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Citation of this paper:

McLean, Cheynne; Wilson, Aze; and Kim, Richard B., "Impact of Transporter Polymorphisms on Drug Development: Is It Clinically Significant?" (2016). *Paediatrics Publications*. 2045. https://ir.lib.uwo.ca/paedpub/2045



Impact of Transporter Polymorphisms on Drug Development: Is It Clinically Significant?

The Journal of Clinical Pharmacology (2016), 56(S7) S40–S58 © 2016, The American College of Clinical Pharmacology DOI: 10.1002/icph.691

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Abstract

Drug transporters are becoming increasingly recognized as relevant to the drug development process. This may be a reflection of increasing target complexity and the need for high-affinity interaction with drug targets that minimize off-target side effects. Moreover, as new molecular entities (NMEs) become larger in size and amphipathic in nature, interaction with drug transporters, both uptake as well as efflux, becomes increasingly likely. In some cases transporters may limit the absorption or organ-specific entry of NMEs, whereas in other cases transporters may enhance their absorption or tissue accumulation. Indeed, in some cases, transporters may prove to be a therapeutic target. Accordingly, a better understanding of potentially clinically relevant drug transporter polymorphisms earlier in the drug development process is highly desirable. In this review we examine key transporters that are important to the absorption, distribution, and excretion of a large number of drugs in clinical use. Importantly, we provide our assessment of the potential impact of known polymorphisms in such transporters and discuss whether there is sufficient evidence to incorporate these polymorphisms in the drug development process.

Keywords

drug transporters, pharmacogenomics, drug development

Transporters are proteins localized to the plasma membrane and membranes of numerous subcellular organelles that function to facilitate the uptake or clearance of toxins, xenobiotics, and endogenous molecules important for cellular homeostasis. Transporters are classified into 2 superfamilies based on sequence homology: the ATP-binding cassette (ABC) family and the solute carrier (SLC) family. ABC transporters function as efflux transporters that directly harness energy produced by ATP hydrolysis. SLC transporters mediate import or, in some cases, export of substrates via an ion gradient established by ATPdependent pumps or an electrochemical gradient.¹ To date, 48 ABC transporters and 350 SLC transporters have been annotated in the human genome, and approximately 30 of these are involved in the disposition of drugs.^{2,3} Moreover, membrane transporters located on the intestinal epithelium, liver, and kidney modulate the overall pharmacokinetic (PK) profiles of new molecular entities (NMEs) and are therefore important considerations in drug development. In addition, membrane transporters expressed in organs that possess a barrier function, such as the blood-brain barrier, have proved to be highly relevant to drug development and therapeutic targeting.⁴ The locations of key transporters are summarized in Figure 1.

The discovery and development of safe and effective NMEs require the consideration of multiple

factors that impact drug disposition. Indeed, until recently, genetic variation in drug-metabolizing enzymes (DMEs) has been the major focus of pharmacogenetics. However, in the past decade, single nucleotide polymorphisms (SNPs) in membrane transporters have been demonstrated to be important, and sometimes rate-limiting, determinants of drug disposition and response.⁵ Recognizing the need to evaluate the impact of membrane transporters during the drug development process, the International Transporter Consortium (ITC) developed guidelines for assessing the importance of emerging key transporters in drug development.⁴ Subsequently, in 2013, the Food and Drug Administration (FDA) published a clinical pharmacogenomics guidance titled, "Clinical Pharmacogenomics: Premarketing Evaluation in Early-Phase Clinical Studies and Recommendations for Labeling."

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Submitted for publication 27 September 2015; accepted 2 December 2015.

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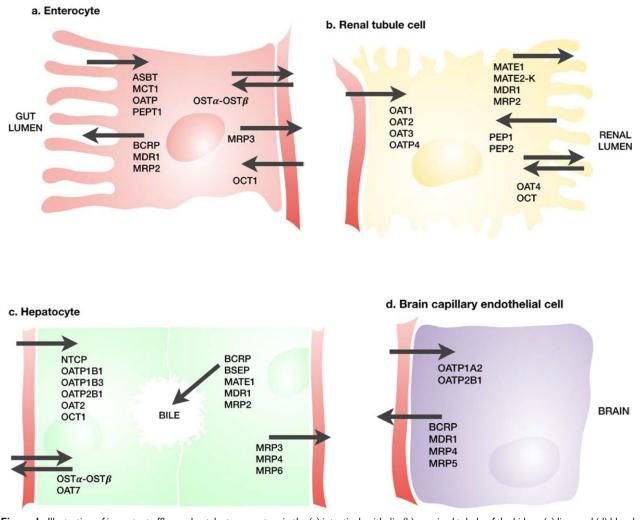


Figure 1. Illustration of important efflux and uptake transporters in the (a) intestinal epithelia, (b) proximal tubule of the kidney, (c) liver, and (d) bloodbrain barrier that may be involved in the absorption, distribution, metabolism, and excretion of a drug. Abbreviations: ASBT, apical sodium-dependent bile acid transporter; MCT, monocarboxylate transporter; OATP, organic anion-transporting polypeptide; PEPT, peptide transporter; BCRP, breast cancer resistance protein; MDR, multidrug resistance; OCT, organic cation transporter; OST, organic solute transporter; OAT, organic anion transporter; MATE, multiantimicrobial extrusion protein; URAT, urate transporter; OCTN, organic zwitterion/cation transporters; NTCP, Na⁺-taurocholate cotransporting polypeptide; BSEP, bile slat export pump. (Adapted from International Transporter Consortium.⁴)

This work sought to provide more specific guidance to the pharmaceutical industry with regard to genetic variants in both DMEs and membrane transporters and emphasized the incorporation of pharmacogenomics into the drug development process.

However, the field of drug transporters, particularly in relation to clinically relevant transporter polymorphisms, continues to evolve rapidly and merits ongoing assessment to aid in future drug development. Although the definition of clinically significant transporter polymorphisms remains debatable, based on observed drug-drug interactions associated with transporters, a 3-fold or higher increase in AUC of transporter substrate drug would fit such a definition. However, for many drugs in clinical use, clinical relevance may occur even at much lower AUC increase of substrate drugs. Accordingly, in this review, we summarize the key drug transporter polymorphisms associated with variations in drug pharmacokinetics and response. Specifically, we focus on SNPs within members of the organic cation transporters (OCTs), organic aniontransporting polypeptides (OATPs) family of SLC transporters, in addition to multidrug-resistant proteins (MRPs), breast cancer resistance protein (BCRP), and P-glycoprotein (P-gp, *MDR1*, *ABCB1*).

SLC Transporter Polymorphisms of Potential Clinical Significance

Organic Anion-Transporting Polypeptides

In humans, 12 organic anion-transporting polypeptides (OATP proteins; encoded by the SLCO gene family) have been identified with a wide substrate specificity for amphipathic molecules, including endogenous compounds such as bile acids and hormones, and xenobiotic substrates including statins, antidiabetics, and chemotherapeutic agents.^{6,7} Of these, OATP1B1 (SLCO1B1), OATP1B3 (SLCO1B3), and OATP2B1 (SLCO2B1) are largely expressed at the sinusoidal endothelium of the liver and mediate hepatocellular uptake of substrates from portal circulation; OATP2B1 is also expressed on intestinal endothelia, where it facilitates the absorption of its substrates.⁸ Interestingly, OATP1A2 (SLCO1A2) is expressed in the distal nephron, apical membrane of cholangiocytes, ciliary body epithelium of the eye, and capillary endothelia of the brain and is now thought to facilitate brain entry of certain medications.9 Select OATP variants and their effects are summarized in Table 1.

OATPIBI. Of the 40+ nonsynonomous variants that have been identified in human SLCO1B1, c.521T>C (rs4149056) in exon 5, first identified by Tirona et al, is the most extensively characterized.^{6,10} SLCO1B1 c.521T>C has an allele frequency of approximately 8% to 20% in European individuals with lower frequencies in East Asian and African-American individuals.¹¹ In vitro work suggests that the c.521T>C variant significantly impairs transport activity of several OATP1B1 substrates through reduced transporter expression at the plasma membrane.¹⁰ Clinically, the significance of SLCO1B1 c.521T>C is best exemplified by its effects on statins. For example, the areas under the plasma drug concentration-time curves (AUCs) for the pharmacologically active form of simvastatin (simvastatin acid) were 120% and 221% higher in c.521C/C healthy volunteers than in those with the c.521C/T and c.521T/T genotypes, respectively.¹² Furthermore, the c.521T>C variant was strongly associated with the risk for simvastatin-induced myopathy in a genome-wide association study (GWAS) that compared 85 patients with myopathy on high-dose simvastatin with 90 matched controls.¹³ These studies, in conjunction with others, spurred the Clinical Pharmacogenetics Implementation Consortium (CPIC) to include recommendations for genotype-informed dosing of statins in the 2014 guideline update.14

Another commonly studied SNP in *SLCO1B1* is c.388A>G (rs2306283), which has been shown to cause increased liver OATP1B1 expression in a white population.¹⁵ Allele frequencies for *SLCO1B1* c.388A>G appear to range from 26% to 77% in European and sub-Saharan individuals, respectively.¹¹ Unlike c.521T>C, c.388A>G is associated with lower plasma concentrations of various statins,^{16,17} and homozygous variant carriers have shown increased lowdensity lipoprotein (LDL) reduction in response to atorvastatin treatment when compared with heterozy-

gous and homozygous wild-type individuals.¹⁸ Interestingly, Teft et al found that although c.521C was significantly associated with increased exposure to the active metabolite of irinotecan, the progression-free survival (PFS) was significantly longer in c.388G/G advanced cancer patients treated with irinotecan-based regimes.¹⁹ Moreover, c.521T>C and c.388A>G can be in linkage disequilibrium and result in 4 functionally distinct haplotypes with varying effects on drug disposition.⁸

OATP1B3. c.334T>G (rs4149117) and c.699G>A (rs7311358) in SCLO1B3 are frequently observed across diverse ethnic populations and have been shown to alter the pharmacokinetics of certain medications.²⁰ For example, both variants have been associated with increased dose-adjusted AUC of the immunosuppressant mycophenolate mofetil (MMF) in Japanese renal transplant patients.²¹ In contrast, Picard et al investigated MMF plasma exposure in kidney transplant patients who were coadministered tacrolimus or sirolimus and found that carriers of at least 1 c.334T allele had a 1.4-fold higher AUC than carriers of the GG genotype.²² More recently, 334T > G has also been associated with increased imatinib clearance and increased accumulation within the leukocytes in chronic myeloid leukemia patients.^{23,24} Interestingly, inactivating mutations of SLCO1B1 and SLCO1B3 result in Rotor syndrome, a benign but very rare autosomal recessive liver disease associated with conjugated hyperbilirubinemia, impaired hepatic uptake of diagnostic dyes, and delayed clearance of certain anionic drugs.25 Clinical impact of SLCO1B3 genetic variation, particularly of relevance to drug development, remains to be clarified.

