

University of Parma Research Repository

Antidepressant-like effects of pharmacological inhibition of FAAH activity in socially isolated female rats

This is the peer reviewd version of the followng article:

Original

Antidepressant-like effects of pharmacological inhibition of FAAH activity in socially isolated female rats / Carnevali, L.; Statello, R.; Vacondio, F.; Ferlenghi, F.; Spadoni, G.; Rivara, S.; Mor, M.; Sgoifo, A.. - In: EUROPEAN NEUROPSYCHOPHARMACOLOGY. - ISSN 0924-977X. - 32:(2020), pp. 77-87. [10.1016/j.euroneuro.2019.12.119]

Availability: This version is available at: 11381/2884825 since: 2020-12-11T10:02:44Z

*Publisher:* Elsevier B.V.

Published DOI:10.1016/j.euroneuro.2019.12.119

Terms of use: openAccess

Anyone can freely access the full text of works made available as "Open Access". Works made available

Publisher copyright

(Article begins on next page)

1	Antidepressant-like effects of pharmacological inhibition of FAAH activity in socially					
2	isolated female rats					
3						
4	Luca Carnevali <sup>a</sup> , Rosario Statello <sup>a</sup> , Federica Vacondio <sup>b</sup> , Francesca Ferlenghi <sup>b</sup> , Gilberto Spadoni <sup>c</sup> ,					
5	Silvia Rivara <sup>b</sup> , Marco Mor <sup>b</sup> , Andrea Sgoifo <sup>a</sup>					
6						
7	<sup>a</sup> Stress Physiology Lab, Department of Chemistry, Life Sciences and Environmental Sustainability,					
8	University of Parma, Parma, Italy					
9	<sup>b</sup> Department of Food and Drug, University of Parma, Parma, Italy					
10	<sup>c</sup> Department of Biomolecular Sciences, University of Urbino "Carlo Bo", Urbino, Italy					
11						
12	Corresponding author: Andrea Sgoifo, Stress Physiology Lab, Department of Chemistry, Life					
13	Sciences and Environmental Sustainability, University of Parma, Parco Area delle Scienze 11/A,					
14	43124, Parma, Italy. Email address: andrea.sgoifo@unipr.it					
15	Total word-count: 5636					

# 17 Abstract

Pharmacological inhibition of the enzyme fatty acid amide hydrolase (FAAH), which terminates 18 signalling of the endocannabinoid N-arachidonoylethanolamine (or anandamide, AEA), exerts 19 20 favourable effects in rodent models of stress-related depression. Yet although depression seems to 21 be more common among women than men and in spite of some evidence of sex differences in 22 treatment efficacy, preclinical development of FAAH inhibitors for the pharmacotherapy of stressrelated depression has been predominantly conducted in male animals. Here, adult female rats were 23 exposed to six weeks of social isolation and, starting from the second week, treated with the FAAH 24 25 inhibitor URB694 (0.3 mg/kg/day, i.p.) or vehicle. Compared to pair-housed females, socially isolated female rats treated with vehicle developed behavioral (mild anhedonia, passive stress coping) and 26 physiological (reduced body weight gain, elevated plasma corticosterone levels) alterations. 27 Moreover, prolonged social isolation provoked a reduction in brain-derived neurotrophic factor 28 29 (BDNF) and AEA levels within the hippocampus. Together, these changes are indicative of an increased risk of developing a depressive-like state. Conversely, pharmacological inhibition of FAAH 30 activity with URB694 restored both AEA and BDNF levels within the hippocampus of socially isolated 31 rats and prevented the development of behavioral and physiological alterations. These results 32 33 suggest a potential interplay between AEA-mediated signaling and hippocampal BDNF in the pathogenesis of depression-relevant behaviors and physiological alterations and antidepressant 34 action of FAAH inhibition in socially isolated female rats. 35

36 Keywords: depression; stress; endocannabinoid; BDNF; females

37

# 39 **1. Introduction**

40 Prolonged or repeated exposure to stressors of psychosocial nature can act as a precipitating factor for the onset of depression (Cohen et al., 2007; Dinan, 2005). One of the most susceptible brain 41 regions to the effects of psychosocial stress is the hippocampus, a component of the limbic system 42 43 that regulates emotional and cognitive processes related to psychiatric disorders (Belleau et al., 44 2019; Sheline et al., 2019). The hippocampus is also a major regulator of the hypothalamic-pituitary-45 adrenal (HPA) axis (Jacobson and Sapolsky, 1991), the neuroendocrine system responsible for the release of glucocorticoid stress hormones (i.e., cortisol in humans, corticosterone in rodents). In 46 47 patients with depression, hippocampal volume is decreased (Sapolsky, 2000; Sheline, 1996) and 48 the HPA axis is dysregulated (Stetler and Miller, 2011). Depletion of hippocampal neurogenesis has 49 been implicated as one of the substrates that may explain the hippocampal volume loss seen in depression (Duman and Monteggia, 2006; Levone et al., 2015). Specifically, the neurotrophic 50 hypothesis of depression proposes that stress-induced reductions in the expression of brain-derived 51 52 neurotrophic factor (BDNF), a member of the neurotrophin family regulating synaptic plasticity (Leal et al., 2017; Lu et al., 2014), occur in key limbic structures, including the hippocampus, to contribute 53 to the pathogenesis of depression (Castren et al., 2007; Duman and Monteggia, 2006). Moreover, 54 several lines of clinical and preclinical evidence indicate that conventional antidepressants (e.g., 55 tricyclics, selective serotonin reuptake inhibitors and norepinephrine reuptake inhibitors) may in part 56 57 exert their effects through BDNF upregulation (Hayley and Anisman, 2013; Pittenger and Duman, 2008; Tardito et al., 2006). 58

The past two decades have witnessed a driven focus on the identification of novel therapeutic targets 59 60 for depression, in an attempt to overcome the notable limitations of conventional antidepressant treatments, poor efficacy being perhaps the most critical (Connolly and Thase, 2012). For example, 61 substantial evidence has accumulated implicating a deficit in endocannabinoid (eCB) 62 neurotransmission in the etiology of depression (for a comprehensive review see Gorzalka and Hill, 63 64 2011). At the preclinical level, a deficiency in the signaling mediated by the eCB Narachidonoylethanolamine (or anandamide, AEA) has been noted in the hippocampus, 65 hypothalamus, ventral striatum, and prefrontal cortex of rats exposed to several stressors (i.e., 66

chronic unpredictable stress and social defeat stress) and presenting a "depressive-like" phenotype 67 (reviewed in Carnevali et al., 2017b). These findings have triggered significant interest in the 68 69 development of eCB-interacting drugs, including direct-acting receptor ligands and catabolism 70 inhibitors for the pharmacotherapy of depression (Micale et al., 2013). Specifically, within preclinical models, facilitation of AEA signaling through pharmacological inhibition of its degrading enzyme (i.e., 71 72 fatty acid amide hydrolase (FAAH)) can enhance monoaminergic transmission, increase cellular 73 plasticity and neurotrophin expression within the hippocampus, dampen HPA axis activity, and evoke 74 antidepressant-like behavioral effects (reviewed in Carnevali et al., 2017b). However, while the 75 literature has been unequivocal in showing that women experience depression at twice the rate of men (e.g., Grigoriadis and Robinson, 2007), very few preclinical studies have been conducted on 76 77 female experimental animals (Beery, 2018; Kokras and Dalla, 2014). Moreover, despite the existence of sex differences in response to antidepressant treatment (Sloan and Kornstein, 2003), 78 79 preclinical research on the antidepressant action of FAAH inhibitors has been predominantly conducted in male rodents (Carnevali et al., 2017b; Fowler, 2015). Therefore, there is a clear need 80 81 to use female animals in preclinical models of stress to either confirm and generalize to females the 82 previously obtained male animal-based findings or underscore potential sex differences in the 83 etiology of depression and/or in the efficacy of new treatments.

