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Current status and advances of fish vaccines in Malaysia

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Abstract

Fish diseases have a significant negative influence on the Malaysian aquaculture industry. Since the 1980s, the sector has grown in size, which has resulted in a rise in the prevalence of infectious outbreaks affecting both freshwater and marine cultured fish species. Demand for commercially available fish vaccinations is predicted to increase as infectious disease outbreaks continue to occur. In Malaysia, aquaculture vaccine research and development (R&D) are still in its infancy, with most efforts concentrating on producing vaccines against bacterial infections, most notably streptococcosis, vibriosis, and motile *Aeromonas* septicemia. Despite several attempts, no homegrown vaccine has been effectively introduced into the manufacturing pipeline to date. At the moment, only three imported aquatic vaccines have received full permission, a far cry from the 314 and 60 vaccines licensed in the poultry and porcine industries, respectively. This review will describe recent findings regarding the development of aquaculture vaccines for certain fish species and diseases in Malaysia. In our opinion, R&D on fish vaccines is critical to the aquaculture industry's viability. © 2022 *Veterinary World*. All rights reserved.

Author Keywords

Aquaculture; Fish; Fish diseases; Malaysia; Vaccine

Index Keywords

immunoglobulin A, recombinant vaccine, triacylglycerol, vaccine, virulence factor; *Aeromonas*, *Aeromonas salmonicida*, antibiotic resistance, Article, autophagosome, bacterial infection, bacteriophage, bacterium culture, *Bifidobacterium bifidum*, biofilm, epidemic, *Epinephelus*, *exophthalmos*, fish, fish disease, freshwater fish, gene deletion, herd immunity, humoral immunity, lamina propria, leishmaniasis, marine environment, *Oreochromis niloticus*, phagocytosis, prevalence, RNA interference, *Salmo salar*, septicemia, upregulation, vaccination, *Vibrio harveyi*, *Vibrio parahaemolyticus*, *Vibrio vulnificus*, vibriosis

References

- (2020) *Annual Fisheries Statistic 2020. Aquaculture Table*, 1. Malaysia: Department of Fisheries Malaysia. Retrieved on 12-09-2021
- Wan Norhana, M.N., Gerald, M.J., Rozana, J. **Aquaculture component of a national action plan on antimicrobial resistance in Malaysia** (2020) *Asian Fish. Sci*, 33S, pp. 90-96. (S1)
- (2019) *Annual Fisheries Statistic 2019*, 1. Department of Fisheries Malaysia, Malaysia. Retrieved on 12-09-2021
- (2016) *Annual Fisheries Statistic 2016-2019*, 1. (-2018) Department of Fisheries Malaysia, Malaysia. Retrieved on 12-09-2021

- (2020) *The State of World Fisheries and Aquaculture 2020. Sustainability in Action*, Food and Agriculture Organization of the United Nations, Rome, Italy
- Tacon, A.G.J.
Trends in global aquaculture and aquafeed production: 2000-2017
(2020) *Rev. Fish. Sci. Aquat*, 28 (1), pp. 43-56.
- Sayuthi, S.
Fish diseases in Malaysia: Status and problems
(1993) *Proceedings of the Aquaculture Workshop for SEAFDEC/AQD Training Alumni*, p. p173.
8-11 Sept. 1992, Iloilo, Philippines. SEAFDEC Aquaculture Department, Iloilo, Philippines
- Siti Hawa, A., Mohd Syafiq, M.R., Siti-Zahrah, A., Nur- Nazifah, M., Firdaus-Nawi, M., Zamri-Saad, M., Amal, M.N.A.
Retrospective identification of bacterial depository revealed that *Streptococcus iniae* was responsible for some of the streptococcosis cases in cultured red tilapia in Malaysia since 2006
(2020) *Pertanika J. Trop. Agric. Sci*, 43 (2), pp. 231-238.
- Aisyhah, M.A.S., Amal, M.N.A., Zamri-Saad, M., Siti-Zahrah, A., Shaqinah, N.N.
***Streptococcus agalactiae* isolates from cultured fishes in Malaysia manifesting low resistance pattern towards selected antibiotics**
(2015) *J. Fish Dis*, 38 (12), pp. 1093-1098.
- Siti-Zahrah, A.
Common bacterial diseases of catfish, *Clarias macrocephelus*, cultured in Malacca
(1992) *Proceedings of the National Intensification of Research in Priority Areas (IRPA) Seminar*, II.
Ministry of Science, Technology, and Environment, Kuala Lumpur, Malaysia
- Abdullah, A., Ramly, R., Ridzwan, M.S.M., Sudirwan, F., Abas, A., Ahmad, K., Murni, M., Kua, B.C.
First detection of tilapia lake virus (TiLV) in wild river carp (*Barbonymus schwanenfeldii*) at Timah Tasoh Lake, Malaysia
(2018) *J. Fish Dis*, 41 (9), pp. 1459-1462.
- Amal, M.N.A., Koh, C.B., Nurliyana, M., Suhaiba, M., Nor-Amalina, Z., Santha, S., Diyana-Nadhirah, K.P., Zamri-Saad, M.
A case of natural co-infection of tilapia lake virus and *Aeromonas veronii* in a Malaysian red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) farm experiencing high mortality
(2018) *Aquaculture*, 485, pp. 12-16.
- Miccoli, A., Manni, M., Picchiatti, S., Scapigliati, G.
State-of-the-art vaccine research for aquaculture use: The case of three economically relevant fish species
(2021) *Vaccines*, 9 (2), p. 140.
- Standish, I.F., Brenden, T.O., Faisal, M.
Does herd immunity exist in aquatic animals?
(2016) *Int. J. Mol. Sci*, 17 (11), p. 1898.

