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# Analysis of the expression of PIWI-interacting RNAs during cardiac differentiation of human pluripotent stem cells --Manuscript Draft--

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Abstract:	PIWI-interacting RNAs (piRNAs) are a class of non-coding RNAs initially thought to be restricted exclusively to germline cells. In recent years, accumulating evidence has demonstrated that piRNAs are actually expressed in pluripotent, neural, cardiac and even cancer cells. However, controversy remains around the existence and function of somatic piRNAs. Using small RNA-seq samples from H9 pluripotent cells differentiated to mesoderm progenitors and cardiomyocytes we identified the expression of 447 piRNAs, of which 241 were detected in pluripotency, 218 in mesoderm and 171 in cardiac cells. The majority of them originated from the sense strand of protein coding and lncRNAs genes in all stages of differentiation, though no evidences for secondary piRNAs (ping-pong) were found. Genes hosting piRNAs in cardiac samples were related to critical biological processes in the heart, like contraction and cardiac muscle development. Our results indicate that somatic piRNAs might have a role in fine-tuning the expression of genes involved in differentiation of pluripotent cells to cardiomyocytes.íí						
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**RE: New submission** 

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Analysis of the expression of PIWI-interacting RNAs during cardiac differentiation of human pluripotent stem cells

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Dear Dr. Editor of Cell Systems,

Hereby we submit the original manuscript entitled "Analysis of the expression of PIWI-interacting RNAs during cardiac differentiation of human pluripotent stem cells" by Alejandro La Greca and collaborators. The manuscript is not under evaluation in any other journal, and it has not been partially or totally published in any peer review form. A pre-liminary version has been uploaded to the preprint server BiorXiv.org and it is publicly available in the following link http://dx.doi.org/10.1101/639906.

In the present work we describe the identification of piRNA transcripts during differentiation of pluripotent stem cells to cardiomyocytes, using small RNA sequencing samples generated in our laboratory. We analyzed abundance of identified piRNAs and determined differentially expressed transcripts among three stages of differentiation (pluripotent, mesoderm progenitor and cardiomyocytes), suggesting piRNAs are actively regulated during this process. Furthermore, the majority of identified piRNAs originated from protein coding and lncRNA genes in sense orientation. Of note, in cardiomyocytes these genes were related to cardiovascular development and function.

Our evidences indicate that somatic piRNAs participate in fine-tuning expression of key genes involved in the differentiation of pluripotent cells to cardiomyocytes.

Raw sequencing data files used in this work are publicly available for download at GEO (Gene Expression Omnibus) under accession number GSE108021 and custom scripting is available at https://github.com/sgmiriuka/piRNA\_custom\_scripting.

Authors declare there are no competing financial interests in relation to this investigation and commit to update bioRxiv entry if manuscript is accepted. Additionally, authors would like to submit the manuscript to Stem Cell Reports for joint consideration.

Regards,

Santiago Miriuka, MD MSc PhD FACC

# Analysis of the expression of PIWI-interacting RNAs during cardiac differentiation of human pluripotent stem cells

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#### Abstract

PIWI-interacting RNAs (piRNAs) are a class of non-coding RNAs initially thought to be restricted exclusively to germline cells. In recent years, accumulating evidence has demonstrated that piRNAs are actually expressed in pluripotent, neural, cardiac and even cancer cells. However, controversy remains around the existence and function of somatic piRNAs. Using small RNA-seq samples from H9 pluripotent cells differentiated to mesoderm progenitors and cardiomyocytes we identified the expression of 447 piRNAs, of which 241 were detected in pluripotency, 218 in mesoderm and 171 in cardiac cells. The majority of them originated from the sense strand of protein coding and lncRNAs genes in all stages of differentiation, though no evidences for secondary piRNAs (ping-pong) were found. Genes hosting piRNAs in cardiac samples were related to critical biological processes in the heart, like contraction and cardiac muscle development. Our results indicate that somatic piRNAs might have a role in fine-tuning the expression of genes involved in differentiation of pluripotent cells to cardiomyocytes.

*Keywords:* piRNA, cardiac differentiation, pluripotent stem cells, somatic cells, non-coding RNAs

#### 1 Introduction

Differentiation of pluripotent stem cells (PSC) to cardiomyocytes (CM) 2 was first reported shortly after the characterization of embryonic stem cells 3 (ESC) (Kehat et al., 2001). Initially, differentiation was non-specific and 4 spontaneously achieved, but in the last 10 years upgraded protocols have been 5 developed significantly improving efficiency and reproducibility in cardiac dif-6 ferentiation (Evseenko et al., 2010; Burridge et al., 2012; Lian et al., 2013). 7 These protocols are based in sequentially adding factors (morphogens) and/or inhibitors that modulate  $Wnt/\beta$ -catenin signaling pathways in pluripotent 9 cells. PSC-based models undergo epithelial-to-mesenchymal transition to an 10 early mesoderm progenitor cell (MPC) (Evseenko et al., 2010; Loh et al., 11 2016) followed by further committing to cardiac mesoderm and later cardiac 12 progenitor cells (CPC), which may eventually adopt more especialized fea-13 tures. Though this is arguably similar to *in vivo* embryo development, they 14 recapitulate hallmark features of differentiation thus becoming well suited 15 tools for disease modelling, drug screening and potential cell-based thera-16 pies. 17

Like many other developmental processes, changes associated with differ-18 entiation to CM are tightly regulated. Only recently, and mostly due to the 19 advent of next generation sequencing technologies, the scientific community 20 is unveiling the complex regulatory networks governing the shifts in gene ex-21 pression profiles. Non-coding RNAs (ncRNAs) are critical players in these 22 networks, regulating almost all cellular processes including proliferation, dif-23 ferentiation and death (Beermann et al., 2016; Devaux et al., 2015). Although 24 microRNAs (miRNAs) are the most extensively studied in a wide variety of 25 organisms (Ha and Kim, 2014; Li and Gregory, 2008; Espinoza-Lewis and 26 Wang, 2012; Garate et al., 2018), other ncRNAs have been identified such 27 as long non-coding RNAs (lncRNAs), small interfering RNAs (siRNAs), cir-28 cular RNAs (circRNAs) and PIWI-interacting RNAs (piRNAs). Much has 29 been published about these ncRNAs, though piRNAs is one of the least un-30 derstood. Thought to be initially confined almost exclusively to germinal cell 31 lines (Girard et al., 2006), piRNAs gained much attention primarily because 32 of an increasing amount of evidence demonstrating that these ncRNAs are 33 not only expressed in somatic cells but they actively participate in gene reg-34 ulation as well (Ro et al., 2007; Malone et al., 2009; Yan et al., 2011; Ishizu 35 et al., 2012). 36

<sup>37</sup> Expression of piRNAs was first described as negatively regulating trans-

position of repetitive elements thus protecting genome integrity and favouring 38 self-renewal (Girard et al., 2006; Aravin et al., 2007). Reports in numerous 39 organisms showed that they exert their regulatory function through bind-40 ing a specific clade of the Argonaute (AGO) family -namely PIWI proteins-, 41 resulting in an association which resembles the well-known AGO/miRNA 42 complex (Lau et al., 2006; Saito et al., 2006; Brennecke et al., 2007; Ha and 43 Kim, 2014). Unlike miRNAs, piRNAs are primarily biosynthesized as single-44 stranded long precursors which are then clived into the 24-34 nucleotide-long 45 mature forms in a Dicer-independent manner (Girard et al., 2006; Han et al., 46 2017; Phay et al., 2018). They show a bias for uridine (U) redidues in 5' 47 ends together with a 2'O-methyl modification at their 3' ends. Germ line 48 piRNAs were also found to be synthesized through a secondary pathway 49 named the Ping-Pong amplification loop, which increases levels of primary 50 piRNAs using target mRNAs as intermediary molecules for processing new 51 piRNA precursors (Brennecke et al., 2007; Sato and Siomi, 2013). Of note, 52 these mechanisms seem to be highly conserved across species (Lau et al., 53 2006; Siomi et al., 2011). 54

In the last few years, many studies have proposed an active participation 55 of PIWI/piRNAs complexes in diverse and critical pathways such as neural 56 development or body regeneration of lower eukaryotes (Rajasethupathy et al., 57 2012; Ross et al., 2014). Furthermore, recent work demonstrated a positive 58 correlation between altered piRNA expression profiles and clinically relevant 59 pathologies. The involvement of specific piRNAs in regulating mRNAs lev-60 els of genes related to Alzheimer's disease was described in 2017 (Roy et al., 61 2017), while other groups implicated piRNAs in cardiac function and regen-62 eration through modulation of AKT pathway (Vella et al., 2016; Rajan et al., 63 2014). However, great controversy still remains around expression, function 64 and biosynthetic pathways of somatic piRNAs. Particularly, the potential 65 role of piRNAs in differentiation of pluripotent stem cells to cardiomyocytes 66 has not been formally addressed. Using small RNAseq data generated in 67 our laboratory (Garate et al., 2018) we characterized the expression profile 68 of small RNAs consistent with piRNAs in three stages of cell differentiation 69 from pluripotency (day 0) to mesoderm (day 3.5) and then contractile car-70 diocytes (day 21). Results presented here provide evidences supporting the 71 existence of somatic piRNA transcripts and their stage-specific pattern as a 72 mechanism for potentially fine-tuning gene expression during cell differenti-73 ation. 74

#### 75 **Results**

#### <sup>76</sup> Detection and characterization of piRNA

Detection of piRNAs was conducted on small RNAseq samples from three 77 independent experiments consisting of pluripotent stem cells (PSC, day 0). 78 early mesoderm progenitor cells (MPC, day 3.5) and cardiac progenitor cells 79 (CPC, day 21). After aligning reads to human reference genome (hg38), 80 we found that more than half of mapped reads were 20 to 23 nucleotides 81 (nt) long, where the abundant miRNAs were included (Figure 1A, (Garate 82 et al., 2018)). Considering that the average length of piRNAs in mammals 83 ranges between 24 and 34 nt (Girard et al., 2006; Ozata et al., 2018; Phay 84 et al., 2018), mapped reads were filtered by length to accommodate to this 85 restriction. Nearly 50-70% of mapped reads were removed from the samples 86 after this initial processing step (Figure S1A and S1B). Then, employing a 87 similar approach as previously published work (Yan et al., 2011), we filtered 88 out any read that mapped on ncRNAs (DASHR (Leung et al., 2016)) besides 89 piRNAs (Figure S1C) given that previous publications emphasized on the 90 fact that many identified piRNAs were actually fragments of other types of 91 ncRNAs (Tosar et al., 2018). Approximately 5-20% of initial mapped reads 92 remained after this step (Figure S1B). Importantly, all nine aligned samples 93 behaved similarly to both filtering steps (Figure 1B), reflecting consistency 94 among experimental replicates. 95

To verify the elimination of potential misleading contaminants in fully 96 processed alignments, we analyzed the distribution of mapped reads over two 97 well-characterized miRNAs, pluripotency-associated miR-302b and cardiac-98 expressed miR-143 (Garate et al., 2018). As expected, expression of miRgc 302b was evident in unfiltered data of PSC and MPC populations while 100 miR-143 showed appreciable coverage in unfiltered data of CPC (Figure 1C, 101 top panels). No signal was detected for either of the two genes in processed 102 alignments (Figure 1C, bottom panels). However, these samples showed a 103 strong and sharp coverage signal on known piRNA loci (Figure 1D) con-104 firming that the pipeline employed successfully enriched for reads mapping 105 to these known piRNAs. Henceforth, all analyses were performed on fully 106 processed alignments unless explicitly specified otherwise. 107

Sequence analysis of reads mapping to known piRNA loci showed that all samples but MPC bore a bias for 5' uridine residues as it usually occurs in germline cells (Figure 2A). We corroborated our proceedings by employing the pipeline described above on two small RNA-seq samples from human

testis downloaded from the ENCODE project. Indeed, there was a marked 112 preference for uridine at 5' ends in testis samples (Figure S2A and S2B), 113 which suggest that putative piRNAs in our model are subjected to similar 114 mechanisms of 5' end formation as in germline cells. However, the substantial 115 difference in frequecy of 5'-U residues between our samples and testis sam-116 ples could be an indicative of unconserved biosynthetic steps (Ross et al., 117 2014). In addition, no secondary piRNA production was detected in any of 118 the replicates of our samples given that we did not found evidences of the 119 characteristic 10 nt overlap (ping-pong signature) between 5' ends of sense 120 and antisense mapped reads (Figure 2B). 121

#### 122 Expression of piRNAs during cardiac differentiation

To study the expression profile of known piRNAs in differentiating PSC 123 we kept those with an average count among replicates higher than or equal to 124 3. Normalization by library depth showed equivalent distribution of relative 125 quantifications between samples (Figure S3A), enabling confident identifica-126 tion of 447 piRNAs considering the three cell differentiation stages investi-127 gated (Table S1). Despite some differences between replicates, each stage 128 of cell differentiation was categorically defined by a specif piRNA expression 129 profile (Figure 3B) which was also reflected in Principal Component Analy-130 sis results (Figure S3C). These identifying profiles preferentially aggregated 131 PSC and MPC together indicating a greater resemblance between samples 132 of these two cell populations than with CPC. 133

Of the 447 identified piRNAs, 241 were expressed in PSC, 218 in MPC 134 and 171 in CPC (Table S1). Differential expression analysis revealed only 135 30 genes with significant shifts in RNA levels  $(-1 > \log 2FC > 1; -\log 10p-$ 136 value > 1.30 for the comparison between PSC and MPC, while 137 were 137 differentially expressed (DE) between PSC and CPC and 153 between MPC 138 and CPC (Figure 3A). Of the total 447 piRNAs, 204 were found to be DE 139 with respect to CPC, 86 of which were shared by MPC and PSC (Figure 140 S3D and Table S1). These results were consistent with correlation analysis 141 that showed a higher Pearson coefficient for the MPC-PSC pair (R=0.5, 142 p < 2.2e-16) than for CPC-PSC (R=-0.0062, p=0.9) (Figure 3B), suggesting 143 that PSC bear a greater resemblance to MPC than to CPC not only in the 144 identity of expressed piRNAs, but in their abundance as well. Upregulated 145 piRNAs accounted for 14% of total DE piRNAs in CPC (Figure 3C and 146 Table S1), far fewer than the downregulated piRNAs (Figure 3D and Table 147 S1). We validated several piRNA transcripts (piR-1919272, piR-2519215 and 148

piR-97458) by qPCR in an independent set of samples from H9 pluripotent
cells and 14 days after the onset of cardiac differentiation, corroborating our
detection pipeline and subsequent DE analysis (Figure S3F).