84 Based on this background, the purpose of the current study was two-fold. First, we aimed at 85 documenting the development of behavioral (passive stress coping, anhedonia) and biological 86 (reduced hippocampal BDNF levels, HPA axis hyperactivity, body weight loss) alterations in adult female rats exposed to prolonged social isolation, a mild chronic social stressor that has been widely 87 used to model symptoms that are often associated with an increased risk of developing a depressive-88 89 like state in rodents (Carnevali et al., 2017a). Second, we tested the hypothesis that pharmacological 90 inhibition of FAAH activity would correct the alterations associated with prolonged social isolation. To this aim, we employed the FAAH inhibitor URB694 (6-hydroxy-[1,1'-biphenyl]-3-yl-91 cyclohexylcarbamate) which was shown to exhibit higher selectivity and more prolonged and 92 profound access to the brain than the standard inhibitor URB597 (Clapper et al., 2009). 93

94

### 95 **2. Experimental procedures**

# 96 2.1. Animals and housing conditions

Four-month-old female wild-type Groningen rats were used in this study. This rat population, 97 originally derived from the University of Groningen (the Netherlands) and currently bred in our 98 99 laboratory under standard conditions, shows considerable individual differences in trait-like patterns 100 of behavioral and physiological responses to environmental challenges (Carnevali et al., 2014; de 101 Boer et al., 2017). After weaning, female animals were housed in same-sex sibling pairs and kept in rooms with controlled temperature (22 ± 2 °C) and humidity (50 ± 10 %), under a reversed light-dark 102 103 cycle (light on from 19:00 to 7:00 h), with food and water ad libitum except when required for the 104 sucrose preference test (see below). A total of 40 pairs were included in the study, but only one 105 female rat from each pair was submitted to the experimental procedures described below. Experiments were performed in accordance with the European Community Council Directive 106 2010/63/UE and approved by the Italian legislation on animal experimentation (D.L. 04/04/2014, n. 107 108 26, authorization n. 449/2017-PR). All efforts were made to reduce sample size and minimize animal suffering. 109

110

## 111 2.2. Experimental design

The experimental timeline is depicted in Figure 1. Specific procedures and data analysis are 112 described in the following sections. On day 0, animals were randomly divided in socially isolated (SI) 113 and paired-housed (PH) groups. Female rats from the SI group were separated from their respective 114 sibling and individually housed in a soundproof room for 6 weeks to avoid any sensory (visual, 115 116 olfactory, and acoustic) contact with their conspecifics. On the contrary, female rats from the PH group were continually housed with their respective sibling and kept in the same room with other 117 pairs. Handling and cage cleaning were matched between the two groups. Starting from the 118 beginning of the third week of the social isolation/pair-housing condition, animals received daily i.p. 119 120 injection of either the FAAH inhibitor URB694 or vehicle (VEH). Thus, four experimental subgroups 121 emerged: (i) SI + VEH (n = 10), (ii) SI + URB694 (n = 10), (iii) PH + VEH (n = 10), and (iv) PH + 122 URB694 (n = 10). Experiments were conducted on separate cohorts of 8 experimental animals each

(n = 4 SI and n = 4 PH rats), starting with the VEH-treated animals. Experimental animals were 123 tested four times in the sucrose preference test and once in the forced swim test during the dark 124 125 phase of the daily cycle between 10.00 and 12.00 h. At sacrifice (day 42), trunk blood, adrenal 126 glands, and hippocampus were harvested. Body weight was measured weekly throughout the study. Moreover, the estrous cycle phase of female rats was determined immediately after each behavioral 127 test and before sacrifice using vaginal smear cytology. Vaginal smears were collected by gently 128 129 introducing a moistened (0.9% NaCl) cotton swab in the rat's vagina. The sample was transferred to 130 a glass slide and examined microscopically following Giemsa staining. The phase of the cycle (metaestrous, diestrous, pro-estrous or estrous) was determined based upon the presence of 131 leukocytes, nucleated epithelial or cornfield epithelial cells (Marcondes et al., 2002). 132

133

# 134 2.3. Drug treatment

URB694 is a carbamate FAAH inhibitor that irreversibly carbamoylates the nucleophile catalytic 135 serine in FAAH active site (Tarzia et al., 2006). URB694 is a second generation inhibitor with 136 137 improved metabolic stability and selectivity for FAAH (Clapper et al., 2009). URB694 was freshly dissolved in VEH containing 5% PEG, 5% Tween 80, and 90% saline. VEH (vol:1 ml/kg) or URB694 138 (0.3 mg/kg, i.p.) were injected i.p. between 11.00 and 13.00 h and, on the days of the sucrose 139 solution and forced swim tests, at least 1 h after the completion of the test. URB694 dose was chosen 140 141 based on our previous studies (Carnevali et al., 2015a; Carnevali et al., 2015b), and a pilot study 142 showing that FAAH activity in the brain of female wild-type Groningen rats was substantially inhibited 143 24 h after administration of this drug dose (Supplemental Figure S1).

144

# 145 2.4. Sucrose preference test

Ad libitum 2% sucrose solution was available for 5 days before the beginning of the experimental procedures to allow adaptation to its taste. Food and water were removed from the cage for 16 hours before each sucrose preference test; moreover, one hour before the test, all experimental animals (paired and isolated) were moved into individual cages to ensure accurate fluid intake measurements of paired animals. Water and 2% sucrose solution were placed in premeasured bottles in the

individual cage, and fluid intake was monitored for 1 hour. Animals were returned to their respective home cages immediately after the test (Grippo et al., 2007). Sucrose preference tests were conducted in baseline conditions (day -3) and after 11, 25, and 39 days of social isolation (Figure 1). Sucrose solution intake was expressed as the relative percentage of the total liquid intake and was taken as an operational index of anhedonia, defined as reduced sucrose preference relative to control animals and baseline values (Grippo et al., 2007).

157

158 2.5. Forced swim test

An adapted version of the forced swim test originally described by Porsolt (Porsolt et al., 1977) was used. On day 35 (Fig. 1), female rats were forced to swim individually for 5 min in a Plexiglas cylinder (height: 40 cm, diameter: 30 cm) filled with water (temperature:  $24 \pm 2^{\circ}$ C; depth: 30 cm). During the test, rats' behavior was video-taped. The overall time spent in immobility (floating and making only those movements necessary to keep the head above water) was scored by a trained experimenter blind to animals' condition and treatment. Immobility during the single session of the forced swim test was used as an index of passive stress coping (Commons et al., 2017).

166

### 167 2.6. Measurements at sacrifice

Twenty-four hours after the last administration of URB694 or VEH (i.e., at 11.00 h; day 42, Figure 1), female rats were euthanized by decapitation under isoflurane anesthesia (2% in 100% oxygen). Trunk blood was collected in EDTA-coated tubes (Sarsted AG, Numbrecht, Germany) and plasma was separated by centrifugation (2600 g, 4°C, 10 min). Brains were immediately removed and the hippocampus rapidly dissected and snap-frozen in nitrogen. All samples were stored at -80°C until further analysis, as described below. Adrenal glands were also removed and weighed.

