



The ecotoxicology of marine tributyltin (TBT) hotspots: A review

Jonny Beyer ^{a,*}, You Song ^a, Knut Erik Tollefsen ^a, John Arthur Berge ^{a,1}, Lise Tveiten ^a,
Aud Helland ^b, Sigurd Øxnevad ^a, Merete Schøyen ^a

^a Norwegian Institute for Water Research (NIVA), Økernveien 94, NO-0579, Oslo, Norway

^b COWI, Karvesvingen 2, NO-0579, Oslo, Norway



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ABSTRACT

Tributyltin (TBT) was widely used as a highly efficient biocide in antifouling paints for ship and boat hulls. Eventually, TBT containing paints became globally banned when TBT was found to cause widespread contamination and non-target adverse effects in sensitive species, with induced pseudohermaphroditism in female neogastropods (imposex) being the best-known example. In this review, we address the history and the status of knowledge regarding TBT pollution and marine TBT hotspots, with a special emphasis on the Norwegian coastline. The review also presents a brief update on knowledge of TBT toxicity in various marine species and humans, highlighting the current understanding of toxicity mechanisms relevant for causing endocrine disruption in marine species. Despite observations of reduced TBT sediment concentrations in many marine sediments over the recent decades, contaminant hotspots are still prevalent worldwide. Consequently, efforts to monitor TBT levels and assessment of potential effects in sentinel species being potentially susceptible to TBT in these locations are still highly warranted.

1. Introduction

The strong antifouling (AF) effect of trivalent organotin (OT) compounds was discovered in the 1950s by a Dutch research group led by [van der Kerk and Luitjen \(1954\)](#) and this led tributyltin (TBT) to become the main biocidal ingredient for marine AF paint products, primarily in the form of bis(tributyltin)oxide (TBTO) ([IPCS, 1990](#)). In the 1960s, early versions of TBT AF paints for ship hulls were of the “free association” type which allowed a free and rapid release of AF agents over a relatively short time. In the “self-polishing copolymer” paints, which were introduced in the 1970s, TBT was chemically bounded to a polymer backbone and this enabled a slow, controlled release of TBT which maintained the AF effect longer, typically several years ([Omae, 2003; Schultz, 2007; Santillo et al., 2008; Amara et al., 2018](#)). In 1980, the annual global use of TBT agents in marine AF paints was about $2\text{-}3 \times 10^3$ tonnes ([IPCS, 1990](#)). TBT was also used in other AF applications which contributed to the total release of TBT in marine ecosystems. For example, in Norway about 14×10^4 kg of OTs were used in 1986 for AF treatment of nets and sea pens at approximately 600 fish farms ([Linden, 1987](#)). In addition, TBT was heavily used as a biocide in a range of other applications such as cooling systems, wood pulping, leather processing,

wood preservation processes, and textile treatments ([IPCS, 1990](#)). The total use of TBT AF applications resulted in significant TBT pollution in coastal areas all over the world ([Hoch, 2001; Sousa et al., 2014](#)) and at the most contaminated sites, the TBT hotspots, the contamination has persisted even until today (see paragraph 4).

TBT has by some ecotoxicologists been ranked as the most hazardous anthropogenic chemical ever to be deliberately released in large quantities to the environment. Thousands of studies about the environmental fate and effects of TBT have been reported during more than 50 years of research, and this field still attracts considerable attention, much due to the actions of TBT as an endocrine disrupting compound with the capacity to influence biological fitness (e.g., growth, development and reproduction) in sensitive species groups. In this regard, focus will be on identifying molecular mechanisms and the Modes of Actions (MoA) which TBT have on sensitive species, and how these mechanisms may be relevant in the toxicity progression also for other marine species.

In this review, we provide a short historical and ecotoxicological summary of the antifoulant biocide TBT, with emphasis on its role as an endocrine disrupting compound. We examine the coastal sites in Norway with the highest known levels of TBT pollution in marine sediments, and we compare these to similar locations found in other countries. We

* Corresponding author.

E-mail address: jonny.beyer@niva.no (J. Beyer).

¹ Retired.

then assess the knowledge status on TBT ecotoxicity in marine systems and in various bioindicator organisms, but with an emphasis to the imposex effect in marine neogastropod snails, as this is the best-known ecotoxicological effect of TBT pollution and the main reason why the use of TBT AF paints became banned globally. In the final part, we examine the possible role that marine TBT hotspots may have for continued, mechanism-oriented ecotoxicological research on TBT, especially with relation to the endocrine disruption effect phenomena in different species, including humans.

2. Fate, effects and ban of TBT antifouling products

The environmental behaviors of TBT AF compounds in marine systems are complex. Being ionizable, their speciation is strongly affected by the pH of seawater, but under the mild basic conditions of normal seawater TBT is expected to occur mainly as uncharged hydroxyl complexes (TBT-OH) and behave relatively similar as hydrophobic organic contaminants (Brändli et al., 2009). In seawater, TBT-OH has an acid-dissociation constant (pK_a) of 6.3 to the TBT⁺ cation (Arnold et al., 1997) and cations of TBT, and other OTs, will be attracted and adsorbed to suspended particles with net negatively charged surfaces, such as organic matter and clay (Arnold et al., 1997; Weidenhaupt et al., 1997; Meador, 2000; Hoch, 2001). These fate processes make TBT accumulate and persist in marine sediments, especially if the sediment is fine-grained, high in organic content (high Total Organic Carbon, TOC) and with low/depleted levels of oxygen (i.e., hypoxic and anoxic sediment conditions). In well-oxygenated surficial sediments, TBT typically has a half-life of 1–5 years, but in fine-grained, O₂-depleted - anoxic sediments the half-life can extend to several decades (Dowson et al., 1993, 1996; De Mora et al., 1995; Berge et al., 2006; Langston et al., 2015). Studies show that the sediment/porewater partition coefficient

(K_d) for TBT correlate positively to the TOC content of the sediments (Langston and Pope, 1995). The TBT content is therefore often normalized to sediment TOC levels, typically 5% TOC (Abraham et al., 2017). As TBT bound to sediments leak slowly back into the seawater, TBT hotspot sediments represent a long-term secondary source for re-contamination of the water column in these areas, unless active sediment remediation measures, such as dredging and capping, are performed. TBT has been shown to bioaccumulate in all aquatic taxa, with molluscs showing highest bioconcentration and bioaccumulation factors that may result in tissue burdens ranging up to 7 µg g⁻¹ (wet weight) (Laughlin, 1996).

The first signs of TBT causing adverse impacts in non-target marine animals came in the early 1970s when scientists started to notice developmental disorders in oysters (*Ostrea edulis* and *Crassostrea gigas*) and in prosobranch neogastropod snails such as dogwhelk (*Nucella lapillus*), mud snails (*Nassarius obsoletus*), and oyster drill (*Ocenebra erinaceus*). The disorders were particularly prevalent at coastal locations near busy boating and shipping activities (Blaber, 1970; Smith, 1971, 1981; Waldock and Thain, 1983; Alzieu et al., 1986; Gibbs, 1993; Matthiessen and Gibbs, 1998; Alzieu, 2000; Sousa et al., 2014). In oysters, a shell thickening and decreased growth phenomenon was noted by oyster farmers in Arcachon Bay (France) as the first place. It gave considerable losses for the local shellfish industry as affected oyster batches were literally unsellable on the market (Waldock and Thain, 1983; Higuera-Ruiz and Elorza, 2011; Horiguchi, 2009). Later studies found that the oyster shell thickening effect occurs when TBT concentrations in seawater exceed 2 ng L⁻¹ (Langston, 2020). In the TBT affected neogastropods, abnormal females exhibited non-functional male-like sexual tissues/organs such as a vas deferens and even a penis-like organ grown superimposed over the female sexual tissues/organs (Fig. 1). This abnormal condition was named “imposex”

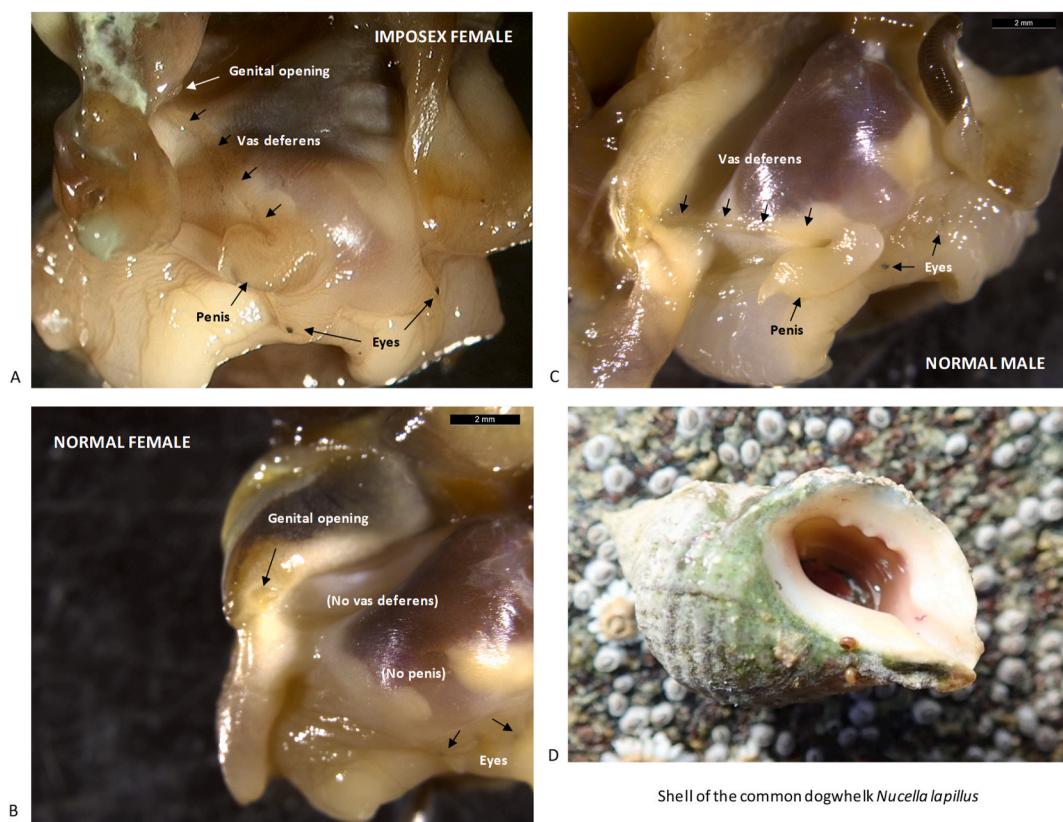


Fig. 1. Imposex in female of common dogwhelk *Nucella lapillus* (A) compared to the normal sexual characteristics of females (B) and males (C) of the same species. Panel A shows imposex at stage 4 according to Oehlmann et al. (1991), with a continuous vas deferens and a penis superimposed on the female sexual characters. Panel D shows a typical dogwhelk shell. Included scale bars are 2 mm. Photos: NIVA.

(Smith, 1971). Detailed studies revealed that the developing females acquired infertility in the advanced stages of imposex (Oehlmann et al., 1991), eventually causing the affected populations to decline and even become extinct (Blaber, 1970; Smith, 1981). Controlled exposure-effect studies in the laboratory found *N. lapillus* to be extremely sensitive with young females developing imposex traits when exposed to TBT concentrations as low as 1 ng.L⁻¹ in seawater (i.e., 0.001 parts per billion) and with affected females becoming infertile at TBT concentrations above 3–5 ng.L⁻¹ (Bryan et al., 1986; Gibbs et al., 1987, 1988; Evans et al., 1996; Matthiessen and Gibbs, 1998; Birchenough et al., 2002b). Field surveys in the 1980s and 90s demonstrated the imposex phenomenon to be surprisingly widespread, occurring even at locations relatively far from the busiest shipping or boating areas, and being found in several hundred neogastropod species worldwide (Vos et al., 2000; Shi et al., 2005; Titley-O'Neal et al., 2011b; Grilo and Rosa, 2017). The sum of studies on marine molluscs and TBT have demonstrated TBT contaminants may cause a range of different toxicities, including increased larval mortality (IPCS, 1990), inhibited egg development (Bryan and Gibbs, 1991), induced vitellogenin gene expression (Park et al., 2012), reduced survival of hatchlings (Leung et al., 2007), altered population sex ratios (Sousa et al., 2005a), induced DNA damage (Gabbianelli et al., 2006; Hagger et al., 2006; Martinovic et al., 2016), increased expression of heat-shock proteins (Clayton et al., 2000), and female “intersex” condition in certain mesogastropoda species such as the common periwinkle *Littorina littorea* (Bauer et al., 1995; Matthiessen and Gibbs, 1998; Oehlmann et al., 1998; Rank, 2009).

The discovery of TBT AF systems causing widespread contamination and adverse effects in nontarget molluscs caused much concern and led to initiatives to initiate restrictions of OT based AF paints in the late 1980s. The earliest ban was implemented in France in 1982 regarding all leisure boats and small commercial vessels shorter than 25 m (Alzieu et al., 1986; Abel, 1990; Santillo et al., 2008). In the subsequent years, OSPAR countries committed themselves to phase out OT based marine AF paints (OSPARCOM, 2009) and in 1999, the Marine Environmental Protection Committee of the International Maritime Organization finally supported the full phase-out strategy worldwide during the five-year implementation period between 2003 and 2008 (Champ, 2000). Documentation from many field surveys and monitoring programs suggest the ban against TBT AF paints has largely been effective, although even after 2008 some suppliers continued to produce and sell these products in several countries (Turner and Glegg, 2014), even at the present time (Paz-Villarraga et al., 2022; Uc-Peraza et al., 2022a).

3. Trends of marine TBT effect monitoring

Organotins in sediments and TBT specific effects became mandatory determinants of the OSPAR Co-ordinated Environmental Monitoring Programme (CEMP) from 2003 onwards (OSPAR Agreement, 2010-1) and guidance for these activities is provided in Technical annex 3 of the JAMP Guidelines for contaminant-specific biological effects monitoring (OSPAR, 2008). OSPAR CEMP (2009) developed a six-class quality scheme (class A-F) for assessing TBT-specific biological effects in five marine gastropods (*Nucella*, *Nassarius*, *Buccinum*, *Neptunea*, *Littorina* spp), including effect parameters (imposex/intersex) and TBT contamination data for seawater and sediments to enable an integrated (exposure-effect) assessment in these bioindicators.

Many marine TBT effect monitoring programs conducted later than the early 1990s have documented the existence of TBT pollution and neogastropod imposex effects in coastline seas in all continents, i.e., in Europe (Ten Hallers-Tjabbes et al., 1994; Følsvik et al., 1999; Svarsson, 2000; Svarsson et al., 2001; Chiavarini et al., 2003; Vasconcelos et al., 2010; Laranjeiro et al., 2018), Africa (Marshall and Rajkumar, 2003), Asia (Horiguchi et al., 1994, 1997; Tan, 1997; Bech, 2002), Oceania (Smith, 1996), North America (Gooding et al., 2003), and South America (Gooding et al., 1999). The imposex effect monitoring programs have most often targeted *N. lapillus* as the monitoring

organism due to its cosmopolitan distribution and its extreme sensitivity and vulnerability to TBT. As *N. lapillus* has internal fertilization, lay eggs in capsules fixed to the bottom, hatch as juvenile miniature versions of the adult (i.e., no free-swimming planktonic larvae), they are particularly vulnerable to reductions of fecundity as local populations have limited potential for recovery based on immigration. Other whelk species closely related to *N. lapillus*, such as netted dogwhelk in the genus *Nassarius*, which also have been extensively used for TBT/imposex studies, e.g. Stroben et al. (1992), Barroso et al. (2002); Cuevas et al. (2014); Rial et al. (2018); Cacciatore et al. (2018), Ruiz et al. (2018), are less vulnerable to local population extinctions as they have planktonic larvae.

