

1 **Growth response of cuttings to drought and intermittent flooding for three *Salix***  
2 **species and implications for riverbank soil bioengineering**

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16 **Abstract**

17 Willows are used as cuttings or in fascines for riverbank soil bioengineering, to control  
18 erosion with their high resprouting ability and rapid growth. However, water availability  
19 is highly variable along riverbanks both in time and space and constitutes a major stress  
20 limiting willow establishment. A species-specific understanding of willow cutting response  
21 to water stress is critical to design successful riverbank soil bioengineering projects given  
22 exclusive use of local species is often recommended. In a three-month greenhouse  
23 experiment, we investigated the effects of three soil moisture treatments (drought – soil  
24 saturation – intermittent flooding) on survival, biomass production and root growth of  
25 cuttings of three willow species used for soil bioengineering along NE American streams  
26 (*Salix discolor* – *S. eriocephala* – *S. interior*). Cutting survival was high for all species and  
27 treatments (>89%). Biomass production and root volume only differed between species. *S.*  
28 *eriocephala* produced the highest biomass and root volume, and *S. discolor* invested more  
29 in below- than aboveground biomass. Root length responded to soil moisture, but  
30 differently between species. Under intermittent flooding, *S. eriocephala* produced shorter  
31 roots, while *S. interior* produced longer roots. For riverbank soil bioengineering, *S.*  
32 *eriocephala* should be favoured at medium elevation and *S. interior* at lower elevation.

33

34 **Keywords**

35 Bioengineering; Biomass allocation; Root growth; *Salicaceae*; Water stress; WinRHIZO

**36 Introduction**

37 Willows (*Salix* spp.) encompass up to 500 species of shrubs and trees that dominate  
38 riparian habitats across different biomes (Rechinger 1992; Malanson 1993; Karrenberg et  
39 al. 2002; Kuzovkina et al. 2009). These pioneer plants are characterized by morphological  
40 and dispersal strategies adapted to fluvial environments (Naiman et al. 2005). One female  
41 willow can produce thousands to millions of seeds annually to be dispersed by wind and  
42 water, which confers these species a high colonization ability in the open, bare and moist  
43 riverbank surfaces that flooding disturbance leaves behind (Karrenberg et al. 2002;  
44 González et al. 2016; Bourgeois and González 2019). Once established, their dense and  
45 relatively deep root system further reduces uprooting risk during flooding and fosters soil  
46 stabilization (Karrenberg et al. 2002; Catford and Jansson 2014). Moreover, willows  
47 possess a rapid growth rate and a high capacity for vegetative propagation, which promotes  
48 their fast re-establishment after disturbance (Krasny et al. 1988; Kollmann et al. 1999; but  
49 see Douhovnikoff et al. 2005). Owing to this extraordinary resprouting ability, willows are  
50 frequently introduced as cuttings or fascines during soil bioengineering projects designed  
51 to control erosion along riverbanks (Evette et al. 2009; Kuzovkina, et al 2009).

52 Along riverbanks, hydrological conditions fluctuate widely both in time following high  
53 flow-baseflow cycles, and in space, as less topographically elevated surfaces (e.g., lower  
54 banks) experience flooding more regularly than more elevated ones (e.g., upper banks).  
55 Tolerance to water stress (drought and flood-induced anoxia) is therefore a major constraint  
56 that can limit willow establishment and survival along bioengineered riverbanks (Francis  
57 et al. 2005; Pezeshki et al. 2007). In that regard, riparian willows have developed specific  
58 strategies to better cope with drought (e.g., by deepening their root system; Garssen et al.

