

1-23-2019

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Kashyap, V.K., Wang, Q., Setua, S. et al. Therapeutic efficacy of a novel β III/ β IV-tubulin inhibitor (VERU-111) in pancreatic cancer. *J Exp Clin Cancer Res* 38, 29 (2019). <https://doi.org/10.1186/s13046-018-1009-7>

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Therapeutic efficacy of a novel β III/ β IV-tubulin inhibitor (VERU-111) in pancreatic cancer

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Abstract

Background: The management of pancreatic cancer (PanCa) is exceptionally difficult due to poor response to available therapeutic modalities. Tubulins play a major role in cell dynamics, thus are important molecular targets for cancer therapy. Among various tubulins, β III and β IV-tubulin isoforms have been primarily implicated in PanCa progression, metastasis and chemo-resistance. However, specific inhibitors of these isoforms that have potent anti-cancer activity with low toxicity are not readily available.

Methods: We determined anti-cancer molecular mechanisms and therapeutic efficacy of a novel small molecule inhibitor (VERU-111) using in vitro (MTS, wound healing, Boyden chamber and real-time xCELLigence assays) and in vivo (xenograft studies) models of PanCa. The effects of VERU-111 treatment on the expression of β -tubulin isoforms, apoptosis, cancer markers and microRNAs were determined by Western blot, immunohistochemistry (IHC), confocal microscopy, qRT-PCR and in situ hybridization (ISH) analyses.

Results: We have identified a novel small molecule inhibitor (VERU-111), which preferentially represses clinically important, β III and β IV tubulin isoforms via restoring the expression of miR-200c. As a result, VERU-111 efficiently inhibited tumorigenic and metastatic characteristics of PanCa cells. VERU-111 arrested the cell cycle in the G2/M phase and induced apoptosis in PanCa cell lines via modulation of cell cycle regulatory (Cdc2, Cdc25c, and Cyclin B1) and apoptosis - associated (Bax, Bad, Bcl-2, and Bcl-xl) proteins. VERU-111 treatment also inhibited tumor growth ($P < 0.01$) in a PanCa xenograft mouse model.

Conclusions: This study has identified an inhibitor of β III/ β IV tubulins, which appears to have excellent potential as monotherapy or in combination with conventional therapeutic regimens for PanCa treatment.

Keywords: VERU-111, Pancreatic cancer, β -tubulins, miR-200c, β III/ β IV-tubulin inhibitor

Background

Pancreatic cancer (PanCa) is one of the most lethal cancers and ranked as the fourth most common cause of cancer-related deaths among both men and women in the United States [1]. The management of PanCa is exceptionally difficult due to poor response to available therapeutic regimens [2]. Thus, the identification of

newer, highly effective therapeutic agents with no or minimal toxicity is highly desirable for the improved management of PanCa.

Tubulins associated cellular structures composed of α - β tubulin heterodimers are critically involved in a wide range of cellular processes, including maintenance of cell shape, cell motility and cell division, thus are considered important molecular targets in cancer therapeutics [3, 4]. Among these, β -tubulins (namely β I, β IIa, β IIb, β III, β IVa, β IVb, β V and β VI) isotypes are reported to have higher expression in several cancer types such as lung, breast, prostate, gastric and melanoma [5–10].

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Moreover, increased expression of β -tubulins is correlated with disease progression, overall survival and resistance to chemotherapeutic agents in different cancers [10, 11]. The aberrant expression of the β -tubulins (such as β IVa and β IVb) increases in metastatic cancer cells [12, 13]. Recent studies have reported major roles of β III and β IV-tubulins in PanCa as these isoforms are highly expressed in pancreatic tumors, while absent in normal pancreas (acinar and pancreatic islets) [14, 15]. The expression of these tubulins has been associated with PanCa progression, metastasis and chemoresistance [13, 16–20]. Additionally, β III-tubulin knockdown reduced the pancreatic tumor growth and metastasis in an orthotopic xenograft mouse model [15]. These studies suggest a major role of β III and β IV-tubulins in PanCa, thus can be precise and logical molecular targets for PanCa treatment. Although numerous tubulin inhibitors have been reported, nevertheless, selective inhibitors of these specific tubulins are rarely identified [21]. In this study, we have shown that a novel synthetic molecule that is known to overcome multidrug resistance preferentially inhibits the expression of β III and β IV-tubulins via restoring the expression of miR-200c in PanCa cells [22–24]. For the first time, we have also demonstrated that VERU-111 effectively inhibits growth and metastatic phenotypes of PanCa cells in vitro and in vivo model systems. Our study suggests potential implications of this novel small molecule in the treatment of PanCa in future.

Methods

Synthesis of VERU-111

Synthesis of VERU-111 was carried out as described in our earlier study [24]. Its synthesis scheme and characterization procedures are shown in Additional file 1: Figure S1.

Antibodies and reagents

Specific monoclonal and polyclonal antibodies of β I-tubulin (cat. # SC-5274), β III-tubulin (cat. # SC-51,670) were obtained from Santa Cruz Biotechnology. β V-tubulin (cat. # ab110592) was obtained from Abcam. β IIb-tubulin (cat. # PA5-60448), β IVb-tubulin (cat. # PA5-25050), β VI-tubulin (cat. # PA5-21826) were obtained from Invitrogen. Antibody β IIa-tubulin (cat. # TA345669) and β IVa-tubulin (cat. # TA-340088) were obtained from OriGene. Cyclin B1 (cat. # 4138), Cdc25C (cat. # 4688), Cdc2 (cat. # 77055), p-Cdc2Tyr15 (cat. # 9111), Bax (cat. # 2772), Bcl-2 (cat. # 2876), Bad (cat. # 9292), Bcl-xL (cat. # 2762), Caspase 3 (cat. # 9665), Cleaved caspase 3 (cat. # 9661), Caspase 9 (cat. # 9508) and PARP (cat. # 9542) were purchased from Cell Signaling Technology. The anti-mouse IgG HRP and rabbit IgG HRP-linked secondary antibodies were procured from Promega (Madison, WI). The hematoxylin stain

was purchased from Fisher Scientific and the Annexin V/FITC apoptosis kit from Bio-Rad (Hercules, CA). MTT(3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2-H-tetrazoliumbromide), Phenylmethanesulfonyl fluoride (PMSF), fetal bovine serum (FBS), eukaryotic protease inhibitor cocktail, pyruvic acid and Propidium iodide (PI), were purchased from Sigma–Aldrich Co. (St. Louis, MO) or Fisher Scientific (Pittsburgh, PA). Pan Caspases inhibitor (Z-VAD-FMK cat. FMK001) was obtained from R&D systems, USA.

Cell lines

Panc-1, AsPC-1, and HPAF-II cells were obtained from ATCC and cultured in their respective media as DMEM, RPMI-1640 and DMEM/F12 containing 10% FBS and 1% antibiotic/antimycotic. Cells were maintained in CO₂ incubator at 37 °C with 98% humidity and 5% CO₂ environment.

Cell proliferation and colony forming assays

The anti-proliferative effect of VERU-111, colchicine, paclitaxel and vinorelbine on PanCa cells was examined by MTT and colony formation assays as described in our previous studies [25].

miR transfection and qRT-PCR

Cells were transfected using Lipofectamine 2000 (Invitrogen) following the manufacturer's protocol. Briefly, Panc-1 and AsPC-1 cells were transiently transfected with miR-200c mimics or non-targeting control mimic (NC) at 100 nM (Applied Biosystems). Total RNA was extracted from control and VERU-111 treated PanCa cells using TRIzol™ reagent (Invitrogen, Life Technologies, Grand Island, NY). cDNAs were prepared by SYBR Green RNA Reverse Transcription kit. The mRNA expression of beta tubulin isoforms were analyzed by qPCR using specific primers sequences as described earlier in Additional file 2: Table S1 [26]. For miRNA detection, 100 ng total RNA was reverse transcribed into cDNA using specific primers designed for miRNA analysis (Applied Biosystems, Foster City, CA). Expression of miRNA 200c was determined by qPCR using the Taqman PCR master mix and specific primers designed for the detection of mi-R200c (Applied Biosystems). The expression of miR-200c was normalized with endogenous control RUN6B [27].

Western blot and confocal microscopy analyses

PanCa cells (1×10^6) were treated with VERU-111, colchicine, and vinorelbine (5–20 nM) for 24 h. Total cell lysates were processed for Western blot analysis for detecting protein levels of various beta tubulin isoforms and other oncoproteins [28, 29]. Confocal immunofluorescence microscopy was performed for microtubule

subcellular localization [28]. Briefly, PanCa cells were seeded on glass coverslips in 6 well plates and incubated overnight. Cells were treated with VERU-111 and tubulins targeting agent, fixed and incubated primary antibodies overnight. This was followed by incubation with or Alexa Fluor 488, donkey secondary antibodies for 1 h. The images were then captured with a Zeiss 710 Confocal microscope and Zen imaging software (Zeiss). To investigate whether caspase 3 and 9 are involved in VERU-111 induced apoptosis, PanCa cells were treated with Z-VAD-FMK (20 μ) for 2 h followed by VERU-111 for 24 h. The protein levels of cleaved and pro-Caspase-3, 9, and PARP protein were analyzed by Western blot analysis.

In situ hybridization (ISH)

To determine the expression of miR-200c, we performed in situ hybridization assays in excised tumor tissues of control and VERU-111 treated mice by Biochain kit (Biochain, San Francisco, CA) as described [28].