- (2019) *Flow Chart of Veterinary Vaccine Registration*, Department of Veterinary Services Malaysia, Malaysia. Retrieved on 12-09-2021
- (2020) *List of Approved Veterinary Vaccines*, Department of Veterinary Services Malaysia, Malaysia. Retrieved on 12-09-2021
- Leong, T.S.
Control of parasites in cultured marine finfishes in Southeast Asia-an overview
(1997) *Int. J. Parasitol*, 27 (10), pp. 1177-1184.
- Chuah, T.T.
Survey of grouper diseases in Malaysia
(2001) *Report and proceeding of APEC FWG Project 02/2000 "Development of a Regional Research Programme on Grouper Virus Transmission and Vaccine Development"*, pp. p38-p40.
Asia Pacific Economic Cooperation (APEC), Aquatic Animal Health Research Institute (AAHRI), Fish Health Section of the Asian Fisheries Society (FHS/AFS), and the Network of Aquaculture Centres in Asia-Pacific (NACA), Bangkok, Thailand
- See-Yong, W., Ong, B., Thia-Eng, C.
Isolation, identification of causative agent of 'Red Boil Disease' in Grouper (*Epinephelus salmoides*) and its possible control by vaccination
(1979) *Proceeding of the International Workshop on Pen Cage Culture of Fish*, pp. p81-p87.
11-12 Feb. 1979, Tigbauan, Iloilo, Philippines. Aquaculture Department, Southeast Asian Fisheries Development Center, Tigbauan, Iloilo, Philippines
- Siti-Zahrah, A., Palanisamy, V., Chuah, T.T., Kua, B.C., Azila, A., Vijayenthly, N.
Experimental vaccine trials using Alpharma vaccine against vibriosis on *Lutjanus argentimaculatus* in laboratory condition
(2003) *FRI Newslett*, 8 (1-2), pp. 14-15.
- Siti-Zahrah, A., Misri, S., Padilah, B., Zulkafli, R., Kua, B.C., Azila, A., Rimatulhana, R.
Pre-disposing factors associated with the outbreak of streptococcal infection in floating cage-cultured tilapia in reservoirs
(2004) *7th Asian Fisheries Forum 04 Abstracts. Triennial Meeting of the Asian Fisheries Society*, p. p420.
Penang, Malaysia
- Siti-Zahrah, A., Padilah, B., Azila, A., Rimatulhana, R., Shahidan, H.
Multiple streptococcal species infection in cage-cultured red tilapia but showing similar clinical signs
(2008) *Diseases in Asian Aquaculture VI. Proceedings of the Sixth Symposium on Asian Aquaculture*, pp. p313-p320.
Bondad-Reantaso, M.G., Mohan, C.V., Crumlish, M. and Subasinghe, R.P., editors. Fish Health Section of the Asian Fisheries Society, Colombo, Sri Lanka
- Pei-Chih, L., Yi-Lun, T., Yao-Chung, C., Pei-Chi, W., Shu-Chu, L., Shih-Chu, C.
Analysis of streptococcal infection and correlation with climatic factors in cultured tilapia *Oreochromis spp.* in Taiwan
(2020) *Appl. Sci*, 10 (11), p. 4018.

