

# Evaluation of newly generated LRP1 antibodies in different cell types: THP1 cancer cell line, human and mouse immune cells

Scott D. Semelsberger<sup>1,2</sup>, Olga Sizova, PhD<sup>2</sup>, Dan Li, PhD<sup>2</sup>, Qing Ma, PhD<sup>2</sup>, Lisa St. John, PhD<sup>2</sup>, Gheath Al-Atrash, DO, PhD<sup>2</sup>, and Jeffrey J Molldrem, MD<sup>2</sup>

<sup>1</sup> Department of Biology, Indiana University of Pennsylvania, Indiana, PA

<sup>2</sup> Department of Hematopoietic Biology and Malignancy, The University of Texas MD Anderson Cancer Center, Houston, TX

# THE UNIVERSITY OF TEXAS MDAnderson Cancer Center

Making Cancer History®

## **Background**

- Enhanced understanding of the basic biology surrounding T-cell activation and proliferation has enabled the use of immunotherapy to combat deadly diseases, such as cancer<sup>1</sup>.
- In our lab's previous findings, Low Density Lipoprotein Receptor Related Protein 1 (LRP1) has been shown to play a direct role in T-cell proliferation<sup>2</sup>
- The use of anti-LRP1 antibody to target T-cells resulted in a significant and permanent reduction of T-cell inhibition of proliferation<sup>2</sup>

## **Hypothesis**

 Exploiting the similarity in LRP1's homology in mouse and human, we hypothesize that newly-generated anti-human LRP1 antibody (6 clones) will bind to mouse LRP1

## **Objective**

 In this project, 6 newly generated purified anti-human LRP1 antibodies will be tested for binding on a monocyte-like cell line (THP1) and on different hematopoietic cell populations from both mice and human to effectively study LRP1's immunological function

### **Materials and Methods**

- 6 novel purified LRP1 antibodies were generated by GenScript (unconjugated)
- Two commercially available LRP1 antibodies (directly conjugated to AF488)
  8G1
  - 001 • EDD2724
  - EPR3724
- Mouse cell surface marker antibodies and secondary antibody
  - 2<sup>nd</sup> IgG antibody (anti-mouse), CD3, CD45, CD11B, Ly6C, Ly6G
- Human cell surface marker antibodies and secondary antibody
- 2<sup>nd</sup> IgG antibody (anti-mouse), CD3, CD14
- THP1 monocyte-like AML cell line
- Human buffy coat
- Mouse splenocytes
- Mouse peripheral blood
- Calculations were performed to ensure use of equal quantities of each antibody
- Flow cytometry was performed to determine binding intensity for each clone

