We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



167,000





Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

### Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



#### Chapter

# Saffron endomycorrhizae: diversity and effect on plant growth and corm formation

Ourras Samah, El Aymani Ismail, Mouden Najoua, El Gabardi Soumaya, Adnani Manal, Selmaoui Karima, Artib Mariam, Benkirane Rachid, El Modafar Cherkaoui, Ouazzani Touhami Amina and Douira Allal

#### Abstract

Saffron cultivation is an important alternative for marginalized areas. Due to low soil fertility and low water availability, arbuscular mycorrhizal fungi are an essential alternative for maintaining fertility and water economy, stimulating growth and protecting plants against soil diseases. Studies on the diversity of endomycorrhizal fungi in the rhizosphere of *Crocus sativus* in Taliouine (Tinfat), located in Morocco, revealed the impact of age saffron plantations. A greater endomycorrhizal fungi density was recorded in the rhizosphere of saffron plants from plots operated for 2 years (138.66/100 g of soil) over that occupied for 10 years. Seventeen morphotypes of collected spores belong to 5 genera: Glomus (seven species), Acaulospora (seven species), Rhizophagus, Densicitata, and Funneliformis (one species). The weak endomycorrhizal species richness can be explained by the accumulation of C. sativus residues over time and its allelopathic effect. The beneficial effect of composite endomycorrhizal inocula, originating from Moroccan saffron plantations, was obvious in the growth of saffron plants, mother bulb number, the leaves length, root, and vegetative masses. These inocula mycorrhized over time saffron plants' roots and can sporulate at the level of the rhizosphere of these plants. The use of composite inocula, as biofertilizers, can be one of the solutions for sustainable agriculture.

**Keywords:** Morocco, saffron, diversity, endomycorrhizal, composite endomycorrhizal inoculums, growth parameters, bulb multiplication

#### 1. Introduction

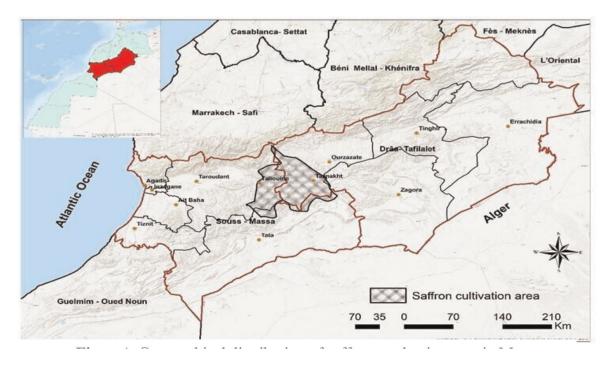
Saffron (*Crocus sativus* L.) is an autumn flowering plant from the *Iridaceae* family, renowned worldwide for its red stigmas, which represent the most precious spice in the world [1], a kilogram of good quality saffron produced from *C. sativus* can cost over 2000 US dollars. Approximately 150,000 flowers are needed to produce 1 kilogram of

dried saffron, and to growth is amount one would typically require some 2000 m<sup>2</sup> under cultivation per kg harvest [2, 3]. It has been cultivated in the Mediterranean area and Near East since the ancient period, used as a condiment for food, as a dye for textiles, and in traditional medicinal preparations [4]. Nowadays, saffron cultivation gaining interest due to its beneficial health effects, including antioxidant, anticancer, anti-inflammatory, and anti-depressive properties [5]. Iran, India, Greece, Morocco, and Spain are the main saffron-producing countries, with respective average annual productions of 180 tons, 9 tons, 5.5 tons, 3 tons, and 1 ton, recorded over the period 2004–2008 [6]. Saffron is also grown in small areas in Azerbaijan, Afghanistan, France, Switzerland, Italy, Turkey, Australia, and China [1, 3]. In recent years the productivity in Iran has increased enormously, Nevertheless, Spain remains the country most associated with this spice, notably thanks to its main production area, this position makes Spain currently control the world market of saffron and more particularly that of the United States, considered to be the main market of Spain [3], packs and re-exports about 40–50% of Iranian saffron. The United Arab Emirates is also an important importer of Iranian saffron [7].

Saffron is a hardy plant that thanks to its morphology and physiology can with stand very severe climatic conditions [8]. In Morocco, the saffron sector represents a major challenge for this local product both economically and socially. This sector is a promising way to reduce poverty and income inequalities in saffron-growing areas.