174 2.6.1. Plasma corticosterone levels

Plasma was deproteinized by addition of two volumes of organic solvent (ice-cold acetonitrile), containing the internal standard dexamethasone (structural analog of corticosterone, 75 nmol/L). After centrifugation (14000 g, 4°C, 10 min), the supernatant was directly injected in the liquid chromatography/tandem mass spectrometry system (HPLC/MS/MS) for quantification of

corticosterone levels, in accordance with previously published analytical methods (Plenis et al.,
2011). A detailed description of the HPLC/MS/MS analytical method and related MS instrumentation
is reported in the Supplemental Material.

182 2.6.2. BDNF hippocampal content

BDNF content in the hippocampus was measured using a commercially available sandwich enzymelinked immune sorbent assay (ELISA) kit (Quantikine ®ELISA-Total BDNF, R&D Systems, Minneapolis, MN, USA) according to the manufacturer's instructions. A detailed description of the experimental procedure is reported in the Supplemental Material. BDNF tissue content was expressed as a percentage of the control group (PH+VEH rats).

188 2.6.3. AEA hippocampal levels

AEA was extracted from 10% w/v hippocampal tissue homogenates employing two volumes of icecold acetonitrile containing the deuterated internal standard AEA-d₄ and quantified by HPLC/MS/MS as previously reported (Carnevali et al., 2015a) The analytical standards AEA and AEA-d₄ were purchased from Cayman Chemical (Ann Arbor, MI, USA) as stock solutions in ethanol. AEA levels were expressed as pmol/g wet weight of tissue. A detailed description of the HPLC/MS/MS analytical method and related MS instrumentation is reported in the Supplemental Material.

195 2.6.4. FAAH activity in the hippocampus

For ex vivo determination of FAAH activity, frozen hippocampi were thawed and homogenized in ice-196 197 cold Tris buffer (10 volumes, 50 mM, pH 7.5) containing 0.32 M sucrose. The homogenates were 198 centrifuged (1000 g, 10 min, 4°C) and total protein content was quantified in the supernatant by the 199 bicinchoninic acid (BCA) protein kit (Pierce Biotechnology, Rockford, IL, USA). FAAH activity was measured at 37°C for 30 min in 0.5 mL Tris buffer (50 mM, pH 7.5) containing fatty acid-free bovine 200 201 serum albumin (BSA) (0.05%, w/v), 50 μg of protein from brain homogenates, 10 μM AEA and [<sup>3</sup>H]-202 AEA (10000 disintegrations per minute) as previously described (Clapper et al., 2009). Briefly, the 203 reactions were stopped with 1 mL chloroform:methanol (1:1). After centrifugation (2000 g, 10 min, 4°C), [<sup>3</sup>H]-ethanolamine was measured in the aqueous phase by liquid scintillation counting. [<sup>3</sup>H]-204 AEA (specific activity: 60 Ci/mmol), employed as a substrate for ex vivo FAAH assay, was purchased 205 from American Radiolabeled Chemicals (St. Louis, MI, USA). 206

# 207 2.7. Statistical analysis

All statistical analyses were performed using SPSS v. 25 (IBM software package). Data are 208 209 presented as mean ± standard error of the mean (SEM). The influence of the estrous cycle phase on behavioral and biochemical measurements was controlled in all statistical analyses. A three-way 210 ANOVA for repeated measures with "condition" (2 levels: isolation, pair-housing) and "treatment" (2 211 levels: VEH, URB694) as the between subject factors, and "time" as the within subject factor (3 212 213 levels: days 11, 25, and 39) was applied on delta changes in sucrose solution preference with respect 214 to baseline. All other data were analyzed with 2 (factor "condition": isolation or pair-housing) x 2 (factor "treatment": URB694 or VEH) factorial design ANOVAs. Follow-up analyses were conducted 215 using Student's "t" tests, with a Bonferroni correction for multiple comparisons. Pearson's r216 correlations were performed to assess the correlation between plasma corticosterone levels, BDNF 217 218 hippocampal content and AEA hippocampal levels. Statistical significance was set at p < 0.05.

219

# 220 **3. Results**

3.1. Body weight

There were no significant differences in body weight among groups at the start of the experiment 222 223 (i.e., when animals were assigned to the different housing conditions) (PH + VEH =  $230 \pm 2$  g; IS +  $VEH = 237 \pm 5 \text{ g}; PH + URB694 = 231 \pm 4 \text{ g}; IS + URB694 = 226 \pm 8 \text{ g}).$  However, a significant time 224 225 x condition interaction emerged on body weight gain calculated as the difference between weight at the end (i.e., immediately before animals were euthanized) and at the start of the experiment (F = 226 7.1, p = .012). As shown in Figure 2, socially isolated female rats treated with VEH gained 227 significantly less weight compared with their respective pair-housed counterparts (p = .002). This 228 effect of social isolation was prevented by URB694 treatment (SI + URB694 vs SI + VEH, p = .012). 229

# 230 3.2. Sucrose preference test

Total fluid intake did not differ among groups at each assessment point (Supplemental Table S1). Also, there were no significant differences among groups in their baseline preference for the consumption of the sucrose solution (PH + VEH =  $85 \pm 2\%$ ; IS + VEH =  $88 \pm 2\%$ ; PH + URB694 =  $83 \pm 3\%$ ; IS + URB694 =  $82 \pm 3\%$ ). Of note, the estrous cycle phase had no effect on baseline

235 sucrose solution preference (F = 0.3, p = .543). However, factorial ANOVA yielded a significant time x condition interaction (F = 5.1, p = .028) on preference changes during the social isolation period 236 237 (calculated as the difference between each assessment point and the baseline), with no significant effects of the estrous cycle phase (F = 0.4, p = .497). Specifically, as shown in Figure 3, no group 238 differences were observed on day 11. However, on day 25, socially-isolated female rats treated with 239 VEH showed a significantly larger reduction in the preference for sucrose solution consumption 240 241 compared with their respective pair-housed counterparts (p = .025). This effect was prevented by 242 URB694 treatment (SI + URB694 vs SI + VEH, p = .003). A similar trend was observed on day 39, although differences did not reach full statistical significance (SI + VEH vs PH + VEH, p = .056; SI + 243 VEH vs SI + URB694, p = .067). 244

245 3.3. Forced swim test

Behavior during the forced swim test is illustrated in Figure 4. Factorial ANOVA yielded a significant effect of treatment (F = 4.9, p = .033) and a strong trend for condition x treatment interaction (F = 3.5, p = .071) on immobility time, with no significant effects of estrous cycle phase (F = 0.2, p = .632). Specifically, socially isolated female rats treated with VEH spent significantly more time in immobility compared with their respective pair-housed counterparts (p = .024). This behavioral effect of social isolation was significantly corrected by URB694 treatment (SI + URB694 vs SI + VEH, p = .007).

#### 252 3.4. Measurements at sacrifice

253 3.4.1. Plasma corticosterone levels and adrenal weight

Factorial ANOVA yielded a significant condition x treatment interaction (F = 7.1, p = .012) on plasma corticosterone levels at the end of the experimental protocol, with no significant effects of the estrous cycle phase (F = 0.6, p = .430). As depicted in Figure 5, socially isolated female rats treated with VEH had significantly higher plasma corticosterone levels than their respective pair-housed counterparts (p = .016). URB694 treatment prevented the effect of social isolation on plasma corticosterone levels (SI + URB694 vs SI + VEH, p = .003).

