

Article Info

Review Article

Impact of carotenoids on gut microbiome: Implications in human health and disease

Pramukh Subrahmanya Hegde

Nitte (Deemed to be University), KS Hegde Medical Academy (KSHEMA), Department of Physiology, Mangalore-575018 (Karnataka), India

Megha Bhat Agni

Nitte (Deemed to be University), KS Hegde Medical Academy (KSHEMA), Department of Physiology, Mangalore-575018 (Karnataka), India

Praveen Rai

Nitte (Deemed to be University), Nitte University Centre for Science Education and Research (NUCSER), Division of Infectious Diseases, Mangalore-575018 (Karnataka), India

Mohana Kumar B

Nitte (Deemed to be University), Nitte University Centre for Stem Cell Research and Regenerative Medicine (NUCSReM), Mangalore-575018 (Karnataka), India

Damodara Gowda KM*

Nitte (Deemed to be University), KS Hegde Medical Academy (KSHEMA), Department of Physiology, Mangalore-575018 (Karnataka), India

*Corresponding Author. E-mail: dr_damodar@nitte.edu.in

How to Cite

Hegde, P. S. et al. (2022). Impact of carotenoids on gut microbiome: Implications in human health and disease. Journal of Applied and Natural Science, 14(3), 1085 - 1099. https://doi.org/10.31018/jans.v14i3.3582

Abstract

Carotenoids are the colored compounds that prominently occur in fruits, vegetables, flowers, algae, fungi, yeast, and marine organisms. The coloration of carotenoids is mainly due to varieties of conjugated double bonds, which act as a light-absorbing chromophores. β -Carotene, α -Carotene, Lycopene, Astaxanthin, Lutein, Zeaxanthin, β -Cryptoxanthin, α -Cryptoxanthin, γ -Carotene and Fucoxanthin are the common carotenoids of the human diet. This review aimed at providing scientific evidence supporting the benefits of nutritional carotenoid intake on gut microbiota modulation in different disease models. Carotenoids have some beneficial effects on human health, and it is due to the activity of pro-vitamin A and antioxidant function. Although mechanisms are under investigation, studies suggest that carotenoid intake may reduce the risk of cancer, cardiovascular disease, eye disease, haematological disease, immune stimulants, and improve cognitive function. Recent studies have shown that carotenoids can modulate gut microbiota composition associated with host health. The human gut harbors a complex community of over 100 trillion microbial cells, influencing human physiology, metabolism, nutrition, and immune function. The combination of extrinsic (lifestyle and medication) and intrinsic (host genetics, immune and metabolic regulations) factors shapes the gut microbiota. Diet is a crucial modifiable factor influencing gut microbiota composition, indicating the potential for therapeutic dietary strategies to manipulate microbial diversity, design, and stability.

Keywords: Astaxanthin, β-carotene, Carotenoids, Fucoxanthin, Gut- microbiome, Gut dysbiosis, Human health, Lycopene.

INTRODUCTION

"Carotenoids" are a collection of red, orange, and yellow-pigmented molecules that are synthesized by photosynthetic plants, algae, bacteria, and fungi. Around 700 specific carotenoids are identified in plant species and some species of algae and fungi. Carotenoid is a common term used to signify the non-nitrogenous biochromes, universally dispersed in residing entities in a lipophilic environment and is commonly insoluble in

water (Zia-ul-haq et al., 2021).

Animals do not often produce carotenoids, although some animals (e.g., aphids) were proven to synthesize them *de novo*. Up to date, ~ 700 Carotenoids have been described, of which approximately 50 turn out to be components of the human diet, while only ~20 are found in human blood and tissues (Fiedor and Burda, 2014). The most important include β -Carotene, α -Carotene, Lycopene (LYC), Lutein, Zeaxanthin, β -Cryptoxanthin, α -Cryptoxanthin, γ -Carotene, Astaxan-

This work is licensed under Attribution-Non Commercial 4.0 International (CC BY-NC 4.0). © : Author (s). Publishing rights @ ANSF.

https://doi.org/10.31018/ jans.v14i3.3582 Received: June 11, 2022 Revised: September 8, 2022 Accepted: September 13, 2022 thin (AsX), Fucoxanthin (Fx), Neurosporene, ζ -Carotene, Phytofluene, and Phytoene.

Structure and classification of carotenoids

The International Union of Pure and Applied Chemistry (IUPAC) defines a carotenoid as a compound consisting of C-20 and C-20' intact methyl groups, with a structure characterized by the conjugated double-bond system that determines the colour (it acts as a lightabsorbing chromophore). As the conjugated doublebond increases, colour varies from light yellow to orange to red. Traditionally, carotenoids are named according to their biological origin.

Carotenoids are classified into two categories according to the presence or absence of oxygen atoms in the molecule (Fig. 1). They are hydrocarbon Carotenes such as β -Carotene, α -Carotene, LYC, and oxygen-containing carotenoids, Xanthophylls, such as Lutein, Zeaxanthin, AsX, etc. Depending on chemical structure, carotenoids can be classified into cyclic carotenoids, acyclic carotenoids, hydroxy carotenoids (or Carotenols), epoxy-carotenoids, and uncommon or species-specific carotenoids. The molecular structure of carotene consists of only carbon and hydrogen and forms a polyunsaturated chain with the chemical formula C₄₀H₅₆ (Olatunde *et al.*, 2020).

Sources of dietary carotenoids

Carotenoids are prominent in algae and photosynthetic bacteria as they can synthesize a variety of carotenoids, and are accumulated in cells called chromatophores (Table 1). Halobacteria (bacteria resistant to high salt concentrations) contains carotenoids in the form of β-carotene and forms a bright red blood cell suspension. Marine organisms, especially microalgae, have become the focus of identifying new biologically active natural substances in the last few years. In particular, it was reported that diatoms are one of the most promising microalgae groups, which possess a series of phytochemicals with beneficial biological activities. For example, the water extract of Haslea ostrearia shows antitumor and antiproliferative effects, while the extract of Chaetoceros calcitrans shows antioxidant effects. Carotenoids are considered one of the main bioactive components in microalgae, however, studies have also shown that other compounds, such as polyphenols and polyunsaturated fatty acids, may impact their health (Galasso et al., 2017).

Microalgae contain many different types of carotenoids. Among them, about 30 species perform functions related to photosynthesis, while other species are intermediates for carotenoid production. Although many microalgae can produce carotenoids, only a few can accumulate sufficient carotenoids suitable for mass production. Only the unicellular green microalga *Haematococcus* pluvialis is cultivated on an industrial scale. These freshwater microalgae exist in temperate regions. Cysts can be formed under certain stress conditions, such as bright light, nitrogen deficiency, and high salt content and a large amount of AsX accumulated in the cytoplasm until the colour changes from green to greenishred. For potential commercial cultivation, Dunaliella salina is another green microalga under study. It adapts to the extreme saline-alkaline conditions where very few organisms can survive and produce a large amount of β -Carotene to protect themselves from the intense penetration and light challenges (Galasso et al., 2017). In marine animals, sponges, sea urchins, krill shells and shrimps, and the scales of various fishes contain the highest amount of carotenoids. The pink to orange coloration of salmon and trout meat is also the result of carotenoid deposition, especially AsX, extracted from krill (Maoka et al., 2011).

Different fungi can produce and store high amounts of carotenoids in cells as microalgae. According to the reports, other species from the phyla *Chytridiomycota, Zygomycota, Chytridiales,* and *Basidiomycotina* have carotenogenesis capacity-like *Cantharellus sp, Phaffia rhodozyma, Blakeslea trispora* and *Rhodotorula rubra.* The mushroom *Cantharellus cinnabarinus* is one of the best examples of the naturally occurring orange-red oxygen-containing carotenoid Canthaxanthin. Another genus, *Rhodotorula* described as a natural source of β -carotene is of interest as a biological source, although its importance is minimal compared to other microorganisms (Echavarri *et al.*, 2002).

Quantitatively, fruits and vegetables contain the maximum amount of natural carotenoids. Carotenoids are abundant in leaves, other green parts, flowers, roots, and seeds in terrestrial plants. Carotenoids in terrestrial plants mainly correspond to red and yellow xanthophylls, such as lutein, zeaxanthin, capsaicin, violaxanthin, or neoxanthin. In addition, marigold flower (*Tagetes standa*) and red pepper (*Capsicum annuum*) are good examples of the natural occurrence of yellow lutein, capsanthin, red xanthophylls, respectively. Many vegetables are rich in carotenes, such as LYC in tomatoes and β -carotene in carrots and sweet potatoes (Mezzomo and Ferreira, 2016).

In the stomach, the carotenoids are d emulsified into small lipid droplets (Fig. 2). From the lipid droplets, carotenoids are transferred to mixed micelles formed by the action of bile salts, biliary phospholipids, dietary lipids, and their hydrolysis products. The mixed micelles migrate to the brush border, where carotenoids are absorbed by the intestinal cells, packed into chylomicrons and secreted to the lymphatic system. The uptake of carotenoids from the intestinal lumen takes place by simple diffusion down a concentration gradient through the brush border membrane into the cytoplasm of the enterocytes mainly through carrier protein SR-BI. The hairpin-like conformation of SR-BI external domain forms a hydrophobic channel that may facilitate the uptake of carotenoids by the enterocytes (Yonekura and Nagao, 2007).