59 2014) or anoxia (e.g., by developing lenticels and adventitious roots; Kozłowski 1997; Li  
60 et al. 2006; Parent et al. 2008), but these vary markedly across species, but also within  
61 species in relation to phenotypic plasticity. Many experiments have been conducted under  
62 controlled conditions to test the survival and growth of *Salix* species along a gradient from  
63 permanent anoxia to extreme drought (Krasny et al. 1988; Jackson and Attwood 1996;  
64 Amlin and Rood 2001), as such responses can be less detectable under field conditions (but  
65 see Shields et al. 1998; Caplan et al. 2013). For example, *Salix eleagnos* tolerates both  
66 water table declines and periodic inundation well (Francis et al. 2005). *S. subfragilis* has  
67 higher tolerance to anoxia than *S. gracilistyla* (Nakai and Kisanuki 2011). Shoot and root  
68 elongation of *S. exigua* increased under inundated conditions but remained constant for *S.*  
69 *lutea* (Amlin and Rood 2001). *S. alba* and *S. matsudana* x *S. alba* hybrids showed reduced  
70 growth after six weeks under drought compared to continuous flooding (Doffo et al. 2017).  
71 Wikberg and Ögren (2004) reported higher tolerance to drought for *S. purpurea*,  
72 intermediate tolerance for *S. viminalis*, and lower tolerance for *S. schwererii* x *S. viminalis*.  
73 Similarly, *S. purpurea* exhibits higher drought tolerance than *S. suchowensis*, as shown by  
74 less growth reduction (Jia et al. 2020). Some species also perform better at field capacity,  
75 such as *S. nigra*, which showed reduced growth under both drought and flooding (Pezeshki  
76 et al. 1998). Given these species-specific responses, carefully selecting local willow  
77 species according to their tolerance to water stress could greatly improve the success of  
78 riverbank soil bioengineering projects (Kuzovkina and Volk 2009). To meet this objective,  
79 a greater understanding of local willow species' response to water stress is required,  
80 notably in regions where such information is scarce, such as North-Eastern America.

81 This study investigated the effect of three soil moisture treatments (drought, soil  
82 saturation, and intermittent flooding) on survival as well as on the biomass production, root  
83 length and root volume of cuttings from three willow species: *Salix discolor* Muhlenberg,  
84 *S. eriocephala* Michaux, and *S. interior* Rowlee. These species, native to North America,  
85 were selected as they are commonly used for riverbank soil bioengineering projects due to  
86 their high biomass production, high resprouting ability and distinct lateral distribution  
87 along riverbanks (Marie-Victorin et al. 2002). We hypothesized that *S. discolor*, which  
88 naturally occurs on upper banks should be more tolerant to drought, while *S. interior*, which  
89 is mostly restricted to lower banks, should be more tolerant to intermittent flooding. As a  
90 generalist species distributed on both upper and lower banks, *S. eriocephala* was expected  
91 to be highly tolerant to both drought and intermittent flooding. Based on results from our  
92 three-month greenhouse experiment, we provide insights for willow species selection in a  
93 context of riverbank soil bioengineering.

94

## 95 **Materials and methods**

### 96 *Cutting collection*

97 Cuttings of *Salix discolor*, *S. eriocephala* and *S. interior* were collected during the  
98 dormancy phase (November 2017) from four natural populations per species established in  
99 seven riparian areas surrounding Quebec City, Canada (supplementary Table S1). For each  
100 species, a total of 12 individuals (from two to five individuals per population) measuring 1  
101 to 2.5 m tall were selected, from which three cuttings were obtained, for a total of 108  
102 cuttings. More precisely, newly produced willow branches, that were 20-cm long and  
103 similar in diameter (around 0.6 cm), were harvested as cuttings. Only young willow

104 branches from the last growing season were therefore selected as their reduced lignin  
105 content promotes resprouting compared to older branches. All these cuttings were  
106 harvested at the apex of parent shoot, given cutting location on the parent shoot (i.e., near  
107 the base of the parent shoot, or near the apex) can influence their rooting ability (Zalesny  
108 and Wiese, 2006). These cuttings were also visually inspected to ensure for their  
109 homogeneity within species and among sites. Each of the three cuttings obtained from a  
110 single individual was dedicated to one of the three soil moisture treatments. After a large-  
111 spectrum fungicide was applied, cuttings were stored in vacuum sealed bags at -4°C until  
112 the start of the experiment (i.e., for three months). Throughout this study, we therefore  
113 ensured to use a harvest and storage protocol similar for all cuttings in order to limit  
114 potential biases related to harvest date, cutting size or differences in cutting location along  
115 the parent shoot (by selecting cuttings only at the apex of the parent shoot).

116

#### 117 *Cutting plantation*

118 After rehydrating cuttings by immersion under water for 24h to promote resprouting,  
119 each willow cutting was planted in a 30 L permeable bag filled with sand, with two-thirds  
120 of its length buried and at least three buds present on the aboveground part. Prior to  
121 planting, the initial diameter of each cutting measured. Cuttings were grown for three  
122 weeks at soil saturation and slightly fertilized with 500 mL of an NPK solution (20-20-20  
123 kg.ha<sup>-1</sup>) to reduce stress and mortality risk upon application of the soil moisture treatments.