#### Results

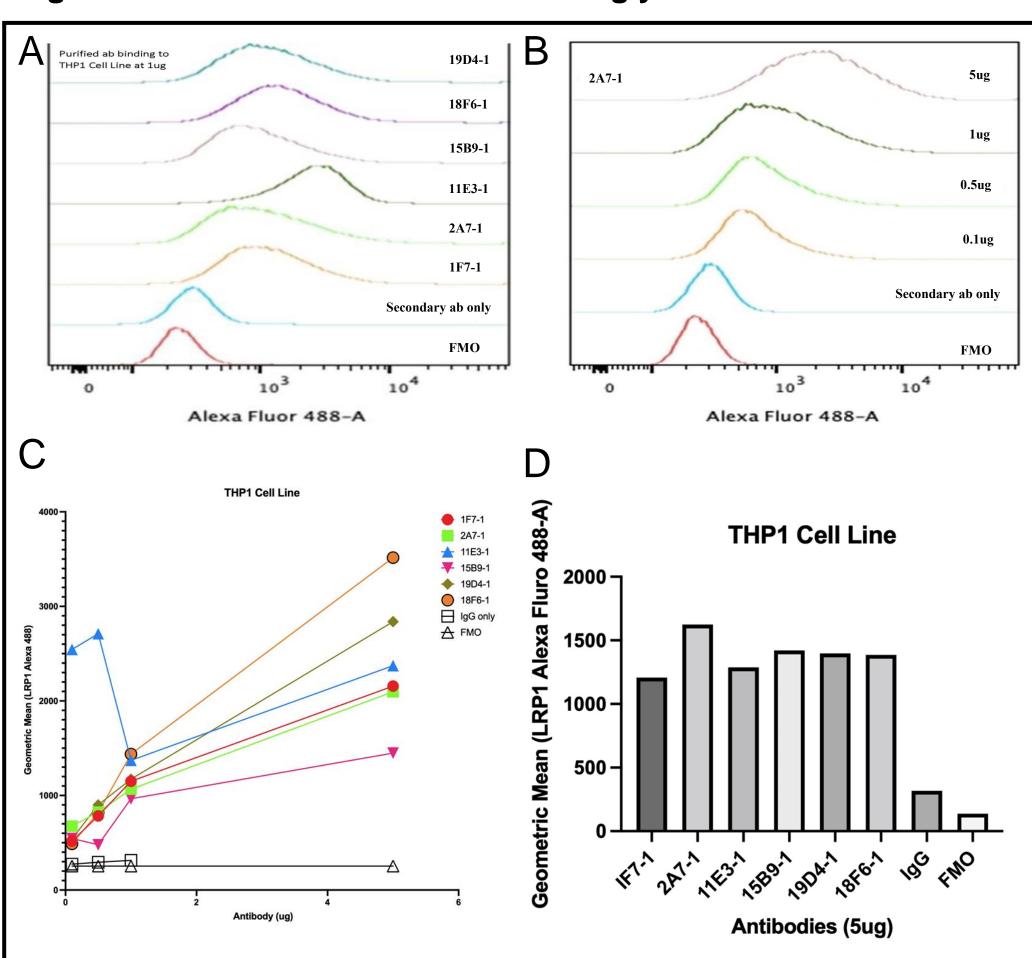
#### **Human Cell Line and Human Peripheral Blood**

- The newly generated LRP1 purified antibody clones bind strongly to the THP1 cancer cell line
- Significant binding was seen on human CD14+ (monocytes)
- No significant binding was seen on human CD3+ (T-cells)

#### **Mouse Splenocytes**

Significant binding occurred in mouse T-cells, monocytes, and neutrophils

#### Figure 1. Purified LRP1 Abs Bind Strongly to THP1 AML Cell Line



**A.** Flow cytometry histograms at 1.0ug for each antibody clone and negative controls. **B.** Antibody binding intensity is dose dependent for clone 2A7-1. **C.** Line graph of different antibody binding intensity at different concentrations. **D.** Bar graph of antibody binding intensity at 5ug.