Dietary carotenoids and human health

Carotenoids are not only colored pigments but also biologically active substances that positively affect human health (Fig. 3). β-carotene, cryptoxanthin, zeaxanthin, lutein, and LYC are common dietary carotenoids. This protective effect is due to the activity of provitamin-A and antioxidant function. Epidemiological studies have revealed that regular intake of carotenoids rich in fruits and vegetables reduces the risk of cancer, cardiovascular disease, eye disease, blood disease and enhances immunity (Yonekura and Nagao, 2007). The potential antioxidant properties of carotenoids can help prevent other diseases caused by free radicals. The oxidants produced during normal metabolism and immune defence against chemical substances and infectious pathogens are responsible for damaging DNA, proteins, and cells in the human body. This damage is considered the leading cause of aging and degenerative diseases such as cancer, cardiovascular disease, weakened immune system, and cataracts. The localization of carotenoid molecules in biological tissues also affects their ability to recognize and scavenge free radicals (Bakan et al., 2014).

Carotenoids and gut microbiota

The human gut harbours a complicated network of over a hundred trillion microbial cells that impact human physiology, metabolism, nutrition, and immune function (Guinane and Cotter, 2013). Firmicutes, Bacteroidetes, Actinobacteria, Proteobacteria, Verrucomicrobia, and Fusobacteria are the most common bacterial phyla observed in the human intestine (Rinninella *et al.*, 2019). Gut microbiota (GM) diversity affects numerous metabolic and immune functions of the host's physiology. The GM is formed through a mixture of extrinsic elements (lifestyle and medication) and intrinsic elements (lifestyle and medication) and intrinsic elements like host genetics, immunity, and metabolic regulations. Diet is a crucial modifiable component influencing the composition of the GM, indicating the potential for therapeutic dietary strategies to manipulate microbial variety, composition, and stability. Consequently, an imbalance of GM or dysbiosis can be the reason for many pathological conditions like infectious diseases, gastrointestinal cancers, inflammatory bowel disorder, or even obesity and diabetes (Carrera-Quintanar *et al.*, 2018).

Microbial dysbiosis indicates the depletion of beneficial microorganisms and the growth of potential pathogens (Feher *et al.*, 2021). Therefore, GM is a suitable target for dietary interventions to boost health. Accordingly, carotenoids that may impact GM have recently been studied as adjuvants to remedy a few persistent and inflammatory diseases. Although the health benefits of carotenoids are well reported, only a small part of dietary carotenoids are absorbed from the intestine and eventually enter the body tissues. The rest enter the intestine and are metabolized by the microbiota (Bas-Bellver *et al.*, 2020).

It has also been reported that carotenoids have potential prebiotic effects. Supplementation of carotenoids can increase the microbial community and diversity, leading to overall changes in the microbial community. Especially in humans, higher dietary carotenoids help inhibit many Firmicutes positively associated with overweight/obesity (Guo *et al.*, 2019). However, there is a lack of information about the interaction between carotenoids and the GM. Based on the findings of recently

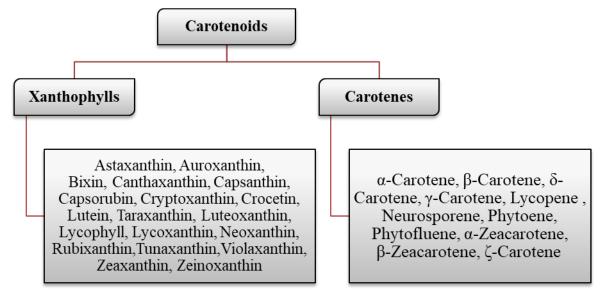


Fig. 1. Classification of carotenoids depending on the presence or absence of oxygen in the molecule (https:// www.tuscanydiet. net/2013/11/02/carotenoids-definition-structure-classification)

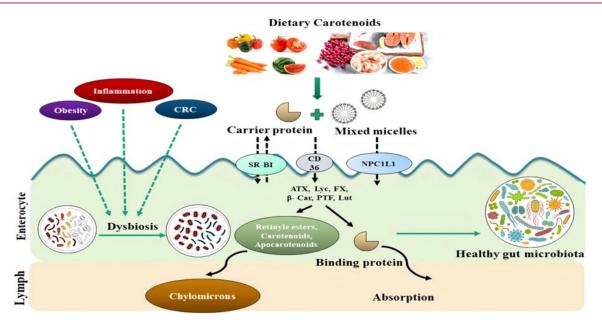


Fig. 2. Absorption of dietary carotenoids and its role on gut dysbiosis (Source: Reboul et al., 2013)

published studies, this review summarizes the interactions between intestinal flora and dietary carotenoids in the course of several disease conditions.

Effect of astaxanthin on gut microbiota

AsX is a dark-red pigment found primarily on marine algae and aquatic animals. It is a ketocarotenoid found in a variety of shellfish and birds, including salmon, trout, red sea bream, shrimp, lobster, flamingos, quails etc. The green algae Haematococcus pluvialis, red yeast Phaffia rhodozyma, Chlorella zofingiensis, Chlorococcum sp., and the marine bacterium Agrobacterium aurantiacum, as well as crustacean by-products, are the familiar sources of natural AsX (Jyotika, et al., 2012). H. pluvialis, a microalga, has the highest ability to accumulate AsX, up to 4-5% of dry cell weight (Yuan et al., 2011). AsX possess several health-promoting effects in the prevention and treatment of a variety of diseases, such as cancer, chronic inflammatory diseases, metabolic syndrome, diabetes, diabetic nephropathy, cardiovascular diseases, gastrointestinal diseases, liver diseases, neurodegenerative diseases, eye diseases, skin diseases, exerciseinduced fatigue, male infertility, and anaemia. Several investigations have shown that AsX possesses antioxidant properties. Regarding free radical antioxidant activity, AsX can outperform vitamin E and β -carotene by up to several times (Shimidzu et al., 1996).

AsX, popularly known as "super vitamin E," contains ten times the antioxidant activity of other carotenoids like Zeaxanthin, Lutein, Canthaxanthin, and β -carotene and 100 times the antioxidant activity of α -tocopherol (Higuera-Ciapara *et al.* 2006). Because of its molecular structure, AsX possesses unique chemical characteristics. The more significant antioxidant activity is due to hydroxyl and keto moieties on each ionone ring (Kishimoto et al., 2010). Besides the health benefits of AsX, recent research has revealed the role of AsX in the gut microbiota linked to various diseases (Table 2). It was reported that AsX affects the activities of the intestinal mucosa in Cyclophosphamide-induced immune-deficient mice (Zhang et al., 2020). They found a change in the composition of the gut flora and its metabolites after administering AsX. Lactobacillus and Bifidobacteria, beneficial probiotic bacteria, were significantly reduced in the control group. At the same time, they were higher in the β carotene control and AsX-treated groups. In contrast, immune-deficient mice had higher levels of commensal bacteria such Clostridium as coccoides, Enterobacteriaceae spp., and Enterococcus spp (Fig. 4).

On the other hand, C. coccoide and Enterobacteriaceae *spp.* were significantly reduced in the β -carotene group and the AM (AsX medium-dose- 60 mg/kg b.w) groups. The AM and AH (AsX high dose- 120 mg/kg b.w) groups had a more similar composition of major intestinal bacteria to the control group than the other groups. Short-chain fatty acids (SCFA) estimation revealed that the control group had the lowest total SCFAs, whereas intervention with β-carotene and AsX increased total SCFAs. Furthermore, propionic acid and butyric acid levels in immune-deficient mice had reduced by 62% and 35%, respectively. They were significantly higher in the β -carotene group and the AH group. These findings suggested that β-carotene or ASX could help to speed up the formation of SCFAs. SCFAs, particularly acetic acid, propionic acid, and butyric acid, are essential bacterial metabolites in modulating intestinal immunity, lowering intestinal inflammation, and improving intestinal

Dietary Carotenoid	Structure	Source	References
Astaxanthin	$H_{3}C \xrightarrow{CH_{3}} CH_{3} \xrightarrow{CH_{3}} CH_{3} \xrightarrow{H_{3}C} H_{3}C \xrightarrow{H_{3}C} OH$	<i>Adonis annua</i> flowers, fungi, algae, bacteria, shrimp	Saini <i>et al</i> ., 2022
β-carotene	H ₃ C H ₃ C CH ₃ CH	Carrots, fungi, algae, bacteria, fruits and all green vegetables (e.g: apricot, cantaloupe, mango, carrots, red peppers, sweet po- tatoes, pumpkins, spinach, broc- coli, chard and kale)	Saini <i>et al</i> ., 2022
α-carotene (combination with β-carotene)	$\begin{array}{c} CH_3 \\ CH_3 \\ CH_3 \\ CH_3 \end{array} \xrightarrow{CH_3} \\ CH_3 \\ CH_$	Green leafy vegetables (e.g. carrots) and some varieties of squash, and pumpkins	Saini <i>et al.</i> , 2022
Canthaxanthin	H_3C CH_3 CH_3 CH_3 H_3C CH_3 H_3C CH_3 H_3C CH_3 H_3C	Crustaceans, fungi (<i>Cantharellus</i> <i>sp.),</i> algae	Takaichi <i>et al</i> ., 2011
Capsanthin	H_3C CH_3 CH_3 CH_3 H_3C H_3C H_3C H_3C CH_3 H_3C	Red pepper (<i>Paprika - Capsicum</i> annuum)	Takaichi <i>et al</i> ., 2011
Cryptoxanthin	$H_{3}C H_{3}C $	Extensively found in low amounts in many tropical or- angefleshed fruits (e.g., oranges, mangos, ripe red and orange peppers, mandarins, papayas, pumpkin, yellow maize, sea- weed, peaches, nuts and per- simmons)	Saini <i>et al.</i> , 2022
Lutein	но Слическо с с с с с с с с с с с с с с с с с с	Green vegetables (e.g., squash, broccoli, peas, brussels sprouts, string, beans)	Saini <i>et al</i> ., 2022
Zeaxanthin	$H_{3}C \xrightarrow{CH_{3}} \xrightarrow{CH_{3}} \xrightarrow{CH_{3}} \xrightarrow{H_{3}C} \xrightarrow{H_{3}$	Egg yolks, certain yellow-orange fruits (e.g. squash, oranges) and dark leafy green vegetables	Takaichi <i>et al</i> ., 2011
Lycopene	H_3C CH_3 CH_3 CH_3 H_3C H_5C	Alfalfa, tomato, watermelon, pink grapefruit, papaya, guava, rose hip, fungi	Takaichi <i>et al</i> ., 2011