- Mishra, A., Gyu-Hwi, N., Jeong-An, G., Hee-Eun, L., Jo, A., Heui-Soo, K.
Current challenges of Streptococcus infection and effective molecular, cellular, and environmental control methods in aquaculture
(2018) *Mol. Cell*, 41 (6), pp. 495-505.
- Zamri-Saad, M., Amal, M.N.A., Siti-Zahrah, A.
Pathological changes in red tilapias (*Oreochromis spp.*) naturally infected by *Streptococcus agalactiae*
(2010) *J. Comp. Pathol*, 143 (2-3), pp. 227-229.
- Laith, A.A., Ambak, M.A., Hassan, M., Sheriff, S.M., Nadirah, M., Draman, A.S., Wahab, W., Najiah, M.
Molecular identification and histopathological study of natural *Streptococcus agalactiae* infection in hybrid tilapia (*Oreochromis niloticus*)
(2017) *Vet. World*, 10 (1), pp. 101-111.
- Amal, M.N.A., Siti-Zahrah, A., Zulkafli, R., Misri, S., Ramley, A., Zamri-Saad, M.
The effect of water temperature on the incidence of *Streptococcus agalactiae* infection in cage-cultured tilapia
(2008) *Proceedings of the International Seminar on Management Strategies on Animal Health and Production Control in Anticipation of Global Warming*, pp. p48-p51.
Surabaya, Indonesia
- Najiah, M., Aqilah, N.I., Lee, K.L., Khairulbariyah, Z., Mithun, S., Jalal, K.C.A., Shaharom-Harrison, F., Nadirah, M.
Massive mortality associated with *Streptococcus agalactiae* infection in cage-cultured red hybrid tilapia *Oreochromis niloticus* in Como River, Kenyir Lake, Malaysia
(2012) *J. Biol. Sci*, 12 (8), pp. 438-442.
- Amal, M.N.A., Zamri-Saad, M., Zulkafli, A.R., Siti-Zahrah, A., Misri, S., Ramley, B., Shahidan, H., Sabri, M.Y.
Water thermocline confirms susceptibility of tilapia cultured in lakes to *Streptococcus agalactiae*
(2010) *J. Anim. Vet. Adv*, 9 (22), pp. 2811-2817.
- Rodkhum, C., Kayansamruaj, P., Pirarat, N.
Effect of water temperature on susceptibility to *Streptococcus agalactiae* serotype Ia infection in Nile tilapia (*Oreochromis niloticus*)
(2011) *Thai J. Vet. Med*, 41 (3), pp. 309-314.
- Zamri-Saad, M., Amal, M.N.A., Siti-Zahrah, A., Zulkafli, A.R.
Control and prevention of streptococcosis in cultured tilapia in Malaysia: A review
(2014) *Pertanika J. Trop. Agric. Sci*, 37 (4), pp. 389-410.
- Syuhada, R., Zamri-Saad, M., Ina-Salwany, M.Y., Mustafa, M., Nasruddin, N.N., Desa, M.N.M., Nordin, S.A., Amal, M.N.A.
Molecular characterisation and pathogenicity of *Streptococcus agalactiae* serotypes Ia ST7 and III ST283 isolated from cultured red hybrid tilapia in Malaysia
(2020) *Aquaculture*, 515, p. 734543.
- Suphia-Amiera, S.
(2019) *Molecular Characterisation of Streptococcus agalactiae Virulence Gene Isolated from Malaysian Hybrid Tilapia (*Oreochromis spp.*)*,

(Master's Thesis, International Islamic University Malaysia, Kuantan, Malaysia). Retrieved on 27-09-2021

- Amal, M.N.A., Zamri-Saad, M., Siti-Zahrah, A., Zulkafli, A.R., Nur-Nazifah, M.
Molecular characterisation of Streptococcus agalactiae strains isolated from fishes in Malaysia
(2013) *J. Appl. Microbiol*, 115 (1), pp. 20-29.
- Sun, J., Fang, W., Ke, B., He, D., Liang, Y., Ning, D., Tan, H., Deng, X.
Inapparent Streptococcus agalactiae infection in adult/commercial tilapia
(2016) *Sci. Rep*, 6, p. 26319.
- Leal, C.A.G., Queiroz, G.A., Pereira, F.L., Tavares, G.C., Figueiredo, H.C.P.
Streptococcus agalactiae sequence type 283 in farmed fish, Brazil
(2019) *Emerg. Infect. Dis*, 25 (4), pp. 776-779.
- Piamsomboon, P., Thanasaksiri, K., Murakami, A., Fukuda, K., Takano, R., Jantrakajorn, S., Wongtavatchai, J.
Streptococcosis in freshwater farmed seabass *Lates calcarifer* and its virulence in Nile tilapia *Oreochromis niloticus*
(2020) *Aquaculture*, 523, p. 735189.
- Al-Harbi, A.H.
Phenotypic and genotypic characterisation of Streptococcus agalactiae isolated from hybrid tilapia (*Oreochromis niloticus* × *O. aureus*)
(2016) *Aquaculture*, 464, pp. 515-520.
- Klesius, P., Shoemaker, C.A., Evans, J.
Streptococcus: A worldwide fish health problem
(2008) *Proceedings of the Eight International Symposium on Tilapia in Aquaculture*, p. p7. 12-14 Oct. 2008, Cairo, Egypt
- Amal, M.N.A., Zamri-Saad, M.
Streptococcosis in tilapia (*Oreochromis niloticus*): A review
(2011) *Pertanika J. Trop. Agric. Sci*, 34 (2), pp. 195-206.
- Firdaus-Nawi, M., Yusoff, S.M., Hanan, Y., Siti-Zahrah, A., Zamri-Saad, M.
Efficacy of feed-based adjuvant vaccine against Streptococcus agalactiae in Oreochromis spp. in Malaysia
(2013) *Aquac. Res*, 45 (1), pp. 87-96.
- Ismail, M.S., Siti-Zahrah, A., Syafiq, M.R.M., Amal, M.N.A., Firdaus-Nawi, M., Zamri-Saad, M.
Feed-based vaccination regime against streptococcosis in red tilapia, *Oreochromis niloticus* × *Oreochromis mossambicus*
(2016) *BMC Vet. Res*, 12 (1), p. 194.
- Ismail, M.S., Syafiq, M.R., Siti-Zahrah, A., Fahmi, S., Shahidan, H., Hanan, Y., Amal, M.N.A., Zamri-Saad, M.
The effect of feed-based vaccination on tilapia farm endemic for streptococcosis
(2016) *Fish Shellfish Immunol*, 60, pp. 21-24.
- Nur-Nazifah, M., Sabri, M.Y., Siti-Zahrah, A.
Development and efficacy of feed-based recombinant vaccine encoding the cell wall surface anchor family protein of Streptococcus agalactiae against streptococcosis