OATP2B1. Compared to the extensive knowledge on the clinical significance of polymorphisms in SLCO1B1, data surrounding the functional consequences of SLCO2B1 polymorphisms are limited. One of the first nonsynonymous variants described in SLCO2B1 was c.1457C>T, which has an allele frequency of approximately 3% and 31% in white and Asian individuals, respectively.⁸ Although c.1457C>T does not appear to affect OATP2B1 protein expression,¹⁵ it has been linked to decreased AUC of both celiprolol (β -blocker),²⁶ and fexofenadine (antihistamine) in healthy volunteers.²⁷ In contrast, Akamine et al found lower levels of S-fexofenadine in healthy Japanese variant carriers following a 60-mg dose of fexofenadine.²⁸ Similarly, SLCO2B1 c.935G>A (rs12422149) was associated with decreased plasma concentrations of montelukast (leukotriene receptor antagonist) and decreased symptom improvement in asthma patients in 1 study,29 and another study found no association between the variant and montelukast pharmacokinetics.³⁰ These conflicting findings

 Table 1. Organic Anion-Transporting Polypeptides

Variant	Effect on mRNA/Protein	Drug Class	In Vitro/ In Vivo	Allele Effect/Association	Drug/Effect Association ^a
Select Variants an	d Variant Effects Withi	n SLCOIA2 (OATPIA	.2)		
113T, 38T>C rs10841795		Antimetabolites	In vitro	Methotrexate: ↑ uptake (Xenopus laevis oocytes) ¹⁰⁴	Low/Preliminary
		Tyrosine kinase inhibitors	In vivo	Imatinib: No effect on average steady-state concentration in white cancer patients (n = 94) ³²	Low/Preliminary
Q172D, 516A>C rs11568563	Variant appears to cause a reduction in cell surface transporter expression ⁹	δ-Opioid receptor agonists	In vitro	Deltorphin II and [D-penicillamine-2,5]- enkephalin (DPDE):↓ uptake (HeLa cells) ³¹	Low/Preliminary
	·	Tyrosine kinase inhibitors	ln vitro	Imatinib : \downarrow uptake (HeLa cells) ³²	Low/Preliminary
			In vivo	Imatinib : No effect on drug levels in white cancer patients (n = 94) ³²	
–1105G>A rs4148977		Tyrosine kinase inhibitors	In vivo	Imatinib : \downarrow clearance in CML patients (n = 34) ³³	Low/Preliminary
–1032G>A rs4148978		Tyrosine kinase inhibitors	In vivo	Imatinib : \downarrow clearance in CML patients (n = 34) ³³	Low/Preliminary
-361G>A rs3764043		Tyrosine kinase inhibitors	In vivo	Imatinib : \uparrow clearance in CML patients $(n = 34)^{33}$	Low/Preliminary
Select Variants an	d Variant Effects Within	n SLCOIBI (OATPIB	I)		
V174A, 521T>C rs4149056	Variant causes reduced membrane transporter expression ¹⁰	Statins	In vivo	Simvastatin Acid: \uparrow AUC and C_{max} (n=32); ¹⁰⁵ variant strongly associated with \uparrow simvastatin-induced myopathy risk (n = 175) ¹³	High
			In vivo	Pravastatin : \uparrow drug plasma concentration in healthy white volunteers (n = 41) ¹⁰⁶	Moderate
			In vitro	Pravastatin : $\downarrow V_{max}$ (HEK 293 cells) ¹⁰⁷	
			In vivo	Atorvastatin and Rosuvastatin: \uparrow AUC in healthy volunteers (n = 32) ¹⁰⁸	Moderate
		Meglitinides	In vivo	Repaglinide: ↑ AUC in both homozygous and heterozygous variant carriers when compared to wild-type individuals (n = 56) ¹⁰⁹	Low/Preliminary
		Antimetabolites	In vivo	Methotrexate: ↓ clearance in children diagnosed with acute lymphoblastic leukaemia (n = 434) ¹¹⁰	Low/Preliminary
		Alkaloids	In vivo	Irinotecan: ↑ plasma exposure of active metabolite in advanced cancer patients (n = 127) ¹⁹	Low/Preliminary
N I 30D, 388A>G rs2306283	Variant causes increased liver protein expression in white population ¹⁵	Statins	In vivo	Pravastatin : \downarrow AUC in healthy Japanese volunteers (n = 23) ¹⁶	Low/Preliminary
			In vivo	Atorvastatin: ↑ LDL reduction for GG variant carriers when compared with GA and AA carriers (n=136) ¹⁸	Low/Preliminary

(Continued)

Table I. Continued

Variant	Effect on mRNA/Protein	Drug Class	In Vitro/ In Vivo	Allele Effect/Association	Drug/Effect Association ^a
			In vivo	Lovastatin Acid : \downarrow AUC in healthy white volunteers $(n = 27)^{17}$	Low/Preliminary
		Alkaloids	In vivo	<pre>Irinotecan: ↑ PFS in advanced cancer patients (n = 103)¹⁹</pre>	Low/Preliminary
		Cholesterol absorption inhibitors	In vivo	Ezetimibe : gene dose \downarrow in AUC in healthy volunteers $(n = 17)^{111}$	Low/Preliminary
		Meglitinides	In vivo	Repaglinide : \downarrow AUC and C _{max} , and \uparrow mean blood glucose concentration in healthy volunteers (n = 8) ¹¹²	Low/Preliminary
P155T, 463C>A rs11045819		Antimycobacterials	In vivo	Rifampin: ↓ AUC in adults with pulmonary tuberculosis from Africa, North America, and Spain (n = 72) ¹¹³	Low/Preliminary
–11187G>A , rs4149015		Statins	In vivo	Pravastatin : heterozygous carriers have a 98% \uparrow drug plasma concentration than homozygous wild-type individuals in healthy white volunteers (n = 41) ¹⁰⁶	Moderate
		Alkaloids	In vivo	Irinotecan: \uparrow AUC of active metabolite in Korean patients with advanced non-small-cell lung cancer (n = 81) ¹¹⁴	Low/Preliminary
Select Variants ar	d Variant Effects Withir	SLCOIB3 (OATPIB	3)		
SII2A, 334T>G rs4I49II7	Does not alter membrane protein localization ¹¹⁵	Immune suppressants	In vivo	$\begin{array}{l} \mbox{Mycophenolate mofetil:} \uparrow \\ \mbox{dose-adjusted AUC and } \downarrow \mbox{oral} \\ \mbox{clearance in Japanese renal} \\ \mbox{transplant recipients } (n = \\ \mbox{87})^{21}; \mbox{in contrast, a study in} \\ \mbox{kidney transplant patients} \\ \mbox{found } \downarrow \mbox{plasma concentrations} \\ \mbox{(n = 70)}^{22} \end{array}$	Low/Preliminary
		Tyrosine kinase inhibitors	In vivo	Imatinib : \uparrow clearance in Japanese CML patients $(n = 34)^{24}$	Low/Preliminary
			In vivo	Imatinib: ↑ intracellular accumulation in leukocytes of CML patients (n = 15) ²³	
M233I, 699G>A rs7311358	Does not alter membrane protein localization ¹¹⁵	Immune suppressants	In vivo	Mycophenolate mofetil: ↑ dose-adjusted AUC and ↓ oral clearance in Japanese renal transplant recipients (n = 87) ²¹	Low/Preliminary
VS12-5676A>G rs11045585	Does not affect protein expression ¹⁵	Mitotic inhibitors	In vivo	Docetaxel : \uparrow AUC, \downarrow oral clearance, and \uparrow risk of docetaxel-induced adverse events in Japanese cancer patients (n = 84) ¹¹⁶	Low/Preliminary
Select Variants ar R312Q, 935G>A	Id Variant Effects Withir Variant does not	Leukotriene	In vivo	Montelukast:↓ plasma	Low/Preliminary
rs12422149	appear to affect protein expression ¹⁵	receptor antagonists		concentrations and \downarrow symptom improvement in asthma patients (n = 489) ²⁹ ; in contrast, a study done in healthy volunteers found no association between the variant and montelukast	
				and montelukast pharmacokinetics $(n = 16)^{30}$	

Table I. Continued

Variant	Effect on mRNA/Protein	Drug Class	In Vitro/ In Vivo	Allele Effect/Association	Drug/Effect Association ^a
			In vitro	Montelukast: ↓ transport (Caco-2 cells) ²⁹	
S486F, 1457C>T rs2306168	Variant does not appear to affect protein expression ¹⁵	β -Blockers	In vivo	Celiprolol : \downarrow AUC in healthy participants (n = 30) ²⁶	Low/Preliminary
		Antihistamine	In vivo	Fexofenadine : \downarrow AUC in healthy individuals (n = 14) ²⁷ ; in contrast, the variant was associated with \uparrow AUC of S-fexofenadine in healthy Japanese volunteers (n = 24) ²⁸	Low/Preliminary

^a High: annotation for a variant-drug combination where most evidence demonstrates an association. The association must be replicated in more than 1 cohort with significant *P*-values. **Moderate**: annotation for a variant-drug combination with moderate evidence of an association. The association must be replicated in more than 1 cohort, but there may be some studies that do not show statistical significance. **Low/Preliminary**: annotation for a variant-drug combination based on a single significant study or annotation for a variant-drug combination evaluated in multiple cohorts but lacking clear evidence of an association; this classification also encompasses associations that have been studied only in vitro.

underscore the need for further investigations into *SLCO2B1* polymorphisms and their effects on drug disposition. Current knowledge does not support a major role of *SLCO2B1* SNPs in drug disposition; thus, a major focus on *SLCO2B1* genetic variation during development is unlikely to more fully predict clinical drug response or variation in the PK profile of a OATP2B1 substrate NME.

OATPIA2. To date, OATP1A2 has been the only human OATP transporter detected in the brain capillary endothelium and thus has the potential to play a role in the central nervous system penetration of certain medications.³¹ Our group had demonstrated decreased transport of δ -opioid receptor agonists (deltorphin II and [D-penicillamine-2,5]-enkephalin, DPDE) in HeLa cells expressing the SLCO1A2 variant c.516A>C (rs11568563).³¹ SLCO1A2 c.516A>C has an allele frequency of approximately 2% and 5% to 7% in African and white individuals, respectively, and appears to cause a reduction in cell surface transporter expression.^{8,9} The effects of SLCO1A2 c.516A>C have also been investigated in relation to imatinib (a tyrosine kinase inhibitor) pharmacokinetics. However, a significant change in imatinib plasma concentrations in 94 white cancer patients was not observed.³² Interestingly, another study found that common promoter polymorphisms in SLCO1A2 were associated with decreased (c.-361G>A, rs3764043) and increased (c.-1105G>A,rs4148977; c.-1032G>A, rs4148978) clearance of imatinib in Japanese myeloid leukemia patients.³³ Taken together, although OATP1A2 is likely to play an important role in the CNS entry of certain drugs, there is no compelling evidence to suggest functional SNPs in this transporter will have a major impact on substrate drug response, particularly in terms of PK.

In general, among the *SLCO* variants, the *SLCO1B1* variants c.521T>C and -11187G>A appear to have the greatest evidence in support of their impact on statin PK and toxicity profiles.

Organic Cation Transporters

The organic cation transporters (OCTs) belong to the SLC22 family.³⁴ In humans, the members of this subgroup include OCT1 (*SLC22A1*), OCT2 (*SLC22A2*), and OCT3 (*SLC22A3*). These proteins share a very similar structure and translocate a number of endogenous and exogenous substrates bidirectionally across cellular membranes, primarily through facilitated diffusion.^{35,36} OCT1 is predominantly expressed on the hepatocyte sinusoidal membrane, whereas OCT2 is positioned mainly within the proximal tubule of the kidney, and OCT3 is more broadly distributed.^{35,37} The OCTs have been implicated in regulating the disposition of numerous drugs including metformin, lamivudine, and numerous chemotherapeutics.³⁵

A number of SNPs in the genes encoding OCT1 and OCT2 have been identified and are associated with changes in the mRNA or protein expression of these transporters.^{38–42} Furthermore, many of these variants appear to be associated with reduced protein and/or mRNA expression resulting in changes in the in vitro and in vivo PK of biguanides, tyrosine kinase inhibitors, and dopamine precursors (Table 2).