The peak global usage of TBT in marine AF products occurred most likely sometime during the 1980s, but significant variation among countries is likely due to variable timing and strictness of TBT regulations. After effectuation of TBT bans, marine effect monitoring programs with dogwhelk as bioindicators have largely shown declining imposex trends as well as positive recoveries of local gastropod populations, e.g., (Smith, 1996; Birchenough et al., 2002a; Schøyen et al., 2019). However, in many coastal TBT hotspots, high pollution levels have been shown to persist, including in Europe (Sousa et al., 2005b, 2009; Smith et al., 2006; Gibbs, 2009; Giltrap et al., 2009; Galante-Oliveira et al., 2011; Furdek et al., 2012; Nicolaus and Barry, 2015; Langston et al., 2015; Anastasiou et al., 2016; Ruiz et al., 2018; Cacciatore et al., 2018; Laranjeiro et al., 2018; Filipkowska and Kowalewska, 2019), North America (Coray and Bard, 2007; Tallmon and Hoferkamp, 2009; Titley-O'Neal et al., 2011a), South America (Toste et al., 2011; Rossato et al., 2016; Batista et al., 2016; Mattos et al., 2017; da Costa et al., 2017; Castro et al., 2018), Asia (Choi et al., 2013; Ho and Leung, 2016; Lam et al., 2017; Kim et al., 2017); and Africa (El Ayari et al., 2018). Both historical and recent uses of organotin based AF have been identified as sources for continued high contamination in these hotspot areas. Hull maintenance of leisure boats that earlier have been treated with TBT paint may also represent an important source, e.g., as shown by Ytreberg et al. (2016). TBT contamination has also been linked to other sources. For example, Diez et al. (2002) found TBT at levels of 244 ng.g⁻¹ in a sewage discharge and pointed to this as an important secondary source of TBT to the marine environment. The study by Cornelissen et al. (2008) found significant levels of TBT (median 140 µg.kg⁻¹ d.w.) in urban runoff particles far away from any yard or maritime activity and they suggested long-lasting house paint and use of TBT as timber preservative to be the possible sources.

4. Marine TBT hotspot locations in Norway versus elsewhere

In Norway, TBT became in the early 1990s one of the mandatory determinants for sediment pollution monitoring surveys (Norwegian Environment Agency, 2018). A considerable amount of sediment TBT data has subsequently been registered in the national Vannmiljø [“Aquatic environment”] database.² Olsen et al. (2021) used this database to conduct a ranking of marine sediment locations along the Norwegian coastline for their levels of different priority pollutants. For TBT, the study identified 95 locations (Fig. 2) for which the highest detected TBT levels in surficial sediments were greater than a maximum tolerable risk (MTR) limit of 35 µg.kg⁻¹ d.w., i.e., a special limit defined for management of TBT polluted sediments (Norwegian Environment Agency, 2018). TBT data from the top nine locations are listed in Table 1. Among these sites, the highest TBT mean concentration values per site exceeded 30 000 µg.kg⁻¹ d.w. whereas the highest TBT concentrations detected exceeded 100 000 µg.kg⁻¹ d.w. (Table 1). For comparison, TBT concentration data from highly polluted sediments in other countries are shown in Table 2, although making simple comparisons of such hotspots can be challenging due to differences in

² The Norwegian Vannmiljø [“Aquatic environment”] database.

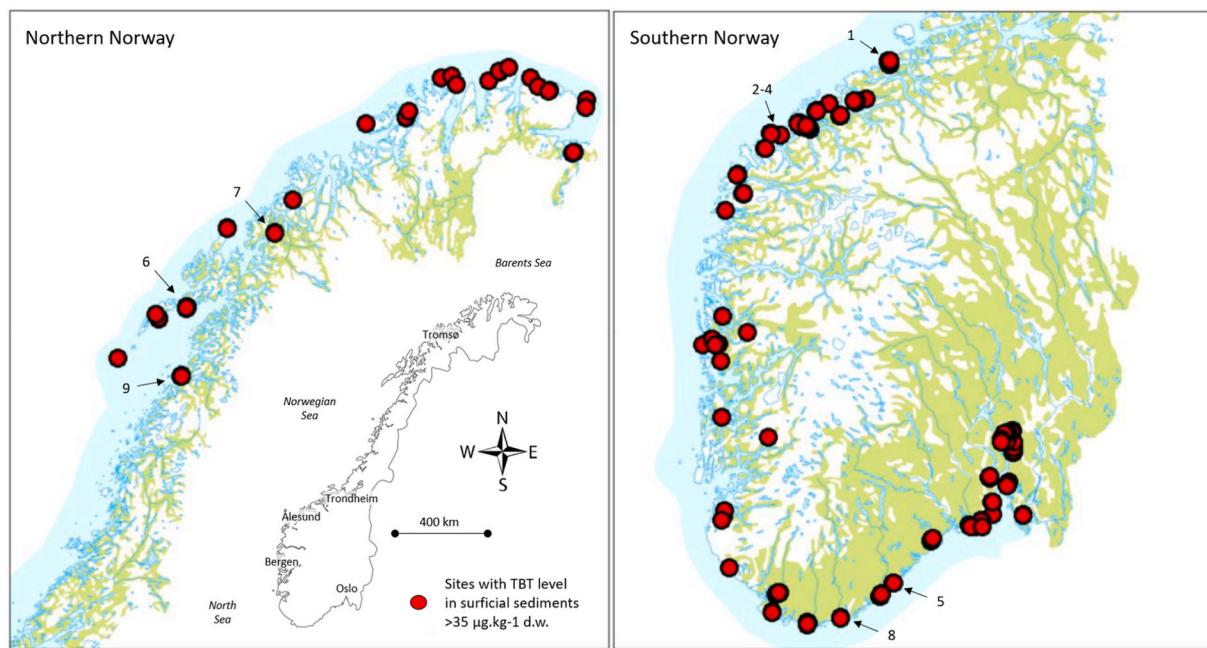


Fig. 2. Map showing the geographical distribution of TBT hotspots along the Norwegian coastline, highlighting 95 costal sites (red dots) where TBT contents in surficial sediments have been quantified (after 2006) at concentrations above the maximum tolerable risk (MTR) limit of $35 \mu\text{g}\cdot\text{kg}^{-1}$ d.w., according to data registered in the *Vannmiljø* database. Arrows and numbers 1–9 indicate the top nine sediment sites listed in Table 1. Data and map reproduced from Olsen et al. (2021) with modifications.

Table 1

The top nine TBT hotspots in Norway based on data registered in the *Vannmiljø* database after 2007. TBT concentrations are shown as min, mean, max measurements data in marine surficial sediments. The data were obtained from the database mid-2021. N = number of samples analyzed per sediment location.

Site*	Location (and city/place) in Norway	Depth in sediment (cm)	Sediment TBT ($\mu\text{g}\cdot\text{kg}^{-1}$ d.w.)			N	Study report
			Min	Mean	Max		
1	Kristiansund – inner harbor	0–10	48.1	30 088	79 800	8	Olsen et al. (2021)
2	By Fiskerstrand yard (Sula, Ålesund)	0–2	1270	29 433	101 000	11	Fagerhaug (2009)
3	Åregjerdevågen (Sula, Ålesund)	2–4	210	25 270	196 600	10	Haveland and Hestholm (2008b)
4	Eidssundet by Vegsund yard (Sula, Ålesund)	2–4	2500	23 962	110 000	8	Haveland et al. (2008a)
5	Vikkilen fjord - inner part	0–5	770	8226	93 000	15	Bakke et al. (2008)
6	Svolvær harbor	0–10	0.04	8160	90 000	89	Olsen et al. (2021)
7	Gisundet by Finnsnes	0–10	0.7	6217	55 900	9	Olsen et al. (2021)
8	Kristiansandsfjord - inner part	0–10	1.0	4365	60 000	19	Olsen et al. (2021)
9	Bodø harbor	0–10	1.0	958	48 600	95	Kaurin et al. (2015)

* Geographical locations of sites 1–9 are indicated at the map in Fig. 2.

methodologies (discussed later). Nevertheless, a comparison of these data suggests Norwegian TBT hotspot sediments to be comparable to the most contaminated TBT hotspots internationally, and not surprisingly, the TBT hotspots are typically located close to shipyards or within harbor/port areas. This spatiality confirms TBT AF paint and ship hulls maintenance operations to be key sources for this contamination. However, it is striking to note that while the TBT hotspots in other countries are found in conjunction with major ports and shipyards, the typical Norwegian hotspots are found in conjunction with relatively small yards and harbor/port locations (e.g., fishing villages), spread literally all along the long coastline (Fig. 2). The surficial sediment strata that have formed after the TBT AF paint ban would be expected to contain lower TBT levels than the older strata found underneath. TBT contamination levels in historical layers of sediment core samples have been studied in the two fjords Oslofjord and Drammensfjord in Norway (Dolven and Alve, 2018; Ekeroth et al., 2020), which both are fjords that have significant shipping activities. The TBT levels showed a gradual increase through the 1960s–70s, towards peak concentrations detected in sediment strata from the 1980s–90s, with Drammensfjord showing TBT levels above $2500 \mu\text{g}\cdot\text{kg}^{-1}$ d.w. at the highest, followed at both

places by significantly declining levels in sediments strata from the recent decades, but with the average TBT concentration in the youngest sediment layers in Drammensfjord still being significantly above the MTR limit of $35 \mu\text{g}\cdot\text{kg}^{-1}$ d.w. (Dolven and Alve, 2018; Ekeroth et al., 2020). The peak TBT concentrations, recorded in Drammensfjord, were at a level similar to the highest concentrations detected in Rotterdam harbor during the 1990s, as reported by Stronkhorst (1996) who found TBT concentrations up to about $2100 \mu\text{g}\cdot\text{kg}^{-1}$ d.w. in sediments from the inner and most polluted part of the harbor.

The presence of abraded TBT antifouling paint particles (TBT APPs) is likely to be a key reason why sediments close to shipyards may have extreme levels of TBT and also why TBT monitoring data from the same geographical area may vary so much (Table 1, Table 2). The relevance of TBT APPs for ecotoxicological studies in marine TBT hotspots was highlighted by Elgethun et al. (2000) and this issue has recently gained increasing attention, e.g. (Turner, 2010; Eklund et al., 2014; Choi et al., 2014; Batista-Andrade et al., 2018; Soroldoni et al., 2018; Muller-Karanassos et al., 2019; Abreu et al., 2020; Alshemmary et al., 2020; Soroldoni et al., 2020; Abreu et al., 2021; Muller-Karanassos et al., 2021; Soon et al., 2021; Soroldoni et al., 2021; Moreira et al., 2021; Torres and

Table 2

Examples of TBT concentration in hotspots from countries outside Norway. The shown locations are ranked from high to low levels based on the maximum levels of TBT or sum butyltins.

Site	Depth in sediment (cm)	Sediment TBT concentrations ($\mu\text{g. kg}^{-1}$ d.w.)	Study report
Hong Kong shipyards	0–2	Maximum 129 320 ^a Average median 1220 ^b	Ko et al. (1995)
Ulsan Bay shipyard, South Korea	0–2	Max 112 484 ^a	Shim et al. (2002)
Kelly Boatworks-Shipyard, USA	Surficial sediment ^c	Max (with paint chips) 68 613 Without paint chips 1196	Elgethun et al. (2000)
Shipyard, Portland and Boothbay Harbor, USA	0–2	Maximum: 30 256 ^a	Page et al. (1996)
Falmouth Dockyard, Port Pendennis, UK (1997 survey)	Surficial sediment ^c	Maximum: 20 700	Langston and Burt (2007)
Caldera Region, Chile	0–2	Means: 4015–16 932 ^b	Mattos et al. (2017)
Korean coastal waters	0–4	348–9576 highest concentrations for each sampling year	Choi et al. (2009)
Port of Gdańsk and Gdynia, Poland	Surficial sediment	14–8100 (sum butyltins), highest mean TBT: 4733	Filipkowska and Kowalewska (2019)
Imbituba Harbor, Brazil	0–10	Highest mean (n = 3): 9183 ^b	de Oliveira et al. (2010)
Toulon Bay, Mediterranean coast of France	0–5	Maximum: 6600 ^a	Pougnet et al. (2014)

^a TBT analysis results originally expressed on the basis of tin (Sn) content are converted to TBT content by multiplication by a factor of 2.44, but not normalized to TOC.

^b TBT values normalized to 5% TOC by multiplying the measured value with 5 and then divided by the actual TOC content of the sediment where TOC was measured.

^c Not specified.

De-la-Torre, 2021; Turner, 2021; Lopez et al., 2022; Sparks and Awe, 2022; Uc-Peraza et al., 2022b). Several studies express interest for TBT APPs as ecotoxicologically relevant forms of secondary microplastic particles, i.e., being synthetic polymers or resins which carry high relative concentrations of toxic biocidal compounds that have the capacity to cause adverse effects in many sediment related species, e.g. (Muller-Karanassos et al., 2021; Torres and De-la-Torre, 2021; Turner, 2021; Sparks and Awe, 2022), making the occurrence of TBT APPs in coastal, marine environments a major environmental concern. Regrettably, surveys in Norwegian hotspots, have typically not systematically considered the issue of TBT APPs, which is a weakness of the TBT data registered in the Vannmiljø database.

5. Environmental regulation and management of marine TBT hotspots

Sediment remediation measures, such as dredging and capping, are common management actions for improving the quality status of marine TBT hotspot sediments. Such operations are very costly, and their implementation must therefore be based on appropriate environmental regulations. In this context, national guidelines which instruct the processes of ecological risk assessment, quality classification and remediation management of polluted sediments are important. These regulations and guidelines at the national level should also comply with transnational agreements and mechanisms such as the EU Water Framework Directive (WFD) and the Marine Strategy Framework Directive (MSFD), which have the superior goals of establishing good environmental status (MSFD) and good ecological status (WFD)

throughout European waters. In Norway, the first quality assessment and classification system for marine systems was implemented in 1997, with a five-class scheme based principally on chemical concentrations of selected priority pollutants quantified in specified marine matrices (water, sediment or biota) (Molvær et al., 1997). In 2007, the Norwegian Environment Agency (NEA) replaced this concentration-based scheme with a risk-based (or effect-based) quality classification scheme, which defined quality classes (Table 3) based on the risk/liability for eco-toxic impacts of priority contaminants and the use of safety factors to compensate for intrinsic uncertainties that existed regarding their possible long term ecological effects. Due to the high toxicity of TBT to several non-target marine organisms, the effect/risk-based quality classes for this sediment contaminant were set to exceptionally low concentration levels (Table 3). These strict quality classes represented a challenge, firstly as analysis of TBT concentrations to such low levels with required precision and accuracy is almost technically unfeasible, as the LOQ for sediment TBT analyses for most chemical analysis laboratories is $1 \mu\text{g. kg}^{-1}$ d.w. Secondly, the classes were defined so low that surficial sediments, obtained even from relatively pristine locations along the Norwegian coastline, would most likely not meet the “good quality” demand based on their TBT content. To untie this Gordian knot, NEA therefore decided to implement a preliminary and more liberal quality classification scheme for “management purposes” especially for TBT concentrations in marine sediments (Table 3), and also to define the special MTR limit for management of TBT polluted sediments at $35 \mu\text{g. kg}^{-1}$ d.w. (Norwegian Environment Agency, 2018). If sediments exceeded this MTR limit, it will trigger demands for a more thorough risk assessment study and secondly potentially lead to demands for active sediment remediation actions. Large scale sediment remediation projects have now been initiated or already performed in some of the worst Norwegian TBT hotspots, while others are being planned. Further descriptions of these efforts fall beyond the scope of this paper. But interestingly, also the EU WFD has seen the rationale for implementing quality standards for TBT pollution in marine surficial sediments that are more realistic. In a report to the EU WFD, Sahlin and Ågerstrand proposed that a sediment based quality standard for TBT of $1.6 \mu\text{g. kg}^{-1}$ d.w. (normalized to 5% TOC level) would most likely be protective for sediment dwelling species in both freshwater and marine systems (Sahlin and Ågerstrand, 2020).

