sup>28,32</sup>.

Zonal and vertical changes in nutrient stoichiometry corroborate the notion of an exogenous source of N to the Angola Gyre. The SAMW-to-thermocline N\* increase in the gyre is consistent with the addition of N in stoichiometric excess of P (i.e., relative to the N:P ratio of the regional nutrient supply, 15.5:1) while the SAMW-to-thermocline N\* decrease in the western tropical basin suggests that sinking OM may be lower in N:P than the regional nutrient supply (SI.4).

Despite the existing data pointing to complete nitrate consumption in the Angola Gyre surface, instances of incomplete nitrate assimilation could occur in this perennially-upwelling feature, and these could cause the loss of high- $\delta^{15}N$  nitrate (by advection), with low- $\delta^{15}$ N OM retained and remineralized in the gyre thermocline<sup>29,50</sup>. However, the regeneration of OM deriving from partial nitrate assimilation, which would have an N:P of ~15.5:1, could not yield the high  $N^*$  observed in the thermocline. Additionally, mixed-layer nitrate was measurable at only three of 19 stations at the time of our sampling, reinforcing the view that nitrate is seldom unconsumed in Angola Gyre surface waters in spring, and likewise in autumn<sup>44</sup>. The  $\delta^{15}$ N of thermocline nitrate could arguably be lowered by the lateral export of high- $\delta^{15}N$ dissolved organic N from the gyre surface. However, high- $\delta^{15}$ N dissolved organic N export cannot account for the regional increase in thermocline N\*. We thus rule out partial nitrate



**Fig. 2 Zonal trends in physical and biogeochemical properties across the tropical South Atlantic.** Gridded section plots across the CoFeMUG (30°W to 12°E; **a-d**) and MET131 (6°S to 15.1°S; **f-h**) transects of **a**, **f** conservative temperature in °C, where the red contour denotes the 12 °C isotherm, the shoaling of which between 7.5°W and 12.2°E marks the approximate zonal extent of the Angola Gyre<sup>40</sup>; **b**, **g** apparent oxygen utilization (AOU) in  $\mu$ M; **c** dissolved iron concentrations in nM<sup>34</sup> overlaid by N\* contours (black) in  $\mu$ M; and **d**, **h** nitrate  $\delta^{15}$ N in %<sub>c</sub> versus N<sub>2</sub> in the air (CoFeMUG data from ref. <sup>14</sup>). **e**, **i** shows a recent modelled water column-integrated N<sub>2</sub> fixation rate along 11°S (**e**) and 11°E (**i**) in mmol N.m<sup>-2</sup>.a<sup>-113</sup>, with the western basin values in blue and the Angola Gyre values in red. Black (grey) contours on **a**, **b**, **d**, **f**, **g**, **h** (**e** and **h**) are potential density surfaces indicating tropical water masses, where TSW is Tropical Surface Water, TW is Thermocline Water, SAMW is Subantarctic Mode Water and IW is Intermediate Water (SI.2). Small grey circles in **c**, **d** and **h** indicate the sampling resolution.

assimilation and dissolved organic N export as explanations for the low  $\delta^{15}N$  of Angola Gyre thermocline nitrate.

Atmospheric deposition can introduce exogenous N to the surface ocean with a  $\delta^{15}N < 0\%^{51}$  and an N:P ratio >1000:1<sup>52</sup>, potentially explaining both the low- $\delta^{15}N$  nitrate and high N\* of the Angola Gyre thermocline. However, estimates of the N deposition flux to the South Atlantic are orders of magnitude too low to account for our observations<sup>52</sup> (SI.5). We thus conclude that the exogenous N in the Angola Gyre thermocline derives from N<sub>2</sub> fixation.