#### Figure 2. Human Buffy Coat Flow Cytometry Gating Strategy

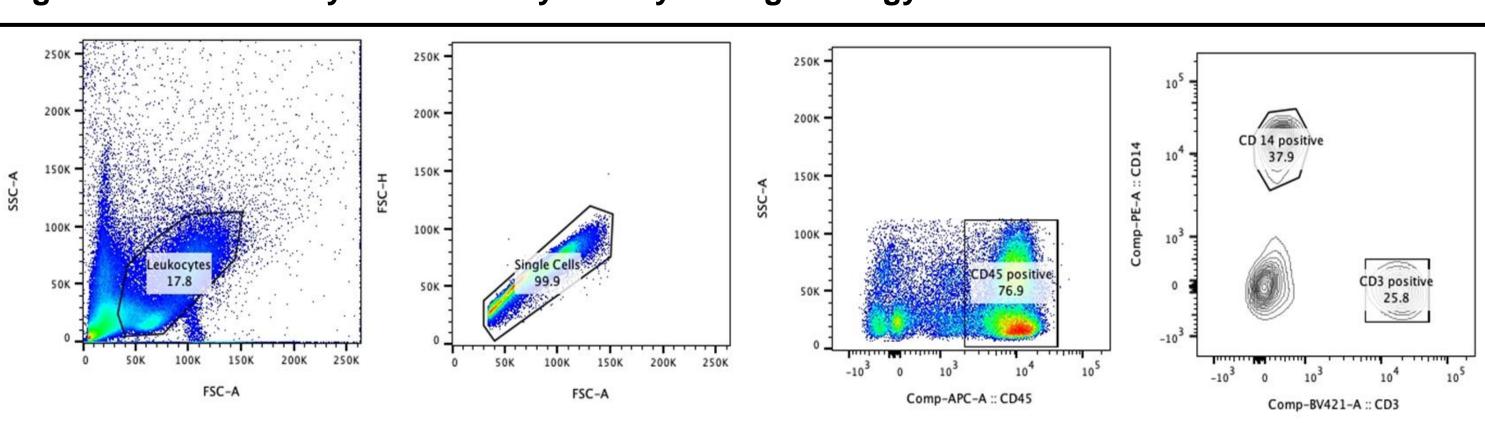
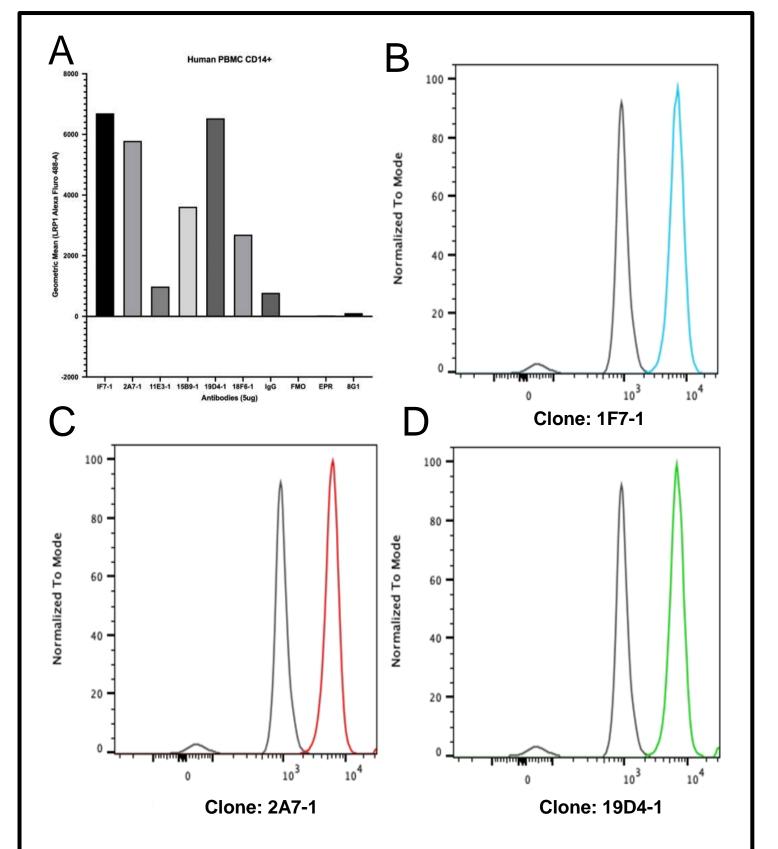
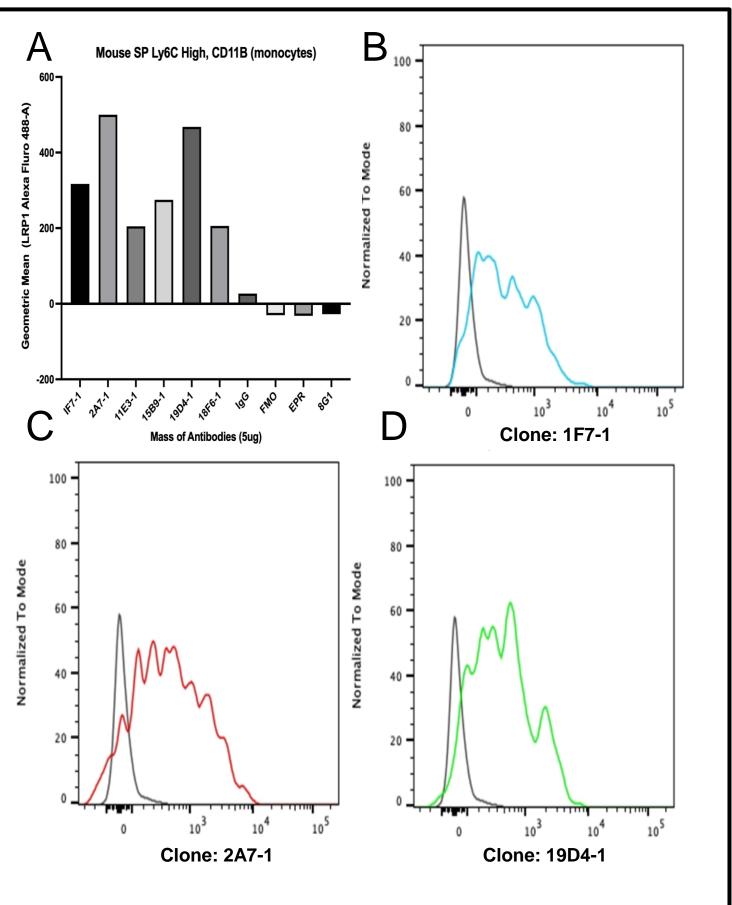