Cell migration and invasion assays

Cell migration assay was performed in Corning's 96-well HTS Transwell as per manufacturer's instructions. Cells were treated with VERU-111 (1.25–10 nM) for 24 h, fixed with 4% para-formaldehyde, and stained with crystal violet. Further, a wound healing assay was also performed to evaluate the effect of VERU-111 on cell migration. Images of the wounds were monitored under a phase-contrast microscope at 10X magnification. For invasion assays, cells were grown on BD Biocoat Matrigel Invasion Chambers (BD Biosciences, Heidelberg, Germany) according to the manufacturer's protocol. Cells were then treated with different concentrations of VERU-111 and incubated for 24 h. Invaded cells were fixed, stained and counted as described in our previous study [30].

Real time cell proliferation, migration and invasion by xCELLigence assays

The effect of VERU-111 on proliferation, migration and invasion of Panc-1 and AsPC-1 cells was investigated by real time xCELLigence technology as described in our previous study [31]. PanCa cells were seeded per chamber for cell proliferation, migration, and invasion assays in E plates following the xCELLigence real time cell analyzer manuals. VERU-111 and vehicle control were added at indicated time and concentrations. The baseline cell index for VERU-111 treated cells compared to control cells was calculated for at least two measurements from three independent experiments.

Cell cycle analysis

Effect of VERU-111 on cell cycle arrest of PanCa (Panc-1 and AsPC-1) cells was analyzed by flow cytometric

analysis. Briefly, approximately 70% confluent cells were synchronized by overnight incubation in FBS free media. Cells were treated with VERU-111 (0, 5, 10, and 20 nM) for 24 h, harvested, fixed overnight in ice-cold ethanol (70%) followed by incubation with RNase and incubated with the DNA staining Propidium iodide (Sigma) for flow cytometry. Data regarding the number of cells in different phases of the cell cycle was analyzed by BD Accuri C6; Becton-Dickinson, Mountain View, CA.

Apoptosis assay

The effect of VERU-111 on apoptosis induction in PanCa cells was analyzed by Annexin V-7AAD staining and mitochondrial membrane potential ($\Delta\Psi_m$). Briefly, PanCa cells (1×10^6) were treated with VERU-111 (5–40 nM) for 24 h. To investigate whether caspase 3 and 9 are involved in VERU-111 induced apoptosis, PanCa cells were treated with Z-VAD-FMK (20 μ M) for 2 h followed by VERU-111 for 24 h. Cells were then collected and stained with Annexin V and 7-AAD (5 μ l/100 μ l of cell suspension) and analyzed by Accuri C6 Flow Cytometer setting FL2 and FL3 channels. Effect of VERU-111 on mitochondrial membrane potential ($\Delta\Psi_m$) in PanCa cells was analyzed by uptake of tetramethylrhodamine (TMRE) staining. Briefly, PanCa cells were treated with VERU-111 (5–20 nM) for 24 h, and further incubated with TMRE (100 nM) for 20 min; Fluorescence intensities of TMRE stained cells were measured by flow cytometry.

Tumor xenograft study

To determine effective tumor growth inhibitory dose of VERU-111 at tumor site, we performed ectopic xenograft studies in athymic nude mice. For that, six-week-old female athymic nude mice (nu/nu) were purchased from Jackson laboratory and maintained in a pathogen-free environment. All procedures were carried out as per the approved UTHSC Institutional Animal Care and Use Committee (UTHSC-IACUC) protocol. To establish ectopic xenograft tumors in mice, AsPC-1 cells (2×10^6 cells) were suspended in phosphate buffer saline (PBS) and Matrigel (BD Biosciences) solution (1:1 ratio) and then injected subcutaneously on the dorsal flanks of each mouse. Tumor growth in mice was monitored using a digital Vernier caliper. When tumor volume reached $\sim 200 \text{ mm}^3$, mice were divided into control ($n = 6$) and VERU-111 ($n = 6$) treatment groups. Mice were administered with VERU-111 (50 μ g/mouse/week for 3 weeks; intra-tumorally) or vehicle control (PBS). Tumor volumes were measured weekly and calculated by the formula $0.5238 \times L \times W \times H$, where L is length, W is width and H is the height of the tumor. Mice were euthanized when tumor volume of control mice reached $\sim 1000 \text{ mm}^3$. At the time of sacrifice, tumors were excised and

processed for RNA, tissue lysates, histopathology and slides preparation (5 μ m section).

Immunohistochemistry

The effect of VERU-111 was determined on the expression of PCNA and tubulin isoforms in excised tumors by immunohistochemistry using Biocare kits (Biocare Medical, Concord, CA) as described previously [28].

Statistical analysis

The data discussed above are presented in terms of mean values and the SEM of several independent experiments. *p*-values < 0.05 were considered statistically significant. All statistical analyses were performed using the Statistical Package for the Social Sciences, version 11.5 (SPSS Inc., Chicago, IL).

Results

VERU-111 inhibits growth and clonogenic potential of PanCa cells

We first assessed the cytotoxic effect of VERU-111 against various human PanCa cell lines (AsPC-1, Panc-1 and HPAF-II). In this experiment, cells were treated with various concentrations of VERU-111 (1.25–160 nM) for 24 and 48 h, and cell viability was determined by MTT assay. VERU-111 inhibited the growth of PanCa cells in a dose and time-dependent manner (Fig. 1Ai-ii). The IC₅₀ of VERU-111 was 25, 35 and 35 nM in Panc-1, AsPC-1 and HPAF-II, respectively after 24 h treatment (Fig. 1Ai), while 48 h post-treatment it was 11.8, 15.5, and 25 nM (Fig. 1Aii). We further evaluated growth inhibitory effect of VERU-111 in real time with the help of xCELLigence system. This system monitors cell growth by measuring electrical impedance, which is expressed as a cell index. A growth curve, recorded as the basal cell index value, showed that VERU-111 significantly reduced the cell index in a dose-dependent manner compared to vehicle treated cells (Fig. 1Bi-ii). To determine the long-term effect of VERU-111 on the growth, we performed colony formation assays in PanCa cells. VERU-111 (1.25–5 nM) treatment significantly reduced the number of colonies in a dose-dependent manner (Fig. 1C-E) as compared to respective control groups.

VERU-111 inhibits mRNA expression and protein stability of β -tubulin isoforms in PanCa cells

It is well documented that β III and β IV-tubulins are involved in pancreatic carcinogenesis and are potential molecular targets for PanCa therapeutics [32]. Thus, we first investigated the effect of VERU-111 on the expression of β III and β IV-tubulins in PanCa cells. VERU-111 (5–20 nM) treatment significantly (*p* < 0.01) inhibited the mRNA expression of β III and β IV-tubulins, in PanCa cells (Fig. 2A) as determined by qRT-PCR. Western blot

analysis results also demonstrated VERU-111 mediated inhibition of the expression of β III and β IV-tubulins (Fig. 2B). We also evaluated the effect of VERU-111 on other tubulin isoforms to determine its specificity at both the mRNA and protein level. We observed differential inhibition in the expression of β I and β VI tubulins protein and mRNA levels (Fig. 2A-B). However, the expression of β IIa, β IIb and β V-tubulins remain unaffected in both the tested cell lines (Fig. 2A-B).

VERU-111 inhibits β III-tubulin and causes disruption of microtubular dynamics in PanCa cells

Next, we compared the effect of VERU-111 on the expression of β III-tubulin with known microtubule targeting agent's colchicine and vinorelbine (destabilizing agent) and paclitaxel (microtubule stabilizing). In this experiment, Panc-1 cells were treated with 5–20 nM of VERU-111, colchicine, vinorelbine and paclitaxel for 24 h, and RNA and protein lysates were prepared to determine the mRNA expression and protein level of β III-tubulin. VERU-111 effectively inhibited mRNA expression (Fig. 3Ai) and protein levels (Fig. 3B) of β III-tubulin. Destabilizing agents (colchicine and vinorelbine) did not show significant effect on the expression of β III-tubulin (Fig. 3Aii-iii and Fig. 3B). Paclitaxel treatment, however, stabilized the expression of β III-tubulin (Fig. 3Aiv and Fig. 3B). To determine effects of VERU-111 on microtubule networks and cell morphology, we utilized confocal microscopy. For that, Panc-1 cells were treated with VERU-111, colchicine, vinorelbine and Paclitaxel at different concentrations for 18 h. The untreated cells appeared typical nuclear and cytoskeleton structures, extending throughout the cell to the cell periphery while, VERU-111 treated cells show reduced and fragmented β III-tubulin microtubule network (Fig. 3C). Colchicine and vinorelbine did not affect the morphological changes in microtubule. Paclitaxel treatment led the formation of shortened but unusually highly defined microtubules bundles concentrated toward the nucleus (Fig. 3C). The microtubule fragmentation with disruption observed by VERU-111 is consistent with its putative mechanism of action (i.e., inhibiting tubulin polymerization).