Differentially expressed piRNAs ranked among the top expressing piR-152 NAs. This is probably due to the fact that highly expressed genes are in-153 herently less sensitive to inter-replicate noise, hence more likely to return a 154 lower p-value for contrasts. Thus, in order to investigate potential patterns 155 underlying expression data which might have been masked from differen-156 tial expression analysis, we implemented a soft clustering algorithm to data. 157 This approach returned 8 different patterns of piRNA expression (Figure 3E 158 and Table S2), or Expression Clusters (EC), that reflected two dynamically 159 relevant tendencies: downregulation of piRNAs towards cardiac differenti-160 ation (cluster 1 to 4 and 7) and upregulation of piRNAs towards cardiac 161 differentiation (cluster 5, 6 and 8). The former, as was previously observed, 162 encompassed the majority of DE piRNAs. Regardless of the condition (up 163 or downregulated) of piRNAs in CPC, it was clear that a fraction of piRNAs 164 sustained early change (PSC to MPC) while others shifted later in the differ-165 entiation process (MPC to CPC). Interestingly, expression profile of human 166 PIWI genes (HIWI, HILI, HIWI2 and HIWI3) changed between day 0 and 167 14 of differentiation (Figure 4a). While no conclusive results were obtained 168 for HIWI and HIWI3, HILI and HIWI2 were upregulated towards day 14 169 suggesting that there might be a connection between these *PIWI* genes and 170 cardiac piRNAs. Specific markers of pluripotency and cardiomyocytes were 171 measure at these timepoints to corroborate cell identity (Figure 4b). In addi-172 tion, analysis of H9 and H1 published RNA-seq data validated upregulation 173 of *HIWI2* with cardiac differentiation (Figure S4, (Liu et al., 2017)). 174

#### <sup>175</sup> Genome distribution of expressed piRNAs

Identified piRNAs were distributed rather uniformly throughout the nu-176 clear genome (Figure 5A), except in chromosome Y for which no data was 177 available given the XX karyotype of H9 embryonic stem cells used in this 178 work. Moreover, Expression Clusters did not seem to follow any particu-179 lar arrangement in these chromosomes as well (Figure 5A, center of circular 180 plot). Inclusion of the mitochondrial chromosome (chrM) in the analysis 181 revealed that 90 of 447 piRNAs originated from the mitochondrial genome 182 (Figure 5B). This was consistent with previous work in human somatic can-183 cer cell lines reporting the synthesis of piRNAs from mitochondrial genome 184

((Kwon et al., 2014)). In fact, the chrM was the major contributor of expressed piRNAs in our samples and was mostly dominated by three EC: a)
cluster 3, with piRNAs highly expressed in PSC; b) cluster 8, with piRNAs
highly expressed in CPC; c) cluster 5, with piRNAs highly expressed in both
PSC and CPC.

We corroborated this finding by evaluating the distribution of mapped 190 reads over chrM, and thus eliminated the possibility of errors during counting 191 of reads per transcript (Figure S5A). Despite our pipeline for identification 192 of expressed piRNAs filtered out all reads that mapped to ncRNAs -other 193 than piRNAs- using DASHR database, 90% of mitochondrial piRNAs (81 out 194 of 90) mapped directly to tRNA and rRNA annotations (GENCODE v29) 195 (Figure S5B). DASHR database showed only one annotation in chrM (LSU-196 rRNA) that corresponded to the large ribosomal subunit RNR2 (Figure S5C). 197 This was not the case in the nuclear genome where no piRNAs were found 198 to map on rRNAs and tRNAs annotated in GENCODE database (Figure 199 S5D). Nonetheless, piRNAs identified in length-filtered data (initial step of 200 filtering, Figure 1B) did not map to nuclear rRNA or tRNA annotations 201 from GENCODE to begin with (Figure S5D), suggesting that this step was 202 sufficient enough to remove reads mapping on them. 203

Regardless of the chromosome distribution, identified piRNAs localized preferentially on gene annotations (Figure 5C). PSC samples showed that 88.5% of piRNAs were generated from gene features, while the percentage was higher in MPC and CPC samples, with 96.2 and 95.5% respectively.

#### <sup>208</sup> Protein coding and lncRNA genes hosting piRNAs

Further analysis on genomic distribution of identified piRNAs revealed 209 that nearly 65% of those intersected to gene features originated from coding 210 (53%) and long non-coding (12%) annotations (Figure S5D). To test whether 211 these events were random, we shuffled our samples 1500 times (bootstraping) 212 and analyzed intersection to these features in sense and antisense orientation. 213 Once data was collected, we calculated enrichment on genes as "sample piR-214 NAs" over "shuffled piRNAs" and determined that sense-oriented piRNAs 215 occurred non-randomly on protein coding and long non-coding (lnc) genes 216 (Figure 6A). On the contrary, intersection in antisense had poor fold enrich-217 ment values suggesting piRNAs were preferentially located elsewhere. We 218 observed similar results for piRNAs identified in all three cell differentiation 219 stages studied in this work, as well as in two samples (isogenic replicates) 220

downloaded from ENCODE project corresponding to H1-derived neural progenitor cells (NPC). Both neural samples were handled following the same steps and criteria described before (Figure S6A).

Taking into consideration that reads originated from protein coding and 224 lncRNA features might have been the result of ordinary transcript degra-225 dation, we investigated the distribution of reads mapped on such piRNA-226 hosting genes. Results showed that the percentage of bases covered by sense-227 oriented reads in these genes was low, with a median value of 0.56% in PSC, 228 1.50% in MPC, 2.08% in CPC and 3.63% in NPC (Figure S6B). Moreover, 220 coverage was localized to a set of specific piRNAs instead of all piRbase anno-230 tations described in any single gene (Figure S6C), proving to be inconsistent 231 with random degradation-produced reads. Coverage by antisense-oriented 232 reads was closer to none (Figure S6D) and significantly lower than sense-233 oriented coverage in all cell population except in MPC (Figure 6B), possibly 234 due to a higher level of dispersion in values of these samples. 235

The wide majority of piRNA-hosting protein coding and lncRNA genes 236 harboured a single piRNA transcript with a tendency to augment the number 237 of piRNAs per gene throughout the differentiation process (Figure 6C, pie 238 charts). Like MPC and CPC, NPC exhibited a wider spectrum of piRNAs 239 per gene than undifferentiated pluripotent cells (PSC). A more detailed ex-240 ploration into CPC results revealed that MALAT1 (lncRNA gene) and TTN 241 (protein coding gene) contained the highest number of piRNAs -12 and 6 242 respectively-, followed by PLN, RPPH1, ACTC1 and AL355075.4 with 4 243 (Figure 6C, bottom panel). Using RNA-seq data from H9 cells differentiated 244 to CM (Liu et al., 2017) we analyzed the expression profile of these piRNA-245 containing genes. Transcript abundance of MALAT1, TTN and RPPH1 246 increased from day 0 (PSC) to day 2/4 (MPC) and then dropped between 247 day 4 and day 30 (CPC), which were inversely correlated with the expres-248 sion dynamics of piRNAs from EC 8 (Figure 6D). PLN and ACTC1 RNA 249 levels increased from day 4 to day 30 practically impervious to piRNA pro-250 duction, though lack of data between day 4 and 30 hindered our analysis 251 for these genes (data not shown). With respect to AL355075.4 gene, we 252 found no count data available in RNA-seq samples. However, this gene over-253 laps *RPPH1* in sense orientation and it partially overlaps protein coding 254 gene PARP2 in antisense orientation, meaning that piRNAs originated from 255 it could be potentially involved in regulating both genes. In fact, PARP2 256 expression dynamic showed a steady descent in transcript levels from day 257 0 to 30, which is also consistent with the fact that the piRNAs originated 258

from AL355075.4 were also detected in MPC (Figure S6E). Similar results were found when we studied two genes with high piRNA content (>3) in MPC (Figure 6E) - APLNR and RMRP- in which almost all piRNAs belong to EC 2. Taken together, these evidences suggest that piRNAs originated from these genes may be implicated in their downregulation or possibly in a moderate fine-tuning as in the case of PLN and ACTC1.

#### <sup>265</sup> Functional analysis on piRNA-hosting genes in differentiated cells

The expression profile of piRNAs proved to be sufficient to clearly dis-266 criminate CPC samples not only from PSC and MPC populations, but from 267 neural progenitors (NPC) as well. The comparison between CPC and NPC 268 samples revealed that 52 piRNAs were expressed in both types of differ-269 entiated cells, but more importantly the majority were not (Figure 7A). 270 Unshared piRNAs constitute a unique repertoire for each cell population 271 which could possibly reflect upon diverse functional processes. To evaluate 272 this notion, we extracted all protein coding genes which were intersected 273 by at least one piRNA and determined their involvement in any biologi-274 cal process (BP). In search for overrepresented terms (BPs with more genes 275 involved than expected), we found that CPC and NPC showed markedly dif-276 ferent terms. The BPs associated to CPC were intimately related to heart 277 development and muscle differentiation and contraction (Figure 7B), while 278 overrepresented BPs in NPC showed a clear inclination towards neurogenesis 270 regulation and neural proliferation and development (Figure 7C). The group 280 of genes intersected by piRNAs shared by both CPC and NPC (52 piRNAs 281 in venn diagram) did not participate in any of the statistically significant 282 overrepresented BPs, meaning that enriched categories for each population 283 are mostly based in their unique collection of piRNAs. 284

#### 285 Discussion

Since the first mechanistic evidences of piRNA biogenesis in ovarian fol-286 licle cells of *D. melanogaster* were published (Malone et al., 2009), much 287 information has emerged on the somatic expression and function of this type 288 of regulatory small RNAs. Several publications demonstrated that piRNAs 289 (or piRNA-like RNAs) originate from discrete genomic regions of somatic 290 cells in a wide diversity of species and tissues (Cichocki et al., 2010; Yan 291 et al., 2011; Vella et al., 2016; Ng et al., 2016; Roy et al., 2017). In agree-292 ment with this line of evidence, we report the expression of 447 small RNAs 293

consistent with piRNAs among three stages of differentiation of pluripotent
stem cells to cardiomyocytes using a database-driven approach.

The pipeline leading to the identification and quantification of piRNAs 296 involved two filtering steps that were implemented to avoid innacurate inter-297 pretation of results. Firstly, aligned reads shorter than 24nt and longer than 298 34nt were discarded from further analyses. However, our results showed a 299 higher-than-basal frequency of 36-37nt-long reads, which prompts the issue 300 if these reads should have been kept for further investigation as potential 301 longer piRNAs or perhaps remnants of piRNA precursors. Length restric-302 tion answers to one of the hallmark attributes of mature piRNA transcripts, 303 albeit the range seems to vary across species. In fact, in C. elegans 21nt-long 304 piRNAs are produced from transcript precursors of 25-27nt in length (Ruby 305 et al., 2006) whose processing machinery is partially unknown. The reason 306 for the range diversity has been convincingly connected to the activity of 307 the proteins involved in their biosynthetic pathway (biogenesis). It is possi-308 ble that they also contribute to explaining the differences between germline 309 and somatic piRNAs considering that diverse sets of enzimes have been re-310 ported to be engaged in piRNA synthesis in these cell lineages (Ruby et al., 311 2006; Zamore, 2010). Moreover, the differential expression profile of HIWI 312 genes observed in our data - and in H9 and H1 external RNA-seq data- points 313 to cell-type specific functions for these proteins and consequently for their 314 piRNA partners. 315

In a second step, length-filtered reads mapping to small ncRNAs other 316 than piRNAs were removed from samples. The fact that remaining reads in 317 PSC and CPC samples showed a moderate 5' U bias, whereas MPC sam-318 ples did not could probably be related to the transitional nature of this cell 319 population. However, none of the samples exhibited the characteristic 10nt-320 overlap signature of secondary piRNAs, which is a generally accepted feature 321 in germline cells of most animals where piRNAs are synthesized through both 322 primary and secondary (ping-pong loop) pathways (Yan et al., 2011; Ross 323 et al., 2014). Despite synthesis in somatic cells has been proposed to produce 324 only primary piRNAs, many of the mechanisms underlying piRNA biogenesis 325 -specially in non-gonadal tisues- are not yet fully understood. Conceivably, 326 the filtering steps eliminated potential piRNAs from the three stages, though 327 it has been argued that a considerable amount of annotated piRNAs are ac-328 tually ncRNA fragments derived from rRNAs, tRNAs and even miRNAs 329 (Tosar et al., 2018). On the matter, an insightful disscussion by Tosar and 330 collaborators (Tosar et al., 2018), advocating for gonadal piRNAs, suggested 331

that somatic piRNAs mapping to ncRNA fragments are not unquestionably 332 wrong, still further biochemical evidence is needed to include them as such. 333 An important aspect of this work lies on the identification of a piRNA 334 expression profile associated to each of the cell populations under study. 335 These expression profiles parallel the embryological connection between the 336 stages, where PSC is more closely related to MPC than to CPC (Evseenko 337 et al., 2010; Sha et al., 2019). Upon this premise, the piRNAs identified 338 as early-changing could potentially be involved in maintaining pluripotency 339 or in the commitment of pluripotent cells to mesoderm progenitors, which 340 might eventually differentiate to multiple lineages. Late-changing piRNAs, 341 on the contrary, would influence further commitment of mesoderm cells to 342 cardiac progenitors. The fact that six times more piRNAs were downregu-343 lated rather than upregulated during differentiation to CPC suggests that 344 piRNA pathways become less relevant in differentiated cells. It is possible 345 that mechanisms evolutionarily linked to the regulation of transposable ele-346 ments are not as critically conserved in differentiated cells as they do in cells 347 with high proliferation rates or reproductive functions, such as pluripotent 348 cells and germ line cells. For example, it has been proposed that cancer 349 cells might promote piRNA biosynthetic pathways as a mechanism to reduce 350 genome instability caused by increased mitotic and transcriptional activities 351 (Liu et al., 2018). 352

The genomic localization of piRNAs included in anyone particular EC was 353 not the same. In fact, piRNA expression appeared to be uniformly scattered 354 across the genome except for the mitochondrial chromosome. The majority of 355 piRNAs identified in the mitochondrial genome mapped to rRNAs or tRNAs 356 and though we have not definitively proved they are truly piRNAs, previous 357 work established a link between tRNA-/rRNA-derived piRNAs, HIWI2 ex-358 pression and regulation of metabolic processes in somatic cells (Keam et al., 359 2014). Analogously, the increased levels of piRNAs from mitochondrial tR-360 NAs/rRNAs and the significant upregulation of *HIWI2* (day 14 v. day 0, 361 and external H9 RNA-seq data) in CPC could seemingly be connected to the 362 large-scale modifications in CM metabolism (Tohyama et al., 2013). 363

Even though identified piRNAs were dispersed throughout the genome, it was clear that the vast majority of them originated from gene loci. However, it is not yet clear the reason why these piRNAs are generated from the sense strand of their hosting genes. One possibility relies on the capacity of PIWI/piRNA complexes to direct recruitment of DNA and histone methyltransferases, modifying accesibility of transcriptional machinery to chromatin

(Rajasethupathy et al., 2012; Pezic et al., 2014). Available data of DNA or 370 histone methylation status in the three stages of cardiac differentiation is 371 scarse or dissimilar, so preliminary correlation analysis were not conclusive 372 at this point (data not shown). Nevertheless, further experiments on pro-373 moter methylation and H3K9me3 mark deposition ought to be performed to 374 pursuit this possibility. Also, considering that antisense transcripts have been 375 described to positively regulate stability of sense RNA (Zong et al., 2016), it 376 is possible that sense-originated piRNAs regulate antisense transcript levels 377 in a miRNA-like mechanism. For instance, TALAM1 - an antisense transcript 378 at the MALAT1 gene locus- promotes stability and maturation of MALAT1 379 RNA by facilitating enzymatic cleavage of its 3' end (Zong et al., 2016), thus 380 a potential piRNA-mediated downregulation of TALAM1 would redound to 381 diminished MALAT1 levels. 382

In sum, the evidences presented here contribute to understanding the dynamic expression of piRNAs during differentiation of pluripotent stem cells to cardiomyocytes and further explore their potential function as posttranscriptional modulators in somatic cells. Together with miRNAs, piRNAs seem to participate in the fine-tuning of transcript levels, adding yet another layer to the complex and intrincated networks governing gene expression.

