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Macrophages as determinants and regulators of fibrosis in systemic sclerosis

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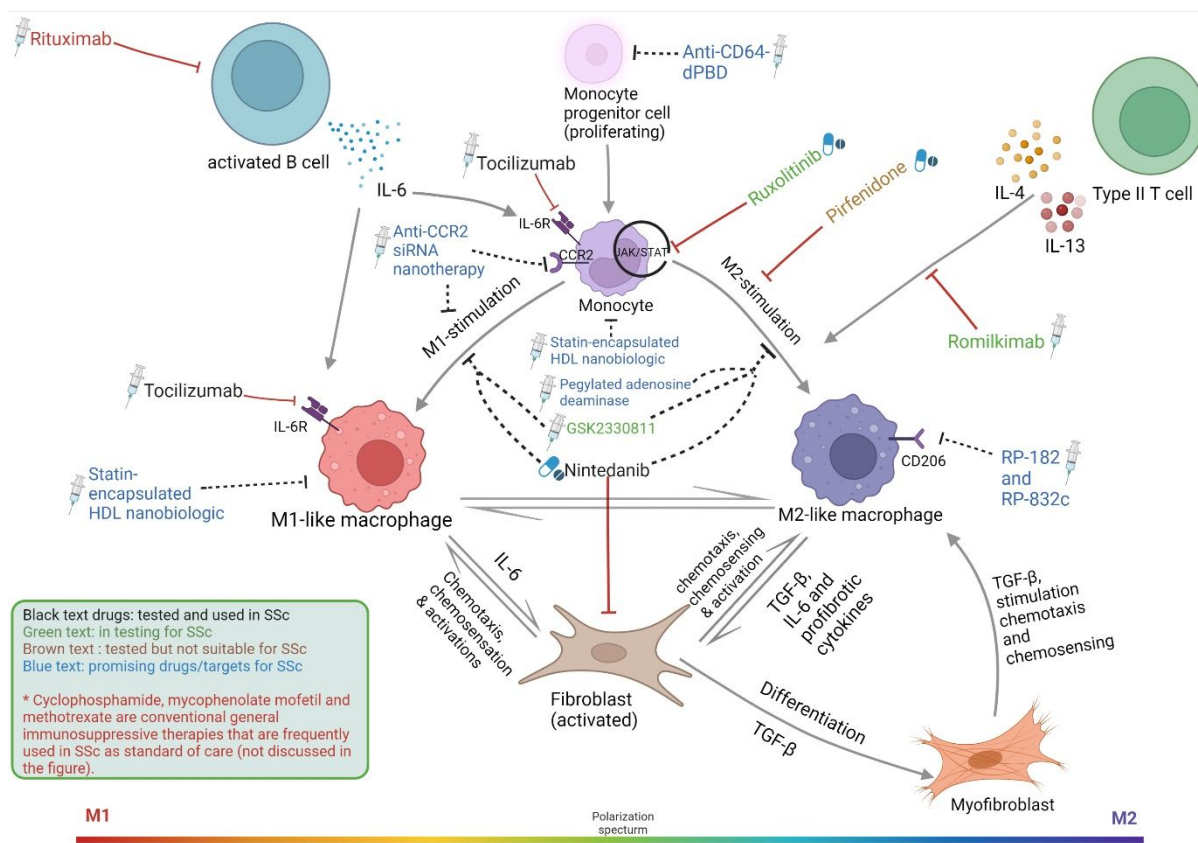
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Abstract

Systemic Sclerosis (SSc) is a multiphase autoimmune disease with a well-known triad of clinical manifestations including vasculopathy, inflammation and fibrosis. Although a plethora of drugs has been suggested as potential candidates to halt SSc progression, nothing has proven clinically efficient. In SSc, both innate and adaptive immune systems are abnormally activated fuelling fibrosis of the skin and other vital organs. Macrophages have been implicated in the pathogenesis of SSc and are thought to be a major source of immune dysregulation. Due to their plasticity, macrophages can initiate and sustain chronic inflammation when classically activated while, simultaneously or parallelly, when alternatively activated they are also capable of secreting fibrotic factors. Here, we briefly explain the polarization process of macrophages. Subsequently, we link the activation of macrophages and monocytes to the molecular pathology of SSc, and illustrate the interplay between macrophages and fibroblasts. Finally, we present recent/near-future clinical trials and discuss novel targets related to macrophages/monocytes activation in SSc.

Graphical Abstract

Created with BioRender.com



Key words: scleroderma, systemic sclerosis, macrophages, monocytes, fibrosis, potential targeted therapeutics.

Key messages:

- Systemic sclerosis (SSc) is a heterogeneous disease and monocytes/macrophages are central within this heterogeneity.
- Plasticity of monocytes/macrophages allow them to reflect and affect all disease phases of SSc.
- There is an urgent unmet need for personalized medicine to treat SSc patients.