Hegde, P. S. *et al. / J. Appl. & Nat. Sci*. 14(3), 1085 - 1099 (2022)

Table 1. Natural sources of different types of carotenoids

Source: Saini et al., 2022; Takaichi et al., 2011

functioning (Zhang et al., 2020).

Another study reported that, while modulating the variety of cecal microbiota, AsX reduced Ochratoxin A (OTA)-induced cecum damage and inflammation in mice (Chen *et al.*, 2021). They observed that OTA decreased gut flora diversity and increased the number of *Firmicutes* and *Lactobacillaceae*. The number of intestinal pathogens is inversely proportional to the abundance of *Firmicutes* (Mulder *et al.*, 2009). Changes in lactobacillus proportions induce the alteration in the abundance of *Firmicutes*, indicating that *lactobacillus* is a primary detoxifying bacteria in the OTA group (Walter,2008). Moreover, OTA decreased the number of *Lachnospiraceae* but recovered with ASX supplementation. AsX treatment improved gut flora and increased gut microbial diversity (especially *Lactobacillaceae* and *Lachnospiraceae*), leading to cecum barrier recovery from OTA-induced injury.

A study investigated the role of AsX on mouse gut mi-

crobiota to inhibit alcoholic fatty liver disease (Liu *et al.*, 2018). They reported that ethanol and AsX supplementation considerably impacted specific bacterial populations. *Bacteroidetes*, including the genera *Bacteroides*, *Butyricimonas*, and *Parabacteroides* were substantially more common in the disease model group (Ethanolinduced Alcoholic Fatty Liver Disease (AFLD) group). *Bacteroides* are the most prevalent genus observed in alcohol-fed mice (Yan *et al.*, 2012; Lowe *et al.*, 2017; Neyrinck *et al.*, 2017)

They also found that AsX intervention significantly reversed the ethanol-induced increase in Bacteroidetes and Proteobacteria, restoring their proportions to the regular diet group. Proteobacteria can grow in the gut as a pro-inflammatory intestinal microorganism in response to an imbalanced microbial composition (Shin et al., 2015), suggesting that AsX's anti-inflammatory properties are likely to be responsible for its protective effects. Furthermore, they observed а drop in Akkermansia (Akk) abundance in ethanol-exposed AFLD mice, which was reversed by AsX treatment, bringing it back to levels even higher than the control group indicating that AsX supplementation help mice with AFLD by enhancing Akk. Lack of Akk is an early indicator of gut dysbiosis caused by alcohol (Liu et al., 2018).

Further, ethanol exposure diminishes the number of *Akk* in both mice and humans, and oral supplementation alleviated the status of AFLD, suggesting the bacterium's protective action on AFLD. They also reported that the ethanol diet considerably increased the species of *Butyricimonas, Oscillospira, Clostridiales, Bilophila* and AsX supplementation significantly reduced it. *Oscillospira* and *Clostridales* increase during the inflammatory response, linked to intestinal mucosa barrier damage (Power *et al.*, 2016; Mastrocola *et al.*, 2018). These findings suggest that AsX alleviates the AFLD phenotype by enriching the essential bacteria for gut integrity and anti-inflammation.

Wu et al., (2020), investigated the effect of AsX on gut microbiota, inflammation, and whole-body metabolic homeostasis in wild-type C57BL/6 J and, β -carotene 9', 10'-dioxygenase (BCO2) knockout (KO) mice. The study demonstrated the interplay between gender, diet, and genotype in gut microbiota homeostasis and host metabolism. 16S rRNA sequencing revealed a genderspecific association of gut microbiota, which might present distinct responses to dietary AsX supplementation. Bacteroidetes and Proteobacteria were significantly higher, whereas the Firmicutes/Bacteroidetes (F/B) ratio and Actinobacteria abundance were substantially lower in females than males. In addition, they found that female mice had higher fecal SCFA concentrations and abundance of Lachnospiraceae bacterium (Firmicutes) and lower Ruminococcus (Firmicutes) than

male mice. Lanchnospiraceae species ferment undigested food fibres to create SCFAs (Tanca et al. 2018). SCFAs decrease systemic inflammation by promoting commensal bacterial growth and host epithelial regeneration (Xu et al., 2018) showed that changes in glucose and lipid metabolism are due to a shift in the gut microbiota. M. schaedleri was significantly enhanced in BCO2-deficient animals, regardless of gender, and was highly related to numerous metabolic parameters, including fasting blood glucose, fat percentage, and cholesterol. According to previous investigations, increased M. schaedleri abundance in the gut is related to local and systemic inflammatory responses (Daniel et al., 2017; Loy et al., 2017). Hence, variations in M. schaedleri abundance could be a biomarker for BCO2 protein expression status in mice.

sX supplementation in male knock-out mice increased the number of Akk. muciniphila (Verrucomicrobia) substantially compared to malewild-type mice. Akk. muciniphila is amucin-degrading bacterium that safeguards the intestinal mucus layer. Indeed, serum carotenoid levels signify the proliferation of A. muciniphila in the intestine. A. muciniphila enrichment protects mouse white adipose tissues from inflammation caused by high saturated lipid levels following high-fat dietinduced (Caesar et al., 2015; Diuric et al., 2018; He et al., 2018), whereas quantity of Proteobacteria is lowered in KO mice. Increased Proteobacteria populations are linked to inflammation and are considered as a hallmark of microbiome imbalance and a contributing factor to cecal inflammation (Carvalho et al., 2012). Thus, AsX promotes gut microbiota homeostasis by increasing the abundance of beneficial bacteria (such as A. muciniphila) while decreasing harmful bacterial growth

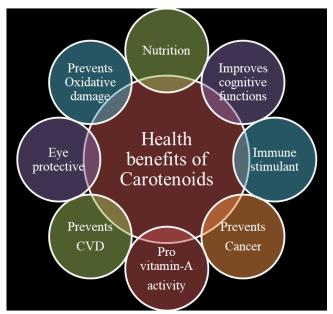


Fig. 3. Role of carotenoids in human health and disease (Source: Olatunde et al., 2020)

Hegde, P. S. et a	al. / J. Appl.	& Nat. Sci.	14(3),	1085 - 1099 (2022)
-------------------	----------------	-------------	--------	--------------------

Dietary Carotenoid	Disease model	Gut microbes altered	References
		Lactobacillus ↑	
	Cyclophosphamide - induced immunodeficient mice	Bifidobacteria ↑ Clostridium coccoides ↓ Enterobacteriaceae spp. ↓ Enterococcus spp. ↓	(Zhang <i>et al</i> ., 2020)
	OTA-induced cecum dam-	Firmicutes \downarrow	
Astaxanthin	age and inflammation in mice	Lactobacillaceae ↑ Lachnospiraceae ↑ Bacteroides ↓	(Chen <i>et al</i> ., 2021)
		Butyricimonas ↓ Parabacteroides ↑ Proteobacteria ↓ Akk ↑	
	AFLD in mice	Butyricimonas ↓ Oscillospira ↓ Clostridiales ↓ Bilophila ↓	(Liu <i>et al.</i> , 2018)
	Wild-type C57BL/6 J and BCO2 KO mice	Akk. muciniphila ↑ Proteobacteria ↓	(Wu <i>et al</i> ., 2020)
	High fat diet (HFD) in- duced obese mice	Firmicutes/Bacteroides ↓ Proteobacteria ↓ Akk ↑	(Backhed <i>et al</i> ., 2004

Table 2. Effect of Astaxanthin on the gut microbiome in different disease models

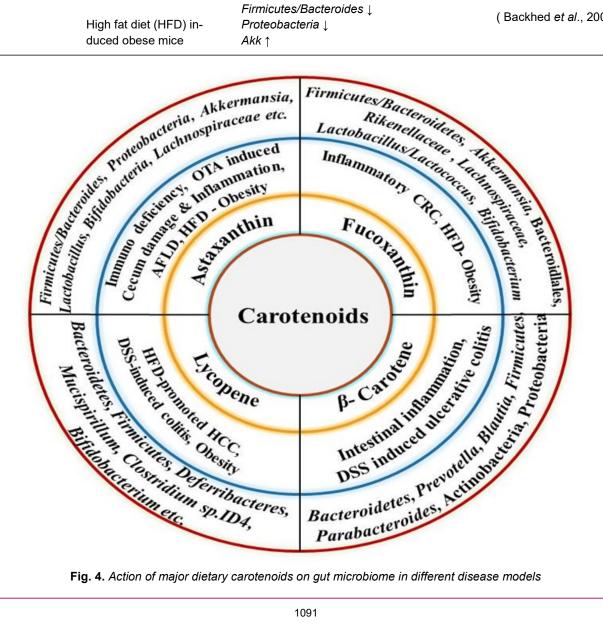


Fig. 4. Action of major dietary carotenoids on gut microbiome in different disease models

(such as *Proteobacteria*). Hence AsX significantly impacts the gut microbiota by reducing local and systemic oxidative stress and inflammation.

