# DOI: http://dx.doi.org/10.18203/2320-6012.ijrms20203668

# **Original Research Article**

# Vepris nobilis plant: a potential source of anticancer agents

# Carolyne Chepkirui<sup>1\*</sup>, Richard Kagia<sup>2</sup>

<sup>1</sup>Department of Physical and Biological Science, Kabarak University, Kenya

Received: 09 July 2020 Accepted: 10 August 2020

## \*Correspondence: Carolyne Chepkirui,

E-mail: carochep2005@gmail.com

**Copyright:** © the author(s), publisher and licensee Medip Academy. This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### **ABSTRACT**

**Background:** Cancer is one of the major causes of death worldwide. Current cancer therapy is costly, it has poor therapeutic outcomes and many side effects. Therefore, new medications are needed. Plants have been used as sources of anticancer drugs. *Vepris* species have anticancer properties. The purpose of this study is to assess *Vepris nobilis*, a plant found in Kenya as a potential source of anticancer drugs.

**Methods:** The dichloromethane/methanol (CH<sub>2</sub>Cl<sub>2</sub>/MeOH) 1:1 extract of the stem bark of *Vepris nobilis* led to the isolation of an alkaloid named, 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b]quinolone. SwissADME online tool was used to assess the compound's pharmacokinetic parameters. Pass online tool identified potential targets while protox server described the toxicity of the compound. Chimera and Avogadro softwares were used for molecular docking studies.

**Results:** In-silico pharmacokinetic studies, showed that the isolated compound complied with Lipinski rule of five, it showed high gastrointestinal activity, and it also inhibits cytochrome P450 (CYP) isoforms 1A2, 2C9 and 2C19. In toxicity studies the compound was relatively safe with a predicted median lethal dose (LD50) of 1600 mg/kg, apart from potential immunotoxicity and mutagenicity. Molecular docking studies demonstrated that, the compound has potential anticancer activity, it interacted with deoxyribonucleic acid (DNA) topoisomerase I in an almost similar manner to camptothecin though it had less binding potential.

**Conclusions:** 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy) furo[2,3-b]quinolone derived from *Vepris nobilis* is a potential drug for the management of cancer which can be administered orally.

Keywords: Cancer, In-silico, Molecular docking, Pharmacokinetic, Vepris nobilis

## **INTRODUCTION**

Cancer is a group of diseases due to abnormal growth of cells, which comprise a complex environment of different types of malignant cells which support its growth and development.<sup>1,2</sup> It is among the leading cause of death worldwide. Globally, 9.8 million deaths were reported in 2018.<sup>3-5</sup> The number of cancer deaths are expected to rise to 13 million by 2030.<sup>6</sup>

In Kenya, cancer is a major health problem, it is a third leading cause of mortality, responsible for 7% of the annual deaths.<sup>7</sup> The current treatment methods are facing

many challenges; they are only effective at early stages, very expensive and resistance to the chemotherapeutic drugs has occurred in some cases.<sup>8-13</sup> In addition, the severe side effects are not effectively mitigated. Hence there is an urgent need for continuous research to find alternative agents to fight against cancer.

Plants have been used as sources of anticancer drugs. For instance, camptothecin, an alkaloid extracted from *Camptotheca acuminata* is a drug used in cancer treatment by Thomas et al. Other anticancer drugs derived from plants include but not limited to vincristine, vinblastine from *Catharanthus roseus* and taxol from *Taxus* 

<sup>&</sup>lt;sup>2</sup>Department of Pharmacology and Pharmacognosy, Kabarak University, Kenya

*brevifolia*. <sup>16</sup> The genus *Vepris* is a rich source of furoquinoline and acridone alkaloids, which have been reported to have anticancer antiplasmodial, antimicrobial and antioxidant activities. <sup>17-20</sup> Therefore, it was worthwhile to isolate the secondary metabolites of this genus.

Drug discovery is very expensive and requires a lot of time. However, *in silico* drug discovery is cheaper and not as time-consuming as conventional methods of drug discovery like *in vitro* and *in vivo* studies. *In silico* drug discovery involves use of softwares and databases to assist in discovery and development of drugs. It facilitates predictions of how ligands and drugs may interact with various targets or receptors.

#### **METHODS**

#### General

The <sup>1</sup>H (200, 600 MHz) and <sup>13</sup>C (50, 150 MHz) were acquired using Varian-Mercury and Bruker instrument using residual solvent signals as reference. Column chromatography was on normal silica gel 60G (Merck, 70-230 mesh) and Sephadex LH-20. Analytical thin layer chromatography (TLC) using silica gel 60 F254 (Merck) pre-coated plates were used to monitor the separation of compounds. For qualitative work, the TLC plates were visualized under ultraviolet (254 and 366 nm) light, exposure to iodine (I<sup>2</sup>) vapor or spraying with Dragendorff reagent.