VERU-111 restores the expression of miR-200c via targeting β III-tubulin

It has been reported that miR-200c directly targets β III-tubulin in PanCa cells. Thus, we sought to determine the effect of VERU-111 on the expression of miR-200c [33]. Interestingly, VERU-111 treatment induced the expression of miR-200c in Panc-1, AsPC-1 and HPAF-II cells, when compared with control treated cells (Fig. 3D). We next determined if inhibition of miR-200c minimizes the effect of VERU-111 on the

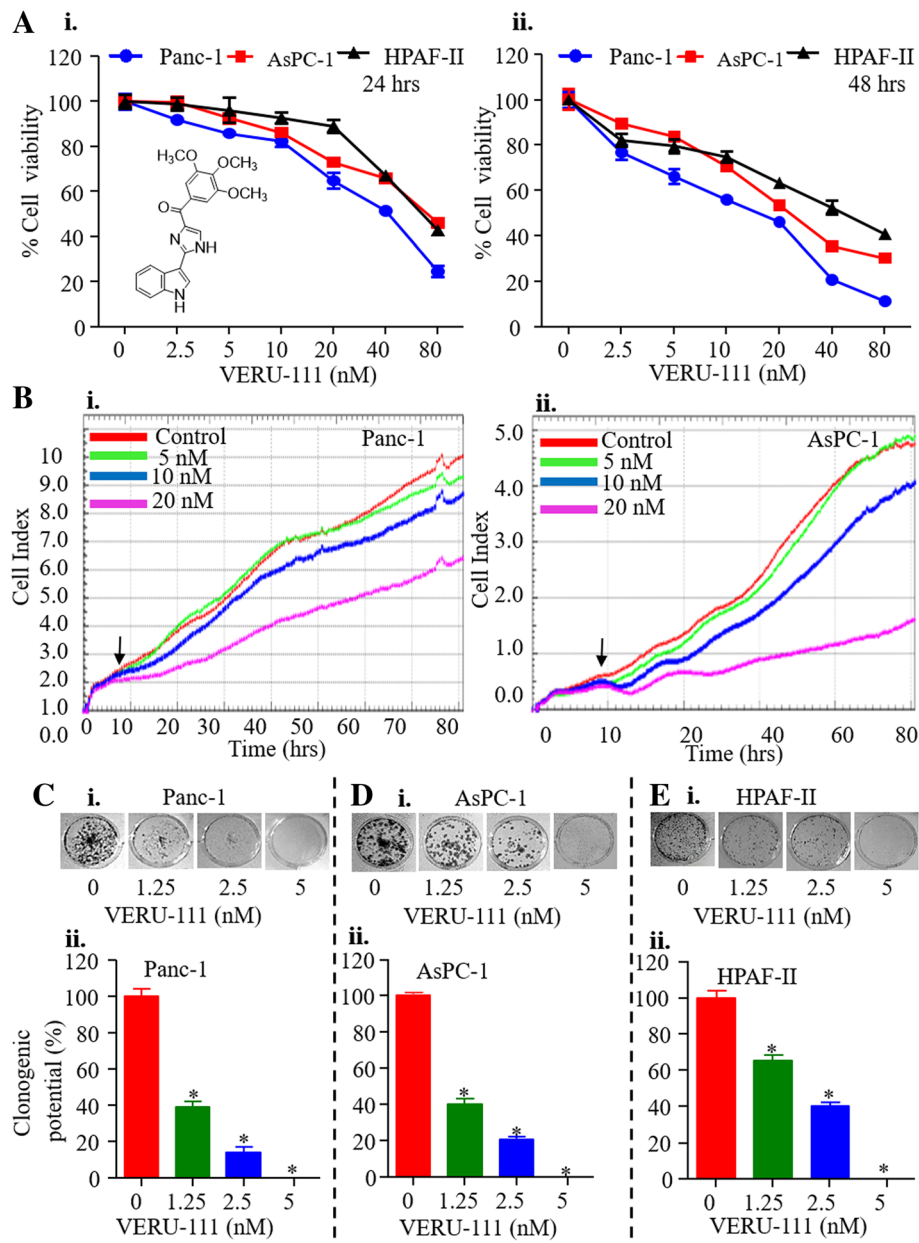
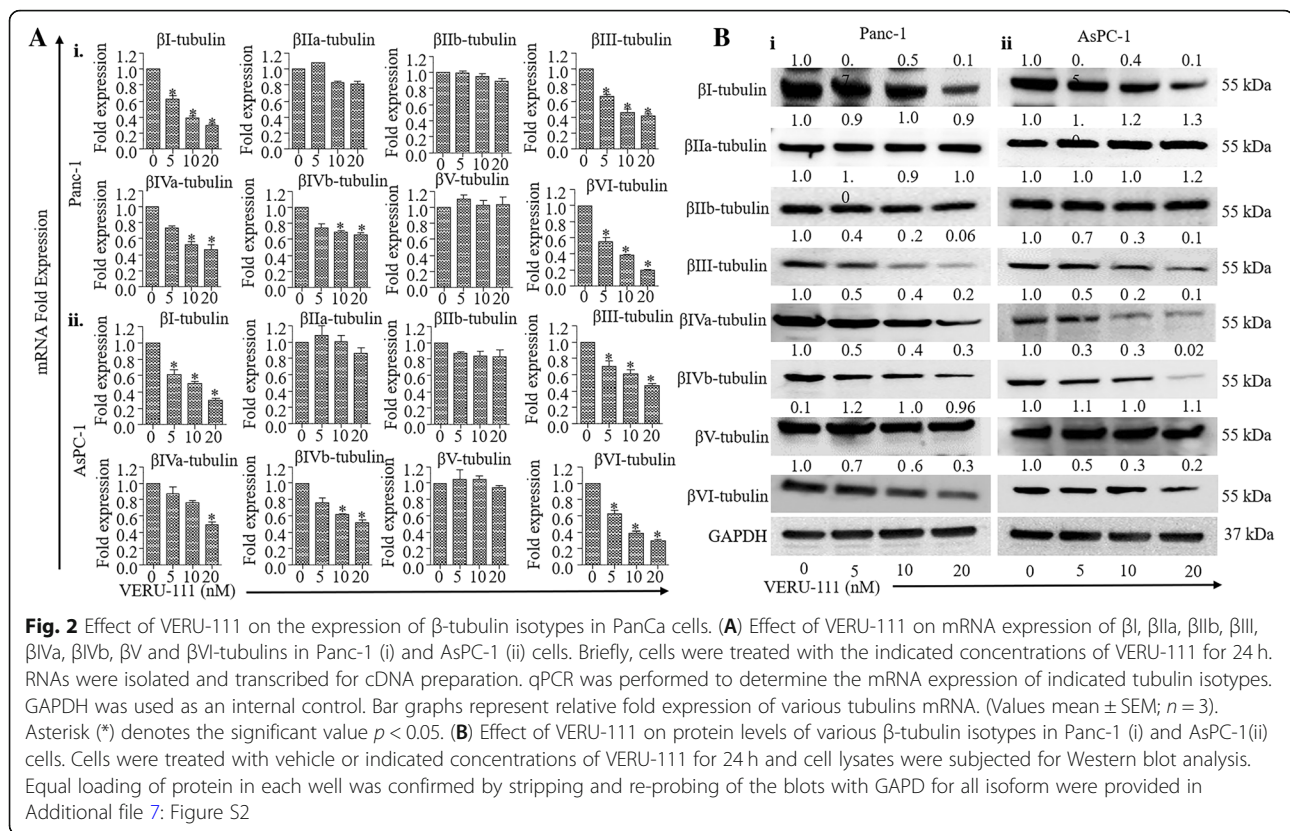


Fig. 1 VERU-111 inhibits growth characteristics of PanCa cells. **(A)** Effect of VERU-111 on viability of Panc-1, AsPC-1, and HPAF-II cells. Structure of VERU-111 (2-(1H-indol-3-yl)-1H-imidazol-4-yl) (3, 4, 5-trimethoxyphenyl) – methanone) is shown as insert. Cells were treated with indicated concentrations of VERU-111 for 24 **(i)** and 48 **(ii)** hrs and cell viability was determined by MTT assay. **(B)** Effect of VERU-111 on PanCa cells in a real time cell proliferation assay. Cells (5000 cells/well) were seeded in E-plate (xCELLigence) and placed into the xCELLigence Real Time Cell Analyzer (RTCA) DP. After 8–10 h, VERU-111 or the vehicle control was added and the experiment was allowed to run for 80 h. Line graphs show the average baseline cell index of control and VERU-111-treated cells. **(C–E)** Effect of VERU-111 on clonogenic potential of PanCa cells as determined by anchorage dependent colony formation assay. Representative images of colony formation assay and bar graphs indicating quantification of cells are shown at indicated concentrations of VERU-111. (Values means \pm SEM; $n = 3$). Asterisk (*) denotes the significant value $p < 0.05$

expression of β III tubulin. For that, cells were transfected with miR-200c mimics in Panc-1 cells that resulted to the inhibition of β III-tubulin expression of, which was rescued with the transfection of miR-200c inhibitor (Fig. 3Ei-ii). A combination of VERU-111 and miR-200c mimic treatment showed a pronounced effect

on the inhibition of β III tubulin expression (Fig. 3E). These results suggest that VERU-111 inhibits β III tubulin expression via restoration of the expression of miR-200c in PanCa cells. Next, we performed comparative anti-proliferative effect of VERU-111, colchicine, vinorelbine and Paclitaxel in PanCa (Panc-1, AsPC-1



and HPAF-II) cells using MTT assays. Interestingly, VERU-111 showed the most potent anti-proliferative activity compared to other agents (Fig. 3F).