56

Systemic Sclerosis

58 Systemic sclerosis (SSc) is an autoimmune disease that involves microangiopathy, early inflammation
59 and progressive fibrosis of the skin and internal organs (1–6). Limited cutaneous (lcSSc), diffuse
60 cutaneous (dcSSc) and sine SSc forms of SSc can present as stable conditions but can also progress to
61 severe disease modes with increased morbidity and mortality (7–9). Although advances have been
62 made in biomarker discovery for SSc, it is still difficult to predict which patients are going to progress
63 to severe fibrotic disease for which no disease-modifying treatment is currently available (9,10).
64 Interstitial lung disease, a result of inflammatory and fibrosing processes, is the leading cause of
65 mortality in SSc patients who develop this complication (~50%) (10,11). Therefore, new treatment
66 strategies are urgently needed to attenuate progression and potentially modify the disease course.
67 Recently, macrophages have captured the interest of the SSc scientific community. This interest is due
68 to the abundance of these cells in affected tissues and their potential in driving both inflammatory, as
69 well as fibrotic processes (12). Macrophages possess great plasticity which allows them to adopt
70 different polarization states.

71

72 This review focuses on the polarization dynamics of macrophages and their precursors (monocytes) in
73 SSc, the influence of monocytes and macrophages on the disease course and the role of cytokines in
74 the activation of macrophages in SSc patients. We also discuss the interplay between activated
75 macrophages and fibroblasts. Furthermore, we describe the heterogeneity of the disease and what a
76 multi-phased disease means for the clinic when it comes to treatment. Finally, we discuss how
77 focussing on macrophage polarization could potentially facilitate novel targeted therapy discovery for
78 SSc patients.

79

Macrophage in health and disease

81 Macrophages play a principal role in maintaining (physiological) homeostasis by engulfing, degrading
82 and clearing of cellular debris, dead cells, and cancer cells (13). Additionally, macrophages function as

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3 83 reparatory machines, playing an essential role in the wound healing process, allowing quicker post-
4 84 insult recovery (14). Their capacity of releasing chemoattractants and cytokines to recruit other
5 85 effector immune cells, make them crucial in terms of host defence response (15).
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11 87 Macrophage tissue infiltration is a known phenomenon in most autoimmune diseases including SSc
12 88 (16–24). Accumulating evidence has revealed a crucial role of innate immune reactions in driving not
13 89 only disease flare ups (causing progression) but also contributing to the ignition processes in such
14 90 diseases (25,26).
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19 91

20 21 92 Macrophage polarization

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23 93 Monocytes are circulating cells, composing around 10% of the cells in healthy peripheral blood, that
24 94 are known to be precursors of macrophages and dendritic cells forming the mononuclear phagocytic
25 95 system. CD14 is expressed on the surface of monocytes and is used as a marker to identify them. A
26 96 complex network of stimuli including cytokines, chemokines and inter-cellular signalling is coordinated
27 97 in a healthy individual to regulate the differentiation of monocytes to macrophages (27,28).
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34 99 Historically, macrophages were classified according to their activation pattern into classically activated
35 100 macrophages (M1) or alternatively activated macrophages (M2). Certain cytokines will
36 101 polarize/differentiate macrophages into a pro-inflammatory phenotype (classical) which handles
37 102 pathogen destruction. On the other side of the spectrum, another set of cytokines, chemokines and
38 103 hormones skew the activation of macrophages into a healing/regenerative phenotype (alternative)
39 104 which are (pro)fibrotic, anti-inflammatory and in charge of tissue repair (29,30). The contribution of
40 105 specific factors will be explored later in this review. Recent scientific observations on macrophage
41 106 classification confirmed that the earlier nomenclature is based on *in vitro* experimentation and does
42 107 not represent *in vivo* scenarios. Thus, macrophage polarization is rather considered as a continuum
43 108 than two distinct populations where classical activation is at one end and alternative activation is at
44 109 the other. Therefore, macrophage phenotypes can slide across this spectrum of the
45 110 classical/alternative paradigm of activations (31).
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51 112 Classically activated macrophages maintain inflammation as a defence mechanism to ward off
52 113 intruders. To be able to function in that manner, classically activated macrophages express a distinct
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3 114 set of surface receptors (see table.1) that allow them to respond adequately to specific stimuli (27,30).
4
5 115 An important stimulus for macrophage polarization towards the classical phenotype is interferon-
6
7 116 gamma (IFN- γ) through its ability to directly activate effector genes including antiviral proteins,
8
9 117 microbicidal molecules, phagocytic receptors, chemokines and cytokines (32). Additionally, IFN- γ can
10
11 118 indirectly activate macrophages by enhancing their reaction to other stimuli through what is known
12
13 119 as “priming” (32). When macrophages are stimulated with IFN- γ , the result is Janus kinase 1 (JAK 1)
14
15 120 and JAK 2 activation, and signal transducer and activator of transcription 1 (STAT1)/interferon
16
17 121 regulatory factors (IRF) signalling, leading to differentiation to the classical phenotype as the product
18
19 122 (33). This activation pathway is not the only means to produce classical macrophages. During bacterial
20
21 123 infections, lipopolysaccharide (LPS) is present abundantly in the body which is well-recognized for
22
23 124 stimulating the classical polarization through its binding with toll-like receptors 2 and 4 (TLRs) which
24
25 125 in turn initiates nuclear factor-light-chain-enhancer of activated B cells (NF- κ B), activator protein-1
26
27 126 (AP-1), IRF and STAT1 signalling (34). Finally, granulocyte-macrophage colony-stimulating factor (GM-
28
29 127 CSF) is capable of inducing the classical phenotype through the activation of the JAK2 pathway (35)
30
31 128 (Fig.1).