Wang et al. (2019) found that Xanthophyllomyces dendrorhous derived AsX had a regulatory influence on lipid metabolism and gut microbiota in high-fat dietinduced obese mice, and adding AsX or X. dendrorhous powder to a high-fat meal lowered the Firmicutes/Bacteroides ratio. The Firmicutes to Bacteroidetes ratio is crucial for bodyweight variations (Backhed et al., 2004) preventing Proteobacteria growth, which is beneficial for health. Increased Proteobacteria prevalence is a possible diagnostic signature of dysbiosis and illness risk (Shin et al., 2015). Proteobacteria flourished in the gut due to a high-fat diet, causing illnesses. Addition of AsX or X. dendrorhous powder to the gut enhanced the amount of Akk. Akk draws energy from the mucin produced by the mucosa, thereby guarding the gut against infections through competition (Belzer and De Vos, 2012) by adhering to the gut mucosa. Previous research has shown that feeding mice live Akk can prevent diet-induced obesity while having little effect on their appetite or eating behaviour (Everard et al., 2013). A high-fat diet and a large amount of alcohol intake will lower the amount of Akk in the gut. In conclusion. AsX and X. dendrorhous powder had a beneficial regulatory effect on gut microbiota, ensuring the health of gut micro-ecology and preventing obesity due to a high-fat diet.

Effect of fucoxanthin on gut microbiota

Fx is an important carotenoid contributing around 10% of the overall production of carotenoids in nature. It has a unique structure containing an uncommon allenic bond, a 5, 6-monoepoxide (Kim and Pangestuti, 2011), and a polyene chain conjugated to a carbonyl group. This unique structural characteristic cause Fx's antioxidant, anti-inflammatory, and anticancer properties. Fx is an exceedingly polar carotenoid commonly present algae, consisting of *Undaria pinnatifida, Laminaria japonica, Phaeodactylum tricornutum,* and *Cylindrotheca closterium* (Zhang *et al.*, 2015). Intestinal microbes hydrolyze Fx into fucoxanthinol, hence enhancing its bioavailability (Liu *et al.*, 2019).

Guo *et al.*, 2019 investigated the impact of Fx on microbial profiles of caecal and faecal samples from BALB/c mice fed with HFD, confirmed the changes in *Firmicutes/Bacteroidetes* ratio and the abundance of S24-7 and *Akk* indicated the anti-obesity activity of Fx. Another group led by Liu *et al.* (2019), reported the impact of Fx isolated from edible algae *Undaria pinnatifida* on pathogenic microorganism *Escherichia coli* and *Lactobacillus* in invitro and invivo models wherein, Fx strongly inhibited the growth of Gram-positive pathogenic microorganisms but was less effective towards Gramnegative microorganisms and promoted the growth of intestinal microbes in mice (Table 3).

Terasaki et al., (2018), investigated the chemoprotective effect of Fx and its impact on gut microbiota in a mouse model of inflammatory colorectal cancers showed the inhibition of multiplicity of colorectal adenocarcinoma upon administration of Fx (30 mg/kg BW) for 14-week. Fx administration notably suppressed Bacteroidales and Rikenellaceae and increased Lachnospiraceae population compared with the control mice. Oral administration of a faecal suspension acquired from Fxtreated mice aimed to increase Lachnospiraceae, suppress the number of colorectal adenocarcinomas in Azoxymethane and Dextran sodium sulphate (AOM/ DSS) treated mice with an increase in Lachnospiraceae in the gut. Their findings suggested that an alteration in gut microbiota by Fx is probably an essential factor in the chemoprotective impact of Fx in AOM/DSS- treated mice (Fig. 4).

Sun *et al.* (2020), designed a mouse model to study the modulation of gut microbiota by Fx to Richards intestinal microbial homeostasis. Fx also reduced HFD-induced gut microbiota dysbiosis by suppressing the growth of obesity/inflammation-related *Lachnospiraceae* and *Erysipelotrichaceae* while boosting the growth of *Lactobacillus / Lactococcus*, *Bifidobacterium*, and some butyrate-producing bacteria as confirmed by its 16S rRNA sequencing.

Effect of lycopene on gut microbiota

LYC is a lipophilic, unsaturated, biologically active acyclic carotenoid found in fruits and vegetables that are red in colours, such as tomato, papaya, pink grapefruit, pink guava, and watermelon. Processed tomato products like ketchup, tomato juice, spaghetti sauce, and pizza sauce account for more than 80% of dietary LYC consumption, as processing generally entails water loss. Hence, the amount of LYC processed foods is often significantly higher than that in fresh foods (Xia et al., 2018). According to Zhao B et al., 2018, LYCcontaining foods, such as tomato products and supplements, affect a wide range of illnesses, including cancer, heart disease, and asthma. LYC exhibits antiatherosclerotic, antioxidant, anti-inflammatory, antihypertensive, anti-platelet, anti-apoptotic, and protective endothelial effects (Table 4). LYC showed the prevention of malignancies such as Osteosarcoma, lung, and prostate cancers due to its antioxidant properties. Unlike most carotenoids, LYC does not convert to Vitamin A in human bodies (Story et al., 2010; Mozos et al., 2018).

LYC-containing dietary tomato powder (TP) feeding showed the suppression of HFD-promoted hepatocellular carcinoma (HCC) development andprotected HFDinduced inflammation with putative gut microbiota modulation (Editorial 2007). A 16s rDNA sequencing study showed that TP feeding boosted the richness of the gut microbiome and a significant increase in microbial diversity in the HFD + DEN (Diethyl Nitrosamine, a liverspecific carcinogen) group in a BCO1/BCO2 double KO mice (Xia et al., 2018). The relative abundance of gram -positive bacteria increased following TP feeding, whereas gram-negative bacteria decreased. According phylum-level analysis, six main phyla to (Bacteroidetes, Firmicutes, Deferribacteres, Proteobacteria, Actinobacteria, and Tenericutes) dominate the fecal microbiota (Fig 4).

Although Gram-positive and gram-negative bacteria coexist in the gut, Gram-negative bacteria overgrowth can increase the generation of hepatotoxic chemicals like lipopolysaccharide (LPS) (Wigg et al., 2001). They observed the species of Deferribacteres and a relative abundance of Mucispirillum and Mucispirillum Schaedler reduced drastically following TP feeding. Mucispirillum cause increased serum leptin levels and inflammatory markers (Loy et al., 2017), suggested that TP feeding could reduce inflammatory cytokines by reducing Mucispirillum's gut microbiota. The Clostridium sp.ID4 and Clostridium sp.Clone.9 populations in the TP feeding group also showed a decline and an increase in the TP feeding group, respectively. Hepatic lipogenesis and fat oxidation have increased by Clostridium species (Nadeau and Conjeevaram, 2017). Together, Gut-liver interaction and TP feeding boosted the gut microbiome's richness and diversity and improved some beneficial bacteria.

Zhao *et al.* (2020), found a correlation between LYC consumption and gut microbiota structure and metabolite production in DSS-induced colitis mice. They reported that *Firmicutes, Actinobacteria,* and *TM*7 had increased relative abundance in both the control and LYC treatment groups compared to DSS group, but *Proteobacteria* and *Deferribacteres* had higher relative abundance in the DSS group. The relative abundance of *Clostridiales* and *Lachnospiraceae* in the SCFAs of gut microbiota in the DSS group was much lower than

in the control group. DSS increased the relative abundance of *Mucispirillum*, whereas the relative abundance of probiotic bacteria (*Lactobacillus* and *Bifido- bacterium*) was decreased. SCFAs like acetate, propionate, isobutyrate, butyrate, valerate and isovalerate are significantly reduced but, LYC increased the concentrations in DSS-induced colitis mice. LYC could influence the relative abundance of gut microbiota and its metabolic products. As a result, LYC affected DSS-induced changes in gut microbiota metabolites, including LPS and SCFAs. The gut microbiota structure of DSSinduced colitis mice was altered by LYC, suggesting that the gut-brain axis balance may also play a role. Wiese *et al.* (2019) explored the systemic effect of LYC and dark chocolate (DC) on gut microbiota blood liver

and dark chocolate (DC) on gut microbiota, blood, liver metabolism, skeletal muscle tissue oxygenation and skin in moderately obese individuals. After four weeks of supplementation with GA-LYC (GAL) (GAL - is added to overcome reduced bioavailability in older people and individuals with metabolic syndrome), followed by 16s rRNA sequencing of gut microbiota from faeces demonstrated a shift in gut microbial communities. They found GAL formulations resulted in a dosedependent increase in members of the phyla Actinobacteria, primarily due to the rise in the relative abundance of Bifidobacterium spp. and Lactobacilli spp. implied the prebiotic potential. They also observed that continuous intervention with GAL, DC, and DCL (DC+LYC) resulted in a considerable reduction in Bacteroidetes abundancedue to the direct impact or its indirect activity via stimulation of Bifidobacteria species or a combination of both.