#### Plant material

The stem bark of *Vepris nobilis* was collected from Kakamega forest, Kenya, in July 2010. The plant was identified at the University Herbarium, School of Biological Sciences, University of Nairobi.

## Extraction and isolation

The dried and ground stem bark (3.2 kg) of *Vepris nobilis* was extracted thrice using dichloromethane/methanol (CH<sub>2</sub>Cl<sub>2</sub>/MeOH) 1:1 by cold percolation. The crude extract (80 g) was subjected to column chromatography on silica gel (600 g). Gradient elution with n-hexane containing increasing amount of ethyl acetate and finally washed with MeOH afforded twenty major fractions (labeled A-T). Fraction M (eluted with 55% CH2Cl2 in n-hexane) was used to obtain compound 1 (24 mg) as colourless solids, after further purification on a silica gel (50 g) column with n-hexane containing increasing amounts of CH<sub>2</sub>Cl<sub>2</sub> (1 to 99% v/v).

# In-silico pharmacokinetic analysis

SwissADME online tool (http://www.swissadme.ch/) was used to predict the pharmacokinetic profile of 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo [2,3 b]quinolone.<sup>23</sup> Canonical SMILES of the compound were uploaded to the SwissADME tool which predicts and

evaluates medicinal chemistry likeness, drug-likeness and pharmacokinetic properties.

#### In-silico toxicity prediction

Canonical SMILES of 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b]quinoline were uploaded to the ProTox server (http://tox.charite.de/protox\_II/) which was used to predict the toxicity profile including hepatotoxicity, cytotoxicity, mutagenicity, immunotoxicity, carcinogenicity, toxicological pathways and toxicity targets.<sup>24</sup>

#### Determination of potential targets

Using the pass online website, potential targets for 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b]quinolone were identified.<sup>25</sup>

#### Molecular docking

4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo [2,3-b]quinolone was drawn using pubchem sketcher. A molfile of the compound was downloaded and converted to 3-D using Avogadro software.<sup>26</sup> Optimisation of the chemical structure to the most stable conformation was done using the Avogadro software. Based on the pass online results, the compound had antineoplasic activity and targeted beta-glucurunidase. Anticancer that are also affected by beta glucuronidase are the topoisomerase inhibitors. Therefore, DNA topoisomerase (PDB ID:1A36) was downloaded from the protein databank. Residues were removed from the DNA topoisomerase enzyme using the chimera software.<sup>27</sup> Molecular docking between DNA topoisomerase enzyme and 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl) oxy)furo[2,3-b]quinolone was done using autodock vina feature in chimera software. The binding energies were compared with the binding between campotothecin and DNA topoisomerase. Ligandreceptor interactions were observed using Discovery studio software.

### **RESULTS**

Compound 1 was isolated as yellow oil with retention factor (Rf) value of 0.5 (1% MeOH in CH<sub>2</sub>Cl<sub>2</sub>). The spot turned orange when sprayed with Dragendorff reagent which is an indication of an alkaloid. The <sup>1</sup>H NMR spectrum revealed the presence of a pair of AB doublets corresponding to the two furan protons (H-2 and H-3) of furoquinoline alkaloids along with a downfield shifted methoxyl which is also characteristic of a methoxyl group at C-4 for furoquinoline alkaloids.<sup>28</sup> The <sup>1</sup>H NMR spectrum further showed a second methoxyl group resonating at δH 4.02, with corresponding carbon resonating at δC 56.1.

The presence of two singlet aromatic protons at 7.52 (H-5,  $\delta$ C 104.6) and 7.32 (H-8,  $\delta$ C 101.2) is consistent with C-6 and C-7 substituted c ring. One of these substituents being

methoxyl ( $\delta$ H 4.02,  $\delta$ C 56.1) groups and was placed at C-6 based on Nodiff experiment. Irradiation of methoxyl at C-4 showed interaction with H-3 and H-5, and irradiation of the methoxyl group resonating at  $\delta$ H 4.02 ppm (6-OMe) showed interaction with H-5.