Figure 3. Purified LRP1 Antibodies Bind Strongly to Human Monocytes



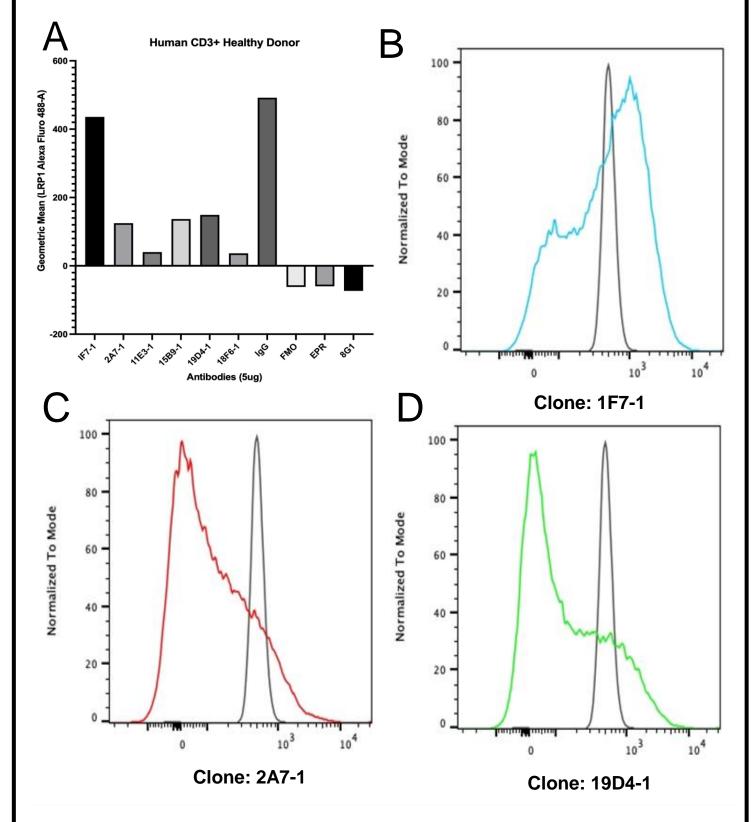
**A.** Flow cytometry fluorescence intensity bar graph at 5.0ug for each antibody clone and negative controls. **B, C, D**. CompAlexa Fluor 488-A:LRP1 is shown on the X axis and has a significant shift compared to the negative control for clone 1F7-1, 2A7-1, and 19D4-1. Grey is the secondary antibody only negative control.