6. Toxicity mechanisms of TBT induced imposex in neogastropoda

The discovery of imposex in the early 1970s created a need for clarifying the effect mechanisms and the Modes of Actions (MoA) involved. The studies were in the earliest phase preferably addressing possible inhibitory and modulative interactions of TBT to metabolic pathways and detoxifying enzyme systems in different model species, particularly including the cytochrome P-450 dependent monooxygenase system and glutathione S-transferases, e.g., (Fish et al., 1976). Eventually, the research narrowed in on endogenous hormones regulation systems, e.g., (Lee, 1991), and later to control mechanisms of gene transcription mediated via intracellular and nuclear receptors, e.g., (Evans, 1988). During the following years of research, three major imposex MoA hypotheses have been described, namely: the increased testosterone MoA, the APGWamide neuropeptide activation MoA, and the retinoid X-receptor (RXR) agonism/activation MoA. An overview of key research reports that have been in support of each respective imposex MoA is shown in Table 4, and a brief outline of these three areas of research is described next.

The first major attempt to elucidate the imposex MoA was the “increased testosterone” or “increased androgen” hypothesis which alleged that TBT acted by inhibiting the enzymatic conversion of androgens to estrogens by the P-450-dependent aromatase system (CYP19A), causing a shift in the androgen-estrogen balance in females, and thereby making them start to develop and grow male sexual

Table 3

Effect/risk-based and management-based quality classification system for TBT concentrations in sediments according to the Norwegian guidelines for risk assessment of contaminated sediments (M-409 & M-1132/2018^a) and the Norwegian quality standards for water, sediment and biota (M-608/2016-revised 2020^b).

Sediment quality scheme on TBT concentration	Unit	Class I Background	Class II Good	Class III Moderate	Class IV Bad	Class V Very bad
Effect/risk-based	µg.kg ⁻¹ d.w.		0–0.002	0.002–0.016	0.016–0.032	>0.032
Management based ^c		0–1	1–5	5–20	20–100	>100

^a Norwegian Environment agency - [Guidelines for risk assessment of contaminated sediments](#).

^b Norwegian Environment agency - [Quality standards for water, sediment and biota](#).

^c The maximum tolerable risk (MTR) limit for management of TBT polluted sediments is set to 35 µg.kg⁻¹ d.w.

Table 4

Research publications that have supported each of the three competing imposex MoA hypotheses in marine neogastropods, all implying a receptor-based gene activation scheme.

Imposex MoA hypothesis	Published reports supporting each respective MoA
Increased testosterone	Bettin et al. (1996); Oehlmann et al. (1996); Ronis and Mason (1996); Matthiessen and Gibbs (1998); Alzieu (2000); Oberdörster and McClellan-Green (2002); Gooding et al. (2003); Santos et al. (2005); LeBlanc et al. (2005); Oehlmann et al. (2007); Stange et al. (2012)
APGWamide neuropeptide activation	Oberdörster and McClellan-Green (2000, 2002, 2003); Oberdörster et al. (2005)
Retinoid X Receptor (RXR) agonism/activation	Nishikawa et al. (2004); Nishikawa (2006); Castro et al. (2007); Iguchi et al. (2007); Nakanishi (2007); Horiguchi et al. (2008); Sternberg et al. (2008); Nakanishi (2008); le Maire et al. (2009); Horiguchi et al. (2010a); Horiguchi et al. (2010b); Sternberg et al. (2010); Lima et al. (2011); Urushitani et al. (2011); Stange et al. (2012); Chapman and Guillette (2013); Pascoal et al. (2013); Dominguez-Ojeda et al. (2014); Urushitani et al. (2018); Lagadic et al. (2018); Giraud-Billoud and Castro-Vazquez (2019); Giulianelli et al. (2020)

characteristics, notably vas deferens and penis analogue tissues (Bettin et al., 1996). A range of the early imposex MoA studies were in support of this theory (see Table 4). This MoA research direction also involved possible involvements of other enzyme targets and systems such as the steroid 5α-reductase 3(α),β-hydroxysteroid dehydrogenase pathway and UDP-glucuronosyltransferase (UGT) phase II enzymes (which are key for glucuronidation and the subsequent excretion of testosterone). However, this hypothesis implies strong similarities of steroidogenesis in neogastropoda and vertebrates and that the different steroid compounds involved have similar sexual regulation roles in gastropods and vertebrates. Most recent research in this area tend to contradict both these assumptions (Mizuta and Kubokawa, 2007; Scott, 2012, 2013; Fodor et al., 2020).

The second MoA theory is known as the APGWamide hypothesis. In molluscs, the neuropeptide neurotransmitter APGWamide (Ala-Pro-Gly-Trp-NH2) is produced and released from neurosecretory cells in the pedal ganglia. In gonochoristic (distinct sexes) neogastropods, APGWamide is key for regulating sexual development and reproductive behaviors in males (Smit et al., 1992; LeBlanc et al., 1999; LaFont, 2000). The APGWamide MoA theory alleges that TBT exposed young female gastropods will develop abnormally high concentrations of APGWamide, making them start developing male-like tissues which subsequently will start producing androgens that further promote the male-like sexual growth (including penis development and spermatogenesis). This theory is sometimes called the penis morphogenic factor (PMF) hypothesis. However, since this MoA theory build on the premise of androgen mediated signaling and regulation in females, it suffers many of the same weaknesses as the increased testosterone hypothesis.

The third imposex MoA theory is the Retinoid X Receptor (RXR) agonist/activation hypothesis, which alleges that TBT causes an inappropriate activation of the RXR signaling pathway, either directly by

ligand binding or indirectly by increasing the concentration of endogenous free retinoid (Sternberg et al., 2010). A key evidence for this MoA hypothesis is the induction of imposex in female neogastropods when treated with the natural RXR ligand 9-cis retinoic acid (9-cis RA), in absence of TBT (Castro et al., 2007). The indirect-effect sub theory suggests that TBT can inhibit acyl coenzyme A:acyltransferase (AXAT) and cause increased endogenous retinoid levels (due to a lowered retinoid AXAT esterification activity). Then, the increased free retinoid level will activate RXR and trigger the development/growth of imposex phenotypes (Sternberg et al., 2010). A recent study by Lesoway and Henry (2021) demonstrated retinoid agonists to promote penis development in the sequentially hermaphroditic *Crepidula* gastropod (which first develops into males, then change sex, and finally matures into females), and that blocking RXR or retinoic acid synthesis decreased penis length in this species. Interestingly, the RXR signaling pathway may also be involved in the organotin induced oyster shell thickening phenomenon. Huang et al. (2020) investigated the possible role of RXR in TBT and triphenyltin (TPhT) induced shell thickening in the Pacific oyster and found that both compounds influenced the shell development process by interaction with RXR and by disruption of this signaling pathway.

RXRs are members of the steroid and thyroid hormone nuclear receptor superfamily and act as dimer complexes, as homodimers or heterodimer partners of class II nuclear receptors, and most often together with the Peroxisome Proliferator-Activated Receptor (PPAR). The PPAR-RXR heterodimer complex regulates a range of cellular processes, including cellular development and differentiation, metabolism, and cell death. A diversity of regulatory functions of PPAR-RXR is possible partly because RXR and PPAR occur in different subtypes (RXR-α, -β, -γ and PPAR-α, -δ, -γ), and with the different combinations of PPAR-RXR subtypes pairs having different functions and roles in transcription of many different genes and gene clusters. Insight on these matters may have great relevance also for deciphering how TBT may exert endocrine disruptive actions to various biological control systems and why certain species and life-stages may have different sensitivities to endocrine disruptive influence from TBT. It should also be noted that an interaction of TBT directly with PPAR may possibly be important for imposex development as well as for other TBT ecotoxicities. For example, the treatment of dogwhelks with the known vertebrate PPAR ligand rosiglitazone has been shown to induce imposex development in absence of TBT, suggesting that PPAR could be more directly involved in modulating this endocrine signaling pathway (Pascoal et al., 2013).

Species variability in toxic sensitivity of TBT can possibly be explained by natural differences in how strong TBT bind to (and activate) RXR. Hence, it is interesting to evaluate RXR protein sequence similarities across species, and compare data from sensitive species, such as the common dogwhelk, with species that have unknown sensitivity to TBT. As part of this study, we therefore did some initial screening tests employing an established *in silico* bioinformatic tool SeqAPASS (Sequence Alignment to Predict Across Species Susceptibility) which is developed by the US EPA (LaLone et al., 2016) to compare RXR protein and conserved ligand-binding domain (LBD) sequences across a range of species, aiming particularly to assess percentage similarity to dogwhelk RXRs. With sequence information from the NCBI protein database (<https://www.ncbi.nlm.nih.gov>)

([://www.ncbi.nlm.nih.gov/guide/proteins/](http://www.ncbi.nlm.nih.gov/guide/proteins/)), we aligned the *N. lapillus* RXR- α (NCBI accession ABS70715.1) and RXR- β (ABS70716.1) protein sequences as well as their conserved LBDs (NCBI accession cd06943) against similar data from a broad suite of other species. Two illustrative examples of this similarity screening are shown below (Fig. 3a and b), ranking species and species groups from highest to lowest on similarity with LBD of RXR- α and RXR- β protein sequences of *N. lapillus*. A large number of species showed high sequence similarities with LBD in *N. lapillus*, for example above 75% similarity, whereas others showed

considerably lower similarity (Fig. 3a and b). Although being preliminary results, we see these findings as quite encouraging as they could mean that SeqAPASS can be used for targeting particularly sensitive and relevant bioindicator species in future effect studies of TBT, especially when effects are expected to be mediated via an RXR agonism/activation MoA, such as RXR- α and/or RXR- β ligand binding or modulations, but which will lead to different effect phenotypes than imposex, as imposex is exclusively found in neogastropoda. A key purpose of using such a SeqAPASS based screening, can be to filter out

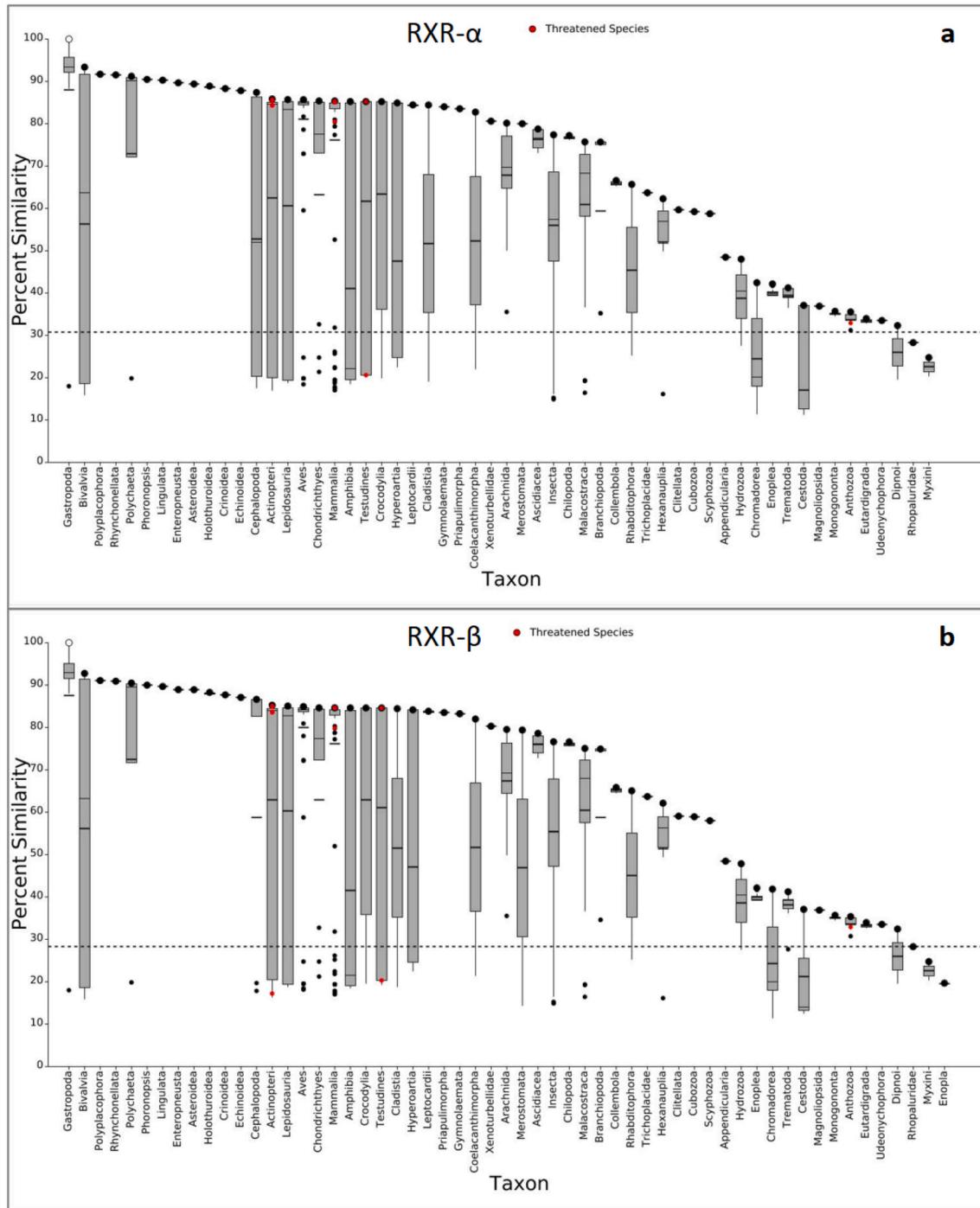


Fig. 3. Protein sequence similarities in the ligand binding domain (LBD) for retinoid X receptor alpha (RXR- α , a) and beta (RXR- β , b) for different species groups compared relatively to *Nucella lapillus*. High degree of similarity could be a screening indicator for the sensitivity to effects of tributyltin (TBT), and similarly acting toxicants, mediated via the RXR signaling pathway. The dashed lines indicate the susceptibility cut-offs (above: susceptible species groups; below: non-susceptible species groups). The red dots indicate species groups that are listed as threatened by the US Fish and Wildlife Service.

particularly sensitive and susceptible taxa, providing improved species targeting and significantly reduced workload for risk assessors. To discuss this issue further, we will in the following paragraph briefly examine the status of knowledge on TBT effects in various non-gastropod taxa.