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33 129
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35 130 On the other side of the spectrum lies the alternatively activated macrophage phenotype. The central
36
37 131 aim of alternative macrophages is to release anti-inflammatory cytokines and recruit specific tissue-
38
39 132 regenerating cells (36,37). Activated adaptive immune cells such as mast cells, basophils and type 2 T
40
41 133 helper (T_{H2}) cells release IL-4/IL-13 which, in turn, stimulate alternative polarization through the JAK 1
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43 134 and JAK3/STAT6 pathway. This pathway is considered to be the canonical pathway for alternative
44
45 135 activation (38). However, to have a more discrete nomenclature within the alternative phenotype, the
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47 136 IL-4/IL-13-induced activation of macrophages is named M2a (39). Other specific alternative subtypes
48
49 137 can be induced by other stimuli such as immune complexes and TLR ligands (40). Such interactions
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51 138 shut down the proinflammatory cytokine IL-12 release and substitute it with the profibrotic cytokine
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53 139 IL-10. This macrophage-activation state involves spleen tyrosine kinase (Syk) and phosphoinositide 3-
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55 140 kinase (PI3K) activation and is known as M2b. The M2b macrophages can release both pro-
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57 141 inflammatory and anti-inflammatory cytokines (41) (see table.1). Both glucocorticoids and IL-10 are
58
59 142 able to induce the third subtype of alternative macrophages; M2c through the glucocorticoid receptor
60
143 (GRC) or IL-10R, respectively. The M2c subtype has a strong fibroproliferative cytokine signature
144 releasing IL-10 and TGF- β cytokines. The fourth subtype is M2d macrophages which are activated
145 through the binding of TLR agonists to the Adenosine 2 receptor. Consequently, significant
146 suppression of pro-inflammatory cytokine release and promotion of anti-inflammatory cytokines

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3 147 production occurs (42) (Fig.1). The detailed macrophage polarization pathways, different cytokine
4
5 148 signatures and distinct surface markers have all been previously described elsewhere (43,44).
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7 149

150 Monocyte/Macrophage signature in Systemic Sclerosis

151 Plasticity allows macrophages to influence all phases of SSc. Although limited in number, several
152 studies have investigated the role of macrophage polarization in SSc pathogenesis.
153

154 Higashi-Kuwata et al. utilized flow cytometry and immunohistochemistry (IHC) techniques to show
155 that the blood and skin of SSc, respectively, have higher expression of macrophage fibrotic markers
156 (CD163 and CD204) compared to healthy controls. Flow cytometry was used on isolated peripheral
157 blood mononuclear cells (PBMCs) where CD14 surface marker was used to gate for monocytes and
158 CD163 and CD204 surface markers were used to detect the fibrogenic phenotype. Skin biopsies were
159 stained with antibodies against the pan-macrophage surface markers CD68, CD163 and CD204. They
160 found an enhanced expression of CD163 and CD204 on the PBMCs and in skin biopsies from SSc
161 patients compared to controls. Consequently, the authors suggested that the activation status of
162 monocytes/macrophages in SSc patients is profibrotic compared to controls. However, they did not
163 investigate M1 markers to detect classical polarization. (45). Mathai and colleagues showed that CD14
164 monocytes isolated from PBMCs of SSc-associated interstitial lung disease (SSc-ILD) patients show
165 higher expression of CD163 compared to controls. Interestingly, when CD14+ monocytes were
166 isolated from PBMCs of SSc-ILD patients and treated *in vitro* with LPS, a classical activation inducer,
167 these monocytes were skewed into a more profibrotic pattern in contrast to a proinflammatory
168 profile, with more CD163+, CCL18 and IL-10 expression compared to controls (46). However, as the
169 primary question in this study was whether monocytes from SSc-ILD patients have higher expression
170 of profibrotic markers, inflammatory markers were not studied.
171