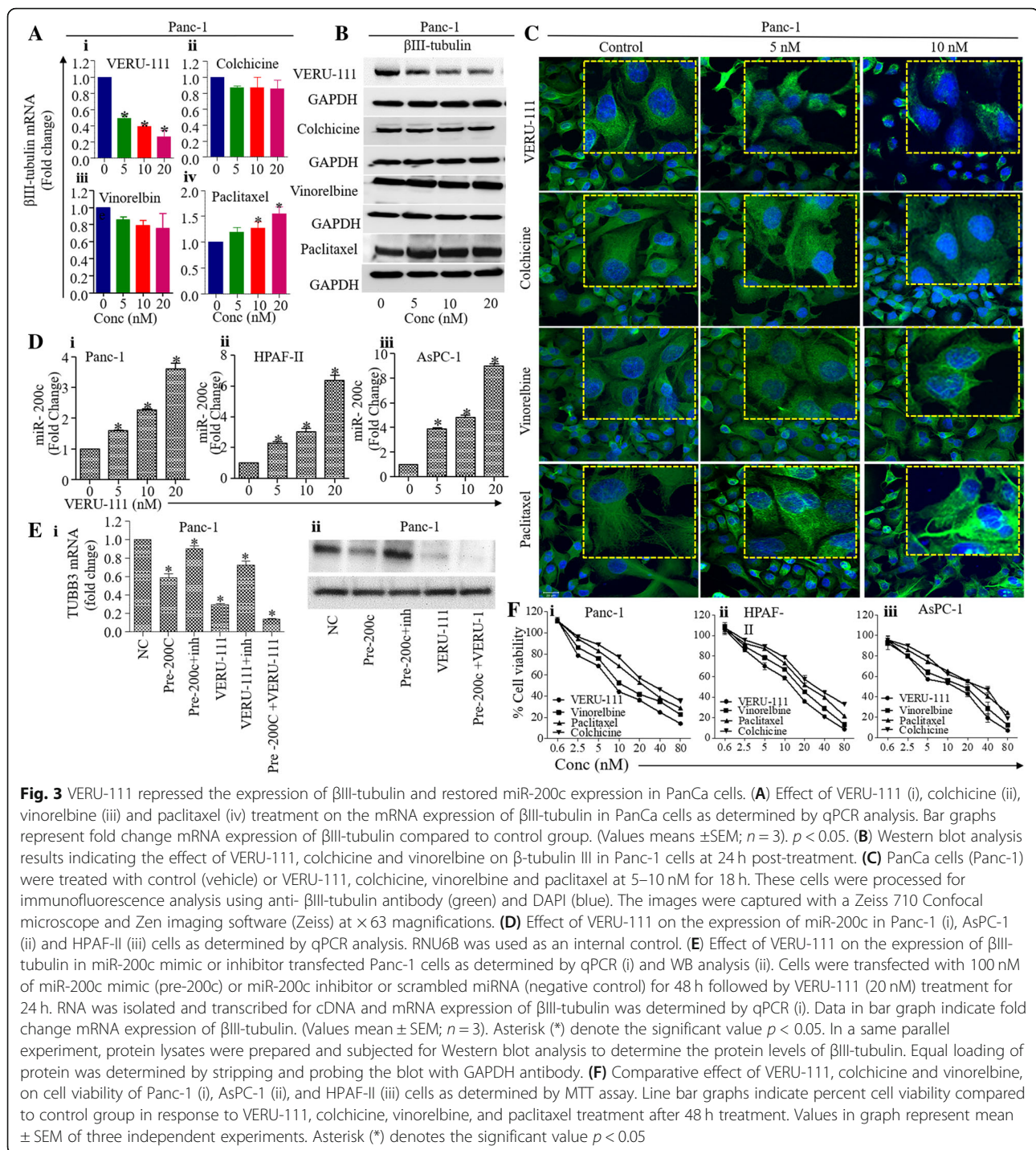
VERU-111 inhibits migration and invasive potential of PanCa cells

In addition to anti-proliferative activity, we elucidated anti-metastatic potential of VERU-111 by determining its effect on the invasive and migratory characteristics of PanCa cells. To address this, we first performed wound healing assays to determine the effect of VERU-111 on the migration of PanCa cells. Our results revealed remarkable inhibition in migration of both Panc-1 (Fig. 4Ai), AsPC-1 (Fig. 4Aii) and HPAF-II (Additional file 3: Figure S3A) cells when treated with sub-lethal concentrations of VERU-111 (1.25 and 2.5 nM). We further evaluated the effect of VERU-111 on PanCa cell migration by transwell assay. VERU-111 (1.25–2.5 nM) also showed significant ($p < 0.01$) inhibition of Panc-1, AsPC-1 (Fig. 4Bi-ii) and HPAF-II (Additional file 3: Figure S3Bi-ii) cell migration in a dose-dependent manner. VERU-111 at sub lethal concentrations (1.25–2.5 nM) also significantly ($P < 0.01$) inhibited invasion of Panc-1, AsPC-1 (Fig. 4Ci-ii) and HPAF-II (Additional file 3: Figure S3 Ci-ii) cells as compared to the vehicle treatment group. The effect of VERU-111 on migration and invasion of PanCa cells was

further confirmed using the xCELLigence system. VERU-111 also dose-dependently (5–20 nM) reduced the baseline cell index of PanCa cells as compared to control, which reflects potent inhibitory effects of VERU-111 on PanCa cells migration (Fig. 4Di) and invasion (Fig. 4Dii).

VERU-111 arrests cell cycle in G2/M phase and induces apoptosis in PanCa cells

It has been reported that tubulin-destabilizing agents block the cell cycle in the G2/M phase due to microtubule depolymerization and cytoskeleton disruption [34]. This prompted us to evaluate the effect of VERU-111 on PanCa cell cycle distribution using flow cytometry. VERU-111 treatment arrested Panc-1 and AsPC-1 cells in G2/M phase in a dose-dependent manner (Fig. 5Ai-ii) and Additional file 4: Figure S4Ai-ii). The complex formation between cdc2 and cyclin B1 is an important event for cell entry into mitosis [35, 36]. Thus, we determined the effect of VERU-111 on cell cycle regulatory proteins. VERU-111 effectively inhibited the expression of cyclin B1, cdc2 and cdc25c in PanCa (Panc-1 and AsPC-1) cells, while enhancing the phosphorylation of cdc2 (Fig. 5Bi-ii). Since, we observed arrest of cell cycle in G2/M phase, we subsequently investigated the effect of VERU-111 on apoptosis induction in PanCa cells by Annexin V-7AAD staining and



mitochondrial membrane potential ($\Delta\Psi_m$) using flow cytometer. As shown in Fig. 5C and (Additional file 5: Figure S5Ai-ii), VERU-111 treatment (5–20 nM) resulted in apoptosis induction in both Panc-1 and AsPC-1 cells. It has been shown that mitochondria plays key role in apoptosis through modulation of intrinsic signaling pathway mechanism and decrease in mitochondrial membrane potential ($\Delta\Psi_m$) has been considered as an

early event of apoptosis [37]. Thus, we examined the effect of VERU-111 on $\Delta\Psi_m$ in PanCa cells using TMRE staining. VERU-111 illustrated a dose-dependent (5–20 nM) decrease of TMRE staining in PanCa cells (Additional file 5: Figure S5Bi-ii). Additionally, we evaluated the effect of VERU-111 on other mitochondrial pro-apoptotic (Bax and Bad) and anti-apoptotic (Bcl2 and Bcl-xL) proteins. VERU-111 (5–20 nM)

