

ZooKeys 554: 1–25 (2016)
doi: 10.3897/zookeys.554.6745
<http://zookeys.pensoft.net>

RESEARCH ARTICLE

A peer-reviewed open-access journal

Launched to accelerate biodiversity research

JellyWeb: an interactive information system on Scyphozoa, Cubozoa and Staurozoa

Stefano Martellos¹, Luca Ukosich¹, Massimo Avian¹

¹ *University of Trieste, Department of Life Sciences, via L. Giorgieri 10, I-34127 Trieste, Italy*

Corresponding author: *Stefano Martellos* (martelst@units.it)

Academic editor: *L. van Ofwegen* | Received 2 October 2015 | Accepted 25 November 2015 | Published 18 January 2016

<http://zoobank.org/AA6565FA-986E-4616-80D3-E15329A3E3BD>

Citation: Martellos S, Ukosich L, Avian M (2016) JellyWeb: an interactive information system on Scyphozoa, Cubozoa and Staurozoa. *ZooKeys* 554: 1–25. doi: 10.3897/zookeys.554.6745

Abstract

Identification of organisms is traditionally based on the use of “classic” identification keys, normally printed on paper. These keys have several drawbacks: they are mainly based on the systematics, requiring identification of orders, families and genera at first; they are written by experts for other experts, in a specific scientific jargon; they have a “frozen” structure (sequence of theses/antitheses); once published, they cannot be changed or updated without printing a new edition. Due to the use of computers, it is now possible to build new digital identification tools, which: 1) can be produced automatically, if the characters are stored in a database; 2) can be freed from the traditional systematics, giving priority to easy-to-observe characters, incl. those usually uncommon to the classical keys, such as ecology and distribution; 3) can be updated in real time once published on-line; 4) can be available on different media, and on mobile devices. An important feature of these new digital tools is their “collaborative” nature. They can be enriched by the contribution of several researchers, which can cooperate while maintaining rights and property of the resources and data they contribute to the system. JellyWeb, the information system on Scyphozoa, Cubozoa and Staurozoa has been developed in Trieste since 2010. The system was created with the aim of – potentially – becoming a starting point for a wide collaborative effort in developing a user-friendly worldwide digital identification system for jellyfishes.

Keywords

Biodiversity informatics, Cnidaria, FRIDA, identification, jellyfish, Medusozoa

Introduction

Since the Rio Earth Summit in 1992, access to biodiversity information has become a fundamental task. Biodiversity data are targeted by several efforts of digitalization and aggregation, most of which focus on primary biodiversity data, i.e. natural history collection specimens and field records. Some of these efforts produced wide global networks, e.g. the GBIF (Global Biodiversity Information Facility; Berendsohn et al. 2010, King et al. 2010), which, together with the BioCASE (Biodiversity Collection Access for Europe, Holetschek et al. 2012), is mobilizing ca. 600 millions of records. Primary biodiversity data are mostly used in modeling the distribution of the taxa, and in predicting the effect of climate changes and anthropic pressure on endangered or alien invasive taxa. Taxon related information (nomenclature, auto-ecology, etc.) become the focus of similar large scale efforts only in the last years (Martellos and Attorre 2012, Martellos 2014). The GBIF itself is starting to aggregate checklist data (GBIF 2010), while other efforts are focused on molecular data (Field et al. 2011, Holetschek et al. 2012, Wieczorek et al. 2012), and to ecological information (Fegraus et al. 2005). In the field of hydrobiology, some recent examples can be Fish-SPRICH (Brosse et al. 2013) and Fish-AMAZBOL (Carvajal-Vallejos et al. 2014). In the case of jellyfishes, online resources are however scarce, but some relevant exceptions (e.g., the Jellyfish Dataset Initiative, <http://www.bco-dmo.org/dataset/526852>).

Digital identification keys are a particular case in the world of biodiversity informatics. Since the development of the DELTA language (Dallwitz 1980), efforts aiming at creating online digital identification keys followed several approaches. The resulting products differ in usability, accessibility, size, etc. (Nimis and Martellos 2009, Hagedorn et al. 2010, Randlane et al. 2010, Martellos and Nimis 2015). With the development of FRIDA (FRiendly IDentificAtion, Martellos 2010), the researchers of the Department of Life Sciences, University of Trieste, aimed at producing a simple but effective instrument for the development of digital identification keys in collaborative efforts. This led to the publication – in the framework of project *Dryades*, and of the EU projects KeyToNature (<http://www.keytonature.eu>), SiiT (<http://www.siiit.eu>) and CSMON-LIFE (LIFE13 ENV/IT/842, <http://www.csmon-life.eu>) – of ca. 600 different digital identification keys for several groups of organisms.

As far as Scyphozoa, Cubozoa and Staurozoa are concerned, there are digital databases hosting taxonomic information, such as WoRMS (World Register of Marine Species, <http://www.marinespecies.org/>), as well as paper printed keys to genera (as an example, see Cornelius 1997; other keys are listed in Morandini et al., 2005). Few examples of digital resources are available in the web, often limited to specific geographic regions, as the Cubozoan and scyphozoan key of the Carolinian Biogeographic Province (Calder 2009), the key to the Scyphozoa and Cubozoa of the South Atlantic Bight (Calder and King 2008), and, as far as the Mediterranean is concerned, the web site *MeteoMeduse* (Boero 2013, <http://meteomeduse.focus.it/>). The latter, however, is an example of citizen science observatory, and does not provide an identification key. To our knowledge, no comprehensive digital identification tools to species of these taxa exist.

By combining taxonomical, ecological, and morphological and anatomical features into an information system, we developed the so called JellyWeb, a simple tool which allow to researchers and laypersons to identify Scyphozoa, Cubozoa and Staurozoa to the species level. This paper presents the results of this effort, available online at the URL <http://dryades.units.it/jelly>.

Methods

Data were collected from several sources in literature. The most relevant are Kramp (1961), WoRMS (<http://www.marinespecies.org/>), the Scyphozoan Wiki (<http://thescyphozoan.ucmerced.edu/>), and Mills (1999-). Further sources are listed in Balboni 2008, Benci 2008, Sarto 2009, Sola 2009, Coral 2012, Benci 2012, Savonitto 2012, Ukosich 2014. Other paper are under consideration, and will lead to adding to the database other species for several genera, such as *Atolla* (*A. russelli*, *A. gigantea*, *A. chuni*), *Aurelia* (*A. marginalis*), *Chironex* (*C. yamaguti*), *Cyanea* (*C. lamarkii*, *C. rosea*, *C. annaskala*, *C. tzetlinii*, and several other species), *Desmonema* (*D. comatum*, *D. scoresbyanna*), *Drymonema* (*D. gorgo*, *D. larsoni*), *Nausithoe* (*N. marginata*), *Pelagia* (*P. benovici*) *Tripedalia* (*T. binata*).

The information system is freely available online at the URL <http://dryades.units.it/jelly>. It organizes data collected in the last five years by the research unit headed by Massimo Avian, at the Dept. of Life Sciences of the University of Trieste. The researchers which contributed to the project agreed on distributing the data under a Creative Commons, share alike, by attribution 3.0 (CC 3.0 by-sa) license.

The software of the information system has been developed in PHP language. The data are stored in a MySQL database. The system is equipped with a multi-entry query interface (Hagedorn et al. 2010), which operates on both a taxonomic database, and a database of nine easily recognizable morphological characters (see below). The multi-entry interface allows complex queries, which can be a first step in the identification of an organism. The multi-entry query system returns lists of taxa, on which the identification process can continue by using a digital identification system. The latter has been developed by using the FRIDA (FRiendly IDentificAtion) package (Martellos 2010). It operates on a morpho-anatomical database, which hosts ca. 200 characters for several infra-generic taxa of Scyphozoa, Cubozoa and Staurozoa (a revision of the content of the database due to recent taxonomic advancements is ongoing). The output of the digital identification system is a digital identification key to the remaining taxa, which can be used by an interactive interface, or printed out as a dichotomous, illustrated key. The whole key can also be exported in a stand-alone version for mobile devices (Nimis et al. 2012).

The query interfaces have been developed according to the results of several usability tests, conducted in the framework of projects KeyToNature and SiiT, as detailed in Martellos and Nimis (2015). The system is under continuous development, following users' input.

Results

JellyWeb hosts several information pages and two query system. The home page (<http://dryades.units.it/jelly>) provides access to several sections: information, describing how the system works; survey area; query (detailed below); checklist, listing all taxa alphabetically by genus and species name, and providing access to their taxon pages; credits.

The query system is made of two parts.

- 1) Multi-entry interface (Fig. 1). The first interface of the query system provides the users with the opportunity of specifying a set of nine easily observable characters, and/or scientific name and family. The morphological characters are:
 - Jellyfish sessile / swimming;
 - Umbrella shaped like a cube or a box / not shaped like a cube or a box;
 - Tentacles present / absent;
 - Tentacles isolated / grouped in clusters;
 - Umbrella with a coronal groove / without a coronal groove;
 - Umbrella flat / not flat;
 - Oral arms absent / 4 / more than 4;
 - Jellyfish with filaments (oral arm appendages) / without filaments;
 - Jellyfish with scapulae / without scapulae.