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#### 392 Author Contributions

A.L.G. and S.G.M. design experiments and wrote manuscript. MAS, M.C.H.C., N.P., S.C., C.C., A.M.M., N.L.S.V. and C.A. performed global bioinformatic analysis. X.G., A.W., L.M., G.S., C.L. and S.G.M. discussed and revised analyses and manuscript. G.S. and S.G.M. provided fundings for this paper.

- 398 Conflict of interest statement
- None declared.

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#### 403 Data Availability

Small RNA sequencing data can be found in GEO under accession numberGSE108021.

#### 406 Figure Legends

Figure 1. Detection of piRNAs in small RNAseq samples. A) 407 Length frequency of unfiltered mapped reads in three samples from pluripo-408 tent cells (PSC), three from mesoderm progenitor cells (MPC) and three 409 from cardiac cells (CPC). Grey areas denote the size range for both microR-410 NAs (miRNA) and putative piwi-interacting RNAs (piRNA). The color key 411 used is indicated in top right corner of the plot. B) Number of mapped reads 412 for all nine samples before processing (unfiltered, grey box), after filtering 413 by read length (23 < RL < 35, coral box) and removing non coding RNA 414 other than piRNAs (-ncRNA, green box). C) Distribution of mapped reads 415 as a function of density on a fragment of chromosome 4 (chr4:100,000,000-416 120,000,000; -) which includes pluripotency miR-302b and a fragment of 417 chromosome 5 (chr5:149,300,000-149,600,000, +) including cardiac miR-143 418 for unprocessed alignments (top panels, unfiltered data) and fully processed 419 samples (bottom panels, length+ncRNA filtered). Alignment files from each 420 experimental replicate were merged into one. Color key for the density curves 421 is shown in the graph. D) Analysis of coverage on all piRNA loci available 422 in piRbase for fully processed normalized (counts per million) samples. 423

Figure 2. Characterization of reads mapped to known piRNAs. A) Frequency of bases per position in reads mapped to known piRNAs after fully processing alignments from PSC, MPC and CPC samples (merged replicates). Position 1 is marked by a vertical grey bar. B) Frequency profile showing overlap between reads mapped to known piRNA loci in sense orientation and complementary reads using an independent approach (ssviz package).

Figure 3. piRNA expression profile in differentiating pluripotent stem cells. A) Differential expression analysis performed on raw

counts with DESeq2 package. Significantly different expression values (-1 433  $>= \log 2FC >= 1$ ;  $-\log 10p$ -value > 1.30) are represented as orange dots in 434 the volcano plots for the three possible comparisons: MPC vs. PSC (left), 435 CPC vs. PSC (center) and CPC vs. MPC (right). B) Scatter plot of nor-436 malized expression data showing Pearson correlation analysis on MPC (green 437 dots) and CPC (red dots) versus PSC. Marginal density plots to the right 438 denote areas of highly abundant data and black arrows mark the positions of 439 the most DE piRNA genes. C) Heatmap shows normalized counts of upreg-440 ulated piRNAs genes in CPC considering the three cell populations (PSC, 441 MPC, CPC). Dendrograms resulted from running hard unsupervised cluster-442 ing algorithms on piRNA genes (left) and samples (top). D) Heatmap as in 443 c showing downregulated piRNA genes in CPC. E) Implementation of soft 444 clustering algorithms (R package MFuzz) produces eight distinct patterns 445 (clusters 1 to 8) of piRNA expression. 446

Figure 4. Expression of human PIWI genes in cardiac differ-447 entiation. A) Transcript levels of *HILI* (PIWIL2) and *HIWI2* (PIWIL4) 448 were measure by qPCR in H9 pluripotent cells (D0) and H9-derived cardiac 449 progenitors (D14). B) Stage-specific markers were evaluated by qPCR in 450 samples from a. Pluripotency genes are shown in the top row and genes 451 for cardiac lineage in the bottom. All results were expressed as mean  $\pm$  se of 452 three independent experiments after normalization by the geometric mean 453 of  $RPL\gamma$  and HPRT1 housekeeping genes. Statistical significance was evalu-454 ated by Student's T test (day14 v. day0) and results are indicated as p-values 455 on the bar plots. 456

Figure 5. Distribution of piRNAs and Expression Clusters in 457 the genome. A) Normalized expression of piRNAs from averaged samples 458 of PSC (outer track), MPC (middle track) and CPC (inner track) displayed 459 lengthwise in a circular representation of human autosomal chromosomes 460 (1 to 22) and chromosome X. Level of expression is depicted as heatmaps 461 inside the tracks and follow a red-green scale for high expression and low 462 expression respectively. The center of the plot shows links between piRNAs 463 that belong to the same Expression Cluster (EC); color key for the links 464 appears at the bottom left corner of the plot. B) Bars represent number 465 of piRNAs expressed in all samples per chromosome. EC membership of 466 piRNAs is indicated with color key as in a (top left corner of barplot). C) 467 Percentage of piRNAs in PSC, MPC and CPC samples that intersected or 468

<sup>469</sup> not with genes from GENCODE database (v29).

Figure 6. Proteing coding and lncRNA-originated piRNAs. A) 470 Fold enrichment was calculated as number of piRNAs intersected to protein 471 coding and lncRNA genes in sense (purple) and antisense (green) orientation 472 over 1500 different random distributions (bootstraping). Red dashed line 473 marks the point of no enrichment  $(\log 2 1=0)$ . NPC: human neural progenitor 474 cells. B) Distribution of percent coverage values on genes in sense (purple) 475 and antisense orientation (green). P-value (p) for statistical analysis is shown 476 in the plots (ANOVA). ns: not significant. C) Pie charts show the proportion 477 of genes containing piRNAs. The number of sense piRNAs per gene in each 478 category is indicated in the charts. Plot at the bottom provides detail on 479 pie chart for CPC labeling genes with two or more piRNAs. Mitochondrial 480 genes are not included in the analysis. D) Normalized counts (FPKM) for 481 three genes from c during differentiation of H9 cells (day 0, day 2, day 4 and 482 day 30) to CM. Data was extracted from previously published total RNA-seq 483 experiments. E) Normalized counts (FPKM) for two genes containg piRNAs 484 in MPC. 485

Figure 7. Functional exploration of piRNAs in differentiated cells. A) Expression profiles of piRNAs (log2 normalized counts) in CPC (cpc1, cpc2 and cpc3) and NPC (neural1 and neural2). Color key for sample clustering is displayed at the top right corner of the heatmap. B) Overrepresented biological processes (BPs) on proteing coding genes harbouring piRNAs from CPC samples. C) Overrepresented BPs on proteing coding genes harbouring piRNAs from NPC samples.

#### 493 Experimental Procedures

#### 494 Small RNAseq data

Data samples used in this work (PSC: H9 human embryonic stem cells, 495 MPC: early mesoderm progenitor and CPC:cardiomyocytes) were generated 496 in our laboratory following previously described protocols and are available 497 under accession code GSE108021. Briefly, H9 cells (H9-hTnnTZ-pGZ-D2 498 obtained from WiCell) were routinely maintained in co-culture with irra-490 diated primary mouse embryonic fibroblasts. Mesoderm induction (MPC 500 population) was performed by initially seeding cells with mTeSR (StemCells 501 Technologies) on plates coated with Geltrex (Thermo Fisher Scientific) and 502

then switching to (Evseenko et al., 2010) StemPro-34 SFM (Thermo Fisher 503 Scientific) supplemented with Activin A only at the first day, BMP4, VEGF 504 and bFGF (Thermo Fisher Scientific) for 3.5 days. At this point, meso-505 derm progenitors were isolated by FACS with anti-CD326 and anti-CD56 506 (Biolegend). CPC population was obtained by formation of embryoid bod-507 ies with H9 cells using BMP4, bFGF and Activin A in StemPro-34 followed 508 by addition of VEGF and Wnt inhibitor, IWR-1. Libraries for small RNA 509 sequencing were prepared with 200 ng of RNA using NEBNext Small RNA 510 Library Prep Set with modified adaptors and primers compatible for Illu-511 mina (New England Biolabs). Single end sequencing was carried out at the 512 TCGB Resources (UCLA Path and Lab Med) using an Illumina HiSeq 2500. 513 Culture conditions and sequencing of small RNAs for these samples are more 514 extensively described in (Garate et al., 2018). 515

#### 516 External data

Human testis small RNA-seq samples from two men of 54 and 37 years old (GSE88414 and GSE88124, respectively) and H1-derived neural progenitor cells (GSM1296459 and GSM1296460) were downloaded from ENCODE (encodeproject.org). RNA-seq data (counts per transcript) from H1 and H9 cardiac differentiation protocols can be found under GEO accession GSE85331.

#### <sup>522</sup> Data processing and analyses

Adapters were removed from raw sequencing reads with cutadapt (v1.9.1) 523 keeping reads with a minimum of 20 and up to 50 nt in length. Quality 524 checked (FastQC) processed reads were mapped to human reference genome 525 (GRCh38/hg38) using STAR aligner (v2.5.3a (Dobin et al., 2013)) under 526 mostly default parameters. Mapped reads in output SAM/BAM files were 527 filtered by read length (23 < RL < 35) with samtools and custom awk script-528 ing. Resulting reads were intersected (bedtools v2.27.1 (Quinlan and Hall, 529 2010)) to ncRNAs in a strand specific manner (DASHR (Leung et al., 2016)) 530 to remove potential misleading alignments. Raw counts on piRNAs were 531 determined with htseq-count matching mapped reads to piRNA coordinates 532 downloaded from piRBase (Wang et al., 2018). Counts were then fed into 533 DESeq2 for differential expression analysis (p < 0.05 and fdr< 0.1). Soft clus-534 tering methods were implemented with R package Mfuzz (v2.42.0 (Futschik 535 and Carlisle, 2005) using parameter m=1.15. In parallel to our customized 536 pipeline, tools for ping-pong signature detection like ssviz R package and 537 PingPongPro (Uhrig and Klein, 2018) were run following recommendations 538

from authors. Graphics and statistical analyses were performed in R software and deepTools (Ramírez et al., 2016). Further details on custom code
is available at https://github.com/sgmiriuka/piRNA\_custom\_scripting.

#### 542 Reverse Transcription of piRNA

To obtain cDNA from piRNA transcripts we adapted a previously de-543 scribed methodology employed for miRNA detection and amplification (Chen 544 et al., 2005). Briefly, stem-loop retrotranscription (RT) primers were gener-545 ated using 6-8nt from the 3' end of every piRNA of interest. Each RT reaction 546 was performed with a maximum of 10 different stem-loop primers including 547 one for RNU6B and hsa-miR-302b as controls. SuperScriptIII retrotranscrip-548 tase (Thermo Fisher Scientific) was used for RT reactions following guidelines 549 from manufacturer. Detection by qPCR involved forward primers matching 550 the sequence of target piRNAs and a reverse universal primer complementary 551 to the stem-loop RT primer. 552

#### 553 Real time PCR

Total RNA was prepared with TRI-Reagent (Sigma Aldrich) following 554 manufacturer's instructions and then reverse transcribed using MMLV re-555 verse transcriptase (Promega) and random primers for detection of polyade-556 nilated transcripts. Quantitative real time PCR (qPCR) was performed in a 557 StepOne Real Time PCR system (Applied Biosystems). Expression was nor-558 malized to the geometrical mean of HPRT1 and RPL7 housekeeping genes 559 and log2 transformed. Statistical significance for qPCR results was analyzed 560 by t test (day14 v. day0). Primers sequences are available on request. 561

Table S2. Related to Figure 3 and 5. piRNAs included in Expression Clusters.

#### 564 References

Aravin, A.A., Sachidanandam, R., Girard, A., Fejes-Toth, K., Hannon, G.J.,
 2007. Developmentally regulated pirna clusters implicate mili in transpo son control. Science 316, 744–7. doi:10.1126/science.1142612.

Beermann, J., Piccoli, M.T., Viereck, J., Thum, T., 2016. Non-coding RNAs
 in Development and Disease: Background, Mechanisms, and Therapeutic
 Approaches. Physiological Reviews .

Brennecke, J., Aravin, A.A., Stark, A., Dus, M., Kellis, M., Sachidanandam, R., Hannon, G.J., 2007. Discrete small rna-generating loci as master regulators of transposon activity in drosophila. Cell 128, 1089–103.
doi:10.1016/j.cell.2007.01.043.

<sup>575</sup> Burridge, P.W., Keller, G., Gold, J.D., Wu, J.C., 2012. Production of De
<sup>576</sup> Novo Cardiomyocytes: Human Pluripotent Stem Cell Differentiation and
<sup>577</sup> Direct Reprogramming. Cell Stem Cell .

<sup>578</sup> Chen, C., Ridzon, D.A., Broomer, A.J., Zhou, Z., Lee, D.H., Nguyen, J.T.,
<sup>579</sup> Barbisin, M., Xu, N.L., Mahuvakar, V.R., Andersen, M.R., Lao, K.Q.,
<sup>580</sup> Livak, K.J., Guegler, K.J., 2005. Real-time quantification of micrornas by
<sup>581</sup> stem-loop rt-pcr. Nucleic Acids Res 33, e179. doi:10.1093/nar/gni178.