Effect of β-carotene on gut microbiota

In certain fruits and vegetables, β -carotene is a retinol precursor (Vitamin A). It is a cyclic carotene with yellow colour. The most active and significant provitamin-A carotenoid has a bright red-orange colour and is commonly found in plants and fruits. It has multiple roles in the biological system as a natural pigment, provitamin-A, plant metabolite, human metabolite, mouse metabo-

Dietary Carotenoid	Disease model	Gut microbes altered	References
	HFD fed BALB/c mice	<i>Firmicutes/Bacteroidetes</i> ↓ S24-7 ↑ <i>Akk</i> ↓	(Guo <i>et al</i> .,2019)
Fucoxanthin	Inflammatory colo- rectal cancers in mouse	Bacteroidlales↓ <i>Rikenellaceae</i> ↓ <i>Lachnospiraceae</i> ↑	(Terasaki <i>et al</i> .,2020)
	HFD induced obese mice	Lachnospiraceae↓ Erysipelotrichaceae↓ Lactobacillus/Lactococcus ↑ Bifidobacterium ↑	(Sun <i>et al</i> ., 2020)

lite, cofactor, andantioxidant (beta-Carotene | C40H56 -PubChem https://pubchem.ncbi.nlm.nih.gov/compound/ beta-Carotene).

β-Carotene is one of the most widely studied carotenoids for its vitamin-A activity, abundant in fruits and vegetables. Apricots, Asparagus, Broccoli, Carrots, Chinese cabbage, Onions, Peas, Peppers, Plums, Pumpkin, Spinach, Squash, and Sweet Potatoes are rich in β-Carotene. β-carotene can also be extracted from halo-tolerant algae (Dunaliella salina) or fungi (Blakeslea trispora). Epidemiological studies have proved the abundance of dietary carotenoids as being protective against many diseases (Table5). β-carotene protects membrane lipids from free radical damage by scavenging singlet oxygen, cleaning peroxide radicals, and interacting directly with peroxy radicals. β-carotene improves cognitive function in addition to its antioxidant activity. Long-term intake of β-carotene for 15 years or more may slow down age-related cognitive decline, but short-term intake has no significant effect (Grodstein et al., 2007; Kristine, 2007). It also helps to improve skin health, which is likely due to its antioxidant effects (Stahl and Sies, 2012). Diets high in carotenoids, such as β -carotene, may aid in eye health and protect against illnesses of the eyes, such as age-related macular degeneration (AMD). High blood levels of carotenoids, such as β -carotene, have been demonstrated in studies to lessen the chance of developing advanced age-related macular degeneration (Wu et al., 2016). Furthermore, diets rich in beta carotene-containing fruits and vegetables may be particularly efficient in lowering the incidence of AMD in smokers (Kim et al., 2017) and preventing lung cancer (Cai et al., 2019); further, evidence of β-carotene's effect on gut microbial dysbiosis exists.

Li et al., (2019) demonstrated that β -carotene reduces weaning-induced intestinal inflammation in piglets by modifying gut flora. Their findings showed that βcarotene improved growth performance, intestinal morphology, and relieving inflammation. Furthermore, compared to the weaning group, β-carotene considerably decreased species of the phyla Bacteroidetes and the genera Prevotella and Blautia, and significantly increased species of the phyla Firmicutes, genera p-75a5, Parabacteroides. Prevotella and Blautia were positively correlated, whereas, Parabacteroides and Synergists were negatively correlated with the levels of IL-1β, IL-6, and TNF-α. At the same time, p-75-a5 showed a negative correlation with IL-6 in piglet serum samples. These data suggest that β-carotene reduces weaninginduced intestinal inflammation in pigs via altering the gut microbiome. Prevotella could target β-carotene in piglets suffering from weaning-induced intestinal inflammation (Fig. 4).

Zhu *et al.*, (2021) investigated the effects of β -carotene

on gut microbiota modulation and its anti-inflammatory effects in a rat model of ulcerative colitis induced by dextran sulphate sodium (DSS). Their 16S rRNA sequencing approach demonstrated that the relative abundance of gut microbiota changed dramatically with DSS administration at the phylum and genus levels. Furthermore, β -carotene treatment enhanced the population of certain beneficial microorganisms and is negatively correlated with inflammatory cytokine levels.

Firmicutes and Actinobacteria had low relative abundance in DSS group, while Bacteroidetes and Proteobacteria were higher. On the other hand, treatment with β -carotene boosted the abundance of Firmicutes and Actinobacteria while decreasing the abundance of Bacteroidetes and Proteobacteria. The Bacteroidetes phylum in humans and animals contains three essential bacteria that can turn dangerous; hence their microecological imbalance may lead to endogenous infection (Gilmore, 2008; Liu *et al.*, 2018). The increase in *Proteobacteria* is a warning sign of disease (Yan *et al.*, 2012).

The correlations between gut bacteria and serum inflammatory cytokine levels revealed that only Faecalibacterium was inversely correlated with inflammatory cytokine levels, whereas the others were favourably correlated. Faecalibacterium is one of the most common butyrate-producing bacteria among gut microbes and has a negative correlation with disease activity in inflammatory bowel disease patients (Lopez-Siles et al., 2017; Pittayanon et al., 2020). Faecalibacterium has anti-inflammatory capabilities and can promote epithelial cell proliferation and repair to sustain the intestinal mucosal barrier's integrity and gut health (Sokol et al., 2017). Staphylococcus, Sutterella, and Parabacteroides were highly expressed in the DSS group and positively associated with TNF-a. Most Staphylococcus species are conditional pathogens that cause infection and are common in IBD patients. Sutterella is commonly observed in patients with gastrointestinal problems linked to inflammation in the intestine. DSS-induced chronic colitis dramatically enhances the abundance of Sutterella in mice (Cai et al., 2019; T. Liu et al., 2019). Alistipes and Phascolarctobacterium were more prevalent in the DSS group, and both had significantly correlated with IL-6 and IL-1 levels. Alistipes have been associated with colon rectum cancer, while Phascolarcto bacterium is associated with diarrhoea (Seishima et al., 2019). Veillonella and Enterococcus were also more prevalent in the DSS group and were strongly related to IFN-y. Both Veillonella and Enterococcus are pathogens, and studies have shown that Veillonella raises the risk of colitis, while Enterococcus can cause colon illness and an imbalance in inflammation factors (Abdel -Aal et al., 2013; Santoru et al., 2017). These findings suggest that β-carotene's anti-inflammatory actions are

Dietary Carotenoid	Disease model	Gut microbes altered	References
		Bacteroidetes↑	
		Firmicutes↑	
		<i>Deferribacteres</i> ↑	
		Proteobacteria↑	
		Actiobacteria↑	
		Tenericutes↑	
	HFD-promoted	Deferribacteres↓	
	Hepato cellular	Mucispirillum ↓	(Xia <i>et al</i> .,2018)
	carcinoma in mice	Mucispirillum schaedleri↓	(Ala et al.,2010)
Lyconono		Clostridium sp.ID4 ↓	
Lycopene		<i>Clostridium sp</i> .Clone.9 ↓ Firmicutes↑	
		Actinobacteria↑	
	DSS-induced colitis	TM7↑	
	in mice	Proteobacteria↓	(Nadeau and Conjeevaram, 2017)
		Deferribacteres↓	
		<i>Bifidobacterium longum</i> ↑	
	Moderately obese individuals	Bifidobacterium adolescentis↑	
		<i>Lactobacilli</i> spp↑	(Wiese <i>et al.</i> , 2019)
		Bacteroidetes↓	

Hegde, P. S. et al. / J. Appl. & Nat. Sci. 14(3), 1085 - 1099 (2022)

Table 5. Effect of β-carotene on the gut microbiome in different disease models

Dietary Carotenoid	Disease model	Gut microbes altered	References
β-carotene	Weaning-induced intestinal inflammation in piglets	Bacteroidetes↓ Prevotella↓ Blautia↓ Firmicutes↑ p-75-a5↑ Parabacteroides↑	(Li <i>et al.</i> ,2019)
	DSS induced ulcerative colitis in rats	Firmicutes↑ Actinobacteria↑ Bacteroidetes↓ Proteobacteria↓	(Zhu <i>et al</i> ., 2021)

mediated by altering the gut flora through the inflammatory factor.

Apart from these carotenoids, some unique carotenoids also substantially affected gut microbiota, such as Leutin and Crocetin. Lutein naturally occurs in several fruits and vegetables, orange, and reddish-yellow colored vegetables like kale, spinach, broccoli, parsley, leek, red pepper, peas, lettuce, and also in egg yolks (Lin et al., 2021). Lutein supplements were used in alternative medicine for eye diseases, such as cataracts and macular degeneration. Alternative medicine proponents claim that lutein supplements can also help prevent colon cancer, breast cancer, diabetes, and heart disease. To date, most of the studies on lutein's health benefits have focused on its dietary intake, suggesting that lutein may help protect against atherosclerosis, age-related macular degeneration, and cataracts.