The substituent at C-7 is 3-methylbut-1,3-dienyloxy, as evidenced by the presence in the  $^1H$  NMR spectrum of a pair of doublets resonating at  $\delta H$  6.85 (J=12.2 Hz, for H-1'), and 6.32 (J=12.2 Hz, H-2'), terminal methylene protons resonating at  $\delta H$  4.94 ( $^1H$ , Br S) and 4.89 ( $^1H$ , Br S), and a methyl group at  $\delta H$  1.91 for Me-5. The corresponding carbon atoms of this group appeared at  $\delta C$  142.3 (C-1'), 115.3 (C-2'), 114.5 (C-3'), 118.9 (C-4') and 18.9 (C-5'). Therefore this compound was characterized as 7-(3-methylbuta-1,3-dienylloxy)-4, 6-dimethoxyfuro [2,3-b]quinolone.

#### **Pharmacokinetics**

Based on SwissADME, 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b]quinolone has high gastrointestinal activity, it crosses the blood brain barrier and inhibits cytochrome P450 (CYP) isoforms 1A2, 2C9 and 2C19. In terms of Pan-assay interefence compounds (PAINS) alert, it had no alert. The compound has a molecular weight of 311.33 g/mol, partition coefficient (log P) of 3.76, 0 hydrogen bond donors and 5 hydrogen bond acceptors. The bioavailability score was 0.55.

#### **Toxicity**

The predicted median lethal dose (LD50) for 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl) oxy) furo[2,3-b] quinolone was 1600 mg/kg which indicates that the compound is in toxicity class 4 (LD50 between 300 and 2000). Assessment of organ toxicity, toxicity end points, toxicological pathways and toxicity targets indicated that the compound can cause immunotoxicity and mutagenicity.

#### Potential targets

Based on Pass online website, the major potential actions of 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy) furo[2,3-b]quinolone were antineoplastic activity, beta glucuronidase inhibitor and gluconate-2-dehydrogenase inhibitor.

#### Molecular docking

4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo [2,3-b]quinolone bound to DNA topoisomerase I as shown in Figure 1. However, the binding energies were less optimal compared to the binding energies of camptothecin and DNA topoisomerase I. The compound intercalated with the DNA of the topoisomerase enzyme.

The compound interacted with DNA topoisomerase via pication interactions with lysine at position 493, hydrogen

bond with threonine at position 501, alkyl interactions with alanine at position 499, arginine at position 364, lysine at position 493 and position 532. More interactions are shown in Figure 2.

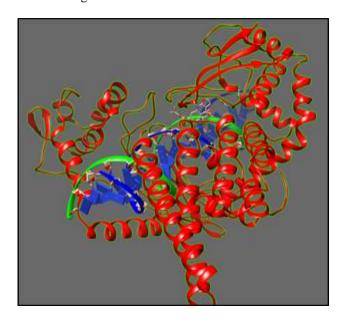


Figure 1: 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl) oxy)furo[2,3-b]quinolone bound to DNA topoisomerase I.

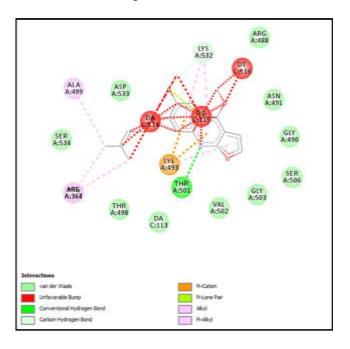


Figure 2: Interactions between DNA topoisomerase I with 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b] quinolone.

## DISCUSSION

4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo [2,3-b]quinolone lacks any PAINS alert and thus is a very good lead compound to be developed to a drug. <sup>29,30</sup> This

compound complies with rules of drug likeness proposed by Lipinski which recommended that a potentially orally active drug has a molecular weight of less than or equal to 500, a log P of less than or equal to 5, less than or equal to 10 hydrogen acceptors and less than or equal to 5 hydrogen bonds. It also complied to Veber's rules on drug likeness which recommended less than or equal to 10 rotatable bonds and less than or equal to 140 angstroms in terms of polar surface area. The compound also has high gastrointestinal activity. This indicates that the compound is a potential drug that can be administered orally.

The compound inhibits cytochrome P450 isoform 1A2 and thus may affect metabolism of caffeine, clozapine, olanzapine, lidocaine, ropivacaine, melatonin, tacrine, tizanidine, triamterene, zolmitriptan and frovatriptan.<sup>33</sup> It also inhibits CYP2C9 which is critical in metabolism of warfarin, phenytoin, tolbutamide, some non-steroidal antiinflammatory drugs, losartan, candesartan, cyclophosphamide, zafirlukast and other drugs.<sup>34</sup> It also inhibits CYP2C19 and thus affects metabolism of carisoprodol, omeprazole, pantoprazole, lansoprazole, moclobemide, diazepam, mephenytoin, mephobarbital and hexobarbital.35 Therefore, this compound has many drugdrug interactions and cost versus benefit analysis should be done for patients with several comorbidities.