124

#### 125 *Experimental design*

126 A greenhouse split-plot experiment was conducted to determine the effect of drought,  
 127 soil saturation and intermittent flooding on the survival and growth (below- and  
 128 aboveground biomass, root length and root volume) of cuttings of the three willow species  
 129 studied. Soil moisture treatments were applied to main plots corresponding to plastic tanks  
 130 (110 x 72 x 100 cm) arranged into six blocks for a total of 18 experimental units. Willow  
 131 species were applied to subplots corresponding to the 30 L bags. One cutting was planted  
 132 in each bag, for a total of six subplots: i.e. two willow cuttings per species and per tank  
 133 (Figure S1).

134 The three soil moisture treatments were applied by using different watering frequency  
 135 and volume. The drought treatment was obtained by irrigating cuttings once a week with  
 136 1-2 L of water per 30 L bag, a water availability ranging between wilting point and  
 137 optimum for plant growth. For the soil saturation treatment, cuttings were irrigated twice a  
 138 week with 3-4 L of water per 30 L bag, which corresponded to field capacity, i.e. the  
 139 amount of water retained in the soil after the excess water has drained away and the rate of  
 140 downward movement has decreased (Veihmeyer and Hendrickson 1931). Under the  
 141 intermittent flooding treatment, water level was maintained at 5 cm above the substrate  
 142 surface for two weeks, then drained away using a faucet installed at the bottom of each  
 143 tank, and soil kept at water saturation for one week; this three-week cycle was repeated  
 144 four times throughout the experiment. Soil water content was further monitored every two  
 145 days using a time-domain-reflectometry moisture sensor in order to maintain the desired  
 146 water availability.

147 Greenhouse temperature, relative humidity and daily light integral respectively varied  
 148 between 17.0 and 28.0 °C (mean: 23.5°C), 35 and 75% (mean: 57.8%), and 6 and 25 mol.m<sup>-2</sup>

149  $^2 \cdot \text{day}^{-1}$  (mean:  $22 \text{ mol} \cdot \text{m}^{-2} \cdot \text{day}^{-1}$ ), during the three months of the experiment (March-May  
 150 2018), excluding the three-week acclimation phase.

151

152 *Growth measurements*

153 At the end of the experiment, cuttings were carefully removed from the substrate. Roots  
 154 were first separated from the aboveground parts (leaves and stem), cleaned with water,  
 155 coloured with a red dye solution, and the complete root system of each plant scanned with  
 156 an image analysis system (WinRHIZO, Regent Instruments) to determine total root length  
 157 as well as the volume of three root types based on a preliminary experiment, i.e. fine  
 158 (diameter: 0.5-1.5 mm), medium (1.5-2.0 mm) and coarse (2.0-2.5 mm). Roots, leaves and  
 159 stem were then dried at  $105^\circ\text{C}$  for 48h and weighed to measure newly produced  
 160 aboveground (leaf + stem) and belowground (root) biomass. Biomass allocation was then  
 161 calculated as the ratio of aboveground biomass over belowground biomass. Cuttings were  
 162 excluded from both biomass and root measurements given that we aimed to evaluate the  
 163 resprouting ability of the studied species under various soil moisture treatments.

164

165 *Statistical analyses*

166 Linear Mixed Models (LMMs) followed by LSD (Fisher's Least Significant Difference)  
 167 were used to assess the effects of species and soil moisture on the five plant growth metrics,  
 168 namely above- and belowground biomass, biomass allocation, root length and volume.  
 169 Initial cutting diameter was included as a covariable while block and source population  
 170 were included as random effects in these LMMs investigating changes in above- and  
 171 belowground biomass and biomass allocation. For root volume, the effect of root type (i.e.,



172 fine, medium or coarse root) was also added to the model in interaction with species and  
 173 soil moisture. Data were log- or square root-transformed when needed to meet residual  
 174 normality and homoscedasticity, and back-transformed for results presentation. Analyses  
 175 were conducted using PROC MIXED on SAS 9.4 (SAS Institute, Cary, NC, USA).

176

## 177 **Results**

### 178 *Survival*

179 High cutting survival was observed for all three species and soil moisture treatments at  
 180 the end of the experiment. All *Salix eriocephala* cuttings survived. Survival varied from  
 181 89% (soil saturation) to 92% (drought) and 94% (intermittent flooding) for *S. discolor*, and  
 182 from 97% (drought and soil saturation) to 100% (intermittent flooding) for *S. interior*.