Indeed, it is one of the pillars of the economy of the Taliouine-Taznakht region, which is characterized by difficult soil and climate conditions [9], high rates of poverty and income inequality, and a high level of the rural exodus [10].

The annual production has increased according to ANDZOA from 3 tons of dry stigmas and an area of 600 ha in 2009 to 6.8 tons and about 1800 ha in 2018. The cultivation of saffron has been extended and intensified in recent years (Green Morocco Plan). The objective is to reach 9 tons in 2020 according to the contract program signed in 2016 between the Moroccan Interprofessional Federation of Saffron and the government. The cultivation area in Morocco is concentrated in the area of Taliouine–Tazenakht (**Figure 1**).



**Figure 1.** *Geographical distribution of saffron production area in Morocco.* 

According to the Regional Office of Agricultural Development (ORMVAO), the region of Souss-Massa ensures about 95% of the national production of saffron, which represents an annual turnover of 75 million DH. Around Two-thirds of the production is destined for the international market.

Moroccan saffron has a great reputation at national and international levels [11]. Data from the 'Office des Changes du Maroc' show that the main destination for "Moroccan" saffron is France. These data also show (mass imports registered with the customs services and which come essentially from the Islamic Republic of Iran or Greece. These quantities are redirected to Qatar or other lesser-known destinations. This intruding mass is cheaper, but affects the reputation of Moroccan saffron and competes with local productions.

Saffron cultivation management depends on the development of new technical production practices adapted to the pedoclimatic conditions of traditional areas for this crop. Among, the major constraints that limit its production and productivity is poor management of saffron cultivation, as it involves in adequate plant population, the incidence of corm rot disease [12], nutrient depletion, and lack of irrigation facilities. Moreover, environment and cultivation management affect strongly flower induction in C. sativus [13]. Accordingly, Increasing saffron yield and quality, reducing production costs, and flowering modulation may need new technology implementation as proposed in the Mediterranean environment viz., soilless cultivation systems were proposed but only limited and controversial reports are present in literature. Another option for saffron plant performance is through the use of biostimulants, or microorganisms applied to plants to enhance nutrition efficiency, abiotic stress tolerance, and/or crop quality traits, regardless of the irnutrient content. Soil microorganisms such as arbuscular mycorrhizal fungi (AMF) can form a mutualistic symbiosis with about 80% of land plant species, including several crops [14]. This association provides ecological stability to the environment [15, 16], and enhances water and nutrient uptakes such as phosphorus, nitrogen, and micronutrients, thus improving plant growth and resistance to biotic and abiotic stresses [17, 18].

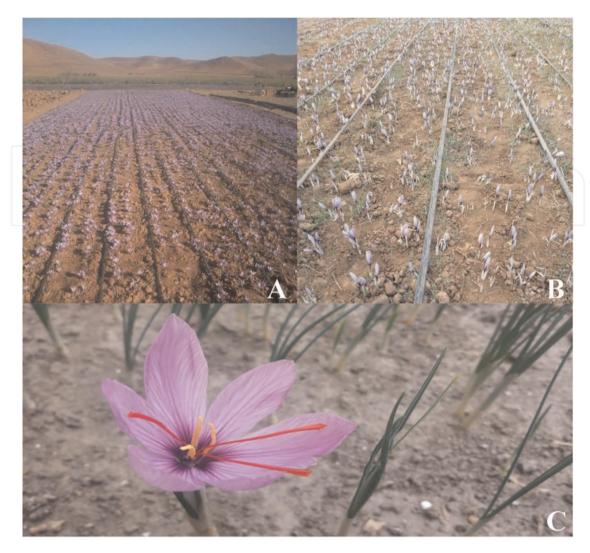
This chapter presents an overview of saffron cultivation in Morocco and information on the diversity of arbuscular mycorrhizal fungi in the Taliouine region. Inocula, based on these fungi, originating from the saffron rhizosphere, were tested to promote plant growth and bulb production and multiplication.

#### 2. Mycotrophic nature of Saffron plants

As saffron does not grow from seeds, reproduction is only possible by vegetative propagation, using the corms which can withstand a long dry dormant period before sprouting. Thus, corms are indispensable to saffron propagation reproducing vegetatively into corm lets that ultimately develop into new plants (**Figure 2**) [19].