OCT1. In vitro work using HEK293 cells transfected with either *SLC22A1* p.M420del (rs35191146)⁴³ or *SLC22A1* c.181C>T (rs12208357)⁴⁴ has demonstrated decreased uptake of metformin, a blood-glucose-lowering medication that exerts its effect by increasing peripheral insulin sensitivity.⁴⁵ Moreover, healthy volunteer data have demonstrated that

Variant	Effect on mRNA/Protein	Drug Class	In Vitro/ In Vivo	Allele Effect/Association	Drug/Effect Association ^a
Select Variants and	Variant Effects Within				
M420del, 1258delG rs35191146	↓ protein expression despite no difference in the amount of mRNA transcript ³⁹	Tyrosine kinase inhibitors	In vivo	Imatinib : \uparrow probability of treatment failure in chronic myeloid leukemia patients $(n = 336)^{39}$	Low/Preliminary
			In vitro	Imatinib: ↓ uptake, but this effect was countered if the M408V (rs628031) SNP was also present (KCL22-hOCT1 cells) ³⁹	
		Biguanides	In vitro	Metformin: ↓ uptake and ↑ sensitivity to metformin transport inhibition by medications such as verapamil and amitriptyline (HEK293 cells) ⁴³	Low/Preliminary
M408V, I 222A>G rs62803 I	↓ hepatic protein expression; no association with mRNA expression ^{38,49}	Tyrosine kinase inhibitors	In vivo	Imatinib: ↓ treatment response, 5-year survival, and event-free survival in chronic myeloid leukemia patients (n = 167) ⁴⁸	Low/Preliminary
		Biguanides	In vivo	Metformin: positive predictor of hematological, cytogenic, and major molecular responses in patients with type 2 diabetes $(n = 33)^{49}$; variant was not associated with glucose or lipid control in patients with type 2 diabetes $(n = 135)^{50}$	Low/Preliminary
			In vitro	Metformin : no change in uptake (HEK293 cells) ⁴³	
R61C, 181C>T rs12208357	↓ protein expression and membrane localization ^{40,42}	Biguanides	In vivo In vitro	Metformin: \uparrow AUC and C _{max} in healthy white volunteers ⁴⁴ ; another study found no association between the variant and metformin pharmacokinetics in patients with type 2 diabetes (n = 120), healthy white individuals (n = 16), or healthy Malaysian individuals (n = 169) ⁴⁶ Metformin: \downarrow uptake	Low/Preliminary
			In vitro	(HEK293 calls) ⁴²	
386C>A rs622342		Biguanides	In vivo	Metformin: \downarrow efficacy in white (n = 102) ⁵¹ and South Indian (n = 122) ¹¹⁷ patients with type 2 diabetes	Moderate
		Dopamine Precursor	In vivo	Levodopa: ↑ doses and ↑ mortality in Parkinson patients (n = 99) ¹¹⁸	Low/Preliminary
160L, 480C>G rs683369	No association with altered mRNA expression ³⁸	Tyrosine kinase inhibitors	In vivo	Imatinib: \downarrow in apparent drug clearance in chronic myeloid leukemia patients carrying at least 1 polymorphic allele (n = 60) ¹¹⁹	Low/Preliminary

Table 2. Organic Cation Transporters

(Continued)

Table 2. Continued

Effect on mRNA/Protein	Drug Class	In Vitro/ In Vivo	Allele Effect/Association	Drug/Effect Association ^a
nd Variant Effects Within	SLC22A2 (OCT2)			
Inconclusive data regarding changes in protein expression; variant may alter substrate recognition and translocation ^{47,120,121}	Alkylating agents	In vivo	Cisplatin : no association between variant and pharmacokinetic parameters in white cancer patients (n = 106) ¹²² ; variant was found to be protective against cisplatin-induced ototoxicity in 64 pediatric patients and 66 adult patients ¹²³ ; variant was found to be protective against nephrotoxicity in cancer patients (n = 53) ¹²⁴	Low/Preliminary
	Biguanides	In vivo In vitro	Metformin: \uparrow AUC/C _{max} and \downarrow renal clearance in healthy Korean individuals (n = 26) ⁵⁴ ; variant associated with \downarrow renal clearance in healthy white and African-American individuals (n = 23) ¹²⁰ Metformin: \downarrow uptake	Low/Preliminary
	mRNA/Protein ad Variant Effects Within a Inconclusive data regarding changes in protein expression; variant may alter substrate recognition and	mRNA/Protein Drug Class d Variant Effects Within SLC22A2 (OCT2) Inconclusive data Alkylating agents regarding changes in protein expression; variant may alter substrate recognition and translocation ^{47,120,121}	mRNA/ProteinDrug ClassIn Vitro/ In VivoIn Vitro/ Un VivoIn vivoIn vivoInconclusive dataAlkylating agentsIn vivoregarding changes in protein expression; variant may alter substrate recognition and translocation47,120,121In vivo	mRNA/ProteinDrug ClassIn Vitro/ In VivoAllele Effect/Associationad Variant Effects Within SLC22A2 (OCT2)Inconclusive dataAlkylating agentsIn vivoCisplatin: no association between variant and pharmacokinetic parameters in white cancer patients (n = 106) ¹²² ; variant was found to be protective against cisplatin-induced ototoxicity in 64 pediatric patients and 66 adult patients ¹²³ ; variant was found to be protective against nephrotoxicity in cancer patients (n = 53) ¹²⁴ BiguanidesIn vivoMetformin: \uparrow AUC/C _{max} and \downarrow renal clearance in healthy Korean individuals (n = 26) ⁵⁴ ; variant associated with \downarrow renal clearance in healthy white and African-American individuals (n = 23) ¹²⁰

^a High: annotation for a variant-drug combination where most evidence demonstrates an association. The association must be replicated in more than 1 cohort with significant *P*-values. **Moderate**: annotation for a variant-drug combination with moderate evidence of an association. The association must be replicated in more than 1 cohort, but there may be some studies that do not show statistical significance. **Low/Preliminary**: annotation for a variant-drug combination evaluated in multiple cohorts but lacking clear evidence of an association; this classification also encompasses associations that have been studied only in vitro.

individuals carrying a reduced-function SLC22A1 c.181C>T allele had an increased AUC and higher maximum plasma concentration of metformin compared to those homozygous wild-type individuals;⁴⁴ however, patient data did not replicate these findings.⁴⁶ The prevalence of these 2 polymorphisms is variable depending on the ethnic group being evaluated. SLC22A1 p.M420del is more commonly seen in white (18.5%) and Mexican (21.4%) Americans than in African Americans (2.9%), whereas c.181C>T is less prevalent with frequencies of 7.2% and 5.6% for white and Mexican Americans, respectively.⁴⁷ Furthermore, the presence of variant SLC22A1 p.M420del in KCL22-hOCT1 cells was also associated with reduced imatinib uptake, although the effect was obliterated in the presence of SLC22A1 c.1222A>G (rs628031).³⁹ This effect translated to findings in imatinib-treated chronic myeloid leukemia (CML) patients, where an increased risk of treatment failure was seen in those carrying the variant allele.48

SLC22A1 c.1222A>G (rs628031) is found in 74% of African American, 60% of white American, and 74% to 80% of Asian individuals.⁴⁷ Although in vitro studies did not correlate with reduced uptake of metformin in transfected HEK293 cells,⁴³ a small study in 33 patients with type 2 diabetes on metformin revealed

that the variant allele was a positive predictor of metformin efficacy.⁴⁹ This finding was not supported by a larger, more recent patient-based study, which showed that the *SLC22A1* variant c.122A>G did not correlate with blood glucose levels or lipid control in 135 diabetic subjects.⁵⁰ Additionally, a fourth *SLC22A1* variant, c.1386C>A (rs622342), has been associated with reduced metformin efficacy in white⁵¹ and south Indian diabetic patients.⁵²

OCT2. In comparison to *SLC22A1*, few variants within *SLC22A2* have been assessed for drug associations. However, *SLC22A2* c.808G>T (rs316019), which has an overall prevalence of approximately 7% to 16%,⁴⁷ has been associated with decreased metformin transport in vitro.⁵³ Moreover, Song et al determined that the *SLC22A2* c.808G>T variant was associated with increased metformin AUC and decreased renal clearance in healthy Korean individuals.⁵⁴ However, this association has yet to be validated in a patient population.

Overall, there is sufficient evidence to suggest that a number of commonly occurring SNPs in *SLC22A1* should be considered early in the drug development process, when an NME is deemed to be a substrate of this transporter. However, evidence does not suggest appreciable drug-variant associations for OCT2. Overall, our understanding of the impact of genetic variation in the OCTs is still in its infancy, with only preliminary or single-study evidence available in most cases. The exception appears to be the *SLC22A1* variant 1386C>A, which appears to have a moderate association with metformin efficacy.

Efflux Transporter Polymorphisms of Potential Clinical Significance

P-Glycoprotein

P-Glycoprotein (P-gp) is highly studied in drug development because of its widely appreciated capability for mediating the efflux transport of a broad array of structurally divergent endogenous and xenobiotic compounds.⁵⁵ P-gp substrates include various antiepileptics, statins, selective estrogen receptor modulators (SERMs), protease inhibitors, and a number of chemotherapeutic agents and immunosuppressants.⁵⁶

The genetic variability of *MDR1* has been extensively studied in vivo and in vitro, and a number of *MDR1* SNPs have been identified (Table 3).⁵⁷ Most notably, nonsynonymous SNPs in exons 12 (c.1236C>T), 21 (g.2677G>T), and 26 (c.3435C>T) have been researched extensively with regard to their impact on P-gp substrate pharmacokinetics.⁵⁸ Although not shown in vitro, of 15 identified MDR1 SNPs, Groups have linked homozygous variant carriers of c.3435T to reduced P-gp tissue expression and higher drug plasma concentrations.^{58,59} Moreover, several groups have shown that c.3435C>T, g.2677G>T, and c.1236C>T are inherited in linkage disequilibrium.^{60,61}

Not all substrates of P-gp are impacted by genetic variation in *MDR1*. Hoffmeyer et al showed an increase in digoxin plasma concentrations in individuals homozygous for the *MDR1* c.3435T variant, but this has not been replicated in other studies.^{56,58} The effect of the *MDR1* c.3435C>T SNP on the efficacy of antiepileptics such as carbamazepine has been extensively reviewed. Although 1 systematic review of 4269 Chinese adult patients revealed an association between the CC genotype and anti-epileptic resistance, similar reviews in larger and more diverse populations have failed to show any association between *MDR1* c.3435C>T and antiepileptic drug response.^{62–64}

Limited data exist regarding the effect of MDR1 polymorphisms on statin pharmacokinetics and patient drug response. Homozygous variant carriers of MDR1 c.3435C>T and g.2677G>T have both been linked to increased statin exposure and reduced statin effect. Both the AUC and maximum concentrations of pivastatin and rosuvastatin were increased in healthy Chinese volunteers with the MDR1 g.2677TT and

c.3435TT genotypes, respectively.^{65,66} Similarly, a small study in Bosnian volunteers found a smaller statin effect in individuals homozygous for the c.3435T variant.⁶⁷

The P2y12 inhibitor clopidogrel has been shown to be a substrate of P-gp in vitro, and its oral absorption is affected by the inhibition or upregulation of P-gp.⁶⁸ The *MDR1* c.3435C>T SNP has been shown to modulate the pharmacokinetics and efficacy in multiple patient cohorts; however, the findings were contradictory to the posited physiologic effect of the variant. In a group of 60 white acute coronary syndrome (ACS) patients, individuals with the TT genotype had a lower clopidogrel AUC and maximal concentration.⁶⁸ This was duplicated in a group of 401 Chinese ACS patients, where the c.3435T variant was associated with lower plasma concentrations of both clopidogrel and its active metabolite and resulted in reduced platelet inhibition.⁶⁹