Origin of the  $N_2$  fixation signal in the Angola Gyre thermocline. The low  $\delta^{15}N$  of Angola Gyre thermocline nitrate could

originate, at least partly, in the tropical North Atlantic basin where N<sub>2</sub> fixation rates are elevated<sup>14,17,28,49</sup>, and be transported into the gyre by equatorial feeder currents (Fig. 1). We address this possibility by estimating the concentration-weighted influx of nitrate  $\delta^{15}N$  by the three Angola Gyre feeder currents: the Equatorial Undercurrent, the South Equatorial Undercurrent, and the South Equatorial Countercurrent<sup>40</sup>. We estimate the  $\delta^{15}N$  of the Equatorial Undercurrent, South Equatorial Undercurrent, and South Equatorial Countercurrent end-members to be 5.7‰, 6.0‰, and 6.3‰, respectively ('Methods'; SI.6). The relative contribution of each current to Angola Gyre thermocline nitrate based on transport volumes<sup>42</sup> yields a mean flux-weighted nitrate- $\delta^{15}N$  of 6.1±0.1‰. While lower than the  $\delta^{15}N$  of underlying SAMW (6.4‰), this end-member is not low enough



**Fig. 3 Characteristics of tropical South Atlantic nitrate.** Density profiles of **a** nitrate concentrations in  $\mu$ M; **b** nitrate  $\delta^{15}$ N in % versus N<sub>2</sub> in air; **c** N<sup>\*</sup> (=  $[NO_3^-] - 15.5 \times [PO_4^{3-}]$ ; SI.1) in  $\mu$ M; and **d** the depth-specific concentration of newly-fixed nitrate in  $\mu$ M in the Angola Gyre. In **a**-**c**, warm (cool) coloured profiles represent stations from the Angola Gyre (tropical western basin). Round markers show data from the CoFeMUG transect while triangle markers represent data from the MET131 transect. The legend provides station longitude. The grey shading on **a**-**d** indicates the density range of TW. In c, the dashed (solid) black line indicates the mean gridded N<sup>\*</sup> for the Angola Gyre (western basin) stations; only CoFeMUG data are shown here because of the higher depth resolution of this sample set. **d** shows the concentration of newly-fixed nitrate in the Angola Gyre, computed by multiplying the depth-specific fraction of newly fixed nitrate ( $f_{depth_specific}$ ; equation S3b) by the corresponding nitrate concentration at each depth. To calculate  $f_{depth_specific}$ , a nitrate  $\delta^{18}$ O-based correction is applied to the  $\delta^{15}$ N data to remove the phytoplankton nitrate assimilation signal from Tropical Surface Water (S1.7). Light grey data show results from scenario 1 while dark grey data show scenario 2 (see text for details). The mean of each scenario is indicated by the bold line of corresponding colour.

to decrease Angola Gyre thermocline nitrate  $\delta^{15}$ N to 5.7‰ (nor to the observed minimum of 4.6‰). Thus, some amount of the low- $\delta^{15}$ N signal must derive from local N<sub>2</sub> fixation.

Our conclusion that N<sub>2</sub> fixation occurs in the Angola Gyre is further supported by the slightly lower  $\delta^{15}$ N of nitrate at the depth of the AOU maximum compared to that of core SAMW (6.2‰ versus 6.4‰; Fig. 2b, g, d, h). Given that the provenance of the high AOU signal is in situ primary production<sup>40</sup>, OM remineralized at the AOU maximum is lower in  $\delta^{15}$ N than SAMW nitrate, consistent with its low  $\delta^{15}$ N deriving from N<sub>2</sub> fixation in Angola Gyre surface waters. Additional support comes from direct rate measurements made during the CoFeMUG cruise<sup>53</sup> and other rate data from near the Angola Gyre<sup>15,54,55</sup>, which indicate N<sub>2</sub> fixation rates ranging from undetectable to ~170 µmol N.m<sup>-2</sup>.d<sup>-1</sup>. Regional observations of diazotrophs (*Trichodesmium* spp. being the most abundant) and diatoms hosting N<sub>2</sub>-fixing symbionts (*Rhizosolenia* and *Chaetoceros* spp.) further substantiate the occurrence of N<sub>2</sub> fixation in the Angola Gyre<sup>53,54,56,57</sup>.