The traditional area of saffron production in Morocco is characterized by very poor soils, however, the rhizosphere of cultivated saffron supports very interesting populations of microorganisms such as endomycorrhizae that are supposed to be essential for plant growth and ecosystem functioning.

Like most geophytes, saffron produces a coarse root system; these root systems have under gone extensive mycorrhization and benefit from this symbiosis [20]. Indeed, field-grown saffron is extensively mycorrhized [21, 22]. The Saffron plant

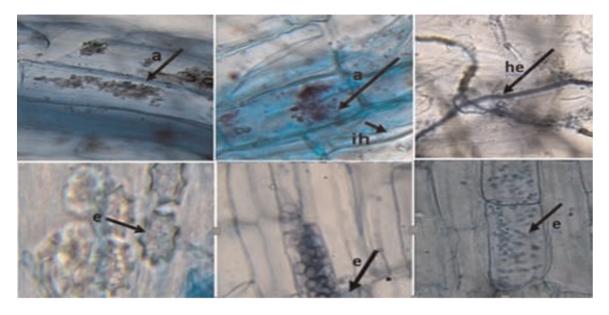


**Figure 2.** *A*, *B*: Saffron fields in Talouine region; C: Saffron plant at flowering stage.

shows a high incidence of AMF colonization. It was observed that the corm, despite being the modified stem, shows AMF colonization [23]. Morphological examination of fine roots comprised in rhizospheric soil samples collected in October 2020 from 15 different plots of saffron located in the Taliouine area (Morocco) under light microscope revealed the presence of AMF structures inside the examined roots which showed different colonization levels (**Figure 3**).

Ourras et al. [24] pointed out a high mycorrhization rate of 95.02% which reflects the inoculum pressure or propagules rate infecting the surrounding substrate. Likewise, El Aymani et al. [12] have reported different root mycorrhizal frequencies of 93.33% and 96.67% associated with the rhizosphere of *C. sativus* grown in the Taliouine region. This mycorrhizal colonization demonstrates relevant interaction between *C. sativus* roots and native endomycorrhizal populations under field conditions through which mechanisms such as modulation of water transport [25], nutrient acquisition [26], and stimulating the exudation of root phosphatase [27, 28] under drought conditions, are active in mycorrhizal plants to alleviate environmental stressors.

With the identification of native arbuscular mycorrhizal fungi and their application, it could be possible to expand the saffron cultivated area and increase the performance of arable lands.



**Figure 3.** AMF structures inside the examined roots of saffron plants grown in fields in Taliouine regions.

## 3. Diversity of arbuscular mycorrhizal fungi from moroccan saffron plantations areas

The knowledge of the current richness of Glomale an AM fungi (Glomerales; Glomeromycota) and biological diversity associated with plants grown in adverse environments could be necessary to expand the cultivated area and increase the performance of arable lands.

AMF diversity has been scarcely studied in the Taliouine-Taznakht region where saffron cultivation is exclusively concentrated [29].

This diversity was unveiled by direct spore isolation and morphological characterization of rhizospheric soil and roots samples from different sites in the main saffron production region. A variety of endomycorrhizal fungi has been noted in the rhizosphere of *C. sativus* grown in the Taliouine region (Tinfat) (**Figure 4**) [30]. These authors have detected 36 morphologically distinctive AMF species that were directly retrieved from field samples. All the encountered species in the different studied sites belong to six genera: *Glomus* (14 species), *Acaulospora* (10 species), *Scutellospora* (6 species), *Gigaspora* (2 species), *Pacispora* (2 species), and *Entrophospora* (1 species). The genus *Glomus* was the most dominant, it occurred in all studied sites with a proportion of distribution that varies between 8.33% and 30.56%.

A low number of AMF species have been reported by Ourras et al. [24]. Chamkhi et al. [31] detected 11 species of AMF spores among which *Funneliformis* and *Rhizoglomus* species were the most abundant (>35%). Yang et al. [32] reported that AMF soil community evolved in the function of environmental conditions. Another factor affects AMF survival and community composition in association with the saffron crop.