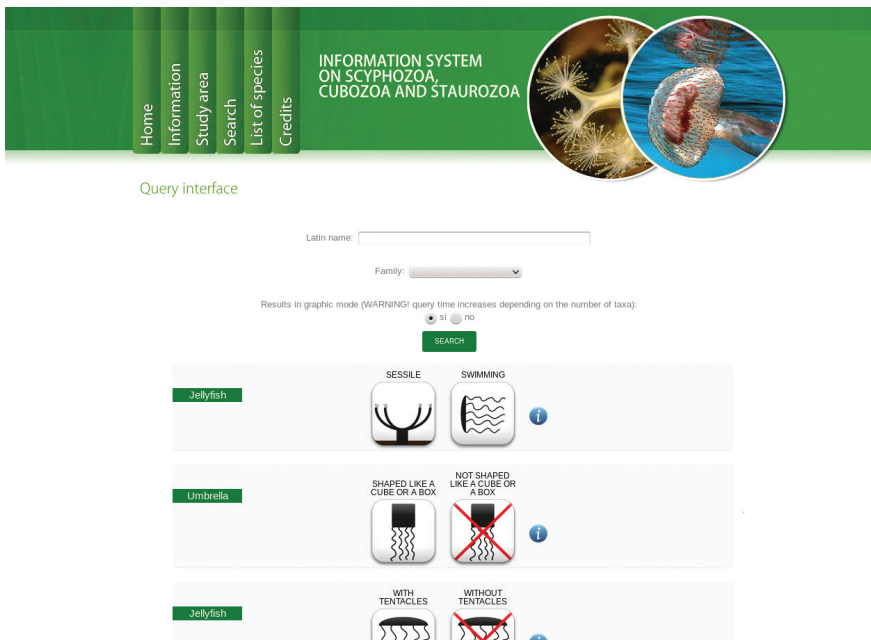


Figure 1. Multi-entry interface. The multi-entry interface allows to combine the states of several morphologic and anatomic characters, together with taxonomic information, to query the database.

For each character, an information popup window with images and text detailing the most relevant features is accessible by clicking on the question mark button. The result of a query is a list of taxa (Fig. 2). For each taxon an image is displayed (if available, see below). A link provides access to the taxon page (Fig. 3), which displays a description, as well as all the images available in the system, with credits and metadata, and other information (when available). Taxon pages can host a virtually unlimited amount of information and images, and/or provide access to external resources through HTML links.

- 2) Digital identification key. The results page of the multi-entry interface allows to generate an interactive identification key to remaining taxa. The key can be used through a simple single entry interface (Fig. 4, Hagedorn et al. 2010), or printed out as a textual, illustrated dichotomous key. At each step of the identification process users can list out the remaining taxa, or print an illustrated key. By clicking on a taxon name, the corresponding taxon page is shown (Fig. 3). Each key generated by this system is different from the others, since they contain a different number of infra-generic taxa. Normally, the lower the number of taxa is, the easier the resulting key. A key to all the taxa currently included in our databases can also be generated, and is provided below.

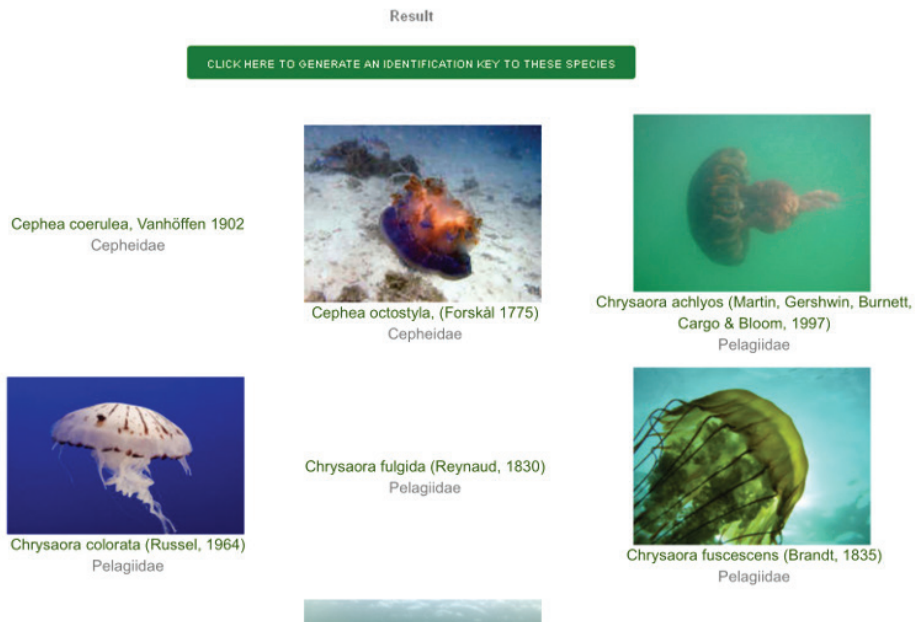


Figure 2. List of taxa. The result of a query made by using the multi-entry interface is an illustrated list of infra-generic taxa.

[Home](#)
[Information](#)
[Study area](#)
[Search](#)
[List of species](#)
[Credits](#)

INFORMATION SYSTEM
ON SCYPHOZOA,
CUBOZOA AND STAUROZOA



Aurelia aurita (Linnaeus, 1758)



© BS Thamer Hof
by BS Thamer Hof

SYSTEMATIC POSITION

Classis: Scyphozoa
 SubClassis: Dicommedusae
 Ordo: Semeaeostomae
 Familia: Ulmaridae (ex Aurellidae)
 SubFamilia: Aurellinae
 Genus: Aurelia (Phalon & Lancelot, 1899)
 Species: A. aurita (Linnaeus, 1775)

SYNONYMS

Aurelia coerulea

COMMON NAMES

Moon Jellyfish

MORPHO-ANATOMICAL CHARACTERISTICS

Genus Aurelia

- tentacles arising from the side of exumbrella, above the margin
- lappet-like structures arising from the side of exumbrella above the margin
- bell margin divided in 8 or 16 broad velar lobes
- 8 rhopalia, each with 2 ocelli
- some or all radial canals with anastomosing branches
- with ring canal
- invaginated gonads with external subgenital pits

Aurelia aurita

- flat thin bell, diameter up to 40 cm, smooth surface
- very transparent
- numerous thin short tentacles of various colours
- 8 broad velar lappets
- 4 linear thick oral arms, with densely crenulated margins, as long as bell's radius
- 4 interradial stomach pouches, each with a line of gastric filaments, just centripetal to gonads
- 8 rhopalar and 8 adradial canals unbranched, others 16 branched with anastomoses
- 4 evident brilliant horseshoe-shaped gonads, lining gastric pouches

GEOGRAPHICAL AND SEASONAL DISTRIBUTION

Coastal species, Circum-Boreal distribution, occurring usually at spring

STING EFFECTS

The venom of A. aurita contains potent lethal dermonecrotic, vasopermeability and hemolytic factors, but great differences have been reported concerning the dangerousness of specimens from different zones of the world. Specimens from Red Sea were stated to be most dangerous, causing local pain, punctions, urticaria, ulcerations. In other zones, usually causing only local itching, but may possible cross-reaction between venom of Aurelia and ones of C. quinquacirris, Chronex fleckeri and Physalia physalis

Figure 3. Taxon page. A typical taxon page displays an image, a description, as well as all the other images available in the system, together with credits and metadata. Taxon pages can host a virtually unlimited amount of data, links and media.

Dichotomous key to all taxa

This key was automatically generated by the system, and contains all the infra-generic taxa currently stored in our databases at the date October 30, 2015. When a taxon is added to the system the key automatically changes. Hence, the key an user will obtain in the future will be slightly – or completely – different. The keys are not the transposition of an existing paper printed key, but are automatically generated by the system from a database for morphological and anatomical characters by using the package FRIDA (Martellos 2010).

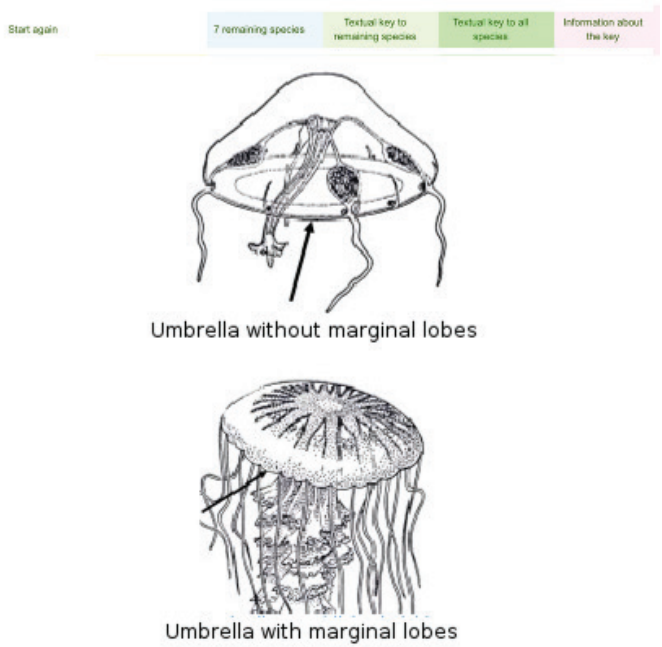


Figure 4. Single entry digital identification key. The digital identification key to remaining taxa is generated from the results of the multi-entry query system. It is used through a single entry interface, and can be printed out as a textual, illustrated dichotomous key as well.

