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Research Article

Microbiological profile of diabetic foot ulcers and its antibiotic susceptibility pattern in a teaching hospital, Gujarat

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ABSTRACT

Background: Diabetic foot lesions are a major medical, social and economic problem and are the leading cause of hospitalization for patients with diabetes worldwide. Infection sometimes leads to amputation of the infected foot if not treated promptly. The present study was conducted to isolate and identify the bacterial pathogens associated with diabetic foot ulcer and to find out its antibiotic susceptibility pattern to reduce the risk of complications.

Methods: Total 100 pus samples were collected from patients having diabetic foot ulcer, during July to October 2012. Samples were processed as per standard guidelines.

Results: Out of 100 pus samples, 73 (73%) yielded growth of organisms making total of 92 isolates. Out of 92 bacterial isolates, 72 were gram negative and 20 were gram positive. *Pseudomonas aeruginosa* 25 (27%) was most common isolate causing diabetic foot infections followed by 20 (22%) *Klebsiella sp.*, 17 (19%) *E. coli*, 15 (17%) *S. aureus*, 6 (7%) *Proteus sp.* and 4(3%) *Enterococci*, 2 (2%) *Acinetobacter sp.* and 2(2%) CONS and 1(1%) *Providencia*. Out of 72 GNB, 50 (69.4%) were extended spectrum β lactamase (ESBL) producer. Most gram negative isolates were resistant to levofloxacin, gentamicin, ampicillin-sulbactam and gatifloxacin. All GNB were sensitive to imipenem. Out of 15 S. aureus, 9 (60%) were Methicillin Resistant *Staphylococcus aureus* (MRSA) and were sensitive to vancomycin and linezolid

Conclusions: *Pseudomonas sp.* was the most common cause of infections. Most isolates were multi drug resistance.

Keywords: Diabetic foot ulcer, Polymicrobial infection, Bacterial isolates, Antibiotic susceptibility pattern

INTRODUCTION

Diabetes is a metabolic disorder of the endocrine system which plagues approximately 17 million people nationwide. Each year over 700,000 new cases are diagnosed; 12,000 to 14,000 of which are children, teenagers and young adults, while this life threatening disease can be controlled. Diabetes is often accompanied by serious complications, and still today there is no cure. ¹

Foot ulceration and infection in diabetic patients is one of the major causes of morbidity, hospitalization and foot amputation.² This complication accounts for approximately 20% of hospital admissions in diabetic patients.³ Diabetic foot infections include cellulitis, abscess, necrotizing fasciitis, septic arthritis, tendonitis and osteomyelitis.⁴

Infections are often polymicrobial, Multi drug resistant and associated with inadequate glycemic control. There is a need for continuous surveillance of resistant bacteria to provide the basis for empirical therapy and reduce the risk of complications.

Aims and objectives

- To isolate and identify the bacterial pathogens associated with Diabetic foot infections.
- To find out its antibiotic susceptibility pattern.

METHODS

The present study was conducted in the Department of Microbiology, Surendranagar during the period of July to October, 2012.

Sample collection

100 Pus samples were collected from patients having Diabetic foot infections, using a sterile disposable swab. Care was taken to avoid contamination of specimen. After collection samples were transported to the Microbiology department. Samples were processed as per standard guidelines.

Isolation and Identification

Samples were subjected to Gram stain to screen for presence of bacterial pathogen. Samples were inoculated on Blood agar, Mac Conkey agar and Nutrient agar. Isolates were identified and confirmed by biochemical reaction.

Antibiotic susceptibility testing

Antibiotic susceptibility testing was performed by Kirby Bauer Disk Diffusion method as per CLSI guidelines. Gram positive isolates were tested for Linezolid, vancomycin, tetracycline, gentamicin, Co-trimoxazole, ampicillin-sulbactam, penicillin, amoxicillin, erythromycin, clindamycin and neomycin. Gram negative isolates were tested for ciprofloxacin, gentamicin, amikacin, piperacillin-tazobactam, ampicillin-sulbactam, amoxicillin-clavulanic cotrimoxazole, chloramphenicol, gatifloxacin, imipenem and polymyxin B.

RESULTS

Out of 100 pus samples obtained, 73 (73%) yielded growth of organisms making total of 92 isolates. Out of 92 bacterial isolates, 72 were Gram negative and 20 were Gram positive.