172 In a monocyte-derived macrophage (MDMs) *in vitro* transcriptomic study, Morano-Moral et al
173 identified 602 genes that were differentially regulated in SSc patients compared to controls.
174 Upregulated genes were related to hypoxia, glycolysis and mTOR pathways while IFN- γ response
175 pathways were downregulated. This study also highlighted gasdermin A specific variant as an SSc risk
176 factor when upregulated, suggesting that MDMs could be the reason behind dysregulated pyroptosis
177 in SSc. This study robustly links SSc pathogenesis with genetic changes and presents a transcriptomic
178 signature in MDMs of SSc patients (47).
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3 180 It is known that the affected skin of SSc patients is infiltrated with immune cells, especially T cells and
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5 181 macrophages. To understand the recruitment of macrophages to the skin, researchers have studied
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7 182 the chemokine gene expression in affected skin from SSc patients. RT-qPCR data of homogenised skin
8
9 183 showed a higher expression of CCL2, CCL5, CCL18, CCL19 and CXCL13 in dcSSc patients when
10
11 184 compared to control skin. Skin biopsies from dcSSc patients exposed a colocalization of CD163+
12
13 185 macrophage subset with CCL19, strongly suggesting the release of CCL19 from the CD163+
14
15 186 macrophage subset. Moreover, not only was CCL2, an important macrophage recruiting chemokine,
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17 187 expression positively correlated with skin thickening in dcSSc skin but also serum levels of CCL2 were
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19 188 elevated in these patients. These localized and systemic correlations are strongly suggestive of the
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21 189 involvement of the alternative macrophage phenotype in the development of skin lesions in SSc.
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23 190 Another group studied patients with SSc-ILD compared to those SSc patients without ILD. There were
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25 191 increased mixed classic (CD80, CD86, TLR2 and TLR4) and alternative (CD206, CD204 and CD163)
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27 192 circulating monocytes in patients with SSc-ILD. (48). Additionally, they found that these markers were
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29 193 significantly elevated on PBMCs isolated from SSc patients compared to healthy controls (49). These
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31 194 results point out that the circulating monocytes from SSc patients have enriched classic and alternative
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33 195 markers compared to controls while this enrichment is even greater when ILD is present.
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35 196

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37 197 On a single-cell level, RNA-sequencing of SSc-ILD lung tissue revealed several monocyte/macrophage
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39 198 subgroups in which SPP1^{hi} proliferating macrophages were more predominant in SSc-ILD lungs
40
41 199 compared to controls (50). SPP1 macrophages have been attributed to lung fibrosis through the
42
43 200 activation of myofibroblasts in idiopathic pulmonary fibrosis (51) which could be having the same role
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45 201 in SSc-ILD. On the same level, RNA-sequencing of dcSSc skin tissues revealed innate immune system
46
47 202 activation(52). Specifically, macrophages highly expressing Fcy receptor IIIA were only associated with
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49 203 dcSSc skin but not in healthy skin. Importantly, the proliferating macrophages were exclusively
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51 204 detected in dcSSc skin but not in healthy ones (52). Thus, it is plausible that proliferating macrophages
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53 205 are fundamental to skin and lung disease progression in SSc.
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55 206

56
57 207 Due to their plasticity, monocytes/macrophages could be an important link for the transition from the
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59 208 inflammatory to the fibrotic phase in SSc pathology. Perhaps, monocyte/macrophage polarization
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209 shifts along the classic/alternative spectrum of activation to a more fibrotic state over time due to
210 intracellular changes and differential presence of cytokines and chemokines in their environment. The
211 change in the cytokine and chemokine profile can be attributed to reactive B cells (IL-6 release) and
212 activated CD4+ T_{H2} cells (IL-13 and IL-4 release) (53,54). Consequently, monocytes/macrophages
213 become profibrotic and start to release fibrotic factors that lead to the activation of more

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3 214 monocyte/macrophages (and of fibroblasts) into the profibrotic phenotype generating an autocrine
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5 215 loop. Moreover, the mounting recruitment of fibrogenesis-effector cells such as fibrocytes and
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7 216 fibroblasts into affected tissues, and their activation by the released fibroproliferative chemokines are
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9 217 key events in SSc- related tissue fibrosis.

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12 13 219 The interplay between fibroblasts and macrophages

14 220 Activation of monocytes/macrophages is crucial for stimulation of the fibrosis effector cells
15 221 (fibroblasts) in affected tissues. Bhandari et al. (55) showed that the activation of fibroblasts is
16 222 dependent on SSc plasma-differentiated macrophages. SSc plasma significantly activated monocytes
17 223 from both control and SSc groups into the profibrotic (alternative) phenotypes when compared to
18 224 monocytes cultured with control plasma. Moreover, significantly higher mRNA and protein expression
19 225 and production of CCL2, IL-6 and TGF- β were reported in SSc plasma-cultured compared to control-
20 226 cultured monocytes. This experiment illustrated that SSc plasma can differentiate control monocytes
21 227 into SSc phenotype macrophages. Additionally, RT-qPCR data from dermal fibroblasts revealed
22 228 overexpression of α -SMA in SSc fibroblasts co-cultured with SSc plasma-differentiated macrophages,
23 229 compared to healthy dermal fibroblasts co-cultured with SSc plasma-differentiated macrophages.
24 230 These data indicate that SSc macrophages induce and activate dermal fibroblasts into becoming
25 231 fibrogenic cells through fibroblast to myofibroblast transdifferentiation (55).

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27 232

28 233 In *in vivo* scenarios, macrophages have to be in close proximity to fibroblasts to stimulate them to
29 234 myofibroblasts (56). Pakshir et al (57) described how mechanosensation and integrins help
30 235 myo/fibroblasts attract macrophages to the vicinity of the fibrotic niche. Fibroblasts establish ECM
31 236 cues through remodeling collagens to form deformation fields in the collagen mesh which guides
32 237 macrophages to come closer to myo/fibroblasts. Importantly, these ECM alterations have more far-
33 238 reaching effects than chemotaxis.

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36 239 Thus, the interaction between macrophages and myo/fibroblasts is necessary to establish a
37 240 progressive fibrotic niche.