There were no significant effects of condition and/or treatment on adrenal weight corrected for body weight at the end of the experiment (PH + VEH =  $0.021 \pm 0.002$  mg/g; IS + VEH =  $0.027 \pm 0.003$ mg/g; PH + URB694 =  $0.027 \pm 0.002$  mg/g; IS + URB694 =  $0.026 \pm 0.002$  mg/g).

# 263 3.4.2. BDNF hippocampal content

Factorial ANOVA yielded a significant effect of treatment (F = 7.3, p = .012) and a significant condition x treatment interaction (F = 6.9, p = .014) on BDNF content in the hippocampus at the end of the experimental protocol. As illustrated in Figure 6A, socially isolated female rats treated with VEH showed a significantly lower BDNF hippocampal content compared with their respective pairhoused counterparts (p = .023). This effect of social isolation was prevented by URB694 treatment (SI + URB694 vs SI + VEH, p = .001). Moreover, we found a negative, although not significant, correlation between plasma corticosterone levels and BDNF hippocampal content (Table 1).

# 271 3.4.3. AEA hippocampal levels

Factorial ANOVA yielded significant effects of condition (F = 19.7, p < .001) and treatment (F = 27.6, 272 p < .001), and a significant condition x treatment interaction (F = 5.3, p = .028) on AEA hippocampal 273 levels at the end of the experimental protocol. As shown in Figure 6B, socially isolated female rats 274 treated with VEH showed significantly lower AEA hippocampal levels compared with their respective 275 pair housed counterpart (p < .001). As expected, URB694-treated groups showed significantly 276 277 greater AEA levels than corresponding VEH-treated groups, both in the social isolation (p < .001) 278 and pair-housing (p = .040) condition. Moreover, we found a significant positive correlation between AEA levels and BDNF content within the hippocampus (Table 1), as well as a strong trend for a 279 negative correlation between AEA hippocampal levels and plasma corticosterone levels (Table 1). 280

# 281 *3.4.4. FAAH activity*

Factorial ANOVA yielded a significant effect of treatment (F = 456.0, p < .001) on FAAH activity in the hippocampus, being, as expected, significantly lower in URB694-treated than VEH-treated rats in both the social isolation (p < .001) and pair-housing (p < .001) condition (Figure 6C).

285

### 286 **4. Discussion**

The major findings of the current investigation are the following. Compared to pair-housed females, socially isolated female rats developed behavioral (mild anhedonic state, passive stress coping) and physiological (reduced body weight gain, elevated plasma corticosterone levels) changes, and showed a reduction in BDNF and AEA levels within the hippocampus. Together, these changes are indicative of an increased risk of developing a depressive-like state. Notably, pharmacological
inhibition of FAAH activity with URB694 restored AEA and BDNF hippocampal levels, and prevented
the development of behavioral and physiological alterations following prolonged social isolation.

4.1. Depressive-like changes in socially isolated female rats

Psychiatric disorders in humans have been linked prevalently with social stress and/or reduced 295 social interaction (Bjorkqvist, 2001; Heinrich and Gullone, 2006). Within preclinical models, the social 296 297 defeat paradigm has been shown to have a substantial impact on depression-relevant behavioral 298 and physiological parameters in adult male rats, while solitary housing is particularly effective in 299 precipitating depressive-like symptoms in previously group-housed female rats (Beery and Kaufer, 2015; Carnevali et al., 2017a). Of note, the social isolation protocol adopted in this study included 300 both solitary housing and long-term deprivation of sensory stimuli originating from the surrounding 301 social environment. Therefore, it is likely that the described effects are due to a combination of both. 302 303 Specifically, female rats showed a reduction in body weight gain, signs of a mild anhedonic-like state 304 (i.e., reduced preference for the consumption of a sucrose solution), passive coping (i.e., increased 305 immobility in the forced swim test), and elevated plasma corticosterone levels. Deficits in body weight gain in isolated rats may be explained by reduced food intake, as previously demonstrated in 306 307 individually housed mice and rats (Izadi et al., 2018; Sun et al., 2014), particularly around light-dark 308 phase transitions (Sun et al., 2014). Interestingly, reductions in heat production and in the respiratory 309 exchange ratio were also found during light-dark transitions in individually housed mice (Sun et al., 310 2014), suggesting that metabolic functions may have been affected also in our socially isolated rats. 311 Moreover, the mild reduction in the preference for the consumption of a palatable solution observed only after 25 days of social isolation resembles the time course of changes reported in female Wistar 312 rats exposed to chronic mild stress (Grippo et al., 2005) and in socially isolated female prairie voles 313 314 (Grippo et al., 2007). However, we acknowledge that the interpretation of this result is limited by the 315 difference, albeit not statistically significant, between the two stressed groups on day 11 (i.e., before the start of the pharmacological treatment). Notably, the estrous cycle phase did not seem to have 316 any effect on any of the behavioral and biological variables assessed in the current study, although 317 318 our analysis is limited by the small sample size given that four different stages were considered.

Nevertheless, this is in line with empirical research across multiple rodent species demonstrating that estrous cyclicity is not a major source of variability in females or, at least, is not greater than intrinsic variability in males (Beery, 2018; Finnell et al., 2018; Kokras et al., 2015).

Animal and human studies have provided support for the role of stress in the pathogenesis of 322 depression via alterations in BDNF-mediated signaling (Hashimoto, 2010; Stepanichev et al., 2014), 323 a neurotrophin that primarily regulates synaptic plasticity (Leal et al., 2017; Lu et al., 2014). In line 324 325 with these findings, we found that BDNF content was reduced in the hippocampus of socially isolated 326 female rats with depressive-like symptoms. Remarkably, such downregulation of hippocampal BDNF 327 was paralleled by a decrease in AEA hippocampal levels. Converging lines of evidence support the possibility that AEA signaling at the cannabinoid receptor 1 (CB1R) may be an important mediator 328 of neuroplastic phenomena within the hippocampus (Aguado et al., 2005; Hashimotodani et al., 329 2007; Hill et al., 2010; Scarante et al., 2017; Burstein et al., 2018). Particularly relevant for the current 330 results are findings of decreased BDNF levels in the hippocampus of CB1R knockout mice (Aso et 331 al., 2008). Thus, we hypothesize that a deficiency in AEA-mediated signaling at the CB1R might be 332 333 implicated in the downregulation of BDNF hippocampal content observed in socially isolated female 334 rats. Moreover, the positive correlation found here between AEA levels and BDNF content further 335 supports a role for the eCB system in adult hippocampal neurogenesis (Scarante et al., 2017). Notably, while one study reported a similar decrease in AEA levels in the hippocampus of chronically 336 337 stressed male rats (Hill et al., 2008), other studies showed no changes in AEA hippocampal levels 338 upon chronic stress exposure (Bortolato et al., 2007; Carnevali et al., 2015a; Hill et al., 2005). Of 339 note, our data suggest that reduced AEA levels in the hippocampus of socially isolated rats were not due to an upregulation of FAAH enzymatic activity. This is in line with previous studies showing that 340 FAAH activity is not affected by chronic stress exposure in rats (Bortolato et al., 2007; Hill et al., 341 342 2008), suggesting that the stress-induced decline in the hippocampal pool of AEA might be due to diminished biosynthetic mechanisms. Empirical evidence indicates the eCB system may be a 343 biochemical effector of glucocorticoids in the brain (Hill and McEwen, 2010). Notably, the 344 hippocampus itself is particularly sensitive to the action of glucocorticoid stress hormones due the 345 rich concentration of receptor sites for glucocorticoids (De Kloet et al., 1998). The negative, although 346