Therefore, it is evident that there is a differential ex-

pression of gut bacteria upon administration of different carotenoids. The differential expression includes alterations in the abundance of certain microbial taxa or structure of the microbial community, decreased bacterial richness and/or diversity and decreased microbial community stability.

Conclusion

The current active field of research in biomedicine is on the human gut microbiome; its dysbiosis in various disease models indicates the importance of the involvement of the gut microbiome in human health and disease. Various studies have revealed the major differences in microbial taxonomic and functional composition in different disease conditions compared to their normal counterparts. It is still under debate, whether dysbiosis precedes the development of disease and sets the inflammatory process or merely reflects the inflamed mucosa's altered immune and metabolic environment. Therefore, it is essential to study newly diagnosed treatment-naïve patients, where the microbiome can be studied at the beginning of the disease.

Future perspectives

Currently, Gut microbiome studies are mainly focused on bacteria, and the gut microbiome is composed of other microorganisms beyond bacteria, such as viruses, fungi or archaea, which play a role in human health and disease other than the bacterial population per se. Therefore, future studies should include a holistic approach with respect to the gut microbiome for a deeper understanding of the specific disease. The available microbiome data represent a valuable data source that can be utilized for identifying associations between the microbiome and specific disease conditions upon administration of specified carotenoids along with their pathophysiological understanding may help in promoting the development of clinical strategies, including disease prevention, treatment, stratification and assessment of the high-risk population. As there is a great diversity in the composition of microbial taxa diversity between individuals and within the same individual over time, the predictive value of these potential indicators has low clinical importance. In order to use the gut microbiome as non-invasive biomarkers in human disease, prospective longitudinal studies on gut dysbiosis in early-onset of specific disease conditions are warranted to enable personalized treatments and to reduce the health care economic burden.

ACKNOWLEDGEMENTS

The authors are grateful to Nitte (Deemed to be University) for providing the necessary research facilities to carry out this work.

Conflict of interest

The authors declare that they have no conflict of interest.

REFERENCES

- Abdel-Aal, E.S.M., Akhtar, H., Zaheer, K. & Ali, R. (2013). Dietary sources of lutein and zeaxanthin carotenoids and their role in eye health. *Nutrients*, 5(4),1169– 1185. https://doi.org/10.3390/nu5041169
- Backhed F., Ding H., Wang, T., Hooper, L. V., Gou, Y.K., Nagy A., Semenkovich C.F. & Gordon JI. (2004). The gut microbiota as an environmental factor that regulates fat storage. *Proceedings of the National Academy of Sciences of the USA*, 101(44),15718–15723. https:// doi.org/10.1073/pnas.0407076101
- Bakan E, Tugba Akbulut, Z., & Levent Inanc, A. (2014). Carotenoids in Foods and their Effects on Human Health.

Academic Food Journal, 12(2) 61-68. http://www.acad emicfoodjournal.com

- Bas-Bellver, C., Andres C., Segui L., Barrera C., Jimenez -Hernandez N., Artacho A., Betoret N., & Gosalbes MJ. (2020). Valorization of Persimmon and Blueberry Byproducts to Obtain Functional Powders: In Vitro Digestion and Fermentation by Gut Microbiota. *Journal of Agricultural and Food Chemistry*, 68(30):8080–8090. https:// doi.org/10.1021/acs.jafc.0c02088
- Beita Zhao., Hua Liu., Jiamin Wang., Pujie Liu., Xintong Tan., Bo Ren., Zhigang Liu., & Xuebo Liu. (2018). Lycopene Supplementation Attenuates Oxidative Stress, Neuroinflammation, and Cognitive Impairment in Aged CD-1 Mice. *Journal of Agricultural and Food Chemistry*. 66,12,3127–3136. https://doi.org/10.1021/acs.jafc.7b 05770
- Belzer C., & De Vos WM. (2012). Microbes insidefrom diversity to function: The case of Akkermansia. *International Society for Microbial Ecology* J. 6(8):1449–1458. https://doi.org/10.1038/ismej.2012.6
- Caesar R., Tremarol I.V., Kovatcheva-Datchary P., Cani P.D., & Backhed F. (2015). Crosstalk between gut microbiota and dietary lipids aggravates WAT inflammation through TLR signaling. *Cell Metabolism*. 22(4):658–668. https://doi.org/10.1016/j.cmet.2015.07.026
- Cai X., Han Y., Gu M., Song M., Wu X., Li Z., Li F., Goulette T., & Xiao H. (2019). Dietary cranberry suppressed colonic inflammation and alleviated gut microbiota dysbiosis in dextran sodium sulfate-treated mice. *Food & Function*. 10(10):6331–6341. https://doi.org/10.1039/ c9fo01537j
- Carrera-Quintanar L., Roa R.I.L., Quintero-Fabian, S., Sanchez-Sanchez M.A., Vizmanos B., & Ortuno-Sahagun D. (2018). Phytochemicals that influence gut microbiota as prophylactics and for the treatment ofobesity and inflammatory diseases. *Mediators of Inflammation*. https://doi.org/10.1155/2018/9734845
- Carvalho, F.A., Koren O., Goodrich. J.K., Johansson M.E.V., Aitken J.D., Su Y., Chassaing B., Walters W. A., Gonzalez A., & Clemente J.C. *et al.*, (2012). Transient Inability to Manage Proteobacteria Promotes Chronic Gut Inflammation in TLR5-Deficient Mice. *Cell Host & Microbe*. 12(2):139–152. https://doi.org/10.1016/ j.chom.2012.07.004.Transient
- Chen Y., Zhao S., Jiao D., Yao B., Yang S., Li P., & Long M. (2021). Astaxanthin Alleviates Ochratoxin A-Induced Cecum Injury and Inflammation in Mice by Regulating the Diversity of Cecal Microbiota and TLR4/MyD88/NF- κ B Signaling Pathway. Oxidative Medicine and Cellular Longevity. https://doi.org/10.1155/2021/8894491
- Daniel S.G., Ball C.L., Besselsen D.G., Doetschman T., & Hurwitz B.L. (2017). Functional Changes in the Gut Microbiome Contribute to Transforming Growth Factor β-Deficient Colon Cancer. *mSystems*. 2(5). https:// doi.org/10.1128/msystems.00065-17
- Djuric Z, Bassis, C.M., Plegue, M.A., Ren J., Chan R., Sidahmed, E.K., Turgeon, D.K., Ruffin, M.T., Kato I., & Sen A. (2018). Colonic Mucosal Bacteria Are Associated with Inter-Individual Variability in Serum Carotenoid Concentrations. *Journal of the Academy of Nutrition and Dietetics*. 118(4):606-616.e3. https://doi.org/10.1016/j.jand.2 017.09.013

- Echavarri-Erasun, C., & Johnson, E.A. 2002. Fungal carotenoids. *Applied Mycology and Biotechnology*. 2 (C):45–85. https://doi.org/10.1016/S1874-5334(02)80006 -5
- Everard A., Belzer C., Geurts L., Ouwerkerk, J.P., Druart C, Bindels, L.B., Guiot Y., Derrien M., Muccioli, G.G., & Delzenne, N.M. *et al.* (2013). Cross-talk between Akkermansia muciniphila and intestinal epithelium controls diet -induced obesity. *Proceedings of the National Academy* of Sciences of the USA, 110(22):9066–9071. https:// doi.org/10.1073/pnas.1219451110
- Feher M., Fauszt P., Tolnai E., Fidler G., Pesti-Asboth, G., Stagel A., Szucs I., Biro S., Remenyik J., Paholcsek M., & Stundl L. (2021). Effects of phytonutrientsupplemented diets on the intestinal microbiota of Cyprinus carpio. *PLoS One*. 16(4 April):1–24. https:// doi.org/10.1371/journal.pone.0248537
- Fiedor J., & Burda K. (2014). Potential role of carotenoids as antioxidants in human health and disease. *Nutrients*. 6 (2):466–488. https://doi.org/10.3390/nu6020466
- Galasso C., Corinaldesi C., & Sansone C. (2017). Carotenoids from marine organisms: Biological functions and industrial applications. *Antioxidants*. 6(4). https:// doi.org/10.3390/antiox6040096
- Grodstein F., Kang, J.H., Glynn, R.J., Cook, N.R., & Gaziano, J.M. (2007). A randomized trial of beta carotene supplementation and cognitive function in men: The physicians' health study II. *Archives of internal medicine*. 167 (20):2184–2190. https://doi.org/10.1001/archinte.167.2 0.2184
- Guinane, C.M., & Cotter, P.D. (2013). Role of the gut microbiota in health and chronic gastrointestinal disease : understanding a hidden metabolic organ. *Therapeutic Advances in Gastroenterology*. 6(4): 295–308. https:// doi.org/10.1177/1756283X13482996
- Guo B., Yang B., Pang X., Chen T., Chen F., & Cheng, K.W. (2019). Fucoxanthin modulates cecal and fecal microbiota differently based on diet. *Food Funct*. 10 (9):5644–5655. https://doi.org/10.1039/c9fo01018a
- He C., Cheng D., Peng C., Li Y., Zhu Y., & Lu N. (2018). High-fat diet induces dysbiosis of gastric microbiota prior to gut microbiota in association with metabolic disorders in mice. *Frontiers in Microbiology*. 9(APR):1–9. https:// doi.org/10.3389/fmicb.2018.00639
- Higuera-Ciapara, I., Felix-Valenzuela, L., & Goycoolea, F.M. (2006). Astaxanthin: A review of its chemistry and applications. *Critical Reviews in Food Science and Nutrition*. 46(2):185–196. https://doi.org/10.1080/1040 86905 90957188
- Jihui Wang., Shiwen Liu., Han Wang., Shan Xiao., Cheng Li., Ying Li., & Bingnan Liu (2019). Xanthophyllomyces dendrorhous-Derived Astaxanthin Regulates Lipid Metabolism and Gut Microbiota in Obese Mice Induced by A High-Fat Diet. *Mar. Drugs*, 17, 337; doi:10.3390/md17060337
- Jyotika Dhankhar., Sumita S., Kadian., & Asha Sharma. (2012). ASTAXANTHIN: A POTENTIAL CAROTENOID. International Journal of Pharmaceutical Sciences and Research, Vol. 3(5): 1246-1259
- 26. Kim SK., & Pangestuti R. Biological activities and potential health benefits of fucoxanthin derived from marine

brown algae. Adv Food Nutr Res. 2011;64:111-28. doi: 10.1016/B978-0-12-387669-0.00009-0. PMID: 22054942.