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44 244 Emerging role of oxidative stress in monocyte/macrophage polarization in SSc

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46 245 Antioxidant/oxidant imbalance is thought to be connected to SSc pathogenesis (58) The nuclear
47 246 factor erythroid 2 (NF-E2)-related factor 2 (Nrf-2) is an important cellular sensing protein for

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3 247 oxidative stress which- in turn- can stimulate the transcription of antioxidants including glutathione
4 248 (GSH). In an SSc mouse model, Nrf^{-/-} knockdown and wild-type (control) mice were intradermally
5 249 injected with hypochloric acid (a substance to induce oxidative stress). The Nrf^{-/-} mice showed more
6 250 severe inflammation and fibrosis than controls. Importantly, in the skin of hypochloric acid-treated
7 251 mice, the Nrf^{-/-} type had a more pronounced M2 polarization marker profile than the wild-type
8 252 mice. This indicates that oxidative stress induces a shift towards M2 polarization and suggests a
9 253 strong link between Nrf-2 function, alternative polarization, fibroblasts activation and fibrogenesis
10 254 (59).

11 255
12 256 **Systemic Sclerosis is a multi-phase disease – Interventional remarks focusing on targeting**
13 257 **monocytes/macrophages** (summary of current and prospective/promising SSc therapeutics
14 258 is stated in table.2)

15 259
16 260 SSc-investigated targeted therapies

17 261 In SSc patients who present early in their disease course, inflammation is generally the predominant
18 262 process activated, especially in progressive dcSSc. As elaborated, M1 phenotype macrophage
19 263 activation may be central at this stage, and it would be reasonable to introduce drugs that target the
20 264 effector pathways early. In patients with SSc-ILD, monocytes are known to produce higher amounts
21 265 of IL-6 compared to healthy controls (60). This high production is strongly associated with SSc
22 266 pathogenesis since it leads to the activation, differentiation and proliferation of T lymphocytes.
23 267 Tocilizumab (anti-IL-6R monoclonal antibody) is of growing interest and use in clinical practice. It is
24 268 FDA-approved to slow the rate of decline of lung function in adult patients with SSc-ILD (61,62).
25 269 Moreover, IL-6 is abundantly produced by activated B cells expressing CD20. The DESIRES RCT showed
26 270 that using rituximab, an anti-CD20 monoclonal antibody, in SSc patients resulted in a significant
27 271 reduction in mRSS compared to the placebo arm (63). Rituximab's positive results can be indirectly
28 272 attributed to the blockage of macrophage polarization leading to mitigated skin and lung disease
29 273 (64,65).

30 274
31 275 An IFN type I serum profile is related to higher mRSS and HAQ-Disability Index. Researchers have
32 276 indirectly targeted IFN type I by targeting CD52. In a translational study, transcriptomic analyses of
33 277 circulating CD14⁺ monocytes obtained from SSc patients revealed enhanced expression of IFN I-
34 278 related genes compared to healthy controls. These monocytes also displayed down-regulated CD52

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3 279 expression which is an important T cell inhibitory antigen. Interestingly, when healthy monocytes were
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5 280 treated with an anti-CD52 antibody, enhanced activation of IFN I pathways were achieved.
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7 281 Consequently, targeting the CD52-IFN I pathway is a promising approach in early SSc patients (66).
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11 283 Targeting the profibrotic cytokines IL-4 and IL-13 to prevent further activation of
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13 284 monocytes/macrophages to the profibrotic forms has shown promising results. Romilkimab was
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15 285 developed as a humanized bispecific mAB against both IL-4 and IL-13. When neutralizing these serum
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17 286 elevated cytokines, the paracrine and autocrine activation loops of macrophages are blocked. Indeed,
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19 287 a phase 2 RCT in early dcSSc was performed where romilkimab efficacy was tested vs placebo. After
20
21 288 24 weeks, patients who have been treated with romilkimab had a significant improvement in their
22
23 289 mRSS compared to the placebo group (67).
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25 290
26 291 Although it is known to work as a multi tyrosine kinase inhibitor, nintedanib also functions by
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28 292 disturbing the expression of surface markers, and/or the chemokine and cytokine signature of
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30 293 monocyte-derived macrophages. It also inhibits the phosphorylation of the colony-stimulating factor
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32 294 1 receptor in monocyte/macrophages which is essential in activation and polarization of these cells.
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34 295 When monocytes were stimulated to polarize to classical or alternative macrophages *-in vitro*-
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36 296 subsequent to treatment with nintedanib, several alterations were observed. First, classical
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38 297 macrophages continued to express classical surface markers at the same level as untreated
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40 298 macrophages but released significantly less proinflammatory cytokines. Second, the alternatively-
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42 299 stimulated macrophages had a significant decrease in their M2 markers while their profibrotic
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44 300 cytokines and chemokines release remained comparable to untreated alternative macrophages
45
46 301 (68,69). Clinically, Azuma et al. (70) performed a phase 3 RCT where SSc patients with at least 10%
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48 302 lung fibrosis on HRCT were included. The primary endpoint was the annual rate of decline in FVC. After
49
50 303 52 weeks, the annual rate of decline in FVC was significantly higher in the placebo arm than in the
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52 304 treatment arm. Based on these data, nintedanib was the first drug to be approved for treating SSc-ILD
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54 305 (71–73).
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58 307 Pirfenidone has shown inhibitory effects on rat alternatively-activated lung macrophages cultured *in*
59
60 308 *vitro*. This was demonstrated by a significant reduction of TGF- β release and lower expression of M2
61
62 309 surface markers when macrophages were treated with pirfenidone. When the supernatant of the
63
64 310 pirfenidone-treated macrophages was used to treat rat lung fibroblasts, suppressed proliferation, and
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66 311 collagen mRNA expression and production were observed in these fibroblasts (74). In light of these