183

### 184 *Biomass production*

185 Biomass production was independent of soil moisture treatment, but strongly varied  
 186 between species for all biomass components investigated (Table 1). *S. eriocephala*  
 187 produced the highest aboveground biomass, namely 3.9 and 1.6 times more than for *S.*  
 188 *interior* and *S. discolor*, respectively (Fig.1). Aboveground biomass production was almost  
 189 equally distributed between leaves and (new) stems for *S. eriocephala* (respectively 55%  
 190 and 45%) while leaf biomass was approximately twice that of stem biomass for the two  
 191 other species studied ( $p = 0.0027$  for species effect on leaves biomass and  $p < 0.0001$  for  
 192 species effect on stem biomass, all species being significantly different from each other for  
 193 these two biomass metrics according to *post-hoc* multiple comparisons). Belowground  
 194 biomass was also maximal for *S. eriocephala*, with 2.8 and 2.3 times more root biomass

195 produced relative to *S. discolor* and *S. interior*, respectively (Fig. 1). Finally, biomass  
 196 allocation differed between the three species: aboveground biomass was 4.4 times higher  
 197 than belowground for *S. interior*, 2.6 times higher for *S. eriocephala* and 1.3 times higher  
 198 for *S. discolor* (Fig. 1).

199

#### 200 *Root length and volume*

201 Soil moisture influenced root length differently for each species (significant interaction;  
 202 Table 2), though on average, root length was maximal for *S. eriocephala* and minimal for  
 203 *S. interior*. Root length increased at soil saturation compared to intermittent flooding for *S.*  
 204 *eriocephala*, while the reverse was observed for *S. interior*. For *S. discolor*, root length was  
 205 independent of soil moisture, with length similar to *S. interior* under intermittent flooding,  
 206 and similar to *S. eriocephala* at soil saturation (Fig. 2).

207 Root volume varied between species, independently of soil moisture (Table 3). The root  
 208 volume of *S. eriocephala* cuttings was indeed almost twice that of *S. discolor* and *S.*  
 209 *interior* (Fig. 3). Yet, for all species, root volume was mostly accounted for by fine roots  
 210 (66%), while medium and coarse roots respectively contributed to 26% and 8% of the total  
 211 root volume (Table 3; Fig. 3).

212

#### 213 **Discussion**

214 This study provides a greater understanding of the establishment and growth potential  
 215 of willow cuttings from *Salix discolor*, *S. eriocephala*, and *S. interior*, under various water  
 216 stress levels. Among the three growth parameters studied here, only root length was  
 217 significantly influenced by soil moisture, with distinct responses among species. *S.*

218 *eriocephala* produced the longest roots under all soil moisture treatments, despite reduced  
 219 length under intermittent flooding. *S. interior* had the shortest roots of the three species,  
 220 and produced longer roots under intermittent flooding. The contrasting effect of soil  
 221 moisture on *S. eriocephala* and *S. interior* concurs with previous studies demonstrating a  
 222 high plasticity of willow root systems in response to drought and flooding (Pezeshki et al.  
 223 1998; Wikberg and Ögren 2007; Parent et al. 2008). Root length was however independent  
 224 of soil moisture for *S. discolor*. These findings partly support our hypotheses: only *S.*  
 225 *interior*, which is usually found on lower banks, performed better under intermittent  
 226 flooding. Conversely, *S. eriocephala* showed a reduced tolerance to intermittent flooding  
 227 despite being a generalist species distributed both on lower and upper banks. Although  
 228 Amlin and Rood (2001) found that *S. discolor* root length decreased with increasing water  
 229 depths in a five-month experiment, no difference in root length was detected here under  
 230 drought compared to soil saturation. Our results suggest that the two other species studied  
 231 may be as well adapted to drought as *S. discolor*, even if this latter species is more  
 232 frequently found on upper banks. Factors other than soil moisture (e.g., soil nutrient  
 233 resources, interspecific competition) may explain the distribution of our willow species  
 234 under natural conditions. Further investigations are therefore required to determine these  
 235 limiting factors.