- Kim, E.K., Kim, H., Vijayakumar A., Kwon O., & Chang N. (2017). Associations between fruit and vegetable, and antioxidant nutrient intake and age-related macular degeneration by smoking status in elderly Korean men. *Nutrition Journal*. 16(1):1–9. https://doi.org/10.1186/ s12937-017-0301-2
- Kristine Yaffe.(2007). Antioxidants and Prevention of Cognitive Decline: Does Duration of Use Matter?. Editorial. Arch intern med 167(20):2167–2168. https:// doi.org/10.1002/14651858.CD002854.10.
- Kishimoto Y., Tani M., Uto-Kondo, H., Iizuka M., Saita E., Sone H., Kurata H., & Kondo K. (2010). Astaxanthin suppresses scavenger receptor expression and matrix metalloproteinase activity in macrophages. *European Journal of Nutrition*. 49(2):119–126. https:// doi.org/10.1007/s00394-009-0056-4
- Li R., Li L., Hong P., Lang W., Hui J., Yang Y., & Zheng X. (2019). β-Carotene prevents weaning-induced intestinal inflammation by modulating gut microbiota in piglets. *Asian-Australasian Journal of Animal Science*. 00(00):1–14. https://doi.org/10.5713/ajas.19.0499
- Lin S., Li Q., Jiang S., Xu Z., Jiang Y., Liu L., Jiang J., Tong Y., & Wang P. (2021). Crocetin ameliorates chronic restraint stress-induced depression-like behaviors in mice by regulating MEK/ERK pathways and gut microbiota. *Journal of Ethnopharmacology*. 268:113608. https:// doi.org/10.1016/j.jep.2020.113608
- Liu H., Liu M., Fu X., Zhang Z., Zhu L., Zheng X., & Liu J. (2018). Astaxanthin prevents alcoholic fatty liver disease by modulating mouse gut microbiota. *Nutrients*. 10(9). https://doi.org/10.3390/nu10091298
- Liu T., Xiong Q., Li L., & Hu Y. (2019). Intestinal microbiota predicts lung cancer patients at risk of immunerelated diarrhea. *Immunotherapy*. 11(5):385–396. https:// doi.org/10.2217/imt-2018-0144
- 34. Liu Z., Sun Xiaowen., Sun Xun., Wang S., & Xu Y. (2019). Fucoxanthin Isolated from Undaria pinnatifida Can Interact with Escherichia coli and lactobacilli in the Intestine and Inhibit the Growth of Pathogenic Bacteria. *Journal of Ocean University of China*. 18(4):926–932. https://doi.org/10.1007/s11802-019-4019-y
- Lopez-Siles, M., Duncan, S.H., Garcia-Gil, L.J., & Martinez-Medina M. (2017). *Faecalibacterium prausnitzii*: From microbiology to diagnostics and prognostics. *International Society for Microbial Ecology*. 11(4):841–852. https://doi.org/10.1038/ismej.2016.176
- Lowe, P.P., Gyongyosi B., Satishchandran A., Iracheta-Vellve, A., Ambade A., Kodys K., Catalano D., Ward, D.V., & Szabo G. (2017). Alcohol-related changes in the intestinal icrobiome influence neutrophil infiltration, inflammation and steatosis in early alcoholic hepatitis in mice. *PLoS One*. 12(3):1–16. https://doi.org/10.1371/ journal.pone.0174544
- Loy A., Pfann C., Steinberger M., Hanson B., Herp S., Brugiroux S., Gomes C., Boekschoten. M.V., Schwab C., & Urich T, *et al.* (2017). schaedleri , Lifestyle and Horizontal Gene Transfer-Mediated Evolution of Mucispirillum schaedleri, a Core Member of the Murine Gut Microbiota. *mSystems.* 2(1):e00171-16

- Maoka T.(2011). Carotenoids in marine animals. *Marine* Drugs. 9(2):278–293. https://doi.org/10.3390/md9020278
- Mastrocola R., Ferrocino I., Liberto E., Chiazza F., Cento, A.S., Collotta D., Querio G., Nigro D., Bitonto V., & Cutrin, J.C. *et al.* (2018). Fructose liquid and solid formulations differently affect gut integrity, microbiota composition and related liver toxicity: a comparative in vivo study. *Journal of Nutritional Biochemistry* . 55:185–199. https://doi.org/10.1016/j.jnutbio.2018.02.003
- Mezzomo N., & Ferreira, S.R.S. (2016). Carotenoids functionality, sources, and processing by supercritical technology: A review. *Journal of Chemistry*. 2016. https:// doi.org/10.1155/2016/3164312
- Mozos I., Stoian D., Caraba A., Malainer C., Horbanczuk, J.O., & Atanasov, A.G. (2018). Lycopene and vascular health. *Frontires in Pharmacology*. 9(MAY):1–16. https:// doi.org/10.3389/fphar.2018.00521
- Mulder. I,E., Schmidt B., Stokes, C.R., Lewis M., Bailey M., Aminov, R.I., Prosser, J.I., Gill, B.P., Pluske, J.R., & Mayer, C.D. *et al.* (2009). Environmentally-acquired bacteria influence microbial diversity and natural innate immune responses at gut surfaces. *BMC Biology*. 7:1–20. https://doi.org/10.1186/1741-7007-7-79
- Nadeau, B.A. & Conjeevaram H. (2017). Nonalcoholic Fatty Liver Disease and the Gut Microbiome. *Clinics in Liver Disease*. 10(5):116–119. https://doi.org/10.1002/ cld.671
- Neeraj K Surana., & Dennis L Kasper. (2012). The yin yang of bacterial polysaccharide: : lessons learned from B. fragilis PSA. Immunological Reviews 245(1):13-26. doi: 10.1111/j.1600-065X.2011.01075.x.
- Neyrinck, A.M., Etxeberria U., Taminiau B., Daube G., Van Hul M., Everard A., Cani, P.D., Bindels, L.B., & Delzenne, N.M. (2017). Rhubarb extract prevents hepatic inflammation induced by acute alcohol intake, an effect related to the modulation of the gut microbiota. *Molecular Nutrition & Food Research*. 61(1):1–12. https:// doi.org/10.1002/mnfr.201500899
- Olatunde A., Tijjani H., Ishola, A.A., Egbuna C., Hassan S., & Akram M. (2020). Carotenoids as Functional Bioactive Compounds. Functional Foods and Nutraceuticals. https://doi.org/10.1007/978-3-030-42319-3_20
- Pittayanon R., Lau, J.T., Leontiadis, G.I., Tse F., Yuan Y., Surette M., & Moayyedi P. (2020). Differences in Gut Microbiota in Patients With vs Without Inflammatory Bowel Diseases: A Systematic Review. *Gastroenterology*. 158(4):930-946.e1. https://doi.org/10.1053/ j.gastro.2019.11.294
- Power, K.A., Lu, J.T., Monk, J.M., Lepp D., Wu W., Zhang C., Liu R., Tsao R., Robinson, L.E., Wood, G.A., & Wolyn, D.J. (2016). Purified rutin and rutin-rich asparagus attenuates disease severity and tissue damage following dextran sodium sulfate-induced colitis. *Molecular Nutrition & Food Research*. 60(11):2396–2412. https:// doi.org/10.1002/mnfr.201500890
- Rao, A.V., Waseem Z., & Agarwal S. (1998). Lycopene content of tomatoes and tomato products and their contribution to dietary lycopene. *Food Research International*. 31(10):737–741. https://doi.org/10.1016/S0963-9969(99) 00053-8
- 50. Reboul E. (2013). Absorption of vitamin A and carotenoids by the enterocyte: Focus on transport proteins.