Quantifying the N<sub>2</sub> fixation rate in the Angola Gyre. Using our nitrate  $\delta^{15}$ N data, we estimate the annual rate of N<sub>2</sub> fixation in the Angola Gyre, considering two scenarios: (1) the low- $\delta^{15}$ N of thermocline nitrate is generated solely from in situ N<sub>2</sub> fixation and superimposed on SAMW nitrate, and (2) local N<sub>2</sub> fixation contributes the fraction of low- $\delta^{15}$ N thermocline nitrate that is not supplied by the equatorial feeder currents. For both scenarios, we estimate the fraction (*f*) of newly fixed nitrate in the Angola Gyre as:

$$f = \frac{\delta^{15} N_{AGmean} - \delta^{15} N_{scenario\_mean}}{\delta^{15} N_{N_2 fix} - \delta^{15} N_{scenario\_mean}}$$
(1)

 $δ^{15}N_{AGmean}$  is the mean concentration-weighted  $δ^{15}N$  of Angola Gyre nitrate for the upper water column ( $σ_θ ≤ 27.0$ ), estimated to be  $5.7 \pm 0.4\%$  (5.5-5.9%; 5th-95th percentiles; Methods),  $δ^{15}N_{N2fix}$  is the  $δ^{15}N$  of nitrate regenerated from diazotrophic OM,  $-1\%^{30,31}$ , and  $δ^{15}N_{scenario}$  mean is the scenario-specific mean concentration-weighted  $δ^{15}N$  of nitrate supplied to the Angola Gyre thermocline ( $6.4 \pm 0.1\%$  and  $6.1 \pm 0.1\%$  for scenarios 1 and 2, respectively).

For scenario 1, f = 9% (6–12%; 5th–95th percentiles) and for scenario 2, f = 6% (2–9%; 5th–95th percentiles). On average, therefore, 6–9% (associated with scenarios 2 and 1, respectively) of the nitrate above SAMW originates from local N<sub>2</sub> fixation. This mass-balance method neglects particulate organic N, which is reasonable given that this pool contributes negligibly to total N, accounting for <1% of the fixed N reservoir above  $\sigma_{\theta} = 27.0$ . Our approach is validated by the concentration-weighted  $\delta^{15}$ N that it yields for nitrate above  $\sigma_{\theta} = 27.0$  in the western tropical basin of  $6.3 \pm 0.1\%$  (6.2–6.4%; 5th–95th percentiles), indistinguishable from that of SAMW (6.2 ± 0.1‰). This correspondence further confirms that N<sub>2</sub> fixation is negligible in the tropical waters west of the Angola Gyre.

To visualize the depth distribution of newly-fixed nitrate in the Angola Gyre, we calculate  $f_{depth\_specific}$  (i.e., the fraction of the

nitrate pool at each depth that is newly fixed) for each sample above  $\sigma_{\theta} = 27.0$ , after removing the signal of phytoplankton assimilation from nitrate  $\delta^{15}$ N using the coincident nitrate  $\delta^{18}$ O data ('Methods'; SI.7). This exercise reveals that a substantial fraction of Tropical Surface Water nitrate is newly-fixed, which is not apparent from the nitrate  $\delta^{15}N$  profiles because fractionation during nitrate assimilation overprints the isotopic signal of newly nitrified N (Fig. 3b, d). It is not appropriate, however, to integrate these depth-specific estimates to yield a measure of the newlyfixed N inventory. This is due to the likelihood of 'doublecounting'—that is, low- $\delta^{15}N$  nitrate assimilated in the surface is returned to the thermocline when the resulting OM is exported and remineralized, such that at the scale of the water column, correction of shallow nitrate- $\delta^{15}N$  for nitrate assimilation is unnecessary<sup>28</sup>. Instead, the mass-balance approach represented by Eq. 1 is most appropriate for estimating the water column burden of newly fixed N.