1	Medusa sessile.....	2
–	Medusa swimming	34
2 (1)	Medusa without aboral peduncle	
 <i>Lucernariopsis vanhoeffeni</i> (Browne, 1910)	
–	Medusa with aboral peduncle	3
3 (2)	Medusa with sense organs: rhopalioids (anchors)	4
–	Medusa without sense organs.....	15
4 (3)	Coronal muscle divided.....	5
–	Coronal muscle unbroken	10
5 (4)	Calyx not conical.....	6
–	Calyx conical	7
6 (5)	Calyx quadro-pyramidal	<i>Halichlystus borealis</i> Uchida, 1933
–	Calyx pyramidal, octangular	<i>Halichlystus salpinx</i> Clark, 1863
7 (5)	Marginal anchors fairly large, egg-shaped.....	
 <i>Halichlystus stejnegeri</i> Kishinouye, 1899	
–	Not as above.....	8

8 (7) Marginal anchors very large, biscuit-shaped..... *Halicylistus antarcticus* Pfeffer, 1889
 9
 – Not as above..... 9
 9 (8) Marginal anchors kidney-shaped, with a short, cylindric stalk
 *Halicylistus auricula* (Rathke, 1806)
 10
 – Marginal anchors small, oval. *Halicylistus kerguelensis* Vanhöffen, 1908
 10 (4) Peduncle single-chambered..... *Manania hexaradiata* (Broch, 1907)
 11
 – Peduncle with 4 perradial chambers..... 11
 11 (10) Gonads not united by a transverse circumferential membrane (claustrum)
 which divide each of the 4 perradial stomach pouches into an outer and
 an inner space..... *Stenoscyphus inabai* (Kishinouye, 1893)
 12
 – Gonads united by a transverse circumferential membrane (claustrum)
 which divide each of the 4 perradial stomach pouches into an outer and
 an inner space..... 12
 12 (11) Calyx as long as wide..... *Manania williamsi* Larson & Fautin, 1989
 13
 – Calyx longer than wide..... 13
 13 (12) Calyx with dark herringbone pattern
 *Manania distincta* (Kishinouye, 1910)
 14
 – Calyx without dark herringbone pattern..... 14
 14 (13) Arms twice as long as broad..... *Halimocyathus platypus* Clark, 1863
 15
 – Arms short..... *Manania handi* Larson & Fautin, 1989
 16
 15 (3) Peduncle with 4 perradial chambers..... 16
 – Peduncle single-chambered..... 22
 16 (15) Peduncle with muscle in the septa 17
 – Peduncle without muscle in the septa 18
 17 (16) On each arm about 9 tentacles *Depastrum cyathiforme* (M. Sars,
 1846)
 18
 – On each arm about 25 tentacles
 *Depastromorpha africana* Carlgren, 1935
 19
 18 (16) Gonads united by a transverse circumferential membrane (claustrum)
 which divide each of the 4 perradial stomach pouches into an outer and
 an inner space..... 19
 – Gonads not united by a transverse circumferential membrane (claustrum)
 which divide each of the 4 perradial stomach pouches into an outer and
 an inner space..... 20
 19 (18) On each arm 60–80 tentacles
 *Craterolophus convolvulus* (Johnston, 1835)
 21
 – On each arm about 30 tentacles
 *Craterolophus macrocystis* von Lendenfeld, 1884
 22
 20 (18) Arms adradial *Kishinouyea nagatensis* (Oka, 1897)
 23
 – Arms interradial..... 21
 21 (20) Arms larger at base than *S. tsingtaoensis*.....
 *Sasakiella cruciformis* Okubo, 1917
 24

–	Arms narrower at base than <i>S. cruciformis</i>
	<i>Sasakiella tsingtaoensis</i> Ling, 1937
22 (15)	Peduncle without muscle in the septa
	<i>Lucernariopsis campanulata</i> (Lamouroux, 1815)
–	Peduncle with muscle in the septa	23
23 (22)	Marginal lobes (arms) faintly developed.....	24
–	Marginal lobes (arms) well developed	26
24 (23)	Tentacles reduced	<i>Lipkea stephensoni</i> Carlgren, 1933
–	Not as above.....	25
25 (24)	Tentacles not true	<i>Lipkea ruspoliana</i> Vogt, 1886
–	Tentacles rudimentary	<i>Lipkea sturdzi</i> (Antipa, 1893)
26 (23)	Tentacles up to 60 on each arm	27
–	Tentacles more than 60 on each arm.....	28
27 (26)	Subumbrellar margin with 4 perradial pigment spots.....
	<i>Stylocoronella riedli</i> Salvini-Plawen, 1966
–	Subumbrellar margin without 4 perradial pigment spots
	<i>Stylocoronella variabilis</i> Salvini-Plawen, 1987
28 (26)	Peduncle rudimentary.....	<i>Lucernaria australis</i> Vanhöffen, 1908
–	Peduncle true.....	29
29 (28)	Peduncle as long or longer than height of calyx.....	30
–	Peduncle shorter than height of calyx.....	31
30 (29)	Tentacles 100–140 on each arm. <i>Lucernaria quadricornis</i> O.F.Müller, 1776	
–	Tentacles 700–850 on each arm.....	<i>Lucernaria walteri</i> (Antipa, 1892)
31 (29)	Tentacles 80 or less on each arm
	<i>Lucernaria infundibulum</i> Haeckel, 1880
–	Tentacles more than 80 on each arm.....	32
32 (31)	Peduncle 1/3 as long as height of calyx
	<i>Lucernaria haeckeli</i> (Antipa, 1892)
–	Not as above.....	33
33 (32)	Peduncle less than 1/3 of the height of calyx.....
	<i>Lucernaria bathyphila</i> Haeckel, 1880
–	Peduncle about half as long as height of calyx
	<i>Lucernaria sainthilairei</i> (Redikorzev, 1925)
34 (1)	Medusa with calix.....
	<i>Tesserantha connectens</i>, Haeckel, 1880 – Warning: some authors debate on the validity of swimming Stauromedusae (see Rodriguez et al. 2011)	
–	Medusa with umbrella	35
35 (34)	Exumbrella divided by a circular and deep coronal groove.....	36
–	Exumbrella not divided by a circular and deep coronal groove.....	64
36 (35)	Tentacles from 4 to 6.....	37
–	Tentacles 8 or more	42
37 (36)	Rhopalia 4.....	38
–	Rhopalia 6.....	39

38 (37)	Gonads almost equidistant	<i>Pericolpa campana</i> (Haeckel, 1880)	
–	Gonads in 4 pairs	<i>Pericolpa quadrigata</i> Haeckel, 1880	
39 (37)	Gonads 6	<i>Atorella arcturi</i> Bigelow, 1928	
–	Not as above		40
40 (39)	Gonads 8	<i>Atorella octogonus</i> Mills, Larson & Young, 1987	
–	Gonads 4		41
41 (40)	Gonads sac-like, swollen	<i>Atorella subglobosa</i> Vanhöffen, 1902	
–	Gonads leaf-shaped	<i>Atorella vanhoeffeni</i> Bigelow, 1909	
42 (36)	Rhopalia up to 6		43
–	Rhopalia more than 6		48
43 (42)	Rhopalia perradial, 4		44
–	Rhopalia interradial, 4		45
44 (43)	Coronal muscle divided	<i>Paraphyllina intermedia</i> Maas, 1903	
–	Coronal muscle unbroken	<i>Paraphyllina ransoni</i> Russel, 1956	
45 (43)	Marginal lappets 32	<i>Nauphantopsis diomedea</i> Fewkes, 1885	
–	Not as above		46
46 (45)	Gonads 4	<i>Periphyllopsis galathea</i> Kramp, 1959	
–	Gonads 8		47
47 (46)	Marginal lappets 16	<i>Periphylla periphylla</i> (Péron & Lesueur, 1809)	
–	Marginal lappets 24	<i>Periphyllopsis braueri</i> Vanhöffen, 1902	
48 (42)	Gonads 4 or 4 pairs		49
–	Gonads 8		53
49 (48)	Stomach pouches break up into numerous ragged-edged branches in the marginal lappets		50
–	Stomach pouches simple, radiating		51
50 (49)	Subumbrellar protuberances in 2 circles	<i>Linuche aquila</i> Mayer 1910	
–	Subumbrellar protuberances in 3 circles	<i>Linuche unguiculata</i> (Schwartz, 1788)	
51 (49)	Gonads bean-shaped	<i>Palephyra indica</i> Vanhöffen, 1902	
–	Gonads crescent-shaped		52
52 (51)	Gonads with horns recurved	<i>Palephyra antiqua</i> Haeckel, 1880	
–	Gonads consisting of 3 swellings	<i>Palephyra pelagica</i> Haeckel, 1880	
53 (48)	Rhopalia > 8		54
–	Rhopalia 8 (Genus <i>Nausithoe</i> . The key refers to free-swimming stages only)		56
54 (53)	Gastric ostia with two pigmented spots	<i>Atolla vanhoeffeni</i> Russell, 1957	
–	Gastric ostia without pigmented spots		55
55 (54)	Species with 20–24 tentacles	<i>Atolla parva</i> Russell, 1958	
–	Species with usually 22, sometimes up to 32 tentacles	<i>Atolla wyvillei</i> Haeckel, 1880	
56 (53)	Central disk with large pits		57
–	Central disk without pits		58
57 (56)	Central disk with radiating furrows .	<i>Nausithoe rubra</i> Vanhöffen, 1902	