in *Oreochromis* sp

(2014) *Fish Shellfish Immunol*, 37 (1), pp. 193-200.

- Nadirah, A.N.
(2015) *Field Evaluation of Feed-Based Recombinant Protein Adjuvanted Vaccine Against Streptococcosis in Red Hybrid Tilapia (*Oreochromis* sp.)*,
(Master's Thesis, Universiti Putra Malaysia, Serdang, Malaysia). Retrieved on 27-09-2021
- Sa'aidatun Asyikin, A., Farina, M.K., Zamri-Saad, M., Siti-Zahrah, A., Mohd Syafiq, M.R., Hanan, M.Y., Shahidan, H., Suphia-Amiera, S.
Effect of incorporating different concentrations of palm oil as an adjuvant in the fish vaccine
(2018) *Int. J. Biosci*, 12 (1), pp. 35-41.
- Noraini, O., Sabri, M.Y., Siti-Zahrah, A.
Efficacy of spray administration of formalin-killed *Streptococcus agalactiae* in hybrid red tilapia
(2013) *J. Aquat. Anim. Health*, 25 (2), pp. 142-148.
- Kahieshesfandiari, M., Sabri, M.Y., Ina-salwany, M.Y., Hassan, M.D., Noraini, O., Ajadi, A.A., Isiaku, A.I.
Streptococcosis in *Oreochromis* sp.: Is feed-based biofilm vaccine of *Streptococcus agalactiae* effective?
(2019) *Aquac. Int*, 27 (3), pp. 817-832.
- Laith, A.A., Abdullah, M.A., Nurhafizah, W.W.I., Hussein, H.A., Aya, J., Effendy, A.W.M., Najiah, M.
Efficacy of live attenuated vaccine derived from the *Streptococcus agalactiae* on the immune responses of *Oreochromis niloticus*
(2019) *Fish Shellfish Immunol*, 90, pp. 235-243.
- Hayat, M., Sabri, M.Y., Intan-Shameha, A.R., Ina-Salwany, M.Y., Thompson, K.D.
Localisation of antigens in the gut post-challenge with *Streptococcus iniae* in vaccinated and non-vaccinated red hybrid tilapia (*Oreochromis* sp.)
(2020) *Aquac. Int*, 28 (4), pp. 1739-1752.
- Hayat, M., Sabri, M., Yusoff, M., Samad, M.J., Shameha, I., Razak, A., Ina Salwany, M.Y., Hasni, K.
Efficacy of feed-based formalin-killed vaccine of *Streptococcus iniae* stimulates the gut-associated lymphoid tissues and immune response of red hybrid tilapia
(2021) *Vaccines*, 9 (1), p. 51.
- Matusin, S.
(2015) *Molecular Characterisation of *Aeromonas hydrophila* and Development of Recombinant cells vaccine expressing outer membrane proteins Against its in African Catfish (*Clarias gariepinus* Burchell)*,
(Master's Thesis, Universiti Putra Malaysia, Serdang, Malaysia). Retrieved on 27-09-2021
- Monir, M.S., Yusoff, S.M., Zulperi, Z.M., Hassim, H.A., Mohamad, A., Hafiz Ngoo, M.S.M., Ina-Salwany, M.Y.
Haemato-immunological responses and effectiveness of the feed-based bivalent vaccine against *Streptococcus iniae* and *Aeromonas hydrophila* infections in hybrid red tilapia (*Oreochromis mossambicus* × *O. niloticus*)
(2020) *BMC Vet. Res*, 16, p. 226.