Multiplying f (Eq. 1) by the mean Angola Gyre nitrate concentration for  $\sigma_{\theta} \leq 27.0$  indicates that local N<sub>2</sub> fixation supplies 313–641 mmol N.m<sup>-2</sup> to the thermocline nitrate reservoir. The residence time of thermocline waters in the Angola Gyre is 4.4–8.5 years<sup>40</sup> and its areal extent is  $2.6 \times 10^{12}$  m<sup>2</sup>. These constraints yield an N<sub>2</sub> fixation rate of 2.8-5.4 Tg N.a<sup>-1</sup> (mean of  $4.1 \pm 2.8$  Tg N.a<sup>-1</sup>) for scenario 1 and 1.4-2.6 Tg N.a<sup>-1</sup> (mean of  $2.0 \pm 3.0$  Tg N.a<sup>-1</sup>) for scenario 2, considering both extrema for residence time ('Methods'; SI.8). Our estimates are equivalent to daily rates of  $102-390 \,\mu$ mol N.m<sup>-2</sup>.d<sup>-1</sup>, comparable to incubation-based N<sub>2</sub> fixation rates measured near the Angola Gyre<sup>15,53–55</sup> (SI.9).

The Angola Gyre is a hotspot for N<sub>2</sub> fixation in the South Atlantic. Our N<sub>2</sub> fixation rate estimates imply that the Angola Gyre accounts for 28–108% of the highly uncertain mean South Atlantic N<sub>2</sub> fixation rate of  $5.0 \pm 3.3$  Tg N.a<sup>-1</sup> (Table 1). Since the latter derives from few observations that are sparsely distributed across the basin, it would be unwise to conclude that the Angola Gyre makes a disproportionate contribution to South Atlantic N<sub>2</sub> fixation. Nonetheless, since the Angola Gyre occupies only ~10% of the South Atlantic by area, our data indicate that it is a hotspot for N<sub>2</sub> fixation. Conditions particular to the Angola Gyre must render this feature favourable for N<sub>2</sub> fixation.

The phosphate-bearing, nitrate-deplete surface waters of the South Atlantic receive a limited supply of aeolian iron<sup>20</sup>, a condition proposed to limit  $N_2$  fixation across the basin<sup>9,13,14,17,38</sup>. The non-aeolian iron supply to the Angola Gyre must thus be substantial. The Congo-shelf-zone near the Angola Gyre has been reported to supply a substantive quantity of iron to the marginal ocean, equivalent to  $40 \pm 15\%$  of the entire South Atlantic aeolian iron flux<sup>58</sup>, and the elevated iron concentrations observed in the vicinity of the Angola Gyre (Fig. 2c) are evidence of this sedimentary iron supply. We hypothesize that the subsurface iron plume, which is trapped by the retentive regional circulation of the Angola Gyre through repeated cycles of upward supply, export, and remineralization<sup>34</sup>, relieves diazotrophs in the overlying surface waters of iron limitation.

We thus conclude that  $N_2$  fixation occurs in the Angola Gyre because of the overlapping biogeography of excess P and bioavailable iron. While the iron supply to the Angola Gyre is local, originating from the nearby low-oxygen margin sediments, the excess P derives from both regional and remote N loss. The apparent importance of margin-derived iron echoes a growing body of work showing that sedimentary iron sources are important to the global ocean iron budget<sup>59–62</sup>. The notion that both iron and excess P together limit N<sub>2</sub> fixation at the regional scale in the South Atlantic is compatible with previous work showing negligible rates of  $N_2$  fixation in the western tropical basin where measurements were made in P-bearing but iron-limited waters<sup>9,17,22</sup>.

The possibility of additional N<sub>2</sub> fixation hotspots. We hypothesize that N<sub>2</sub> fixation hotspots are inherent to ocean regions characterized by biogeochemical conditions analogous to those encountered in the Angola Gyre, possibly enhanced by retentive circulation features (SI.10). In the subtropical South Atlantic where the Brazil Current recirculation forms a subgyre adjacent to the continental shelf<sup>41</sup> (Fig. 1), an N<sub>2</sub> fixation hotspot should result as waters bearing a remote (i.e., Southern Ocean-derived) and regional (i.e., NBUS-derived) supply of excess P<sup>6</sup> (Fig. 1) encounter a local supply of sediment-derived iron<sup>62</sup>. Observations of diazotroph blooms<sup>63</sup> and low- $\delta^{15}$ N thermocline nitrate<sup>64</sup> in the Brazil subgyre validate this prediction. In addition, the cyclonic Guinea Dome in the eastern tropical North Atlantic receives excess P from local upwelling of P-rich subsurface waters<sup>8</sup> and iron from dust deposition, possibly augmented by an iron flux from the nearby low-oxygen margin sediments<sup>60</sup>. Direct  $N_2$  fixation rate measurements<sup>15,16</sup>, the presence of diazotrophs<sup>16,65</sup>, and low- $\delta^{15}$ N thermocline nitrate<sup>66</sup> in the vicinity of the Guinea Dome are consistent with this region also being an N<sub>2</sub> fixation hotspot.