7. Ecotoxicity of TBT in other species than neogastropods

Marine crustaceans constitute a large group, >50 000 known species, which include many bioindicator species possibly suitable for TBT effect studies (Verslycke et al., 2005; Parmentier et al., 2019). For example, McClellan-Green et al. (2007) pointed to commonly used crustacean bioindicator/bioassay species, including copepods like *Amphiascus tenuiremis*, *Tisbe bulbisetosa*, and *Acartia tonsa*, as well as the grass shrimp *Palaemonetes pugio* as particularly relevant bioindicator species in studies of TBT toxicokinetics and TBT effects on toxicological endpoints such as development, morphology, behavior and survival. The endocrinology of crustaceans often concerns the roles of ecdysone and peptide hormones in processes of molting and vitellogenesis. The main neuroendocrine center of crustaceans is the X organ–sinus gland system, which is located inside decapods' eyestalk and which produce a family of large peptide hormones including the crustacean hyperglycemic hormone (CHH), putative molt-inhibiting hormone (MIH) and vitellogenesis-inhibiting hormone (VIH) (Keller, 1992). The growth process in crustaceans, as in other ecdysozoans, occurs by the repeated molting process regulated by a negative feedback mechanism involving CHH, MIH, and ecdysteroids, and which potentially can be influenced by TBT toxicity (Vogt et al., 2018). For example, the study of Kusk and Petersen (1997) found larval development in *Acartia tonsa* to be extremely sensitive to TBT with inhibition of larval development in a standard 48h biotest occurring already at a TBT concentration of 1 ng. L⁻¹, indicating a similar sensitivity as for imposex in *N. lapillus*. Besides ecdysteroids, the sesquiterpenoids methyl farnesoate (MF) and juvenile hormone are also important during the growth and metamorphosis of crustaceans. In crustaceans, TBT has been shown to impair reproductive performance (IPCS, 1990), decrease neonate survival (Kusk and Petersen, 1997), inhibit larvae developmental ratios (Waldock et al., 1999), decrease juvenile growth rates (Dahllof et al., 2001), and induce changes to the local community structure (Takeuchi et al., 2001; Aono and Takeuchi, 2008). By interacting with RXR and CHH signaling, TBT can influence carbohydrate and lipid homeostasis and may also disrupt methyl farnesoate and ecdysteroid signaling via interactions with other nuclear receptors and thereby alter growth and sexual maturity, respectively, and also influence reproduction via interference with cytochrome P450 system mediated steroid metabolism (Vogt et al., 2018). The brown shrimp *C. crangon* may be considered as a particularly relevant species for assessing endocrine disruption effects in crustaceans in coastal TBT hotspot locations. Studies have shown that TBT concentrations in *C. crangon* in estuarine, coastal and offshore areas are clearly related to sediment concentrations (Verhaegen et al., 2012; Parmentier et al., 2019). Research into possible endocrine disruption impacts of TBT in *C. crangon* have focused on agonistic interference of TBT with natural ecdysteroid hormones at the metabolic pathways that regulates growth and reproduction (Verhaegen et al., 2011).

Ascidians, or sea squirts, are sessile, filter-feeding tunicates that receive increasing attention for their possible role as bioindicators in endocrine disruption effect studies of TBT and other organotins. These animals are close evolutionary relatives to vertebrates as are demonstrated by their free-swimming, tadpole like, lecithotrophic larvae (which has a notochord in the tail and a dorsal hollow nerve cord). The phylogenetic position of tunicates, located between invertebrates and vertebrates, makes them particularly interesting as possible models and bioindicators for endocrine disruption effect studies in the laboratory as well as in marine surveys, e.g. Osugi et al. (2020); Eliso et al. (2020); Sekiguchi et al. (2020). Developmental and endocrine disruption effects of TBT in ascidian test species have been reported in several studies, e.g.

(Gianguzza et al., 1996; Patricolo et al., 2001; Dolcemascolo et al., 2005; Cangialosi et al., 2008, 2009, 2010; Mansueto et al., 2011). For example, exposure of *Ciona* to TBT was found to yield changes to sterols and sex steroid levels as well as ovarian morphology (Cangialosi et al., 2010).

Fish have frequently been used as test animals and bioindicators in effect studies with TBT and other organotins, e.g. including commercial species such as Japanese medaka (*Oryzias latipes*), (Horie et al., 2018), red seabream (*Pagrus major*) and black rockfish (*Sebastodes melanops*) (Min et al., 2018), and model species such as zebrafish (*Danio rerio*) (Li and Li, 2020a, b; Li and Li, 2021). Exposure of fish to TBT, or other toxic trivalent OTs, leads to bioaccumulation *in vivo* (Miki et al., 2011; Ashraf et al., 2017) and have been linked to a series of toxic impacts, including inhibited growth (IPCS, 1990), induced masculinization (Fent, 1996; Shimasaki et al., 2003), increased sperm abnormalities (McAllister and Kime, 2003), reduced fecundity (Shimasaki et al., 2003), inhibited cytochrome P450 activities (Zhang et al., 2009), impaired ovarian development (Zheng et al., 2005), embryo abnormalities (Zhang et al., 2007), larval malformations (Zhang et al., 2008), increased liver vacuolation (Zuo et al., 2009), induced hematopoietic tissue hyperplasia (Zuo et al., 2012), neurotoxicity (e.g. modulation of the glutamate signaling pathway) (Zhang et al., 2011), increased DNA damage (Zhang et al., 2013), impaired thyroid function (Wu et al., 2020; Li and Li, 2020a, 2021), and induced lipotoxicity (Zhang et al., 2013, 2016; Lyssimachou et al., 2015). Fish species that could be particularly relevant for effect surveys in TBT hotspot field site will be common and shallow-living species that also are relatively territorial/non-migratory and species that because of their feeding or other behaviors are in frequent and direct contact with TBT contaminated surficial sediments, including for example various kinds of Gobiidae species, e.g., (Shimizu and Kimura, 1992; Louiz et al., 2009, 2018).

Marine mammals bioaccumulate organotins via food web accommodated uptake (Berge et al., 2004) potentially leading to several toxic and endocrine-disrupting effects that are known to or expected to be mediated via an RXR agonism/activation MoA (Grün et al., 2006; Evans and Mangelsdorf, 2014). Reported TBT effects in mammals include reproductive anomalies (e.g. reduced spermatogenesis and embryo malformations) (IPCS, 1990), neurobehavioral alterations (Yonezawa et al., 2007), immunological disorders (Antizar-Ladislao, 2008), abnormal adipose tissue differentiation (obesity) (Grün and Blumberg, 2006; Ohtaki et al., 2007), induced cardiovascular toxicity (Chen et al., 2008), inhibited osteoclast differentiation through a retinoic acid receptor-dependent signaling pathway (Yonezawa et al., 2007; Nath, 2008), inhibited mitochondrial ATP synthase activity and disturbed biosynthesis and degradation of steroids (Kotake, 2012), induced behavioral abnormality and toxicity to the developing central nervous system (da Silva et al., 2018).

Human health risks, associated with TBT and other OTs, is a side-issue in the present review, as our focus is predominantly on marine organisms. Exposure, toxicity and risk assessments of TBT/OTs in humans are covered in detail by others, e.g., Antizar-Ladislao (2008) and Heindel and Blumberg (2019). Human risks of TBT are most often associated with consumption of contaminated seafoods. Food safety authorities in Norway and other European countries normally find seafoods to be within food safety limits for TBT, unless the seafoods have been collected from TBT hotspot areas, which generally is not recommended (EU, 2003; EFSA, 2004; EU, 2006; VKM, 2007). In humans (and other vertebrates), endocrine disruption phenomena of TBT may more likely involve increased testosterone as a MoA, in contrast to in molluscs and less advanced taxa (Thibaut and Porte, 2004; Lv et al., 2021). In humans, an issue of particular importance is the potential of TBT/OTs to act as obesogens and to interfere with the endocrine regulation of adipogenesis (Darbre, 2017). Both TBT and TPhT have been demonstrated to act as nanomolar affinity ligands for the PPAR γ -RXR heterodimer and to stimulate preadipocytes to differentiate into adipocytes in a PPAR γ -dependent manner (Kanayama et al., 2005; Grün et al., 2006; Grün and Blumberg, 2006; Li et al., 2011). Much research is presently

ongoing on possible roles of TBT and OT pollutants in conjunction with human obesity.

8. Summary and remaining knowledge needs

TBT would possibly still have been the key marine antifoulant globally if ecotoxicologists hadn't discovered its many non-target impacts in marine systems. This review provides a brief outline of the large study field that concerns TBT. We examine the knowledge status and discuss the rationale for why it can be worthwhile to address TBT impacts, especially in marine TBT hotspots, of which Norway have many. Away from TBT hotspots, the adverse impacts of TBT have fortunately been in decline, e.g., (Schøyen et al., 2019), but within hotspots the recovery process is apparently far slower and the possible ecological impacts in biota have been investigated only to a limited degree. In our group, we consider such TBT hotspots almost as "natural laboratories" in which the fate and effect aspects of TBT, and co-occurring contaminants, can be thoroughly investigated. Key fate issues that can be studied may for example include aspects like the bioavailability of legacy TBT to different local species and species groups, the efficiency of measures to reduce pollution exposures (sediment remediation operations), natural recovery processes of hotspot sediments under different local conditions, and the role of TBT APPs at sites highly littered with such particles. Studies in hotspots will possibly be particularly suitable for elucidating the MoAs and toxic mechanisms of TBT in various bio-indicator species (including both invertebrates and vertebrates). Since the TBT contamination in hotspot sites is preferably embedded within sediments, it is particularly sediment-associated species that are likely to be most relevant as bioindicator study species. Such species may for example include the netted dogwhelk *Nassarius reticulatus*, crustaceans like the brown shrimp *C. crangon* and benthic fish species such as gobies (e.g., like the sand goby *Pomatoschistus minutus*). While endocrine disruption studies in vertebrates are generally benefited by the broad knowledge base that is available on vertebrate endocrine systems, similar knowledge is much more limited for invertebrates, possibly with pest insects as an exception. As discussed by Fodor et al. (2020), effect studies of ED toxicants such as TBT in invertebrates must not be focused on effect pathways that are valid for vertebrates but not for invertebrates.

The most extreme TBT hotspots in Norway are located close to shipyards or in busy fishery harbors (Olsen et al., 2021), where active sediment remediations are likely to be required, if not already performed. A current study in our group has addressed spatiotemporal fate and effect issues in the fjord Vikkilen, i.e., one of the TBT hotspots listed in Table 1 (Schøyen et al., 2022). It is important that such studies are designed and performed so they can provide improved insights into the remaining unknowns of TBT as an endocrine disrupting compound. Such improved insights are important for assessing the operability and realism of sediment quality and risk assessment standards that are authorized by international and/or national bodies. Such standards are the benchmarks that should signal the need for remediation measures and need to be set right in order not to be under-protective nor over-protective. However, the many TBT hotspot sites along the Norwegian coast that show extreme TBT levels, even on a global context, indicates a need for thorough quality verification of TBT data that have been registered in national databases. And the circumglobal presence of TBT contamination and TBT hotspots suggests that this issue has international relevance.

Effect studies in TBT hotspots offer opportunities for detailed studies of endocrine effect mechanisms in various groups of marine bioindicator species, especially regarding nuclear receptor effect pathways. MoAs for TBT induced endocrine disruption effects in invertebrate taxa warrant further investigations, both regarding the critical biological events leading to known phenomena such as imposex in neogastropods, but also for other endocrine effect phenotypes that may causally be linked to TBT pollution in hotspot situations. Whether such endocrine effects can

be forecasted at an early stage using New Approach Methodologies (NAMs), such as high-throughput biomarker screening, broad-content ("OMICS") screening and computational predictions, is a topic relevant to the contamination situations in TBT hotspots and may have a broader importance for next generation environmental risk assessment. Similarly, there is still only limited insight on the possible long-term ecological impacts on populations and communities of benthic invertebrates that inhabit TBT hotspots, as well as on combined toxicity phenomena caused by TBT in combination with other inorganic/organic pollutants commonly occurring in hotspot areas. In these studies, different combinations of TBTs and other pollutants, a greater diversity of nuclear receptors and toxicity pathways, and combined toxicities expressed at different levels of biological organization (i.e., molecular, cellular, individuals and population effects) can and should be assessed. Exposure time is believed to influence the nature and intensity of effect interactions as well as the type and quantity of metabolic products which may have relevance for combined toxicity effects. Lastly, from a general perspective, it is essential that TBT fate and effect studies can be performed under exposure scenarios that are potent but still realistic, and such conditions are present in coastal TBT hotspots. Any improved insights from such studies will have relevance in conjunction with future quality verification, refinement and internationally harmonization of risk based environmental quality standards that should serve as regulatory trigger values for sediment remediation requirements in TBT polluted coastal sediments.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- Abel, R., 1990. The development of legislation on TBT antifoulants in the UK and Europe. *J. Oil Colour Chemists' Assoc.* 73, 332–&.
- Abraham, M., Westphal, L., Hand, I., Lerz, A., Jeschek, J., Bunke, D., Leipe, T., Schulz-Bull, D., 2017. TBT and its metabolites in sediments: survey at a German coastal site and the central Baltic Sea. *Mar. Pollut. Bull.* 121, 404–410.
- Abreu, F.E.L., da Silva, J.N.L., Castro, I.B., Fillmann, G., 2020. Are antifouling residues a matter of concern in the largest South American port? *J. Hazard Mater.* 398, 11.
- Abreu, F.E.L., Batista, R.M., Castro, I.B., Fillmann, G., 2021. Legacy and emerging antifouling biocide residues in a tropical estuarine system (Espirito Santo state, SE, Brazil). *Mar. Pollut. Bull.* 166, 10.
- Alshemmar, H., Al-Awadi, M., Karam, Q., Talebi, L., 2020. Sedimentary butyltin compounds and sediment transport model at the Shuaikh Port, Kuwait Bay. *Arabian J. Geosci.* 13, 11.
- Alzieu, C., 2000. Impact of tributyltin on marine invertebrates. *Ecotoxicology* 9, 71–76.
- Alzieu, C., Sanjuan, J., Deltreil, J.P., Borel, M., 1986. Tin contamination in Arcachon Bay – effects on oyster shell anomalies. *Mar. Pollut. Bull.* 17, 494–498.
- Amara, I., Miled, W., Ben Slama, R., Ladha, N., 2018. Antifouling processes and toxicity effects of antifouling paints on marine environment. A review. *Environ. Toxicol. Pharmacol.* 57, 115–130.
- Anastasiou, T.I., Chatzinikolaou, E., Mandalakis, M., Arvanitidis, C., 2016. Imposex and organotin compounds in ports of the Mediterranean and the Atlantic: is the story over? *Sci. Total Environ.* 569, 1315–1329.
- Antizar-Ladislao, B., 2008. Environmental levels, toxicity and human exposure to tributyltin (TBT)-contaminated marine environment. A review. *Environ. Int.* 34, 292–308.