236 Interestingly, the two other growth parameters studied only differed between species,  
 237 independently of soil moisture. Biomass production (both above- and belowground) and  
 238 root volume were higher for *S. eriocephala* compared to *S. discolor* and *S. interior*. Higher  
 239 rooting ability was also previously observed in *S. eriocephala* compared to *S. discolor* in  
 240 highly disturbed coal mines (Mosseler et al. 2014). Differences in biomass allocation were

241 also evidenced between species: *S. interior* and *S. eriocephala* mostly invested resources  
 242 in aboveground biomass, while biomass was equally distributed in *S. discolor*. While  
 243 riparian plants are generally considered to invest resources firstly in anchoring to the soil  
 244 and secondly in developing aboveground parts, the three species studied here mostly  
 245 allocated resources to stems and leaves which could possibly be due to the absence of shear  
 246 stress and scouring in greenhouse. The absence of soil moisture effect on biomass  
 247 allocation contrasts with previous studies reporting that willow root/shoot ratio increased  
 248 with drought and decreased with flooding (Kozłowski 1997; Lavaine et al. 2015, Markus-  
 249 Michalczyk et al. 2016). As suggested by Li et al. (2004), more prolonged or pronounced  
 250 water stress might be required to observe differences in biomass allocation. Even though  
 251 we did not find differences in biomass production and allocation between soil moisture  
 252 treatments, we believe that the findings related to allocation to aboveground biomass,  
 253 which were higher for *S. interior* and lower for *S. discolor* are consistent with their relative  
 254 positions on riverbanks. *S. interior* on lower banks regularly experience anoxia and invest  
 255 more in lateral spread through root suckering from an extensive lateral root system, hence  
 256 promoting higher aboveground biomass (Mosseler et al. 2014), whereas the other two  
 257 species do not share this ability. This ability might have promoted higher *S. interior* root  
 258 length under intermittent flooding. *S. discolor* usually grows on the upper parts of the  
 259 bank, where it has greater access to unsaturated soil. A different root architecture would be  
 260 expected in plants in a lower topographic position, which also need to cope with shear  
 261 stress and scouring through anchoring.

262 The differential response of root length, root volume and biomass production to water  
 263 stress suggests high physio-morphological adaptability of willow species to environmental

264 conditions in relation to complex trade-offs between plant traits (Kozłowski 1997).  
265 Previous studies have shown that drought and flooding modulate leaf area, wood density,  
266 stomatal conductance or xylem cavitation among some willow species (Wikberg and Ögren  
267 2007; Savage and Cavender-Bares 2011, Doffo et al. 2017). Such physiological  
268 adjustments may therefore help cuttings to maintain similar levels of biomass production  
269 despite varying soil moisture. As shown here, investigating multiple plant traits is required  
270 to fully characterize willow response to environmental stressors.

271 In a context of riverbank stabilization by soil bioengineering, our study suggests that *S.*  
272 *eriocephala* should be preferred to ensure rapid erosion control through its higher biomass  
273 and tolerance to changing water availability. When drought and flooding are less frequent,  
274 *S. discolor* also seems a good candidate species, as it produced similar root length to *S.*  
275 *eriocephala* at soil saturation. The higher investment of *S. discolor* in belowground  
276 biomass relative to aboveground biomass is also known to be a plant trait fostering soil  
277 stabilization (Gray and Sotir 1996). Introducing a mix of both *S. eriocephala* and *S.*  
278 *discolor* cuttings could also be an interesting alternative strategy to provide higher  
279 resilience to changing environmental conditions (such as climate-induced increases in  
280 summer drought; Garssen et al. 2014; Bourgeois and González 2019) or optimized  
281 ecosystem functions (such as biodiversity support; Fang et al., 2015), and hence promote  
282 the success of bioengineering projects. While *S. interior* might appear less suitable for soil  
283 bioengineering given its lower relative investment in belowground biomass, it may be of  
284 interest for vegetating the lower parts of the banks. Indeed, compared to the two other  
285 species, its lower position on the bank seems to indicate higher tolerance to flooding. *S.*  
286 *interior* also has notable regeneration abilities, including root suckering and short, flexible

287 multistems, suitable traits in a context of soil bioengineering (Gray and Sotir 1996). Longer  
288 experiments including a larger number of *Salix* populations might however be required to  
289 confirm these recommendations. Further investigations that would include additional  
290 species could also help expand the list of willow species suitable for riverbank stabilization  
291 by soil bioengineering. Determining the response of willow species to additional flood- or  
292 ice-induced disturbances such as resistance to shear stress and scouring (e.g., Kui and Stella  
293 2016) should also be considered in future experiments to gain further insight into species  
294 performance on riverbanks and improve soil bioengineering techniques.