The surface waters above or just offshore of the ocean's major suboxic zones would, in general, appear well-suited to host hotspots of N<sub>2</sub> fixation. Denitrification in both the shallow subsurface water column and the adjacent sediments generates an N deficit (i.e., a P excess)<sup>6</sup>. At the same time, the adjacency of these zones to low-oxygen margin sediments, coupled with water column suboxia overlying some of the sediments, supplies iron to the water column<sup>59,67,68</sup>. These regions are also characterized by upwelling, which would transport both the excess P and the iron into the surface mixed layer. In these surface waters or just offshore, phytoplankton assimilation leaves nitrate-free but Pand iron-bearing surface waters<sup>6,67–69</sup>, a seemingly ideal recipe for N<sub>2</sub> fixation.

However, the existing data are ambiguous as to the rates of N<sub>2</sub> fixation above and nearby the suboxic zones. Observations from the Arabian Sea support elevated N<sub>2</sub> fixation<sup>70-72</sup> while incubation-based measurements in the eastern tropical Pacific suggest negligible to extremely high rates<sup>38,73-78</sup>. Notably, incubation experiments suggest that N2 fixation occurs in the Costa Rica Dome<sup>75</sup>, a retentive feature embedded within the eastern tropical Pacific suboxic zone that is characterized by similar hydrography to the Angola Gyre. In the water column of all the suboxic zones, a decrease in nitrate  $\delta^{15}N$  is observed above the very high nitrate  $\delta^{15}$ N associated with denitrification<sup>79–82</sup>; this upward  $\delta^{15}$ N decline has been interpreted as evidence of local N<sub>2</sub> fixation<sup>79</sup>. Additionally coupled nitrate  $\delta^{15}N$  and  $\delta^{18}O$ measurements from the eastern tropical North Pacific indicated a lower nitrate  $\delta^{15}$ N than is expected from nitrate  $\delta^{18}$ O in the upper portion of the suboxic zone, also consistent with a role for  $N_2$  fixation<sup>80</sup>. However, N cycling processes occurring within suboxic waters (e.g., nitrite oxidation) provide an alternative explanation for the observed  $\delta^{15}$ N- $\delta^{18}$ O decoupling<sup>80-84</sup>, complicating the use of the nitrate isotopes to evaluate N<sub>2</sub> fixation above and near the major suboxic zones. Importantly, the Angola Gyre is not complicated by the same suboxic zone N cycling processes, allowing us to pinpoint the role of N<sub>2</sub> fixation in this system. Our findings for the Angola Gyre thus suggest that the nitrate  $\delta^{15}$ N decline in the shallow subsurface of the suboxic zones may have a similar origin. The nitrate isotope data of the major ocean suboxic zones should be revisited with this new perspective.