Cichocki, F., Lenvik, T., Sharma, N., Yun, G., Anderson, S.K., Miller,
J.S., 2010. Cutting edge: Kir antisense transcripts are processed into
a 28-base piwi-like rna in human nk cells. J Immunol 185, 2009–12.
doi:10.4049/jimmunol.1000855.

Devaux, Y., Zangrando, J., Schroen, B., Creemers, E.E., Pedrazzini, T.,
Chang, C.P., Dorn, 2nd, G.W., Thum, T., Heymans, S., Cardiolinc network, 2015. Long noncoding rnas in cardiac development and ageing. Nat
Rev Cardiol 12, 415–25. doi:10.1038/nrcardio.2015.55.

Davis, C.A., Schlesinger, F., Drenkow, J., Zaleski. Dobin. Α., 590 Jha, S., Batut, P., Chaisson, М., Gingeras, T.R., 2013.С., 591 Star: ultrafast universal rna-seq aligner. Bioinformatics 29, 15–21. 592 doi:10.1093/bioinformatics/bts635. 593

Espinoza-Lewis, R.A., Wang, D.Z., 2012. MicroRNAs in heart development.
 Current topics in developmental biology .

Evseenko, D., Zhu, Y., Schenke-Layland, K., Kuo, J., Latour, B., Ge, S.,
Scholes, J., Dravid, G., Li, X., Maclellan, W.R., Crooks, G.M., 2010.
Mapping the first stages of mesoderm commitment during differentiation
of human embryonic stem cells. Proceedings of the National Academy of
Sciences of the United States of America .

Futschik, M.E., Carlisle, B., 2005. Noise-robust soft clustering of gene expression time-course data. J Bioinform Comput Biol 3, 965–88.

- Garate, X., La Greca, A., Neiman, G., Blüguermann, C., Santín Velazque,
  N.L., Moro, L.N., Luzzani, C., Scassa, M.E., Sevlever, G.E., Romorini,
  L., Miriuka, S.G., 2018. Identification of the mirnaome of early mesoderm
  progenitor cells and cardiomyocytes derived from human pluripotent stem
  cells. Sci Rep 8, 8072. doi:10.1038/s41598-018-26156-3.
- Girard, A., Sachidanandam, R., Hannon, G.J., Carmell, M.A., 2006. A
  germline-specific class of small rnas binds mammalian piwi proteins. Nature 442, 199–202. doi:10.1038/nature04917.
- Ha, M., Kim, V.N., 2014. Regulation of microRNA biogenesis. Nature
   reviews. Molecular cell biology .
- Han, Y.N., Li, Y., Xia, S.Q., Zhang, Y.Y., Zheng, J.H., Li, W., 2017. Piwi
  proteins and piwi-interacting rna: Emerging roles in cancer. Cell Physiol
  Biochem 44, 1–20. doi:10.1159/000484541.
- Ishizu, H., Siomi, H., Siomi, M.C., 2012. Biology of piwi-interacting rnas:
  new insights into biogenesis and function inside and outside of germlines.
  Genes Dev 26, 2361–73. doi:10.1101/gad.203786.112.
- Keam, S.P., Young, P.E., McCorkindale, A.L., Dang, T.H.Y., Clancy, J.L.,
  Humphreys, D.T., Preiss, T., Hutvagner, G., Martin, D.I.K., Cropley,
  J.E., Suter, C.M., 2014. The human piwi protein hiwi2 associates with
  trna-derived pirnas in somatic cells. Nucleic Acids Res 42, 8984–95.
  doi:10.1093/nar/gku620.
- Kehat, I., Kenyagin-Karsenti, D., Snir, M., Segev, H., Amit, M., Gepstein,
  A., Livne, E., Binah, O., Itskovitz-Eldor, J., Gepstein, L., 2001. Human
  embryonic stem cells can differentiate into myocytes with structural and
  functional properties of cardiomyocytes. The Journal of clinical investigation .
- Kwon, C., Tak, H., Rho, M., Chang, H.R., Kim, Y.H., Kim, K.T., Balch, C.,
  Lee, E.K., Nam, S., 2014. Detection of piwi and pirnas in the mitochondria
  of mammalian cancer cells. Biochem Biophys Res Commun 446, 218–23.
  doi:10.1016/j.bbrc.2014.02.112.
- Lau, N.C., Seto, A.G., Kim, J., Kuramochi-Miyagawa, S., Nakano, T., Bartel, D.P., Kingston, R.E., 2006. Characterization of the pirna complex
  from rat testes. Science 313, 363–7. doi:10.1126/science.1130164.

- Leung, Y.Y., Kuksa, P.P., Amlie-Wolf, A., Valladares, O., Ungar, L.H., Kannan, S., Gregory, B.D., Wang, L.S., 2016. Dashr: database of small human
  noncoding rnas. Nucleic Acids Res 44, D216–22. doi:10.1093/nar/gkv1188.
- Li, Q., Gregory, R.I., 2008. MicroRNA regulation of stem cell fate. Cell Stem
   Cell .
- Lian, X., Zhang, J., Azarin, S.M., Zhu, K., Hazeltine, L.B., Bao, X.,
  Hsiao, C., Kamp, T.J., Palecek, S.P., 2013. Directed cardiomyocyte differentiation from human pluripotent stem cells by modulating wnt/betacatenin signaling under fully defined conditions. Nat Protoc 8, 162–75.
  doi:10.1038/nprot.2012.150.
- Liu, Q., Jiang, C., Xu, J., Zhao, M.T., Van Bortle, K., Cheng, X., Wang,
  G., Chang, H.Y., Wu, J.C., Snyder, M.P., 2017. Genome-wide temporal profiling of transcriptome and open chromatin of early cardiomyocyte
  differentiation derived from hipscs and hescs. Circ Res 121, 376–391.
  doi:10.1161/CIRCRESAHA.116.310456.
- Liu, Y., Zhang, J., Li, A., Liu, Z., He, Z., Yuan, X., Tuo, S., 2018. Prediction of cancer-associated pirna-mrna and pirna-lncrna interactions by
  integrated analysis of expression and sequence data. Tsinghua Science and
  Technology 23, 115–125. doi:10.26599/TST.2018.9010056.
- Loh, K.M., Chen, A., Koh, P.W., Deng, T.Z., Sinha, R., Tsai, J.M., Barkal,
  A.A., Shen, K.Y., Jain, R., Morganti, R.M., Shyh-Chang, N., Fernhoff,
  N.B., George, B.M., Wernig, G., Salomon, R.E.A., Chen, Z., Vogel, H.,
  Epstein, J.A., Kundaje, A., Talbot, W.S., Beachy, P.A., Ang, L.T., Weissman, I.L., 2016. Mapping the pairwise choices leading from pluripotency
  to human bone, heart, and other mesoderm cell types. Cell 166, 451–67.
  doi:10.1016/j.cell.2016.06.011.
- Malone, C.D., Brennecke, J., Dus, M., Stark, A., McCombie, W.R., Sachidanandam, R., Hannon, G.J., 2009. Specialized pirna pathways act in
  germline and somatic tissues of the drosophila ovary. Cell 137, 522–35.
  doi:10.1016/j.cell.2009.03.040.
- Ng, K.W., Anderson, C., Marshall, E.A., Minatel, B.C., Enfield, K.S.S.,
   Saprunoff, H.L., Lam, W.L., Martinez, V.D., 2016. Piwi-interacting rnas

in cancer: emerging functions and clinical utility. Mol Cancer 15, 5.
 doi:10.1186/s12943-016-0491-9.

- Ozata, D.M., Gainetdinov, I., Zoch, A., O'Carroll, D., Zamore, P.D., 2018.
  Piwi-interacting rnas: small rnas with big functions. Nat Rev Genet doi:10.1038/s41576-018-0073-3.
- Pezic, D., Manakov, S.A., Sachidanandam, R., Aravin, A.A., 2014. pirna
  pathway targets active line1 elements to establish the repressive h3k9me3
  mark in germ cells. Genes Dev 28, 1410–28. doi:10.1101/gad.240895.114.
- Phay, M., Kim, H.H., Yoo, S., 2018. Analysis of pirna-like small non-coding
  rnas present in axons of adult sensory neurons. Mol Neurobiol 55, 483–494.
  doi:10.1007/s12035-016-0340-2.
- Quinlan, A.R., Hall, I.M., 2010. Bedtools: a flexible suite of utilities for comparing genomic features. Bioinformatics 26, 841–2.
  doi:10.1093/bioinformatics/btq033.
- Rajan, K.S., Velmurugan, G., Pandi, G., Ramasamy, S., 2014. mirna and
  pirna mediated akt pathway in heart: antisense expands to survive. Int J
  Biochem Cell Biol 55, 153–6. doi:10.1016/j.biocel.2014.09.001.
- Rajasethupathy, P., Antonov, I., Sheridan, R., Frey, S., Sander, C., Tuschl,
  T., Kandel, E.R., 2012. A role for neuronal pirnas in the epigenetic control of memory-related synaptic plasticity. Cell 149, 693–707.
  doi:10.1016/j.cell.2012.02.057.
- Ramírez, F., Ryan, D.P., Grüning, B., Bhardwaj, V., Kilpert, F., Richter,
  A.S., Heyne, S., Dündar, F., Manke, T., 2016. deeptools2: a next generation web server for deep-sequencing data analysis. Nucleic Acids Res 44,
  W160-5. doi:10.1093/nar/gkw257.
- Ro, S., Park, C., Song, R., Nguyen, D., Jin, J., Sanders, K.M., McCarrey,
   J.R., Yan, W., 2007. Cloning and expression profiling of testis-expressed
   pirna-like rnas. RNA 13, 1693–702. doi:10.1261/rna.640307.
- Ross, R.J., Weiner, M.M., Lin, H., 2014. Piwi proteins and piwi-interacting
  rnas in the soma. Nature 505, 353–359. doi:10.1038/nature12987.

Roy, J., Sarkar, A., Parida, S., Ghosh, Z., Mallick, B., 2017. Small
rna sequencing revealed dysregulated pirnas in alzheimer's disease
and their probable role in pathogenesis. Mol Biosyst 13, 565–576.
doi:10.1039/c6mb00699j.

Ruby, J.G., Jan, C., Player, C., Axtell, M.J., Lee, W., Nusbaum, C., Ge,
H., Bartel, D.P., 2006. Large-scale sequencing reveals 21u-rnas and additional micrornas and endogenous sirnas in c. elegans. Cell 127, 1193–207.
doi:10.1016/j.cell.2006.10.040.

Saito, K., Nishida, K.M., Mori, T., Kawamura, Y., Miyoshi, K., Nagami, T.,
Siomi, H., Siomi, M.C., 2006. Specific association of piwi with rasirnas derived from retrotransposon and heterochromatic regions in the drosophila
genome. Genes Dev 20, 2214–22. doi:10.1101/gad.1454806.

Sato, K., Siomi, M.C., 2013. Piwi-interacting rnas: biological functions and
biogenesis. Essays Biochem 54, 39–52. doi:10.1042/bse0540039.

Sha, Y., Haensel, D., Gutierrez, G., Du, H., Dai, X., Nie, Q., 2019. Intermediate cell states in epithelial-to-mesenchymal transition. Phys Biol 16, 021001. doi:10.1088/1478-3975/aaf928.

Siomi, M.C., Sato, K., Pezic, D., Aravin, A.A., 2011. Piwi-interacting small
rnas: the vanguard of genome defence. Nat Rev Mol Cell Biol 12, 246–58.
doi:10.1038/nrm3089.

Tohyama, S., Hattori, F., Sano, M., Hishiki, T., Nagahata, Y., Matsuura,
T., Hashimoto, H., Suzuki, T., Yamashita, H., Satoh, Y., Egashira, T.,
Seki, T., Muraoka, N., Yamakawa, H., Ohgino, Y., Tanaka, T., Yoichi,
M., Yuasa, S., Murata, M., Suematsu, M., Fukuda, K., 2013. Distinct metabolic flow enables large-scale purification of mouse and human
pluripotent stem cell-derived cardiomyocytes. Cell Stem Cell 12, 127–37.
doi:10.1016/j.stem.2012.09.013.

- Tosar, J.P., Rovira, C., Cayota, A., 2018. Non-coding rna fragments account
  for the majority of annotated pirnas expressed in somatic non-gonadal
  tissues. Commun Biol 1, 2. doi:10.1038/s42003-017-0001-7.
- <sup>728</sup> Uhrig, S., Klein, H., 2018. Pingpongpro: a tool for the detection of
   <sup>729</sup> pirna-mediated transposon silencing in small rna-seq data. Bioinformatics
   <sup>730</sup> doi:10.1093/bioinformatics/bty578.

Vella, S., Gallo, A., Lo Nigro, A., Galvagno, D., Raffa, G.M., Pilato,
M., Conaldi, P.G., 2016. Piwi-interacting rna (pirna) signatures in
human cardiac progenitor cells. Int J Biochem Cell Biol 76, 1–11.
doi:10.1016/j.biocel.2016.04.012.

Wang, J., Zhang, P., Lu, Y., Li, Y., Zheng, Y., Kan, Y., Chen, R., He, S.,
2018. pirbase: a comprehensive database of pirna sequences. Nucleic Acids
Res doi:10.1093/nar/gky1043.

Yan, Z., Hu, H.Y., Jiang, X., Maierhofer, V., Neb, E., He, L., Hu, Y., Hu,
H., Li, N., Chen, W., Khaitovich, P., 2011. Widespread expression of
pirna-like molecules in somatic tissues. Nucleic Acids Res 39, 6596–607.
doi:10.1093/nar/gkr298.

Zamore, P.D., 2010. Somatic pirna biogenesis. EMBO J 29, 3219–21.
doi:10.1038/emboj.2010.232.

Zong, X., Nakagawa, S., Freier, S.M., Fei, J., Ha, T., Prasanth, S.G.,
Prasanth, K.V., 2016. Natural antisense rna promotes 3' end processing and maturation of malat1 lncrna. Nucleic Acids Res 44, 2898–908.
doi:10.1093/nar/gkw047.



AFigure\_2



























		intered (2)	intered (3)		
psc1	13,475,096	5,622,650	$1,\!272,\!076$	41.73	9.44
psc2	8,979,026	$2,\!481,\!733$	656,781	27.64	7.31
psc3	$9,\!306,\!433$	$3,\!418,\!745$	844,994	36.742	9.08
mpc1	$18,\!690,\!310$	8,922,435	$3,\!494,\!534$	47.74	18.705
mpc2	$13,\!137,\!228$	5,017,796	1,090,201	38.20	8.30
mpc3	$14,\!426,\!323$	$5,\!582,\!968$	$1,\!921,\!400$	38.70	13.32
cpc1	$10,\!550,\!374$	$3,\!362,\!057$	$635,\!333$	31.87	6.02
$\mathbf{cpc2}$	$10,\!863,\!324$	4,833,395	882,061	44.49	8.12
cpc3	8,045,810	3,770,766	$969,\!896$	46.87	12.05

FIGURE S1: Read length control in filtered samples and mapping to other non coding RNAs. Related to Figure 1. A) Read length expressed as a function of density for pluripotent (PSC1, PSC2 and PSC3), mesoderm progenitor (MPC1, MPC2 and MPC3) and cardiomyocytes (CPC1, CPC2 and CPC3) samples before filtering (unfiltered) and after filtering (23 < RL < 35 and -ncRNA). Color key is indicated in the plot. B) Number of mapped reads. Reads were counted before processing (1) and after being filtered by length (2) and other ncRNAs (3). Remaining reads after processing are expressed as percentage(%) of unfiltered reads (2/1 and 3/1). C) Analysis of coverage on non coding RNAs loci from DASHR database for fully processed normalized (counts per million) samples.