#### 3.1 Age of plantation

Age of plantation seemed to be affected the richness of AM fungi, spore density and root colonization rates [12, 24, 31]. Indeed, the greatest richness of AM fungi was registered in the site at 4 years of successive exploitation by saffron (24 species),

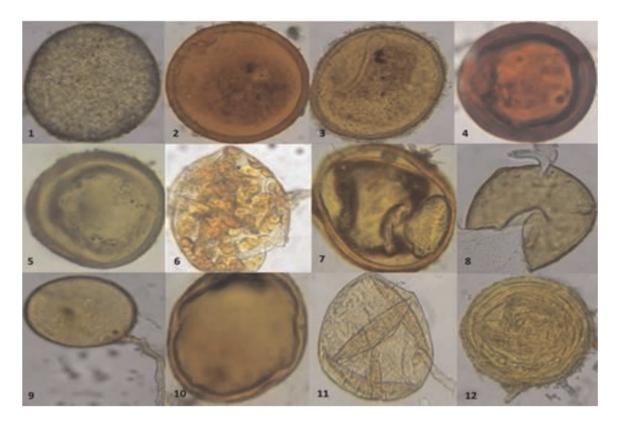


Figure 4.

Some indigenous endomycorrhizal fungals pecies isolated from the rhizosphere of Crocus sativus grown in Taliouine areas.

followed by the site at 6 years of occupation by saffron (21 species), while the lowest number of species was recorded at the sites of two, three and 10 years of soil use by saffron [33]. According to Yu et al. [34], stand age significantly changed the structure of the AM fungal community. The increase of spore density with field age has been cited in previous literature reviews [35, 36]. Rengifo-Del Aguila et al. [37] found that intra-radical colonization and AM fungal phylogenetic diversity increased with plantation age, while AM fungal richness was still constant across time but a significant compositional turnover was detected.

Many studies have demonstrated that the shift in the AM fungal community over ecological succession is associated with variations in soil properties induced by stand age, such as soil texture [38], pH [39], and nutrient availability [40]. Krüger et al. [41] pertain that P has become increasingly limited in old soils as the amount and availability of P declines over time. The changes in soil's chemical and physical properties can play an important role in decreasing saffron yields even after 6 years of cultivation [42].

#### 4. AMF as biofertilizers on Saffron

Regarding the biological activity, as biofertilizers, AMF offers a way to replace, at least partially, the use of chemical fertilizers and pesticides seeing their fatal effects on the environment and health. In low soil fertility of saffron cultivation areas [43, 44], they can serve as an AMF inoculum which may be part of the solution for sustainable agriculture. Cases in point showed successful application on saffron production under controlled conditions.

#### 4.1 Corm production

In addition to the spice yield, another economically important attribute of saffron is the number of replacement corms. Corms inoculation by composite AMF inoculum significantly initiated and stimulated the production of new corms (**Figure 5**) [30].

Such promoting effect of AMF on initiating the new underground stem propagules was observed previously too [18, 45–47]. Considering that tuber initiation in potato is hormonally mediated [48], it may be that in the present study too AMF affected hormone balance in the saffron plant, leading to earlier initiation and production of corms. Inoculation of corms with spores has enhanced the growth of corms and the number of spores per 10 g soil and results recorded as 13–52 with the highest occurrence in December–January [49]. Caser et al. [50] approved the fact that AMF symbiosis enhanced the production of replacement corms and reduced the occurrence of fungal diseases.

#### 4.2 Growth response

Although AMF increased the number of corms produced, it increased also shoot fresh weight and root dry weight [51]. A significant increment of the overall growth parameters such as length of aerial and below ground of saffron plants occurred too after AMF colonization [30].

The use of arbuscular mycorrhizal symbionts as biostimulants positively affected saffron cultivation, improving the crop performances and the content of important nutraceutical compounds. In particular, the inoculum composed by *R. intraradices* and *F. mosseae* increased flower production and saffron yield [52]. Such compatibility between AMF fungi and host plant was previously observed in other plants also such as onion [47] and Potato [18] cultivars.

Van der Heijden et al. [53], found that in the grassland ecosystem, inoculation of 14 AM fungal species increased plant diversity by 105% and plant productivity by 42% compared to that in the ecosystem inoculated with only 1 AM fungal species. A study has also revealed that different AM fungi can absorb P at different distances from the



**Figure 5.** *New bulbs production;* **M:** *Inoculated plant;* **C:** *Control.* 

root of the plant, making the host plants inoculated with mixed AM fungal species grow better than those inoculated with AM fungal species.

Variations in the growth of saffron plants are possibly observed as a function of the age of saffron plantation as seen below (**Figure 6**).



#### Figure 6.

Performance of Crocus sativus plants as affected by the presence of AMF and age of saffron plantations.