- Aono, A., Takeuchi, I., 2008. Effects of tributyltin at concentrations below ambient levels in seawater on *Caprella danilevskii* (Crustacea : Amphipoda : caprellidae). Mar. Pollut. Bull. 57, 515–523.
- Arnold, C.G., Weidenhaupt, A., David, M.M., Muller, S.R., Haderlein, S.B., Schwarzenbach, R.P., 1997. Aqueous speciation and 1-octanol-water partitioning of tributyl- and triphenyltin: effect of pH and ion composition. Environ. Sci. Technol. 31, 2596–2602.
- Ashraf, M.W., Salam, A., Mian, A., 2017. Levels of organotin compounds in selected fish species from the arabian gulf. Bull. Environ. Contam. Toxicol. 98, 811–816.
- Bakke, T., Håvardstun, J., Nes, K., Schaanning, M., Oug, E., Rygg, B., 2008. Miljøtekniske undersøkelser ved Nymo AS i Vikkilen. Supplerende undersøkelser, risiko- og tiltaksverdning. Norsk Institutt for Vannforskning, Oslo, Norway, p. 82.
- Barroso, C.M., Moreira, M.H., Bebianno, M.J., 2002. Imposex, female sterility and organotin contamination of the prosobranch *Nassarius reticulatus* from the Portuguese coast. Mar. Ecol. Prog. Ser. 230, 127–135.
- Batista, R.M., Castro, I.B., Fillmann, G., 2016. Imposex and butyltin contamination still evident in Chile after TBT global ban. Sci. Total Environ. 566, 446–453.
- Batista-Andrade, J.A., Caldas, S.S., Batista, R.M., Castro, I.B., Fillmann, G., Primel, E.G., 2018. From TBT to booster biocides: levels and impacts of antifouling along coastal areas of Panama. Environ. Pollut. 234, 243–252.
- Bauer, B., Fioroni, P., Ide, I., Liebe, S., Oehlmann, J., Stroben, E., Watermann, B., 1995. TBT effects on the female genital system of *Littorina littorea* - a possible indicator of tributyltin pollution. Hydrobiologia 309, 15–27.
- Bech, M., 2002. A survey of impudicid mureids from 1996 to 2000 and identification of optimal indicators of tributyltin contamination along the east coast of Phuket Island, Thailand. Mar. Pollut. Bull. 44, 887–896.
- Berge, J.A., Brevik, E., Bjørge, A., Følsvik, N., Gabrielsen, G.W., Wolkers, H., 2004. Organotins in marine mammals and seabirds from Norwegian territory. J. Environ. Monit. 6, pp. 108–112.
- Berge, J.A., Amundsen, C.E., Eggen, T., Hylland, K., Bøe, E., 2006. Naturlig nedbryting og biotilgjengelighet av tinnorganiske forbinder i marine sedimenter. Norwegian Institute for Water Research (NIVA), Oslo, Norway, p. 72.
- Bettin, C., Oehlmann, J., Stroben, E., 1996. TBT-induced imposex in marine neogastropods is mediated by an increasing androgen level. Helgol. Meeresunters. 50, 299–317.
- Birchenough, A.C., Barnes, N., Evans, S.M., Hinz, H., Kronke, I., Moss, C., 2002a. A review and assessment of tributyltin contamination in the North Sea, based on surveys of butyltin tissue burdens and imposex/intersex in four species of neogastropods. Mar. Pollut. Bull. 44, 534–543.
- Birchenough, A.C., Evans, S.M., Moss, C., Welch, R., 2002b. Re-colonisation and recovery of populations of dogwhelks *Nucella lapillus* (L.) on shores formerly subject to severe TBT contamination. Mar. Pollut. Bull. 44, 652–659.
- Blaber, S.J.M., 1970. The Occurrence of a Penis-like Outgrowth behind the Right Tentacle in Spent Females of *Nucella lapillus* (L.). Proceedings of the Malacological Society of London. Dulau & Co., London, UK, pp. 231–233.
- Brändli, R.C., Breedveld, G.D., Cornelissen, G., 2009. Tributyltin sorption to marine sedimentary black carbon and to amended activated carbon. Environ. Toxicol. Chem. 28, 503–508.
- Bryan, G., Gibbs, P., 1991. Impact of low concentrations of tributyltin (TBT) on marine organisms: a review. In: Newman, M.C., McIntosh, A.W. (Eds.), Metal Ecotoxicology: Concepts and Applications. Lewis Publishers, Boca Raton, pp. 323–361.
- Bryan, G.W., Gibbs, P.E., Hummerstone, L.G., Burt, G.R., 1986. The decline of the gastropod *Nucella lapillus* around southwest England - evidence for the effect of tributyltin from antifouling paints. J. Mar. Biol. Assoc. U. K. 66, 611–640.
- Cacciatori, F., Noventa, S., Antoninini, C., Formalewicz, M., Gion, C., Berto, D., Gabellini, M., Brusa, R.B., 2018. Imposex in *Nassarius nitidus* (Jeffreys, 1867) as a possible investigative tool to monitor butyltin contamination according to the Water Framework Directive: a case study in the Venice Lagoon (Italy). Ecotoxicol. Environ. Saf. 148, 1078–1089.
- Cangialosi, M.V., Puccia, E., D'Agati, P., Saporito, L., Mazzola, A., Mansueto, C., 2008. Evidence for endocrine disruption of tributyltin chloride in ovary of *Gona intestinalis* (Ascidiae, Urochordata). Mar. Environ. Res. 66, 123–123.
- Cangialosi, M.V., Mansueto, V., Puccia, E., Corsi, I., Bonacci, S., Focardi, S., Mazzola, A., 2009. A biochemical study of the effects of tributyltin on unfertilized eggs, embryos and larvae of the sea squirt *Ciona intestinalis*. Caryologia 62, 309–315.
- Cangialosi, M.V., Puccia, E., Mazzola, A., Mansueto, V., Arukwe, A., 2010. Screening of ovarian steroidogenic pathway in *Ciona intestinalis* and its modulation after tributyltin exposure. Toxicol. Appl. Pharmacol. 245, 124–133.
- Castro, L.F.C., Lima, D., Machado, A., Melo, C., Hiromori, Y., Nishikawa, J., Nakanishi, T., Reis-Henriques, M.A., Santos, M.M., 2007. Imposex induction is mediated through the Retinoid X Receptor signalling pathway in the neogastropod *Nucella lapillus*. Aquat. Toxicol. 85, 57–66.
- Castro, I.B., Iannacone, J., Santos, S., Fillmann, G., 2018. TBT is still a matter of concern in Peru. Chemosphere 205, 253–259.
- Champ, M.A., 2000. A review of organotin regulatory strategies, pending actions, related costs and benefits. Sci. Total Environ. 258, 21–71.
- Chapman, R.W., Guillette, L.J., 2013. Contaminants and imposEX: transcriptomics of contaminant-induced sex change. Mol. Ecol. 22, 1485–1487.
- Chen, Y.F., Zuo, Z.H., Chen, S.Z., Yan, F.H., Chen, Y.X., Yang, Z.M., Wang, C.G., 2008. Reduction of spermatogenesis in mice after tributyltin administration. Toxicology 251, 21–27.
- Chiavarini, S., Massanisso, P., Nicolai, P., Nobili, C., Morabito, R., 2003. Butyltins concentration levels and imposex occurrence in snails from the Sicilian coasts (Italy). Chemosphere 50, 311–319.
- Choi, M., Choi, H.G., Moon, H.B., Kim, G.Y., 2009. Spatial and temporal distribution of tributyltin (TBT) in seawater, sediments and bivalves from coastal areas of Korea during 2001–2005. Environ. Monit. Assess. 151, 301–310.
- Choi, M., Moon, H.B., Yu, J., Cho, H., Choi, H.G., 2013. Temporal trends (2004–2009) of imposex in rock shells *Thais clavigera* collected along the Korean coast associated with tributyltin regulation in 2003 and 2008. Arch. Environ. Contam. Toxicol. 64, 448–455.
- Choi, J.Y., Hong, G.H., Ra, K., Kim, K.T., Kim, K., 2014. Magnetic characteristics of sediment grains concurrently contaminated with TBT and metals near a shipyard in Busan, Korea. Mar. Pollut. Bull. 85, 679–685.
- Clayton, M.E., Steinmann, R., Fent, K., 2000. Different expression patterns of heat shock proteins hsp 60 and hsp 70 in zebra mussels (*Dreissena polymorpha*) exposed to copper and tributyltin. Aquat. Toxicol. 47, 213–226.
- Coray, C., Bard, S.M., 2007. Persistence of tributyltin-induced imposex in dogwhelks (*Nucella lapillus*) and intersex in periwinkles (*Littorina littorea*) in atlantic Canada. Water Qual. Res. J. Can. 42, 111–122.
- Cornelissen, G., Pettersen, A., Nesse, E., Eek, E., Helland, A., Breedveld, G.D., 2008. The contribution of urban runoff to organic contaminant levels in harbour sediments near two Norwegian cities. Mar. Pollut. Bull. 56, 565–573.
- Cuevas, N., Alonso, J.I.G., Larreta, J., Rodriguez, J.G., Sariego, C., Zorita, I., 2014. Monitoring the effectiveness of the European tributyltin regulation on the Basque coast (northern Spain) by assessing imposex in two gastropod species (*Nassarius reticulatus* and *Nassarius nitidus*). Chem. Ecol. 30, 636–642.
- da Costa, M.B., Zamprogno, G.C., Otegui, M.B.P., de Moraes, L., Pedruzzi, F.C., Dalbem, G.B., da Silva, D.A., Resende, A.C.B., 2017. A temporal and spatial monitoring of organotin pollution in a harborside region of Brazil by imposex and ecological quality ratio using *Leucozonia nassa*. Environ. Monit. Assess. 189, 16.
- da Silva, I.F., Freitas-Lima, L.C., Graceli, J.B., Rodrigues, L.C.D., 2018. Organotins in neuronal damage, brain function, and behavior: a short review. Front. Endocrinol. 8, 6.
- Dahllof, I., Agrenius, S., Blanck, H., Hall, P., Magnusson, K., Molander, S., 2001. The effect of TBT on the structure of a marine sediment community - a boxcosm study. Mar. Pollut. Bull. 42, 689–695.
- Darbre, P.D., 2017. Endocrine disruptors and obesity. Curr. Obes. Rep. 6, 18–27.
- De Mora, S.J., Stewart, C., Phillips, D., 1995. Sources and rate of degradation of tri(n-butyltin in marine-sediments near Auckland, New-Zealand. Mar. Pollut. Bull. 30, 50–57.
- de Oliveira, C.R., dos Santos, D., Madureira, L.A.D., de Marchi, M.R.R., 2010. Speciation of butyltin derivatives in surface sediments of three southern Brazilian harbors. J. Hazard Mater. 181, 851–856.
- Diez, S., Abalos, M., Bayona, J.M., 2002. Organotin contamination in sediments from the Western Mediterranean enclosures following 10 years of TBT regulation. Water Res. 36, 905–918.
- Dolcemascolo, G., Gianguzza, P., Pellerito, C., Pellerito, L., Gianguzza, M., 2005. Effects of tri-n-butyltin(IV) chloride on neurulation of *Ciona intestinalis* (Tunicata, Asciidae): an ultrastructural study. Appl. Organomet. Chem. 19, 11–22.
- Dolven, J., Alve, E., 2018. Miljøgifter i indre Oslofjord: Kartlegging av historisk forløp gjennom analyser av utvalgte miljøgifter i daterte sedimentkjerner. Norconsult/Uio/USN, Oslo, Norway, p. 29.
- Dominguez-Ojeda, D., Rojas-Garcia, A.E., Robledo-Marenco, M.L., Barron-Vivanco, B.S., Medina-Diaz, I.M., 2014. Exposure to tributyltin chloride induces penis and vas deferens development and increases RXR expression in females of the purple snail (*Plicopurpura pansa*). Isj-Invertebrate Survival Journal 11, 204–212.
- Dowson, P.H., Bubb, J.M., Lester, J.N., 1993. Depositional profiles and relationships between organotin compounds in fresh-water and estuarine sediment cores. Environ. Monit. Assess. 28, 145–160.
- Dowson, P.H., Bubb, J.M., Lester, J.N., 1996. Persistence and degradation pathways of tributyltin in freshwater and estuarine sediments. Estuar. Coast Shelf Sci. 42, 551–562.
- EFSA, 2004. Opinion of the Scientific Panel on Contaminants in the Food Chain on a request from the Commission to assess the health risks to consumers associated with exposure to organotins in foodstuffs. Eur. Food Saf. Author. (EFSA), EFSA J. 1–119.
- Ekereth, N., Hjort, T., Stubø, E., 2020. Ren Drammensfjord - Sediment undersøkelse 2019. NIRAS, Drammen, Norway, p. 233.
- Eklund, B., Johansson, L., Ytreberg, E., 2014. Contamination of a boatyard for maintenance of pleasure boats. J. Soils Sediments 14, 955–967.
- El Ayari, T., Bierne, N., El Menif, N.T., 2018. Imposex incidence in *Stramonita haemastoma* (gastropoda: muricidae) from the mediterranean and atlantic coast after tributyltin global ban. J. Sea Res. 134, 10–15.
- Elgethun, K., Neumann, C., Blake, P., 2000. Butyltins in shellfish, finfish, water and sediment from the Coos Bay estuary (Oregon, USA). Chemosphere 41, 953–964.
- Eliso, M.C., Manfra, L., Savorelli, F., Tornambe, A., Spagnuolo, A., 2020. New approaches on the use of tunicates (*Ciona robusta*) for toxicity assessments. Environ. Sci. Pollut. Res. 27, 32132–32138.
- EU, 2003. Assessment of the Dietary Exposure to Organotin Compounds of the Population of the EU Member States EU Commission, p. 237.
- EU, 2006. Revised Assessment of the Risks to Health and the Environment Associated with the Use of the Four Organotin Compounds TBT, DBT, DOT and TPT. European Commission - SCHER, p. 27.
- Evans, R.M., 1988. The steroid and thyroid-hormone receptor superfamily. Science 240, 889–895.
- Evans, R.M., Mangelsdorf, D.J., 2014. Nuclear receptors, RXR, and the big bang. Cell 157, 255–266.
- Evans, S.M., Evans, P.M., Leksono, T., 1996. Widespread recovery of dogwhelks, *Nucella lapillus* (L.), from tributyltin contamination in the north sea and clyde sea. Mar. Pollut. Bull. 32, 263–269.