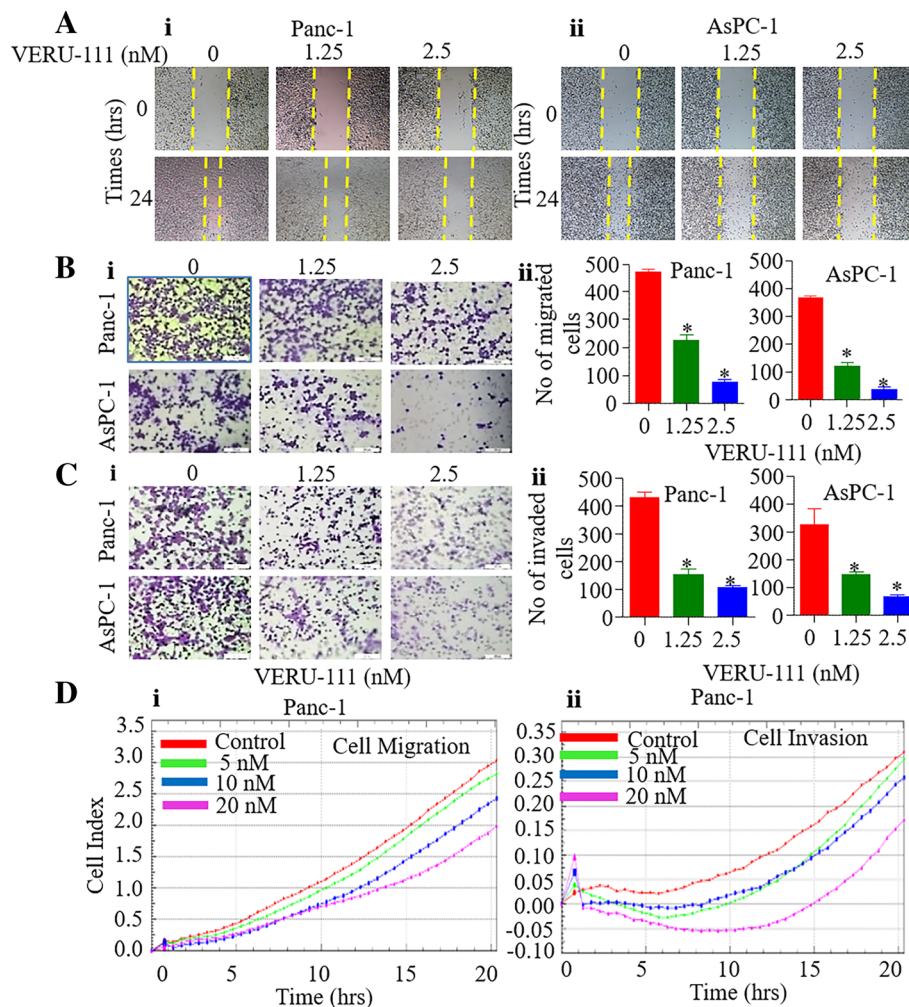
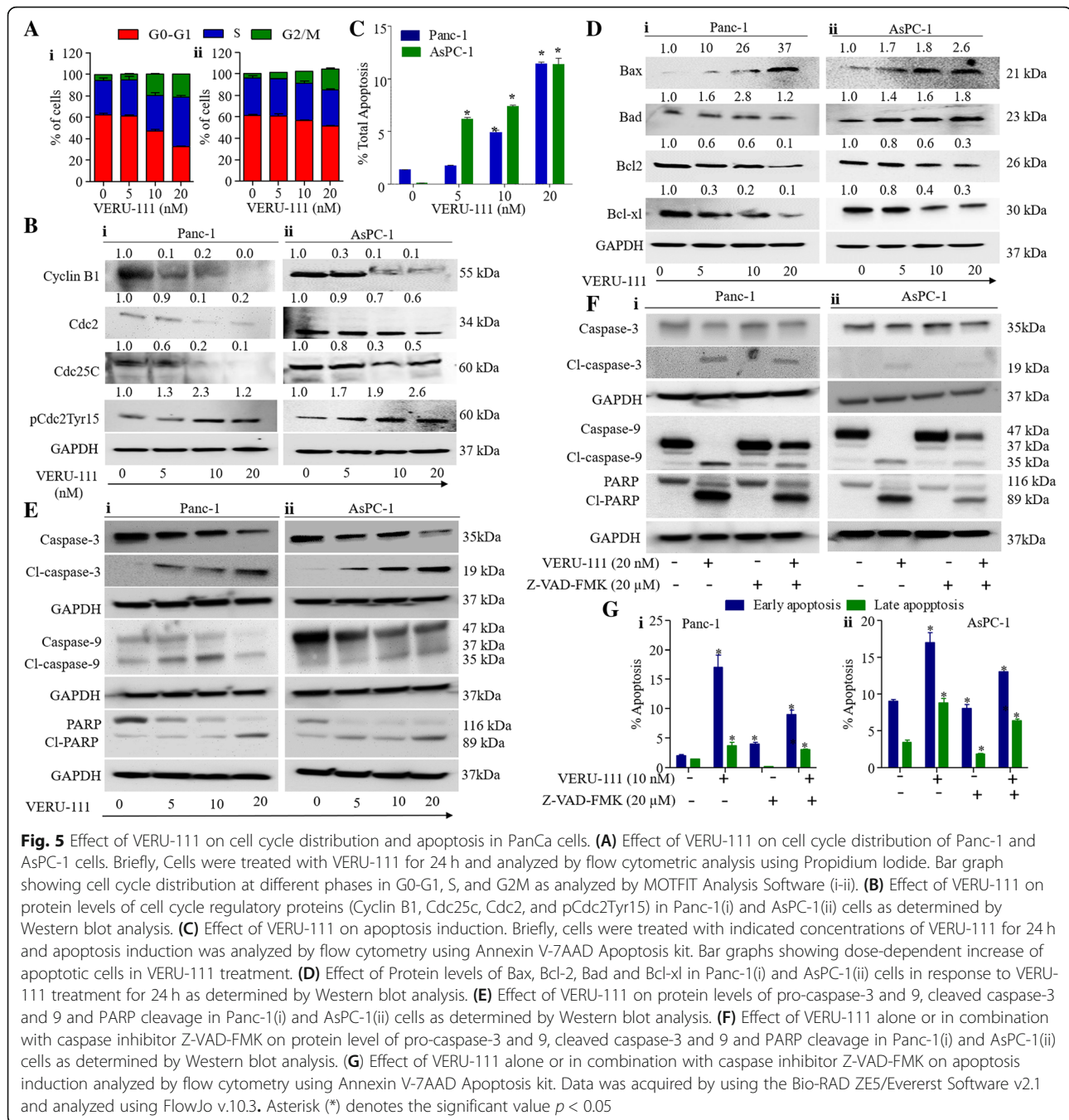


Fig. 4 Effect of VERU-111 on invasion and migration of PanCa cells. **(A)** Effect of VERU-111 on migration of Panc-1 and AsPC-1 cells determined by wound healing assays. The wounded monolayer was incubated in different concentrations of VERU-111 for 24 h. Images of wound healing assays (magnification, $\times 10$) of Panc-1 (i) and AsPC-1 (ii) cells of control and VERU-111 treatment groups after 24 h. **(B)** Effect of VERU-111 on migration of AsPC-1 and Panc-1 cells using 96-transwell chamber plate. Representative images of migratory Panc-1 and AsPC-1 cells (i) of control and VERU-111 treatment groups after 24 h. Bar graphs (ii) indicating number of migratory Panc-1 and AsPC-1 cells in control and VERU-111 treatment groups. **(C)** Effect of VERU-111 on invasion of Panc-1 and AsPC-1 cells (i) as determined by Boyden's Chamber assay. Representative images of control and VERU-111 treatment groups were captured at 20 \times magnification after 24 h. Bar graphs (ii) indicate number of invaded Panc-1 and AsPC-1 cells. **(D)** Effect of VERU-111 on real time migration (i) and invasion (ii) of Panc-1 cells using xCELLigence assay. Results are presented as means \pm SEM ($n = 3$). Asterisk (*) denotes the significant value $p < 0.05$

induced the expression of Bax and Bad and inhibited the expression of Bcl-2 and Bcl-xl proteins (Fig. 5Di-ii). We also investigated the effect of VERU-111 on pro-caspase-3 and 9, cleaved caspase-3 and 9 and PARP cleavage. VERU-111 treatment showed dose-dependent inhibition of pro-Caspase 3 and 9 and activation of Caspase-3 and 9 in both AsPC-1 and Panc-1 cells (Fig. 5Ei-ii). VERU-111 also showed cleavage of PARP protein in pancreatic cancer cells (Fig. 5Ei-ii). These results clearly indicate that VERU-111 induces apoptosis in pancreatic cancer cells. We further confirmed the involvement of caspase 3 and caspase 9 in VERU-111 induced apoptosis in PanCa cells.

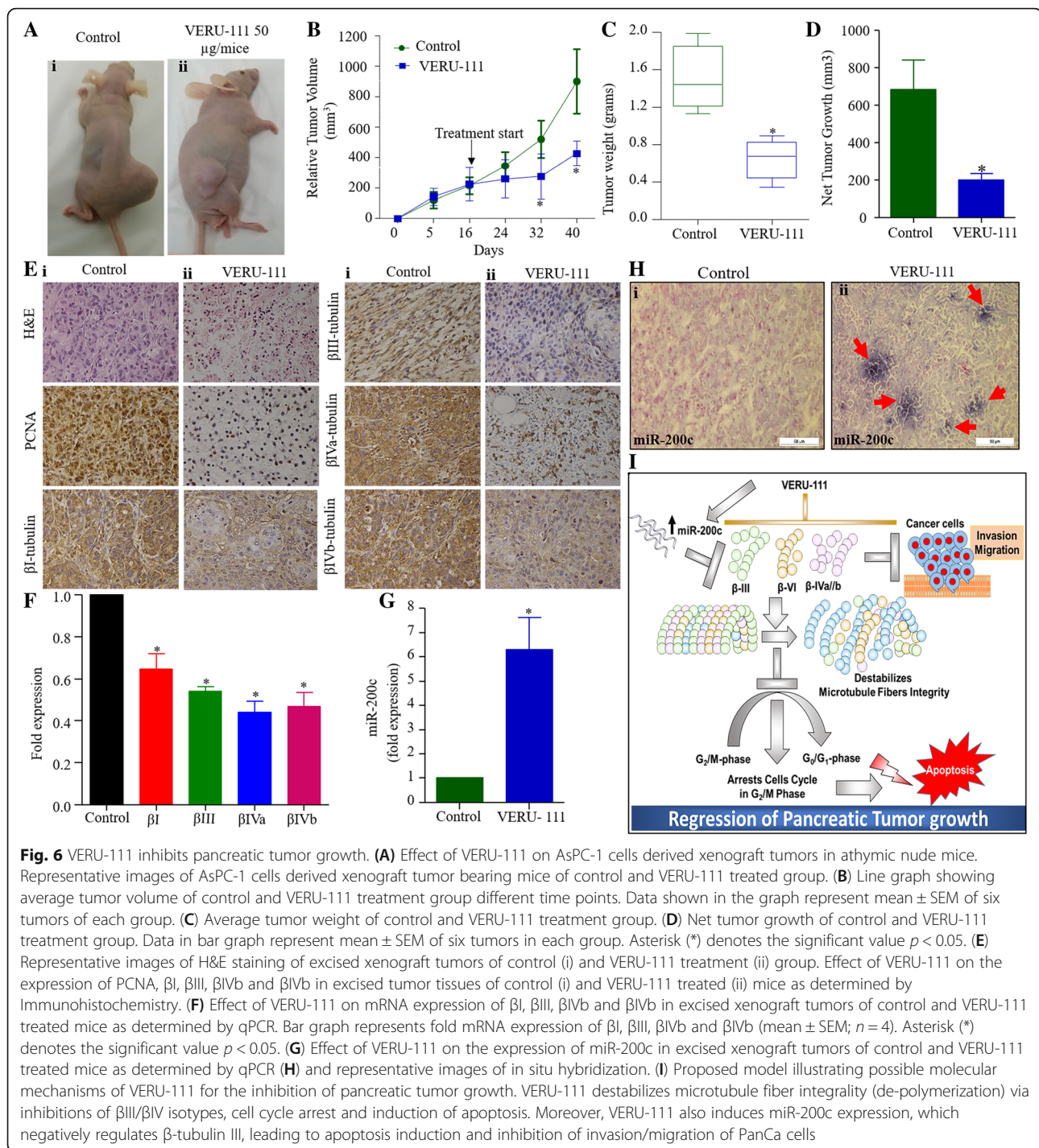
Our results illustrated that treatment of general caspase inhibitor Z-VAD-FMK rescued the VERU-111 induced apoptosis of PanCa cells (Fig. 5F-G). Treatment of PanCa cells with VERU-111 (20 nM) showed increase expression of cleaved caspase 3, 9 and PARP proteins which was decreased with the addition of Z-VAD-FMK (Fig. 5Fi-ii). We also observed significant decrease number of apoptotic PanCa cells with the treatment of Z-VAD-FMK when compared to VERU-111 treatment alone (Fig. 5Gi-ii and Additional file 5: Figure S5 Ci-ii). These results clearly suggest the involvement of caspase 3 and 9 in VERU-111 induced apoptosis of PanCa cells.