Nutrients. 5(9):3563–3581. https://doi.org/10.3390/ nu5093563

- Richards DJ., Li Y., Kerr CM., Yao J., Beeson GC., Coyle RC., Chen X., Jia J., Damon B., Wilson R., & Starr Hazard E.. Human cardiac organoids for the modelling of myocardial infarction and drug cardiotoxicity. Nature Biomedical Engineering. 2020 Apr;4(4):446-62.
- Rinninella E., Raoul P., Cintoni M., Franceschi F., Abele G., Miggiano D., Gasbarrini A., & Mele MC. (2019). What is the Healthy Gut Microbiota Composition ? A Changing Ecosystem across Age , Environment,Diet,and Diseases.*Microorganisms*.Jan;7(1):14.https://doi.org/10.3390/ microorganisms7010014
- Saini, R.K., Prasad P., Lokesh V., Shang X., Shin J., Keum, Y.S., & Lee, J.H. (2022). Carotenoids: Dietary Sources, Extraction, Encapsulation, Bioavailability, and Health Benefits—A Review of Recent Advancements. *Antioxidants*. 11(4). https://doi.org/10.3390/antiox1 1040795
- Santoru, M.L., Piras C., Murgia A., Palmas V., Camboni T., Liggi S., Ibba I., Lai, M.A., Orru S., & Blois S, *et al.* (2017). Cross sectional evaluation of the gut-microbiome metabolome axis in an Italian cohort of IBD patients. *Scientific Reports*. 7(1):1–14. https://doi.org/10.1038/ s41598-017-10034-5
- 55. Seishima J., Iida N., Kitamura K., Yutani M., Wang Z., Seki A., Yamashita Taro., Sakai Y., Honda M., & Yamashita Tatsuya *et al.* (2019). Gut-derived Enterococcus faecium from ulcerative colitis patients promotes colitis in a genetically susceptible mouse host. *Genome Biology*. 20(1):1–18. https://doi.org/10.1186/s13059-019-1879-9
- Se-Kwon, Kim., & Ratih Pangestuti (2011). Biological Activities and Potential Health Benefits of Fucoxanthin Derived from Marine Brown Algae . Advances in Food and Nutrition Research. Volume 64, Pages 111-128. https://doi.org/10.1016/B978-0-12-387669-0.00009-0
- Shimidzu N., Goto M., & Miki W. (1996). Carotenoids as Singlet Oxygen Quenchers in Marine Organisms. *Fisheries Sciences*. 62(1):134–137. https://doi.org/10.2331/ fishsci.62.134
- Shin, N.R., Whon, T.W., & Bae, J.W. (2015). Proteobacteria: Microbial signature of dysbiosis in gut microbiota. *Trends in Biotechnology*. 33(9):496–503. https://doi.org/10.1016/j.tibtech.2015.06.011
- Sokol H., Leducq V., Aschard H., Pham, H.P., Jegou S., Landman C., Cohen D., Liguori G., Bourrier A., & Nion-Larmurier I, *et al.* (2017). Fungal microbiota dysbiosis in IBD. *Gut.* 66(6):1039–1048. https://doi.org/10.1136/gutjnl -2015-310746
- Stahl W., & Sies H. (2012). β-Carotene and other carotenoids in protection from sunlight. *The American Journal* of *Clinical Nutrition*. 96(5):1179–1184. https:// doi.org/10.3945/ajcn.112.034819
- Story E.N., Kopec, R.E., Schwartz, S.J., & Keith Harris G. (2010). An update on the health effects of tomato lycopene. *Annual Review of Food Science and Technology*. 1(1):189–210. https://doi.org/10.1146/annurev.fo od.102308.124120
- Sun Xiaowen., Zhao H., Liu Z., Sun Xun., Zhang D., Wang S., Xu Y., Zhang G., & Wang D. (2020). Modulation of Gut Microbiota by Fucoxanthin during Alleviation of Obesity in High-Fat Diet-Fed Mice. *Journal of Agricul-*

tural and Food Chemistry. 68(18):5118–5128. https:// doi.org/10.1021/acs.jafc.0c01467

- Takaichi S. (2011). Carotenoids in algae: Distributions, biosyntheses and functions. *Marine Drugs*. 9(6):1101– 1118. https://doi.org/10.3390/md9061101
- Tanca A., Palomba A., Fraumene C., Manghina V., Silverman M., & Uzzau S. (2018). Clostridial Butyrate Biosynthesis Enzymes Are Significantly Depleted in the Gut Microbiota of Nonobese Diabetic Mice. *mSphere*. 3(5):1–5. https://doi.org/10.1128/msphere.00492-18
- Terasaki., Masaru., Osamu Uehara., Shinya Ogasa., Taishi Sano., & Atsuhito Kubota *et al.*, (2021). *Carcinogenesis*, Volume 42, Issue 2, Pages 210–219, https:// doi.org/10.1093/carcin/bgaa100
- Walter J. (2008). Ecological role of lactobacilli in the gastrointestinal tract: Implications for fundamental and biomedical research. *Applied and Environmental Microbiology*. 74(16):4985–4996. https://doi.org/10.1128/AEM.00 753-08
- Wang J., Liu S., Wang H., Xiao S., Li C., Li Y., & Liu B. *Xanthophyllomyces dendrorhous*-Derived Astaxanthin Regulates Lipid Metabolism and Gut Microbiota in Obese Mice Induced by A High-Fat Diet. Mar Drugs. 2019 Jun 5;17(6):337. doi: 10.3390/md17060337. PMID: 31195737; PMCID: PMC6627754.
- Wiese M., Bashmakov Y., Chalyk N., Nielsen, D.S., Krych L., Kot W., Klochkov V., Pristensky D., Bandaletova T., & Chernyshova M, *et al.* (2019). Prebiotic Effect of Lycopene and Dark Chocolate on Gut Microbiome with Systemic Changes in Liver Metabolism, Skeletal Muscles and Skin in Moderately Obese Persons. *Bio-Med Research International.* 2019. https:// doi.org/10.1155/2019/4625279
- 69. Wigg, A.J., Roberts-Thomson, I.C., Grose, R.H., Cummins, A.G., Dymock, R.B., & McCarthy, P.J. (2001). The role of small intestinal bacterial overgrowth, intestinal permeability, endotoxaemia, and tumour necrosis factor α in the pathogenesis of non-alcoholic steatohepatitis. *Gut.* 48(2):206–211. https://doi.org/10.1136/gut.48.2.206
- Wu L., Lyu Y., Srinivasagan R., Wu J., Ojo B., Tang M., El-Rassi, G.D., Metzinger K., Smith, B.J., & Lucas, E.A, *et al.* (2020). Astaxanthin-Shifted gut microbiota is associated with inflammation and metabolic homeostasis in mice. *Journal of Nutrition*. 150(10):2687–2698. https:// doi.org/10.1093/jn/nxaa222
- Xia H., Liu C., Li, C.C., Fu M., Takahashi S., Hu, K.Q., Aizawa K., Hiroyuki S., Wu G., Zhao L., & Wang XD. (2018). Dietary tomato powder inhibits high-fat diet-

promoted hepatocellular carcinoma with alteration of gut microbiota in mice lacking carotenoid cleavage enzymes. *Cancer Prevention Research*. 11(12):797–810. https://doi.org/10.1158/1940-6207.CAPR-18-0188

- Xu, Y.H., Gao, C.L., Guo, H.L., Zhang, W.Q., Huang W., Tang, S.S., Gan, W.J., Xu Y., Zhou H., & Zhu Q. (2018). Sodium butyrate supplementation ameliorates diabetic inflammation in db/db mice. *Journal of Endocrinology*. 238(3):231–244. https://doi.org/10.1530/JOE-18-0137
- Yan, A.W., Fouts, D.E., Brandl J., Starkel P., Torralba M., Schott E., Tsukamoto H., Nelson, K.E., & Brenner DA. (2012). *NIH Public Access*. 53(1):96–105. https:// doi.org/10.1002/hep.24018.Enteric
- Yonekura L & Nagao A. (2007). Intestinal absorption of dietary carotenoids. *Molecular Nutrition & Food Research*. 51(1):107–115. https://doi.org/10.1002/ mnfr.200600145
- Yuan, J.P., Peng J., Yin K., & Wang, J.H. (2011). Potential health-promoting effects of astaxanthin: A high-value carotenoid mostly from microalgae. *Molecular Nutrition & Food Research*.55(1):150–165. https://doi.org/10.1002/ mnfr.201000414
- Zhang H., Tang Y., Zhang Y., Zhang S., Qu J., Wang X., Kong R., Han C., & Liu Z. (2015). Fucoxanthin: A Promising Medicinal and Nutritional Ingredient. *Evidence-Based Complementary and Alternative Medicine*. https:// doi.org/10.1155/2015/723515
- Zhang L., Cao W., Gao Y., Yang R., Zhang X., Xu J., & Tang Q. (2020). Astaxanthin (ATX) enhances the intestinal mucosal functions in immunodeficient mice. *Food & Function*. 11(4):3371–3381. https://doi.org/10.1039/ c9fo02555c
- Zhao B., Wu J., Li J., Bai Y., Luo Y., Ji B., Xia B., Liu Z., Tan X., Lv J., & Liu X. (2019). Lycopene Alleviates DSS-Induced Colitis and Behavioral Disorders via Mediating Microbes-Gut-Brain Axis Balance. *Journal of Agricultural and Food Chemistry*. 68(13):3963–3975. https:// doi.org/10.1021/acs.jafc.0c00196
- Zhu L., Song Y., Liu H., Wu M., Gong H., Lan H., & Zheng X. (2021). Gut microbiota regulation and antiinflammatory effect of β-carotene in dextran sulfate sodium-stimulated ulcerative colitis in rats. *Journal of Food Science*. 86(5):2118–2130. https://doi.org/10.1111/1750-3841.15684
- Zia-ul-haq M. (2021). Carotenoids: Structure and Function in the Human Body. Springer publication. https:// doi.org/10.1007/978-3-030-46459-2