# Figure 5. Purified LRP1 Antibodies Bind to Mouse Monocytes



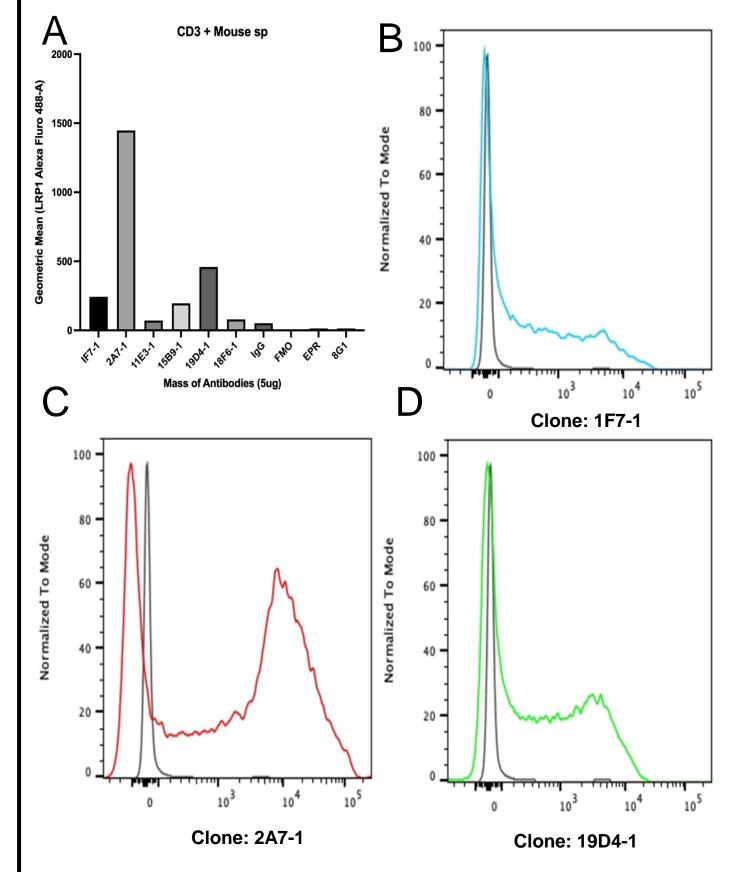
**A.** Flow cytometry fluorescence intensity bar graph at 5.0ug for each antibody clone and negative controls. **B, C, D**. CompAlexa Fluor 488-A:LRP1 is shown on the X axis and has a significant shift compared to the negative control for clone 1F7-1, 2A7-1, and 19D4-1. Grey is the secondary antibody only (negative control).

Figure 4. Purified LRP1 Antibodies Do Not Bind Strongly to Human T-Cells



**A.** Flow cytometry fluorescence intensity bar graph at 5.0ug for each antibody clone and negative controls. **B, C, D**. CompAlexa Fluor 488-A:LRP1 is shown on the X axis and does not have a significant shift compared to the negative control for clone 1F7-1, 2A7-1, and 19D4-1. Grey is the secondary antibody only negative control.