- Aslah, M., Zamri-Saad, M., Amal, M.N.A., Al-saari, N., Monir, M.S., Chin, Y.K., Ina-Salwany, M.Y.
Vaccine efficacy of a newly developed feed-based whole-cell polyvalent vaccine against vibriosis, streptococcosis and motile aeromonad septicemia in Asian seabass, *Lates calcarifer*
(2021) *Vaccines*, 9 (4), p. 368.
- Mohd-Aris, A., Zamri-Saad, M., Hassan, M.D., Yusof, M.T., Ina-Salwany, M.Y.
Vibrio harveyi* protease deletion mutant as a live attenuated vaccine candidate against vibriosis and transcriptome profiling following vaccination for *Epinephelus fuscoguttatus
(2019) *Aquac. Int*, 27 (1), pp. 125-140.
- Chin, Y.K., Al-saari, N., Zulperi, Z., Mohd-Aris, A., Salleh, A., Silvaraj, S., Aslah, M., Ina-Salwany, M.Y.
Efficacy of bath vaccination with a live attenuated *Vibrio harveyi* against vibriosis in Asian seabass fingerling, *Lates calcarifer*
(2020) *Aquac. Res*, 51 (1), pp. 389-399.
- Nehlah, R., Ina-Salwany, M.Y., Zulperi, Z.
Antigenicity analysis and molecular characterisation of two outer membrane proteins of *Vibrio alginolyticus* strain VA2 as vaccine candidates in tiger grouper culture
(2016) *J. Biol. Sci*, 16 (1-2), pp. 1-11.
- Nehlah, R., Firdaus-Nawi, M., Nik-Haiha, N.Y., Karim, M., Zamri-Saad, M., Ina-Salwany, M.Y.
Recombinant vaccine protects juvenile hybrid grouper, *Epinephelus fuscoguttatus* × *Epinephelus lanceolatus*, against infection by *Vibrio alginolyticus*
(2017) *Aquac. Int*, 25 (6), pp. 2047-2059.
- Silvaraj, S., Md Yasin, I.S., Karim, A., Saad, M.Z.
Elucidating the efficacy of vaccination against vibriosis in *Lates calcarifer* using two recombinant protein vaccines containing the outer membrane protein K (r-OmpK) of *Vibrio alginolyticus* and the DNA Chaperone J (r-DnaJ) of *Vibrio harveyi*
(2020) *Vaccines*, 8 (4), p. 660.
M.M. and
- Galindo-Villegas, J., Tobar, J.A., Mutoloki, S., Mweemba Munang'andu, H., Evensen, Ø.
Oral vaccination of fish-antigen preparations, uptake, and immune induction
(2015) *Front. Immunol*, 6, p. 519.
- Somamoto, T., Nakanishi, T.
Mucosal delivery of fish vaccines: Local and systemic immunity following mucosal immunizations
(2020) *Fish Shellfish Immunol*, 99, pp. 199-207.
- Sharma, A., Gautam, S., Bandyopadhyay, N.
Enzyme immunoassays: Overview
(1999) *Encyclopedia of Food Microbiology*, pp. p625-p633.
Batt, C.A. and Tortorello, M.L., editor. 2nd ed. Academic Press, Massachusetts

- de las Heras, A.I., Rodríguez Saint-Jean, S., Pérez- Prieto, S.I.
Immunogenic and protective effects of an oral DNA vaccine against infectious pancreatic necrosis virus in fish
(2010) *Fish Shellfish Immunol*, 28 (4), pp. 562-570.
- Zhang, W., Zhu, C., Chi, H., Liu, X., Gong, H., Xie, A., Zheng, W., Wu, Y.
Early immune response in large yellow croaker (*Larimichthys crocea*) after immunization with the oral vaccine
(2021) *Mol. Cell. Probes*, 56, p. 101708.
- Ramos-Espinoza, F.C., Cueva-Quiroz, V.A., Yunis- Aguinaga, J., Alvarez-Rubio, N.C., Paganoti de Mello, N., Engrácia de Moraes, J.R.
Efficacy of two adjuvants administrated with a novel hydrogen peroxide-inactivated vaccine against *Streptococcus agalactiae* in Nile tilapia fingerlings
(2020) *Fish Shellfish Immunol*, 105 (4), pp. 350-358.
- Sun, B., Yu, S., Zhao, D., Guo, S., Wang, X., Zhao, K.
Polysaccharides as vaccine adjuvants
(2018) *Vaccine*, 36 (35), pp. 5226-5234.
- Firdaus-Nawi, M., Noraini, O., Sabri, M.Y., Siti-Zahrah, A., Zamri-Saad, M., Latifah, H.
The effects of oral vaccination of *Streptococcus agalactiae* on stimulating gut-associated lymphoid tissues (GALTs) in tilapia (*Oreochromis spp.*)
(2011) *Pertanika J. Trop. Agric. Sci*, 34 (1), pp. 137-143.
- Aiyer Harini, P., Ashok Kumar, H., Gupta Praveen, K., Shivakumar, N.
An overview of immunologic adjuvants-a review
(2013) *J. Vaccines Vaccin*, 4 (1), p. 1000167.
- Mufti, A., Kumolosaso, E., Shamsuddin, A.F.
Physicochemical characterization and stability studies of duck pasteurellosis nanovaccine using palm oil (MCT-LCT) lipid emulsion as an adjuvant
(2017) *Acta Pol. Pharm*, 74 (4), pp. 1167-1175.
- Wang, Q., Ji, W., Xu, Z.
Current use and development of fish vaccines in China
(2020) *Fish Shellfish Immunol*, 96, pp. 223-234.
- Yardimci, R.E., Turgay, E.
Diagnosis of *Aeromonas sobria* and *Saprolegnia sp.* co-infection in rainbow trout fry (*Oncorhynchus mykiss*)
(2021) *Aquat. Res*, 4 (1), pp. 65-72.
- Fowoyo, P.T., Achimugu, F.
Virulence of *Aeromonas hydrophila* isolated from freshwater catfish
(2019) *J. Biosci. Med*, 7 (1), pp. 1-12.
- Majtán, J., Černý, J., Ofúkana, A., Takáč, P., Kozánek, M.
Mortality of therapeutic fish garra rufa caused by *Aeromonas sobria*
(2012) *Asian Pac. J. Trop. Biomed*, 2 (2), pp. 85-87.
- Shameena, S.S., Kumar, K., Kumar, S., Kumar, S., Rathore, G.
Virulence characteristics of *Aeromonas veronii* biovars isolated from infected freshwater goldfish (*Carassius auratus*)
(2020) *Aquaculture*, 518, p. 734819.