FIGURE S2: **Processing of testis samples. Related to Figure 2.** a) Number of mapped reads after employing the pipeline described in Figure 1 in human testis samples downloaded from ENCODE (merged replicates). b) Frequency of bases per position in processed mapped reads.



FIGURE S3: Analysis of expression data and DE results. Related to Figure 3. A) Boxplot showing reads for the nine samples normalized by library depth and expressed as log2 counts per million (CPM). B) Heatmap in log2 CPM of piRNAs from a. Hard unsupervised clustering was performed on rows (piRNA ID) and columns (sample ID), and is shown as dendrograms. Color keys for heatmap and phenotype are indicated to left and in the top right corner of the graph, respectively. C) Principal Component Analysis performed on DESeq2 normalized counts. The color key is indicated to the right of the plot. D) Overlap of differentially expressed piRNAs in CPC versus MPC (153; green circle) and PSC (137; purple circle). E) Normalized expression of piRNAs upregulated, dowregulated and non-regulated with respect to CPC. F) Three piRNA transcripts (piR-1919272, piR-2519215 and piR-97458) were evaluated by qPCR using a specific retrotranscription protocol designed for small RNAs in day 0 and 14 of cardiac differentiation. Expression of mir302b -marker of pluripotency- was analyzed to assess protocol success. All results were expressed as mean±se of two independent experiments after normalization by small RNA RNU6B.



FIGURE S4: Expression profile of human Piwi genes in H1 and H9 embryonic stem cell lines. Related to Figure 4. Normalized RNA-seq counts (FPKM) from H1 and H9 cell lines were downloaded from GEO.



FIGURE S5: Expression of piRNAs in mitochondrial chromosome. Related to Figure 5. a) Distribution of PSC, MPC and CPC mapped reads (merged replicates) as a function of density over a fraction of mitochondrial chromosome (chrM:1-4000). Profiles above zero correspond to plus strand and below zero to the minus strand. Color key is located at the bottom left corner of the plot. b) Coverage profiles in counts per million mapped reads (CPM) on the entire mitochondrial rRNA extension (MT-rRNA) and center of tRNA (MT-tRNA). Direction of rRNA genes are indicated by 5' and 3'. c) Image captured from IGV software over a portion of the human mitochondrial chromosome (chrM:2,184-2,780). The tracks from top to bottom are: piRbase annotated piRNAs (piRbase), piRNAs identified in our samples (piRNAs), coverage profiles of PSC, MPC and CPC, DASHR database ncRNA annotations (DASHR) and GENCODE v29 gene annotations (GENCODE v29). d) Fraction of piRNAs mapped to genomic features annotated in GENCODE v29 database in length-filtered samples (length) compared to length+ncRNA-filtered (length+ncRNA) samples. Color key for features is indicated to the rigth of the bars.



FIGURE S6: Coverage on protein coding and lncRNA genes. Related to Figure 6 and 7. A) NPC sample processing. Number of mapped reads after employing the pipeline described in Figure 1 in neural progenitor samples downloaded from ENCODE project. B) Desity estimation of percent (%) coverage on protein coding genes intersected by piRNAs in sense orientation. Vertical lines indicate medians of each curve. C) Image captured from IGV software portraying mapped reads of PSC, MPC, CPC and NPC samples on identified piRNAs. Tracks for piRNA annotation database (piRbase) and gene features (GENCODE v29) are shown. D) Density estimation as in a, in antisense orientation. E) Left panel shows IGV capture depicting piRNAs in PARP2 vecinity. Expression dynamic of PARP2 gene in RNA-seq samples from H9 cells differentiated to CM is shown to the right.

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
1	piR-hsa-1389062	252.26	1263.26	1335.60	666.72	757.88	83.67	110.68	260.73	280.30
2	piR-hsa-4381848 2	86.38	70.78	77.08	78.63	235.99	77.84	81.81	31.18	79.49
3	piR-hsa-3732777	1749	35.39	110.80	$105 \ 40$	106.53	60.32	24.06	55 58	73 21
4	piR-hsa-1904126	12.72	40.01	49.38	96.20	72.82	83.67	57.75	39.76	89.25
5	piR-hsa-1919272	50.88	26.16	33.72	10.20	72.02 72.82	3.89	178.05	41 12	60.66
6	piR-hsa-3660133	88.50	20.10 67 70	77.08	14.34	64.73	35.03	1.60	0.00	0.00
7	piR-lisa-3000133	25.00	60.24	10.38	93 49	04.75 31.09	60 32	0.00	1.36	2.00
0	piR-lisa-3617530	25.91	15.24	26 50	18.40	12.40	0.02	44.01	16.97	2.19
0	pin-lisa-151400	20.44 20.14	10.09	20.00	10.40 0.51	10.49	1.05	44.91 01 01	10.27	11.40 11.14
9	piR-lisa-2559702	20.14 22.70	9.20 119.29	21.00	2.01 5.02	6.74	25.20	01.01	22.14	41.14 0.70
10	pin-lisa-4091200	22.19	0.00	26.90	148.00	0.74	20.00	0.00	0.00 6 79	0.70
11	pin-lisa-1959065	40.20	0.00	0.00	140.90	2.10	0.72	0.00	0.70	2.09
12	pin-lisa-109217	49.29	20.10	261	19.24	29.07 E7.00	9.73	0.00	0.90	2.09
13	pin-lisa-97456	1.06	0.00	0.01 1.00	22.09	0.00	93.40	14.44	14.01	121.00
14	piR-fisa-000910	10.61	0.00	1.20	2.01	0.00	1.10	14.44	14.01	151.08
10 10	pin-fisa-2540970	19.01	00.01	30.11 2.61	22.09	22.95	9.75	0.00	0.00	0.70
10	plR-nsa-1955270	2.00	0.00	3.01 0.42	107.08	30.41 10.70	13.02	105.07	0.45	10.59
10	p1R-nsa-3034065	5.30	0.15	8.43	0.00	10.79	0.00	105.87	0.78	19.52
18	piR-hsa-2489909	0.00	3.08	3.61	0.69	5.39	114.81	4.81	3.01	7.07
19	p1R-hsa-3658275	14.84	7.69	3.61	1.07	16.18	3.89	67.37	14.91	18.83
20	piR-hsa-2832647	7.42	46.16	6.02	7.53	2.70	27.24	20.85	14.91	5.58
21	p1R-hsa-3352181	12.72	16.93	10.84	89.51	2.70	1.95	0.00	0.00	2.09
22	p1R-hsa-151249	7.95	10.77	19.27	40.15	16.18	15.57	1.60	18.98	4.18
23	piR-hsa-316012	4.77	13.85	6.02	2.51	10.79	31.13	20.85	14.46	26.50
24	piR-hsa-2479371	21.73	10.77	14.45	5.02	24.27	1.95	30.48	8.59	10.46
25	piR-hsa-611204	6.89	0.00	8.43	51.87	45.85	9.73	1.60	0.00	1.39
26	piR-hsa-2513278	10.07	9.23	19.27	27.61	39.11	13.62	3.21	0.00	1.39
27	piR-hsa-3974794	32.86	18.46	21.68	15.89	6.74	15.57	3.21	1.81	4.88
28	piR-hsa-2780538	15.37	35.39	24.09	7.53	9.44	25.30	0.00	0.90	2.79
29	piR-hsa-1872085_4	4.24	26.16	8.43	25.10	4.05	31.13	3.21	5.87	8.37
30	piR-hsa-2526525	6.36	4.62	6.02	0.00	5.39	0.00	73.79	2.26	11.16
31	piR-hsa-745484	2.12	3.08	0.00	62.74	2.70	0.00	33.69	3.61	0.70
32	piR-hsa-147461	5.83	3.08	6.02	32.62	18.88	29.19	1.60	7.68	0.70
33	piR-hsa-4460706	11.66	4.62	1.20	0.00	10.79	1.95	41.71	15.36	16.73
34	piR-hsa-1927965	11.13	12.31	3.61	0.00	10.79	0.00	51.33	2.26	9.76
35	piR-hsa-4303719	13.78	24.62	36.13	14.22	2.70	9.73	0.00	0.00	0.00
36	piR-hsa-114666	18.02	15.39	26.50	1.67	9.44	29.19	0.00	0.45	0.00
37	piR-hsa-1870588	18.02	1.54	9.63	1.67	16.18	0.00	35.29	9.49	8.37
38	piR-hsa-7760463	1.59	0.00	0.00	33.46	55.29	3.89	1.60	0.00	4.18
39	piR-hsa-1706026	1.06	4.62	0.00	74.45	9.44	0.00	8.02	0.00	1.39
40	piR-hsa-2856604	4.24	23.08	24.09	25.93	9.44	7.78	0.00	0.45	2.09
41	piR-hsa-1233052	17.49	3.08	7.23	0.00	12.14	0.00	41.71	9.04	4.18
42	piR-hsa-2482189	2.12	10.77	0.00	49.36	6.74	3.89	14.44	3.16	3.49
43	piR-hsa-2519215	15.90	18.46	37.33	7.53	5.39	7.78	0.00	0.00	0.00
44	piR-hsa-2863156	5.30	15.39	6.02	20.08	4.05	35.03	4.81	0.90	0.70
45	piR-hsa-151136	8.48	21.54	14.45	16.73	5.39	11.68	3.21	8.13	0.70
46	piR-hsa-214132	12.72	23.08	19.27	8.37	12.14	13.62	0.00	0.45	0.00
47	piR-hsa-1332287	6.89	52.32	13.25	0.00	5.39	5.84	0.00	2.26	1.39
48	piR-hsa-1923208	6.89	7.69	2.41	3.35	2.70	1.95	35.29	5.87	19.52
49	piR-hsa-1463989	2.65	21.54	27.70	6.69	5.39	19.46	0.00	0.00	0.00
50	piR-hsa-2413094	12.72	30.77	20.47	0.84	10.79	7.78	0.00	0.00	0.00
51	piR-hsa-2299252	14.84	4.62	28.90	10.04	8.09	15.57	0.00	0.00	0.00
52	piR-hsa-2831593	6.89	1.54	27.70	11.71	14.83	1.95	0.00	11.75	5.58
53	piR-hsa-1429070	2.12	4.62	7.23	3.35	6.74	56.43	0.00	0.00	0.00
54	piR-hsa-2268195	14.84	4.62	32.52	1.67	4.05	13.62	1.60	1.36	4.18
55	piR-hsa-4020841	5.83	23.08	30.11	1.67	5.39	11.68	0.00	0.00	0.00
56	piR-hsa-1647694	9.54	12.31	30.11	8.37	6.74	7.78	0.00	0.00	1.39
57	piR-hsa-359160_2	2.12	7.69	3.61	25.10	8.09	9.73	1.60	8.59	9.06
58	piR-hsa-1001021	4.77	4.62	1.20	23.42	6.74	17.51	11.23	0.00	5.58
59	piR-hsa-772699	0.00	0.00	0.00	53.54	8.09	7.78	3.21	0.90	1.39