# Figure 6. Purified LRP1 Antibodies Bind to Mouse T-Cells

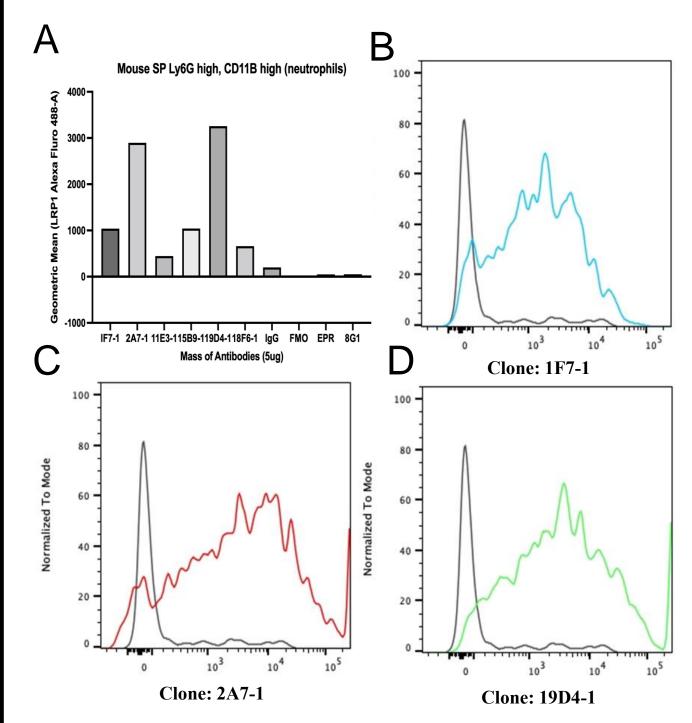


A. Flow cytometry fluorescence intensity bar graph at 5.0ug for each antibody clone and negative controls. **B, C, D**. Comp-Alexa Fluor 488-A:LRP1 is shown on the X axis and has a significant shift compared to the negative control for clone 1F7-

(negative control).

1, 2A7-1, and 19D4-1. Grey is the secondary antibody only

## Figure 7. Purified LRP1 Antibodies Bind to Mouse Neutrophils



**A.** Flow cytometry bar graph at 5.0ug for each antibody clone and negative controls. **B, C, D** Antibody fluorescence intensity is having a significant shift compared to the negative control for clone 1F7-1, 2A7-1, and 19D4-1.

# **Conclusion and Future Directions**

- Three antibody clones exhibit strong binding to LRP1 in the THP1 cancer cell line, human monocytes, and mouse splenocytes (T-cells, monocytes, and neutrophils)
  - 1F7-1
  - 2A7-1
  - 19D4-1
- The next steps include the following:
  - Utilizing various assays, determine which antibody has the strongest affinity to mouse and human LRP1
  - 2. Directly conjugate the antibody with fluorochrome to exclude using the secondary antibody
  - 3. Exploiting the ability of LRP1 to increase surface expression upon T-cell stimulation, test the LRP1 ab clones on CD3/CD28-stimulated human and mouse T-cells
  - 4. Validate LRP1 KO mouse T-cells model (negative control) with the chosen LRP1 ab clones

#### Acknowledgements

Scott Semelsberger was sponsored by the Partnership for Careers in Cancer Science and Cancer Medicine and as a Dr. John J Kopchick Fellow to participate in the 2022 Summer Experience.

### **Grants/Funding**

The Advanced Cytometry & Sorting Core Facility is supported by NCI P30CA016672

#### References

- Carreno BM, Magrini V, Becker-Hapak M, Kaabinejadian S, Hundal J, Petti AA, Ly A, Lie WR, Hildebrand WH, Mardis ER, Linette GP. Cancer immunotherapy. A dendritic cell vaccine increases the breadth and diversity of melanoma neoantigen-specific T cells. Science. 2015 May 15;348(6236):803-8. doi: 10.1126/science.aaa3828. Epub 2015 Apr 2. PMID: 25837513; PMCID: PMC4549796.
- Yang TH, St John LS, Garber HR, Kerros C, Ruisaard KE, Clise-Dwyer K, Alatrash G, Ma Q, Molldrem JJ. Membrane-Associated Proteinase 3 on Granulocytes and Acute Myeloid Leukemia Inhibits T Cell Proliferation. J Immunol. 2018 Sep 1;201(5):1389-1399. doi: 10.4049/jimmunol.1800324. Epub 2018 Jul 18. PMID: 30021768; PMCID: PMC6099529