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DNA cyclization and looping in the wormlike limit: normal modes and the validity of the harmonic approximation

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Abstract

For much of the last three decades Monte Carlo-simulation methods have been the standard approach for accurately calculating the cyclization probability, J, or J factor, for DNA models having sequence-dependent bends or inhomogeneous bending flexibility. Within the last ten years approaches based on harmonic analysis of semi-flexible polymer models have been introduced, which offer much greater computational efficiency than Monte Carlo techniques. These methods consider the ensemble of molecular conformations in terms of harmonic fluctuations about a welldefined elastic-energy minimum. However, the harmonic approximation is only applicable for small systems, because the accessible conformation space of larger systems is increasingly dominated by anharmonic contributions. In the case of computed values of the J factor, deviations of the harmonic approximation from the exact value of J as a function of DNA length have not been characterized. Using a recent, numerically exact method that accounts for both anharmonic and harmonic contributions to J for wormlike chains of arbitrary size, we report here the apparent error that results from neglecting anharmonic behavior. For wormlike chains having contour lengths less than four times the persistence length the error in Jarising from the harmonic approximation is generally small, amounting to free energies less than the thermal energy, k_BT . For larger systems, however, the deviations between harmonic and exact J values increase approximately linearly with size.

I. INTRODUCTION

We recount in this section some key developments in the theoretical treatment of DNA cyclization since the early 1980s, emphasizing the role that Don Crothers and his collaborators played in advancing this field. Although the Crothers group also contributed greatly to the experimental DNA-cyclization literature (sometimes in papers that combined

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experiments and theory), we elected not to review those important contributions here in order to avoid a lengthy digression.

Rigorous experimental measurements of the cyclization free energy of DNA molecules have been carried out since the ground-breaking work of Shore *et al.*¹ and Shore and Baldwin.^{2,3} At that time experiments were well ahead of available theory for the dependence of the cyclization probability on DNA size and helical phase. The lack of a useful theoretical framework was due to the inherently challenging problem of modeling a stiff polymer chain having chain-end constraints in a way that also accounts for thermal fluctuations in such systems. The treatment of conformational fluctuations is important because they are significant even for DNA molecules with contour lengths substantially less than the polymer's persistence length.^{4,5}

A major theoretical advance came from the work of Shimada and Yamakawa,^{6,7} who developed a semi-numerical approach based on a series approximation to the cyclization probability of a uniform helical wormlike chain. The rigor and numerical accuracy of this theory permitted values of the persistence length, torsional rigidity, and helical repeat to be extracted from Shore and Baldwin's experimental data with confidence. However, there was a significant limitation to this approach, namely that the theory applied to an isotropically flexible polymer whose minimum-energy conformation in the linear state is that of a straight rod. By the early- to mid-1980s experiments showed that sequence-dependent conformational properties of DNA can contribute to deviations from the straight-rod minimum-energy state.^{8,9} These observations motivated the development of Monte Carlobased computational tools that could account for sequence-dependent effects on the cyclization probability.

Levene and Crothers^{10,11} used Monte Carlo simulation to compute the distributions of chain end-to-end distance, mutual orientations of chain termini, and an estimate of the writhe distribution in closed chains required to compute the cyclization probability.¹² These calculations were combined, under the assumption of independent twist and writhe variables, with an analytical treatment of the closed-chain twist distribution in order to arrive at a comprehensive treatment of the cyclization problem. The problem of sample attrition, which is significant in the case of limited Monte Carlo ensembles of chain conformations, was dealt with by using a combinatorial chain-dimerization method, first explored by Alexandrowicz.¹³ Around the same time, Hagerman,¹⁴ and later Ramadevi and Hagerman,¹⁵ published a Monte Carlo method that was based mainly on the contributions to the cyclization probability derived from the chain's axial degrees of freedom.

Motivated by the discovery of sequence-dependent intrinsic bends in DNA,⁸ the Monte Carlo method showed that an intrinsic DNA bend comparable in magnitude to one that was experimentally characterized¹⁶ can affect the value of the torsion-independent cyclization probability by fifty fold or more.¹⁰ This result suggested that cyclization measurements are a powerful method for measuring the extent of intrinsic DNA bending due either to DNA-sequence-dependent conformational preferences or bending distortion induced by site-specific protein binding. Measurements of DNA bending by ligase-catalyzed cyclization was

used with great effectiveness by Don Crothers and numerous coworkers during the last two decades of his career.^{17–23}

As with most simulation-based methods, computing cyclization probabilities by Monte Carlo techniques involves a trade-off between speed and accuracy. The large amounts of computer time required to calculate cyclization probabilities for any specified DNA conformation limited the theory's usefulness as a tool for fitting experimental data. This limitation led Zhang and Crothers to develop a new statistical-mechanical approach in the early 2000s that was based on approximating the chain-conformation distribution in terms of harmonic fluctuations about the chain's mechanical-energy minimum.²⁴ Termed the harmonic approximation (HA), this method was the natural successor to the Shimada-Yamakawa theory, but had the virtue that calculations could be done for inhomogeneous DNA conformations having non-zero values of the helical parameters tilt and roll, or sequence-dependent elastic-energy constants. The harmonic approximation was subsequently extended to the problem of protein-mediated DNA looping, wherein the protein assembly mediating the loop was treated as a connected set of rigid subunits interacting through the same harmonic expression that governs interactions between base pairs in the DNA loop.²⁵ The entire looped structure, DNA and associated protein subunits, was treated as a circular polymer with the virtual chain segments defining protein-DNA and protein-protein contacts assumed to adopt non-canonical helical parameters. HA calculations are about four orders of magnitude faster than Monte Carlo simulations of the same system, which made it possible to carry out systematic analyses of lac-repressor-mediated DNA looping in vitro²⁶ and in vivo.²⁷

The applicability of HA is limited to systems for which the accessible conformation space is dominated by harmonic fluctuations about a well-defined elastic-energy minimum. This assumption is justified, *e.g.*, for biopolymers with $L \ll P$ where *L* is the contour length and *P* is the persistence length. However, for larger systems HA results are expected to deviate from exact behavior because for increasing ratios L/P the accessible conformation space is increasingly dominated by anharmonic contributions. For example, in the limit of long, flexible polymer chains the entropy of a circular chain of contour length *L decreases* relative to the entropy of a linear chain of same length as $-3/2 \ln (L)$;^{28,29} this logarithmic decrease as a function of *L* is absent in the HA result for the entropy of this system. The accuracy and applicability of HA as a function of system size, for example the contour length *L* of a cyclized DNA or a DNA loop, have not been systematically investigated. HA therefore remains an uncontrolled approximation for most systems.