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
60	piR-hsa-3916570	15.90	7.69	28.90	11.71	6.74	3.89	0.00	0.00	0.00
61	piR-hsa-1916259	3.18	3.08	1.20	23.42	8.09	3.89	25.67	4.52	0.70
62	piR-hsa-4322932	6.89	24.62	28.90	4.18	4.05	3.89	0.00	0.45	0.70
63	piR-hsa-2521457	3.71	4.62	3.61	1.67	4.05	0.00	41.71	5.42	6.97
64	piR-hsa-1903779	1.59	3.08	7.23	4.18	6.74	42.81	4.81	0.00	0.70
65	piR-hsa-2646470	0.53	4.62	8.43	22.59	21.58	3.89	1.60	2.26	4.88
66	piR-hsa-2497478	24.38	4.62	8.43	1.67	5.39	0.00	19.25	2.26	3.49
67	piB-hsa-1875212	4.24	3.08	1.20	0.00	4.05	0.00	38.50	8.13	9.76
68	piR-hsa-4387218	0.00	0.00	0.00	0.84	1.35	0.00	4.81	27.11	31.38
69	piR-hsa-6913457	10.60	16.93	7.23	20.91	5.39	1.95	1.60	0.00	0.00
70	piB-hsa-1756618	2.65	0.00	3.61	19.24	21.58	3.89	9.62	0.45	3.49
71	piB-hsa-2592846	0.00	0.00	0.00	0.00	0.00	0.00	60.96	0.90	2.09
72	piR-hsa-1908839	6.36	0.00	2.41	1.67	17.53	0.00	16.04	10.84	7.67
73	piR-hsa-1576285	0.00	3.08	0.00	1.67	1.35	13.62	3.21	20.33	18.83
74	piR-hsa-1862211	0.00	7.69	0.00	12.55	0.00	21.40	11.23	3.61	2.79
75	piR-hsa-3658742	6.89	1.54	4.82	0.00	24.27	0.00	16.04	2.26	2.79
76	piR-hsa-1872235	6.89	3.08	9.63	0.00	8.09	1.95	16.04	7.23	4.88
77	piR-hsa-1277994	3.71	0.00	8.43	2.51	29.67	1.95	4.81	0.00	6.28
78	piR-hsa-343382	4.24	4.62	2.41	1.67	4.05	1.95	19.25	9.04	9.76
79	piR-hsa-2027369	14.84	16.93	12.04	4.18	2.70	5.84	0.00	0.00	0.00
80	piR-hsa-1399886	2.12	24.62	4.82	15.89	8.09	0.00	0.00	0.00	0.00
81	piR-hsa-1798104	7.42	15.39	24.09	2.51	1.35	3.89	0.00	0.00	0.00
82	piR-hsa-1875155	2.12	21.54	21.68	3.35	0.00	5.84	0.00	0.00	0.00
83	piR-hsa-1548068	1.59	6.15	13.25	4.18	10.79	0.00	1.60	8.13	8.37
84	piR-hsa-721859 9	2.12	9.23	8.43	6.69	4.05	17.51	3.21	0.90	1.39
85	piR-hsa-3732088	16.43	3.08	20.47	1.67	1.35	1.95	0.00	3.16	4.88
86	piR-hsa-1303811	1.59	4.62	0.00	31.79	1.35	3.89	9.62	0.00	0.00
87	piR-hsa-2481097	6.89	3.08	6.02	0.00	12.14	0.00	19.25	4.07	1.39
88	piR-hsa-4028152	2.12	15.39	0.00	5.02	2.70	25.30	1.60	0.00	0.70
89	piR-hsa-389007	2.65	4.62	2.41	9.20	10.79	19.46	3.21	0.00	0.00
90	piR-hsa-2477264	5.30	6.15	6.02	0.00	10.79	0.00	17.64	4.07	2.09
91	piR-hsa-255984	2.12	3.08	12.04	5.86	2.70	23.35	0.00	0.00	2.09
92	piR-hsa-137136	7.42	7.69	9.63	8.37	8.09	9.73	0.00	0.00	0.00
93	piR-hsa-3807498	7.95	13.85	15.66	1.67	6.74	3.89	0.00	0.00	0.70
94	piR-hsa-783698	15.37	7.69	6.02	5.02	13.49	1.95	0.00	0.00	0.00
95	piR-hsa-1988800	7.95	4.62	12.04	3.35	14.83	5.84	0.00	0.00	0.00
96	piR-hsa-1941780	3.71	6.15	3.61	1.67	2.70	1.95	19.25	1.36	7.67
97	piR-hsa-3875265	10.60	9.23	10.84	5.86	9.44	1.95	0.00	0.00	0.00
98	piR-hsa-1876265	4.24	4.62	3.61	0.00	0.00	0.00	19.25	6.33	9.76
99	piR-hsa-368987	6.36	4.62	4.82	5.02	4.05	3.89	9.62	5.87	3.49
100	piR-hsa-1890632	6.36	4.62	7.23	0.00	5.39	0.00	11.23	4.52	8.37
101	piR-hsa-4078407	2.12	20.00	9.63	10.04	1.35	3.89	0.00	0.00	0.00
102	piR-hsa-76848	4.24	4.62	1.20	1.67	1.35	25.30	1.60	1.36	5.58
103	piR-hsa-2565910	0.53	12.31	4.82	5.02	5.39	15.57	0.00	1.81	1.39
104	piR-hsa-1205256	3.18	4.62	1.20	21.75	0.00	11.68	3.21	0.45	0.00
105	piR-hsa-2840936	10.60	0.00	1.20	0.00	6.74	0.00	19.25	4.97	2.79
106	piR-hsa-20628	3.71	9.23	12.04	2.51	8.09	9.73	0.00	0.00	0.00
107	piR-hsa-753191	0.00	0.00	0.00	0.00	0.00	0.00	35.29	5.42	4.18
108	piR-hsa-3683883	2.12	3.08	6.02	20.08	1.35	1.95	6.42	0.00	2.79
109	piR-hsa-1690788	1.59	4.62	4.82	5.86	6.74	17.51	0.00	0.00	2.09
110	piR-hsa-2530015	6.89	3.08	1.20	2.51	0.00	0.00	16.04	2.26	11.16
111	piR-hsa-346271	3.71	3.08	15.66	1.67	2.70	0.00	3.21	1.81	10.46
112	piR-hsa-3546008	6.89	3.08	8.43	5.02	14.83	3.89	0.00	0.00	0.00
113	piR-hsa-5982715	7.42	13.85	2.41	6.69	2.70	7.78	0.00	0.90	0.00
114	piR-hsa-2398570	1.06	6.15	3.61	2.51	12.14	15.57	0.00	0.00	0.70
115	piR-hsa-1883972	6.36	0.00	0.00	0.00	2.70	0.00	19.25	3.61	9.76
116	piR-hsa-3718263_3	0.00	0.00	0.00	0.00	0.00	0.00	33.69	5.87	2.09
117	piR-hsa-4472891	2.12	3.08	9.63	11.71	6.74	5.84	0.00	1.81	0.70
118	piR-hsa-2208850	4.24	15.39	7.23	5.02	1.35	7.78	0.00	0.00	0.00

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
119	piR-hsa-785969	2.12	3.08	0.00	6.69	0.00	1.95	24.06	1.36	1.39
120	piR-hsa-163695	6.36	10.77	7.23	0.84	6.74	7.78	0.00	0.00	0.70
121	piR-hsa-2490897	6.89	0.00	1.20	1.67	0.00	0.00	11.23	5.42	13.95
122	piR-hsa-2151268	1.59	7.69	3.61	7.53	8.09	11.68	0.00	0.00	0.00
123	piR-hsa-771714_3	3.71	4.62	2.41	0.00	0.00	3.89	17.64	0.90	6.97
124	piR-hsa-1920687	4.24	4.62	1.20	1.67	4.05	0.00	12.83	5.42	5.58
125	piR-hsa-2351941	5.30	23.08	2.41	2.51	2.70	1.95	1.60	0.00	0.00
126	piR-hsa-3265318	3.18	13.85	6.02	1.67	2.70	11.68	0.00	0.00	0.00
127	piR-hsa-2426792	2.12	24.62	4.82	6.69	0.00	0.00	0.00	0.00	0.70
128	piR-hsa-2503702	13.25	0.00	8.43	0.00	4.05	0.00	6.42	3.16	2.79
129	piR-hsa-161264_3	1.06	0.00	1.20	8.37	5.39	21.40	0.00	0.00	0.00
130	piR-hsa-1912443	1.59	0.00	1.20	0.00	1.35	0.00	28.87	2.26	2.09
131	piR-hsa-2230204	8.48	6.15	15.66	3.35	2.70	0.00	0.00	0.00	0.70
132	piR-hsa-2090264	7.42	1.54	25.29	0.84	1.35	0.00	0.00	0.00	0.00
133	piR-hsa-1242358	0.00	0.00	0.00	0.00	0.00	0.00	32.08	2.26	2.09
134	piR-hsa-724912	2.65	1.54	3.61	22.59	1.35	0.00	3.21	0.45	0.70
135	piR-hsa-7765828	1.06	7.69	3.61	13.38	4.05	5.84	0.00	0.00	0.00
136	piR-hsa-2172087	1.59	10.77	3.61	5.02	2.70	7.78	3.21	0.90	0.00
137	piR-hsa-1425899	0.00	4.62	1.20	6.69	2.70	17.51	0.00	1.36	1.39
138	piR-hsa-1921188	1.59	3.08	1.20	0.00	10.79	0.00	14.44	3.61	0.70
139	piR-hsa-1917139	2.65	1.54	1.20	0.00	4.05	0.00	20.85	1.36	3.49
140	piR-hsa-3829948	0.53	3.08	0.00	13.38	4.05	13.62	0.00	0.00	0.00
141	piR-hsa-3735787	7.42	3.08	2.41	0.00	2.70	0.00	12.83	4.07	2.09
142	piR-hsa-5996985	0.00	0.00	0.00	0.00	0.00	0.00	28.87	5.42	0.00
143	piR-hsa-2989729	6.36	6.15	13.25	0.84	2.70	3.89	0.00	0.00	0.00
144	piR-hsa-21839	0.53	3.08	0.00	24.26	2.70	1.95	0.00	0.45	0.00
145	piR-hsa-1872463	0.00	4.62	1.20	1.67	0.00	19.46	0.00	3.16	2.79
146	p1R-hsa-508592	2.12	13.85	7.23	2.51	1.35	5.84	0.00	0.00	0.00
147	piR-hsa-2252211	1.59	0.15	3.61	0.84	20.23	0.00	0.00	0.45	0.00
148	piR-hsa-4110708	2.65	27.70	1.20	0.84	0.00	0.00	0.00	0.00	0.00
149	p1R-nsa-298158	4.24	0.15	0.02	(.53	4.05	3.89	0.00	0.00	0.00
150	piR-fisa-7892900	0.30	1.54	3.01 2.61	4.18	2.70	1.95	3.21	4.52	3.49
151	pin-fisa-2595910	2.05	0.00 10.21	3.01 2.61	5.50 0.97	1.50	2 80	0.00	0.00	0.00
152	pin-fisa-2550290	0.00	12.31 10.77	5.01 6.02	0.07 E 00	$1.50 \\ 2.70$	0.09 2.00	0.00	0.45	0.70
153	pin-lisa-2152776	2.05	10.77	0.02 19.04	0.02 0.07	2.70 5.20	0.09	0.00	0.00	0.00
154	piR-lisa-3077723	$0.00 \\ 7.42$	4.02 6.15	12.04	0.57 2.51	1.39	0.00	0.00	0.00	1.30
156	piR-lisa-1291010 piR-hsa-1001070	1.42	$0.13 \\ 7.60$	0.04	$\frac{2.01}{3.35}$	1.30 5.30	1.05	0.00	0.90	0.00
$150 \\ 157$	piR-hsa-2450089-2	12.13 0.00	1.03	4.82	3 35	1.35	19.46	0.00	0.00	0.00
157	piR-hsa-108574	3.71	$1.04 \\ 7.69$	4.82	3 35	1.55 8.09	10.40 1.05	0.00	0.00	0.00
150	piR-hsa-1259653	2.65	1.53	$\frac{4.02}{2.41}$	0.84	1.35	1.95	16.04	1.81	0.70
160	piR-hsa-3136454	2.00 2.65	16.93	2.11 2.41	1.67	2.70	1.95	0.00	0.00	0.70
161	piR-hsa-2286229	4.24	3.08	9.63	4.18	5.39	1.95	0.00	0.45	0.00
162	piR-hsa-2209630	2.65	0.00	0.00	23.42	2.70	0.00	0.00	0.00	0.00
163	piR-hsa-1696540	0.00	0.00	0.00	0.00	0.00	1.95	24.06	1.81	0.70
164	piR-hsa-6245615	3.71	4.62	7.23	0.00	10.79	1.95	0.00	0.00	0.00
165	piR-hsa-2072163	1.06	4.62	2.41	1.67	2.70	5.84	8.02	1.81	0.00
166	piR-hsa-1557538	0.00	0.00	2.41	8.37	9.44	7.78	0.00	0.00	0.00
167	piR-hsa-2844156	7.42	3.08	4.82	5.86	0.00	3.89	0.00	1.36	1.39
168	piR-hsa-6744266	0.00	1.54	1.20	5.86	0.00	5.84	9.62	2.26	1.39
169	piR-hsa-1882039	1.06	0.00	0.00	0.00	0.00	0.00	17.64	2.71	6.28
170	piR-hsa-2137611	5.30	15.39	6.02	0.84	0.00	0.00	0.00	0.00	0.00
171	piR-hsa-58291	3.71	6.15	8.43	0.84	4.05	3.89	0.00	0.45	0.00
172	piR-hsa-4408495	0.00	0.00	2.41	5.86	5.39	11.68	1.60	0.45	0.00
173	piR-hsa-374600	1.06	0.00	1.20	10.87	1.35	0.00	12.83	0.00	0.00
174	piR-hsa-2829712	3.71	0.00	1.20	0.00	5.39	0.00	9.62	3.16	4.18
175	piR-hsa-3177742	3.18	4.62	7.23	1.67	0.00	9.73	0.00	0.00	0.70
176	piR-hsa-1585146	2.12	0.00	4.82	9.20	6.74	3.89	0.00	0.00	0.00
177	piR-hsa-4403577	2.65	4.62	7.23	0.84	5.39	3.89	0.00	1.36	0.70