VERU-111 effectively inhibits the growth of pancreatic tumors in a xenograft mouse model

We next evaluated the therapeutic effect of VERU-111 in a pre-clinical mouse model of PanCa. In this experiment, highly aggressive AsPC-1 cells (2×10^6) were ectopically injected in athymic nude mice to generate xenograft tumors. VERU-111 (50 $\mu\text{g}/\text{mice}$) and its respective vehicle controls (PBS) were administered intra-tumorally once tumor volume reached $\sim 200 \text{ mm}^3$ (3 times per week for 3 weeks). VERU-111 treatment

effectively inhibited tumor growth as compared to vehicle-treated group (Fig. 6A-D). The average tumor volume in control mice reached to 900 mm^3 within 5 weeks, while at this time it was only 400 mm^3 in VERU-111 treated mice (Fig. 6B). None of the mouse showed any apparent toxicity as we observed constant increase of body weight in VERU-111 treated mice. (Additional file 6: Figure S6). PCNA is one of the markers of cell proliferation, which is upregulated in tumor cells, thus we evaluated if VERU-111 treatment



inhibits PCNA expression in tumor tissues. Our IHC analysis demonstrated considerable inhibition of PCNA expression in VERU-111 treated tumor tissues compared to vehicle control treated tumors (Fig. 6E). In accordance with our in vitro data, we also observed the repression of β III and β IVa and β IVb-tubulins in VERU-111 treated tumors both protein (determined by IHC) and mRNA levels (Fig. 6E, F). Interestingly, VERU-111

treatment also restored the expression of miR-200c in the excised xenograft tumors as determined by qPCR (Fig. 6G) and in situ hybridization (Fig. 6H) assays. In conclusion, we find that VERU-111 disrupts the microtubular dynamics via differential inhibition of β I, β III, β IVa, β IVb, and β VI isotopes, G2/M cell cycle arrest, induction of apoptosis and restoration of the miR-200c expression. Overall, these results determined the

therapeutic efficacy of a novel tubulin inhibitor (VERU-111) in PanCa models and elucidated its putative cellular and molecular mechanisms (Fig. 6I).

Discussion

Therapeutic options for pancreatic cancer (PanCa) are very limited, thus it is one of the leading causes of cancer-related mortalities in the United States [38]. Gemcitabine is a Food and Drug Administration (FDA) approved drug for the treatment of local and advanced metastatic PanCa, however its therapeutic efficacy in clinical setting is limited due to emergence of chemoresistance and systemic toxicity [39, 40]. The combination treatment strategy regimen of albumin bound paclitaxel (Abraxane®) along with gemcitabine has led to a 2.5 month increase in overall survival of PanCa patients [41]. It has been shown that Abraxane® increases tumor uptake of gemcitabine via inhibiting gemcitabine metabolizing enzyme (cytidine deaminase) [42]. In spite of these encouraging results, unwanted systemic toxicities of this combination therapy is an unmet clinical challenge.

Tubulin targeting taxanes (such as paclitaxel, docetaxel) have shown great clinical success as anti-cancer agents, which suggest importance of tubulins in cancer therapeutics [43]. However, being Pan-tubulin modulators (stabilizers or inhibitors), these agents suffer from significant systemic toxicities. To address this important issue, investigations are being carried out towards identification of cancer specific molecular targets. Like other cancers, tubulins play a major role in the development and progression of PanCa [10]. However, among all tubulins, β III and β IV isotypes play major role in PanCa progression, metastasis and chemoresistance [32]. Therefore, selective targeting of β III and β IV-tubulins may improve the therapeutic response of PanCa. In this study, we have identified a novel β III and β IV inhibitor (VERU-111), determined its therapeutic efficacy and investigated underlying cellular/molecular mechanisms using clinically relevant cell lines and preclinical mice PanCa models. Our results demonstrate that VERU-111 can effectively inhibit the growth of highly aggressive PanCa cells. To determine the specificity of β -tubulins targeting, we evaluated effect of VERU-111 on the expression of all β -tubulin isotypes by qPCR and Western blot analysis. These analyses suggest predominant inhibitory effect of this small molecule on the expression of β III and β IV isotypes and differential inhibition of (Fig. 2A-B), which are known to be involved in PanCa growth, metastasis and chemoresistance [44]. Our data also indicate that VERU-111 also differentially inhibits the expression of β I, and β VI isoforms. However, role of these tubulins in PanCa is not defined yet. Studies have also shown that the β III tubulin expression level alters the sensitivity of microtubule destabilizing

agents [45, 46]. Notably, available microtubule destabilizing agents such as vincristine, vinorelbine and nocodazole interact very weakly with β III tubulin compared with other β -tubulin isotypes [47], which is clearly reflected in our comparative analysis of VERU-111 with colchicine, and vinorelbine. Our results demonstrated that VERU-111 is a more potent destabilizing agent of β III/ β IV tubulin isotypes thus has profound therapeutic efficacy in PanCa models.

We also elucidated underlying cellular/molecular mechanisms of VERU-111 mediated targeting of PanCa associated β -tubulin isotypes (β III/ β IV). It has been shown that β III-tubulin is a direct target of miR-200c, thus its overexpression inhibits the metastatic phenotype of cancer cells and enhances sensitivity to chemotherapeutic agents [33]. Our results indicate that VERU-111 significantly ($p < 0.01$) restores the expression of miR-200c in PanCa cells/tumors, while miR-200c inhibitor diminish the effect of VERU-111 on the expression of β -III tubulin. These results strongly suggest that VERU-111 targets β III-tubulin via replenishment of the expression of miR-200c. The invasive and migratory characteristics of cancer cells represent their metastatic phenotypes, which is an important step in cancer metastasis [48]. It has been shown that the overexpression of β -tubulins, specifically β III and β IV, are involved in enhancing the metastatic potential of PanCa cells [44]. Our functional assays indicate that VERU-111 effectively inhibits invasive and migratory potential of PanCa cells. The effect of VERU-111 on cell cycle analysis was evaluated as it has been well documented that tubulin targeting drugs arrest cell cycle in the G2/M phase. G2 arrest of cell cycle prevent cancer cells from repairing DNA damage, forcing them into M phase; thus, the G2/M checkpoint is an ideal therapeutic target for anti-cancer drugs [49]. Our result indicates that VERU-111 arrested the cell cycle in the G2/M phase, which is in accordance with other tubulin targeting agents [49]. It has been reported that different class of cyclins and their cyclin-dependent kinases such as cyclin B1, Cdc2 and Cdc25C are involved in cell cycle progression [50, 51]. The inhibitory effect of VERU-111 on the expression of cyclin B1, Cdc2 and Cdc25C kinases explained its putative molecular effects on the cell cycle. Accumulating evidence suggests that G2/M cell cycle arrest leads to the induction of apoptosis, a mechanism of cell death. Our flow cytometry data indicate enhanced apoptosis in PanCa cells with the treatment of VERU-111. Bcl2 is an upstream effector molecule in the apoptotic pathway and has been recognized as a potent inhibitor of apoptosis and shown to be overexpressed in various types of cancer including PanCa [32, 52]. It forms a heterodimer with the apoptotic protein Bax, thereby neutralizing its apoptotic effects. Therefore, alteration in the ratio of Bax/Bcl2 is a crucial factor that plays an important role to determine whether cells will undergo

apoptosis. Our results indicate that VERU-111 induces apoptosis via modulating mitochondrial proteins (Bcl2, Bcl-xL, Bax and Bad), activation of caspase 3 and 9 and PARP protein cleavage suggesting the involvement of an intrinsic apoptotic pathway. Our results also demonstrate that Z-VAD-FMK treatment rescue the PanCa cells from VERU-111 induce apoptosis. These results clearly suggest involvement of caspase 3 and 9 in VERU-111 induced apoptosis of PanCa cells. Our xenograft mouse model studies demonstrate that VERU-111 (50 µg/mice) effectively inhibits tumor growth with concomitant inhibition of βI, βIII and βIV tubulins and restoring expression of miR-200c. These results indicate that VERU-111 inhibits pancreatic tumor growth via inducing cell cycle arrest, apoptosis and restoration of miR-200c in PanCa cells. These results strongly suggest potent therapeutic efficacy of VERU-111 for PanCa treatment and further studies are warranted to translate this novel molecule for clinical use in future.

Conclusion

Specific inhibitors of βIII and βIV isoforms could be an ideal candidate for the management of PanCa and overcoming chemo-resistance. We for the first time, demonstrate that VERU-111 is a novel synthetic compound, which preferentially targets βIII and βIV tubulin isotypes via restoration of miR-200c in PanCa cells. Overall, this study suggests that VERU-111 is a novel promising small molecule inhibitor of βIII and βIV tubulin isotypes, which could be useful as a monotherapy or in combination with conventional therapeutic regimens for the treatment of PanCa.