Implications of regional N2 fixation hotspots. N2 fixation has been proposed to be spatially and/or quantitatively coupled to N loss at the basin scale<sup>6,10,85</sup>. Our estimate of N<sub>2</sub> fixation in the Angola Gyre is notably comparable to N loss from the NBUS  $(1.4-2.5 \text{ Tg N.a}^{-1})^{11,86}$ . In this regard, the generation and advection of excess-P waters from the NBUS have been hypothesized to fuel N<sub>2</sub> fixation in the vicinity of the Angola Gyre<sup>10</sup>, prompting the suggestion that N sources and sinks in the southeastern Atlantic are coupled. However, surface P\* is perennially elevated across the (sub)tropical South Atlantic, including in Angola Gyre and Brazil subgyre surface waters<sup>6,9</sup> (Fig. 1). Accordingly, it is not clear that the additional flux of excess P from the NBUS, by itself, stimulates N<sub>2</sub> fixation in the Angola Gyre. Rather, N<sub>2</sub> fixation would likely occur in the Angola Gyre regardless of whether it receives excess P from the NBUS, provided that the iron supply is sustained. That is, the greatest influence of the NBUS on N<sub>2</sub> fixation in the Angola Gyre may be through its augmentation of the iron supply. In summary, our findings argue that a P-driven coupling of N<sub>2</sub> fixation to denitrification is contingent on a supply of iron, which is most easily achieved along coastal suboxic zones<sup>59</sup>.

Because the South Atlantic hosts a basin-wide surface P excess, a future increase in the iron supply to the south-eastern Atlantic due to expanding low-oxygen margins<sup>34,62,87</sup> and/or a climate change-driven increase in aeolian deposition<sup>88</sup> could enhance  $N_2$  fixation in and beyond the Angola Gyre. Consequently, the meridional flux of excess P across the Atlantic could decrease, engendering a shift in the dominance of  $N_2$  fixation from the North to the South Atlantic.

Lessons for estimating basin-scale N2 fixation rates. Our observations contradict diagnoses of the regional distribution of N<sub>2</sub> fixation in the South Atlantic computed from nutrient climatologies and velocity fields from ocean general circulation models<sup>6,13</sup> (e.g., P\* convergence). This inconsistency highlights the current inadequacy of nutrient fields to deliver regional representations of N2 fixation. Modelled distributions suggest that N<sub>2</sub> fixation predominantly occurs in the western and central tropical South Atlantic at a rate of 7.5-8.3 Tg N.a<sup>-1</sup>, yet the nitrate- $\delta^{15}$ N data from these longitudes bear no evidence of the remineralization of low- $\delta^{15}$ N, diazotroph-derived OM (Figs. 2d and 3b). Thus, while the model diagnoses offer a powerful framework from which to estimate basin-scale N2 fixation rates and conceptualize the global drivers thereof<sup>6</sup>, some of the fundamental assumptions inherent to this approach can lead to erroneous diagnoses, potentially underpinning the discrepancy between our observations and existing model solutions. First, coarse resolution (2-4°) general circulation models used to simulate current velocities and transports typically resolve only gross circulation features. Although this limitation is widely acknowledged<sup>6,12,13</sup>, the implications for estimates of N<sub>2</sub> fixation rates and distributions may be important, particularly at the regional scale. In particular, such general circulation models are unable to reliably simulate ocean-margin environments<sup>12,37,61</sup>. Second, the available South Atlantic nutrient climatologies include few data, particularly in the central basin, such that the choice of data set can considerably alter the modelled regional distribution of N<sub>2</sub> fixation<sup>6</sup>. Third, diagnoses of N<sub>2</sub> fixation from inverse models are highly sensitive to the N:P ratios of plankton biomass and exported OM. These properties are difficult to measure and are thought to be dynamic and sensitive to environmental conditions<sup>25–27,89</sup>. Elemental ratios of OM are thus typically parameterized<sup>6,13</sup> and may not accurately capture the plasticity of OM stoichiometry. Fourth, to parameterize dissolved OM in models, its production and degradation rates also have to be prescribed. However, few measurements of such rates exist particularly with respect to dissolved organic phosphorus, which diazotrophs can assimilate when ambient phosphate concentrations are low<sup>90,91</sup>—resulting in modelled dissolved OM dynamics that are poorly constrained and potentially erroneous<sup>92</sup>. Finally, the analytical limit of quantification for  $[PO_4^{3-}]$  and  $[NO_3^{-}]$  is such that the propagated error on P\* is  $\geq 0.1 \,\mu M^{93}$  (SI.11). Interpretations of  $\leq 0.1 \,\mu M$  differences in P\* may thus inaccurately diagnose the rate and/or distribution of N<sub>2</sub> fixation<sup>94</sup>.