Pseudomonas aeruginosa 25 (27%) was most common isolate causing diabetic foot infections followed by 20 (22%) Klebsiella sp., 17 (19%) E. coli, 15 (17%) S. aureus, 6 (7%) Proteus sp. and 4 (3%) Enterococci, 2 (2%) Acinetobacter sp. and 2 (2%) CONS and 1 (1%) providencia (Figure 1).

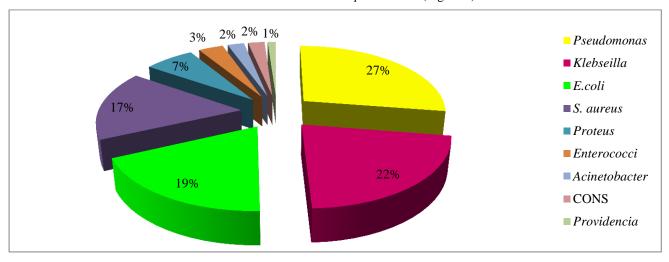


Figure 1: Distribution of various bacterial isolates.

Out of 72 Gram negative isolates, 50 (69.4%) were extended spectrum β lactamase (ESBL) producer. Out of 15 S. aureus, 9 (60%) were Methicillin Resistance Staphylococcus aureus (MRSA) (Figure 2). Gram negative isolates were found to be susceptible to imipenem (100%) followed by polymyxin b (88%), piperacillin/ tazobactam (38%), amikacin (38%), gatifloxacin (38%), ampicillin/ sulbactam (25%) and gentamicin (25%) (Figure 3). Gram positive isolates were found to be susceptible to vancomycin (100%), linezolid (100%)followed tetracycline by (90%),ampicillin/sulbactam (70%) and neomycin (70%), amoxicillin (40%), cotrimoxazole (40%) (Figure 4).

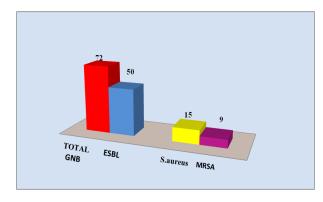


Figure 2: Special resistance pattern of GNB and *S. aureus*.

 $ESBL:\ Extended\ spectrum\ \beta\ lactamase.$ Resistant to all Penicillins and Cephalosporins. MRSA: Methicillin Resistant Staphylococcus aureus

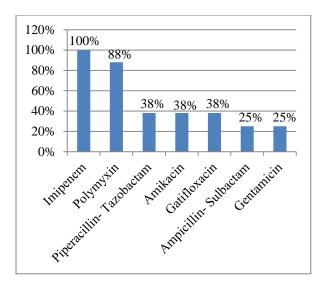


Figure 3: Antibiotic sensitivity pattern of gram negative isolates.

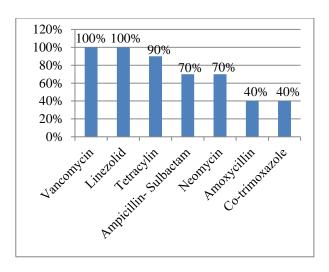


Figure 4: Antibiotic sensitivity pattern of gram positive isolates.

Table 1: Comparison of various bacterial isolates.

Study	Pseudomonas sp.	Klebsiella sp.	E. coli	Proteus sp.	S. aureus	Acinetobacter sp.	Enterococci
Ozer et al	18.90%	36.50%	36.50%	-	10.80%	2.70%	14.90%
Asha et al	23%	17%	12.00%	17%	21%	6%	0
J Viliam et al	24.30%	9%	15.30%	0	42.30%	0.00%	0.00%
Present study	34.70%	27.70%	23.60%	8.33%	75%	2.77%	15%

DISCUSSION

Various authors have reported different bacterial isolates associated with diabetic foot ulcer (Table 1). *Pseudomonas sp.* was the most common cause of diabetic foot infections. Most isolates were multi drug resistance.

Various factors like age, sex, type of diabetes, smoking, immunocompromised status, duration of diabetes, injury to the foot, duration of ulcer, neuropathy, peripheral vascular disease and resistance to ongoing treatment are responsible for aggravation of diabetic foot ulcer. Proper treatment of diabetes, Proper care of foot, and rigorous adherence to the principles of asepsis is the foundation of ulceration site infection prevention.