–	Central disk without radiating furrows....	<i>Nausithoe atlantica</i> Broch, 1914
58 (56)	Gonads very small	<i>Nausithoe clausi</i> Vanhöffen, 1892
–	Not as above.....	59
59 (58)	Gonads of normal dimensions	<i>Nausithoe albatrossi</i> (Maas, 1897)
–	Gonads large.....	60
60 (59)	Central disk not thick nor finely punctured.....	<i>Nausithoe globifera</i> Broch, 1914
–	Central disk thick, finely punctured.....	61
61 (60)	Central disk with radiating furrows	<i>Nausithoe challengerii</i> (Haeckel, 1880)
–	Central disk without radiating furrows	62
62 (61)	Medusa with chocolate brown or carmine gonads and blue gastric cirri ..	<i>Nausithoe picta</i> Agassisi & Mayer, 1902
–	Medusa without chocolate brown or carmine gonads and blue gastric cirri.....	63
63 (62)	Gastric cirri not grouped in clusters.....	<i>Nausithoe punctata</i> (Kölliker, 1853)
–	Gastric cirri grouped in clusters	<i>Nausithoe limpida</i> Hartlaub, 1909
64 (35)	Opening of the subumbrellar cavity partly closed by an annular diaphragm (velarium).....	65
–	Opening of the subumbrellar cavity not closed by an annular diaphragm (velarium).....	88
65 (64)	Tentacles 8 or more	66
–	Tentacles from 4 to 6.....	76
66 (65)	Stomach pouches without diverticula	<i>Tripedalia cystophora</i> Conant, 1897
–	Stomach pouches with 8 diverticula.....	67
67 (66)	Gonads not four-leaved	<i>Chiroadectes maculatus</i> (Cornelius, Fenner & Hore, 2005)
–	Gonads four-leaved.....	68
68 (67)	Medusa with nematocysts on bell	69
–	Medusa without nematocysts on bell	71
69 (68)	Each pedaliu with more than 4 fingers and tentacles.....	<i>Chiropsalmus quadrumanus</i> Müller, 1859
–	Each pedaliu with 4 or less fingers and tentacles	70
70 (69)	Each pedaliu with 2 fingers and tentacles.....	<i>Chiropsalmus zygonema</i> Haeckel, 1880
–	Each pedaliu with 3–4 fingers and tentacles.....	<i>Chiropsalmus alipes</i> Gershwin, 2006
71 (68)	Medusa with mesenteries poorly developed	<i>Chirosella bronzie</i> Gershwin, 2006
–	Not as above.....	72
72 (71)	Gastric saccules are functioning gonads....	<i>Chironex fleckeri</i> Southcott, 1956

–	Gastric saccules are not functioning gonads.....	73
73 (72)	Stomach pouches with 2 branched or feathered saccules.....	74
–	Stomach pouches with 2 unbranched saccules.....	75
74 (73)	Each pedaliium with 9–11 fingers and tentacles.....	
 <i>Chirodropus gorilla</i> Haeckel, 1880	
–	Each pedaliium with 21 fingers and tentacles.....	
 <i>Chirodropus palmatus</i> Haeckel, 1880	
75 (73)	Tentacles and fingers irregularly placed.....	
 <i>Chiropsoides buitendijki</i> (van der Horst, 1907)	
–	Tentacles and fingers not irregularly placed.....	
 <i>Chiropsoides quadrigatus</i> (Haeckel, 1880)	
76 (65)	Tentacles branched.....	<i>Manokia stiasnyi</i> Bigelow, 1938
–	Tentacles simple.....	77
77 (76)	Stomach with weakly developed mesenteries.....	78
–	Not as above.....	80
78 (77)	Sensory niches without well developed covering scale.....	
 <i>Copula sivickisi</i> Stiasny, 1926	
–	Sensory niches with covering scale above.....	79
79 (78)	Velarial canals 3–4 per octant... <i>Carybdea marsupialis</i> (Linnaeus, 1758)	
–	Velarial canals 2 per octant.....	<i>Carybdea rastonii</i> Haacke, 1886
80 (77)	Stomach without mesenteries.....	81
–	Stomach with well developed mesenteries.....	84
81 (80)	Exumbrella without nematocyst-warts.... <i>Alatina moseri</i> (Mayer, 1906)	
–	Exumbrella with nematocyst-warts.....	82
82 (81)	Velarial canals 3 per octant.....	<i>Alatina rainensis</i> Gershwin, 2005
–	Velarial canals 4–5 per octant.....	83
83 (82)	Medusa with 6 eyes per rhopalium.... <i>Alatina madraspartana</i> Menon, 1930	
–	Medusa with 1 eye per rhopalium.. <i>Alatina tetraptera</i> (Haeckel, 1880)	
84 (80)	Medusa with phacellae.....	<i>Tamoya haplonema</i> Müller, 1859
–	Medusa without phacellae.....	85
85 (84)	Velarial canals 1 per octant.....	<i>Carukia shinju</i> Gershwin, 2005
–	Not as above.....	86
86 (85)	Velarial canals 2 per octant.....	<i>Carukia barnesi</i> Southcott, 1966
–	Not as above.....	87
87 (86)	Velarial canals 4–5 per octant.....	<i>Malo maxima</i> Gershwin, 2005
–	Velarial canals more than 5 per octant.....	
 <i>Gerongia rifkinae</i> Gershwin & Alderslade, 2005	
88 (64)	Medusa with a permanent primary mouth opening in adult specimens....	89
–	Medusa without a permanent primary mouth opening in adult specimens.....	135
89 (88)	Medusa without tentacles.....	90
–	Medusa with tentacles.....	94
90 (89)	marginal lappets very shallow, or entirely lacking.....	91

–	marginal lappets evident	92
91 (90)	Exumbrella transparent white, sometimes with brown nuances on margins.....	<i>Deepstaria enigmatica</i> (Russel, 1967)
–	Exumbrella reddish-brown, with stomach margin lighter brown.....	<i>Deepstaria reticulum</i> (Lerson, Madin & Harbison, 1988)
92 (90)	Rhopalia from 24 to more than 50, one in every cleft between the lappets.....	<i>Tiburonia granrojo</i> (Matsumoto, Raskoff & Lindsay, 2003)
–	Not as above.....	93
93 (92)	Rhopalia 20.....	<i>Stygiomedusa gigantea</i> (Browne, 1910)
–	Rhopalia 8.....	<i>Stellamedusa ventana</i> (Raskoff & Matsumoto, 2004)
94 (89)	Medusa with ring-canal	95
–	Medusa without ring-canal	113
95 (94)	Tentacles arising from umbrella's margin.....	96
–	Tentacles not arising from umbrella's margin	104
96 (95)	Marginal lappets 48..... <i>Undosa undulata</i> (Stiasny, 1935) – Warning: dubious species, some authors suggest it is a juvenile stage of <i>Discomedusa lobata</i> Claus 1877
–	Not as above.....	97
97 (96)	Marginal lappets 16.....	98
–	Not as above.....	99
98 (97)	Oral arms broad, egg-shaped	<i>Ulmaris prototypus</i> (Haeckel, 1880)
–	Oral arms narrow and pointed.....	<i>Ulmaris snelliusi</i> (Stiasny, 1935)
99 (97)	Marginal lappets 32.....	100
–	Marginal lappets 64.....	102
100 (99)	Tentacles 32 or 48	<i>Discomedusa lobata</i> (Claus, 1877)
–	Tentacles 24.....	101
101 (100)	Perradial canals branched.....	<i>Discomedusa philippina</i> (Mayer, 1910)
–	Perradial canals not branched	<i>Floresca parthenia</i> (Haeckel, 1880)
102 (99)	Tentacles 24.....	<i>Parumbrosa polylobata</i> (Kishinouye, 1910)
–	Tentacles 16.....	103
103 (102)	Anastomoses absent.....	<i>Diplulmaris antarctica</i> (Maas, 1908)
–	Anastomoses present.....	<i>Diplulmaris malayensis</i> (Stiasny, 1935)
104 (95)	Tentacles arising from subumbrella.....	105
–	Tentacles arising from exumbrella.....	107
105 (104)	Gonads 8.....	<i>Poralia rufescens</i> (Vanhöffen, 1902)
–	Gonads 4.....	106
106 (105)	Rhopalia 8.....	<i>Sthenonia albida</i> (Eschscholtz, 1829)
–	Rhopalia 16.....	<i>Phacellophora camtschatica</i> (Brandt, 1835)
107 (104)	Oral arms bifurcated.....	<i>Aurosa furcata</i> (Haeckel, 1880)
–	Oral arms not bifurcated	108
108 (107)	Marginal lappets 16.....	<i>Aurelia labiata</i> (Chamisso & Eysenhardt, 1821)
–	Marginal lappets 8.....	109