Additional files

Additional file 1: Figure S1. (A) Synthesis and chemical structure of VERU-111. (B) ¹H NMR characterization of VERU-111 using CD₂Cl₂ as the NMR solvent and full assignment of proton chemical shifts of VERU-111 (EtOAc: ethyl acetate). Spectrum was acquired on a Bruker Avance III 400 MHz NMR spectrometer equipped with a BBO probe at room temperature. (TIF 1059 kb)

Additional file 2: Table S1. List of Human β-Tubulin Isoforms qRT-PCR Primers used in this study. ^aIn a number of locations Inosine is inserted to break up runs of four or more Gs that might lead to unusually stable secondary structure.

Additional file 3: Figure S3. Effect of VERU-111 on invasion and migration of HPAF-II cells (A) Effect of VERU-111 on migration of HPAF-II cells as determined by scratch wound assay. Briefly, cells were seeded into 12-well plates and cultured up to 90% confluency. Cell scratch wound line in each well was generated using 200 µl pipette tip. The wounded cells monolayer were treated with indicated concentrations of VERU-111 for 24 h. Representative images (10x magnification) of HPAF-II cells were captured by phase contrast microscope at 0 and 24 h. (B) Effect of VERU-111 on migration of HPAF-II cells using 96-transwell chamber plate. Representative images of migratory HPAF-II cells of control and VERU-111 treatment groups after 24 h (i). Bar graphs (ii) indicating number of migratory HPAF-II cells in control and VERU-111 treatment groups. (C) Effect of VERU-111 on invasion of HPAF-II cells (i) as determined by Matrigel

Invasion assay. Representative images of control and VERU-111 treatment groups were captured at 10x magnification after 24 h. Bar graphs (ii) indicate number of invaded HPAF-II cells. Results are presented as means ± SEM of three independent experiments. Asterisk (*) denotes the significant value $P < 0.05$. (TIF 3005 kb)

Additional file 4: Figure S4. Effect of VERU-111 on cells cycle distribution. (A) Effect of VERU-111 on cell cycle distribution of Panc-1 and AsPC-1 cells. Briefly, cells were treated with VERU-111 for 24 h. Cells in different phase was analyzed by flow cytometric analysis using Propidium Iodide. Representative images of histogram showing cell cycle distribution at different phases in Panc-1 (i) and AsPC-1(ii) cells. (TIF 1759 kb)

Additional file 5: Figure S5. Effect of VERU-111 on apoptosis induction in PanCa. (A) Effect of VERU-111 on apoptosis induction of Panc-1 and AsPC-1 cells. Briefly, cells were treated with indicated concentrations of VERU-111 for 24 h and apoptosis induction was analyzed by flow cytometry using Annexin V-7AAD Apoptosis kit. Data was acquired by using the Bio-RAD ZES/Everest Software v2.1 and analyzed using FlowJo v.10.3. (B) Effect of VERU-111 on mitochondrial membrane potential ($\Delta\Psi_m$) in Panc-1 and AsPC-1 cells as determined by TMRE staining. Representative images from three independent experiments are showing dose-dependent decrease of TMRE staining in Panc-1 and AsPC-1 cells (i). Bar graph showing dose-dependent decrease of $\Delta\Psi_m$ as determined by quantitative analysis of TMRE staining by flow cytometry in Panc-10 and AsPC-1 (ii). Data represented as mean ± SEM of 3 independent experiments. Asterisk (*) denotes the significant value $p < 0.05$. C. Effect of VERU-111 alone or in combination with Z-VAD-FMK on apoptosis of PanCa. The cells were pretreated with Z-VAD-FMK for 2 h followed by VERU-111 (20 µM) for 24 h and apoptosis induction was analyzed by flow cytometry using Annexin V-7AAD Apoptosis kit. Representative images of histogram showing increase of apoptotic cells and data was acquired by using the Bio-RAD ZES/Everest Software v2.1 and analyzed using FlowJo v.10.3. (D) Quantitation of Western blots indicated in Fig. 5 E and F. The density ratio of pro-caspase-3 and 9, cleaved caspase-3 and 9 and PARP cleavage treated with different concentrations of VERU-111 (i) and general caspase inhibitor Z-VAD-FMK (20 µM for 2 h) followed by VERU-111 (20 nM) treatment for 24 h in PanCa cells (ii). Values are expressed as means ± SD. Experiments were repeated 3 times. Asterisk (*) denotes the significant value $P < 0.05$. (TIF 1351 kb)

Additional file 6: Figure S6. Effect of VERU-111 on weight of mice. AsPC-1 cells (2×10^6 cells) were injected subcutaneously on the dorsal flanks of each mice. Mice were administered with VERU-111 (50 µg/mouse/week for three weeks i.e. 3 times per week for 3 weeks). Control group mice were administered with vehicle. Body weight of both the groups' mice was recorded once in a week. Line graph representing constant increase in body weight of both the groups' mice. Data represent mean ± SD value of $n = 6$ mice in each group. (TIF 464 kb)

Additional file 7: Figure S2. Western blot internal control (GAPDH) of various β-tubulin isotypes treated with VERU-111 in PanCa cells. Panc-1 (i) and AsPC-1(ii) cells. Were treated with vehicle or indicated concentrations of VERU-111 for 24 h. Cell lysates were prepared and 40 µg protein was subjected for Western blot analysis. Equal loading of protein in each well was confirmed by stripping and re-probing the blots with GAPDH. Each experiment was repeated two times and similar results were obtained. (TIF 1222 kb)

Abbreviations

FBS: Fetal bovine serum; FDA: Food and Drug Administration; IHC: Immunohistochemistry; ISH: In situ hybridization; PanCa: Pancreatic Cancer; PBS: Phosphate saline buffer; PI: Propidium iodide; PMSF: Phenylmethanesulfonyl fluoride; qRT-PCR: Quantitative-Reverse Transcription Polymerase Chain Reaction; TMRE: tetramethylrhodamine; UTHSC-IACUC: UTHSC Institutional Animal Care and Use Committee and

Acknowledgements

None.

Funding

This work was supported by NIH/NC1 grants (R01 CA210192, R01 CA206069 and R01 CA204552) to SCC, R01 CA148706 to WL and DDM, 1510OD010678

and RR026377 to WL, and College of Pharmacy/University of Tennessee Health Science Center Seed Grant to MJ and BBH.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Author's contributions

Conceived and designed the experiments: SCC, WL, and BBH conceived the idea. SCC, VKK, NC, BBH, WL, DDM performed the experiments. VKK, BBH, QW, PS, SS, SK PC, and MJ analyzed the data. VKK, BBH, QW, SSC, WL wrote the paper. SCC, MJ and BBH supervised the research. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The research protocol was reviewed and approved by the UTHSC-IACUC protocol.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 31 August 2018 Accepted: 17 December 2018