The objective of the present study is to test the validity of HA for the cyclization probability or J factor of a simple homogeneous wormlike chain without torsional elastic energy. To this end, we compare values of the J factor computed by the method of normal-mode analysis (NMA), which is rigorously equivalent to HA computations,²⁴ with the corresponding exact result for the J factor computed by a new method that combines NMA with thermodynamic integration, a technique we denote TI-NMA.³⁰ This allows us to characterize the validity of HA (and NMA) in terms of a universal, model-independent function which depends only on the ratio L/P, where L is the contour length and P is the persistence length of the semiflexible chain. Although we consider here the simple case of a homogeneous wormlike

Page 4

chain without torsional elastic energy, we argue that our result for the deviation of HA (and NMA) from the exact behavior as a function of L/P qualitatively holds for any semiflexible macromolecular system that can be characterized by a contour length L and a persistence length P, including helical wormlike chains, looped DNA, and DNA having intrinsic bends or other local defects.

The rationale of this hypothesis is as follows. As discussed above, the validity of HA (and NMA) is based on the assumption that the system undergoes harmonic fluctuations about a well-defined elastic-energy minimum. For increasing system size, *e.g.*, increasing contour length *L* of a semiflexible biopolymer, it is the *large-scale conformational fluctuations* of the system, *i.e.*, those occurring on length scales large compared with the persistence length *P*, that generate the anharmonic contributions to the accessible conformation space and result in deviations of HA (and NMA) from the exact behavior. The precise form of the conformational fluctuations, and thus the free energy, of a complex macromolecular system depends of course on details such as monomer composition and solution conditions; however, the length scale on which these conformational fluctuations result in anharmonic behavior is set, by definition, by the persistence length *P*. This implies that the range of validity of HA and NMA is controlled by a single parameter, namely the ratio of system size to persistence length, or *L/P*. Due to the universal nature of our results for the deviation of HA (and NMA) from the exact behavior, as a function of *L/P*, we expect our results to remain qualitatively valid for more complex macromolecular systems.

II. THEORY AND RESULTS

1. The cyclization probability, or J factor

Cyclization probabilities are expressed in terms of a thermodynamic quantity *J*, also called the J factor. *J* is defined as a ratio of equilibrium constants for intra- and intermolecular synapsis reactions of chains with *N* vertices (or monomers) (Fig. 1),

$$J(N) = \frac{K_c(N)}{K_b}.$$

(1)

In this expression,

$$K_c(N) = \frac{Q_{cir}(N)}{Q_{lin}(N)}$$

is the equilibrium constant for an intramolecular cyclization reaction in which the two vertices at the ends of a linear chain with *N* vertices associate to form a circular chain with *N* vertices. $Q_{cii}(N)$ and $Q_{lin}(N)$ are conformational partition functions of a circular chain (cir) and a linear chain (lin) with *N* vertices, respectively. Partition functions are unitless by definition, which implies that the equilibrium constant $K_c(N)$ in Eq. (2) is unitless. In Eq. (1), the quantity

$$K_b = \frac{VQ_{lin}(2N)}{Q_{lin}(N) \cdot Q_{lin}(N)}$$

(3)

is the equilibrium constant for an intermolecular reaction in which two vertices at the ends of two different linear chains with N vertices associate to form a linear chain with 2N vertices. The equilibrium constant K_b is independent of N because the dependence of Q_{lin} on N cancels in Eq. (3), reflecting the fact that for linear chains the conformational degrees of freedom of vertices are independent from each other. Only the enthalpy of the additional bond in the 2N- chain enters K_b . Because the reaction corresponding to Eq. (3) is bimolecular, the quotient of partition functions in Eq. (3) is proportional to c=1/V where c is the concentration of linear chains and V is the available volume per chain.³¹ The dependence on the concentration c=1/V is eliminated by the factor V in the numerator of Eq. (3). This implies that K_b has units of volume, the J factor in Eq. (1) has units of concentration, and both K_b and J are independent of c=1/V. The J factor may be defined as the concentration of one end vertex in the vicinity of the other end vertex of the same linear chain L in the cyclization reaction shown in Fig. 1.^{32,33}

The enthalpy of interaction between chain termini contributes similarly to $K_c(N)$ and K_b , which implies that the J factor is independent of the interaction enthalpy and depends only on elastic properties of the chain (the cancelation of the interaction enthalpy between chain termini is based on the well-supported assumption that the equilibrium constants for cyclization and bimolecular ligation depend on the details of the joining reaction in the same way). Therefore, testing the validity of NMA using the J factor yields a universal, model-independent result for the validity of NMA which depends only on the ratio L/P, where L is the contour length and P is the persistence length of the semiflexible chain (cf. Section 4). Conversely, the free energy of cyclization

$$\Delta F = F_{cir} - F_{lin} = -k_B T \ln \left(\frac{Q_{cir}}{Q_{lin}}\right)$$

(4)

associated with the reaction $L \rightarrow C$ shown in Fig. 1 includes the association enthalpy and therefore depends on details of the interaction between chain termini.

In what follows we calculate the equilibrium constants $K_c(N)$ and K_b in Eq. (1) for a semiflexible harmonic model chain. In Section 2 we derive an explicit expression for the partition function Q_{lin} for a linear chain. We also obtain an explicit expression for the harmonic approximation $Q_{lin}^{(NMA)}$ of Q_{lin} , which is equivalent to calculating Q_{lin} using normal-mode analysis (NMA). The equilibrium constants K_b and $K_b^{(NMA)}$ are then calculated using Eq. (3). In Section 3 we discuss the numerical computation of the partition function Q_{cir} for a circular chain using the TI-NMA method³⁰ and obtain the corresponding NMA result $Q_{cir}^{(NMA)}$. In Section 4 we obtain the J factor, J, as a universal function of L/P and compare the exact result for J with the corresponding NMA result, J_{NMA} . This allows us to assess the validity of NMA in terms of a universal function of L/P which is free of microscopic details of our model chain. We argue that our results for the validity of NMA obtained here for a simple homogeneous wormlike chain characterize the validity of NMA for any semiflexible macromolecular system which can be characterized by a contour length L and a persistence length P.