- 109 (108) Oral arms short, thick and curved, much folded, extending laterally against subumbrellar surface *Aurelia limbata* (Brandt, 1835)
 – Not as above 110
- 110 (109) Oral arms linear, thick and stiff, with densely crenulated margins, as long as bell's radius *Aurelia aurita* (Linnaeus, 1758)
 – Not as above 111
- 111 (110) Oral arms narrow and thin, with slightly folded margins only in proximal part *Aurelia solida* (Browne, 1905)
 – Oral arms long and broad, curtain-like, with densely crenulated margins.. 112
- 112 (111) Adradial canals not branched *Aurelia maldivensis* (Bigelow, 1904)
 – Adradial canals branched *Aurelia colpata* (Brandt, 1838)
- 113 (94) Tentacles arising from the subumbrella at some distance from the margin 114
 – Tentacles arising from the exumbrellar margin 122
- 114 (113) Tentacles not arranged in tufts
 *Drymonema dalmatinum* (Haeckel, 1880)
 – Tentacles arranged in tufts 115
- 115 (114) Medusa without radial musculature in the subumbrella 116
 – Medusa with radial musculature in the subumbrella 118
- 116 (115) Medusa with few broad canals in the lappets
 *Desmonema gaudichaudi* (Lesson, 1830)
 – Medusa with numerous narrow canals in the lappets 117
- 117 (116) Tentacles not ribbon-like
 *Desmonema chierchianum* (Vanhöffen, 1888)
 – Tentacles ribbon-like *Desmonema glaciale* (Larson, 1986)
- 118 (115) Rhopalar and tentacular stomach pouches completely separated 119
 – Rhopalar and tentacular stomach pouches connected by anastomoses 120
- 119 (118) Peripheral canals without, or with few anastomoses
 *Cyanea capillata* (Linnaeus, 1758)
 – Peripheral canals with numerous anastomoses
 *Cyanea purpurea* (Kishinouye, 1910)
- 120 (118) Peripheral canals with numerous anastomoses
 *Cyanea nozakii* (Kishinouye, 1891)
 – Peripheral canals without, or with few anastomoses 121
- 121 (120) Radial muscles originating from the outer side of coronal muscle
 *Cyanea buitendijki* (Stiasny, 1919)
 – Radial muscles originating from the middle of coronal muscle
 *Cyanea mjobergi* (Stiasny, 1921)
- 122 (113) Stomach pouches 32 123
 – Stomach pouches 16 124
- 123 (122) Subgenital pits heart-shaped *Sanderia malayensis* (Goette, 1886)
 – Subgenital pits horseshoe-shaped
 *Sanderia pampinosus* (Gershwin & Zeidler, 2008)

124 (122)	Marginal lappets 16.....	125
–	Not as above.....	126
125 (124)	Nematocyst warts about as long as wide	
 <i>Pelagia noctiluca</i> (Forsskål, 1775)	
–	Nematocyst warts highly protrusive, more long than wide.....	
 <i>Pelagia flaveola</i> (Eschscholtz, 1829)	
126 (124)	Marginal lappets 48.....	127
–	Marginal lappets 32.....	129
127 (126)	Tentacles all alike.....	<i>Chrysaora fulgida</i> (Reynaud, 1830)
–	Tentacles different in length.....	128
128 (127)	Tentacles usually 5 per octant, 1 central primary, 2 lateral secondary about half in length, 2 tertiary, between former two types, about 1/4 as long as the median.....	<i>Chrysaora lactea</i> (Eschscholtz, 1829)
–	Tentacles 5 per octant, 3 primary arising from deep cleft between tentacular lappets and 2 lateral and shorter secondary, arising from subumbrellar side of rhopalar lappets	<i>Chrysaora quinquecirrha</i> (Desor, 1848)
129 (126)	Tentacles 8.....	<i>Chrysaora colorata</i> (Russel, 1964)
–	Tentacles 24.....	130
130 (129)	Exumbrella yellowish-brown or reddish-yellow with 32-rayed chestnut-brown star	<i>Chrysaora fuscescens</i> (Brandt, 1835)
–	Not as above.....	131
131 (130)	Exumbrella reddish-brown or purplish-pink with 16 broad, darker radial bands and numerous light spots	<i>Chrysaora plocamia</i> (Lesson, 1830)
–	Not as above.....	132
132 (131)	Oral arms extremely large with frilly margins, hardly coiled to form a dense mass	<i>Chrysaora achlyos</i> (Martin, Gershwin, Burnett, Cargo & Bloom, 1997)
–	Oral arms linear, with broad frilly margins, more or less coiled around central body.....	133
133 (132)	Exumbrella golden-brown, with darker margins, sometimes with 16-32 lighter radial stripes	<i>Chrysaora fuscescens</i> (Brandt, 1835)
–	Not as above.....	134
134 (133)	Stomach pouches all-alike.....	<i>Chrysaora hysoscella</i> (Linnaeus, 1766)
–	Stomach pouches unequal, tentacular ones slightly broader proximally and distally than rhopalar ones.....	<i>Chrysaora melanaster</i> (Brandt, 1838)
135 (88)	Umbrella with papillar knobs	136
–	Umbrella without papillar knobs	138
136 (135)	Oral arms without filaments	<i>Lobonemoides sewelli</i> Rao, 1931
–	Oral arms with filaments	137
137 (136)	Intracircular anastomosing network in communication with the inter-rhopalar canals.....	<i>Lobonema smithii</i> Mayer, 1910
–	Intracircular anastomosing network not in communication with the inter-rhopalar canals	<i>Lobonemoides robustus</i> Stiasny, 1920

138 (135)	Oral arms dichotomous	139
–	Oral arms three-winged	159
139 (138)	Medusa with 4 completely separated subgenital cavities.....	140
–	Medusa with 4 not completely separated subgenital cavities	147
140 (139)	Oral arms 3/4 the length of bell radius.....	<i>Cassiopea frondosa</i> (Pallas, 1774)
–	Not as above.....	141
141 (140)	Oral arms cylindrical, slender, somewhat longer than bell radius	
	<i>Cassiopea ornata</i> Haeckel, 1880
–	Not as above.....	142
142 (141)	Oral arms very large, flat, with 6-8 short, wide-spreading main branches ...	
	<i>Cassiopea depressa</i> Haeckel, 1880
–	Not as above.....	143
143 (142)	Oral arms 1 1/4 times the length of bell radius, triangular in cross-section, aboral surface broad and flat, with 10–15 alternate primary branches.....	
	<i>Cassiopea xamachana</i> Bigelow, 1892
–	Not as above.....	144
144 (143)	Oral arms wide, flat, with 4–6 flat, short tree-shaped side branches	
	<i>Cassiopea andromeda</i> (Forskål, 1775)
–	Not as above.....	145
145 (144)	Oral arms with numerous small lateral branches in their proximal portion.....	<i>Cassiopea medusa</i> Light, 1914
–	Oral arms cylindrical, 1 1/2 times as long as bell radius, branched tree-like	146
146 (145)	Species with numerous large club-shaped vesicles	
	<i>Cassiopea mertensi</i> Brandt, 1838
–	Species without ribbon-like filaments	
	<i>Cassiopea ndrosia</i> Agassiz & Mayer, 1899
147 (139)	Oral arms without filaments	148
–	Oral arms with filaments	150
148 (147)	Exumbrella without a central raised dome.....	
	<i>Marivagia stellata</i> (Galil & Gershwin, 2010)
–	Exumbrella with a central raised dome	149
149 (148)	More than 1 cupolar warts.....	
	<i>Netrostoma dumokuroa</i> (Agassiz & Mayer, 1899)
–	1 cupolar wart	<i>Netrostoma nuda</i> (Gershwin & Zeidler, 2008)
150 (147)	In each octant 3 radial canals	151
–	In each octant more than 3 radial canals.....	153
151 (150)	Between the mouths two kinds of appendages	
	<i>Netrostoma coeruleescens</i> Maas, 1903
–	Between the mouths numerous appendages.....	152
152 (151)	Exumbrella with a central raised dome	
	<i>Netrostoma setouchianum</i> (Kishinouye, 1902)