#### Figure 7.

Different structures of endomycorrhizal fungi colonizing the roots of Saffron plants 4 months after corms inoculation: Arbuscules (a), Spores (s) and intra-radicular hyphae (h) ( $G \times 400$ ).

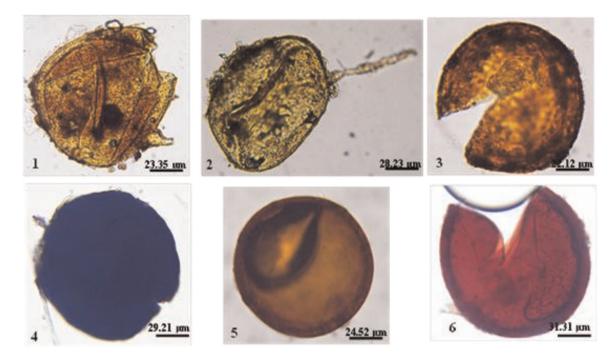


Figure 8.

Quelques morphotypes de spores rencontrés au niveau de la rhizosphère des plants du safran provenant des cormes traitées avec un inoculum endomycorhiziens (S4), après quatre mois de culture. 1: Rhizophagus intraradices, 2: Glomus lamellosum, 3: Funneliformis geosporum, 4: Densicitata nigra, 5: Glomus microcarpum;, 6: Glomus deserticola.

#### 4.3 Root colonization with AMF after corms inoculation

The positive influence of AMF on plant is otherwise viewed in roots, at this level, mycorrhized plants are shown if they have been able to establish a strong mutualistic relationship with composite AMF inoculum. Four months after inoculation in potted culture, the roots of the plants from the inoculated corms are richly mycorrhized and showed numerous arbuscules and vesicles with extended hyphal colonizing groots. Arbuscules and spores were the most dominant structures (**Figures 7** and **8**).

#### 5. Conclusion

The growing interest in knowing and safeguarding the biological diversity associated with certain arid environments may contribute to improving ecologically the yields of agricultural crops. In the case of saffron, the rhizospheric soil harbored a diversified AMF community and its application as composite inoculum in substrate culture pertains a potential performance on growth and plant succession. As a perspective, future studies should be focused to know the influence of AMF on the growth of the saffron plant in open field conditions.

#### Acknowledgements

This study was conducted under the research project of the 2nd PMA 2020/11, entitled 'Effect of endomycorrhizal fungi inoculum on agro morphological behavior

and productivity of saffron (*C. sativus*) under water and salinity stress' financed by 'Agence Nationale des Plantes Médicinales et Aromatique' et l'Université Ibn Tofail, Kénitra, Morocco'.

# IntechOpen

#### Author details

Ourras Samah<sup>1</sup>, El Aymani Ismail<sup>1</sup>, Mouden Najoua<sup>2</sup>, El Gabardi Soumaya<sup>1</sup>, Adnani Manal<sup>1</sup>, Selmaoui Karima<sup>1</sup>, Artib Mariam<sup>1</sup>, Benkirane Rachid<sup>1</sup>, El Modafar Cherkaoui<sup>3</sup>, Ouazzani Touhami Amina<sup>1</sup> and Douira Allal<sup>1\*</sup>

1 Laboratory of Plant, Animal and Agro-Industry Productions, Botany, Biotechnology and Plant Protection Team, Faculty of Sciences, Ibn Tofail University, Kenitra, Morocco

2 Laboratory of Molecular Chemistry and Environmental Molecules, Multidisciplinary Faculty of Nador-Mohammed 1st University Oujda, Morocco

3 Center of Agrobiotechnology, Research Unit Labeled CNRST (URL-CNRST05), Faculty of Sciences and Techniques Guéliz, Cadi Ayyad University, Marrakech, Morocco

\*Address all correspondence to: douiraallal@hotmail.com

#### IntechOpen

© 2022 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### References

[1] Gresta F, Lombardo GM, Siracusa L, Ruberto G. Saffron, an alternative crop for sustainable agricultural systems. A review. Agronomy for Sustainable Development. Springer Verlag/EDP Sciences/INRA. 2008;**28**(1):95-112

[2] Aboudrare A, AW Hassan A, Lybbert TJ. Importance socio-économique du Safran pour les ménages des zones de montagne de la région de Taliouine-Taznakht au Maroc. Rev. Mar. Sci. Agron. Vét. 2014;**2**:5-14