2. Partition function for a linear chain

We consider a semi-flexible harmonic chain as a coarse-grained mesoscopic model for duplex DNA. Chain elements are extensible segments with equilibrium length b_0 connected end-to-end by semi-flexible joints, or vertices, at positions \mathbf{r}_i , i=1, ..., N (Fig. 2). The conformational partition function of a linear chain with N vertices (including the 2 vertices at the chain ends) and N-1 segments suspended in a volume V is given by

$$Q_{lin}(N) = \int_{V} \frac{d^3 r_1}{a^3} \dots \int_{V} \frac{d^3 r_N}{a^3} \exp\left[-\beta U_{lin}(\vec{r})\right]$$

(5)

where $U_{lin}(\vec{r})$ is the total potential energy for a chain conformation $\vec{r} = (\mathbf{r}_1, ..., \mathbf{r}_N)$ and $\boldsymbol{\beta} = (k_B T)^{-1}$ (*T* is the temperature in Kelvin and k_B is the Boltzmann constant). The constant *a* in Eq. (5) is a microscopic length required to make the partition function unitless. For a system of massive point particles undergoing Newtonian dynamics, the length *a* corresponds to the thermal wavelength;³⁴ however, in this work we are concerned only with conformational degrees of freedom, and consider *a* as a non-universal microscopic length much shorter than any other length scale associated with the chain. Essentially, the length *a* corresponds to the lattice constant of an underlying lattice needed to obtain a finite number of accessible conformations. The results for the J factor obtained in this work are independent of the length *a*, and thus largely independent of the discretization of our model chain.

Segments are described by displacement vectors $\mathbf{b}_i = \mathbf{r}_{i+1} - \mathbf{r}_i$ with length b_i and unit-length direction vectors $\hat{\mathbf{b}}_i = \mathbf{b}_i / b_i$. The bending angle θ_i at vertex \mathbf{r}_i between segments \mathbf{b}_{i-1} and \mathbf{b}_i is given by $\cos(\theta_i) = \hat{\mathbf{b}}_{i-1} \cdot \hat{\mathbf{b}}_i$ (Fig. 2). The total potential energy of the chain is given by

$$U_{lin}(\vec{r}) = k_B T c_b \sum_{i=2}^{N-1} \left[1 - \cos(\theta_i) \right] + k_B T \frac{c_s}{2} \sum_{i=1}^{N-1} \left(\frac{b_i}{b_0} - 1 \right)^2,$$

(6)

where c_b , c_s are bending and stretching elastic constants, respectively. In this work we neglect excluded volume interactions between chain segments, so that $U_{lin}(\vec{r})$ only includes the elastic potential energy of the chain. For a linear chain there is no elastic energy of bending associated with the end vertices \mathbf{r}_1 and \mathbf{r}_N which implies that the first sum in Eq. (6) only includes the N-2 inner vertices i = 2, ..., N-1 (Fig. 2).

The bending energy constant c_b in Eq. (6) is chosen such that the chain has a given persistence length *P*. Thus, c_b is implicitly determined by the equation

$$\langle \cos (\theta) \rangle = \exp \left(-\frac{b_0}{P}\right) = \exp \left(-\frac{1}{n}\right)$$

1	7	1
C	1)

where $n = P/b_0$ is the number of segments with equilibrium length b_0 per persistence length *P*. The thermal average $\langle \cos(\theta) \rangle$ is given by

$$\langle \cos (\theta) \rangle = \frac{\int_0^{\pi} d\theta \sin (\theta) \cos (\theta) \exp \left[-c_b (1 - \cos (\theta)) \right]}{\int_0^{\pi} d\theta \sin (\theta) \exp \left[-c_b (1 - \cos (\theta)) \right]}$$

(8)

In this work we consider chains for which $n = \{150, 50, 25\}$. These *n* values correspond to equilibrium segment lengths $b_0 = \{1, 3, 6\}$ ℓ where ℓ is the DNA axial rise corresponding to a single base pair (using P = 50 nm for DNA under physiological conditions we obtain $\ell = P$ 150 = 0.3333nm). Numerically solving Eq. (7) for c_b we find $c_b = \{150.5006, 50.5017, 25.5033\}$. The stretching energy constant c_s in Eq. (6) is defined as $c_s = K_s b_0/(k_B T)$ where K_s is the stretching modulus. Using $b_0 = \{0.3333, 1, 2\}$ nm, T = 300 K, and the approximate

value $K_s = 1000$ pN for DNA under physiological conditions, we find $c_s = \{80.4775, 241.4324, 482.8648\}.$

The reason for the above choice of the three *n* values is that numerical simulations used to obtain exact results for the free energy of a circular chain (Section 3) are computationally most efficient for chains having a number of vertices N between 50 and 300. For fixed *n* this corresponds to a range of values L/P = N/n spanning only a factor of 6 (e.g., 0.3333 L/P2 for n = 150). Given such a restricted range of N, choosing different values for n allows us to vary L/P over a much wider range, namely between 0.3333 (N = 50, n = 150) and 12 (N= 300, n = 25), corresponding to a 36-fold range of L/P. In addition, and equally important, using different *n* values allows us to compute the J factor for the same value of *L/P* using chains with different discretization (i.e., different segment lengths b_0 and associated elastic constants c_b , c_s). Because for a homogeneous chain we expect the J factor to depend only on the ratio L/P on general grounds, we expect that the results for the J factor computed for chains with different *n* collapse onto a single curve as a function of L/P. Using different *n* values therefore provides not only an opportunity to test the expected universal behavior of the J factor for a homogeneous chain (namely its dependence on L/P only, independent of details of the model chain), but also constitutes an important test of the validity and accuracy of our computational method.