This compound is generally safe since it does not affect toxicological pathways, toxicity targets and hepatotoxicity. However, it causes immunotoxocity and mutagenicity. A number of anticancer drugs like doxorubicin, cyclophosphamide, busulphan and mercaptopurine are also mutagenic.<sup>36</sup> This still indicates the potential of this compound as an anticancer agent.

4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo [2,3-b]quinolone intercalates the DNA in a similar manner to camptothecin. It also interacts with arginine amino acid at position 364 similar to camptothecin. However, camptothecin also interacts with aspartate at position 533 and asparagine at position 722 unlike this compound. Therefore, camptothecin has better binding capacity to DNA topoisomerase I compared to 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b]quinolone. However, it can still interact with DNA topoisomerase I and thus has potential in management of cancer.

#### CONCLUSION

4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo [2,3-b]quinolone derived from *Vepris nobilis* is a potential drug for the management of cancer which can be administered orally. However, it has many drug-drug interactions.

#### Recommendations

*In vitro* and *in vivo* studies are needed to test 4,6-dimethoxy-7-((3-methylbuta-1,3-dien-1-yl)oxy)furo[2,3-b] quinolone for management of cancer.

#### **ACKNOWLEDGEMENTS**

Authors would like to acknowledge Professor Abiy Yenesew for the supervision of isolation and characterization of compound 1.

Funding: No funding sources Conflict of interest: None declared Ethical approval: Not required

#### REFERENCES

- 1. Adem FA. Phytochemical analysis of selected plants in the leguminosae and moraceae families for anticancer principles. University Nairobi. 2019;315.
- Parkin A, Man J, Timpson P, Pajic M. Targeting the complexity of Src signalling in the tumour microenvironment of pancreatic cancer: from mechanism to therapy. FEBS J. 2019;286:3510-39.
- 3. Block KI, Gyllenhaal C, Lowe L, Amedei A, Amin ARMR, Amin A, et al. A broad-spectrum integrative design for cancer prevention and therapy. Semin Cancer Biol. 2015;35:276.
- 4. Seebacher NA, Stacy AE, Porter GM, Merlot AM. Clinical development of targeted and immune based anti-cancer therapies. J Exp Clin Cancer Res. 2019;38:156.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. Cancer J Clin. 2018;68:394-424.
- 6. Zarocostas J. Global cancer cases and deaths are set to rise by 70% in next 20 years. Br Med J Online. 2010;340.
- Topazian H, Cira M, Dawsey SM, Kibachio J, Kocholla L, Wangai M, Welch J, et al. Joining forces to overcome cancer: The Kenya cancer research and control stakeholder program. J. Cancer Policy. 2016;7:36-41.
- 8. Siddiqui M, Rajkumar SV. The high cost of cancer drugs and what we can do about it. Mayo Clin Proc. 2012:87:935-43.
- 9. Faden RR, Chalkidou K, Appleby J, Waters HR, Leider JP. Expensive cancer drugs: a comparison between the United States and the United Kingdom. Milbank Q. 2009;87:789-819.
- 10. Lichtenberg FR. How cost-effective are new cancer drugs in the U.S.? Expert Rev Pharmacoecon Outcomes Res. 2020;1-17.
- 11. Sharma SV, Lee DY, Li B, Quinlan MP, Takahashi F, Maheswaran S, McDermott U, Azizian N, Zou L, et al. A chromatin-mediated reversible drug-tolerant state in cancer cell subpopulations. Cell. 2010;141:69-80.
- 12. Wilson C, Nicholes K, Bustos D, Lin E, Song Q, Stephan JP, et al. Overcoming EMT-associated resistance to anti-cancer drugs via Src/FAK pathway inhibition. Oncotarget. 2014;5:7328-41.