295

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302

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413 **Table 1.** Effect of soil moisture treatments (drought – soil saturation – intermittent  
 414 flooding) and species (*Salix discolor* – *S. eriocephala* – *S. interior*) on the aboveground  
 415 (leaf + stem) and belowground (root) biomass production and biomass allocation (ratio of  
 416 aboveground biomass over belowground biomass) of willow plants obtained from cuttings,  
 417 assessed by linear mixed models. P-values for initial cutting diameter used as a covariate  
 418 were 0.9901 for aboveground biomass production, 0.0058 for belowground biomass  
 419 production, and 0.0567 for biomass allocation. Significant p-values are indicated in bold.  
 420

	df	Aboveground biomass		Belowground biomass		Biomass allocation	
		F	p-value	F	p-value	F	p-value
Soil moisture	2	0.52	0.6091	0.85	0.4554	0.05	0.9522
Species	2	25.80	<b>0.0011</b>	20.68	< <b>0.0001</b>	39.12	< <b>0.0001</b>
421 Soil moisture x Species	4	0.23	0.9121	1.24	0.3422	0.44	0.7765

422 **Table 2.** Effect of soil moisture treatments (drought – soil saturation – intermittent  
 423 flooding) and species (*Salix discolor* – *S. eriocephala* – *S. interior*) on the root length of  
 424 willow plants obtained from cuttings, assessed by a linear mixed model. The p-value for  
 425 initial cutting diameter used as a covariate was 0.9800. Significant p-values are indicated  
 426 in bold.

	df	F	p-value
Soil moisture	2	0.51	0.6174
Species	2	52.82	< <b>0.0001</b>
428 Soil moisture x Species	4	3.20	<b>0.0489</b>

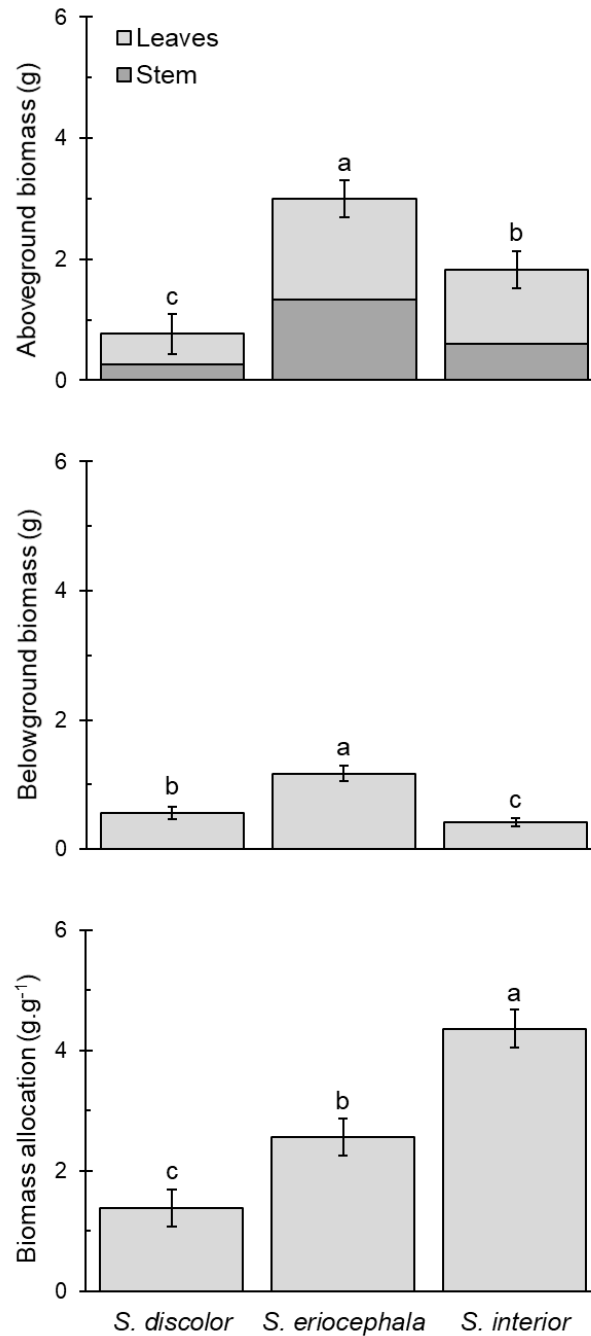
429 **Table 3.** Effect of soil moisture treatments (drought – soil saturation – intermittent  
 430 flooding), species (*Salix discolor* – *S. eriocephala* – *S. interior*) and root type (fine –  
 431 medium – coarse) on the root volume of willow plants obtained from cuttings, assessed by  
 432 a linear mixed model. The p-value for initial cutting diameter used as a covariate was  
 433 0.9735. Significant p-values are indicated in bold.