The partition function $Q_{lin}(N)$ in Eq. (5) may be calculated explicitly by using spherical polar coordinates and carrying out the N-2 integrations over polar angles θ_i and N-1 radial integrations of segment lengths b_i iteratively. We find

$$Q_{lin}(N) = \frac{V}{a^3} \left(\frac{b_0^3}{a^3}\right)^{N-1} 2(2\pi)^{3(N-1)/2} \left(\frac{1-e^{-2c_b}}{c_b}\right)^{N-2} \left(\frac{1+c_s}{c_s^{3/2}}\right)^{N-1}$$



To obtain Eq. (9) we used the approximation $\int_0^\infty db_i \simeq \int_{-\infty}^\infty db_i$, i.e., extending the lower integration limits for integrations of b_i from 0 to $-\infty$; the error of this approximation is of order $\exp(-c_s/2)$ and is completely negligible for the values of c_s used in this work. The harmonic approximation to $Q_{lin}(N)$, equivalent to calculating $Q_{lin}(N)$ by normal model analysis (NMA), may formally be obtained as the leading contribution of $Q_{lin}(N)$ in Eq. (9) in a Taylor expansion of $1/c_b$ and $1/c_s$ about $1/c_b = 0$ and $1/c_s = 0$, which is given by

$$Q_{lin}^{(NMA)}(N) = \frac{V}{a^3} \left(\frac{b_0^3}{a^3}\right)^{N-1} 2(2\pi)^{3(N-1)/2} c_b^{-(N-2)} c_s^{-(N-1)/2}.$$

(10)

(This is formally equivalent to keeping for the exponent $\beta U_{lin}(\vec{r})$ of the Boltzmann factor in Eq. (5) only the leading, quadratic terms $\sim c_b \theta_i^2$ and $\sim c_s (b_i - b_0)^2$, and evaluating the resulting Gaussian integrals using $\int_0^\infty db_i b_i^2 \simeq \int_{-\infty}^\infty db_i b_i^2 \simeq b_0^2 \int_{-\infty}^\infty db_i$ in leading order.) The equilibrium constant K_b is found by inserting Q_{lin} from Eq. (9) in Eq. (3), resulting in

$$K_b = b_0^3 2^{1/2} \pi^{3/2} \left(\frac{1 - e^{-2c_b}}{c_b} \right)^2 \frac{1 + c_s}{c_s^{3/2}}.$$

(11)

Similarly, the NMA result $K_b^{(NMA)}$ is found by inserting $Q_{lin}^{(NMA)}$ from Eq. (10) in Eq. (3), resulting in

$$K_b^{(NMA)} = b_0^3 2^{1/2} \pi^{3/2} c_b^{-2} c_s^{-1/2}$$
(12)

As noted above, K_b has units of volume, but is independent of the concentration c = 1/V of linear chains; in the present case, K_b has units of volume because of the term b_0^3 in Eqs. (11), (12), where b_0 is the equilibrium length of a segment in our model chain.

3. Partition function and free energy for a circular chain

3.1. TI-NMA method—The partition function $Q_{cit}(N)$ for a circular chain with N vertices and N segments, corresponding to the circular state C shown in Fig. 1, is given by the expression in Eq. (5) replacing $U_{lin}(\vec{r})$ with

$$U_{el}(\vec{r}) = k_B T c_b \sum_{i=1}^{N} \left[1 - \cos(\theta_i) \right] + k_B T \frac{c_s}{2} \sum_{i=1}^{N} \left(\frac{b_i}{b_0} - 1 \right)^2$$

(13)

In this work, we neglect excluded volume interactions between chain segments and consider a phantom chain without topological constraint, i.e., we consider ensembles of circular chains which include all knot types. Furthermore, we consider the elastic energy due to bending and stretching of the chain only, as appropriate for nicked DNA. The only contribution to the potential energy of the chain is thus given by the elastic potential energy in Eq. (13) where the elastic constants c_b , c_s are the same as in Section 2. We compute the partition function Q_{cir} and the free energy $F_{cir} = -k_BT \ln(Q_{cir})$ using the TI-NMA method presented in reference ³⁰ and summarized for the present case in Figure 3. In this method, a circular molecular state, C, is gradually transformed into a harmonically constrained reference state C_0 which corresponds to the minimum energy conformation. The associated change in free energy, $F_0^{(TD)}$, is computed by thermodynamic integration (TI). The absolute free energy, $F_0^{(NMA)}$, of the reference state C_0 is computed separately by using NMA. The TI-NMA method yields the absolute free energy of the circular state C as (Fig. 3)

$$F_{cir} = F_0^{(NMA)} = \Delta F^{(TI)}.$$

(14)

3.2. Normal-mode Analysis—Applying normal-mode analysis (NMA) to the partition function Q_{cir} yields an approximation, $Q_{cir}^{(NMA)}$, which is expressed in terms of the eigenvalues of the Hessian matrix associated with the potential function $U_{el}(\vec{r})$ in Eq. (13). The Hessian matrix is calculated at the minimum energy conformation \vec{r}_0 of the circular chain, which consists of a regular polygon with *N* sides of length b_0 (Fig. 1). Following the procedure outlined in ³⁰ we obtain

$$Q_{cir}^{(NMA)}(N) = \exp\left(-\beta E_0\right) \frac{V}{a^3} \left(\frac{b_0^3}{a^3}\right)^{N-1} N^{3/2} 8\pi^2 \sqrt{I_x I_y I_z} \prod_{m=7}^{3N} \left(\frac{2\pi}{\nu_m}\right)^{1/2}$$

where $E_0 = k_B T c_b N[1 - \cos(2\pi/N)]$ is the energy of the minimum conformation \vec{r}_0 and I_x , I_y , I_z are the principal moments of inertia of \vec{r}_0 in units of b_0 (we here assume that each vertex of the chain is associated with a unit mass). The unitless quantities v_m in Eq. (15) are the eigenvalues of the Hessian matrix in units of $k_B T$ and b_0 . Assuming that the eigenvalues are ordered such that $v_1 \quad v_2 \quad \dots \quad v_{3N}$ one finds $v_m = 0$ for $m = 1, \dots, 6$ and $v_m > 0$ for $m = 7, \dots, 3N$. The 3N-6 nonzero eigenvalues are associated with internal vibrations of the chain about the minimum conformation \vec{r}_0 which incur a finite energetic cost. Conversely, the 6 zero eigenvalues $v_m = 0$ for $m = 1, \dots, 6$ are associated with rigid translations and rotations of the chain which do not incur any energetic cost. The corresponding eigenmodes contribute to $Q_{cir}^{(NMA)}(N)$ in Eq. (15) in terms of the number N of "particles" (vertices) and on the shape of the energy-minimized conformation, \vec{r}_0 , in terms of the principal moments I_x , I_y , I_y , I_z .