–	Exumbrella without a central rised dome.....
	<i>Cephea octostyla</i> (Forskål, 1775)
153 (150)	Exumbrella without a central rised dome.....
	<i>Polyrhiza vesiculosa</i> (Agassiz, 1862)
–	Exumbrella with a central rised dome.....	154
154 (153)	Medusa with warts on the central portion of the exumbrella.....	155
–	Medusa without warts on the central portion of the exumbrella.....	156
155 (154)	Radial canals 5–6 per ottante.....	<i>Cephea cephea</i> (Forskål, 1775)
–	Radial canals 7 per ottante.....	<i>Cephea coerulea</i> Vanhöffen, 1902
156 (154)	In each octant 4–6 radial canals....	<i>Cotylorhiza erythraea</i> Stiasny, 1920
–	Not as above.....	157
157 (156)	In each octant 7–9 radial canals....	<i>Cotylorhiza tuberculata</i> (Macri, 1778)
–	In each octant more than 11 radial canals.....	158
158 (157)	Radial canals 11–13 per ottante.....	<i>Cotylorhiza ambulacrata</i> Haeckel, 1880
–	Radial canals 16–17 per ottante.....	<i>Cotylorhiza pacifica</i> (Mayer, 1915)
159 (138)	Oral arms triangular.....	160
–	Oral arms not triangular.....	162
160 (159)	Oral arms terminate in a short, oval knob.....
	<i>Thysanostoma loriferum</i> (Ehrenberg, 1835)
–	Not as above.....	161
161 (160)	Oral arms terminate in a long, tapering filament.....
	<i>Thysanostoma flagellatum</i> (Haeckel, 1880)
–	Oral arms without a terminal portion.....
	<i>Thysanostoma thysanura</i> Haeckel, 1880
162 (159)	Oral arms not pyramidal.....	163
–	Oral arms pyramidal.....	181
163 (162)	Oral arms broad.....	164
–	Oral arms of normal width.....	169
164 (163)	Oral arms without filaments.....	165
–	Oral arms with filaments.....	166
165 (164)	Oral arms without terminal clubs....	<i>Lychnorhiza malayensis</i> Stiasny, 1920
–	Oral arms with terminal clubs....	<i>Pseudorhiza aurosa</i> von Lendenfeld, 1882
166 (164)	Oral arms with terminal clubs.....	167
–	Oral arms without terminal clubs.....	168
167 (166)	Medusa without a single filament at the distal end of one of the oral arms.....	<i>Anomalorhiza shawi</i> Light, 1921
–	Medusa with a single filament at the distal end of one of the oral arms...	<i>Pseudorhiza haeckeli</i> Haacke, 1884
168 (166)	In each octant 8 velar lappets.....	<i>Lychnorhiza arubae</i> Stiasny, 1920
–	In each octant 4 velar lappets.....	<i>Lychnorhiza lucerna</i> Haeckel, 1880
169 (163)	Oral arms coalesced throughout their entire length.....	170
–	Oral arms coalesced in proximal portion only.....	171

170 (169)	Velar lappets about 14 in each octant.....	<i>Stomolophus meleagris</i> Agassiz, 1862
–	Velar lappets about 24 in each octant.....	<i>Stomolophus fritillaria</i> Haeckel, 1880
171 (169)	Oral arms with filaments	172
–	Oral arms without filaments	175
172 (171)	Umbrella more than 100 cm wide	<i>Nemopilema nomurai</i> (Kishinouye, 1922)
–	Umbrella less than 100 cm wide	173
173 (172)	Velar lappets 14–20 in each octant	<i>Rhopilema esculentum</i> Kishinouye, 1891
–	Velar lappets 8 in each octant	174
174 (173)	Exumbrella with sharply conical warts	<i>Rhopilema hispidum</i> (Vanhöffen, 1888)
–	Exumbrella with blunt tuberculation	<i>Rhopilema nomadica</i> Galil, Spanier & Ferguson, 1990
175 (171)	Oral arms with clubs	176
–	Oral arms without clubs	177
176 (175)	Velar lappets 14–20 in each octant	<i>Rhopilema rhopalophorum</i> Haeckel, 1880
–	Velar lappets 6 in each octant	<i>Rhopilema verrilli</i> (Fewkes, 1887)
177 (175)	Oral arms without terminal clubs	178
–	Oral arms with terminal clubs	179
178 (177)	Umbrella ca. 150 mm wide; marginal lobes rectangular in shape	<i>Eupilema scapulare</i> Haeckel, 1880
–	Umbrella ca. 400 mm wide; marginal lobes triangular in shape	<i>Eupilema inexpectata</i> (Pages, Gili & Bouillon, 1992)
179 (177)	Proximal portion of oral arms considerably longer than distal portion	<i>Rhizostoma luteum</i> (Quoy & Gaimard, 1827)
–	Proximal portion of oral arms about as long as distal portion.....	180
180 (179)	Taxon present in the Mediterranean and in the Atlantic Ocean	<i>Rhizostoma pulmo</i> (Macri, 1778)
–	Taxon present in the North Sea only.....	<i>Rhizostoma octopus</i> (Macri, 1778)
181 (162)	Oral arms shorter than usual.....	182
–	Oral arms of normal length	195
182 (181)	Oral arms without terminal appendages	183
–	Oral arms with terminal appendages.....	184
183 (182)	Medusa with rhopalar canals with anastomoses throughout thier length	<i>Mastigietta palmipes</i> (Haeckel, 1880)
–	Medusa with perradial rhopalar canals without anastomoses, interrarial canals with anastomoses.....	<i>Versuriga anadyomene</i> (Maas, 1903)
184 (182)	Intracircular mesh-work of canals never communicating with the rhopalar canals	185

–	Intracircular mesh-work of canals usually communicating with the rhopalar canals.....	187
185 (184)	Terminal appendages nearly as long as the oral arms.....	
 <i>Phyllorhiza pacifica</i> (Light, 1921)	
–	Terminal appendages very long, with distal expansion.....	186
186 (185)	Oral filaments without a triple heart-shaped knob; bell diameter far larger than 25 cm.....	
 <i>Phyllorhiza punctata</i> (von Lendenfeld, 1884)	
–	Oral filaments with a triple heart-shaped knob; bell of ca. 25 cm of diameter.....	
 <i>Phyllorhiza peronlesueuri</i> (Goy, 1990)	
187 (184)	Mouth arms twice as long as disk radius.....	188
–	Not as above.....	189
188 (187)	In each octant more than 10 canal-roots.....	
 <i>Mastigias pantherinus</i> Haeckel, 1880	
–	In each octant up to 10 canal-roots.....	
 <i>Mastigias siderea</i> Chun, 1896	
189 (187)	Mouth arms shorter than disk radius.....	190
–	Mouth arms long as disk radius.....	192
190 (189)	In each octant more than 10 canal-roots.....	
 <i>Mastigias ocellatus</i> (Modeer, 1791)	
–	In each octant up to 10 canal-roots.....	191
191 (190)	Vaulted bell, thin at margin but very thick at apex.....	
 <i>Mastigias gracilis</i> (Vanhöffen, 1888)	
–	Doubtful species, flat and hat-shaped bell, average size unknown.....	
 <i>Mastigias roseus</i> (Reynaud, 1830)	
192 (189)	In each octant up to 10 canal-roots.....	193
–	In each octant more than 10 canal-roots.....	194
193 (192)	Umbrella not flat.....	
 <i>Mastigias papua</i> (Lesson, 1830)	
–	Umbrella flat, disk-shaped.....	
 <i>Phyllorhiza luzoni</i> Mayer, 1915	
194 (192)	Perradial rhopalar canals not bottle-shaped.....	
 <i>Mastigias albipunctatus</i> Stiasny, 1920	
–	Perradial rhopalar canals bottle-shaped.....	
 <i>Mastigias andersoni</i> Stiasny, 1926	
195 (181)	Oral arms with filaments.....	196
–	Oral arms without filaments.....	199
196 (195)	Intracircular anastomosing network not in communication with the rhopalar canals.....	197
–	Intracircular anastomosing network in communication with the rhopalar canals.....	198
197 (196)	Distal three-winged portion of oral arms about twice as long as proximal simple portion.....	
 <i>Crambione bartschi</i> (Mayer, 1910)	
–	Distal three-winged portion of oral arms as long as proximal simple portion.....	
 <i>Crambione mastigophora</i> Maas, 1903	
198 (196)	Oral arms narrow with short filaments.....	
 <i>Acromitus flagellatus</i> (Maas, 1903)	