434

	df	F	p-value
Soil moisture	2	0.67	0.5320
Species	2	7.77	<b>0.0006</b>
Root type	2	190.40	< <b>0.0001</b>
Soil moisture x Species	4	0.50	0.7358
Species x Root type	4	1.59	0.1796
Root type x Soil moisture	4	0.66	0.6203
Soil moisture x Species x Root type	8	0.63	0.7486

435

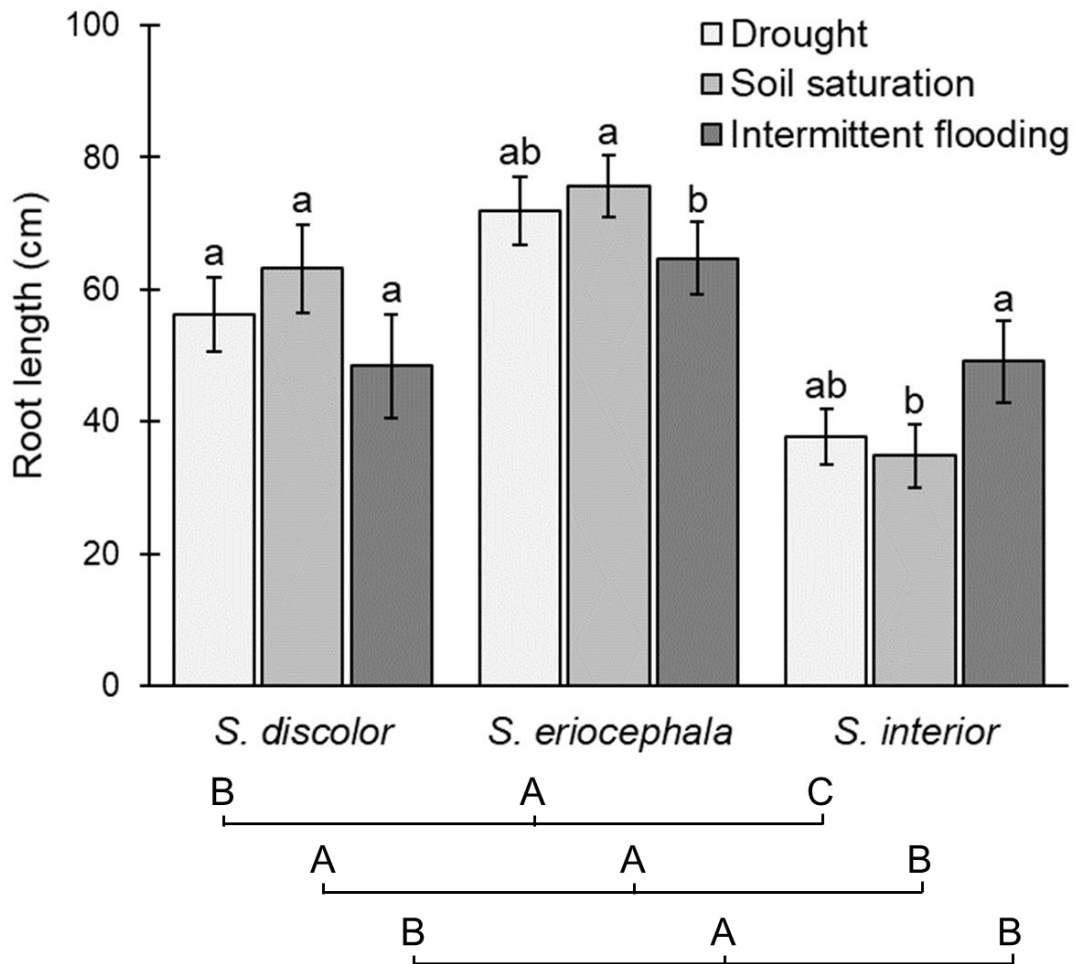
436 **Fig. 1.** Variations in aboveground biomass production, belowground biomass production,  
 437 and biomass allocation (ratio of aboveground biomass over belowground biomass)  
 438 between plants of three willow species obtained from cuttings (*Salix discolor* – *S.*  
 439 *eriocephala* – *S. interior*; mean  $\pm$  standard error; n = 36 cuttings per species). Letters  
 440 indicate significant differences obtained by LSD (Fisher's Least Significant Difference).