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3 312 data, Khanna et al. (75) and Sharma et al. (76) performed phase 2 clinical trials to assess the efficacy
4 313 of pirfenidone in SSc-ILD patients. Although the data from these trials have not shown significant
5 314 differences in lung function decline between the treatment and placebo groups, pirfenidone was well
6 315 tolerable and appeared likely to maintain lung function better than the placebo.
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10 316

11 317 Current studies

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13 318 Several studies are currently investigating pharmaceutical agents that could hinder the activation
14 319 and/or the release of cytokines/chemokines from monocytes/macrophages. For example, in the “Hit
15 320 hard and early” study [NCT03059979], very early diagnosed SSc (VEDOSS) patients are being treated
16 321 with high dose methylprednisolone, potentially preventing early vasculopathy by forcing attenuation
17 322 of inflammation (77). It is also highly plausible that the mechanism behind this strategy is mitigating
18 323 the polarization of macrophages towards the classical inflammatory phenotype, as this is a known
19 324 effect of prednisolone (78,79). Another strategy is being investigated using upfront autologous
20 325 hematopoietic stem cell transplantation (AHSCT) in early dcSSc patients with the aim of resetting the
21 326 immune system (the UPSIDE study; [NCT04464434]) (80). Monocytes derived from myeloid
22 327 progenitors are activated and play a role in igniting and perpetuating the inflammatory, and thereafter
23 328 the fibrosis processes, in SSc patients. Studies show that dcSSc patients have higher expression of
24 329 CD16+ monocytes which are known to have enhanced pro-inflammatory activation (81). The upfront
25 330 depletion of these cells, coupled with replacing them with “normal/undiseased” precursors through
26 331 AHSCT could yield monocytes that can suppress the pathogenetic pathways of SSc by enhancing T_{regs}
27 332 cell production, inhibiting fibroblast to myofibroblast transdifferentiation and suppressing CD4 T cells
28 333 proliferation (82).
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43 335 The SCLERO JAK [NCT04206644] study is investigating the efficacy of the JAK 1/2 inhibitor ruxolitinib
44 336 in SSc patients. one of the outcomes aims to gather a greater understanding of the impact of this drug
45 337 on the activation states of monocyte-derived macrophages obtained from SSc patients. It is
46 338 hypothesized that blocking the JAK-STAT pathway would attenuate the profibrotic properties of
47 339 monocyte-derived macrophages. This will be tested in an *in vitro* model by measuring CCL18 levels in
48 340 the culture media of ruxolitinib-treated SSc macrophages compared to untreated (as a primary
49 341 outcome). In addition, macrophage surface markers studies will be performed.
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58 343 GSK2330811 is a humanized monoclonal antibody against the oncostatin M (OSM) protein, which is
59 344 implicated in inflammation, fibrosis and vasculopathy, typical features of SSc pathogenesis. Activated

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3 345 monocytes/macrophages are known to produce OSM which alters fibroblasts -among other
4 346 connective tissue cells- production of cytokines and chemokines such as MCP-1 and IL-6 which in turn
5 347 affects the polarization of macrophages (paracrine activation loop) (83,84). After showing a well-
6 348 tolerated safety profile in phase I clinical trial (85), proof of mechanism phase II randomized clinical
7 349 trial [NCT03041025] in dcSSc patients is currently being undertaken and it is hoped the data will be
8 350 available soon.

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14 351

15
16 352 Future perspectives – promising therapeutics/pathways

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18 353 SSc upstream processes involve both the innate and adaptive immune systems. repurposing drugs
19 354 from other medical fields such as oncology and haematology to the field of autoimmune diseases is
20 355 not unusual. Therefore, we are suggesting the following potential drugs abide the recent
21 356 understanding of SSc pathogenesis.

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23 357

24
25 358 Targeting purinergic signalling may ameliorate fibrosis. As explained above, adenosine can skew
26 359 macrophages towards the alternative phenotype. Degradation of adenosine using pegylated
27 360 recombinant adenosine deaminase reduced fibrogenesis in SSc preclinical models. Adenosine
28 361 deaminase has also shown promising results regarding vasculopathy and inflammation in a mouse
29 362 model of SSc (86). The effects of such a drug should be examined in a clinical trial to better
30 363 comprehend its potential efficacy in SSc patients.