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REFERENCES

- Vimalin Hena J, Growther L. Studies on Bacterial Infections of Diabetic Foot Ulcer. Afr J Cln Exper Microbiol. 2010;11(3):146-9.
- 2. Lipsky BA, Pecoraro RE, Larson SA, Hanley ME, Ahroni JH. Outpatient Management of uncomplicted lower –extremity infection in diabetic patients. Arch Indian Med. 1990;150:790-7.
- 3. Abdulrazak A, Bitar ZI, Al-Shamali AA, Mobasher LA. Bacteriological study of diabetic foot infections. J Diabetes Complications. 2005;19(3):138-41.
- 4. Asha Konipparambil Pappu, Aprna Sinha, Aravind Johnson: Microbiological profile of Diabetic Foot Ulcer. Calicut Medical Journal. 2011;9(3):e2.
- 5. Clinical and Laboratory Standards Institute. Performance standard for antimicrobial susceptibility testing; 22 informational supplement. Clinical and Laboratory Standards Institute, Wayne, PA, USA. 2012;32(3):M100-S22.
- Ozer B, Kalaci A, Semerci E, Duran N, Davul S, Yanat A N: Infections and aerobic bacterial pathogens in diabetic foot, African Journal of Microbiology Research. 2010;4(20):2153-60.

- 7. J Vimalian Hena, Lali Growther. Studies on bacterial infections of Diabetic foot Ulcer, Afr J Cln Exper. Microbiol. 2010;11(3):146-9.
- 8. Goldstein EJ, Citron DM, Nesbit CA. Diabetic foot infections: bacteriology and activity of oral antimicrobial agents against bacteria isolated from consecutive cases. Diabetes Care. 1996;19:638–41.
- 9. Tentolouris N, Jude EB, Smirnof I, Knowles EA, Boulton AJ. Methicillin-resistant *Staphylococcus aureus*: an increasing problem in a diabetic foot clinic. Diabet Med. 1999;16:767–71.
- Shanker EM, Mohan V, Premlatha G, Srinivasan RS, Usha AR: Bacterial etiology of diabetic foot infections in South India. Eur J Intern Med. 2005;16:567–70.
- 11. Hartemann-Heurtier A, Robert J, Jacqueminet S, Ha Van G, Golmard JL, Jarlier V, Grimaldi A. Diabetic foot ulcer and multidrug-resistant organisms: risk factors and impact. Diabet Med. 2004;21:710 –15.
- 12. Wagner FW. The avascular foot: a system of diagnosis and treatment. Foot Ankle. 1981;2:64 –122.
- 13. M, Caillaux M, Yazdanpanah Y, Mouton Y. Culture of percutaneous bone biopsy specimens for diagnosis of diabetic foot osteomyelitis: concordance with ulcer swab cultures. Clin Infect Dis. 2006;42:57–62.
- 14. Erle G, de Lalla F. Deep tissue biopsy vs. superficial swab culture monitoring in the microbiological assessment of limb threatening diabetic foot infection. Diabet Med. 2001;18:822–7.
- Mantey I, Hill RL, Foster AV, Wilson S, Wade JJ, Edmonds ME. Infection of foot ulcers with Staphylococcus aureus associated with increased

- mortality in diabetic patients. Commun Dis Public Health. 2000 Dec;3(4):288-90.
- Fejfarova V, Jerkovska A, Skiboia J, Petkov V. Pathogen resistance and other risk factors in the frequency of lower limb amputation in patients with the diabetic foot syndrome. Vnitr Lek. 2002;48:302– 6.
- 17. Dang CN, Prasad YD, Boulton AJ, Jude EB. Methicillin-resistant Staphylococcus aureus in the diabetic foot clinic: a worsening problem. Diabet Med. 2003;20:159 –61.
- 18. Viswanathan V, Jasmine JJ, Snehalatha C, Ramachandran A: Prevalence of pathogen in diabetic foot infection in South Indian type 2 diabetic patients. J Assoc Physicians India. 2002;50:1013–16.
- 19. Lipsky BA, Berendt AR. Principles and practice of antibiotic therapy of diabetic foot infections. Diabetes Metab Res. 2000;16(1):42-6.
- 20. Mohanty S, Kapil A, Dhawan B, Das BK. Bacteriological and antimicrobial Susceptibility profile of soft tissue infections from Northern India. Indian J Med Sci. 2004;58;10-5.
- 21. Ikem RT, Kolawole BA, Ikem IC. The prevalence, presentation and outcome of diabetic foot lesions in a Nigerian teaching hospital. Trop Doct. 2002;32:226-7.

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