441

442 **Fig. 2.** Variations in root length (mean  $\pm$  standard error) between the three soil moisture  
 443 treatments (drought – soil saturation – intermittent flooding) and the three willow species  
 444 studied (*Salix discolor* – *S. eriocephala* – *S. interior*; n = 12 cuttings per species and soil  
 445 moisture treatments). Lowercase letters indicate significant differences between soil  
 446 moisture treatments for a given species, and uppercase letters indicate significant  
 447 differences between species for a given soil moisture treatment, obtained by LSD (Fisher’s  
 448 Least Significant Difference).

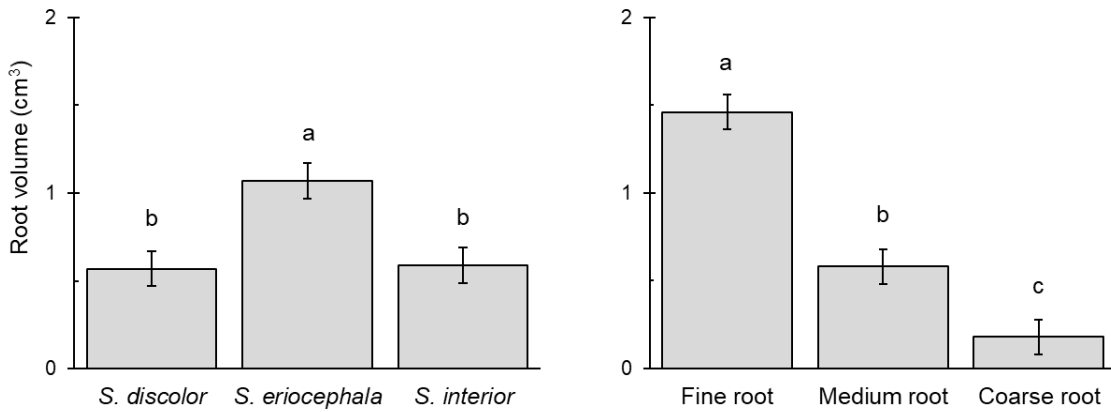
449



450

451 **Fig. 3.** Variations in root volume (mean  $\pm$  standard error) between the three willow species  
452 studied (*Salix discolor* – *S. eriocephala* – *S. interior*; n = 36 cuttings per species) and  
453 between the three categories of root size (all species pooled; n = 36 root subsample per  
454 category). Letters indicate significant differences obtained by LSD (Fisher’s Least  
455 Significant Difference).

456



457



458 **Supplementary material**

459 **Table S1.** Location of the natural willow populations sampled for cutting collection. For  
 460 each species and site, figures indicate the number of cuttings harvested / of individuals  
 461 sampled.

462

463

Location	Coordinates	Species collected		
		<i>S. discolor</i>	<i>S. eriocephala</i>	<i>S. interior</i>
Base de plein air de Sainte-Foy	46°47'03.9"N 71°20'20.2"W	12 / 4	15 / 5	9 / 3
Rivière Cap-Rouge	46°45'57.1"N 71°21'23.2"W	6 / 2	6 / 2	
Domaine Pointe-de-Saint-Vallier	46°54'29.5"N 70°47'27.6"W	12 / 4		
Trait-Carré	46°51'34.4"N 71°16'26.2"W	6 / 2	6 / 2	
Parc des Chutes-de-la-Chaudière	46°43'02.1"N 71°16'55.3"W		9 / 3	6 / 2
Parc de Duberger	46°49'01.3"N			3 / 1
Rivière Saint-Charles	46°49'54.9"N			18 / 6

464

465 **Supplementary material**

466 **Figure S1.** Illustrations of the greenhouse split-plot experimental design. Cuttings were  
467 planted into 30L bag (subplot) and grouped by six into plastic tanks (main plot) to which  
468 soil moisture treatments were applied, for a total of two cuttings per species and per tank.  
469



470