347 only marginally significant, correlation found between plasma corticosterone levels and AEA hippocampal levels prompts further investigation into the specific mechanisms underlying the effects 348 349 of stress exposure on AEA metabolism and their causal relationship with BDNF hippocampal downregulation. Interestingly, sex-specific mechanisms of eCB-mediated synaptic modulation within 350 the hippocampus have been proposed to partly explain sex disparities in prevalence of depression 351 (Huang and Woolley, 2012; Tabatadze et al., 2015). Decreased levels of BDNF may contribute to 352 353 the atrophy of the hippocampus that has been observed in patients with depression (Sheline, 1996; 354 Sheline et al., 2019). Recently, Belleau and colleagues (Belleau et al., 2019) proposed a model according to which chronic life stress can trigger the initial development of hippocampal volume 355 reduction. However, this reduction would be neither necessary nor sufficient to produce a major 356 depressive episode (Belleau et al., 2019). On the other hand, stress also initiates a set of neurotoxic 357 processes (HPA axis dysregulation, inflammation, and neurotransmitter disturbances) that interact 358 and may drive the development of a more chronic type of depression marked by further hippocampal 359 volume reduction (Belleau et al., 2019). Although hippocampal volume was not assessed in the 360 361 current study, we speculate that AEA-BDNF interactions might be implicated in the development of 362 depressive symptoms and hippocampal volume decline under chronic life stress. Future longitudinal 363 studies in rodent models of social stress may be informative in this regard.

4.2. Antidepressant-like effects of the FAAH inhibitor URB694

365 In an attempt to replicate findings of our previous study demonstrating antidepressant-like effects of 366 the FAAH inhibitor URB694 in chronically stressed male rats (Carnevali et al., 2015a), pharmacological treatment with URB694 started after two weeks of social isolation (i.e., we 367 anticipated that depressive-like behaviors would already have begun to manifest by then). However, 368 contrary to our expectations, we failed to conclusively demonstrate the onset of an anhedonic-like 369 370 state before the start of the treatment. Thus, the fact that URB694-treated females did not show depressive-like behavioral and biological symptoms after a prolonged period of social isolation 371 suggests, more cautiously, that inhibition of FAAH activity represents an effective preventive 372 measure in this animal model. These results are in line with a growing body of evidence 373 demonstrating that pharmacological inhibition of FAAH activity produces an antidepressant response 374

375 in chronically stressed male rodents (Carnevali et al., 2017b). Interestingly, FAAH inhibitors have been shown to increase hippocampal neurogenesis in adult rats (Goncalves et al., 2008; Hill et al., 376 377 2006; Marchalant et al., 2009) and prevent stress-induced BDNF downregulation in the brain (Burstein et al., 2018), supposedly via facilitation of CB1R-mediated activation of the extracellular 378 signal-regulated kinase signaling pathway (Derkinderen et al., 2003; Rubino et al., 2006). Therefore, 379 given that CB1Rs are highly abundant in the rodent (and human) hippocampus (Mackie, 2005), we 380 381 hypothesize that the antidepressant-like action of the FAAH inhibitor URB694 in socially isolated 382 female rats may be partly mediated by a preservation of hippocampal BDNF content via enhancement of AEA signaling at the CB1R. However, the antidepressant-like effects of URB694 383 may also be interpreted in light of experimental evidence showing that AEA-signaling enhancement 384 at the CB1R facilitates adaptive stress coping behaviors (Haller et al., 2013) and attenuates the 385 neuroendocrine response to psychological stressors (Gorzalka et al., 2008). Moreover, given that 386 387 FAAH inhibitors also increase the levels of other fatty acid amines with activity at peroxisome proliferator activated receptor- $\alpha$  (N-oleoylethanolamine (OEA) and N-palmitoylethanolamine (PEA)), 388 389 the possibility of other non-cannabinoid receptor-mediated mechanisms cannot be completely ruled out. For example, a growing body of preclinical evidence suggests that PEA could have 390 antidepressant-like activity (De Gregorio et al., 2019). On the other hand, increases in the 391 endogenous levels of OEA may reduce food intake by regulating systems that control hunger and 392 393 satiety in the brain (Romano et al., 2015). However, these compounds might also prolong and 394 enhance AEA biological activity by competing with AEA for FAAH-mediated degradation (Petrosino 395 et al., 2009). Of note, the current drug regimen had no effects on control animals, suggesting that the FAAH inhibitor did not affect normal biological processes and behavioral responses. 396

397

# 398 4.3. Conclusion

The results of this study suggest a potential interplay between AEA-mediated signaling and BDNF at the level of the hippocampus in the development of depression-relevant behaviors and physiological changes in female rats exposed to prolonged social isolation. Moreover, the current results document the ability of the FAAH inhibitor URB694 to correct the alterations associated with

prolonged social isolation. One should note, however, that the current results are merely suggestive, 403 and their translational implications for depression should be interpreted within the context of their 404 405 limitations. First, the behavioral and biological changes described here after social isolation are often 406 associated with an increased risk of developing major depression, but are not clinical symptoms of major depression per se. Second, we adopted a rodent model of prolonged social isolation, which 407 should not be intended as a diagnostic model, but rather as a model of risk and vulnerability factor 408 409 of stress-related depression. Moreover, we must acknowledge that, at present, clinical research on 410 FAAH inhibitors has been slowed down by the serious adverse effects caused by the FAAH inhibitor BIA 10-2474 for the treatment of pain (von Schaper, 2016), which displayed both intrinsic toxic 411 effects at high doses and off-targets effects (van Esbroeck et al., 2017). Investigations conducted by 412 a Temporary Specialist Scientific Committee concluded that the toxicity of BIA 10-2474 is unlikely 413 due to FAAH inhibition (Temporary Specialist Scientific Committee, 2016). A communication from 414 the U.S. Food and Drug Administration also reported that the unique toxicity of this drug does not 415 extend to other FAAH inhibitors (Food and Drug Administration, 2016), which are well tolerated by 416 417 patients enrolled in clinical trials, and remarkably lack of the common adverse events elicited by 418 exogenous cannabinoid-like compounds, including impairment in cognition, motor coordination, and 419 psychoses (Mallet et al., 2016). The disorders for which these agents are being tested are mostly neuropsychiatric, such as pain conditions, depression, anxiety disorders, and phobias (Mallet et al., 420 421 2016). Nevertheless, the current results in female rats and previous research in male rodents using 422 the carbamate FAAH inhibitors URB597 (e.g. Bortolato et al., 2007) and URB694 (Carnevali et al., 423 2015a) warrant more translational studies to examine the mood-modulating properties of this class of FAAH inhibitors (Gururajan et al., 2019). Recently, sex differences in hippocampal response to 424 pharmacological inhibition of FAAH activity have been reported in rats after acute intense stress 425 426 (Zer-Aviv and Akirav, 2016). This suggests that preclinical development of FAAH inhibitors for the pharmacotherapy of stress-related depression should aim at comparing the underlying 427 428 neurobiological mechanisms between males and females.

Table 1 Correlation matrix between plasma corticosterone levels, brain-derived neurotrophic factor
(BDNF) hippocampal content, and anandamide (AEA) hippocampal levels at the end of the
experimental protocol.

		Corticosterone	BDNF	AEA
Corticosterone	r	-		
	р			
BDNF	r	32	-	
	р	.082		
AEA	r	31	.44	-
	р	.068	.015	

434 Figure legends

435 **Figure 1.** Timeline of experimental procedures.