[3] Wyeth P, Malik N. A Strategy for Promoting Afghan Saffron Exports. Report. Afghanistan: ICARDA and Washington State University; 2008

[4] Giupponi L, Ceciliani G, Leoni V, Panseri S, Pavlovic R, Guido L, et al. Quality traits of saffron produced in Italy: Geographical area effect and good practices. Journal of Applied Botany and Food Quality. 2019;**92**:336-342

[5] Mykhailenko O, Desenko V, Ivanauskas L, Georgiyants V. Standard operating procedure of Ukrainian saffron cultivation according to good agricultural and collection practices to assure quality and traceability. Industrial Crops and Products. 2020;**151**:112376

[6] Dubois, A., (2010). Analyse de la filière du safran au Maroc: Quelle perspective pour la mise en place d'une indication géographique? Thèse de Master of Science, N 107. Montpellier: CIHEAM-IAMM

[7] Ghorbani M. The efficiency of saffron's marketing channel in Iran.World Applied Sciences Journal. 2008;4 (4):523-527

[8] Sepaskhah AR, Yarami N. Interaction effects of irrigation regime and salinity

on flower yield and growth of saffron. The Journal of Horticultural Science and Biotechnology. 2009;**84**(2):216-222

[9] Garcin GD, Carral S. Le safran marocain entre tradition et marché. Etude de la filière du safran au Maroc, en particulier dans la région de Taliouine, province de Taroudant. Rapport de consultation. Maroc: Projet FAO/ Association Migrations et Développement; 2007 http://www.mpdiscussion.org/casablanca/doc/zaf.pdf

[10] Bouchelkha M. Etude Sociologique sur le Safran. *Rapport de consultation*. Maroc: Projet FAO/TCP/MOR/3201. Rome, Italie; 2009

[11] Lage M, Cantrell CL. Quantification of saffron (Crocus sativus L.) metabolites crocins, picrocrocin and safranal for quality determination of the spice grown under different environmental Moroccan conditions. Scientia Horticulturae. 2009; **121**(3):366-373

[12] El Aymani I, Qostal S, Mouden N,
Selmaoui K, Touhami AO, Benkirane R,
et al. Fungi associated with saffron (*Crocus sativus*) in Morocco. Plant Cell
Biotechnology and Molecular Biology.
2019;20:1180-1188

[13] De Juan JA, Córcoles HL, Muñoz RM, Picornell MR. Yield and yield components of saffron under different cropping systems. Industrial Crops and Products. 2009;**30**(2):212-219

[14] Berruti A, Lumini E, Balestrini R, Bianciotto V. Arbuscular mycorrhizal fungi as natural biofertilizers: Let's benefit from past successes. Frontiers in Microbiology. 2016;**6**:1559

[15] Harley JL, Smith SE. Symbiose Mycorhizienne. Londres, Royaume-Uni: Academic Press; 1983 [16] Strullu DG. Les mycorhizes des arbres et des plantes cultivées. Lavoisier Paris. 3éme edit ed1991. p. 249

[17] Goussous SJ, Mohammad MJ. Comparative effect of two arbuscular mycorrhizae and N and P fertilizers on growth and nutrient uptake of onions. International Journal of Agriculture & Biology. 2009;**11**:463-467

[18] Lone R, Shuab R, Wani KA, Ganaie MA, Tiwari AK, Koul KK. Mycorrhizal influence on metabolites, indigestible oligosaccharides, mineral nutrition and phytochemical constituents in onion (*Alliumcepa L.*) plant. ScientiaHorti. 2015;**193**:55-61

[19] Hossein Zadeh H, Nassiri-Asl M.
Avicenna's (Ibn Sina) the canon of medicine and saffron (Crocus sativus): A review. Phytotherapy Research. 2013;27
(4):475-483

[20] Smith SE, Read DJ. Mycorrhizal Symbiosis. 2nd ed. London: Academic Press; 1997

[21] Kianmehr H. Vesicular arbuscular mycorrhizal spore population and infectivity of saffron (*Crocus sativus*) in Iran. New Phytologist. 1981;**88**(1):79-82

[22] Lone R, Razia SR, Koul KK. AMF association and their effect on metabolite mobilization, mineral nutrition and nitrogen assimilating enzymes in saffron (*Crocus sativus*) plant. Journal of Plant Nutrition. 2016;**39**:131852-131862