All 3 exonic SNPs, *MDR1* c.3435C>T, c.1236C>T, and g.2677G>T, have been linked to changes in protease inhibitor pharmacokinetics and effect, although the evidence is not robust. In vitro, HEK293T cells carrying the g.2677T variant did not transport telaprevir, and higher telaprevir trough plasma concentrations were seen among 29 hepatitis C patients carrying a variant allele.⁷⁰ A Brazilian study of 187 HIV patients, among whom 27 were protease inhibitor non-responders, were assessed for their *MDR1* c.3435C>T genotype. Conversely, individuals carrying the c.3435T variant were more likely to fail first-line HIV treatment regimens compared to those carrying a wild-type allele.⁷¹

Moreover, the immunosuppressants and chemotherapeutics cyclosporine, methotrexate, and imantinib have been shown to be affected by genetic variation in MDR1. Renal transplant patients homozygous for the MDR1 c.3435T or the g.2677T variant had higher cyclosporine trough plasma concentrations compared to individuals carrying a wild-type allele.⁷² In a small Korean population of post-stem-cell-transplant patients, individuals carrying 2 variant MDR1 c.3435T alleles had lower methotrexate clearance.⁷³ Similarly, a small Spanish cohort showed that the MDR1 c.34335CC genotype was associated with primary failure of imantinib treatment in CML.74 A larger Chinese metaanalysis contradictorily found that the MDR1 c.1236T variant was a possible risk factor for nonoptimal clinical response to imantinib treatment in Asian patients with CML.75

Overall, although the c.3435T variant appears to be linked to digoxin and chemotherapeutic pharmacokinetics, the evidence for other *MDR1* variant-drug associations needs more study in larger cohorts.

Table 3. P-Glycoprotein

Variant	Effect on mRNA/Protein	Drug Class	In Vitro/In Vivo	Allele Effect/Association	Drug/Effect Association ^a
Select Variants a	nd Variant Effects Within	MDR/ (P-gp)			
3435C>T rs1045642	↓ duodenal protein expression ⁵⁸	Glycosides	In vivo	Digoxin: ↑ AUC in healthy white volunteers (n = 21) ⁵⁸	Moderate
		Antiepileptics	In vivo	Multiple AEDs: CC genotype associated with drug resistance in a Chinese population (n = 4269) ⁶³	Low/Preliminary
		HMG-CoA reductase inhibitors	In vivo	Simvastain and Atorvastatin:↓ benefit of statin therapy in dyslipidemic patients (n = 180) ⁶⁷	Low/Preliminary
		P2y12 inhibitor	In vivo	Clopiogrel : \downarrow AUC in patients with acute coronary syndrome $(n = 401)^{69}$	Low/Preliminary
		Glucocorticoid	In vivo	Prednisone: variant associated with ↑ risk for steroid-induced osteonecrosis of the femoral head in Chinese population (n = 200) ¹²⁵	Low/Preliminary
		Protease inhibitors	In vivo	Multiple Protease Inhibitors: variant associated with ↑ failure of first-line regimes in Brazilian HIV-infected patients (n = 187) ⁷¹	Moderate
		Antimetabolites	In vivo	Methotrexate: \downarrow clearance in Korean patients after hematopoietic stem cell transplantation (n = 20) ⁷³	Moderate
		Tyrosine kinase inhibitors	In vivo	Imatinib: CC genotype associated with primary failure in chronic myeloid leukemia patients $(n = 65)^{74}$	Low/Preliminary
S893A/T, 2677G>T/A rs2032582		Protease inhibitors	In vivo	Telprevir : ↑ trough concentrations in hepatitis C patients (n = 29) ⁷⁰	Low/Preliminary
		Immune suppressant	In vivo	Cyclosporine: ↑ trough concentrations in TT vs GG/GT renal transplant recipients (n = 101) ⁷²	Low/Preliminary
		Behavior modifier	In vivo	OROS-methylphenidate: TT genotype found to be an independent determinant of adverse drug reactions in children with attention-deficit hyperactivity disorder (n = 134) ¹²⁶	Low/Preliminary
		HMG-CoA reductase inhibitors	In vivo	Pivastatin : \uparrow AUC and C _{max} in non-G carriers $(n = 12)^{66}$	Low/Preliminary
1236C>T rs1128503		Protease inhibitors	In vivo	Telprevir : ↑ trough concentrations in hepatitis C patients (n = 29) ⁷⁰	Low/Preliminary
		Tyrosine kinase inhibitors	In vivo	Imatinib: T variant associated with nonoptimal clinical response in Asian chronic myeloid leukemia patients (meta-analysis) ⁷⁵	Moderate

^a**High**: annotation for a variant-drug combination where most evidence demonstrates an association. The association must be replicated in more than 1 cohort with significant *P*-values. **Moderate**: annotation for a variant-drug combination with moderate evidence of an association. The association must be replicated in more than 1 cohort, but there may be some studies that do not show statistical significance. **Low/Preliminary**: annotation for a variant-drug combination based on a single significant study or annotation for a variant-drug combination evaluated in multiple cohorts but lacking clear evidence of an association. This classification also encompasses associations that have been studied only in vitro.

The Multidrug Resistance Proteins MRP2, MRP3, and MRP4 $% \left({{\left({{{\rm{MRP3}}} \right)}_{\rm{max}}} \right)$

Common genetic variants in *ABCC2* (MRP2), *ABCC3* (MRP3), and *ABCC4* (MRP4) have been linked to changes in mRNA and/or protein expression as well to changes in the disposition of many drugs in vivo including statins and antimetabolites such as methotrexate and azathiopine/6-mp (Table 4). However, genetic variation within *ABCC2* is most widely studied. This may be due, in part, to the propensity of *ABCC2* to tolerate major loss of function with genetic variation. Indeed, complete loss of MRP2 function is the molecular basis for Dubin-Johnson syndrome.⁷⁶

Interestingly, *ABCC2* c.1249G>A (rs2273697) is associated with reduced mRNA expression in human placenta but increased mRNA expression in human liver.^{77,78} *ABCC2* c.1249G>A is found in 9% of Asian, 22% of sub-Saharan African, and 24% of white individuals.⁷⁹ Conversely, *ABCC2* variant, rs717620 is associated with reduced *ABCC2* mRNA in healthy kidney but not in liver or intestine, and variant rs4793665 is associated with reduced *ABCC3* mRNA expression in liver tissue samples.^{77,80} The frequency of these variants is 20% in Asian, white, and African populations for the former and 56% to 86% across these groups for the latter.^{81,82}

Genetic variations in *ABCC2* and *ABCC4* have been linked to alterations in methotrexate pharmacokinetics and efficacy. Specifically, *ABCC2* variants c.1249G>A (rs2273697) and c.3972C>T (rs3740066) were associated with an increased risk of gastrointestinal toxicity such as nausea and vomiting as well as hepatotoxicity in a cohort of 65 children suffering from acute lymphoblastic leukemia (ALL).⁸³ Similarly, in a cohort of 112 Chinese children with ALL, the *ABCC2* variant c.-24C>T (rs717620) was associated with increased plasma concentrations as well as an increased risk of clinical adverse drug reactions.⁸⁴ The *ABCC4* variant c.1372A>C (rs9516519) is also associated with elevated methotrexate plasma concentrations as well as drug toxicity in children suffering from ALL.⁸⁵

Taken together, SNPs in *ABCC2* and possibly *ABCC4* should be considered as potentially relevant during the drug development process if these transporters are considered to be essential or rate limiting to the elimination of a NME in development.

Breast Cancer Resistance Protein

This efflux transporter, encoded by *ABCG2* gene, is an ATP-binding cassette efflux half transporter that likely functions as a homodimer.⁴ Similar to P-glycoprotein, breast cancer resistance protein (BCRP) appears to serve as a barrier to limit the absorption, entry, or retention of diverse substrates including both endogenous (eg, dietary flavonoids, porphyrins, estrone 3-sulfate)

and exogenous (eg, statins, antineoplastics) molecules into various tissue compartments.⁸⁶

Pharmacogenetic studies have identified a number of common SNPs in ABCG2 that are associated with changes in the PK parameters for medications (Table 5). Among the SNPs identified to date, a nonsynonymous SNP c.421C>A (rs22331142) in exon 5 of BCRP, which results in a glutamine-to-lysine change at amino acid position 141, is the most extensively studied.⁸⁷ ABCG2 c.421C>A has an allele frequency of approximately 10% with higher frequencies in Asian individuals and lower frequencies in sub-Sahra African and African-American individuals.^{88,89} In vitro work suggests that the c.421C>A variant impairs protein expression by 30% to 40% but maintains mRNA expression when compared with wild-type controls.⁹⁰ Reduced BCRP expression is possibly due to enhanced susceptibility to ubiquitin-mediated proteasomal degradation and incomplete trafficking to the plasma membrane.^{91,92}

Carriers of the c.421C>A polymorphism have demonstrated increased bioavailability of marketed BCRP substrates. For example, the AUC for atorvastatin and fluvastatin were 72% greater, for simvastatin lactone 111% greater, and for rosuvastatin 144% greater in ABCG2 c.421A/A individuals than in c.421C/C individuals.93,94 Furthermore, the ABCG2 c.421C>A variant has been found to correlate in a gene-dosedependent manner with reductions in LDL cholesterol in individuals taking rosuvastatin to manage their hypercholesterolemia.^{95,96} The c.421C>A polymorphism also appears to impact the disposition and efficacy of antineoplastic medications. For example, Sparreboom et al determined that heterozygous cancer patients receiving a 20-minute infusion of diflomotecan had significantly higher plasma concentrations of the medication when compared to homozygous wild-type individuals.⁹⁷ Similar effects were also seen in vitro for another topoisomerase inhibitor, topotecan, where expression of the variant was associated with 30% reduction in efflux transport of topotecan in HEK293 cells when compared to wild type.⁹⁸ Similarly, ABCG2 c.421C>A has also been associated with increased accumulation of imatinib, a tyrosine kinase inhibitor, in HEK293 cells.99 More recently, Koo et al investigated the 5-year PFS rate in 209 gastrointestinal stromal tumor patients being treated with imatinib and found that PFS was vastly superior in AA-carrying individuals when compared with patients carrying the CC or CA genotypes.¹⁰⁰ Interestingly, another recent study suggested that ABCG2 c.421C>A polymorphism may affect substrate distribution to the brain, as the mean cerebrospinal fluid concentration of raltegravir, an anti-HIV medication, was significantly lower among AA and CA patients when compared with those individuals with the CC genotype.¹⁰¹