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32 364

33 365 Direct targeting of proliferating monocyte progenitor cells without affecting other progenitor cells or
34 366 mature monocytes could be an approach to diminish monocytes' contribution to pathophysiology in
35 367 SSc patients. Using dimeric pyrrolobenzodiazepine (dPBD)-conjugated anti-CD64 antibody (anti-CD64-
36 368 dPBD), Izumi et al (87) were able to selectively induce apoptosis in proliferating human monocyte
37 369 progenitors.

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40 371 Targeting the migration of inflammatory monocytes to sites of injury using small interfering RNA
41 372 (siRNA) is another promising approach that could benefit SSc patients. CCR2 chemokine receptor is
42 373 known to be over-expressed on inflammatory monocytes. Targeting cells with high levels of this
43 374 receptor with nanoparticles-containing anti-CCR2 siRNA showed promising results in several
44 375 inflammatory diseases in preclinical settings. In these preclinical models, anti-CCR2 siRNA was able to
45 376 silence CCR2 mRNA of inflammatory monocytes and consequently reduced migration as well as
46 377 numbers of monocyte-derived macrophages without affecting other healing, physiologically essential
47 378 functions of monocytes and associated macrophages (88).

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380 Non-specific memory of the innate immune system, also as known as “Trained immunity”, is thought
381 to be part of the enhanced and continuity of cytokines and chemokines production by monocytes in
382 SSc patients. Pharmacological blocking of upstream processes of trained immunity using NOD2 and
383 dectin 1 inhibitors, GSK669 or laminarin, respectively could be beneficial (89). Additionally,
384 nanomedicine could offer another novel approach for directly targeting and skewing localized
385 inflammatory monocytes to a less inflammatory phenotype through limiting epigenetic and metabolic
386 changes (89). Statin-encapsulated reconstituted high-density lipoprotein (HDL) nanobiologic is a
387 promising tool targeting inflammatory monocytes and macrophages. Such a drug has shown
388 promising results in inflammatory atherosclerotic plaques (90) but has not yet been applied to
389 autoimmune diseases including SSc.

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391 Reprogramming alternatively-activated macrophages towards apoptosis or classical polarization is a
392 well-characterized strategy in tumour research. RP-182 and RP-832c, host immune peptides, can
393 target CD206 alternative MDMs in lung fibrosis leading to alleviation of fibrogenesis. This mechanism
394 could be beneficial for SSc patients, especially for those who are suffering from dermal and lung
395 fibrosis (91,92).

396

397 Conclusion

398 SSc is a multi-organ, multi-phase disease with various potential pharmaceutical interventions. In order
399 to combat its complications, it is first necessary to identify the phase of the disease. Interpretations of
400 literature and previous research highlight monocytes/macrophages as promising biomarkers that
401 dynamically change according to disease progression reflecting disease status. They can also be
402 considered as potential therapeutic targets through modulation of their polarization.

403

404 Due to the heterogeneity of SSc pathogenesis, examination of SSc patients must recognise that each
405 patient is a unique case. This is a unique opportunity to address the unmet need for personalized
406 medicine in treating SSc patients. Most SSc complications share similar phenotypical and molecular
407 characteristics, however, several important differences have been observed when it comes to
408 progression and initiating factors. Finally, although this personalized approach is still under
409 development, each SSc patient requires a special set of therapeutics according to their disease phase,
410 active pathogenesis pathway, and number and type of complications.

411

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11
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15 420 Data availability statement

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17 421 Data are available upon reasonable request.
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28 427 *Table.1:* Differences in surface markers and cytokine signatures in classical and alternative
29 428 (phenotypes) macrophages
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Macrophage phenotype	Surface markers	Cytokine signature
Classical	CD86, CD68, CD80, MHC-II, TLR-2, TLR-4, IL-1R	IL-1 β , IL-6, IL-12, IL-18, IL-23, TNF- α
Alternative (M2a)	CD206, CD36, CD163, IL-1R	IL-10, TGF- β
Alternative (M2b)	CD86, MHC-II	TNF- α , IL-1 β , IL-6, IL-10
Alternative (M2c)	CD206, TLR-1, TLR-8, CD163	IL-10, TGF- β
Alternative (M2d)	CD206, CD204, CD163	IL-10, VEGF