436

**Figure 2.** Body weight gain of paired-housed (PH) and socially isolated (SI) female rats treated with vehicle (VEH) or URB694, calculated as the difference between weight at the end (immediately before animals were euthanized) and at the start (when animals were assigned to the different housing conditions) of the experiment (n = 10 per group). Data are expressed mean±SEM. \* = significantly different from corresponding PH + VEH group; # = significantly different from corresponding SI + VEH group (p values are reported in the text).

443

Figure 3. Changes in sucrose solution preference in paired-housed (PH) and socially isolated (SI) female rats treated with vehicle (VEH) or URB694, calculated as the difference between each assessment point during the social isolation period and the baseline (n = 10 per group). Data are expressed mean $\pm$ SEM. \* = significantly different from corresponding PH + VEH group; # = significantly different from corresponding SI + VEH group (p values are reported in the text).

449

Figure 4. Time spent in immobility during the forced swim test by paired-housed (PH) and socially isolated (SI) female rats treated with vehicle (VEH) or URB694 (n = 10 per group). Data are expressed mean $\pm$ SEM. \* = significantly different from corresponding PH + VEH group; # = significantly different from corresponding SI + VEH group (p values are reported in the text).

454

Figure 5. Plasma corticosterone levels at the end of the experimental protocol in paired-housed (PH)
and socially isolated (SI) female rats treated with vehicle (VEH) or URB694 (n = 10 per group). Data
are expressed mean±SEM. \* = significantly different from corresponding PH + VEH group; # =
significantly different from corresponding SI + VEH group (p values are reported in the text).

459

Figure 6. Brain-derived neurotrophic factor (BDNF; panel A) and anandamide (panel B) levels, and
fatty acid amide hydrolase (FAAH) activity (panel C) in the hippocampus of paired-housed (PH)
and socially isolated (SI) female rats treated with vehicle (VEH) or URB694 (n = 10 per group).

- 463 Data are expressed mean±SEM. BDNF values are expressed as a percentage of the control group
- 464 (PH+VEH rats). \* = significantly different from corresponding PH + VEH group; # = significantly
- different from corresponding VEH group (p values are reported in the text).

### 467 **References**

- 468 Aguado, T., Monory, K., Palazuelos, J., Stella, N., Cravatt, B., Lutz, B., Marsicano, G., Kokaia, Z.,
- 469 Guzman, M., Galve-Roperh, I., 2005. The endocannabinoid system drives neural progenitor
- 470 proliferation. Faseb J. 19, 1704-1706.
- 471 Aso, E., Ozaita, A., Valdizan, E.M., Ledent, C., Pazos, A., Maldonado, R., Valverde, O., 2008. BDNF
- 472 impairment in the hippocampus is related to enhanced despair behavior in CB1 knockout mice. J.
- 473 Neurochem. 105, 565-572.
- Beery, A.K., 2018. Inclusion of females does not increase variability in rodent research studies. Curr.
  Opin. Behav. Sci. 23, 143-149.
- Beery, A.K., Kaufer, D., 2015. Stress, social behavior, and resilience: insights from rodents.
  Neurobiol. Stress 1, 116-127.
- 478 Belleau, E.L., Treadway, M.T., Pizzagalli, D.A., 2019. The Impact of Stress and Major Depressive
- Disorder on Hippocampal and Medial Prefrontal Cortex Morphology. Biol Psychiatry 85, 443-453.
- Bjorkqvist, K., 2001. Social defeat as a stressor in humans. Physiol. Behav. 73, 435-442.
- Bortolato, M., Mangieri, R.A., Fu, J., Kim, J.H., Arguello, O., Duranti, A., Tontini, A., Mor, M., Tarzia,
- 482 G., Piomelli, D., 2007. Antidepressant-like activity of the fatty acid amide hydrolase inhibitor URB597
- in a rat model of chronic mild stress. Biol. Psychiatry 62, 1103-1110.
- Burstein, O., Shoshan, N., Doron, R., Akirav, I., 2018. Cannabinoids prevent depressive-like symptoms and alterations in BDNF expression in a rat model of PTSD. Prog. Neuropsychopharmacol. Biol. Psychiatry, 84, 129–139.
- Carnevali, L., Montano, N., Statello, R., Sgoifo, A., 2017a. Rodent models of depressioncardiovascular comorbidity: Bridging the known to the new. Neurosci. Biobehav. Rev. 76, 144-153.
  Carnevali, L., Nalivaiko, E., Sgoifo, A., 2014. Respiratory patterns reflect different levels of
  aggressiveness and emotionality in Wild-type Groningen rats. Respir. Physiol. Neurobiol. 204, 2835.
- Carnevali, L., Rivara, S., Nalivaiko, E., Thayer, J.F., Vacondio, F., Mor, M., Sgoifo, A., 2017b.
  Pharmacological inhibition of FAAH activity in rodents: A promising pharmacological approach for
  psychological-cardiac comorbidity? Neurosci. Biobehav. Rev. 74, 444-452.

- 495 Carnevali, L., Vacondio, F., Rossi, S., Callegari, S., Macchi, E., Spadoni, G., Bedini, A., Rivara, S., Mor, M., Sgoifo, A., 2015a. Antidepressant-like activity and cardioprotective effects of fatty acid 496 497 amide hydrolase inhibitor URB694 socially stressed Wistar Kyoto rats. in Eur. Neuropsychopharmacol. 25, 2157-2169. 498
- Carnevali, L., Vacondio, F., Rossi, S., Macchi, E., Spadoni, G., Bedini, A., Neumann, I.D., Rivara,
  S., Mor, M., Sgoifo, A., 2015b. Cardioprotective effects of fatty acid amide hydrolase inhibitor
  URB694, in a rodent model of trait anxiety. Sci. Rep. 5, 18218.
- Castren, E., Voikar, V., Rantamaki, T., 2007. Role of neurotrophic factors in depression. Curr. Opin.
  Pharmacol. 7, 18-21.
- Clapper, J.R., Vacondio, F., King, A.R., Duranti, A., Tontini, A., Silva, C., Sanchini, S., Tarzia, G.,
  Mor, M., Piomelli, D., 2009. A second generation of carbamate-based fatty acid amide hydrolase
  inhibitors with improved activity in vivo. ChemMedChem 4, 1505-1513.
- 507 Cohen, S., Janicki-Deverts, D., Miller, G.E., 2007. Psychological stress and disease. Jama 298, 508 1685-1687.
- 509 Commons, K.G., Cholanians, A.B., Babb, J.A., Ehlinger, D.G., 2017. The Rodent Forced Swim Test
- 510 Measures Stress-Coping Strategy, Not Depression-like Behavior. ACS Chem. Neurosci. 8, 955-960.
- 511 Connolly, K.R., Thase, M.E., 2012. Emerging drugs for major depressive disorder. Expert. Opin. 512 Emerg. Drugs 17, 105-126.
- de Boer, S.F., Buwalda, B., Koolhaas, J.M., 2017. Untangling the neurobiology of coping styles in
  rodents: Towards neural mechanisms underlying individual differences in disease susceptibility.
  Neurosci. Biobehav. Rev. 74(Pt B), 401-422.
- 516 De Gregorio, D., Manchia, M., Carpiniello, B., Valtorta, F., Nobile, M., Gobbi, G., Comai, S., 2019. 517 Role of palmitoylethanolamide (PEA) in depression: Translational evidence: Special Section on 518 "Translational and Neuroscience Studies in Affective Disorders". Section Editor, Maria Nobile MD, 519 PhD. This Section of JAD focuses on the relevance of translational and neuroscience studies in 520 providing a better understanding of the neural basis of affective disorders. The main aim is to briefly 521 summaries relevant research findings in clinical neuroscience with particular regards to specific 522 innovative topics in mood and anxiety disorders. J. Affect. Disord. 255, 195-200.