–	Oral arms thick and broad with long filaments.....
	<i>Acromitus maculosus</i> Light, 1914
199 (195)	Oral arms with terminal clubs	200
–	Oral arms without terminal clubs	202
200 (199)	In each octant 10 velar lappets.....
	<i>Leptobrachia leptopus</i> (Chamisso & Eysenhardt, 1821)
–	Not as above.....	201
201 (200)	In each octant 16 velar lappets <i>Crambionella orsini</i> (Vanhoffen, 1888)	
–	In each octant 12 velar lappets.....	<i>Crambionella stublmanni</i> (Chun, 1896)
202 (199)	Intracircular anastomosing network not in communication with the rhopalar canals	203
–	Intracircular anastomosing network in communication with the rhopalar canals.....	204
203 (202)	In each octant 4 cleft velar lappets
	<i>Acromitoides purpurus</i> (Mayer, 1910)
–	In each octant at least 5 cleft velar lappets.....
	<i>Acromitoides stipbropterus</i> (Schultze, 1897)
204 (202)	Distal three-winged portion of oral arms 1/6 as long as proximal simple portion	<i>Catostylus mosaicus</i> (Quoy & Gaimard, 1824)
–	Not as above.....	205
205 (204)	Distal three-winged portion of oral arms 6 times as long as proximal simple portion.....	<i>Catostylus perezii</i> Ranson, 1945
–	Not as above.....	206
206 (205)	Distal three-winged portion of oral arms 5 times as long as proximal simple portion.....	<i>Catostylus viridescens</i> (Chun, 1896)
–	Not as above.....	207
207 (206)	Distal three-winged portion of oral arms half as long as proximal simple portion	<i>Catostylus tripterus</i> (Haeckel, 1880)
–	Not as above.....	208
208 (207)	Distal three-winged portion of oral arms as long as proximal simple portion.....	<i>Catostylus ornatellus</i> (Vanhöffen, 1888)
–	Distal three-winged portion of oral arms 2–4 times as long as proximal simple portion	209
209 (208)	Oral arms 2/3 the length of bell diameter
	<i>Catostylus townsendi</i> Mayer, 1915
–	Not as above.....	210
210 (209)	Oral arms 1–1,5 times the length of bell radius
	<i>Catostylus cruciatus</i> (Lesson, 1830)
–	Oral arms as long as bell diameter.....	<i>Catostylus tagi</i> (Haeckel, 1869)

Discussion

Digital resources on biodiversity can be relevant not only to researchers, but also to laypeople, such as tourists or citizen scientists. The importance of involving citizens in understanding, monitoring and protecting biodiversity has been recently expressed by the European Commission, in the document “Establishing Horizon 2020” (EU Regulation no. 1291/2013). However, most of the biodiversity-related resources available in the Web – especially the ones dedicated to “difficult” groups, such as jellyfish – are normally devoted almost exclusively to experts (Martellos and Nimis 2015). Exposing scientific information in a form which can be accessible to everybody – without losing its content and informative value – can be a true revolution. Many citizens, especially if already interested in nature and aware of environmental issues (e.g. the presence of invasive alien species), are potentially interested in similar resources. Hence digital resources can be used to involve a wider amount of citizens in scientific tasks, such as the collection of those “big data” which are nowadays fundamental to researchers. The examples of OPAL initiative in the British Isles (<http://www.opalexplornature.org>; accessed 08 August 2015) or, in the field of jellyfish, of MeteoMedusa (Boero 2013, Boero et al. 2013), and JellyWatch (<http://www.jellywatch.org/>; accessed 08 August 2015) are demonstrating the effectiveness of a citizen science approach in collecting scientific data.

JellyWeb is based on morpho-anatomic and taxonomic data, collected and organized in ca. 10 years of research. The development of the portal (Martellos and Nimis 2015) was based upon the experience of the European project KeyToNature (mainly devoted to digital identification) and of the project *Dryades* (devoted to the publication of biodiversity data in the web). This is the first portal devoted to organisms other than vascular plants developed by the research unit of the Dept. of Life Science of the University of Trieste. During its development, a particular attention was paid to user interfaces, in order to provide high quality scientific information in the most straightforward way, and to make it useable by the wider audience as possible.

The multi-entry interface can be useful to both researchers (whom can simply type the name of a taxon to retrieve related information or generate an identification key), and laypeople (whom can use it to start the identification of a jellyfish they have just seen on the seashore). As a further help, interactive keys are enriched by images and drawings of the most relevant characters. Since digital keys are generated in real time, on the basis of the list of remaining organisms, each query produces a different identification key.

Since identification is nowadays often based on molecular analysis, the system has been developed to host molecular data as well. In fact, several attempts to revise the taxonomy of the various taxa like the *Discomedusae* on the basis of morphological observations integrated with genetic analysis are underway, highlighting several critical points, such as the recognition of cryptic species in the *Aurelia* complex within the “traditional” species *Aurelia aurita* (Dawson and Jacobs 2001, Dawson and Martin 2001, Dawson 2003, Dawson et al. 2005, Ramšak et al. 2012), or even at higher taxonomic levels like the proposition of at least two new families within the *Semaeostomeae*

(Bayha and Dawson 2010, Strahler-Pohl et al. 2011). The integration of molecular information in a digital identification system by using the FRIDA software was studied by Bruni et al. (2012) for vascular plants.

Conclusion

JellyWeb is an accumulative system, which can potentially host all data on Scyphozoa, Cubozoa and Staurozoa, and even extend its aim to other groups of the phylum Cnidaria. However, a research group alone can hardly complete such a challenging task. The research unit at the University of Trieste plans to maintain and enrich JellyWeb, but its growth could be faster, if other research groups join this effort. A researcher, or a research group, can contribute to the system by:

- *Fostering a taxon* (such as a genus, or a family). This can be done by managing an instance of the FRIDA system. FRIDA allows to different authors to independently manage separate instances, while at the same time contributing to the same database of morphological and anatomical data, hence, generating updated multi-authored keys to any subset of taxa in the whole system (for a complete description see Martellos 2010). All the digital keys which are generated by the system give credit to the authors of all the data. The keys and all the data and images in JellyWeb are always distributed under a Creative Commons share alike, by attribution 3.0 license (CC 3.0 by-sa).
- *Contributing to the image archive*. High quality images of morphological and anatomical characters and of the whole organisms are probably the most relevant bottlenecks in the process of creating a portal such as JellyWeb. Especially when identifying a taxon, digital images are of capital relevance, both for choosing among the leads of each choice, and as visual census when an identification has been achieved. Several species of Scyphozoa, Cubozoa and Staurozoa are known for one or few specimens, and, even when the taxa are well known, high quality images are, however, scarce. JellyWeb was developed to host a virtually unlimited number of images for each taxon. Each image is displayed with credits to the author(s) and owner(s), institution(s), other metadata, and license.
- *Producing descriptions*. Another relevant bottleneck in developing digital identification keys and portals to one or more groups of organisms are their descriptions. While taxonomic descriptions can be found in books and papers, descriptions which could be actually useful to people other than researchers are difficult to produce. In our experience, to be appreciated by a wider audience, they should mix different sources of information, from ecology to taxonomy, from distribution to human uses, relevance for economy, etymology of the name, etc. Hence, their production is not a simple cut and paste, but a relevant effort of analysis and synthesis.

Potential contributor can contact Massimo Avian (avian@units.it), to define the extent of their participation.

Acknowledgements

The authors are grateful to all the researchers and students (Balboni, Benci, Coral, Sarto, Savonitto, Sola) who contributed to this study. We are also grateful to Dr. Rodolfo Riccamboni, who developed the code of the multi-entry interface of the portal, and to Prof. Pierluigi Nimis, for his suggestions and comments. We are also grateful to Mrs. Sara Triulzi for English language check.

References

- Balboni G (2008) Medusaweb: progettazione e realizzazione di un database e di strumenti digitali interattivi per l'identificazione di Scyphozoa e Cubozoa. Master Thesis, University of Trieste, Trieste.
- Bayha KM, Dawson MN (2010) New family of allomorphic jellyfishes, Drymonematidae (Scyphozoa, Discomedusae), emphasizes evolution in functional morphology and trophic ecology of gelatinous zooplankton. *Biological Bulletin* 219: 249–267.
- Benci E (2008) Progettazione e sviluppo di un database consultabile in rete su Scyphozoa e Cubozoa. Undergr. Thesis, University of Trieste, Trieste.
- Benci C (2012) Analisi critica ed aggiornamento del database Medusaweb. Scyphozoa: ordine Kolpophorae, famiglia Mastigiidae. Undergr. Thesis, University of Trieste, Trieste.
- Berendsohn WG, Chavan V, Macklin JA (2010) Recommendations of the GBIF task group on the global strategy and action plan for the mobilisation of natural history collections data. *Journal of Biodiversity Informatics* 7: 67–71.
- Boero F (2013) Review of jellyfish blooms in the Mediterranean and Black Sea. *GFCM Studies and Reviews* 92: 1–53.
- Boero F, Belmonte G, Bracale R, Frascetti S, Piraino S, Zampardi S (2013) A salp bloom (*Tunicata*, *Thaliacea*) along the Apulian coast and in the Otranto Channel between March–May 2013. *F1000Research* 2: 181. doi: 10.12688/f1000research.2-181.v1
- Brosse S, Beauchard O, Blanchet S, Dürr HH, Grenouillet G, Hugueny B, Lauzeral C, Leprieur F, Tedesco PA, Villéger S, Oberdorff T (2013) Fish-SPRICH: a database of freshwater fish species richness throughout the World. *Hydrobiologia* 700(1): 343–349. doi: 10.1007/s10750-012-1242-6
- Bruni I, De Mattia F, Martellos S, Galimberti A, Savadori P, Casiraghi M, Nimis PL, Labra M (2012) DNA Barcoding as an Effective Tool in Improving a Digital Plant Identification System: A Case Study for the Area of Mt. Valerio, Trieste (NE Italy). *PLoS ONE* 7(9): e43256. doi: 10.1371/journal.pone.0043256
- Calder DR, King RA (2008) An illustrated key to the Scyphozoa and Cubozoa of the South Atlantic Bight. Southeastern Regional Taxonomic Center, South Carolina Department of Natural Resources, Charleston, South Carolina, 18 pp. http://www.dnr.sc.gov/marine/sertc/SAB_jellies_key.pdf [accessed 12 April 2014]
- Calder DR (2009) Cubozoan and scyphozoan jellyfishes of the Carolinian Biogeographic Province, southeastern USA. *Royal Ontario Museum Contributions in Science* 3: 1–58.