3.3. Thermodynamic integration—The objective of thermodynamic integration (TI) is to gradually transform the original circular state C into a harmonically constrained reference state C₀ for which the corresponding free energy, $F_0^{(NMA)}$, may be computed accurately by applying NMA to C₀. To this end, we gradually replace the energy function U_{el} of the original, semi-flexible circular chain C in Eq. (13) with a potential function corresponding to C₀ and to calculate the associated change in free energy, $F^{(TI)}(Fig. 3)$. A switching parameter λ is used to effect this change in chain properties according to the following scheme

$$U(\lambda) = \begin{cases} \lambda U_{ha} + (1 - \lambda) U_{el}, & 0 \le \lambda \le 1 \\ \lambda U_{ha}, & 1 \le \lambda \le \lambda_{max} \end{cases}.$$

(16)

The auxiliary elastic potential energy U_{ha} serves to constrain the system to a predefined reference conformation, which is here given by the minimum energy conformation \vec{r}_0 .³⁰ The auxiliary energy U_{ha} is defined by implementing phased intrinsic bends at the vertices of the chain in such a way that for the given reference conformation \vec{r}_0 every chain segment points in its preferred direction relative to the preceding segment.

TI is carried out in two phases, i.e., $\Delta F^{(TI)} = \Delta F_1^{(TI)} + \Delta F_2^{(TI)}$, with

(17)

$$\Delta F_2^{(TI)} = \int_{1}^{\lambda_{max}} d\lambda \Big\langle \frac{dU}{d\lambda} \Big\rangle_{\lambda} = \int_{1}^{\lambda_{max}} d\lambda \big\langle U_{ha} \big\rangle_{\lambda},$$

(18)

Here $U = U(\lambda)$ is given by Eq. (16) and the symbol $\langle \rangle_{\lambda}$ indicates an ensemble average taken at a specific value of λ . Values for $\langle U_{ha}-U_{el}\rangle_{\lambda}$ in Eq. (17) and $\langle U_{ha}\rangle_{\lambda}$ in Eq. (18) were obtained by Monte Carlo simulation for 11 equally spaced values $\lambda = \{0, 0.1, 0.2, ..., 1.0\}$ and for exponentially increasing λ values from 1 to a maximum value λ_{max} (see ³⁰ for details of the Monte Carlo simulation procedure). Starting with $\lambda = 1.0$, the values of λ were increased according to $\lambda_{i+1} = 1.05\lambda_i$ until λ was large enough to satisfy the criterion $\langle U \rangle_{\lambda} =$ $0.5 \cdot (3N-6)k_BT$ which holds in the harmonic regime due to the equipartition theorem (Fig. 4). The results were linearly interpolated and integrated according to Eqs. (17), (18). Each simulation was started at the minimum energy conformation \vec{r}_0 and an initial 1×10^6 trial moves were made to equilibrate the system. After initial equilibration, a new conformation was saved after each 1000 trial moves to produce a final ensemble. Fig. 4 shows $\langle U_{ha}-U_{el}\rangle_{\lambda}$ for $0 \quad \lambda \quad 1$ and $\langle \lambda U_{ha} \rangle_{\lambda} = \langle U \rangle_{\lambda}$ for $1 \quad \lambda \quad \lambda_{max}$ for all values of λ quoted above. Each data point represents the average obtained from an ensemble of 10^5 conformations.

4. Comparison of J-factor values obtained by NMA and TI-NMA

Using Eq. (1) one may express the J factor as

$$-\ln\left(\frac{J(N)}{c_0}\right) = \frac{\Delta F(N)}{k_B T} + \ln\left(c_0 K_b\right)$$

(19)

where the free energy of cyclization $F = F_{cir} - F_{lin}$ is calculated using Eqs. (4), (9), (14). In Eq. (14), the absolute free energy $F_0^{(NMA)}$ of the reference state C_0 is calculated using

$$F_0^{(NMA)} = -k_B T \ln \left(Q_{cir}^{(NMA)}\right) \text{ with } Q_{cir}^{(NMA)} \text{ from Eq. (15) (cf. Fig. 3). Note that the common prefactor } \frac{V}{a^3} \left(\frac{b_0^3}{a^3}\right)^{N-1} \text{ in Eqs. (9) and (15) cancels in the difference } F = F_{cir} - F_{lin}, \text{ which}$$

implies that the J factor is independent of the volume V and of the microscopic length a introduced in Eq. (5). The equilibrium constant K_b in Eq. (19) is given by Eq. (11). The reference concentration c_0 in Eq. (19) is required to make the arguments of the logarithms in Eq. (19) unitless. Note that c_0 merely defines the units scale in terms of which J and K_b are expressed; the quantities J and K_b themselves are independent of the concentrations c_0 and c = 1/V. In this work we express J in units of the standard molar concentration $c_0 = 1$ M. Eq. (19) shows that the negative logarithm of the length-dependent J-factor J(N) is a measure of the length-dependent cyclization free energy F(N) modulo an additive constant. Both terms on the right hand side of Eq. (19) are non-universal, i.e., depend on details of the model, but J on the left hand side of Eq. (19) is universal.

Similarly, the NMA result for the J factor is given by

$$-\ln\left(\frac{J_{NMA}}{c_0}\right) = \frac{\Delta F_{NMA}}{k_B T} + \ln\left(c_0 K_b^{(NMA)}\right),$$

(20)

where the NMA result for the free energy of cyclization, F_{NMA} , is obtained using $\Delta F_{NMA} = -k_B T \ln \left(Q_{cir}^{(NMA)} / Q_{lin}^{(NMA)} \right)$ with $Q_{lin}^{(NMA)}$ in Eq. (10). The NMA result $Q_{cir}^{(NMA)}$ is obtained by applying NMA directly to the circular state C (see Eq. (15) and Fig. 3). The equilibrium constant $K_h^{(NMA)}$ in Eq. (20) is given by Eq. (12).

J and J_{NMA} in Eqs. (19), (20) were calculated for chains with $N = \{50, 75, 100, 125, 150, 175, 200, 225, 250, 275, 300\}$ vertices and segments of equilibrium length $b_0 = \{1, 3, 6\}$ where l is the DNA axial rise corresponding to a single base pair (Fig. 5). The J factor, J, is universal in the sense that J depends only on the ratio L/P, but not on microscopic details of the model, such as the segment length b_0 . That is, when calculated using different values of b_0 , the resulting J factor collapses to a single function of L/P. This collapse confirms the expected universal behavior of the J factor for a homogeneous chain and constitutes an acid test of the validity and accuracy of our computational method (Fig. 5).