- 523 De Kloet, E.R., Vreugdenhil, E., Oitzl, M.S., Joels, M., 1998. Brain corticosteroid receptor balance in 524 health and disease. Endocr. Rev. 19, 269-301.
- 525 Derkinderen, P., Valjent, E., Toutant, M., Corvol, J.C., Enslen, H., Ledent, C., Trzaskos, J., Caboche,
- 526 J., Girault, J.A., 2003. Regulation of extracellular signal-regulated kinase by cannabinoids in 527 hippocampus. J. Neurosci. 23, 2371–2382.
- Dinan, T.G., 2005. Stress: the shared common component in major mental illnesses. Eur. Psychiatry
  20 Suppl 3, S326-328.
- Duman, R.S., Monteggia, L.M., 2006. A neurotrophic model for stress-related mood disorders. Biol.
  Psychiatry 59, 1116-1127.
- 532 Finnell, J.E., Muniz, B.L., Padi, A.R., Lombard, C.M., Moffitt, C.M., Wood, C.S., Wilson, L.B.,
- 533 Reagan, L.P., Wilson, M.A., Wood, S.K., 2018. Essential Role of Ovarian Hormones in Susceptibility
- to the Consequences of Witnessing Social Defeat in Female Rats. Biol. Psychiatry 84, 372-382.
- 535 Food and Drug Administration, Drug Safety and Availability report, 2016. FDA finds drugs under
- 536 investigation in the U.S. related to French BIA 10-2474 drug do not pose similar safety risks.
- 537 https://www.fda.gov/drugs/drug-safety-and-availability/fda-finds-drugs-under-investigation-us-
- related-french-bia-10-2474-drug-do-not-pose-similar-safety.
- 539 Fowler, C.J., 2015. The potential of inhibitors of endocannabinoid metabolism as anxiolytic and 540 antidepressive drugs--A practical view. Eur. Neuropsychopharmacol. 25, 749-762.
- 541 Goncalves, M.B., Suetterlin, P., Yip, P., Molina-Holgado, F., Walker, D.J., Oudin, M.J., Zentar, M.P.,
- 542 Pollard, S., Yanez-Munoz, R.J., Williams, G., Walsh, F.S., Pangalos, M.N., Doherty, P., 2008. A
- diacylglycerol lipase-CB2 cannabinoid pathway regulates adult subventricular zone neurogenesis in
  an age-dependent manner. Mol. Cell. Neurosci. 38, 526-536.
- Gorzalka, B.B., Hill, M.N., 2011. Putative role of endocannabinoid signaling in the etiology of
  depression and actions of antidepressants. Prog. Neuropsychopharmacol. Biol. Psychiatry 35, 15751585.
- 548 Gorzalka, B.B., Hill, M.N., Hillard, C.J., 2008. Regulation of endocannabinoid signaling by stress:
- 549 implications for stress-related affective disorders. Neurosci. Biobehav. Rev. 32, 1152-1160.

- 550 Grigoriadis, S., Robinson, G.E., 2007. Gender issues in depression. Ann. Clin. Psychiatry 19, 247-551 255.
- 552 Grippo, A.J., Cushing, B.S., Carter, C.S., 2007. Depression-like behavior and stressor-induced 553 neuroendocrine activation in female prairie voles exposed to chronic social isolation. Psychosom. 554 Med. 69, 149-157.
- 555 Grippo, A.J., Sullivan, N.R., Damjanoska, K.J., Crane, J.W., Carrasco, G.A., Shi, J., Chen, Z.,
- 556 Garcia, F., Muma, N.A., Van de Kar, L.D., 2005. Chronic mild stress induces behavioral and
- 557 physiological changes, and may alter serotonin 1A receptor function, in male and cycling female rats.
- 558 Psychopharmacology (Berl) 179, 769-780.
- Gururajan, A., Reif, A., Cryan, J.F., Slattery, D.A., 2019. The future of rodent models in depression
  research. Nat. Rev. Neurosci., in press.
- 561 Haller, J., Goldberg, S.R., Pelczer, K.G., Aliczki, M., Panlilio, L.V., 2013. The effects of anandamide
- signaling enhanced by the FAAH inhibitor URB597 on coping styles in rats. Psychopharmacology
  (Berl) 230, 353-362.
- Hashimoto, K., 2010. Brain-derived neurotrophic factor as a biomarker for mood disorders: an
  historical overview and future directions. Psychiatry Clin. Neurosci. 64, 341-357.
- Hashimotodani, Y., Ohno-Shosaku, T., Kano, M., 2007. Endocannabinoids and synaptic function in
  the CNS. Neuroscientist 13, 127-137.
- Hayley, S., Anisman, H., 2013. Neurotrophic paths in the treatment of depression. J. Psychiatry
  Neurosci. 38, 291-293.
- Heinrich, L.M., Gullone, E., 2006. The clinical significance of loneliness: a literature review. Clin.
  Psychol. Rev. 26, 695-718.
- Hill, M.N., Carrier, E.J., McLaughlin, R.J., Morrish, A.C., Meier, S.E., Hillard, C.J., Gorzalka, B.B.,
- 573 2008. Regional alterations in the endocannabinoid system in an animal model of depression: effects
- of concurrent antidepressant treatment. J. Neurochem. 106, 2322-2336.
- 575 Hill, M.N., Kambo, J.S., Sun, J.C., Gorzalka, B.B., Galea, L.A., 2006. Endocannabinoids modulate
- 576 stress-induced suppression of hippocampal cell proliferation and activation of defensive behaviours.
- 577 Eur. J. Neurosci. 24, 1845-1849.

- Hill, M.N., McEwen, B.S., 2010. Involvement of the endocannabinoid system in the neurobehavioural
  effects of stress and glucocorticoids. Prog. Neuropsychopharmacol. Biol. Psychiatry 34, 791-797.
- Hill, M.N., Patel, S., Carrier, E.J., Rademacher, D.J., Ormerod, B.K., Hillard, C.J., Gorzalka, B.B.,
  2005. Downregulation of endocannabinoid signaling in the hippocampus following chronic
  unpredictable stress. Neuropsychopharmacology 30, 508-515.
- Hill, M.N., Titterness, A.K., Morrish, A.C., Carrier, E.J., Lee, T.T., Gil-Mohapel, J., Gorzalka, B.B.,
  Hillard, C.J., Christie, B.R., 2010. Endogenous cannabinoid signaling is required for voluntary
  exercise-induced enhancement of progenitor cell proliferation in the hippocampus. Hippocampus 20,
  513-523.
- 587 Huang, G.Z., Woolley, C.S., 2012. Estradiol acutely suppresses inhibition in the hippocampus 588 through a sex-specific endocannabinoid and mGluR-dependent mechanism. Neuron 74, 801-808.
- 589 Izadi, M.S., Radahmadi, M., Ghasemi, M., Rayatpour, A., 2018. Effects of Isolation and Social
- 590 Subchronic Stresses on Food Intake and Levels of Leptin, Ghrelin, and Glucose in Male Rats. Adv.
- 591 Biomed. Res. 7, 118.
- Jacobson, L., Sapolsky, R., 1991. The role of the hippocampus in feedback regulation of the hypothalamic-pituitary-adrenocortical axis. Endocr. Rev. 12, 118-134.
- Kokras, N., Antoniou, K., Mikail, H.G., Kafetzopoulos, V., Papadopoulou-Daifoti, Z., Dalla, C., 2015.
  Forced swim test: What about females? Neuropharmacology 99, 408-421.
- 596 Kokras, N., Dalla, C., 2014. Sex differences in animal models of psychiatric disorders. Br. J. 597 Pharmacol. 171, 4595-4619.
- Leal, G., Bramham, C.R., Duarte, C.B., 2017. BDNF and Hippocampal Synaptic Plasticity. Vitam.
  Horm. 104, 153-195.
- Levone, B.R., Cryan, J.F., O'Leary, O.F., 2015. Role of adult hippocampal neurogenesis in stress
  resilience. Neurobiol. Stress 1, 147-155.
- Lu, B., Nagappan, G., Lu, Y., 2014. BDNF and synaptic plasticity, cognitive function, and dysfunction.
- 603 Handb. Exp. Pharmacol. 220, 223-250.
- Mackie, K., 2005. Distribution of cannabinoid receptors in the central and peripheral nervous system.
- 605 Handb. Exp. Pharmacol., 299-325.