Table 4. Multidrug-Resistant Proteins

Variant	Effect on mRNA/Protein	Drug Class	In Vitro/ In Vivo	Allele Effect/Association	Drug/Effect Association ^a
Select variants and	variant effects within AE	CC2 (MRP2)			
V4171, 1249G>A rs2273697	Variant energies within A2 with reduced mRNA expression in human preterm placenta ⁷⁸ and associated with increased mRNA expression in human hepatic tissue ⁷⁷	Antimetabolites	In vivo	Methotrexate: variant associated with \uparrow GI toxicity in children with acute lymphoblastic leukemia $(n = 65)^{83}$	Low/Preliminary
		Antiepileptics	In vivo	Multiple antiepileptics: meta-analysis indicated that the variant genotypes (in Asian or white populations) were associated with a significantly \downarrow risk of antiepileptic resistance ¹²⁷ ; no association between the variant and antiepileptic response was found in Croatian epilepsy patients (n = 97) ¹²⁸	Low/Preliminary
–24C>T rs717620	Variant associated with lower ABCC2 mRNA levels in the normal human kidney ¹²⁹ but not in the intestine ¹³⁰ or liver ⁷⁷	Antimetabolites	In vivo	Methotrexate: ↑ plasma concentrations ↑ risk of hematologic and nonhematologic adverse events in Chinese children with acute lymphoblastic leukemia (n = 112) ⁸⁴	Low/Preliminary
	or liver	Statins	In vivo	Simvastatin and atorvastatin: variant associated with a dose \downarrow or switches to other cholesterol-lowering drugs in simvastatin, but not atorvastatin, users (n = 1014) ¹³¹	Low/Preliminary
11 3241, 3972C>T rs3740066		Antimetabolites	In vivo	Methotrexate: variant associated with ↑ hepatotoxicity in children with acute lymphoblastic leukemia (n = 65) ⁸³	Low/Preliminary
	Variant Effects Within A	. ,	1	Manual tax	L. (D.).
-211C>T rs4793665	Lower mRNA expression in liver samples ^{81,132}	Opiates	In vivo	Morphine: associated with ↓ levels of morphine-6-glucoronide and morphine-3-glucoronide in children undergoing outpatient adenotonsillectomy (n = 220) ¹³³	Low/Preliminary
	Variant Effects Within	. ,	In vive	Azathionwine/6 MD: *	low/Prolimine
E857K, 2269G>A rs3765534	Impaired membrane localization ¹³⁴	Immune suppressants/ antimetabolites	In vivo	Azathioprine/6-MP:↑ hematopoitic toxicity in IBD patients (n = 235) ¹³⁵	Low/Preliminary
3225+1243C>A rs9561778		Alkylating agents	In vivo	Cyclophosphamide : ↑ adverse drug reactions (ie, gastrointestinal toxicity and leukopenia/neutropenia) in 403 breast cancer patients ¹³⁶	Low/Preliminary

Table 4.	Continued
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	Effect on		In Vitro/ In		Drug/Effect
Variant	mRNA/Protein	Drug Class	Vivo	Allele Effect/Association	Association ^a
1372A>C rs9516519		Antimetabolites	In vivo	Methotrexate: variant associated with ↑ methotrexate levels and toxicity in children with acute lymphoblastic leukemia	Low/Preliminary

^aHigh: annotation for a variant-drug combination where most evidence demonstrates an association. The association must be replicated in more than 1 cohort with significant P-values. Moderate: annotation for a variant-drug combination with moderate evidence of an association. The association must be replicated in more than I cohort, but there may be some studies that do not show statistical significance. Low/Preliminary: annotation for a variant-drug combination based on a single significant study or annotation for a variant-drug combination evaluated in multiple cohorts but lacking clear evidence of an association. This classification also encompasses associations that have been studied only in vitro.

Of particular importance to drug development is the recent association of Jr(a-) blood group with nullallelic mutations in ABCG2.^{102,103} In essence, those who have the Jr(a-) could be considered ABCG2 knockout humans. The Jr(a-) phenotype is mainly detected in Japanese individuals with region-dependent frequencies between 0.3% and 1.7%.¹⁰³ In the Jr(a-) population, the production of anti-Jr(a) antibodies is known to be involved in hemolytic disease, but little is known about the effect the nonfunctional BCRP protein has on drug disposition and efficacy. The Jr(a-) population may aid in establishing the precise role of ABCG2 in the pharmacokinetics of a NME.

Therefore, given the presence of common as well as rare functional genetic variations in BCRP in the human population, and the presence of moderate to strong variant-drug associations, SNPs in ABCG2 should most certainly be considered as a part of the overall drug development process for a substrate NME.

Drug Discovery and Development Strategies

There have been a number of excellent publications in the past 5 years, in terms of in vitro as well as in vivo guidance for transporter-related studies during drug discovery and development.²⁻¹⁰ Shown below is a very brief 6-step summary of what should be viewed as our opinion, with regard to drug transporters and clinically relevant polymorphisms.

Step 1: Figure Out Extent of Metabolism vs Transport

In vitro or liver microsome studies should quickly demonstrate the likely extent of metabolism. If the NME is shown to be metabolically stable, anionic, cationic, zwitterionic, or amphipathic, with a molecular weight between 300 and 1000 daltons, transporters are likely to be relevant in vivo.

Step 2: Transporter(s) to Screen

It is not possible to screen all the transporters. The decision with regard to which subset of transporter to screen should be based on therapeutic target and likely organ(s) involved in the elimination of the NME. For CNS-active compounds, P-gp and BCRP should be considered. For those cleared by liver or targeting liver, consider OATP1B1, P-gp, or BCRP. If the NME is cleared by the kidney, particularly if the renal clearance appears to exceed GFR, consider OAT1/3 for anionic NMEs and OCT2 and MATE2-K for cationic NMEs in addition to P-gp.

Step 3: OATPIBI, P-gp, and BCRP

These are the top 3 transporters to keep in mind if a broader screening effort for various NMEs is to be carried out earlier in the discovery process. These transporters have shown the most clinical relevance to date. Additional transporters might be considered later in discovery or during development.

Step 4: Determine Transporter Inhibition-Associated DDI Risk

There is now significant guidance with regard to in vitro transporter studies and prediction of in vivo relevance. In vitro inhibition data could be used as a surrogate for loss of function polymorphism(s) in such transporters.

Step 5: If Worried About OATPIBI, P-gp, or BCRP Polymorphisms

A number of functional SNPs in these transporters result in loss of cell surface trafficking of the variant transporter. Thus, instead of worrying about whether there may be polymorphism-dependent differences in substrate specificity, it may be easier to skip in vitro analysis of variant transporters and directly carry out in vivo assessment in relevant rodent knockout models.

Table 5. Breast Cancer Resistance Protein

Variant	Effect on mRNA/Protein		In Vitro/In Vivo	Allele Effect/Association	Drug/Effect Association ^a
Variant	mkinA/Protein	Drug Class	¥1VO	Allele Effect/Association	Association
Select Variants ar	nd Variant Effects Within A	ABCG2 (BCRP)			
Q141K,421C>A rs2231142	↓protein expression and trafficking to plasma membrane ^{90,91}	HMG-CoA reductase inhibitors	In vivo	Rosuvastatin : \uparrow AUC in healthy Chinese $(n = 7)^{137}$ and Finnish $(n = 32)^{94}$ volunteers; superior LDL-C lowering in Chinese patients with hypercholesterolemia (n = 305) ⁹⁶ and white patients who had recently suffered a	High
				myocardial infarction $(n = 601)^{95}$	
			In vivo	Atorvastatin: \uparrow AUC in healthy Finnish (n = 32) ⁹⁴ volunteers	Moderate
		Topoisomerase inhibitors	In vivo	$\begin{array}{l} \mbox{Difflomotecan:} \uparrow AUC \mbox{ and } C_{max} \\ \mbox{ in white cancer patients} \\ \mbox{ receiving 20-minute infusion} \\ (n=22)^{97} \end{array}$	Low/Preliminary
			In vitro	Topotecan : ↓ efflux transport (HEK293 cells) ⁹⁸	Low/Preliminary
		Tyrosine kinase inhibitors	In vivo	Imatinib: 5-year PFS rate in gastrointestinal stromal tumor patients with the AA variant significantly superior to that of patients with CC/CA	Low/Preliminary
			In vitro	genotypes (n = 209) ¹⁰⁰ Imatinib: ↑ drug accumulation (HEK293 cells) ⁹⁹	
		Anivirals	In vivo	Raltegravir: \downarrow CSF concentrations in HIV-infected patients (n = 14) ¹⁰¹	Low/Preliminary
V12M, 34G>A	Possible \downarrow	Tyrosine kinase	In vivo		Low/Preliminary
rs2231137	localization to apical plasma membrane ¹³⁸	inhibitors		Gefitinib, Erlotinib, Icotinib : \uparrow median overall survival of Chinese advanced non-small-cell lung cancer patients (n = 100) ¹³⁹ Imatinib : homozygous wild-type carriers showed impaired cytogenetic response when compared to variant carriers (n = 229) ¹⁴⁰	
-15994C>T rs7699188	↑ multitissue protein expression ¹⁴¹	Tyrosine kinase inhibitors	In vivo	Imatinib: ↑ oral clearance in gastrointestinal stromal tumor patients (n = 82) ¹⁴¹	Low/Preliminary
18271G>A rs1564481	No association with mRNA expression ³⁸	CDK9 kinase inhibitor	In vivo	Flavopiridol: \uparrow AUC and improved response to treatment (n = 51) ¹⁴²	Low/Preliminary

^a**High**: annotation for a variant-drug combination where most evidence demonstrates an association. The association must be replicated in more than 1 cohort with significant *P*-values. **Moderate**: annotation for a variant-drug combination with moderate evidence of an association. The association must be replicated in more than 1 cohort, but there may be some studies that do not show statistical significance. **Low/Preliminary**: annotation for a variant-drug combination based on a single significant study or annotation for a variant-drug combination evaluated in multiple cohorts but lacking clear evidence of an association. This classification also encompasses associations that have been studied only in vitro.

Step 6: In Phase I and 2 Studies, Collect DNA and Assess Impact on PK Relative to Transporter Genotype(s)

For NMEs that are subject to transport, having genotype-vs-PK data as a part of the overall

development process will provide meaningful data in terms of likely clinical impact. The key will be to include DNA collection and genotyping strategies as a part of the development process. Note that effect of genotype and DDI may provide additional insights relating to the overall clinical impact because many drugs are substrates of multiple transporters, and a number of clinical DDI studies focus specifically on key efflux transporters such as P-gp.

Conclusion

Predicting all the factors that can account for interpatient variation in drug response is an important goal of the drug development process. It is now becoming clear that genetic variations within drug transporters are clinically relevant. Specifically, in 2014, international statin-dosing guidelines were changed to reflect the impact of OATP1B1 variants on statin PK profile and toxicity. Similar to CYP enzymes, it appears that a handful of key drug uptake and efflux transporters account for the absorption, distribution, and elimination of the majority of currently prescribed drugs as well as NMEs. Systematic inclusion of transporter studies, including the consideration of polymorphisms in relevant transporters, will result in better candidate NME selection as well as a more robust prediction of intersubject variation in the response to the NME when it is released for general use.

Acknowledgments

R.B.K. is supported by the Wolfe Medical Research Chair in Pharmacogenomics and by grants from the Canadian Institutes of Health Research (MOP-89753) and the Drug Safety and Effectiveness Network (DSEN-PREVENT, FRN-117588), Academic Medical Organization of Southwestern Ontario Alternate Funding Plan Innovation Fund, the Cancer Care Ontario (CCO) Research Chair Award (Tier-1) in Experimental Therapeutics, and the Ontario Institute for Cancer Research (OICR) Translational Research Team grant. C.M. is supported by the Canadian Institutes of Health Research. A.W. is supported by the Clinician-Investigator Program at Schulich School of Medicine & Dentistry at Western and the Canadian Institutes of Health Research/Crohn's Colitis Canada/Canadian Association of Gastroenterology Joint Research Fellowship (inflammatory bowel disease priority area) (2014111BD) as well as by a joint inflammatory bowel disease/clinical pharmacology research grant from Janssen Incorporated.

References

- Lin L, Yee SW, Kim RB, Giacomini KM. SLC transporters as therapeutic targets: emerging opportunities. *Nat Rev Drug Discov*. 2015;14(8):543–560.
- DeGorter MK, Kim RB. Introduction to pharmacogenomics of drug transporters. In: Ishikawa T, Kim RB, Konig J. eds. *Pharmacogenomics of Human Drug Transporters: Clinical Impacts*. Hoboken, NJ: John Wiley & Sons; 2013: 1–11.