- 13. Brown R, Links M. Clinical relevance of the molecular mechanisms of resistance to anti-cancer drugs. Expert Rev Mol Med. 1999;1:1-21.
- 14. Nurgali K, Jagoe RT, Abalo R. Adverse effects of cancer chemotherapy: Anything new to improve tolerance and reduce sequelae? Front Pharmacol. 2018;9:245.
- 15. Thomas CJ, Rahier NJ, Hecht SM. Camptothecin: current perspectives. Bioorg Med Chem. 2004;12:1585-604.
- Kumar A, Patil D, Rajamohanan PR, Ahmad A. Isolation, purification and characterization of vinblastine and vincristine from endophytic fungus Fusarium oxysporum isolated from Catharanthus roseus. PLoS one. 2013;8.
- 17. Islam KJJ, Koorbanally NA. A novel flavonoid and furoquinoline alkaloids from Vepris glomerata and their antioxidant activity. Nat Prod Commun. 2011;6.
- Chaturvedula PVS, Schilling JK, Miller JS, Andriantsiferana R, Rasamison VE, Kingston DGI. New cytotoxic alkaloids from the wood of vepris punctata from the Madagascar rainforest. J Nat Prod. 2003;66:532-4.
- Maggi F, Randriana FR, Rasoanaivo P, Nicoletti M, Quassinti L, Bramucci M, et al. Chemical composition and in vitro biological activities of the essential oil of Vepris macrophylla (Baker) I.Verd. Endemic to Madagascar. Chem Biodivers. 2013;10:356-66.
- 20. Imbenzi PS, Osoro EK, Aboud NS, Omollo J, Cheplogoi PK. A review on chemistry of some species of genus Vepris (Rutaceae family). 2014;3(3):357-62.
- 21. Geldenhuys WJ, Gaasch KE, Watson M, Allen DD, Van der Schyf CJ. Optimizing the use of open-source software applications in drug discovery. Drug Discov Today. 2006;11:127-32.
- Ekins S, Mestres J, Testa B. In silico pharmacology for drug discovery: methods for virtual ligand screening and profiling. Br J Pharmacol. 2007;152:9-20.
- 23. Daina A, Michielin O, Zoete V. SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. Sci Rep. 2017;7:42717.
- 24. Banerjee P, Eckert AO, Schrey AK, Preissner R. ProTox-II: A webserver for the prediction of toxicity of chemicals. Nucleic Acids Res. 2018;46:257-63.
- 25. Filimonov DA, Lagunin AA, Gloriozova TA, Rudik AV, Druzhilovskii DS, Pogodin PV, Poroikov VV. Prediction of the biological activity spectra of organic compounds using the pass online web resource. Chem Heterocycl Comp. 2014;50:444-57.
- Hanwell MD, Curtis DE, Lonie DC, Vandermeerschd T, Zurek, E, Hutchison GR.

- Avogadro: an advanced semantic chemical editor, visualization, and analysis platform. J Cheminformatics. 2012;4:4-17.
- Yang Z, Lasker K, Schneidman-Duhovny D, Webb B, Huang CC, Pettersen EF, Goddard TD, Meng EC, et al. UCSF Chimera, MODELLER, and IMP: An integrated modeling system. J Struct Biol. 2012;179:269-78.
- 28. Ayafor JF, Okogun JI. Nkolbisine, a New Furoquinoline Alkaloid, and 7-Deacetylazadirone From Teclea verdoorniana. Available at: https://pubs.acs.org/doi/pdf/10.1021/np50020a012. Accessed on: 26 May 2020.
- Baell JB, Holloway GA. New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays. J Med Chem. 2010;53:2719-40.
- 30. Capuzzi SJ, Muratov EN, Tropsha A. Phantom PAINS: Problems with the Utility of Alerts for Pan-Assay in terference Compound S. J Chem Inf Model. 2017;57:417-27.
- 31. Lipinski CA. Lead- and drug-like compounds: The rule-of-five revolution. Drug Discov Today Technol. 2004;1:337-41.
- 32. Veber DF, Johnson SR, Cheng HY, Smith BR, Ward KW, Kopple KD. Molecular properties that influence the oral bioavailability of drug candidates. J Med Chem. 2002;45:2615-23.
- 33. Faber MS, Jetter A, Fuhr U. Assessment of CYP1A2 activity in clinical practice: why, how, and when? Basic Clin Pharmacol Toxicol. 2005;97:125-34.
- 34. Rettie AE, Jones JP. Clinical and toxicological relevance of CYP2C9: Drug-Drug Interactions and Pharmacogenetics. Annu Rev Pharmacol Toxicol. 2005;45:477-94.
- 35. Wedlund PJ. The CYP2C19 enzyme polymorphism. Pharmacol. 2000;61:174-83.
- Seino Y, Nagao M, Yahagi T, Hoshi A, Kawachi T, Sugimura T. Mutagenicity of several classes of antitumor agents to Salmonella typhimurium TA98, TA100, and TA92. Cancer Res. 1978;38:2148-56.
- 37. Wang X, Zhou X, Hecht SM. Role of the 20-hydroxyl group in camptothecin binding by the topoisomerase I-DNA binary complex. Biochem. 1999;38:4374-81.
- 38. Pommier Y. Topoisomerase I inhibitors: camptothecins and beyond. Nat Rev Cancer. 2006;6:789-802.

**Cite this article as:** Chepkirui C, Kagia R. *Vepris nobilis* plant: a potential source of anticancer agents. Int J Res Med Sci 2020;8:3203-7.