Published online: 23 January 2019

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin*. 2016; 66:7–30.
2. Ansari D, Gustafsson A, Andersson R. Update on the management of pancreatic cancer: surgery is not enough. *World J Gastroenterol*. 2015;21: 3157–65.
3. Sorger PK, Dobles M, Tournebise R, Hyman AA. Coupling cell division and cell death to microtubule dynamics. *Curr Opin Cell Biol*. 1997;9:807–14.
4. Horio T, Murata T. The role of dynamic instability in microtubule organization. *Front Plant Sci*. 2014;5:511.
5. Bernard-Marty C, Treilleux I, Dumontet C, Cardoso F, Fellous A, Gancberg D, Bissery MC, Paesmans M, Larsimont D, Piccart MJ, Di Leo A. Microtubule-associated parameters as predictive markers of docetaxel activity in advanced breast cancer patients: results of a pilot study. *Clin Breast Cancer*. 2002;3:341–5.
6. Moiseyenko VM, Volkov NM, Suspistin EN, Yanus GA, Iyevleva AG, Kuligina E, Togo AV, Kornilov AV, Ivantsov AO, Imyanitov EN. Evidence for predictive role of BRCA1 and bTUBIII in gastric cancer. *Med Oncol*. 2013;30:545.
7. Mozzetti S, Ferlini C, Concolino P, Filippetti F, Raspaglio G, Prislei S, Gallo D, Martinelli E, Ranelletti FO, Ferrandina G, Scambia G. Class III beta-tubulin overexpression is a prominent mechanism of paclitaxel resistance in ovarian cancer patients. *Clin Cancer Res*. 2005;11:298–305.
8. Seve P, Dumontet C. Is class III beta-tubulin a predictive factor in patients receiving tubulin-binding agents? *Lancet Oncol*. 2008;9:168–75.
9. Seve P, Isaac S, Tredan O, Souquet PJ, Pacheco Y, Perol M, Lafanechere L, Penet A, Peiller EL, Dumontet C. Expression of class III {beta}-tubulin is predictive of patient outcome in patients with non-small cell lung cancer receiving vinorelbine-based chemotherapy. *Clin Cancer Res*. 2005;11:5481–6.
10. Kavallaris M. Microtubules and resistance to tubulin-binding agents. *Nat Rev Cancer*. 2010;10:194–204.
11. Karki R, Mariani M, Andreoli M, He S, Scambia G, Shahabi S, Ferlini C. betaIII-Tubulin: biomarker of taxane resistance or drug target? *Expert Opin Ther Targets*. 2013;17:461–72.
12. Atjanasupatt K, Lirdprapamongkol K, Jantaree P, Svasti J. Non-adherent culture induces paclitaxel resistance in H460 lung cancer cells via ERK-mediated up-regulation of betaIVa-tubulin. *Biochem Biophys Res Commun*. 2015;466:493–8.
13. Shalli K, Brown I, Heys SD, Schofield AC. Alterations of beta-tubulin isotypes in breast cancer cells resistant to docetaxel. *FASEB J*. 2005;19:1299–301.
14. Lee KM, Cao D, Itami A, Pour PM, Hruban RH, Maitra A, Ouellette MM. Class III beta-tubulin, a marker of resistance to paclitaxel, is overexpressed in pancreatic ductal adenocarcinoma and intraepithelial neoplasia. *Histopathology*. 2007;51:539–46.
15. McCarroll JA, Sharbeen G, Liu J, Youkhana J, Goldstein D, McCarthy N, Limbri LF, Dischl D, Ceyhan GO, Erkan M, et al. betaIII-tubulin: a novel mediator of chemoresistance and metastases in pancreatic cancer. *Oncotarget*. 2015;6:2235–49.
16. Kavallaris M, Kuo DY, Burkhart CA, Regl DL, Norris MD, Haber M, Horwitz SB. Taxol-resistant epithelial ovarian tumors are associated with altered expression of specific beta-tubulin isotypes. *J Clin Invest*. 1997;100:1282–93.
17. Ranganathan S, Benetatos CA, Colarusso PJ, Dexter DW, Hudes GR. Altered beta-tubulin isotype expression in paclitaxel-resistant human prostate carcinoma cells. *Br J Cancer*. 1998;77:562–6.
18. Gan PP, Kavallaris M. Tubulin-targeted drug action: functional significance of class II and class IVb beta-tubulin in vinca alkaloid sensitivity. *Cancer Res*. 2008;68:9817–24.
19. Miller LM, Menthen A, Chatterjee C, Verdier-Pinard P, Novikoff PM, Horwitz SB, Angeletti RH. Increased levels of a unique post-translationally modified betaIVb-tubulin isotype in liver cancer. *Biochemistry*. 2008;47:7572–82.
20. Ranganathan S, Dexter DW, Benetatos CA, Chapman AE, Tew KD, Hudes GR. Increase of beta (III)- and beta (IVa)-tubulin isotypes in human prostate carcinoma cells as a result of estramustine resistance. *Cancer Res*. 1996;56:2584–9.
21. Wu X, Wang Q, Li W. Recent advances in heterocyclic tubulin inhibitors targeting the colchicine binding site. *Anti Cancer Agents Med Chem*. 2016;16:1325–38.
22. Chen J, Wang Z, Li CM, Lu Y, Vaddady PK, Meibohm B, Dalton JT, Miller DD, Li W. Discovery of novel 2-aryl-4-benzoyl-imidazoles targeting the colchicines binding site in tubulin as potential anticancer agents. *J Med Chem*. 2010;53:7414–27.
23. Chen J, Li CM, Wang J, Ahn S, Wang Z, Lu Y, Dalton JT, Miller DD, Li W. Synthesis and antiproliferative activity of novel 2-aryl-4-benzoyl-imidazole derivatives targeting tubulin polymerization. *Bioorg Med Chem*. 2011;19:4782–95.
24. Chen J, Ahn S, Wang J, Lu Y, Dalton JT, Miller DD, Li W. Discovery of novel 2-aryl-4-benzoyl-imidazole (ABI-III) analogues targeting tubulin polymerization as antiproliferative agents. *J Med Chem*. 2012;55:7285–9.
25. Nagesh PKB, Hatami E, Chowdhury P, Kashyap VK, Khan S, Hafeez BB, Chauhan SC, Jaggi M, Yallapu MM. Tannic acid induces endoplasmic reticulum stress-mediated apoptosis in prostate Cancer. *Cancers (Basel)*. 2018;10.
26. Lobert S, Hiser L, Correia JJ. Expression profiling of tubulin isotypes and microtubule-interacting proteins using real-time polymerase chain reaction. *Methods Cell Biol*. 2010;95:47–58.
27. Kumari S, Khan S, Gupta SC, Kashyap VK, Yallapu MM, Chauhan SC, Jaggi M. MUC13 contributes to rewiring of glucose metabolism in pancreatic cancer. *Oncogenesis*. 2018;7:19.
28. Khan S, Ebeling MC, Chauhan N, Thompson PA, Gara RK, Ganju A, Yallapu MM, Behrman SW, Zhao H, Zafar N, et al. Ormeloxifene suppresses desmoplasia and enhances sensitivity of gemcitabine in pancreatic cancer. *Cancer Res*. 2015;75:2292–304.
29. Bharti R, Dey G, Banerjee I, Dey KK, Parida S, Kumar BN, Das CK, Pal I, Mukherjee M, Misra M, et al. Somatostatin receptor targeted liposomes with Diacerein inhibit IL-6 for breast cancer therapy. *Cancer Lett*. 2017;388:292–302.
30. Chauhan SC, Ebeling MC, Maher DM, Koch MD, Watanabe A, Aburatani H, Lio Y, Jaggi M. MUC13 mucin augments pancreatic tumorigenesis. *Mol Cancer Ther*. 2012;11:24–33.
31. Hafeez BB, Ganju A, Sikander M, Kashyap VK, Hafeez ZB, Chauhan N, Malik S, Massey AE, Tripathi MK, Halawish FT, et al. Ormeloxifene suppresses prostate tumor growth and metastatic phenotypes via inhibition of oncogenic beta-catenin signaling and EMT progression. *Mol Cancer Ther*. 2017;16:2267–80.
32. Parker AL, Teo WS, McCarroll JA, Kavallaris M. An emerging role for tubulin isotypes in modulating Cancer biology and chemotherapy resistance. *Int J Mol Sci*. 2017;18.
33. Cochrane DR, Spoelstra NS, Howe EN, Nordeen SK, Richer JK. MicroRNA-200c mitigates invasiveness and restores sensitivity to microtubule-targeting chemotherapeutic agents. *Mol Cancer Ther*. 2009;8:1055–66.
34. Clarke PR, Allan LA. Cell-cycle control in the face of damage—a matter of life or death. *Trends Cell Biol*. 2009;19:89–98.

35. Kiyokawa H, Ray D. In vivo roles of CDC25 phosphatases: biological insight into the anti-cancer therapeutic targets. *Anti Cancer Agents Med Chem.* 2008;8:832–6.
36. Afshari CA, Barrett JC. Cell cycle controls: potential targets for chemical carcinogens? *Environ Health Perspect.* 1993;101(Suppl 5):9–14.
37. Green DR, Kroemer G. The pathophysiology of mitochondrial cell death. *Science.* 2004;305:626–9.
38. Merika EE, Syrigos KN, Saif MW. Desmoplasia in pancreatic cancer. Can we fight it? *Gastroenterol Res Pract.* 2012;2012:781765.
39. Amrutkar M, Gladhaug IP. Pancreatic Cancer Chemoresistance to gemcitabine. *Cancers (Basel).* 2017;9.
40. Burris HA 3rd, Moore MJ, Andersen J, Green MR, Rothenberg ML, Modiano MR, Cripps MC, Portenoy RK, Storniolo AM, Tarassoff P, et al. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J Clin Oncol.* 1997;15:2403–13.
41. Pedersen AG. Phase I studies of gemcitabine combined with carboplatin or paclitaxel. *Semin Oncol.* 1997;24:57-64-567-68.
42. Frese KK, Neesse A, Cook N, Bapiro TE, Lolkema MP, Jodrell DI, Tuveson DA. Nab-paclitaxel potentiates gemcitabine activity by reducing cytidine deaminase levels in a mouse model of pancreatic cancer. *Cancer Discov.* 2012;2:260–9.
43. Ojima I, Lichtenthal B, Lee S, Wang C, Wang X. Taxane anticancer agents: a patent perspective. *Expert Opin Ther Pat.* 2016;26:1–20.
44. Sharbeen G, McCarroll J, Liu J, Youkhana J, Limbri LF, Biankin AV, Johns A, Kavallaris M, Goldstein D, Phillips PA. Delineating the role of betaIV-tubulins in pancreatic Cancer: betaIVb-tubulin inhibition sensitizes pancreatic Cancer cells to Vinca alkaloids. *Neoplasia.* 2016;18:753–64.
45. Banerjee A, D'Hoore A, Engelborghs Y. Interaction of desacetamidocolchicine, a fast binding analogue of colchicine with isotypically pure tubulin dimers alpha beta II, alpha beta III, and alpha beta IV. *J Biol Chem.* 1994;269:10324–9.
46. Banerjee A, Engelborghs Y, D'Hoore A, Fitzgerald TJ. Interactions of a bicyclic analog of colchicine with beta-tubulin isoforms alphabeta (II), alphabeta (III) and alphabeta (IV). *Eur J Biochem.* 1997;246:420–4.
47. Banerjee A, Luduena RF. Kinetics of colchicine binding to purified beta-tubulin isotypes from bovine brain. *J Biol Chem.* 1992;267:13335–9.
48. van Zijl F, Krupitza G, Mikulits W. Initial steps of metastasis: cell invasion and endothelial transmigration. *Mutat Res.* 2011;728:23–34.
49. Bucher N, Britten CD. G2 checkpoint abrogation and checkpoint kinase-1 targeting in the treatment of cancer. *Br J Cancer.* 2008;98:523–8.
50. Kumagai A, Dunphy WG. The cdc25 protein controls tyrosine dephosphorylation of the cdc2 protein in a cell-free system. *Cell.* 1991;64:903–14.
51. Strausfeld U, Labbe JC, Fesquet D, Cavadore JC, Picard A, Sadhu K, Russell P, Doree M. Dephosphorylation and activation of a p34cdc2/cyclin B complex in vitro by human CDC25 protein. *Nature.* 1991;351:242–5.
52. Murphy KM, Ranganathan V, Farnsworth ML, Kavallaris M, Lock RB. Bcl-2 inhibits Bax translocation from cytosol to mitochondria during drug-induced apoptosis of human tumor cells. *Cell Death Differ.* 2000;7:102–11.

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