- Mallet, C., Dubray, C., Dualé, C., 2016. FAAH inhibitors in the limelight, but regrettably. Int. J. Clin.
  Pharmacol. Ther. 54, 498-501.
- Marchalant, Y., Brothers, H.M., Wenk, G.L., 2009. Cannabinoid agonist WIN-55,212-2 partially restores neurogenesis in the aged rat brain. Mol. Psychiatry 14, 1068-1069.
- Marcondes, F.K., Bianchi, F.J., Tanno, A.P., 2002. Determination of the estrous cycle phases of rats:
- some helpful considerations. Braz. J. Biol. 62, 609-614.
- Micale, V., Di Marzo, V., Sulcova, A., Wotjak, C.T., Drago, F., 2013. Endocannabinoid system and
  mood disorders: priming a target for new therapies. Pharmacol. Ther. 138, 18-37.
- Petrosino, S., Ligresti, A., Di Marzo, V., 2009. Endocannabinoid chemical biology: a tool for the
- development of novel therapies. Curr. Opin. Chem. Biol. 13, 309-320.
- 616 Pittenger, C., Duman, R.S., 2008. Stress, depression, and neuroplasticity: a convergence of 617 mechanisms. Neuropsychopharmacology 33, 88-109.
- Plenis, A., Konieczna, L., Oledzka, I., Kowalski, P., Baczek, T., 2011. Simultaneous determination
  of urinary cortisol, cortisone and corticosterone in parachutists, depressed patients and healthy
  controls in view of biomedical and pharmacokinetic studies. Mol. Biosyst. 7, 1487-1500.
- Porsolt, R.D., Le Pichon, M., Jalfre, M., 1977. Depression: a new animal model sensitive to antidepressant treatments. Nature 266, 730-732.
- Romano, A., Tempesta, B., Provensi, G., Passani, M.B., Gaetani, S., 2015. Central mechanisms
- 624 mediating the hypophagic effects of oleoylethanolamide and N-acylphosphatidylethanolamines:
- 625 different lipid signals? Front. Pharmacol. 6, 137.
- Rubino, T., Forlani, G., Vigano, D., Zippel, R., Parolaro, D., 2005. Ras/ERK signalling in cannabinoid
  tolerance: from behaviour to cellular aspects. J. Neurochem. 93, 984–991.
- Sapolsky, R.M., 2000. The possibility of neurotoxicity in the hippocampus in major depression: a
  primer on neuron death. Biol. Psychiatry 48, 755-765.
- 630 Scarante, F.F., Vila-Verde, C., Detoni, V.L., Ferreira-Junior, N.C., Guimaraes, F.S., Campos, A.C.,
- 631 2017. Cannabinoid Modulation of the Stressed Hippocampus. Front. Mol. Neurosci. 10, 411.
- 632 Sheline, Y.I., 1996. Hippocampal atrophy in major depression: a result of depression-induced
- 633 neurotoxicity? Mol. Psychiatry 1, 298-299.

- Sheline, Y.I., Liston, C., McEwen, B.S., 2019. Parsing the Hippocampus in Depression: Chronic
  Stress, Hippocampal Volume, and Major Depressive Disorder. Biol. Psychiatry 85, 436-438.
- 636 Sloan, D.M., Kornstein, S.G., 2003. Gender differences in depression and response to 637 antidepressant treatment. Psychiatr. Clin. North. Am. 26, 581-594.
- 638 Stepanichev, M., Dygalo, N.N., Grigoryan, G., Shishkina, G.T., Gulyaeva, N., 2014. Rodent models
- of depression: neurotrophic and neuroinflammatory biomarkers. Biomed. Res. Int. 2014, 932757.
- 640 Stetler, C., Miller, G.E., 2011. Depression and hypothalamic-pituitary-adrenal activation: a 641 quantitative summary of four decades of research. Psychosom. Med. 73, 114-126.
- Sun, M., Choi, E.Y., Magee, D.J., Stets, C.W., During, M.J., Lin, E.J., 2014. Metabolic Effects of
  Social Isolation in Adult C57BL/6 Mice. Int. Sch. Res. Notices 2014, 690950.
- Tabatadze, N., Huang, G., May, R.M., Jain, A., Woolley, C.S., 2015. Sex Differences in Molecular
  Signaling at Inhibitory Synapses in the Hippocampus. J. Neurosci. 35, 11252-11265.
- Tardito, D., Perez, J., Tiraboschi, E., Musazzi, L., Racagni, G., Popoli, M., 2006. Signaling pathways
  regulating gene expression, neuroplasticity, and neurotrophic mechanisms in the action of
  antidepressants: a critical overview. Pharmacol Rev. 58, 115-134.
- Tarzia, G., Duranti, A., Gatti, G., Piersanti, G., Tontini, A., Rivara, S., Lodola, A., Plazzi, P.V., Mor,
  M., Kathuria, S., Piomelli, D., 2006. Synthesis and structure-activity relationships of FAAH inhibitors:
  cyclohexylcarbamic acid biphenyl esters with chemical modulation at the proximal phenyl ring.
  ChemMedChem 1, 130-139.
- Temporary Specialist Scientific Committee, 2016. FAAH (Fatty Acid Amide Hydrolase), on the causes of the accident during a Phase 1 clinical trial in Rennes in January 2016. https://ansm.sante.fr/var/ansm\_site/storage/original/application/744c7c6daf96b141bc9509e2f85c2 27e.pdf
- van Esbroeck, A.C.M., Janssen, A.P.A., Cognetta, A.B., Ogasawara, D., Shpak, G., van der Kroeg,
  M., Kantae, V., Baggelaar, M.P., de Vrij, F.M.S., Deng, H., Allarà, M., Fezza, F., Lin, Z., van der Wel,
  T., Soethoudt, M., Mock, E.D., den Dulk, H., Baak, I.L., Florea, B.I., Hendriks, G., De Petrocellis, L.,
  Overkleeft, H.S., Hankemeier, T., De Zeeuw, C.I., Di Marzo, V., Maccarrone, M., Cravatt, B.F.,

- Kushner, S.A., van der Stelt, M., 2017. Activity-based protein profiling reveals off-target proteins of
- 662 the FAAH inhibitor BIA 10-2474. Science 356, 1084–1087.
- von Schaper, E., 2016. Bial incident raises FAAH suspicions. Nat. Biotechnol. 34, 223.
- 664 Zer-Aviv, T.M., Akirav, I., 2016. Sex differences in hippocampal response to endocannabinoids after
- 665 exposure to severe stress. Hippocampus 26, 947-957.















FIGURE 5



FIGURE 6