- Carvajal-Vallejos FM, Bigorne R, Zeballos Fernández AJ, Sarmiento J, Barrera S, Yunoki T, Pouilly M, Zubieta J, Barra E, Jegú M, Maldonado M, Damme P, Céspedes R, Oberdorff T (2014) Fish-AMAZBOL: a database on freshwater fishes of the Bolivian Amazon. *Hydrobiologia* 732(1): 19–27. doi: 10.1007/s10750-014-1841-5
- Coral C (2012) Analisi critica ed aggiornamento del database “Medusaweb”. “Scyphozoa – Discomedusae – Rhizostomeae – Kolpophorae – Cepheidae”. Undergr. Thesis, University of Trieste, Trieste.
- Cornelius PFS (1997) Keys to the genera of Cubomedusae and Scyphomedusae (Cnidaria). In: den Hartog JC (Ed.) Proceedings of the 6th International Conference on Coelenterate Biology, 1995. Nationaal Natuurhistorisch Museum, Leiden, 109–122.
- Dallwitz MJ (1980) A general system for coding taxonomic descriptions. *Taxon* 29: 41–6. doi: 10.2307/1219595
- Dawson MN (2003) Macro-morphological variation among cryptic species of the moon jellyfish, *Aurelia* (Cnidaria: Scyphozoa). *Marine Biology* 143: 369–379. doi: 10.1007/s00227-003-1070-3
- Dawson MN, Jacobs DK (2001) Molecular evidence for cryptic species of *Aurelia aurita* (Cnidaria, Scyphozoa). *Biological Bulletin* 200: 92–96. doi: 10.2307/1543089
- Dawson MN, Martin LE (2001) Geographic variation and ecological adaptation in *Aurelia* (Scyphozoa: Semaestomeae): some implications from molecular phylogenetics. *Hydrobiologia* 451: 259–273. doi: 10.1023/A:1011869215330
- Dawson MN, Gupta AS, England MH (2005) Coupled biophysical global ocean model and molecular genetic analyses identify multiple introductions of cryptogenic species. *Proceedings of the National Academy of Sciences of the United States of America* 102: 11968–11973. doi: 10.1073/pnas.0503811102
- Fegraus EH, Andelman S, Jones MB, Schildhauer M (2005) Maximizing the value of ecological data with structured metadata: an introduction to ecological metadata language (EML) and principles for metadata creation. *Bulletin of the Ecological Society of America* 86(3): 158–168. doi: 10.1890/0012-9623(2005)86[158:MTVOED]2.0.CO;2
- Field D, Amaral-Zettler L, Cochrane G, Cole JR, Dawyndt P, Garrity GM, Gilbert J, Glöckner FO, Hirschman L, Karsch-Mizrachi J, Klenk HP, Knight R, Kottmann R, Kyrpidis N, Meyer F, San Gil I, Sansone SA, Schriml LM, Sterk P, Tatusova T, Ussery DW, White O, Wooley J (2011) The Genomic Standards Consortium. *PLoS Biol* 9: 6. doi: 10.1371/journal.pbio.1001088
- GBIF (2010) Best Practices in Publishing Species Checklists, 10 pp. http://www.gbif.org/orc/?doc_id=2869 [accessed 12 April 2014]
- Hagedorn G, Rambold G, Martellos S (2010) Types of identification keys. In: Nimis PL, Vignes Lebbe R (Eds) *Tools for Identifying Biodiversity: Progress and Problems*, 59–64.
- Holetschek J, Dröge G, Güntsch A, Berendsohn WG (2012) The ABCD of rich data access to Natural History Collections. *Plant Biosystems* 146(4): 771–779. doi: 10.1080/11263504.2012.740085
- King N, Krishtalka L, Chavan V (2010) Thoughts on implementation of the recommendations of the GBIF task group on a global strategy and action plan for mobilisation of natural history collections data. *Journal of Biodiversity Informatics* 7: 72–76. doi: 10.17161/bi.v7i2.4019

- Martellos S (2010) Multi-authored interactive identification keys: The FRIDA (FRiendly IDentificAtion) package. *Taxon* 59: 922–929.
- Martellos S (2012) From a textual checklist to an information system: The case study of ITALIC, the Information System on Italian Lichens. *Plant Biosystems* 146: 764–770. doi: 10.1080/11263504.2012.740088
- Martellos S (2014) A federated database of taxon pages in the Italian Biodiversity Network. *Plant Biosystems*. doi: 10.1080/11263504.2014.988191
- Martellos S, Attorre F (2012) New Trends in Biodiversity Informatics. *Plant Biosystems* 146(4): 749–751.
- Martellos S, Nimis PL (2015) From Local Checklists to Online Identification Portals: A case study on vascular plants. *PLoS ONE* 10(3): e0120970. doi: 10.1371/journal.pone.0120970
- Mills CE (1999) Stauromedusae: list of all valid species names. <http://faculty.washington.edu/cemills/Staurolist.html> [accessed 12 April 2015]
- Morandini AC, Ascher D, Stampar SN, Ferreira JFV (2005) Cubozoa e Scyphozoa (Cnidaria: Medusozoa) de águas costeiras do Brasil. *Iheringia (Série Zoologia)* 95: 281–294.
- Nimis PL, Martellos S (2009) Computer-aided tools for identifying organisms and their importance for protected areas. *Journal on Protected Mountain Areas Research* 1: 55–60.
- Nimis PL, Riccamboni R, Martellos S (2012) Identification keys on mobile devices: the *Dryades* experience. *Plant Biosystems* 146: 783–788. doi: 10.1080/11263504.2012.740089
- Ramšak A, Stopar K, Malej A (2012) Comparative phylogeography of meroplanktonic species, *Aurelia* spp. and *Rhizostoma pulmo* (Cnidaria: Scyphozoa) in European Seas. *Hydrobiologia* 690: 69–80. doi: 10.1080/11263504.2012.740092
- Randlane T, Saag A, Martellos S, Nimis PL (2010) Computer-aided, interactive keys to lichens in the EU project KeyToNature, and related resources. *Bibliotheca Lichenologica* 105: 37–42.
- Rodriguez C, Marques AC, Stampar SN, Morandini AC, Christiansen E, Genzano G, Mianzan HW (2011) The taxonomic position of the pelagic ‘staurozoan’ *Tessera gemmaria* as a ceriantharian larva. *Zootaxa* 2971: 49–58.
- Sarto R (2009) Analisi critica ed aggiornamento del database Medusaweb. Cubozoa: Carybdeidae e Chirodropidae. Undergr. Thesis, University of Trieste, Trieste.
- Savonitto M (2012) Analisi critica e aggiornamento del database Medusaweb. Scyphozoa: ordine Daktyliophorae, famiglia Rhizostomatidae e Stomolophidae. Undergr. Thesis, University of Trieste, Trieste.
- Sola M (2009) Sviluppo della chiave digitale interattiva Medusaweb: Ord. Semaestomeae. Master thesis. University of Trieste, Trieste, Italy.
- Straehler-Pohl L, Widmer CD, Morandini A (2011) Characterizations of juvenile stages of some semaeostome Scyphozoa (Cnidaria), with recognition of a new family (Phacellophoridae). *Zootaxa* 2741: 1–37.
- Ukosich L (2014) Una panoramica su un portale interattivo di Scyphozoa e Cubozoa. Undergr. thesis. Thesis, University of Trieste, Trieste.
- Wieczorek J, Bloom D, Guralnick R, Blum S, Döring M, Giovanni R, Robertson T, Vieglais D (2012) Darwin Core: An Evolving Community-Developed Biodiversity Data Standard. *PLoS ONE* 7(1): e29715. doi: 10.1371/journal.pone.0029715