Fig. 6 shows the deviation $\ln (J[M]) - \ln (J_{NMA}[M]) \ln (J/J_{NMA})$ as a universal function of L/P. Note the deviation $\ln (J[M]) - \ln (J_{NMA}[M])$ essentially corresponds to the difference in cyclization free energies F_{NMA} - F in units of k_BT , modulo an additive constant. The length dependence of this positive deviation is essentially linear in the ratio L/P and exceeds 1 k_BT only for $L/P \gtrsim 4$, approximately 600 bp (assuming that P = 50 nm). Thus, using the harmonic approximation or NMA for systems with smaller chain lengths yields a slightly

underestimated J factor. Given that these results are a universal function of L/P, they should hold both for DNA cyclization and the more general problem of DNA looping.

III. CONCLUSIONS

There is increasing interest in the phenomenon of DNA and chromatin looping as a common mechanism of biological regulation.^{35–44} Together with recognition that cyclization J-factor measurements are exquisitely sensitive to helical parameters and conformational properties of DNA molecules, there has been strong motivation to develop advanced statistical-mechanical models of DNA-loop formation (with cyclization as a special case). Although Monte Carlo methods for computing *J* have been the standard for problems involving sequence-dependent bending and flexibility, they remain challenged by finite computing resources. HA and NMA-based J-factor calculations are up to four orders of magnitude more efficient and therefore more suitable for analyzing experimental looping and cyclization data. As attractive as these approaches are, they have been approximations of unknown extent to the actual physical behavior of DNA rings and loops.

We have sought here to estimate the error in the J factor computed by HA/NMA techniques, which arise from anharmonic contributions to the behavior of wormlike chains and increase with chain size. For L/P 4, HA/NMA systematically underestimate the configurational free-energy cost of cyclization by an amount less than or equal to the thermal energy k_BT , as determined by rigorously computing the exact free energy using a thermodynamic integration technique. Whether this error is tolerable without a TI correction or not may depend on the accuracy needed for a given analysis. For J-factor measurements over a narrow range of DNA sizes, it may be sufficient to assume that a small, multiplicative factor greater than unity can be applied to J_{NMA} in order to correct for the deviation. We suggest, however, that such an approach should be used with caution in the case of larger chains.

Although we consider here the simple test case of a homogeneous wormlike chain without torsional elastic energy, we argue that our result for the deviation of the HA (and NMA) from the exact behavior as a function of L/P (Fig. 6) qualitatively holds for any semiflexible macromolecular system which can be characterized by a contour length L and a persistence length P, including helical wormlike chains, looped DNA, and DNA having intrinsic bends or other locally flexible defects (cf. the end of Section I). It would be interesting to test this hypothesis by systematically testing the validity of HA and NMA for more complex systems.

A hallmark of Don Crothers' approach to science was his fearlessness in adapting, improving, or devising whatever mathematical or computational tools were needed to more effectively analyze experimental results. We offer the present theoretical treatment in the spirit of Don's legacy.

IV. METHODS

Monte Carlo simulations were carried out using Fortran 90 and the same algorithm as in³⁰ but without monitoring excluded volume or knot checking. Averages $\langle U \rangle_{\lambda}$ and standard deviations σ were obtained from ensembles of 10⁵ conformations each. In order to estimate

the number of trial moves between conformations required to generate an ensemble of independent conformations, we calculated the autocorrelation function $\operatorname{acf}(\tau) = \langle (U(t) - \langle U \rangle) (U(t+\tau) - \langle U \rangle) \rangle$ and fit $\operatorname{acf}(\tau)$ to an exponential decay function $\exp(-\tau/k)$ to estimate the value of the wait time *k*. Waiting times were measured in units of 1000 trial moves, *i.e.*, k = 1 corresponds to a waiting time of 1000 trial moves. If k > 1/3 was found, the number η of independent conformations in the ensemble generated by the Monte Carlo simulation was estimated as $\eta = 10^{5}/(3k)$; if k = 3 was found, we used $\eta = 10^{5}$, i.e., considered the entire ensemble of 10^{5} conformations as independent. Standard errors of the mean (sem) for each simulation was estimated as sem $= \sigma/\sqrt{\eta}$. Error in the TI procedure was estimated according to standard error-propagation analysis.

NMA calculations were performed using Python and Fortran 90. CPU time for this calculation scales as $O(N^2)$; for a chain of N = 300 an NMA calculation takes under a minute on a single CPU. TI-NMA calculations were performed on a 32-CPU computing cluster. With all processors occupied a calculation for N = 50 takes about one hour and scales linearly with large N(i.e., N = 300 takes six hours). This linear scaling applies only to calculations omitting excluded-volume- and knot-type-checking steps. Source code is available upon request.

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References

- Shore D, Langowski J, Baldwin RL. Proc Natl Acad Sci USA. 1981; 78:4833–4837. [PubMed: 6272277]
- 2. Shore D, Baldwin RL. J Mol Biol. 1983; 170:957-981. [PubMed: 6315955]
- 3. Shore D, Baldwin RL. J Mol Biol. 1983; 170:983–1007. [PubMed: 6644817]
- Olson WK, Marky NL, Jernigan RL, Zhurkin VB. J Mol Biol. 1993; 232:530–554. [PubMed: 8345522]
- 5. Olson WK, Zhurkin VB. Curr Opinion Struct Biol. 2000; 10:286–297.
- 6. Shimada J, Yamakawa H. Macromolecules. 1984; 17:689-698.
- 7. Shimada J, Yamakawa H. J Mol Biol. 1985; 184:319-329. [PubMed: 4032481]
- Marini JC, Levene SD, Crothers DM, Englund PT. Proc Natl Acad Sci USA. 1982; 79:7664–7668. [PubMed: 16593261]
- 9. Wu HM, Crothers DM. Nature. 1984; 308:509-513. [PubMed: 6323997]
- 10. Levene SD, Crothers DM. J Mol Biol. 1986; 189:61-72. [PubMed: 3783680]
- 11. Levene SD, Crothers DM. J Mol Biol. 1986; 189:73-83. [PubMed: 3783681]
- 12. Flory PJ, Suter UW, Mutter M. J Am Chem Soc. 1976; 98:5733-5739.
- 13. Alexandrowicz Z. J Chem Phys. 1969; 51:561-565.
- 14. Hagerman PJ. Biopolymers. 1985; 24:1881-1897. [PubMed: 4074845]
- 15. Hagerman PJ, Ramadevi VA. J Mol Biol. 1990; 212:351-362. [PubMed: 2319603]
- 16. Levene SD, Wu HM, Crothers DM. Biochemistry. 1986; 25:3988–3995. [PubMed: 3017412]
- Hockings SC, Kahn JD, Crothers DM. Proc Natl Acad Sci USA. 1998; 95:1410–1415. [PubMed: 9465028]
- 18. Kahn JD, Crothers DM. J Mol Biol. 1998; 276:287-309. [PubMed: 9514724]
- 19. Kahn JD, Yun E, Crothers DM. Nature. 1994; 368:163–166. [PubMed: 8139661]