- Fagerhaug, A., 2009. Miljøgeologiske undersøkelser av verftsområde. Feltundersøkelse, risiko- og tiltaksutvurderinger. Multiconsult, Ålesund, Norway, p. 39.
- Fent, K., 1996. Ecotoxicology of organotin compounds. Crit. Rev. Toxicol. 26, 3–117.
- Filipkowska, A., Kowalewska, G., 2019. Butyltins in sediments from the Southern Baltic coastal zone: is it still a matter of concern, 10 years after implementation of the total ban? Mar. Pollut. Bull. 146, 343–348.
- Fish, R.H., Kimmel, E.C., Casida, J.E., 1976. Bio-organotin chemistry - reactions of tributyltin derivatives with a cytochrome-P-450 dependent monooxygenase enzyme system. J. Organomet. Chem. 118, 41–54.
- Fodor, I., Urban, P., Scott, A.P., Pirger, Z., 2020. A critical evaluation of some of the recent so-called 'evidence' for the involvement of vertebrate-type sex steroids in the reproduction of mollusks. Mol. Cell. Endocrinol. 516, 10.
- Folsvik, N., Berge, J.A., Brevik, E.M., Walday, M., 1999. Quantification of organotin compounds and determination of imposex in populations of dogwhelks (*Nucella lapillus*) from Norway. Chemosphere 38, 681–691.
- Furdek, M., Vahetic, M., Scancar, J., Milacic, R., Kniewald, G., Mikac, N., 2012. Organotin compounds in seawater and *Mytilus galloprovincialis* mussels along the Croatian Adriatic Coast. Mar. Pollut. Bull. 64, 189–199.
- Gabbianelli, R., Moretti, M., Carpene, E., Falcioni, G., 2006. Effect of different organotins on DNA of mollusk (*Scapharca inaequivalvis*) erythrocytes assessed by the comet assay. Sci. Total Environ. 367, 163–169.
- Galante-Oliveira, S., Oliveira, I., Ferreira, N., Santos, J.A., Pacheco, M., Barroso, C., 2011. *Nucella lapillus* L. imposex levels after legislation prohibiting TBT antifoulants: temporal trends from 2003 to 2008 along the Portuguese coast. J. Environ. Monit. 13, 304–312.
- Gianguzza, M., Dolcemascolo, G., Mansueti, C., Pellerito, L., 1996. Effects of tributyltin (IV) chloride exposure on larvae of *Ciona intestinalis* (Urochordata): an ultrastructural study. Appl. Organomet. Chem. 10, 405–413.
- Gibbs, P.E., 1993. A male genital defect in the dog-whelk, *Nucella lapillus* (Neogastropoda), favouring survival in a TBT-polluted area. J. Mar. Biol. Assoc. U. K. 73, 667–678.
- Gibbs, P.E., 2009. Long-term tributyltin (TBT)-induced sterilization of neogastropods: persistence of effects in *Ocenebra erinacea* over 20 years in the vicinity of Falmouth (Cornwall, UK). J. Mar. Biol. Assoc. U. K. 89, 135–138.
- Gibbs, P.E., Bryan, G.W., Pascoe, P.L., Burt, G.R., 1987. The use of the dog-whelk, *Nucella lapillus*, as an indicator of tributyltin (TBT) contamination. J. Mar. Biol. Assoc. U. K. 67, 507–523.
- Gibbs, P.E., Pascoe, P.L., Burt, G.R., 1988. Sex change in the female dog-whelk, *Nucella lapillus*, induced by tributyltin from antifouling paints. J. Mar. Biol. Assoc. U. K. 68, 715–731.
- Giltrap, M., Macken, A., Davoren, M., Minchin, D., McGovern, E., Foley, B., Strand, J., McHugh, B., 2009. Use of caged *Nucella lapillus* and *Crassostrea gigas* to monitor tributyltin-induced bioeffects in Irish coastal waters. Environ. Toxicol. Chem. 28, 1671–1678.
- Giraud-Billoud, M., Castro-Vazquez, A., 2019. Aging and retinoid X receptor agonists on masculinization of female *Pomacea canaliculata*, with a critical appraisal of imposex evaluation in the Ampullariidae. Ecotoxicol. Environ. Saf. 169, 573–582.
- Giulianelli, S., Primost, M.A., Lanari, C., Bigatti, G., 2020. RXR expression in marine gastropods with different sensitivity to imposex development. Sci Rep-Uk 10, 8.
- Gooding, M., Gallardo, C., Leblanc, G., 1999. Imposex in three marine gastropod species in Chile and potential impact on muriculture. Mar. Pollut. Bull. 38, 1227–1231.
- Gooding, M.P., Wilson, V.S., Folmar, L.C., Marcovich, D.T., LeBlanc, G.A., 2003. The biocide tributyltin reduces the accumulation of testosterone as fatty acid esters in the mud snail (*Ilyanassa obsoleta*). Environ. Health Perspect. 111, 426–430.
- Grilo, T.F., Rosa, R., 2017. Intersexuality in aquatic invertebrates: prevalence and causes. Sci. Total Environ. 592, 714–728.
- Grin, F., Blumberg, B., 2006. Environmental obesogens: organotins and endocrine disruption via nuclear receptor signaling. Endocrinology 147, S50–S55.
- Grin, F., Watanabe, H., Zamanian, Z., Maeda, L., Arima, K., Chubacha, R., Gardiner, D. M., Kanno, J., Iguchi, T., Blumberg, B., 2006. Endocrine-disrupting organotin compounds are potent inducers of adipogenesis in vertebrates. Mol. Endocrinol. 20, 2141–2155.
- Hagger, J.A., Depledge, M.H., Oehlmann, J., Jobling, S., Galloway, T.S., 2006. Is there a causal association between genotoxicity and the imposex effect? Environ. Health Perspect. 114, 20–26.
- Haveland, F., Hestholm, G., 2008. Risikovurdering av forurensa grunn og sediment. Fyllingen slipp AS. Resipientanalyse, p. 43.
- Haveland, F., Dyp, J., Dronnen, N.K., 2008. Risikovurdering av forurensa grunn og sediment. Vegsund slipp AS. Resipientanalyse, p. 58.
- Heindel, J.J., Blumberg, B., 2019. Environmental obesogens: mechanisms and controversies. In: Insel, P.A. (Ed.), Annual Review of Pharmacology and Toxicology, vol. 59. Annual Reviews, Palo Alto, pp. 89–106.
- Higuera-Ruiz, R., Elorza, J., 2011. Shell thickening and chambering in the oyster *Crassostrea gigas*: natural and anthropogenic influence of tributyltin contamination. Environ. Technol. 32, 583–591.
- Ho, K.K.Y., Leung, K.M.Y., 2016. Imposex status associated with organotin contamination in *Reishia clavigera* after reciprocal transplantation between clean and polluted sites in Hong Kong. Region. Stud. Marine Sci. 8, 480–486.
- Hoch, M., 2001. Organotin compounds in the environment - an overview. Appl. Geochem. 16, 719–743.
- Horie, Y., Yamagishi, T., Shintaku, Y., Iguchi, T., Tatarazako, N., 2018. Effects of tributyltin on early life-stage, reproduction, and gonadal sex differentiation in Japanese medaka (*Oryzias latipes*). Chemosphere 203, 418–425.
- Horiguchi, T., 2009. The endocrine-disrupting effect of organotin compounds for aquatic organisms. In: Takaomi, A., Harino, H., Ohji, M., Langston, W.J. (Eds.), Ecotoxicology of Antifouling Biocides, pp. 125–146.
- Horiguchi, T., Shiraishi, H., Shimizu, M., Morita, M., 1994. Imposex and organotin compounds in *Thais-clavigera* and *T-bronni* in Japan. J. Mar. Biol. Assoc. U. K. 74, 651–669.
- Horiguchi, T., Shiraishi, H., Shimizu, M., Morita, M., 1997. Imposex in sea snails, caused by organotin (tributyltin and triphenyltin) pollution in Japan: a survey. Appl. Organomet. Chem. 11, 451–455.
- Horiguchi, T., Ohta, Y., Nishikawa, T., Shiraishi, F., Shiraishi, H., Morita, M., 2008. Exposure to 9-cis retinoic acid induces penis and vas deferens development in the female rock shell. *Thais Clavigera*. Cell Biol. Toxicol. 24, 553–562.
- Horiguchi, T., Nishikawa, T., Ohta, Y., Shiraishi, H., Morita, M., 2010a. Time course of expression of the retinoid X receptor gene and induction of imposex in the rock shell, *Thais clavigera*, exposed to triphenyltin chloride. Anal. Bioanal. Chem. 396, 597–607.
- Horiguchi, T., Urushitani, H., Ohta, Y., Iguchi, T., Shiraishi, H., 2010b. Establishment of a polyclonal antibody against the retinoid X receptor of the rock shell *Thais clavigera* and its application to rock shell tissues for imposex research. Ecotoxicology 19, 571–576.
- Huang, W., Wu, Q., Xu, F., Li, L., Li, L., Que, H., Zhang, G., 2020. Functional characterization of retinoid X receptor with an emphasis on the mediation of organotin poisoning in the Pacific oyster (*Crassostrea gigas*). Gene 753, 144780.
- Iguchi, T., Katsu, Y., Horiguchi, T., Watanabe, H., Blumberg, B., Ohta, Y., 2007. Endocrine disrupting organotin compounds are potent inducers of imposex in gastropods and adipogenesis in vertebrates. Mole. Cell. Toxicol. 3, 1–10.
- IPCS, 1990. Environmental Health Criteria for Tributyltin Compounds, Environmental Health Criteria No. 116. World Health Organization, Geneva, p. 277.
- Kanayama, T., Kobayashi, N., Mamiya, S., Nakanishi, T., Nishikawa, J., 2005. Organotin compounds promote adipocyte differentiation as agonists of the peroxisome proliferator-activated receptor gamma/retinoid x receptor pathway. Mol. Pharmacol. 67, 766–774.
- Kaurin, M., Aakre, E., Jahren, T., Helland, A., 2015. Sedimenter i Bodø havn: Risikovurdering og tiltaksplan. Rambøll, Oslo, Norway, p. 53 (+ appendix).
- Keller, R., 1992. Crustacean neuropeptides - structures, functions and comparative aspects. Experientia 48, 439–448.
- Kim, N.S., Hong, S.H., Shin, K.H., Shim, W.J., 2017. Imposex in *Reishia clavigera* as an indicator to assess recovery of TBT pollution after a total ban in South Korea. Arch. Environ. Contam. Toxicol. 73, 301–309.
- Ko, M.M.C., Bradley, G.C., Neller, A.H., Broom, M.J., 1995. Tributyltin contamination of marine sediments of Hong Kong. Mar. Pollut. Bull. 31, 249–253.
- Kotake, Y., 2012. Molecular mechanisms of environmental organotin toxicity in mammals. Biol. Pharm. Bull. 35, 1876–1880.
- Kusk, K.O., Petersen, S., 1997. Acute and chronic toxicity of tributyltin and linear alkylbenzenesulfonate to the marine copepod *Acartia tonsa*. Environ. Toxicol. Chem. 16, 1629–1633.
- LaFont, R., 2000. Endocrinol. Invertebr. Ecotoxicol. 9, 41–57.
- Lagadic, L., Katsiadaki, I., Biever, R., Guiney, P.D., Karouna-Renier, N., Schwarz, T., Meador, J.P., 2018. Tributyltin: advancing the science on assessing endocrine disruption with an unconventional endocrine-disrupting compound. Rev. Environ. Contam. Toxicol. 65–127.
- LaLone, C.A., Villeneuve, D.L., Lyons, D., Helgen, H.W., Robinson, S.L., Swintek, J.A., Saari, T.W., Ankley, G.T., 2016. Sequence alignment to Predict across species susceptibility (SeqAPASS): a web-based tool for addressing the challenges of cross-species extrapolation of chemical toxicity. Toxicol. Sci. 153, 228–245.
- Lam, N.H., Jeong, H.H., Kang, S.D., Kim, D.J., Ju, M.J., Horiguchi, T., Cho, H.S., 2017. Organotins and new antifouling biocides in water and sediments from three Korean Special Management Sea Areas following ten years of tributyltin regulation: contamination profiles and risk assessment. Mar. Pollut. Bull. 121, 302–312.
- Langston, W.J., 2020. Endocrine disruption and altered sexual development in aquatic organisms: an invertebrate perspective. J. Mar. Biol. Assoc. U. K. 100, 495–515.
- Langston, W.J., Burt, G.R., 2007. A review of TBT sediment data in the Fal and Helford SAC. In: Marine Biological Association. The Environment Agency, p. 18.
- Langston, W.J., Pope, N.D., 1995. Determinants of TBT adsorption and desorption in estuarine sediments. Mar. Pollut. Bull. 31, 32–43.
- Langston, W.J., Pope, N.D., Davey, M., Langston, K.M., Hara, S.C.M., Gibbs, P.E., Pascoe, P.L., 2015. Recovery from TBT pollution in English Channel environments: a problem solved? Mar. Pollut. Bull. 95, 551–564.
- Laranjeiro, F., Sanchez-Marin, P., Oliveira, I.B., Galante-Oliveira, S., Barroso, C., 2018. Fifteen years of imposex and tributyltin pollution monitoring along the Portuguese coast. Environ. Pollut. 232, 411–421.
- Laughlin, R.B., 1996. Bioaccumulation of TBT by aquatic organisms. In: Champ, M.A., e, S.P.F. (Eds.), Organotin. Springer, Dordrecht.
- Le Maire, A., Grimaldi, M., Roecklin, D., Dagnino, S., Vivat-Hannah, V., Balaguer, P., Bourguet, W., 2009. Activation of RXR-PPAR heterodimers by organotin environmental endocrine disruptors. EMBO Rep. 10, 367–373.
- LeBlanc, G.A., Campbell, P.M., den Besten, P., Brown, R.P., Chang, E.S., Coats, J.R., deFur, P.L., Dhadialla, T., Edwards, J., Riddiford, L.M., Simpson, M.G., Snell, T.W., Thorndyke, M., Matsunura, F., 1999. The endocrinology of invertebrates. In: al, P.L. d.e (Ed.), Endocrine Disruption in Invertebrates: Endocrinology, Testing and Assessment. SETAC, pp. 23–106.
- LeBlanc, G.A., Gooding, M.P., Sternberg, R.M., 2005. Testosterone-fatty acid esterification: a unique target for the endocrine toxicity of tributyltin to gastropods. Integr. Comp. Biol. 45, 81–87.
- Lee, R.F., 1991. Metabolism of tributyltin by marine animals and possible linkages to effects. Mar. Environ. Res. 32, 29–35.
- Lesoway, M.P., Henry, J.Q., 2021. Retinoids promote penis development in sequentially hermaphroditic snails. Dev. Biol. 478, 122–132.