- Li, T., Raza, S.H.A., Yang, B., Sun, Y., Wang, G., Sun, W., Qian, A., Shan, X.
Aeromonas veronii Infection in commercial freshwater fish: A potential threat to public health
(2020) *Animals*, 10 (4), p. 608.
- Roy, A., Singha, J., Abraham, T.J.
Histopathology of Aeromonas caviae infection in challenged Nile tilapia Oreochromis niloticus (Linnaeus, 1758)
(2018) *Int. J. Aquac*, 8 (20), pp. 151-155.
- Stratev, D., Odeyemi, O.A.
An overview of motile Aeromonas septicaemia management
(2017) *Aquac. Int*, 25 (3), pp. 1095-1105.
- Laith, A.A., Najiah, M.
Aeromonas hydrophila: Antimicrobial susceptibility and histopathology of isolates from diseased catfish, Clarias gariepinus (Burchell)
(2013) *J. Aquac. Res. Dev*, 5 (2), p. 1000215.
- Mahmood, S., Mansor, N.N., Abdullah, S., Chowdhury, A., Ramly, R., Amal, M.N.A.
Assessment of bacteria and water quality parameters in cage cultured Pangasius hypophthalmus in Temerloh, Pahang River, Malaysia
(2019) *Nat. Environ. Pollut. Technol*, 18 (5), pp. 1479-1486.
- Pauzi, N.A., Mohamad, N., Azzam-Sayuti, M., Ina- Salwany, M.Y., Zamri-Saad, M., Nasruddin, N.S., Amal, M.N.A.
Antibiotic susceptibility and pathogenicity of Aeromonas hydrophila isolated from red hybrid tilapia (Oreochromis niloticus×Oreochromis mossambicus) in Malaysia
(2020) *Vet. World*, 13 (10), pp. 2166-2171.
- Baumgartner, W.A., Lorelei, F., Hanson, L.
Lesions caused by virulent Aeromonas hydrophila in farmed catfish (Ictalurus punctatus and I. punctatus×I. furcatus) in Mississippi
(2017) *J. Vet. Diagn. Invest*, 29 (5), pp. 747-751.
- Roges, E.M., Gonçalves, V.D., Cardoso, M.D., Festivo, M.L., Siciliano, S., Berto, L.H., Pereira, V.L.A., de Aquino, M.H.C.
Virulence-associated genes and antimicrobial resistance of Aeromonas hydrophila isolates from animal, food, and human sources in Brazil
(2020) *Biomed Res. Int*, 2020, p. 1052607.
- Ji, Y., Li, J., Qin, Z., Li, A., Gu, Z., Liu, X., Lin, L., Zhou, Y.
Contribution of nuclease to the pathogenesis of Aeromonas hydrophila
(2015) *Virulence*, 6 (5), pp. 515-522.
- Yadav, S.K., Meena, J.K., Sharma, M., Dixit, A.
Recombinant outer membrane protein C of Aeromonas hydrophila elicits mixed immune response and generates agglutinating antibodies
(2016) *Immunol. Res*, 64 (4), pp. 1087-1099.
- da Silva, B.C., Mouriño, J.L.P., Vieira, F.N., Jatobá, A., Seiffert, W.Q., Martins, M.L.
Haemorrhagic septicaemia in the hybrid surubim (Pseudoplatystoma corruscans×Pseudoplatystoma fasciatum) caused by Aeromonas hydrophila
(2012) *Aquac. Res*, 43 (6), pp. 908-916.