- 20. Koo HS, Drak J, Rice JA, Crothers DM. Biochemistry. 1990; 29:4227-4234. [PubMed: 2361140]
- 21. Nathan D, Crothers DM. J Mol Biol. 2002; 316:7–17. [PubMed: 11829499]
- Roychoudhury M, Sitlani A, Lapham J, Crothers DM. Proc Natl Acad Sci USA. 2000; 97:13608– 13613. [PubMed: 11095739]
- 23. Zhang Y, Crothers DM. Proc Natl Acad Sci USA. 2003; 100:3161–3166. [PubMed: 12629220]
- 24. Zhang Y, Crothers DM. Biophys J. 2003; 84:136–153. [PubMed: 12524271]
- 25. Zhang Y, McEwen AE, Crothers DM, Levene SD. Biophys J. 2006; 90:1903–1912. [PubMed: 16361335]
- Swigon D, Coleman BD, Olson WK. Proc Natl Acad Sci USA. 2006; 103:9879–9884. [PubMed: 16785444]
- 27. Zhang Y, McEwen AE, Crothers DM, Levene SD. PLoS ONE. 2006; 1:e136. [PubMed: 17205140]
- 28. Flory, PJ. Principles of Polymer Chemistry. Cornell University Press; Ithaca, NY: 1953.
- 29. de Gennes, PG. Scaling Concepts in Polymer Physics. Cornell University Press; Ithaca, NY: 1979.
- Giovan SM, Scharein RG, Hanke A, Levene SD. J Chem Phys. 2014; 141:174902. [PubMed: 25381542]
- 31. Hill, TL. Introduction to Statistical Thermodynamics. Addison-Wesley; Reading, MA: 1960.
- Crothers DM, Drak J, Kahn JD, Levene SD. Methods Enzymol. 1992; 212:3–29. [PubMed: 1518450]
- Levene SD, Giovan SM, Hanke A, Shoura MJ. Biochem Soc Trans. 2013; 41:513–518. [PubMed: 23514145]
- 34. Reichl, LE. A Modern Course in Statistical Physics. John Wiley & Sons; New York, NY: 1998.
- Polikanov YS, Bondarenko VA, Tchernaenko V, Jiang YI, Lutter LC, Vologodskii A, Studitsky VM. Biophys J. 2007; 93:2726–2731. [PubMed: 17573434]
- 36. Hou C, Dale R, Dean A. Proc Natl Acad Sci USA. 2010; 107:3651-3656. [PubMed: 20133600]
- 37. Tokuda N, Sasai M, Chikenji G. Biophys J. 2011; 100:126–134. [PubMed: 21190664]
- Guo Y, Monahan K, Wu H, Gertz J, Varley KE, Li W, Myers RM, Maniatis T, Wu Q. Proc Natl Acad Sci USA. 2012; 109:21081–21086. [PubMed: 23204437]
- Kulaeva OI, Nizovtseva EV, Polikanov YS, Ulianov SV, Studitsky VM. Mol Cell Biol. 2012; 32:4892–4897. [PubMed: 23045397]
- 40. Le May N, Fradin D, Iltis I, Bougnères P, Egly JM. Mol Cell. 2012; 47:622–632. [PubMed: 22771116]
- 41. Sanyal A, Lajoie BR, Jain G, Dekker J. Nature. 2012; 489:109–113. [PubMed: 22955621]
- 42. Brackley CA, Taylor S, Papantonis A, Cook PR, Marenduzzo D. Proc Natl Acad Sci USA. 2013; 110:E3605–E3611. [PubMed: 24003126]
- 43. Chaumeil J, Micsinai M, Ntziachristos P, Deriano L, Wang JM, Ji Y, Nora EP, Rodesch MJ, Jeddeloh JA, Aifantis I, Kluger Y, Schatz DG, Skok JA. Cell Rep. 2013; 3:359–370. [PubMed: 23416051]
- 44. Hensel Z, Weng X, Lagda AC, Xiao J. PLoS Biol. 2013; 11:e1001591. [PubMed: 23853547]

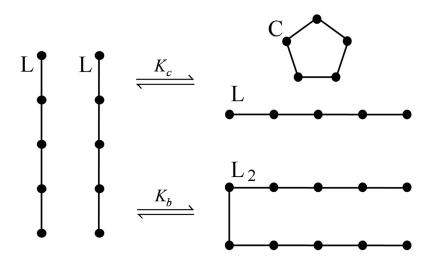


Fig. 1.

Intra- and intermolecular synapsis reactions. For the intramolecular reaction (equilibrium constant K_c) the end vertices of a linear chain, L, with N vertices and N-1 segments bind to form a circular chain, C, with N vertices and N segments (here N=5). The added segment in the circular chain corresponds to the new chemical bond formed in the cyclization reaction. For the intermolecular reaction (equilibrium constant K_b) the end vertices of two different linear chains L associate to form a linear chain with 2N vertices (L₂).

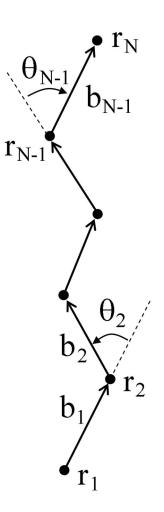


Fig. 2.

Linear semi-flexible harmonic chain with *N* vertices at positions $\vec{r} = (\mathbf{r}_1, ..., \mathbf{r}_N)$ and *N*-1 segments described by displacement vectors $\mathbf{b}_i = \mathbf{r}_{i+1} - \mathbf{r}_i$, i = 1, ..., N-1 (here *N*=6). Bending of the chain is described by *N*-2 polar angles θ_i located at inner vertices \mathbf{r}_i between segments \mathbf{b}_{i-1} and \mathbf{b}_i , i = 2, ..., N= 1.