- Leung, K.M.Y., Grist, E.P.M., Morley, N.J., Morritt, D., Crane, M., 2007. Chronic toxicity of tributyltin to development and reproduction of the European freshwater snail *Lymnaea stagnalis* (L.). *Chemosphere* 66, 1358–1366.
- Li, P., Li, Z.H., 2020a. Environmental co-exposure to TBT and Cd caused neurotoxicity and thyroid endocrine disruption in zebrafish, a three-generation study in a simulated environment. *Environ. Pollut.* 259.
- Li, P., Li, Z.H., 2020b. Toxicity evaluation of triphenyltin in zebrafish larvae by embryonic malformation, retinal development, and GH/IGF axis. *Fish Physiol. Biochem.* 46, 2101–2107.
- Li, Z.H., Li, P., 2021. Effects of the tributyltin on the blood parameters, immune responses and thyroid hormone system in zebrafish. *Environ. Pollut.* 268.
- Li, X., Ycaza, J., Blumberg, B., 2011. The environmental obesogen tributyltin chloride acts via peroxisome proliferator activated receptor gamma to induce adipogenesis in murine 3T3-L1 preadipocytes. *J. Steroid Biochem. Mol. Biol.* 127, 9–15.
- Lima, D., Reis-Henriques, M.A., Silva, R., Santos, A.I., Castro, L.F.C., Santos, M.M., 2011. Tributyltin-induced imposex in marine gastropods involves tissue-specific modulation of the retinoid X receptor. *Aquat. Toxicol.* 101, 221–227.
- Linden, O., 1987. The Scope of the Organotin Issue in Scandinavia, Organotin Symposium, Oceans '87 Conference. The Institute of Electrical and Electronics Engineers, New York, USA, Halifax, Nova Scotia, Canada, pp. 320–1323.
- Lopez, M., Lilao, A.L., Ribalta, C., Martinez, Y., Pina, N., Ballesteros, A., Fito, C., Koehler, K., Newton, A., Monfort, E., Viana, M., 2022. Particle release from refit operations in shipyards: exposure, toxicity and environmental implications. *Sci. Total Environ.* 804, 12.
- Louiz, I., Ben-Attia, M., Ben-Hassine, O.K., 2009. Gonadosomatic index and gonad histopathology of *Gobius niger* (Gobiidae, Teleost) from Bizerta lagoon (Tunisia): evidence of reproduction disturbance. *Fish. Res.* 100, 266–273.
- Louiz, I., Ben-Attia, M., Ben Hassine, O.K., 2018. Reproductive disorders in *Zosterisessor ophiocephalus* (pisces, Gobiidae) in a polluted lagoon (bizerta, Tunisia). *Rev. Ecol. Terre Vie* 73, 227–241.
- Lv, H., Wang, J.J., Wang, M.Y., Shen, L., Xiao, L., Chen, T.J., Sun, T.Z., Li, W.J., Zhu, L.L., Zhang, X.K., 2021. Potent inhibition of tributyltin (TBT) and triphenyltin (TPT) against multiple UDP-glucuronosyltransferases (UGT): a new potential mechanism underlying endocrine disrupting actions. *Food Chem. Toxicol.* 149, 9.
- Lyssimachou, A., Santos, J.G., Andre, A., Soares, J., Lima, D., Guimaraes, L., Almeida, C. M.R., Teixeira, C., Castro, L.F.C., Santos, M.M., 2015. The mammalian "obesogen" tributyltin targets hepatic triglyceride accumulation and the transcriptional regulation of lipid metabolism in the liver and brain of zebrafish. *PLoS One* 10.
- Mansueto, V., Cangialosi, M.V., Faqi, A.S., 2011. Post-embryonic development effect of Bisphenol A and Tributyltin effects in *Ciona intestinalis*. *Caryologia* 64, 478–484.
- Marshall, D.J., Rajkumar, A., 2003. Imposex in the indigenous *Nassarius kraussianus* (Mollusca : neogastropoda) from South African harbours. *Mar. Pollut. Bull.* 46, 1150–1155.
- Martinovic, R., Kolarevic, S., Kracun-Kolarevic, M., Kostic, J., Jokanovic, S., Gacic, Z., Joksimovic, D., Durovic, M., Kljajic, Z., Vukovic-Gacic, B., 2016. Comparative assessment of cardiac activity and DNA damage in haemocytes of the Mediterranean mussel *Mytilus galloprovincialis* in exposure to tributyltin chloride. *Environ. Toxicol. Pharmacol.* 47, 165–174.
- Matthiessen, P., Gibbs, P.E., 1998. Critical appraisal of the evidence for tributyltin-mediated endocrine disruption in mollusks. *Environ. Toxicol. Chem.* 17, 37–43.
- Mattos, Y., Stotz, W.B., Romero, M.S., Bravo, M., Fillmann, G., Castro, I.B., 2017. Butyltin contamination in Northern Chilean coast: is there a potential risk for consumers? *Sci. Total Environ.* 595, 209–217.
- McAllister, B.G., Kime, D.E., 2003. Early life exposure to environmental levels of the aromatase inhibitor tributyltin causes masculinisation and irreversible sperm damage in zebrafish (*Danio rerio*). *Aquat. Toxicol.* 65, 309–316.
- McClellan-Green, P., Romano, J., Oberdorster, E., 2007. Does gender really matter in contaminant exposure? A case study using invertebrate models. *Environ. Res.* 104, 183–191.
- Meador, J.P., 2000. Predicting the fate and effects of tributyltin in marine systems. In: Ware, G.W. (Ed.), *Reviews of Environmental Contamination and Toxicology*, vol. 166. Springer, New York, pp. 1–48.
- Miki, S., Ikeda, K., Oba, Y., Satone, H., Honda, M., Shimasaki, Y., Onikura, N., Arakawa, O., Oshima, Y., 2011. Tributyltin in blood of marine fish collected from a coastal area of northern Kyushu, Japan. *Mar. Pollut. Bull.* 62, 2533–2536.
- Min, B.H., Kim, B.M., Kim, M., Kang, J.H., Jung, J.H., Rhee, J.S., 2018. Plasma biomarkers in juvenile marine fish provide evidence for endocrine modulation potential of organotin compounds. *Comp. Biochem. Physiol. C-Toxicol. Pharmacol.* 210, 35–43.
- Mizuta, T., Kubokawa, K., 2007. Presence of sex steroids and cytochrome P450 genes in amphioxus. *Endocrinology* 148, 3554–3565.
- Molvær, J., Knutzen, J., Magnusson, J., Rygg, B., Skei, J., Sørensen, J., 1997. Klassifisering av miljøkvalitet i fjorder og kystfarvann - Veileder 97:03 - 97:03 (TA-1467/1997). SFT, Oslo, Norway, p. 34.
- Moreira, L.B., Castro, I.B., Fillmann, G., Peres, T.F., Belmino, I.K.C., Sasaki, S.T., Taniguchi, S., Bicego, M.C., Marins, R.V., de Lacerda, L.D., Costa-Lotufo, L.V., Abessa, D.M.D., 2021. Dredging impacts on the toxicity and development of sediment quality values in a semi-arid region (Ceará state, NE Brazil). *Environ. Res.* 193, 11.
- Muller-Karanassos, C., Turner, A., Arundel, W., Vance, T., Lindeque, P.K., Cole, M., 2019. Antifouling paint particles in intertidal estuarine sediments from southwest England and their ingestion by the harbour ragworm, *Hediste diversicolor*. *Environ. Pollut.* 249, 163–170.
- Muller-Karanassos, C., Arundel, W., Lindeque, P.K., Vance, T., Turner, A., Cole, M., 2021. Environmental concentrations of antifouling paint particles are toxic to sediment-dwelling invertebrates. *Environ. Pollut.* 268, 11.
- Nakanishi, T., 2007. Potential toxicity of organotin compounds via nuclear receptor signaling in mammals. *J. Health Sci.* 53, 1–9.
- Nakanishi, T., 2008. Endocrine disruption induced by organotin compounds; organotins function as a powerful agonist for nuclear receptors rather than an aromatase inhibitor. *J. Toxicol. Sci.* 33, 269–276.
- Nath, M., 2008. Toxicity and the cardiovascular activity of organotin compounds: a review. *Appl. Organomet. Chem.* 22, 598–612.
- Nicolaus, E.E.M., Barry, J., 2015. Imposex in the dogwhelk (*Nucella lapillus*): 22-year monitoring around England and Wales. *Environ. Monit. Assess.* 187, 14.
- Nishikawa, J., 2006. Imposex in marine gastropods may be caused by binding of organotins to retinoid X receptor. *Mar. Biol.* 149, 117–124.
- Nishikawa, J., Mamiya, S., Kanayama, T., Nishikawa, T., Shiraishi, F., Horiguchi, T., 2004. Involvement of the retinoid X receptor in the development of imposex caused by organotins in gastropods. *Environ. Sci. Technol.* 38, 6271–6276.
- Norwegian Environment Agency, 2018. Guidelines for Risk Assessment of Contaminated Sediments. The Norwegian Environment Agency (NEA)/Norwegian Geotechnical Institute (NGI)/Norwegian Institute for Water Research (NIVA), Oslo, Norway, p. 115.
- Oberdörster, E., McClellan-Green, P., 2000. The neuropeptide APGWamide induces imposex in the mud snail, *Ilyanassa obsoleta*. *Peptides* 21, 1323–1330.
- Oberdörster, E., McClellan-Green, P., 2002. Mechanisms of imposex induction in the mud snail, *Ilyanassa obsoleta*: TBT as a neurotoxin and aromatase inhibitor. *Mar. Environ. Res.* 54, 715–718.
- Oberdörster, E., McClellan-Green, P., 2003. Expression of the peptide hormone APGWamide in imposex and normal snails: a neurotoxicity mechanism for imposex induction. *Integr. Comp. Biol.* 43, 896–896.
- Oberdörster, E., Romano, J., McClellan-Green, P., 2005. The neuropeptide APGWamide as a penis morphogenic factor (PMF) in gastropod mollusks. *Integr. Comp. Biol.* 45, 28–32.
- Oehlmann, J., Stroben, E., Fioroni, P., 1991. The morphological expression of imposex in *Nucella lapillus* (Linnaeus) (gastropoda, muricidae). *J. Molluscan Stud.* 57, 375–390.
- Oehlmann, J., Stroben, E., Schulte-Oehlmann, U., Bauer, B., Fioroni, P., Markert, B., 1996. Tributyltin biomonitoring using prosobranchs as sentinel organisms. *Fresen. J. Anal. Chem.* 354, 540–545.
- Oehlmann, J., Bauer, B., Minchin, D., Schulte-Oehlmann, U., Fioroni, P., Markert, B., 1998. Imposex in *Nucella lapillus* and intersex in *Littorina littorea*: interspecific comparison of two TBT-induced effects and their geographical uniformity. *Hydrobiologia* 378, 199–213.
- Oehlmann, J., Di Benedetto, P., Tillmann, M., Duft, M., Oetken, M., Schulte-Oehlmann, U., 2007. Endocrine disruption in prosobranch molluscs: evidence and ecological relevance. *Ecotoxicology* 16, 29–43.
- Ohtaki, K., Aihara, M., Takahashi, H., Fujita, H., Takahashi, K., Funabashi, T., Hirasawa, T., Ikezawa, Z., 2007. Effects of tributyltin on the emotional behavior of C57BL/6 mice and the development of atopic dermatitis-like lesions in DS-Nh mice. *J. Dermatol. Sci.* 47, 209–216.
- Olsen, M., Ranneklev, S., Selvik, J.R., Evensen, A., Pedersen, K.B., Håvardstun, J., Ørnnevad, S., Green, N., Tartu, V., 2021. Where along the Coast, Contaminated Seabed Today Poses the Greatest Risk to Health and the Environment: Knowledge Compilation, Assessment and Ranking of Areas. Norwegian Institute for Water Research, Oslo, Norway, p. 119.
- Omae, I., 2003. Organotin antifouling paints and their alternatives. *Appl. Organomet. Chem.* 17, 81–105.
- OSPAR, 2008. JAMP Guidelines for Contaminant-specific Biological Effects (OSPAR Agreement 2008-09) Monitoring Guidelines. OSPAR Commission, p. 48.
- OSPAR, 2009. CEMP Assessment Report: 2008/2009 - Assessment of Trends and Concentrations of Selected Hazardous Substances in Sediments and Biota, Monitoring and Assessment Series. OSPAR commission, p. 80.
- OSPARCOM, 2009. Assessment of the impacts of shipping on the marine environment. Monitoring and Assessment Series 34.
- Osugi, T., Sasakura, Y., Satake, H., 2020. The ventral peptidergic system of the adult ascidian *Ciona robusta* (*Ciona intestinalis* Type A) insights from a transgenic animal model. *Sci. Rep.-UK* 10.
- Page, D.S., Ozbal, C.C., Lanphear, M.E., 1996. Concentration of butyltin species in sediments associated with shipyard activity. *Environ. Pollut.* 91, 237–243.
- Park, K., Kim, R., Park, J.J., Shin, H.C., Lee, J.S., Cho, H.S., Lee, Y.G., Kim, J., Kwak, I.S., 2012. Ecotoxicological evaluation of tributyltin toxicity to the equilateral venus clam, *Gomphina veneriformis* (Bivalvia: veneridae). *Fish Shellfish Immunol.* 32, 426–433.
- Parmentier, K.F.V., Verhaegen, Y., De Witt, B.P., Hoffman, S., Delbare, D.H.R., Roose, P. M., Hylland, K.D.E., Burgeot, T., Smagghe, G.J., Cooreman, K., 2019. Tributyltin: a bottom-up regulator of the *Crangon crangon* population? *Front. Mar. Sci.* 6, 14.
- Pascoal, S., Carvalho, G., Vasieva, O., Hughes, R., Cossins, A., Fang, Y.X., Ashelford, K., Olohan, L., Barroso, C., Mendo, S., Creer, S., 2013. Transcriptomics and in vivo tests reveal novel mechanisms underlying endocrine disruption in an ecological sentinel, *Nucella lapillus*, Molecular Ecology 22, 1589–1608.
- Patricolo, E., Mansueto, C., D'Agati, P., Pellerito, L., 2001. Organometallic complexes with biological molecules: XVI. Endocrine disruption effects of tributyltin(IV) chloride on metamorphosis of the ascidian larva. *Appl. Organomet. Chem.* 15, 916–923.
- Paz-Villarraga, C.A., Castro, I.B., Fillmann, G., 2022. Biocides in antifouling paint formulations currently registered for use. *Environ. Sci. Pollut. Res.* 29, 30090–30101.
- Pougnet, F., Schafer, J., Dutruch, L., Garnier, C., Tessier, E., Dang, D.H., Lanceleur, L., Mullot, J.U., Lenoble, V., Blanc, G., 2014. Sources and historical record of tin and butyl-tin species in a Mediterranean bay (Toulon Bay, France). *Environ. Sci. Pollut. Res.* 21, 6640–6651.