- Zhang L, Huang SM, Lesko LJ, et al. Transporter-mediated drug-drug interactions. *Clin Pharmacol Ther*. 2011;89(4):481– 484.
- International Transporter Consortium, Giacomini KM, Huang SM, et al. Membrane transporters in drug development. *Nat Rev Drug Discov*. 2010;9(3):215–236.
- Yee SW, Chen L, Giacomini KM, et al. Pharmacogenomics of membrane transporters: past, present and future. *Pharmacogenomics*. 2010;11(4):475–479.
- DeGorter MK, Xia CQ, Yang JJ, Kim RB. Drug transporters in drug efficacy and toxicity. *Annu Rev Pharmacol Toxicol*. 2012;52:249–273.
- Nakanishi T, Tamai I. Genetic polymorphisms of OATP transporters and their impact on intestinal absorption and hepatic disposition of drugs. *Drug Metab Pharmacokinet*. 2012;27(1):106–121.
- Gong IY, Kim RB. Impact of genetic variation in OATP transporters to drug disposition and response. *Drug Metab Pharmacokinet*. 2013;28(1):4–18.
- Tirona RG. OATP1A2, OAT1, and OAT3. In: Ishikawa T, Kim RB, Konig J. eds. *Pharmacogenomics of Human Drug Transporters: Clinical Impacts.* Hoboken, NJ: John Wiley & Sons; 2013: 125–139.
- Tirona RG, Leake BF, Merino G, Kim RB. Polymorphisms in OATP-C: identification of multiple allelic variants associated with altered transport activity among European- and African-Americans. J Biol Chem. 2001;276(38):35669–35675.
- Pasanen MK, Neuvonen PJ, Niemi M, et al. Global analysis of genetic variation in SLCO1B1. *Pharmacogenomics*. 2008;9(1):19–33.
- Pasanen MK, Backman JT, Neuvonen PJ, Niemi M. Frequencies of single nucleotide polymorphisms and haplotypes of organic anion transporting polypeptide 1B1 SLCO1B1 gene in a Finnish population. *Eur J Clin Pharmacol.* 2006;62(6):409–415.
- Group SC, Link E, Parish S, et al. SLCO1B1 variants and statin-induced myopathy—a genomewide study. *N Engl J Med.* 2008;359(8):789–799.
- Ramsey LB, Johnson SG, Caudle KE, et al. The Clinical Pharmacogenetics Implementation Consortium guideline for SLCO1B1 and simvastatin-induced myopathy: 2014 update. *Clin Pharmacol Ther.* 2014;96(4):423–428.
- Nies AT, Niemi M, Burk O, et al. Genetics is a major determinant of expression of the human hepatic uptake transporter OATP1B1, but not of OATP1B3 and OATP2B1. *Genome Med.* 2013;5(1):1.
- Maeda K, Ieiri I, Yasuda K, et al. Effects of organic anion transporting polypeptide 1B1 haplotype on pharmacokinetics of pravastatin, valsartan, and temocapril. *Clin Pharmacol Ther*. 2006;79(5):427–439.
- Tornio A, Vakkilainen J, Neuvonen M, Backman JT, Neuvonen PJ, Niemi M. SLCO1B1 polymorphism markedly affects the pharmacokinetics of lovastatin acid. *Pharmacogenet Genomics*. 2015;25(8):382–387.
- Rodrigues AC, Perin PM, Purim SG, et al. Pharmacogenetics of OATP transporters reveals that SLCO1B1 c.388A>G variant is determinant of increased atorvastatin response. *Int J Mol Sci.* 2011;12(9):5815–5827.
- Teft WA, Welch S, Lenehan J, et al. OATP1B1 and tumour OATP1B3 modulate exposure, toxicity, and survival after irinotecan-based chemotherapy. *Br J Cancer*. 2015;112(5):857– 865.
- Maeda K. Organic anion transporting polypeptide (OATP)1B1 and OATP1B3 as important regulators of the pharmacokinetics of substrate drugs. *Biol Pharm Bull*. 2015;38(2):155–168.

- Miura M, Satoh S, Inoue K, et al. Influence of SLCO1B1, 1B3, 2B1 and ABCC2 genetic polymorphisms on mycophenolic acid pharmacokinetics in Japanese renal transplant recipients. *Eur J Clin Pharmacol.* 2007;63(12):1161–1169.
- Picard N, Yee SW, Woillard JB, et al. The role of organic aniontransporting polypeptides and their common genetic variants in mycophenolic acid pharmacokinetics. *Clin Pharmacol Ther*. 2010;87(1):100–108.
- Nambu T, Hamada A, Nakashima R, et al. Association of SLCO1B3 polymorphism with intracellular accumulation of imatinib in leukocytes in patients with chronic myeloid leukemia. *Biol Pharm Bull*. 2011;34(1):114–119.
- 24. Yamakawa Y, Hamada A, Nakashima R, et al. Association of genetic polymorphisms in the influx transporter SLCO1B3 and the efflux transporter ABCB1 with imatinib pharmacokinetics in patients with chronic myeloid leukemia. *Ther Drug Monit*. 2011;33(2):244–250.
- van de Steeg E, Stranecky V, Hartmannova H, et al. Complete OATP1B1 and OATP1B3 deficiency causes human Rotor syndrome by interrupting conjugated bilirubin reuptake into the liver. J Clin Invest. 2012;122(2):519–528.
- Ieiri I, Doi Y, Maeda K, et al. Microdosing clinical study: pharmacokinetic, pharmacogenomic (SLCO2B1), and interaction (grapefruit juice) profiles of celiprolol following the oral microdose and therapeutic dose. *J Clin Pharmacol*. 2012;52(7):1078– 1089.
- Imanaga J, Kotegawa T, Imai H, et al. The effects of the SLCO2B1 c.1457C >T polymorphism and apple juice on the pharmacokinetics of fexofenadine and midazolam in humans. *Pharmacogenet Genom.* 2011;21(2):84–93.
- Akamine Y, Miura M, Sunagawa S, Kagaya H, Yasui-Furukori N, Uno T. Influence of drug-transporter polymorphisms on the pharmacokinetics of fexofenadine enantiomers. *Xenobiotica*. 2010;40(11):782–789.
- Mougey EB, Feng H, Castro M, Irvin CG, Lima JJ. Absorption of montelukast is transporter mediated: a common variant of OATP2B1 is associated with reduced plasma concentrations and poor response. *Pharmacogenet Genom.* 2009;19(2):129– 138.
- Tapaninen T, Karonen T, Backman JT, Neuvonen PJ, Niemi M. SLCO2B1 c.935G>A single nucleotide polymorphism has no effect on the pharmacokinetics of montelukast and aliskiren. *Pharmacogenet Genom.* 2013;23(1):19–24.
- Lee W, Glaeser H, Smith LH, et al. Polymorphisms in human organic anion-transporting polypeptide 1A2 (OATP1A2): implications for altered drug disposition and central nervous system drug entry. J Biol Chem. 2005;280(10):9610– 9617.
- 32. Eechoute K, Franke RM, Loos WJ, et al. Environmental and genetic factors affecting transport of imatinib by OATP1A2. *Clin Pharmacol Ther.* 2011;89(6):816–820.
- Yamakawa Y, Hamada A, Shuto T, et al. Pharmacokinetic impact of SLCO1A2 polymorphisms on imatinib disposition in patients with chronic myeloid leukemia. *Clin Pharmacol Ther*. 2011;90(1):157–163.
- Koepsell H, Endou H. The SLC22 drug transporter family. *Pflugers Arch.* 2004;447(5):666–676.
- Koepsell H. The SLC22 family with transporters of organic cations, anions and zwitterions. *Mol Aspects Med.* 2013;34(2-3):413–435.
- Nies A, Koepsell H, Damme K, Schwab M. Organic cation transporters (OCTs, MATEs), in vitro and in vivo evidence for the importance in drug therapy. In: Fromm MF, Kim RB. eds. *Drug Transporters*. Vol 201. Berlin, Heidelberg: Springer; 2011: 105–167.

- Motohashi H, Sakurai Y, Saito H, et al. Gene expression levels and immunolocalization of organic ion transporters in the human kidney. J Am Soc Nephrol. 2002;13(4):866–874.
- de Lima LT, Vivona D, Bueno CT, et al. Reduced ABCG2 and increased SLC22A1 mRNA expression are associated with imatinib response in chronic myeloid leukemia. *Med Oncol.* 2014;31(3):851-014-0851-0855. Epub Jan 2014.
- Giannoudis A, Wang L, Jorgensen AL, et al. The hOCT1 SNPs M420del and M408V alter imatinib uptake and M420del modifies clinical outcome in imatinib-treated chronic myeloid leukemia. *Blood.* 2013;121(4):628–637.
- Nies AT, Koepsell H, Winter S, et al. Expression of organic cation transporters OCT1 (SLC22A1) and OCT3 (SLC22A3) is affected by genetic factors and cholestasis in human liver. *Hepatology (Baltimore, MD)*. 2009;50(4):1227–1240.
- 41. Shitara Y, Sugiyama Y. Pharmacokinetic and pharmacodynamic alterations of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors: drug-drug interactions and interindividual differences in transporter and metabolic enzyme functions. *Pharmacol Ther*. 2006;112(1):71–105.
- 42. Shu Y, Sheardown SA, Brown C, et al. Effect of genetic variation in the organic cation transporter 1 (OCT1) on metformin action. *J Clin Invest*. 2007;117(5):1422–1431.
- Ahlin G, Chen L, Lazorova L, et al. Genotype-dependent effects of inhibitors of the organic cation transporter, OCT1: predictions of metformin interactions. *Pharmacogenom J*. 2011;11(6):400–411.
- 44. Shu Y, Brown C, Castro RA, et al. Effect of genetic variation in the organic cation transporter 1, OCT1, on metformin pharmacokinetics. *Clin Pharmacol Ther.* 2008;83(2):273–280.
- Sirtori CR, Pasik C. Re-evaluation of a biguanide, metformin: mechanism of action and tolerability. *Pharmacol Res.* 1994;30(3):187–228.
- 46. Duong JK, Kumar SS, Kirkpatrick CM, et al. Population pharmacokinetics of metformin in healthy subjects and patients with type 2 diabetes mellitus: simulation of doses according to renal function. *Clin Pharmacokinet*. 2013;52(5):373–384.
- Stocker SL, Riedmaier AE, Schwab M, Giacomini KM. OCT (SLC22A) and OCTN Family. In: Ishikawa T, Kim RB, Konig J. eds. *Pharmacogenomics of Human Drug Transporters: Clinical Impacts*. Hoboken, NJ: John Wiley & Sons; 2013: 171–208.
- Koren-Michowitz M, Buzaglo Z, Ribakovsky E, et al. OCT1 genetic variants are associated with long term outcomes in imatinib treated chronic myeloid leukemia patients. *Eur J Haematol.* 2014;92(4):283–288.
- Shikata E, Yamamoto R, Takane H, et al. Human organic cation transporter (OCT1 and OCT2) gene polymorphisms and therapeutic effects of metformin. *J Hum Genet*. 2007;52(2):117– 122.
- 50. Klen J, Goričar K, Janez A, et al. The role of genetic factors and kidney and liver function in glycemic control in type 2 diabetes patients on long-term metformin and sulphonylurea cotreatment. *BioMed Res Intl.* 2014;2014:7.
- Becker ML, Visser LE, van Schaik RH, Hofman A, Uitterlinden AG, Stricker BH. Genetic variation in the multidrug and toxin extrusion 1 transporter protein influences the glucoselowering effect of metformin in patients with diabetes: a preliminary study. *Diabetes*. 2009;58(3):745–749.
- Umamaheswaran GPR, Arunkumar AS, Das AK, Shewade DG, Adithan C. Genetic analysis of OCT1 gene polymorphisms in an Indian population. *Indian J Hum Genet*. 2011;17:164–168.
- Song IS, Shin HJ, Shin JG. Genetic variants of organic cation transporter 2 (OCT2) significantly reduce metformin uptake in oocytes. *Xenobiotica*. 2008;38(9):1252–1262.