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
178	piR-hsa-1528884	4.24	10.77	4.82	2.51	1.35	1.95	0.00	0.00	0.70
179	piR-hsa-2494226	4.77	3.08	0.00	0.00	2.70	0.00	8.02	2.71	4.88
180	piR-hsa-229786	0.00	4.62	0.00	2.51	2.70	15.57	0.00	0.00	0.70
181	piR-hsa-1492262	3.71	4.62	1.20	0.84	12.14	0.00	1.60	0.45	1.39
182	piR-hsa-132896	1.59	4.62	4.82	4.18	8.09	1.95	0.00	0.00	0.70
183	piR-hsa-1376916	2.12	10.77	2.41	3.35	2.70	3.89	0.00	0.45	0.00
184	piR-hsa-160969	0.53	0.00	6.02	3.35	2.70	11.68	0.00	0.00	1.39
185	piR-hsa-768321	8.48	3.08	1.20	0.84	0.00	0.00	4.81	0.90	6.28
186	piR-hsa-3842249	1.59	0.00	2.41	10.04	1.35	5.84	3.21	0.45	0.70
187	piR-hsa-362913	1.06	1.54	1.20	21.75	0.00	0.00	0.00	0.00	0.00
188	piR-hsa-1870459	3.18	0.00	4.82	0.84	6.74	0.00	4.81	2.71	2.09
189	piR-hsa-3634880	2.12	0.00	0.00	5.02	6.74	3.89	3.21	0.00	4.18
190	piR-hsa-1941637	2.65	0.00	0.00	16.73	1.35	0.00	3.21	0.45	0.70
191	piR-hsa-834074	0.00	0.00	0.00	0.00	0.00	0.00	20.85	2.71	1.39
192	piR-hsa-1900529	5.83	3.08	3.61	0.00	4.05	1.95	0.00	2.26	4.18
193	piR-hsa-2220917	0.00	3.08	1.20	11.71	8.09	0.00	0.00	0.00	0.70
194	p1R-hsa-3739406	3.18	1.54	2.41	5.86	2.70	0.00	1.60	0.45	6.97
195	p1R-hsa-7308134	0.00	0.00	0.00	14.22	2.70	7.78	0.00	0.00	0.00
190	p1R-nsa-1708978	0.53	0.00	0.00	11.(1	2.70	9.73	0.00	0.00	0.00
197	p1R-nsa-3231825	3.71	12.31	3.01	1.07	1.35	1.95	0.00	0.00	0.00
198	p1R-fisa-2398119	4.11	3.08	0.00	1.07	0.39 4.05	0.00	9.02	0.00	0.00
200	pin-lisa-2002000	5.71 4.77	4.02	10.00	$\frac{0.80}{1.67}$	4.00	1.95	1.00	1.50	1.59
200	pin-lisa-313904	4.11	1.04	10.64 2.41	1.07 1.67	$\frac{0.39}{2.70}$	5.84	0.00	0.00	0.00
$\frac{201}{202}$	piR-lisa-4050155	0.09 3.71	4.02	2.41 9.41	10.87	2.70	0.04	0.00	0.00	0.00
202	piR-hsa-2010104	1.50	0.00	2.41 2 41	10.07 12 55	6.74	0.00	0.00	0.00	0.00
203 204	piR-hsa-1843231	1.05	4.62	2.41 4.82	4 18	5 39	3.89	0.00	0.00	0.10
204	piR-hsa-2670375	1.00	1.54	$\frac{4.02}{2.41}$	11 71	1.35	5.05 5.84	0.00	0.00	0.00
200 206	piR-hsa-2464166	1.06	3.08	0.00	0.84	1.35	17.51	0.00	0.00	0.00
207	piR-hsa-2589139	1.59	1.54	4.82	12.55	1.35	1.95	0.00	0.00	0.00
208	piR-hsa-3119265	4.24	1.54	4.82	2.51	6.74	3.89	0.00	0.00	0.00
209	piR-hsa-1531418	1.06	3.08	3.61	6.69	5.39	3.89	0.00	0.00	0.00
210	piR-hsa-2253283	2.65	4.62	0.00	11.71	2.70	1.95	0.00	0.00	0.00
211	piR-hsa-2148238	1.59	7.69	8.43	2.51	1.35	1.95	0.00	0.00	0.00
212	piR-hsa-1595580	2.12	6.15	7.23	2.51	5.39	0.00	0.00	0.00	0.00
213	piR-hsa-3978322	0.00	0.00	0.00	10.04	6.74	3.89	1.60	0.90	0.00
214	piR-hsa-1773241	2.65	1.54	3.61	3.35	8.09	3.89	0.00	0.00	0.00
215	piR-hsa-1434629	0.53	1.54	3.61	7.53	4.05	5.84	0.00	0.00	0.00
216	piR-hsa-1340768	0.53	1.54	1.20	3.35	6.74	7.78	0.00	0.45	1.39
217	piR-hsa-333507	1.59	4.62	8.43	1.67	2.70	3.89	0.00	0.00	0.00
218	piR-hsa-118348	3.18	6.15	3.61	3.35	4.05	0.00	0.00	0.45	2.09
219	p1R-hsa-3623001	4.24	1.54	1.20	0.00	9.44	0.00	1.60	1.36	3.49
220	piR-hsa-2090890	2.65	4.62	4.82	4.18	2.70	3.89	0.00	0.00	0.00
221	p1R-nsa-1929007	2.12	9.23	1.20	2.51	0.00	1.18	0.00	0.00	0.00
222	plR-fisa-2827579	1.00	0.00	0.00	0.00	1.30	1.95	0.42	0.90	0.70
220	pin-fisa-1707105	0.00	1.04	0.00	0.00	0.00	0.00	10.04	4.02	0.70
$\frac{224}{225}$	piR-lisa-120000	6.80	1.54	0.00 2 41	9.20	2.70	9.75	0.00	0.45	0.70 2.00
220	piR-hsa-1607096	1.06	12.04	1.20	0.00	2.70	3.80	4.81	0.00	2.09
220 227	piR-hsa-2515454	5.30	3.08	0.00	0.04	5 39	0.00	6.42	1.81	0.70
228	piR-hsa-368381	0.50 0.53	1.53	1.20	2.50	0.00	1.95	4 81	3 16	6.97
229	piR-hsa-4379982	4.24	4.62	1.20	0.84	9.44	0.00	1.60	0.00	0.70
230	piR-hsa-2240007	3.71	7.69	4.82	1.67	2.70	1.95	0.00	0.00	0.00
231	piR-hsa-4416099_9	0.00	0.00	1.20	0.84	0.00	19.46	0.00	0.90	0.00
232	piR-hsa-7106256	1.59	0.00	1.20	10.87	5.39	1.95	0.00	0.45	0.70
233	piR-hsa-1481120	2.12	3.08	1.20	8.37	5.39	1.95	0.00	0.00	0.00
234	piR-hsa-1284504	0.53	1.54	0.00	4.18	0.00	3.89	6.42	2.71	2.79
235	piR-hsa-2505515	0.00	0.00	0.00	7.53	6.74	7.78	0.00	0.00	0.00
236	piR-hsa-1927627	0.00	0.00	0.00	0.00	0.00	0.00	16.04	1.81	4.18

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
237	piR-hsa-1921551	2.12	1.54	1.20	12.55	2.70	0.00	0.00	0.45	1.39
238	piR-hsa-163499	4.77	3.08	3.61	1.67	2.70	1.95	1.60	0.45	2.09
239	piR-hsa-4202081	1.59	1.54	4.82	4.18	5.39	3.89	0.00	0.45	0.00
240	piR-hsa-147696	1.59	0.00	0.00	0.84	1.35	0.00	9.62	4.07	4.18
241	piR-hsa-2353109	0.00	12.31	1.20	0.84	1.35	5.84	0.00	0.00	0.00
242	piR-hsa-8270846	3.71	4.62	7.23	5.86	0.00	0.00	0.00	0.00	0.00
243	piR-hsa-1905680	0.53	1.54	3.61	0.84	0.00	0.00	8.02	4.07	2.79
244	piR-hsa-307961	2.12	6.15	0.00	3.35	0.00	9.73	0.00	0.00	0.00
245	piR-hsa-2333057	4.77	0.00	3.61	0.84	9.44	1.95	0.00	0.00	0.70
246	piR-hsa-1938524	0.53	0.00	1.20	0.84	1.35	1.95	11.23	1.36	2.79
247	piR-hsa-3232943	3.18	0.00	3.61	5.02	5.39	3.89	0.00	0.00	0.00
248	piR-hsa-3513154	5.30	1.54	7.23	4.18	2.70	0.00	0.00	0.00	0.00
249	piR-hsa-3674332	14.31	0.00	0.00	0.00	2.70	0.00	1.60	0.90	1.39
250	piR-hsa-2213434	6.36	3.08	7.23	4.18	0.00	0.00	0.00	0.00	0.00
251	piR-hsa-2308163	2.65	7.69	3.61	4.18	2.70	0.00	0.00	0.00	0.00
252	piR-hsa-1748898	0.00	7.69	4.82	5.02	1.35	1.95	0.00	0.00	0.00
253	piR-hsa-2525461	0.00	3.08	3.61	6.69	2.70	1.95	0.00	0.00	2.79
254	piR-hsa-1296118	3.18	0.00	2.41	0.00	2.70	0.00	8.02	3.61	0.70
255	p1R-hsa-1632961	0.53	12.31	3.61	0.84	1.35	1.95	0.00	0.00	0.00
256	p1R-hsa-728085	0.53	1.54	0.00	0.84	0.00	0.00	17.64	0.00	0.00
257	p1R-nsa-023353	0.00	1.54	0.00	0.84	0.00	15.57	0.00	0.45	2.09
258	p1R-nsa-3527815	1.00	13.85	3.01	0.00	0.00	1.95	0.00	0.00	0.00
209	plR-fisa-2478880_2	0.00	1.54	0.00	0.00	2.70	0.00	8.02	3.10	4.88
200 261	pin-lisa-5558751	4.24 5.20	0.10	4.02 7.93	1.07 1.67	$1.00 \\ 1.25$	1.95	0.00	0.00	0.00
201	pin-lisa-4176299	0.00 9.65	4.02 6.15	1.20 9.41	1.07	1.55	0.00	0.00	0.00	0.00
202	piR-lisa-0411007	2.00	7.60	0.00	5.86	4.00	3.89	0.00	1.81	0.00
203 264	piR-hsa-27/22//	1.50	0.00	0.00	0.00	4.05	0.00	0.00	1.01	2.70
$204 \\ 265$	piR-hsa-669874	1.59 0.53	0.00	0.00	19.24	4.00	0.00	9.02	1.01	2.19
$\frac{266}{266}$	piR-hsa-645846	3.18	4.62	3.61	0.84	1.35	5.84	0.00	0.00	0.00
$\frac{260}{267}$	piR-hsa-3776081	$0.10 \\ 0.53$	1.54	0.00	9.20	8.09	0.00	0.00	0.00	0.00
268	piR-hsa-211123	2.12	0.00	1.20	5.02	5.39	3.89	0.00	0.90	0.70
269	piR-hsa-2248086	0.53	4.62	6.02	3.35	2.70	1.95	0.00	0.00	0.00
270	piR-hsa-1909905	0.53	0.00	0.00	1.67	1.35	15.57	0.00	0.00	0.00
271	piR-hsa-1593307	3.18	4.62	6.02	1.67	1.35	1.95	0.00	0.00	0.00
272	piR-hsa-2042088	0.00	0.00	0.00	0.00	0.00	0.00	12.83	4.52	1.39
273	piR-hsa-2490509	0.00	1.54	0.00	0.00	1.35	0.00	12.83	0.90	2.09
274	piR-hsa-144277_2	0.00	0.00	0.00	0.00	5.39	0.00	11.23	1.36	0.70
275	piR-hsa-4131663	0.00	0.00	0.00	4.18	4.05	1.95	8.02	0.45	0.00
276	piR-hsa-2542835	1.59	0.00	1.20	0.00	2.70	0.00	11.23	0.45	1.39
277	piR-hsa-1229611	2.12	1.54	1.20	4.18	2.70	5.84	0.00	0.90	0.00
278	piR-hsa-2423519	1.06	0.00	0.00	0.00	4.05	11.68	1.60	0.00	0.00
279	piR-hsa-3021684	4.24	1.54	3.61	7.53	1.35	0.00	0.00	0.00	0.00
280	piR-hsa-2490287	0.00	1.54	0.00	3.35	1.35	1.95	6.42	2.26	1.39
281	piR-hsa-1259933	1.06	1.54	0.00	8.37	2.70	0.00	1.60	2.26	0.70
282	piR-hsa-1302552	0.53	0.00	0.00	5.86	6.74	0.00	0.00	0.00	4.88
283	p1R-hsa-67957	1.06	13.85	0.00	0.00	0.00	0.00	0.00	0.90	2.09
284	p1R-nsa-1726249	0.89	1.54	0.42	0.84	1.35	0.00	0.00	0.00	0.00
280 286	p1R-nsa-1409954	1.59	1.54	$\frac{8.43}{7.92}$	0.84	0.39 0.70	0.00	0.00	0.00	0.00
$200 \\ 287$	pin-118a-4000010	2.00	1.04	1.23	5.02	2.70	1.99	0.00	1.00	1 80
201 289	pin-115a-2420220	0.00 9.19	2.02	1.20 2.61	J.UZ / 19	2.00	1.05	0.21	1.01	4.00
200 280	pin-115a-1000000 piR_hea_2310750	2.12 2.65	0.00	5.01 7.92	4.10 1.67	2.70	$1.90 \\ 1.05$	0.00	0.00	0.00
209	piit-115a-2019/00 piB_hsa_/20728/	2.00	0.00	1.20 3.61	1.07	0.00	0.72	0.00	0.00	0.00
290 201	piR-hsa-9890/12	1.06	0.00	1.90	1.10 1.10	1 35	0.00	8.02	0.00	1 88
292	piR-hsa-3830126	0.00	0.00	0.00	7.53	4.05	5.84	0.02	0.00	4.00 0.00
292	piR-hsa-3944431	3.71	3.08	3.61	1.67	1.35	3.89	0.00	0.00	0.00
294	piR-hsa-363100 2	2.65	1.54	0.00	1.67	4.05	0.00	3.21	1.36	2.79
295	piR-hsa-6482184	0.53	10.77	1.20	0.84	0.00	3.89	0.00	0.00	0.00

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
296	piR-hsa-642866	0.00	0.00	0.00	0.00	0.00	0.00	12.83	2.26	2.09
297	piR-hsa-1922210	0.53	0.00	1.20	9.20	0.00	3.89	0.00	0.90	1.39
298	piR-hsa-2436454	3.18	3.08	4.82	1.67	0.00	3.89	0.00	0.45	0.00
299	piR-hsa-7821967_3	0.00	0.00	1.20	0.00	0.00	0.00	11.23	3.16	1.39
300	piR-hsa-4424378	0.00	0.00	0.00	0.00	1.35	0.00	12.83	1.36	1.39
301	piR-hsa-237221	1.59	1.54	0.00	10.87	2.70	0.00	0.00	0.00	0.00
302	piR-hsa-7833890	1.59	1.54	0.00	5.02	2.70	5.84	0.00	0.00	0.00
303	piR-hsa-1905329	0.00	3.08	0.00	2.51	1.35	9.73	0.00	0.00	0.00
304	piR-hsa-4450044	0.53	1.54	1.20	13.38	0.00	0.00	0.00	0.00	0.00
305	piR-hsa-3741185	1.59	0.00	0.00	0.00	1.35	1.95	8.02	0.90	2.79
306	piR-hsa-4144265	1.59	3.08	6.02	0.84	0.00	3.89	0.00	0.45	0.70
307	piR-hsa-4198101	1.59	7.69	1.20	0.84	1.35	3.89	0.00	0.00	0.00
308	piR-hsa-8117137	0.00	0.00	2.41	0.84	0.00	1.95	0.00	0.90	10.46
309	piR-hsa-4100164	1.06	4.62	4.82	1.67	2.70	0.00	1.60	0.00	0.00
310	piR-hsa-3280518	6.36	3.08	3.61	3.35	0.00	0.00	0.00	0.00	0.00
311	piR-hsa-3710717	0.00	0.00	0.00	3.35	1.35	11.68	0.00	0.00	0.00
312	piR-hsa-848451	0.00	0.00	0.00	2.51	4.05	9.73	0.00	0.00	0.00
313	piR-hsa-1057272	0.00	0.00	0.00	0.00	1.35	0.00	11.23	2.26	1.39
314	piR-hsa-343616	0.00	0.00	0.00	2.51	0.00	5.84	1.60	1.36	4.88
315	piR-hsa-2832439	1.59	1.54	1.20	0.84	0.00	1.95	4.81	1.36	2.79
316	piR-hsa-2889978	1.06	1.54	1.20	5.02	1.35	5.84	0.00	0.00	0.00
317	piR-hsa-2529368	3.18	1.54	1.20	0.00	0.00	0.00	1.60	1.36	6.97
318	piR-hsa-3161050	0.00	0.00	0.00	5.86	4.05	3.89	1.60	0.45	0.00
319	piR-hsa-1688824	2.12	1.54	1.20	5.86	2.70	1.95	0.00	0.45	0.00
320	piR-hsa-3638679	4.24	4.62	0.00	1.67	1.35	3.89	0.00	0.00	0.00
321	piR-hsa-2281305	0.00	0.00	1.20	0.00	0.00	0.00	9.62	1.36	3.49
322	piR-hsa-2427082	3.18	4.62	4.82	1.67	1.35	0.00	0.00	0.00	0.00
323	piR-hsa-5062317	0.00	4.62	0.00	0.84	0.00	0.00	0.00	1.81	8.37
324	piR-hsa-1555218	1.59	4.62	6.02	3.35	0.00	0.00	0.00	0.00	0.00
325	piR-hsa-4391981_9	0.00	0.00	0.00	3.35	0.00	3.89	1.60	3.16	3.49
326	piR-hsa-4403628	4.77	3.08	0.00	0.84	0.00	0.00	4.81	0.45	1.39
327	piR-hsa-153942	1.06	1.54	1.20	0.00	0.00	0.00	8.02	1.36	2.09
328	piR-hsa-2856544	4.77	1.54	2.41	1.67	0.00	0.00	3.21	0.90	0.70
329	piR-hsa-1894768	1.59	1.54	1.20	0.00	0.00	1.95	3.21	3.61	2.09
330	p1R-hsa-2495779	2.65	1.54	1.20	0.00	4.05	0.00	1.60	2.71	1.39
331	piR-hsa-3706918	0.00	0.00	0.00	3.35	0.00	0.00	6.42	1.81	3.49
332	piR-hsa-4157592	2.12	1.54	1.20	4.18	4.05	1.95	0.00	0.00	0.00
333	piR-nsa-4175180	0.53	1.54	9.63	0.00	1.35	1.95	0.00	0.00	0.00
334	piR-nsa-3714350	0.53	0.00	0.00	9.20	1.35	0.00	3.21	0.00	0.70
330 226	p1R-fisa-354004	1.00	10.77	1.20	0.00	0.00	1.95	0.00	0.00	0.00
000 007	pin-fisa-5094141	0.00	1.04	1.20	10.87	1.55	0.00	0.00	0.00	0.00
220	pin-lisa-4405202	0.00	0.00	0.00	0.00	1.00	1.05	12.65	0.00	2.09
330 330	pin-lisa-5565102	1.09	$\frac{5.00}{7.60}$	1.20	4.10	4.00 1.25	1.95	0.00	0.00	0.00
340	piR-lisa-5557505	0.53	4.62	1.20	0.84 8.37	0.00	0.00	0.00	0.00	0.00
340 341	pin-118a-213629	0.55	4.02	0.00	0.57	8.00	1.05	1.60	0.00	0.00
341	piR-lisa-7050007	0.00	0.00	0.00	2.01	0.09	1.90	12.83	0.00	1.30
342	piR-hsa-255205	0.00	0.00	0.00	11 71	1.35	0.00	12.00 1.60	0.45	0.00
344	piR-hsa-1410500	2.00	0.00	0.00	0.00	1.55	1.05	1.00	0.00	0.00
345	piR-hsa-5124632	1.59	7.20	1.20	0.84	1.35	1.95	0.00	0.00	0.00
346	piR-hsa-3365644	0.00	0.00	2.41	7.53	2.70	1.05 1.95	0.00	0.00	0.00
347	piR-hsa-1886018	0.00	0.00	0.00	0.00	0.00	0.00	11 23	1.36	1.39
348	piR-hsa-3657078	6.89	0.00	2.41	0.84	1.35	0.00	0.00	0.90	2.09
349	piR-hsa-3104125	4.24	1.54	4.82	2.51	1.35	0.00	0.00	0.00	0.00
350	piR-hsa-4378137 4	0.53	1.54	0.00	4.18	2.70	3.89	0.00	0.90	0.70
351	piR-hsa-1592257	0.00	0.00	0.00	0.00	0.00	0.00	11.23	1.81	1.39
352	piR-hsa-3818788	4.24	0.00	6.02	0.00	4.05	0.00	0.00	0.00	0.00
353	piR-hsa-2839864	1.06	0.00	0.00	0.84	0.00	0.00	6.42	3.16	2.79
354	piR-hsa-1678085	0.00	0.00	1.20	1.67	9.44	1.95	0.00	0.00	0.00