Pending enhanced observational coverage and improved model simulations of margin environments, convergent  $N_2$  fixation distributions for the South Atlantic and other dynamic oceanmargin regions will remain elusive. Our work highlights the utility of nitrate isotope ratios for identifying and estimating  $N_2$ fixation, and underscores the importance of regional observations, which are necessary for understanding the regional controls on  $N_2$  fixation that ultimately determine its global distribution.

#### Materials and methods

Sample and data provenance. The CoFeMUG nutrient and nitrate isotope data were first published in refs. <sup>34,14</sup>, respectively. Samples from the CoFeMUG cruise were collected onboard the R/V *Knorr* in November and December 2007 (Fig. 1). Seawater samples were measured for nitrate isotopes at the Woods Hole Oceanographic Institution using the denitrifier method<sup>95,96</sup>. Nitrate concentration and isotope data were first published from the CLIVAR A16N cruise in refs. <sup>49,66</sup>. The samples were collected onboard the NOAA R/V *Ronald H. Brown* between August and October 2013 as part of the GO-SHIP and CLIVAR program. The shipboard-ADCP data from this cruise are provided in Firing & Hummon (2010)<sup>97</sup>. The MET131 cruise data have not been previously published. This cruise was undertaken onboard the R/V *Meteor* in October 2016, with nine stations sampled between 6°S-15°S and 7°E-12°E. Twenty-four 12 L Niskin bottles attached to a Sea-Bird rosette with conductivity-temperature-depth (CTD) and oxygen sensors were remotely fired over the upper 1000 m to collect seawater samples for nutrient and nitrate isotope analysis. Samples were collected in thoroughly rinsed 30 ml HDPE bottles, filtered (0.4 µm), and immediately frozen at -20°C until analysis.

#### Analysis of MET131 cruise nutrient concentrations and nitrate isotopes.

Nitrate and phosphate concentrations ([NO<sub>3</sub><sup>-</sup>], [PO<sub>4</sub><sup>3-</sup>]) and the dual isotopes of nitrate were analysed at the Helmholtz–Zentrum Hereon in Germany. The concentrations of PO<sub>4</sub><sup>3-</sup>, nitrite ([NO<sub>2</sub><sup>-</sup>]) and nitrate plus nitrite ([NO<sub>3</sub><sup>-</sup>] + [NO<sub>2</sub><sup>-</sup>]) were measured colourimetrically using an automated continuous flow system (AA3, Seal Analytical, Germany) with a detection limit of 0.01  $\mu$ M<sup>98</sup>; [NO<sub>3</sub><sup>-</sup>] was then determined by subtraction. The dual isotopes of nitrate, following nitrite removal<sup>99</sup>, were measured using the denitrifier method<sup>95,96</sup> whereby nitrate is quantitatively converted to nitrous oxide gas (N<sub>2</sub>O) by denitrifying bacteria that lack an N<sub>2</sub>O reductase enzyme. The N and O isotope ratios of the N<sub>2</sub>O were measured using a Thermo Delta Plus XP isotope ratio mass spectrometer in-line with a GasBench II. The international reference materials, IAEA-N3<sup>100</sup> and USGS 34<sup>101</sup>, were included in each run to enable post-run calibration of the N<sub>2</sub>O measurements to N<sub>2</sub> in the air ( $\delta$ <sup>15</sup>N and  $\delta$ <sup>18</sup>O was <0.2‰ and <0.4‰, respectively.

**Data gridding and interpolation**. The CoFeMUG and MET131 data were gridded prior to the generation of section plots and the calculation of mean water-column values (Figs. 2 and 3c, d). The *z*-grid (pressure) bins focused on the upper 1000 m. Python 3.6 Xarray linear method was used as the interpolation scheme.