- Song IS, Shin HJ, Shim EJ, et al. Genetic variants of the organic cation transporter 2 influence the disposition of metformin. *Clin Pharmacol Ther*. 2008;84(5):559–562.
- Schwab M, Eichelbaum M, Fromm MF. Genetic polymorphisms of the human MDR1 drug transporter. *Annu Rev Pharmacol Toxicol.* 2003;43:285–307.
- Cascorbi I. P-Glycoprotein: tissue distribution, substrates, and functional consequences of genetic variations. *Handb Exp Pharmacol.* 2011;201:261–283.
- Kroetz DL, Pauli-Magnus C, Hodges LM, et al. Sequence diversity and haplotype structure in the human ABCB1 (MDR1, multidrug resistance transporter) gene. [Erratum appears in *Pharmacogenetics*. 2003;13(11):701.] *Pharmacogenetics*. 2003;13(8):481–494.
- Hoffmeyer S, Burk O, von Richter O, et al. Functional polymorphisms of the human multidrug-resistance gene: multiple sequence variations and correlation of one allele with P-glycoprotein expression and activity in vivo. *Proc Natl Acad Sci USA*. 2000;97(7):3473–3478.
- Fung KL, Gottesman MM. A synonymous polymorphism in a common MDR1 (ABCB1) haplotype shapes protein function. *Biochim Biophys Acta*. 2009;1794(5):860–871.
- Kim RB, Leake BF, Choo EF, et al. Identification of functionally variant MDR1 alleles among European Americans and African Americans. *Clin Pharmacol Ther*. 2001;70(2):189–199.
- Tanabe M, Ieiri I, Nagata N, et al. Expression of P-glycoprotein in human placenta: relation to genetic polymorphism of the multidrug resistance (MDR)-1 gene. J Pharmacol Exp Ther. 2001;297(3):1137–1143.
- Bournissen FG, Moretti ME, Juurlink DN, Koren G, Walker M, Finkelstein Y. Polymorphism of the MDR1/ABCB1 C3435T drug-transporter and resistance to anticonvulsant drugs: A meta-analysis. *Epilepsia*. 2009;50(4):898–903.
- 63. Cheng J-W, Zhang L-J, Hou Y-Q, et al. Association between MDR1 C3435T polymorphism and refractory epilepsy in the Chinese population: a systematic review and meta-analysis. *Epilepsy Behav.* 2014;36:173–179.
- Haerian B, Roslan H, Raymond A, et al. ABCB1 C3435T polymorphism and the risk of resistance to antiepileptic drugs in epilepsy: a systematic review and meta-analysis. *Seizure*. 2010;19(6):339–346.
- Zhou Q, Chen Q-x, Ruan Z-r, Yuan H, Xu H-m, Zeng S. CYP2C9* 3 (1075A>C), ABCB1 and SLCO1B1 genetic polymorphisms and gender are determinants of intersubject variability in pitavastatin pharmacokinetics. *Pharmazie*. 2013;68(3):187–194.
- 66. Zhou Q, Ruan Z-R, Yuan H, Xu D-H, Zeng S. ABCB1 gene polymorphisms, ABCB1 haplotypes and ABCG2 c. 421c>A are determinants of inter-subject variability in rosuvastatin pharmacokinetics. *Pharmazie*. 2013;68(2):129– 134.
- Sałacka A, Bińczak-Kuleta A, Kaczmarczyk M, Hornowska I, Safranow K, Clark JS. Possible association of ABCB1:
 c. 3435T>C polymorphism with high-density-lipoproteincholesterol response to statin treatment—a pilot study. *Bosnian J Basic Med Sci.* 2014;14(3):144.
- Taubert D, Beckerath N, Grimberg G, et al. Impact of Pglycoprotein on clopidogrel absorption. *Clin Pharmacol Ther*. 2006;80(5):486–501.
- Wang X-Q, Shen C-L, Wang B-N, Huang X-H, Li J. Genetic polymorphisms of CYP2C19* 2 and ABCB1 C3435T affect the pharmacokinetic and pharmacodynamic responses to clopidogrel in 401 patients with acute coronary syndrome. *Gene*. 2015;558(2):200–207.

- Cusato J, Allegra S, De Nicolò A, et al. ABCB11 and ABCB1 gene polymorphisms impact on telaprevir pharmacokinetic at one month of therapy. *Biomed Pharmacother*. 2015;69:63–69.
- Coelho AV, Silva SP, de Alencar LC, et al. ABCB1 and ABCC1 variants associated with virological failure of first-line protease inhibitors antiretroviral regimens in Northeast Brazil patients. *J Clin Pharmacol.* 2013;53(12):1286–1293.
- Zhang Y, Li J-L, Fu Q, et al. Associations of ABCB1, NFKB1, CYP3A, and NR112 polymorphisms with cyclosporine trough concentrations in Chinese renal transplant recipients. *Acta Pharmacol Sin*. 2013;34(4):555–560.
- Kim I-W, Yun H-Y, Choi B, et al. ABCB1 C3435T genetic polymorphism on population pharmacokinetics of methotrexate after hematopoietic stem cell transplantation in Korean patients: a prospective analysis. *Clin Ther.* 2012;34(8):1816– 1826.
- 74. Maffioli M, Camós M, Gaya A, et al. Correlation between genetic polymorphisms of the hOCT1 and MDR1 genes and the response to imatinib in patients newly diagnosed with chronic-phase chronic myeloid leukemia. *Leukemia Res.* 2011;35(8):1014–1019.
- Zu B, Li Y, Wang X, He D, Huang Z, Feng W. MDR1 gene polymorphisms and imatinib response in chronic myeloid leukemia: a meta-analysis. *Pharmacogenomics*. 2014;15(5):667– 677.
- Su CLG. Sleisenger and Fordtran's Gastrointestinal and Liver Diseases: Pathophysiology/Diagnosis/Management. Vol 2. 8th ed. Philadelphia, PA: Saunders Elsevier; 2006.
- Deo AK, Prasad B, Balogh L, Lai Y, Unadkat JD. Interindividual variability in hepatic expression of the multidrug resistanceassociated protein 2 (MRP2/ABCC2): quantification by liquid chromatography/tandem mass spectrometry. *Drug Metab Dispos.* 2012;40(5):852–855.
- Meyer zu Schwabedissen HE, Jedlitschky G, Gratz M, et al. Variable expression of MRP2 (ABCC2) in human placenta: influence of gestational age and cellular differentiation. *Drug Metab Dispos*. 2005;33(7):896–904.
- Nies AT. MRP2 (ABCC2) and MRP3 (ABCC3). In: Ishikawa T, Kim RB, Konig J. eds. *Pharmacogenomics of Human Drug Transporters: Clinical Impacts*. Hoboken, NJ: John Wiley & Sons; 2013: 345–364.
- Haenisch S, Zimmermann U, Dazert E, et al. Influence of polymorphisms of ABCB1 and ABCC2 on mRNA and protein expression in normal and cancerous kidney cortex. *Pharmacogenomics J.* 2006;7(1):56–65.
- Sasaki T, Hirota T, Ryokai Y, et al. Systematic screening of human ABCC3 polymorphisms and their effects on MRP3 expression and function. *Drug Metab Pharmacokinet*. 2011;26(4):374– 386.
- Simon N, Marsot A, Villard E, et al. Impact of ABCC2 polymorphisms on high-dose methotrexate pharmacokinetics in patients with lymphoid malignancy. *Pharmacogenomics J*. 2013;13(6):507–513.
- 83. Sharifi MJ, Bahoush G, Zaker F, Ansari S, Rafsanjani KA, Sharafi H. Association of -24CT, 1249GA, and 3972CT ABCC2 gene polymorphisms with methotrexate serum levels and toxic side effects in children with acute lymphoblastic leukemia. *Pediatr Hematol Oncol.* 2014;31(2):169–177.
- Liu Y, Yin Y, Sheng Q, et al. Association of ABCC2 24C>T polymorphism with high-dose methotrexate plasma concentrations and toxicities in childhood acute lymphoblastic leukemia. *PLoS One*. 2014;9(1):e82681.
- Lopez-Lopez E, Ballesteros J, Pinan MA, et al. Polymorphisms in the methotrexate transport pathway: a new tool for

MTX plasma level prediction in pediatric acute lymphoblastic leukemia. *Pharmacogenet Genom.* 2013;23(2):53–61.

- Mao Q, Unadkat JD. Role of the breast cancer resistance protein (BCRP/ABCG2) in drug transport—an update. *AAPS* J. 2015;17(1):65–82.
- Hu M, To KK, Mak VW, Tomlinson B. The ABCG2 transporter and its relations with the pharmacokinetics, drug interaction and lipid-lowering effects of statins. *Expert Opin Drug Metab Toxicol*. 2011;7(1):49–62.
- Chen L, Polli JW. ADME pharmacogenomics in drug development. In: Ishikawa T, Kim RB, Konig J, (eds). *Pharmacogenomics of Human Drug Transporters: Clinical Impacts*. Hoboken, NJ: John Wiley & Sons; 2013: 13–37.
- Honjo Y, Morisaki K, Huff LM, et al. Single-nucleotide polymorphism (SNP) analysis in the ABC half-transporter ABCG2 (MXR/BCRP/ABCP1). *Cancer Biol Ther.* 2002;1(6):696–702.
- Kondo C, Suzuki H, Itoda M, et al. Functional analysis of SNPs variants of BCRP/ABCG2. *Pharm Res.* 2004; 21(10):1895–1903.
- Basseville A, Bates SE, Figg WD, Sparreboom A. BCRP(ABCG2). In: Ishikawa T, Kim RB, Konig J. Eds. *Pharmacogenomics of Human Drug Transporters: Clinical Impacts.* Hoboken, NJ: John Wiley & Sons; 2013: 311–343.
- Furukawa T, Wakabayashi K, Tamura A, et al. Major SNP (Q141K) variant of human ABC transporter ABCG2 undergoes lysosomal and proteasomal degradations. *Pharm Res.* 2009;26(2):469–479.
- Keskitalo JE, Pasanen MK, Neuvonen PJ, Niemi M. Different effects of the ABCG2 c.421C>A SNP on the pharmacokinetics of fluvastatin, pravastatin and simvastatin. *Pharmacogenomics*. 2009;10(10):1617–1624.
- Keskitalo JE, Zolk O, Fromm MF, Kurkinen KJ, Neuvonen PJ, Niemi M. ABCG2 polymorphism markedly affects the pharmacokinetics of atorvastatin and rosuvastatin. *Clin Pharmacol Ther*. 2009;86(2):197–203.
- Bailey KM, Romaine SP, Jackson BM, et al. Hepatic metabolism and transporter gene variants enhance response to rosuvastatin in patients with acute myocardial infarction: the GEOSTAT-1 Study. *Circ Cardiovasc Genet*. 2010;3(3):276–285.
- Tomlinson B, Hu M, Lee VW, et al. ABCG2 polymorphism is associated with the low-density lipoprotein cholesterol response to rosuvastatin. *Clin Pharmacol Ther.* 2010;87(5):558–562.
- Sparreboom A, Gelderblom H, Marsh S, et al. Diflomotecan pharmacokinetics in relation to ABCG2 421C>A genotype. *Clin Pharmacol Ther*. 2004;76(1):38–44.
- Sparreboom A, Loos WJ, Burger H, et al. Effect of ABCG2 genotype on the oral bioavailability of topotecan. *Cancer Biol Ther*. 2005;4(6):650–658.
- Gardner ER, Burger H, van Schaik RH, et al. Association of enzyme and transporter genotypes with the pharmacokinetics of imatinib. *Clin Pharmacol Ther*. 2006;80(2):192–201.
- Koo DH, Ryu MH, Ryoo BY, et al. Association of ABCG2 polymorphism with clinical efficacy of imatinib in patients with gastrointestinal stromal tumor. *Cancer Chemother Pharmacol.* 2015;75(1):173–182.
- 101. Tsuchiya K, Hayashida T, Hamada A, Kato S, Oka S, Gatanaga H. Low raltegravir concentration in cerebrospinal fluid in patients with ABCG2 genetic variants. J AIDS. 2014;66(5):484–486.
- 102. Saison C, Helias V, Ballif BA, et al. Null alleles of ABCG2 encoding the breast cancer resistance protein define the new blood group system Junior. *Nat Genet*. 2012;44(2):174–177.
- Zelinski T, Coghlan G, Liu XQ, Reid ME. ABCG2 null alleles define the Jr(a–) blood group phenotype. *Nat Genet*. 2012;44(2):131–132.