[23] Shuab R, Malla NA, Ahmad J, Lone R, Koul KK. Arbuscular mycorrhizal fungal symbiosis with saffron (Crocus sativus L.) Plant. J. New Biol. Rep. 2016; 5(8)

[24] Ourras S, El Gabardi S, Mouden N, Chliyeh M, Selmaoui K, Benkirane R, et al. Diversity of arbuscular mycorrhizal fungi in the rhizosphere of saffron (Crocus sativus) plants along with age of plantation in Taliouine region in Morocco. Acta Biologica Szegediensis. 2021;**2**(65):199-209

[25] He JD, Zou YN, Wu QS, Kuča K. Mycorrhizas enhance drought tolerance of trifoliate orange by enhancing activities and gene expression of antioxidant enzymes. Scientia Horticulturae. 2020;**262**:108745

[26] Boutasknit A, Baslam M, Ait-El-Mokhtar M, Anli M, Ben-Laouane R, Douira A, et al. Arbuscular mycorrhizal fungi mediate drought tolerance and recovery in two contrasting carob (*Ceratonia siliqua L.*) ecotypes by regulating stomatal, water relations, and (in) organic adjustments. Plants. 2020;**9**(1):80

[27] Chen W, Meng P, Feng H, Wang C.
Effects of arbuscular mycorrhizal fungi on growth and physiological performance of Catalpa bungei CA Mey.
Under Drought Stress. Forests. 2020;11: 1117

[28] Cheng HQ, Zou YN, Wu QS, Kuča K. Arbuscular mycorrhizal fungi alleviate drought stress in trifoliate orange by regulating H+-ATPase activity and gene expression. Frontiers in Plant Science. 2021;**12**:659694

[29] Aziz L, Sadok W. Stratégies d'adaptation des producteurs du Safran de Taliouine (Maroc) face au changement climatique. Journal of Alpine Research| Revue de géographie alpine. 2015:103-102

[30] El Aymani I, Chliyeh M, Selmaoui K, Mouden M, El Gabardi S, Ouazzani Touhami A, et al. Effect of a composite endomycorrhizal inoculum on the growth of saffron plants (*Crocus sativus L*.) and the multiplication of

corms. Plant Cell Biotechnology and Molecular Biology. 2019c;**20**(23&24): 1122-1136

[31] Chamkhi I, Sbabou L, Aurag J. Endophytic fungi isolated from *Crocus sativus L.* (saffron) as a source of bioactive secondary metabolites. Pharmacognosy Journal. 2018;**10**(6)

[32] Yang C, Hamel C, Schellenberg MP, Perez JC, Berbara RL. Diversity and functionality of arbuscular mycorrhizal fungi in three plant communities in semi arid Grasslands National Park, Canada. Microbial Ecology. 2010;**59**(4):724-733

[33] El Aymani I, El Gabardi S, Artib M, Chliyeh M, Selmaoui K, Ouazzani TA, et al. Effect of the number of years of soil exploitation by saffron cultivation in Morocco on the diversity of endomycorrhizal fungi. Acta Phytopathologica et Entomologica Hungarica. 2019b;**54**(1):71-86

[34] Yu Z, Liang K, Wang X, Huang G, Lin M, Zhou Z, et al. Alterations in arbuscular mycorrhizal community along a chronosequence of teak (Tectona grandis) plantations in tropical forests of China. Frontiers in Microbiology. 2021;**12** 

[35] Birhane E, Gebremedihin KM, Tadesse T, Hailemariam M, Solomon N. Exclosures restored the density and root colonization of arbuscular mycorrhizal fungi in Tigray, Northern Ethiopia. Ecol Process. 2017;**6**:33

[36] Burni T, Hussain F, Sharief M. Arbuscular mycorrhizal fungi (amf) associated with the rhizosphere of Mentha arvensis I., and M. longifolia huds. Pakistan Journal of Botany. 2011; **43**(6):3013-3019

[37] Aguila SRD, la Sota-Ricaldi D, Maria A, Corazon-Guivin MA, López-García Á.