Nitrate  $\delta^{15}$ N transported into the Angola Gyre. Cruise A16N sampled both the Equatorial Undercurrent (EUC) and South Equatorial Undercurrent (SEUC) at 0.5°S and 3.6°S, respectively, while the CoFeMUG cruise sampled the South Equatorial Countercurrent (SECC) at 27.5–30°E (Fig. 1 and SI.6). The mean [NO<sub>3</sub><sup>-</sup>]-weighted  $\delta^{15}$ N for the EUC, SEUC, and SECC were estimated to be 5.7‰, 6.0‰ and 6.3‰, respectively. To appropriately weight the nitrate flux of each feeder current, two transport volume scenarios were considered due to the uncertainty in the contribution of the EUC to the Angola Gyre<sup>42</sup>. Scenario A considered the net transport across the northern limb of the Angola Gyre, of 8 Sv, to be apportioned between the SEUC (59%) and the SECC (41%), with no contribution from the EUC<sup>42</sup>. Scenario B considered the net transport across the eastern limb of the Angola Gyre, of 11 Sv, to be apportioned between the SEUC (43%), the SECC (30%), and the EUC (27%)<sup>42</sup>. The mean flux-weighted nitrate  $\delta^{15}$ N resulting from both transport scenarios was  $6.1 \pm 0.1‰$ ; this value is used in scenario 2 throughout the study.

Assimilation-corrected, depth-specific fraction of newly fixed nitrate. At the base of the euphotic zone, co-occurring nitrate assimilation and nitrification of

low- $\delta^{15}$ N OM deriving from N<sub>2</sub> fixation raises the  $\delta^{15}$ N of nitrate, potentially yielding an underestimation of the depth-specific fraction of newly-fixed nitrate inferred from the  $\delta^{15}$ N data (SI.7). During nitrate assimilation, nitrate  $\delta^{18}$ O and  $\delta^{15}$ N rise in a 1:1 ratio as nitrate consumption proceeds<sup>45,46</sup> while during nitrification, nitrate  $\delta^{18}$ O and  $\delta^{15}$ N become decoupled<sup>48,102,103</sup>. The nitrate  $\delta^{18}$ O date thus reveal (1) the depth at which nitrate assimilation begins to alter to isotopic composition of the nitrate pool and (2) the magnitude of the nitrate assimilation signal, two insights that are not apparent from the nitrate  $\delta^{15}$ N data. Assuming that isotopic fractionation associated with nitrate assimilation similarly alters nitrate  $\delta^{15}$ N and  $\delta^{18}$ O as nitrate consumption proceeds<sup>45,46</sup>, we quantified the nitrate assimilation signal at each depth in the water column and subtracted it from the nitrate  $\delta^{15}$ N data, beginning at the depth where nitrate  $\delta^{18}$ O starts to rise above its mean deep-ocean value. Following the removal of the nitrate assimilation signal, the fraction of newly fixed nitrate at each depth was calculated.

**Statistical analyses**. To estimate uncertainty on the mean  $\delta^{15}$ N of Angola Gyre and western basin nitrate ( $\sigma_0 \le 27.0$ ), we calculated the 95% confidence interval associated with a *t*-distribution (for n < 30). The same method was adopted to estimate uncertainty on f (Eq. 1) for scenarios 1 and 2. To characterize the uncertainty associated with the Angola Gyre N<sub>2</sub> fixation rates, we ran a Monte Carlo simulation of 100k steps assuming a normal probability distribution. We report the standard deviation of the output as the error associated with the rates.

#### **Data availability**

Data from the CoFeMUG and A16N cruises can be found in the BCO-DMO database (http://www.bco-dmo.org/dataset/630246/data and http://www.bcodmo.org/dataset/699160/data, respectively). Data from the MET131 cruise can be found in the PANGAEA database (https://doi.org/10.1594/PANGAEA.929671).

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#### Author contributions

T.M., M.R.M. and D.M. made the nitrate isotope measurements. M.A.S. and A.E.N. collected and measured the CoFeMUG cruise iron samples. K.L.C., K.D. and D.M.S. supported the nitrate isotope analyses in their laboratories. D.M., D.M.S. and K.-C.E. provided data and samples. T.M., J.G., D.M.S. and S.E.F. led the data interpretation. All authors contributed to the paper, led by T.M., with major contributions from J.G., D.M.S. and S.E.F. All authors approved the final submission.

#### **Competing interests**

The authors declare no competing interests.

#### **Additional information**

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