- 104. Badagnani I, Castro RA, Taylor TR, et al. Interaction of methotrexate with organic-anion transporting polypeptide 1A2 and its genetic variants. *J Pharmacol Exp Ther*. 2006;318(2):521–529.
- Pasanen MK, Neuvonen M, Neuvonen PJ, Niemi M. SLCO1B1 polymorphism markedly affects the pharmacokinetics of simvastatin acid. *Pharmacogenet Genom.* 2006;16(12): 873–879.
- 106. Niemi M, Schaeffeler E, Lang T, et al. High plasma pravastatin concentrations are associated with single nucleotide polymorphisms and haplotypes of organic anion transporting polypeptide-C (OATP-C, SLCO1B1). *Pharmacogenetics*. 2004;14(7):429–440.
- 107. Kameyama Y, Yamashita K, Kobayashi K, Hosokawa M, Chiba K. Functional characterization of SLCO1B1 (OATP-C) variants, SLCO1B1*5, SLCO1B1*15 and SLCO1B1*15+C1007G, by using transient expression systems of HeLa and HEK293 cells. *Pharmacogenet Genom.* 2005;15(7): 513–522.
- Pasanen MK, Fredrikson H, Neuvonen PJ, Niemi M. Different effects of SLCO1B1 polymorphism on the pharmacokinetics of atorvastatin and rosuvastatin. *Clin Pharmacol Ther*. 2007;82(6):726–733.
- 109. Niemi M, Backman JT, Kajosaari LI, et al. Polymorphic organic anion transporting polypeptide 1B1 is a major determinant of repaglinide pharmacokinetics. *Clin Pharmacol Ther*. 2005;77(6):468–478.
- Trevino LR, Shimasaki N, Yang W, et al. Germline genetic variation in an organic anion transporter polypeptide associated with methotrexate pharmacokinetics and clinical effects. *J Clin Oncol.* 2009;27(35):5972–5978.
- Oswald S, Konig J, Lutjohann D, et al. Disposition of ezetimibe is influenced by polymorphisms of the hepatic uptake carrier OATP1B1. *Pharmacogenet Genom.* 2008;18(7):559–568.
- 112. Kalliokoski A, Backman JT, Neuvonen PJ, Niemi M. Effects of the SLCO1B1*1B haplotype on the pharmacokinetics and pharmacodynamics of repaglinide and nateglinide. *Pharmaco*genet Genom. 2008;18(11):937–942.
- 113. Weiner M, Peloquin C, Burman W, et al. Effects of tuberculosis, race, and human gene SLCO1B1 polymorphisms on rifampin concentrations. *Antimicrob Agents Chemother*. 2010;54(10):4192–4200.
- 114. Han JY, Lim HS, Shin ES, et al. Influence of the organic aniontransporting polypeptide 1B1 (OATP1B1) polymorphisms on irinotecan-pharmacokinetics and clinical outcome of patients with advanced non-small cell lung cancer. *Lung Cancer*. 2008;59(1):69–75.
- Letschert K, Keppler D, Konig J. Mutations in the SLCO1B3 gene affecting the substrate specificity of the hepatocellular uptake transporter OATP1B3 (OATP8). *Pharmacogenetics*. 2004;14(7):441–452.
- 116. Kiyotani K, Mushiroda T, Kubo M, Zembutsu H, Sugiyama Y, Nakamura Y. Association of genetic polymorphisms in SLCO1B3 and ABCC2 with docetaxel-induced leukopenia. *Cancer Sci.* 2008;99(5):967–972.
- 117. Umamaheswaran G, Praveen RG, Damodaran SE, Das AK, Adithan C. Influence of SLC22A1 rs622342 genetic polymorphism on metformin response in South Indian type 2 diabetes mellitus patients. *Clin Exp Med.* 2015;15(4):511–517.
- 118. Becker ML, Visser LE, van Schaik RH, Hofman A, Uitterlinden AG, Stricker BHC. OCT1 polymorphism is associated with response and survival time in anti-Parkinsonian drug users. *Neurogenetics*. 2011;12(1):79–82.
- 119. Di Paolo A, Polillo M, Capecchi M, et al. The c.480C>G polymorphism of hOCT1 influences imatinib clearance in

patients affected by chronic myeloid leukemia. *Pharmacogenom J.* 2014;14(4):328–335.

- Chen Y, Li S, Brown C, et al. Effect of genetic variation in the organic cation transporter 2 on the renal elimination of metformin. *Pharmacogenet Genom.* 2009;19(7):497–504.
- 121. Kang HJ, Song IS, Shin HJ, et al. Identification and functional characterization of genetic variants of human organic cation transporters in a Korean population. *Drug Metab Dispos*. 2007;35(4):667–675.
- Filipski KK, Loos WJ, Verweij J, Sparreboom A. Interaction of cisplatin with the human organic cation transporter 2. *Clin Cancer Res.* 2008;14(12):3875–3880.
- Lanvers-Kaminsky C, Sprowl JA, Malath I, et al. Human OCT2 variant c.808G>T confers protection effect against cisplatininduced ototoxicity. *Pharmacogenomics*. 2015;16(4):323–332.
- 124. Iwata K, Aizawa K, Kamitsu S, et al. Effects of genetic variants in SLC22A2 organic cation transporter 2 and SLC47A1 multidrug and toxin extrusion 1 transporter on cisplatin-induced adverse events. *Clin Exp Nephrol*. 2012;16(6):843–851.
- 125. Zhang Y, Kong X, Wang R, et al. Genetic association of the P-glycoprotein gene ABCB1 polymorphisms with the risk for steroid-induced osteonecrosis of the femoral head in Chinese population. *Mol Biol Rep.* 2014;41(5):3135–3146.
- 126. Kim SW, Lee JH, Lee SH, Hong HJ, Lee MG, Yook K-H. ABCB1 c. 2677G>T variation is associated with adverse reactions of OROS-methylphenidate in children and adolescents with ADHD. J Clin Psychopharmacol. 2013;33(4):491–498.
- 127. Chen P, Yan Q, Xu H, Lu A, Zhao P. The effects of ABCC2 G1249A polymorphism on the risk of resistance to antiepileptic drugs: a meta-analysis of the literature. *Genet Testing Mol Biomarkers*. 2014;18(2):106–111.
- Sporis D, Bozina N, Basic S, et al. Lack of association between polymorphism in ABCC2 gene and response to antiepileptic drug treatment in Croatian patients with epilepsy. *Colleg Antropol.* 2013;37(1):41–45.
- Haenisch S, Zimmermann U, Dazert E, et al. Influence of polymorphisms of ABCB1 and ABCC2 on mRNA and protein expression in normal and cancerous kidney cortex. *Pharmacogenom J.* 2007;7(1):56–65.
- 130. Haenisch S, May K, Wegner D, Caliebe A, Cascorbi I, Siegmund W. Influence of genetic polymorphisms on intestinal expression and rifampicin-type induction of ABCC2 and on bioavailability of talinolol. *Pharmacogenet Genom*. 2008;18(4):357–365.
- 131. Becker ML, Elens LL, Visser LE, et al. Genetic variation in

the ABCC2 gene is associated with dose decreases or switches to other cholesterol-lowering drugs during simvastatin and atorvastatin therapy. *Pharmacogenom J.* 2013;13(3):251–256.

- 132. Lang T, Hitzl M, Burk O, et al. Genetic polymorphisms in the multidrug resistance-associated protein 3 (ABCC3, MRP3) gene and relationship to its mRNA and protein expression in human liver. *Pharmacogenetics*. 2004;14(3):155–164.
- 133. Venkatasubramanian R, Fukuda T, Niu J, et al. ABCC3 and OCT1 genotypes influence pharmacokinetics of morphine in children. *Pharmacogenomics*. 2014;15(10):1297–1309.
- 134. Cheepala SB, Sukthankar M, Schuetz JD. Role of genetic polymorphisms of MRP4. In: Ishikawa T, Kim RB, Konig J. eds. *Pharmacogenomics of Human Drug Transporters: Clinical Impacts.* Hoboken, NJ: John Wiley & Sons; 2013: 365–385.
- 135. Ban H, Andoh A, Imaeda H, et al. The multidrug-resistance protein 4 polymorphism is a new factor accounting for thiopurine sensitivity in Japanese patients with inflammatory bowel disease. J Gastroenterol. 2010;45(10):1014–1021.
- Low SK, Kiyotani K, Mushiroda T, Daigo Y, Nakamura Y, Zembutsu H. Association study of genetic polymorphism in ABCC4 with cyclophosphamide-induced adverse drug reactions in breast cancer patients. *J Hum Genet*. 2009;54(10):564– 571.
- 137. Zhang W, Yu BN, He YJ, et al. Role of BCRP 421C>A polymorphism on rosuvastatin pharmacokinetics in healthy Chinese males. *Clin Chim Acta*. 2006;373(1-2):99–103.
- Mizuarai S, Aozasa N, Kotani H. Single nucleotide polymorphisms result in impaired membrane localization and reduced ATPase activity in multidrug transporter ABCG2. *Int J Cancer*. 2004;109(2):238–246.
- 139. Chen X, Chen D, Yang S, et al. Impact of ABCG2 polymorphisms on the clinical outcome of TKIs therapy in Chinese advanced non-small-cell lung cancer patients. *Cancer Cell Int.* 2015;15:43-015-0191-0193. eCollection 2015.
- 140. Kim DH, Sriharsha L, Xu W, et al. Clinical relevance of a pharmacogenetic approach using multiple candidate genes to predict response and resistance to imatinib therapy in chronic myeloid leukemia. *Clin Cancer Res.* 2009;15(14):4750–4758.
- 141. Poonkuzhali B, Lamba J, Strom S, et al. Association of breast cancer resistance protein/ABCG2 phenotypes and novel promoter and intron 1 single nucleotide polymorphisms. *Drug Metab Dispos*. 2008;36(4):780–795.
- 142. Ni W, Ji J, Dai Z, et al. Flavopiridol pharmacogenetics: clinical and functional evidence for the role of SLCO1B1/OATP1B1 in flavopiridol disposition. *PLoS ONE*. 2010;5(11):e13792.