- Rank, J., 2009. Intersex in *Littorina littorea* and DNA damage in *Mytilus edulis* as indicators of harbour pollution. *Ecotoxicol. Environ. Saf.* 72, 1271–1277.
- Rial, D., Bellas, J., Ruiz, J.M., 2018. Critical analysis of the relationship between imposex and butyltin body burden in *Nassarius reticulatus* and *Nucella lapillus*. *Environ. Pollut.* 237, 523–530.
- Ronis, M.J.J., Mason, A.Z., 1996. The metabolism of testosterone by the periwinkle (*Littorina littorea*) in vitro and in vivo: effects of tributyl tin. *Mar. Environ. Res.* 42, 161–166.
- Rossato, M., Castro, I.B., Paganini, C.L., Colares, E.P., Fillmann, G., Pinho, G.L.L., 2016. Sex steroid imbalances in the muricid *Stramonita haemastoma* from TBT contaminated sites. *Environ. Sci. Pollut. Res.* 23, 7861–7868.
- Ruiz, J.M., Carro, B., Albaina, N., Barreiro, R., Rial, D., Bellas, J., 2018. Extended imposex monitoring in N Atlantic Spain confirms punctual attainment of European environmental objectives for TBT. *Mar. Pollut. Bull.* 126, 462–466.
- Sahlén, S., Ågerstrand, M., 2020. Background Document on Tributyltin (TBT) in Sediment, Swedish Quality Standard, Hazardous Substances and Eutrophication Series. Stockholm University, Department of Environmental Science and Analytical Chemistry (ACES), London, UK, p. 40.
- Santillo, D., Johnston, P., Langston, W.J., 2008. 13. Tributyltin (TBT) antifoulants: a tale of ships, snails and imposex. In: Harremoës, P., Gee, D., MacGarvin, M., Stirling, A., Keys, J., Wynne, B., Vaz, S. (Eds.), *Late Lessons from Early Warnings: the Precautionary Principle 1896–2000*. European Environment Agency, pp. 135–148.
- Santos, M.M., Castro, L.F.C., Vieira, M.N., Micael, J., Morabito, R., Massanisso, P., Reis-Henriques, M.A., 2005. New insights into the mechanism of imposex induction in the dogwhelk *Nucella lapillus*. *Comparative Biochemistry and Physiology C-Toxicology & Pharmacology* 141, 101–109.
- Schøyen, M., Green, N.W., Hjermann, D.O., Tveiten, L., Beylich, B., Øxnevad, S., Beyer, J., 2019. Levels and trends of tributyltin (TBT) and imposex in dogwhelk (*Nucella lapillus*) along the Norwegian coastline from 1991 to 2017. *Mar. Environ. Res.* 144, 1–8.
- Schultz, M.P., 2007. Effects of coating roughness and biofouling on ship resistance and powering. *Biofouling* 23, 331–341.
- Scott, A.P., 2012. Do mollusks use vertebrate sex steroids as reproductive hormones? Part I: critical appraisal of the evidence for the presence, biosynthesis and uptake of steroids. *Steroids* 77, 1450–1468.
- Scott, A.P., 2013. Do mollusks use vertebrate sex steroids as reproductive hormones? II. Critical review of the evidence that steroids have biological effects. *Steroids* 78, 268–281.
- Seiguchi, T., Akitaya, H., Nakayama, S., Yazawa, T., Ogasawara, M., Suzuki, N., Hayakawa, K., Wada, S., 2020. Effect of polycyclic aromatic hydrocarbons on development of the ascidian *Ciona intestinalis* type A. *Int. J. Environ. Res. Publ. Health* 17.
- Shi, H.H., Huang, C.J., Zhu, S.X., Yu, X.J., Xie, W.Y., 2005. Generalized system of imposex and reproductive failure in female gastropods of coastal waters of mainland China. *Mar. Ecol. Prog. Ser.* 304, 179–189.
- Shim, W.H., Hong, S.H., Yim, U.H., Kim, N.S., Oh, J.R., 2002. Horizontal and vertical distribution of butyltin compounds in sediments from shipyards in Korea. *Arch. Environ. Contam. Toxicol.* 43, 277–283.
- Shimasaki, Y., Kitano, T., Oshima, Y., Inoue, S., Imada, N., Honjo, T., 2003. Tributyltin causes masculinization in fish. *Environ. Toxicol. Chem.* 22, 141–144.
- Shimizu, A., Kimura, S., 1992. Long-term effects of bis (normal-tributyltin) oxide (TBTO) on salt-water goby *Chasmichthys-dolichognathus*. *Nippon Suisan Gakkaishi* 58, 1595–1602.
- Smith, A.B., Jimenez, C.R., Dirks, R.W., Croll, R.P., Geraerts, W.P.M., 1992. Characterization of a cDNA clone encoding multiple copies of the neuropeptide APGWamide in the mollusk *Lymnaea stagnalis*. *J. Neurosci.* 12, 1709–1715.
- Smith, B.S., 1971. Sexuality in the American mud snail, *Nassarius obsoletus* say. *J. Molluscan Stud.* 39, 377–378.
- Smith, B.S., 1981. Male characteristics on female mud snails caused by antifouling bottom paints. *J. Appl. Toxicol.* 1, 22–25.
- Smith, P.J., 1996. Selective decline in imposex levels in the dogwhelk *Lepsiella scobina* following a ban on the use of TBT antifoulants in New Zealand. *Mar. Pollut. Bull.* 32, 362–365.
- Smith, A.J., Thain, J.E., Barry, J., 2006. Exploring the use of caged *Nucella lapillus* to monitor changes to TBT hotspot areas: a trial in the River Tyne estuary (UK). *Mar. Environ. Res.* 62, 149–163.
- Soon, Z.Y., Jung, J.H., Loh, A., Yoon, C., Shin, D., Kim, M., 2021. Seawater contamination associated with in-water cleaning of ship hulls and the potential risk to the marine environment. *Mar. Pollut. Bull.* 171, 11.
- Soroldoni, S., Castro, I.B., Abreu, F., Duarte, F.A., Choueri, R.B., Moller, O.O., Fillmann, G., Pinho, G.L.L., 2018. Antifouling paint particles: sources, occurrence, composition and dynamics. *Water Res.* 137, 47–56.
- Soroldoni, S., da Silva, S.V., Castro, I.B., Martins, C.D.G., Pinho, G.L.L., 2020. Antifouling paint particles cause toxicity to benthic organisms: effects on two species with different feeding modes. *Chemosphere* 238, 9.
- Soroldoni, S., Honscha, L.C., Reis, F.O., Duarte, F.A., da Silva, F.M.R., Pinho, G.L.L., 2021. Antifouling paint particles in soils: toxic impact that goes beyond the aquatic environment. *Ecotoxicology* 30, 1161–1169.
- Sousa, A., Genio, L., Mendo, S., Barroso, C., 2005a. Comparison of the acute toxicity of tributyltin and copper to veliger larvae off *Nassarius reticulatus* (L.). *Appl. Organomet. Chem.* 19, 324–328.
- Sousa, A., Mendo, S., Barroso, C., 2005b. Imposex and organotin contamination in *Nassarius reticulatus* (L.) along the Portuguese coast. *Appl. Organomet. Chem.* 19, 315–323.
- Sousa, A., Laranjeiro, F., Takahashi, S., Tanabe, S., Barroso, C.M., 2009. Imposex and organotin prevalence in a European post-legislative scenario: temporal trends from 2003 to 2008. *Chemosphere* 77, 566–573.
- Sousa, A.C.A., Pastorinho, M.R., Takahashi, S., Tanabe, S., 2014. History on organotin compounds, from snails to humans. *Environ. Chem. Lett.* 12, 117–137.
- Sparks, C., Awe, A., 2022. Concentrations and risk assessment of metals and microplastics from antifouling paint particles in the coastal sediment of a marina in Simon's Town, South Africa. *Environ. Sci. Pollut. Res.* 16.
- Stange, D., Sieratowicz, A., Oehlmann, J., 2012. Imposex development in *Nucella lapillus* - evidence for the involvement of retinoid X receptor and androgen signalling pathways *in vivo*. *Aquat. Toxicol.* 106, 20–24.
- Sternberg, R.M., Hotchkiss, A.K., LeBlanc, G.A., 2008. Synchronized expression of retinoid X receptor mRNA with reproductive tract recrudescence in an imposex-susceptible mollusc. *Environ. Sci. Technol.* 42, 1345–1351.
- Sternberg, R.M., Gooding, M.P., Hotchkiss, A.K., LeBlanc, G.A., 2010. Environmental-endocrine control of reproductive maturation in gastropods: implications for the mechanism of tributyltin-induced imposex in prosobranchs. *Ecotoxicology* 19, 4–23.
- Stroben, E., Oehlmann, J., Fioroni, P., 1992. *Hinia-reticulata* and *Nucella-lapillus* - comparison of two gastropod tributyltin bioindicators. *Mar. Biol.* 114, 289–296.
- Stronkhorst, J., 1996. TBT Contamination and toxicity of sediments: a persistent problem. In: Stewen, U. (Ed.), *The Present Status of TBT-Copolymer Antifouling Paints. Proceeding of International One Day Symposium on Antifouling Paints for Ocean-Going Vessels*. Ministry of Transport, Public Works and Water Management, Rijkswaterstaat, DGSM. ORTEP Association, The Hague, Netherlands.
- Svavarsson, J., 2000. Imposex in the dogwhelk (*Nucella lapillus*) due to TBT contamination: improvement at high latitudes. *Mar. Pollut. Bull.* 40, 893–897.
- Svavarsson, J., Granmo, A., Ekelund, R., Szpunar, J., 2001. Occurrence and effects of organotins on adult common whelk (*Buccinum undatum*) (Mollusca, Gastropoda) in harbours and in a simulated dredging situation. *Mar. Pollut. Bull.* 42, 370–376.
- Takeuchi, A., Takahashi, S., Tanabe, S., Miyazaki, N., 2001. Caprella watch: a new approach for monitoring butyltin residues in the ocean. *Mar. Environ. Res.* 52, 97–113.
- Tallmon, D.A., Hoferkamp, L., 2009. Long-term changes in imposex frequency in file dogwinkles, *Nucella lima* G., and tributyltin concentrations in bay mussels, *Mytilus trossulus* G. *Bull. Environ. Contam. Toxicol.* 83, 235–238.
- Tan, K.S., 1997. Imposex in three species of *Thais* from Singapore, with additional observations on *T-clavigera* (Kuster) from Japan. *Mar. Pollut. Bull.* 34, 577–581.
- Ten Hallers-Tjabbes, C.C., Kemp, J.F., Boon, J.P., 1994. Imposex in whelks (*Buccinum undatum*) from the open north-sea - relation to shipping traffic intensities. *Mar. Pollut. Bull.* 28, 311–313.
- Thibaut, R., Porte, C., 2004. Effects of endocrine disruptors on sex steroid synthesis and metabolism pathways in fish. *J. Steroid Biochem. Mol. Biol.* 92, 485–494.
- Titley-O'Neal, C.P., MacDonald, B.A., Pelletier, E., Saint-Louis, R., 2011a. Using *Nucella lapillus* (L.) as a bioindicator of tributyltin (TBT) pollution in eastern Canada: a historical perspective. *Water Qual. Res. J. Can.* 46, 74–86.
- Titley-O'Neal, C.P., Munkittrick, K.R., MacDonald, B.A., 2011b. The effects of organotin on female gastropods. *J. Environ. Monit.* 13, 2360–2388.
- Torres, F.G., De-la-Torre, G.E., 2021. Environmental pollution with antifouling paint particles: distribution, ecotoxicology, and sustainable alternatives. *Mar. Pollut. Bull.* 169, 10.
- Toste, R., Fernandez, M.A., Pessoa, I.D., Parahyba, M.A., Dore, M.P., 2011. Organotin pollution at arraial do cabo, rio De Janeiro state, Brazil: increasing levels after the TBT ban. *Braz. J. Oceanogr.* 59, 111–117.
- Schøyen, M., Beyer, J., Tveiten, L., Hjermann, D., Håvardstun, J., Berge, J.A., Øxnevad, S., 2022. Levels and Trends of Tributyltin (TBT) and Intersex in Common Periwinkle (*Littorina littorea*) in the Fjord Vikkilen, Norway, from 2005 to 2021. In prep.
- Turner, A., 2010. Marine pollution from antifouling paint particles, *Mar. Pollut. Bull.* 60, 159–171.
- Turner, A., 2021. Paint particles in the marine environment: an overlooked component of microplastics. *Water Res.* X 12, 11.
- Turner, A., Glegg, G., 2014. TBT-based antifouling paints remain on sale. *Mar. Pollut. Bull.* 88, 398–400.
- Uc-Peraza, R.G., Castro, I.B., Fillmann, G., 2022a. An absurd scenario in 2021: banned TBT-based antifouling products still available on the market. *Sci. Total Environ.* 805, 8.
- Uc-Peraza, R.G., Delgado-Blas, V.H., Rendon-von Osten, J., Castro, I.B., Proietti, M.C., Fillmann, G., 2022b. Mexican paradise under threat: the impact of antifouling biocides along the Yucatán acute accent an Peninsula. *J. Hazard Mater.* 427, 13.
- Urushitani, H., Katsu, Y., Ohta, Y., Shiraiishi, H., Iguchi, T., Horiguchi, T., 2011. Cloning and characterization of retinoid X receptor (RXR) isoforms in the rock shell. *Thais clavigera*, *Aquatic Toxicology* 103, 101–111.
- Urushitani, H., Katsu, Y., Kagechika, H., Sousa, A.C.A., Barroso, C.M., Ohta, Y., Shiraiishi, H., Iguchi, T., Horiguchi, T., 2018. Characterization and comparison of transcriptional activities of the retinoid X receptors by various organotin compounds in three prosobranch gastropods; *Thais clavigera*, *Nucella lapillus* and *Babylonia japonica*. *Aquat. Toxicol.* 199, 103–115.
- van der Kerk, G.J.M., Luitjen, J.G.A., 1954. The biocidal properties of organotin compounds. *J. Appl. Chem.* 4, 314–319.
- Vasconcelos, P., Gaspar, M.B., Barroso, C.M., 2010. Imposex in *Bolinus brandaris* from the Ria formosa lagoon (southern Portugal): usefulness of "single-site baselines" for environmental monitoring. *J. Environ. Monit.* 12, 1823–1832.
- Verhaegen, Y., Parmentier, K., Swevers, L., Renders, E., Rouge, P., De Coen, W., Cooreman, K., Smagghe, G., 2011. The heterodimeric ecdysteroid receptor complex in the brown shrimp *Crangon crangon*: EcR and RXR isoform characteristics and sensitivity towards the marine pollutant tributyltin. *Gen. Comp. Endocrinol.* 172, 158–169.

- Verhaegen, Y., Monteyne, E., Neudecker, T., Tulp, I., Smaghe, G., Cooreman, K., Roose, P., Parmentier, K., 2012. Organotins in North Sea brown shrimp (*Crangon crangon* L.) after implementation of the TBT ban. Chemosphere 86, 979–984.
- Verslycke, T.A., Vethaak, A.D., Aris, K., Janssen, C.R., 2005. Flame retardants, surfactants and organotins in sediment and mysid shrimp of the Scheldt estuary (The Netherlands). Environ. Pollut. 136, 19–31.
- VKM, 2007. Risikovurdering av organiske tinnforbindelser i sjømat. Vitenskapskomiteen for mattrygghet (VKM), Oslo, Norway, p. 13.
- Vogt, E.L., Model, J.F.A., Vinagre, A.S., 2018. Effect of organotins on Crustaceans: update and perspective. Front. Endocrinol. 9, 8.
- Vos, J.G., Dybing, E., Greim, H.A., Ladefoged, O., Lambre, C., Tarazona, J.V., Brandt, I., Vethaak, A.D., 2000. Health effects of endocrine-disrupting chemicals on wildlife, with special reference to the European situation. Crit. Rev. Toxicol. 30, 71–133.
- Waldock, M.J., Thain, J.E., 1983. Shell thickening in *Crassostrea gigas* - organotin anti-fouling or sediment induced. Mar. Pollut. Bull. 14, 411–415.
- Waldock, R., Rees, H.L., Matthiessen, P., Pendle, M.A., 1999. Surveys of the benthic fauna of the Crouch Estuary (UK) in relation to TBT contamination. J. Mar. Biol. Assoc. U. K. 79, 225–232.
- Weidenhaupt, A., Arnold, C., Muller, S.R., Haderlein, S.B., Schwarzenbach, R.P., 1997. Sorption of organotin biocides to mineral surfaces. Environ. Sci. Technol. 31, 2603–2609.
- Wu, L.Y., Chen, H.G., Ru, H.J., Li, Y.F., Yao, F., Ni, Z.H., Zhong, L.Q., 2020. Sex-specific effects of triphenyltin chloride (TPT) on thyroid disruption and metabolizing enzymes in adult zebrafish (*Danio rerio*). Toxicol. Lett. 331, 143–151.
- Yonezawa, T., Hasegawa, S.I., Ahn, J.Y., Cha, B.Y., Teruya, T., Hagiwara, H., Nagai, K., Woo, J.T., 2007. Tributyltin and triphenyltin inhibit osteoclast differentiation through a retinoic acid receptor-dependent signaling pathway. Biochem. Biophys. Res. Commun. 355, 10–15.
- Ytreberg, E., Bighiu, M.A., Lundgren, L., Eklund, B., 2016. XRF measurements of tin, copper and zinc in antifouling paints coated on leisure boats. Environ. Pollut. 213, 594–599.
- Zhang, J.L., Zuo, Z.H., Chen, Y.X., Zhao, Y., Hu, S., Wang, C.G., 2007. Effect of tributyltin on the development of ovary in female cuvier (*Sebastiscus marmoratus*). Aquat. Toxicol. 83, 174–179.
- Zhang, J.L., Zuo, Z.H., Chen, R., Chen, Y.X., Wang, C.G., 2008. Tributyltin exposure causes brain damage in *Sebastiscus marmoratus*. Chemosphere 73, 337–343.
- Zhang, J.L., Zuo, Z.H., He, C.Y., Cai, J.L., Wang, Y.Q., Chen, Y.X., Wang, C.G., 2009. Effect of tributyltin on testicular development in *Sebastiscus marmoratus* and the mechanism involved. Environ. Toxicol. Chem. 28, 1528–1535.
- Zhang, J.L., Zuo, Z.H., Wang, Y.Q., Yu, A., Chen, Y.X., Wang, C.G., 2011. Tributyltin chloride results in dorsal curvature in embryo development of *Sebastiscus marmoratus* via apoptosis pathway. Chemosphere 82, 437–442.
- Zhang, J.L., Zuo, Z.H., Xiong, J.L., Sun, P., Chen, Y.X., Wang, C.G., 2013. Tributyltin exposure causes lipotoxicity responses in the ovaries of rockfish, *Sebastiscus marmoratus*. Chemosphere 90, 1294–1299.
- Zhang, J.L., Sun, P., Kong, T., Yang, F., Guan, W.C., 2016. Tributyltin promoted hepatic steatosis in zebrafish (*Danio rerio*) and the molecular pathogenesis involved. Aquat. Toxicol. 170, 208–215.
- Zheng, R.H., Wang, C.G., Zhao, Y., Zu, Z.H., Chen, Y.X., 2005. Effect of tributyltin, benzo(a)pyrene and their mixture exposure on the sex hormone levels in gonads of cuvier (*Sebastiscus marmoratus*). Environ. Toxicol. Pharmacol. 20, 361–367.
- Zuo, Z.H., Cai, J.L., Wang, X.L., Li, B.W., Wang, C.G., Chen, Y.X., 2009. Acute administration of tributyltin and trimethyltin modulate glutamate and N-methyl-D-aspartate receptor signaling pathway in *Sebastiscus marmoratus*. Aquat. Toxicol. 92, 44–49.
- Zuo, Z.H., Wang, C.G., Wu, M.F., Wang, Y.Q., Chen, Y.X., 2012. Exposure to tributyltin and triphenyltin induces DNA damage and alters nucleotide excision repair gene transcription in *Sebastiscus marmoratus* liver. Aquat. Toxicol. 122, 106–112.