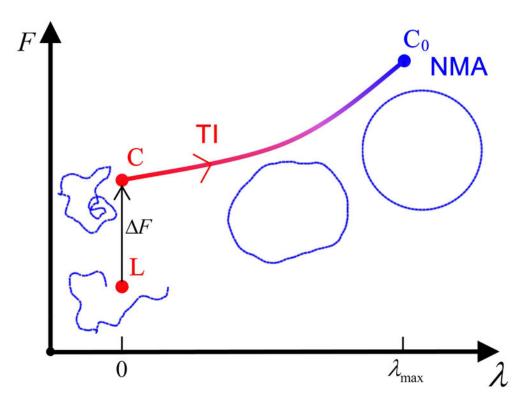


Fig. 3.

Calculation of the cyclization free energy $F = F_{cir} - F_{lin}$ between a circular molecular state C and a linear molecular state L (cf. Fig. 1). Thermodynamic integration (TI) yields the change in free energy, $F^{(TI)}$, as the circular state C is transformed into the reference state C₀ (red section of the curve). Normal mode analysis (NMA) yields the absolute free energy $F_0^{(NMA)}$ of C₀ (blue section). The absolute free energy of the circular state C is then obtained as $F_{cir} = F_0^{(NMA)} - \Delta F^{(TI)}$. Typical Monte Carlo conformations of the circular state C are shown for the TI portion. The absolute free energy F_{lin} of the linear state L is given by $F_{lin} = -k_B T \ln(Q_{lin})$ with Q_{lin} given in Eq. (9).

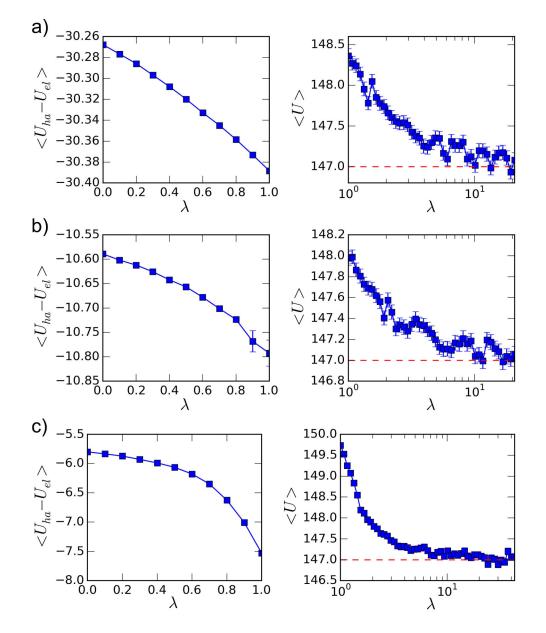


Fig. 4.

 $\langle U_{ha}-U_{el}\rangle_{\lambda}$ and $\langle \lambda U_{ha}\rangle_{\lambda} = \langle U \rangle_{\lambda}$ in units of k_BT for all λ values given in the text, for chains with N = 100 segments and equilibrium segment lengths a) $b_0 = l$ b) $b_0 = 3 l$ and c) $b_0 = 6 l$ where l is the axial rise per base pair in duplex DNA. In the harmonic regime obtained for large λ , $\langle U \rangle_{\lambda} / (k_BT)$ converges to $0.5 \cdot (3N-6) = 147$ for N = 100 due to the equipartition theorem (dashed lines).

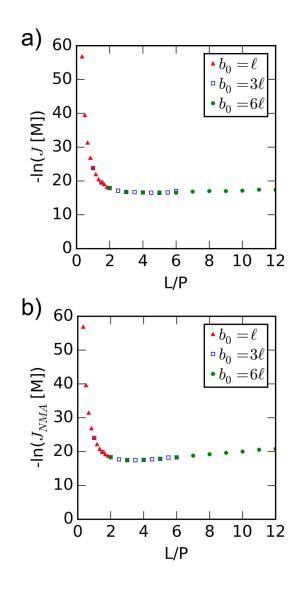


Fig. 5.

a) $-\ln(J[M])$ and b) $-\ln(J_{NMA}[M])$ as universal functions of L/P, where L is the contour length and P the persistence length of the chains in the cyclization reaction (cf. Fig. 1). Because the J factor, J, is a universal function of L/P, results obtained using chains with different values of the segment length b_0 (here $b_0 = \{1, 3, 6\}l$ where l is the rise per base pair in duplex DNA) collapse onto a single curve as a function of L/P.

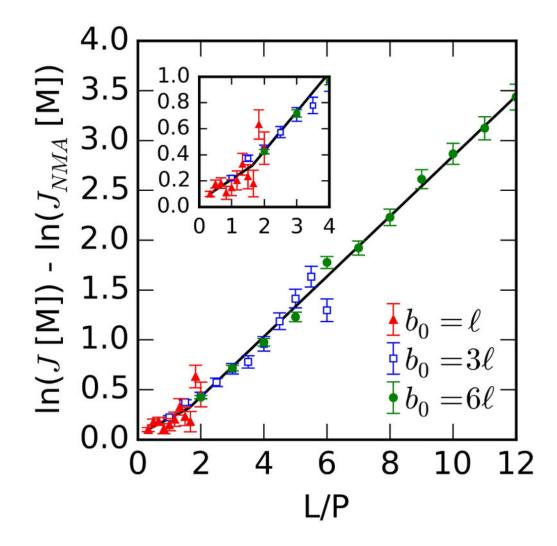


Fig. 6.

Deviation $\ln (J[M]) - \ln(J_{NMA}[M])$ as a universal function of L/P (cf. Fig. 5) to assess the validity of the harmonic approximation (HA) and normal mode analysis (NMA). The solid curve is a piecewise-linear fit given by $y = 0.166 \ x+0.0458 \ (x < 1.63)$ and $y = 0.302 \ x-0.176 \ (x \ 1.63)$, where x = L/P and $y = \ln(J[M])/J_{NMA}[M]$. The fit for $x \ 4$ is also shown on an expanded scale in the inset to the figure. Since the deviation is obtained as a universal function of L/P, we expect similar behavior for any semiflexible macromolecular system which can be characterized by a contour length L and persistence length P.