- Peterman, M.A., Posadas, B.C.
Direct economic impact of fish diseases on the East Mississippi catfish industry
(2019) *N. Am. J. Aquac*, 81 (3), pp. 222-229.
- Stratev, D., Odeyemi, O.A.
Antimicrobial resistance of *Aeromonas hydrophila* isolated from different food sources: A mini-review
(2016) *J. Infect. Public Health*, 9 (5), pp. 535-544.
- Saengsitthisak, B., Chaisri, W., Punyapornwithaya, V., Mektrirat, R., Klayraung, S., Bernard, J.K., Pikulkaew, S.
Occurrence and antimicrobial susceptibility profiles of multidrug-resistant aeromonads isolated from freshwater ornamental fish in Chiang Mai province
(2020) *Pathogens*, 9 (11), p. 973.
- Nayak, S.K.
Current prospects and challenges in fish vaccine development in India with special reference to *Aeromonas hydrophila* vaccine
(2020) *Fish Shellfish Immunol*, 100, pp. 283-299.
- Mzula, A., Wambura, P.N., Mdegela, R.H., Shirima, G.M.
Current state of modern biotechnological-based *Aeromonas hydrophila* vaccines for aquaculture: A systematic review
(2019) *Biomed Res. Int*, 2019, p. 3768948.
- Wong, S.Y., Leong, T.S.
A comparative study of *Vibrio* infections in healthy and diseased marine finfishes cultured in floating cages near Penang, Malaysia
(1990) *Asian Fish. Sci*, 3 (3), pp. 353-359.
- Ransangan, J., Mustafa, S.
Identification of *Vibrio harveyi* isolated from diseased Asian seabass *Lates calcarifer* using 16S ribosomal DNA sequencing
(2009) *J. Aquat. Anim. Health*, 21 (3), pp. 150-155.
- Ransangan, J., Lal, T.M., Al-Harbi, A.H.
Characterization and experimental infection of *Vibrio harveyi* isolated from diseased Asian seabass (*Lates calcarifer*)
(2012) *Malays. J. Microbiol*, 8 (2), pp. 104-115.
- Nurliyana, M., Aidil Roseli, M.F., Amal, M.N.A., Zamri-Saad, M., Ina-Salwany, M.Y., Zulkipli, N.A., Nasruddin, N.S.
Natural concurrent infection of *Vibrio harveyi* and *V. alginolyticus* in cultured hybrid groupers in Malaysia
(2019) *J. Aquat. Anim. Health*, 31 (1), pp. 88-96.
- Amalina, N.Z., Dzarifah, Z., Amal, M.N.A., Yusof, M.T., Zamri-Saad, M., Al-saari, N., Tanaka, M., Ina-Salwany, M.Y.
Recent update on the prevalence of *Vibrio* species among cultured grouper in Peninsular Malaysia
(2019) *Aquac. Res*, 50 (11), pp. 3202-3210.

- Norhariyani, M.N., Siti Hajar, M.Y., Hassan, M.D., Amal, M.N.A., Nurliyana, M.
Costs of management practices of Asian seabass (*Lates calcarifer* Bloch, 1790) cage culture in Malaysia using a stochastic model that includes uncertainty in mortality
(2019) *Aquaculture*, 510, pp. 347-352.
- Mohd-Aris, A., Ina-Salwany, M.Y., Zamri-Saad, M., Hassan, M.D., Norfarrah, M.A.
Molecular characterization of *Vibrio harveyi* virulence-associated serine protease and outer membrane protein genes for vaccine development
(2016) *Int. J. Biosci*, 8 (3), pp. 10-28.
- Abu Nor, N., Zamri-Saad, M., Ina-Salwany, M.Y., Salleh, A., Mustaffa-Kamal, F., Matori, M.F., Amal, M.N.A.
Efficacy of the whole cell inactivated *Vibrio harveyi* vaccine against vibriosis in a marine red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) model
(2020) *Vaccines*, 8 (4), p. 734.
- Maiti, B., Dubey, S., Munang'andu, H.M., Karunasagar, I., Karunasagar, I., Evensen, Ø.
Application of outer membrane protein-based vaccines against major bacterial fish pathogens in India
(2020) *Front. Immunol*, 11, p. 1362.
- Ismail, N., Nor, M.J.M., Sidek, S.
A framework for a successful research products commercialization: A case of Malaysian academic researchers
(2015) *Proc. Soc. Behav. Sci*, 195, pp. 283-292.
- Khademi, T., Ismail, K., Tin Lee, C., Shafaghat, A.
Enhancing commercialisation level of academic research outputs in a research university
(2015) *J. Teknol*, 74 (4), pp. 141-151.
- Ismail, N., Nor, M.J.M., Sidek, S.
Challenges for research product commercialisation: A case of Malaysian academic researchers
(2017) *J. Eng. Appl. Sci*, 12 (6), pp. 1543-1550.
- Heng, L.H., Rasli, A.M., Senin, A.A.
Knowledge determinant in university commercialisation: A case study of Malaysia public university
(2012) *Proc. Soc. Behav. Sci*, 40, pp. 251-257.
- Latif, N.S.A., Abdullah, A., Jan, N.M.
A pilot study of entrepreneurial orientation towards commercialization of university research products
(2016) *Proc. Econ. Finance*, 37, pp. 93-99.
- **Annex 3 WHO good manufacturing practices for biological products**
(2016) *WHO Technical Report Series No. 999*,
World Health Organization, Geneva. Retrieved on 12-09-2021
- (2017) *Southeast Asian State of Fisheries and Aquaculture 2017*,
Southeast Asian Fisheries Development Center, Bangkok, Thailand. Retrieved on 12-09-2021