Phylogenetic diversity of arbuscular mycorrhizal fungal communities increases with crop age in coffea arabica plantations. Journal of Soil Science and Plant Nutrition. 2022:1-13

[38] Herrmann L, Lesueur D, Bräu L, Davison J, Jairus T, Robain H, et al. Diversity of root-associated arbuscular mycorrhizal fungal communities in a rubber tree plantation chronosequence in Northeast Thailand. Mycorrhiza. 2016;**26**(8):863-877

[39] Reyes HA, Ferreira PFA, Silva LC, da Costa MG, Nobre CP, Gehring C. Arbuscular mycorrhizal fungi along secondary forest succession at the eastern periphery of Amazonia: Seasonal variability and impacts of soil fertility. Applied Soil Ecology. 2019;**136**:1-10

[40] Wang Q, Ma M, Jiang X, Guan D,
Wei D, Cao F, et al. Influence of 37 years of nitrogen and phosphorus fertilization on composition of rhizosphere arbuscular mycorrhizal fungi communities in black soil of Northeast China. Frontiers in Microbiology. 2020; 11:539669

[41] Krüger M, Teste FP, Laliberté E, Lambers H, Coghlan M, Zemunik G, et al. The rise and fall of arbuscular mycorrhizal fungal diversity during ecosystem retrogression. Molecular Ecology. 2015;**24**(19):4912-4930

[42] Khozaei M, Kamgar Haghighi AA, Sepaskhah AR, Karimian NA. Effect of 10-year continuous saffron cultivation on physical and chemical properties of soil. Iran Agricultural Research. 2015; **34**(1):46-55

[43] Chamkhi I, Sbabou L, Aurag J. Application of Plant Growth Promoting Rhizobacteria (PGPR) as Biological Fertilizers for Improving the Growth and Yield of Saffron (*Crocus sativus L.*). In: 1st Mediterranean Forum for Ph.D. Students and Young Researchers Designing Sustainable Agricultural and Food Production Systems under Global Changes in the Mediterranean 18-19 July 2016. Montpellier, France; 2016

[44] Chamkhi I, Abbas Y, Tarmoun K, Aurag J, Sbabou L. Morphological and molecular characterization of arbuscular mycorrhizal fungal communities inhabiting the roots and the soil of saffron (Crocus sativus L.) under different agricultural management practices. Archives of Agronomy and Soil Science. 2019;**65**(8):1035-1048

[45] Graham SO, Green NE, Hendrix JW. The influence of vesiculararbuscularmycorrhizalfungi on growth and tuberization of potatoes. Mycologia. 1976;**68**(4):925-929

[46] Niemira BA, Safir GR, Hammerschmidt R, Bird GW. Production of prenuclear minitubers of potato with peat-based arbuscular mycorrhizal fungal inoculum. Agronomy Journal. 1995;**87**(5):942-946

[47] Shuab R, Rafiq LR, Jayanti NJ, Sharma V, Imtiyaz S, Koul KK. Benefits of inoculation of arbuscular mycorrhizal fungi on growth and development of onion (*Allium cepa*) plant. American-Eurasian J. Agric. & Environ. Sci. 2014; **14**(6):527-535

[48] Ewing EE. The role of hormones in potato (*Solanum tuberosum* L.) tuberization. In: Plant Hormones. Dordrecht: Springer; 1995. pp. 698-724

[49] Kianmehr H.

Endotrophicmicorehyza of Saffron in Khorasan and its possible application. Gonabad, Iran: Proceeding of the Second National Symposium on Saffron and Medicinal plants; 1994 [50] Caser M, Victorino IMM, Demasi S, Berruti A, Lumini E, Bianciotto V, et al. In: XXX International 60 Horticultural Congress IHC2018: 61 International Symposium on Medicinal 62 and Aromatic Plants, Culinary Herbs and IV International Jujube Symposium and VI International Symposium on Saffron Biology and Technology. 2018b;**1287**: 441-446

[51] Mohebi-Anabat M, Riahi H, Zanganeh S, Sadeghnezhad E. Effects of arbuscular mycorrhizal inoculation on the growth, photosynthetic pigments and soluble sugar of Crocus sativus (saffron) in autoclaved soil. Int. J. Agron. Agric. Res. 2015;**6**:296-304

[52] Caser M, Victorino ÍMM, Demasi S, Berruti A, Donno D, Lumini E, et al. Saffron cultivation in marginal alpine environments: How AMF inoculation modulates yield and bioactive compounds. Agronomy. 2018a;**9**(1):12

[53] Van Der Heijden MG, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, Boller T, et al. Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. Nature. 1998;**396**(6706): 69-72