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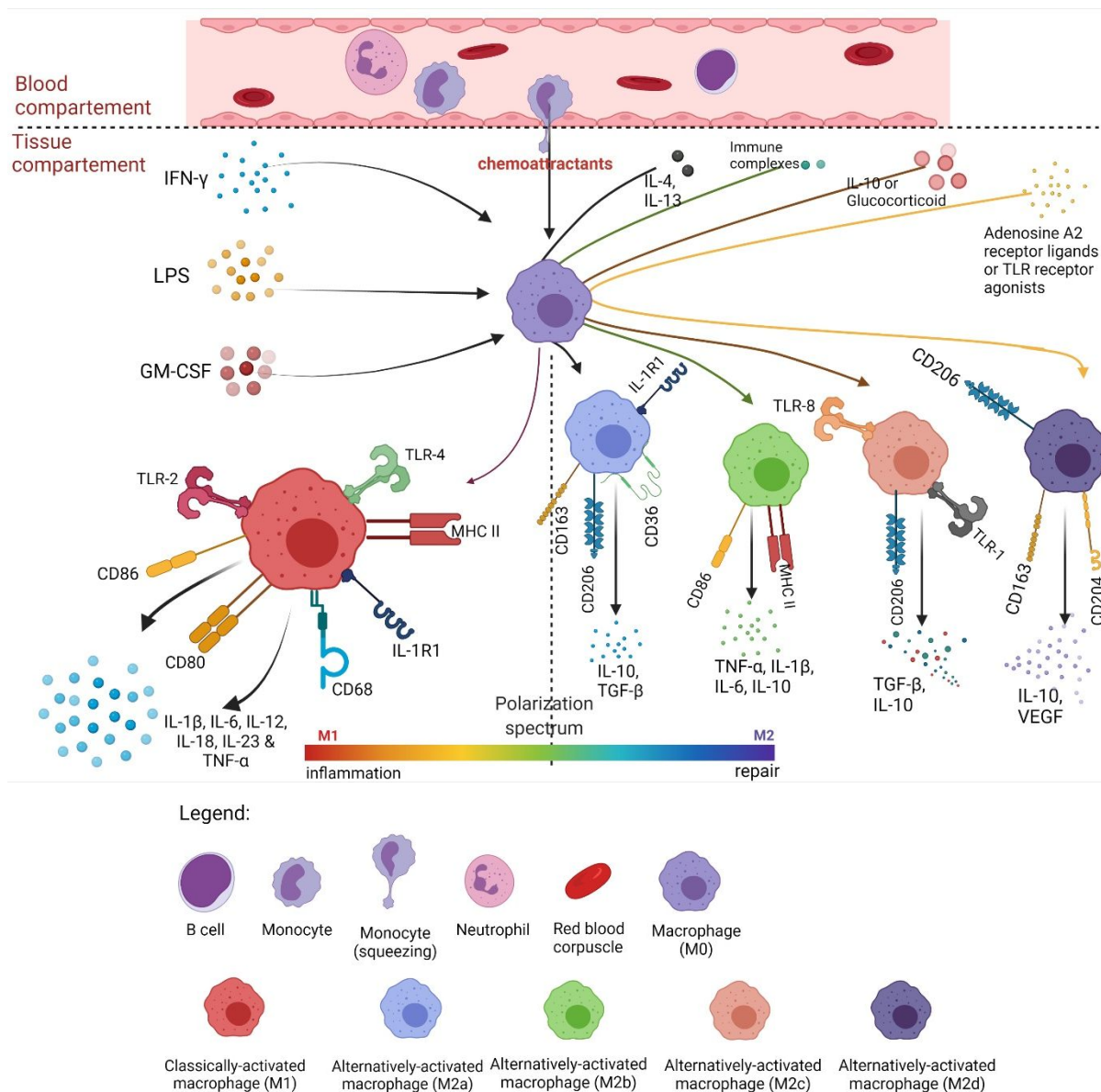
57 437 *Table.2:* SSc-investigated targeted therapies and suggested novel monocyte/macrophage targeted
58 438 therapies in the treatment of systemic sclerosis and systemic sclerosis-associated interstitial lung
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Therapeutic	Target (action)
AHSCT	resetting myeloid progenitor cells including monocytes
Tocilizumab	Anti-IL-6 receptor α -subunit (attenuates monocyte downstream effects)
Rituximab	Anti-CD20 "B cells depletion" (attenuates downstream macrophage polarization)
Romilkimab	Anti- IL-4 and IL-13 cytokines (blocks alternative activation)
Nintedanib	Multi-tyrosine kinase inhibitor (disturbs classical and alternative activations)
Pirfenidone	Blocks alternative activation
Ruxolitinib	JAK 1/2 inhibitor (proposed to attenuate alternative activation)
GSK2330811	Anti-oncostatin M protein (attenuates monocytes downstream effects)
Pegylated adenosine deaminase	adenosine molecules (blocks alternative polarization)
anti-CD64-dPBD	proliferating monocyte progenitors
anti-CCR2 siRNA nanotherapy	inflammatory monocytes migration
Statin-encapsulated HDL nanobiologic	inflammatory monocytes systemically and inflammatory macrophages locally
RP-182 and RP-832c	CD206+ cells "alternatively-activated macrophages"

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Figure 1: Macrophage polarization.

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Monocytes are attracted to injury sites (e.g., infected tissue) by chemoattractants. Thereafter, macrophages are activated dependent on the cytokines/stimuli in the milieu to M1 (classic activation) or M2 (alternative activation). Polarization state of macrophages is rather a dynamic process where macrophages can shift along a polarization spectrum. Activated macrophages express a specific set of (surface) markers and release particular collection of cytokines according to the activation pattern which in turn affects the milieu. CD: Cluster of Differentiation, IL: Interleukin, TGF- β : Transforming Growth Factor-beta, TLR: Toll-Like Receptor, IFN: Interferon, MHC II: Major Histocompatibility Complex class II, VEGF: Vascular Endothelial Growth Factor, TNF- α : Tumor Necrosis Factor-alpha, LPS: Lipopolysaccharide, GM-CSF: Granulocyte-Macrophage Colony-Stimulating Factor. Created with BioRender.com

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