- Mitchell, H.
The pitfalls of field trials in fish vaccinology
(1997) *Dev. Biol. Stand*, 90, pp. 321-332.
- Midtlyng, P.J.
Methods for measuring efficacy, safety and potency of fish vaccines
(2016) *Fish Vaccines*, pp. p119-p141.
Adams, A., editor. 1st ed. Springer, Basel
- Knight-Jones, T.J.D., Edmond, K., Gubbins, S., Paton, D.J.
Veterinary and human vaccine evaluation methods
(2014) *Proc. Biol. Sci*, 281 (1784), p. 20132839.
- Crane, M., Hyatt, A.
Viruses of fish: An overview of significant pathogens
(2011) *Viruses*, 3 (11), pp. 2025-2046.
- Stepien, C.A., Pierce, L.R., Leaman, D.W., Niner, M.D., Shepherd, B.S.
Gene diversification of an emerging pathogen: A decade of mutation in a novel fish Viral Hemorrhagic Septicemia (VHS) substrain since its first appearance in the Laurentian Great Lakes
(2015) *PLoS One*, 10 (8), p. E0135146.
- Kennedy, D.A., Read, A.F.
Why the evolution of vaccine resistance is less of a concern than the evolution of drug resistance
(2018) *Proc. Natl. Acad. Sci. U. S. A*, 115 (51), pp. 12878-12886.
- Naylor, R.L., Hardy, R.W., Buschmann, A.H., Bush, S.R., Cao, L., Klinger, D.H., Little, D.C., Troell, M.
A 20-year retrospective review of global aquaculture
(2021) *Nature*, 591 (7851), pp. 551-563.
- Lihan, S., Jamil, N.A., Jamian, M.A.H., Chiew, T.S., Ajibola, O.O., Justin, S., Benet, F., Kion, L.N.
Distribution and prevalence of antibiotic-resistant bacteria in fish farms in East Malaysia
(2020) *Malays. J. Microbiol*, 16 (4), pp. 263-274.
- Fauzi, N.N.F., Hamda, R.H., Mohamed, M., Ismail, A., Zin, A.A.M., Mohamad, N.F.A.
Prevalence, antibiotic susceptibility, and presence of drug resistance genes in *Aeromonas* spp. isolated from freshwater fish in Kelantan and Terengganu states, Malaysia
(2021) *Vet. World*, 14 (8), pp. 2064-2072.
- Atyah, M.A.S., Zamri-Saad, M., Siti-Zahrah, A.
First report of methicillin-resistant *Staphylococcus aureus* from cage-cultured tilapia (*Oreochromis niloticus*)
(2010) *Vet. Microbiol*, 144 (3-4), pp. 502-504.
- Gudding, R., van Muiswinkel, W.B.
A history of fish vaccination: Science-based disease prevention in aquaculture
(2013) *Fish Shellfish Immunol*, 35 (6), pp. 1683-1688.

- Eggset, G., Mikkelsen, H., Killie, J.E.A.
Immunocompetence and duration of immunity against *Vibrio salmonicida* and *Aeromonas salmonicida* after vaccination of Atlantic salmon (*Salmo salar* L.) at low and high temperatures
(1997) *Fish Shellfish Immunol*, 7 (4), pp. 247-260.
- Midtlyng, P.J.
A field study on intraperitoneal vaccination of Atlantic salmon (*Salmo salar* L.) against furunculosis
(1996) *Fish Shellfish Immunol*, 6 (8), pp. 553-565.
- Kashulin, A., Seredkina, N., Sørum, H.
Cold-water vibriosis. The current status of knowledge
(2017) *J. Fish Dis*, 40 (1), pp. 119-126.
- Midtlyng, P.J., Grave, K., Horsberg, T.E.
What has been done to minimise the use of antibacterial and antiparasitic drugs in Norwegian aquaculture?
(2011) *Aquac. Res*, 42 (s1), pp. 28-34.

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