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
355	piR-hsa-4193743	0.00	0.00	0.00	0.00	0.00	0.00	11.23	0.90	2.09
356	piR-hsa-2537452	0.00	0.00	0.00	14.22	0.00	0.00	0.00	0.00	0.00
357	piR-hsa-4507261	0.00	1.54	0.00	0.00	0.00	0.00	0.00	2.71	9.76
358	piR-hsa-1263612	6.89	0.00	4.82	0.00	0.00	0.00	0.00	0.90	1.39
359	piR-hsa-778924	0.53	1.54	0.00	7.53	2.70	0.00	1.60	0.00	0.00
360	piR-hsa-2165649	1.06	4.62	4.82	0.00	1.35	1.95	0.00	0.00	0.00
361	piR-hsa-1274138	1.06	0.00	0.00	0.00	1.35	1.95	6.42	0.90	2.09
362	piR-hsa-4111185	1.06	3.08	1.20	7.53	0.00	0.00	0.00	0.00	0.70
363	piR-hsa-3693411	0.00	0.00	0.00	5.86	2.70	1.95	1.60	0.00	1.39
364	piR-hsa-1917013	0.53	3.08	0.00	0.00	0.00	0.00	6.42	2.71	0.70
365	piR-hsa-4467055_8	0.00	0.00	2.41	5.86	0.00	3.89	0.00	0.45	0.70
366	piR-hsa-4402141	0.00	0.00	0.00	0.00	0.00	0.00	11.23	1.36	0.70
367	piR-hsa-942735	0.00	0.00	0.00	0.00	0.00	0.00	11.23	1.36	0.70
368	piR-hsa-1574545	0.00	0.00	0.00	0.84	0.00	0.00	9.62	0.00	2.79
369	piR-hsa-2161668	0.00	0.00	1.20	4.18	5.39	1.95	0.00	0.45	0.00
370	piR-hsa-771001	0.53	0.00	0.00	9.20	1.35	0.00	1.60	0.45	0.00
371	piR-hsa-2502273	3.71	0.00	4.82	2.51	0.00	1.95	0.00	0.00	0.00
372	piR-hsa-77303	2.65	1.54	6.02	0.00	1.35	0.00	0.00	0.00	1.39
373	piR-hsa-2853562	0.00	0.00	1.20	8.37	1.35	1.95	0.00	0.00	0.00
374	piR-hsa-2851130	1.59	0.00	0.00	0.00	0.00	0.00	4.81	2.26	4.18
375	piR-hsa-2538984	7.42	0.00	0.00	0.84	1.35	0.00	1.60	0.90	0.70
376	piR-hsa-748358_5	0.00	0.00	0.00	0.00	0.00	0.00	9.62	3.16	0.00
377	piR-hsa-2400882	0.53	0.00	0.00	2.51	5.39	3.89	0.00	0.45	0.00
378	piR-hsa-3610092	5.30	1.54	2.41	0.00	1.35	1.95	0.00	0.00	0.00
379	piR-hsa-727158	5.83	1.54	3.61	0.84	0.00	0.00	0.00	0.00	0.70
380	piR-hsa-1530800	0.00	0.00	0.00	5.86	2.70	3.89	0.00	0.00	0.00
381	piR-hsa-1603046	0.00	0.00	0.00	2.51	4.05	5.84	0.00	0.00	0.00
382	piR-hsa-2518229	0.00	1.54	0.00	0.00	1.35	0.00	3.21	1.81	4.18
383	piR-hsa-3642754	4.77	1.54	4.82	0.84	0.00	0.00	0.00	0.00	0.00
384	piR-hsa-5039329_2	0.00	0.00	0.00	0.00	0.00	0.00	9.62	0.90	1.39
385	piR-hsa-4120912	0.53	0.00	0.00	3.35	5.39	1.95	0.00	0.00	0.70
386	piR-hsa-3263398	5.83	3.08	0.00	0.00	0.00	0.00	0.00	2.26	0.70
387	piR-hsa-2484024	0.00	0.00	0.00	0.84	0.00	0.00	8.02	0.90	2.09
388	piR-hsa-4388882	0.53	1.54	0.00	8.37	1.35	0.00	0.00	0.00	0.00
389	piR-hsa-2499630	0.53	0.00	0.00	0.00	0.00	0.00	8.02	0.90	2.09
390	piR-hsa-2515298	4.24	1.54	0.00	0.00	0.00	0.00	3.21	0.45	2.09
391	piR-hsa-65029	2.12	1.54	4.82	1.67	1.35	0.00	0.00	0.00	0.00
392	piR-hsa-271367	3.71	3.08	2.41	0.84	1.35	0.00	0.00	0.00	0.00
393	piR-hsa-2501382	1.06	0.00	1.20	0.84	0.00	0.00	3.21	2.26	2.79
394	piR-hsa-3345389	0.00	0.00	0.00	3.35	4.05	3.89	0.00	0.00	0.00
395	piR-hsa-1881756	0.00	0.00	0.00	0.00	1.35	0.00	6.42	0.00	3.49
396	piR-hsa-1593686	3.71	3.08	1.20	0.84	0.00	1.95	0.00	0.45	0.00
397	piR-hsa-4359698	2.12	4.62	2.41	0.00	0.00	1.95	0.00	0.00	0.00
398	piR-hsa-1866304	3.18	3.08	4.82	0.00	0.00	0.00	0.00	0.00	0.00
399	piR-hsa-1550295	2.12	7.69	1.20	0.00	0.00	0.00	0.00	0.00	0.00
400	piR-hsa-550050	0.00	0.00	0.00	0.00	0.00	0.00	6.42	3.16	1.39
401	piR-hsa-76694	3.71	0.00	7.23	0.00	0.00	0.00	0.00	0.00	0.00
402	piR-hsa-2491463	4.24	3.08	0.00	0.00	0.00	1.95	0.00	0.90	0.70
403	piR-hsa-2534860	1.06	0.00	0.00	0.00	1.35	3.89	0.00	3.16	1.39
404	piR-hsa-2715002	0.00	0.00	0.00	0.00	0.00	0.00	8.02	1.36	1.39
405	piR-hsa-2511205	0.53	0.00	0.00	0.84	0.00	0.00	3.21	4.07	2.09
406	piR-hsa-4271500	0.53	1.54	0.00	6.69	0.00	1.95	0.00	0.00	0.00
407	- piR-hsa-4150185	1.06	0.00	1.20	8.37	0.00	0.00	0.00	0.00	0.00
408	- piR-hsa-1943369	0.00	0.00	0.00	5.02	5.39	0.00	0.00	0.00	0.00
409	piR-hsa-2447480	1.59	3.08	4.82	0.84	0.00	0.00	0.00	0.00	0.00
410	piR-hsa-1416960	2.65	4.62	1.20	1.67	0.00	0.00	0.00	0.00	0.00
411	piR-hsa-699024	0.00	0.00	1.20	0.00	0.00	0.00	6.42	1.81	0.70
412	- piR-hsa-6764147	0.00	0.00	1.20	0.84	0.00	0.00	0.00	1.81	6.28
413	piR-hsa-644441_3	0.53	0.00	0.00	0.00	0.00	0.00	6.42	3.16	0.00

	piRNA	psc1	psc2	psc3	mpc1	mpc2	mpc3	cpc1	cpc2	cpc3
414	piR-hsa-2500917	0.53	0.00	1.20	0.00	0.00	0.00	4.81	1.36	2.09
415	piR-hsa-1160813	0.00	0.00	0.00	6.69	1.35	1.95	0.00	0.00	0.00
416	piR-hsa-628215	0.00	0.00	0.00	0.00	0.00	0.00	6.42	0.00	3.49
417	piR-hsa-5313694	0.00	0.00	0.00	0.00	0.00	0.00	4.81	4.97	0.00
418	piR-hsa-778446	0.53	0.00	0.00	0.00	0.00	0.00	1.60	1.36	6.28
419	piR-hsa-313568	0.00	0.00	0.00	8.37	1.35	0.00	0.00	0.00	0.00
420	piR-hsa-3153428	2.65	4.62	2.41	0.00	0.00	0.00	0.00	0.00	0.00
421	piR-hsa-2365951	0.00	0.00	0.00	5.86	1.35	1.95	0.00	0.45	0.00
422	piR-hsa-4398177	0.00	0.00	0.00	8.37	0.00	0.00	0.00	0.45	0.70
423	piR-hsa-5884872	1.59	3.08	4.82	0.00	0.00	0.00	0.00	0.00	0.00
424	piR-hsa-851842	3.71	0.00	3.61	0.00	0.00	1.95	0.00	0.00	0.00
425	piR-hsa-4175241	0.53	0.00	1.20	7.53	0.00	0.00	0.00	0.00	0.00
426	piR-hsa-612387	3.71	3.08	2.41	0.00	0.00	0.00	0.00	0.00	0.00
427	piR-hsa-1551388	0.00	0.00	0.00	0.00	0.00	0.00	4.81	2.26	2.09
428	piR-hsa-4049552	0.00	0.00	0.00	0.00	1.35	0.00	3.21	1.81	2.79
429	piR-hsa-42060	1.06	0.00	0.00	6.69	1.35	0.00	0.00	0.00	0.00
430	piR-hsa-338024	3.18	1.54	2.41	0.00	0.00	0.00	0.00	0.45	1.39
431	piR-hsa-2030201	3.71	1.54	3.61	0.00	0.00	0.00	0.00	0.00	0.00
432	piR-hsa-4135859	0.00	0.00	0.00	0.84	0.00	0.00	3.21	4.07	0.70
433	piR-hsa-1692624	0.00	0.00	0.00	6.69	0.00	1.95	0.00	0.00	0.00
434	piR-hsa-1534285	2.12	1.54	4.82	0.00	0.00	0.00	0.00	0.00	0.00
435	piR-hsa-2832794	0.00	0.00	0.00	0.00	0.00	0.00	4.81	2.26	1.39
436	piR-hsa-1370047	0.00	0.00	0.00	0.84	0.00	0.00	0.00	5.87	1.39
437	piR-hsa-632933	0.00	0.00	0.00	0.00	1.35	0.00	1.60	2.26	2.79
438	piR-hsa-2069180	4.24	0.00	1.20	0.84	1.35	0.00	0.00	0.00	0.00
439	piR-hsa-1524373	0.00	0.00	0.00	0.00	0.00	0.00	1.60	0.90	4.88
440	piR-hsa-2487038_7	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.81	5.58
441	piR-hsa-5568482	0.00	0.00	0.00	0.00	0.00	0.00	0.00	3.61	3.49
442	piR-hsa-4346643	0.00	0.00	0.00	0.84	0.00	0.00	0.00	4.07	2.09
443	piR-hsa-7313613	0.53	1.54	0.00	0.00	0.00	0.00	0.00	3.61	0.70
444	piR-hsa-4398796	4.77	0.00	0.00	0.00	0.00	0.00	0.00	1.36	0.00
445	piR-hsa-2369565	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.90	4.88
446	piR-hsa-7098338	0.00	0.00	0.00	0.00	0.00	0.00	1.60	3.16	0.70
447	piR-hsa-7670531_2	0.00	0.00	0.00	0.00	0.00	0.00	0.00	3.61	1.39

Table S1: Related to Figure 3. Normalized counts for the 447 identified piRNAs arranged in descending order expression (row mean). Differentially regulated piRNAs in CPC are indicated in light blue shading (downregulated) and light red shading (upregulated).

cluster1	cluster2	cluster3	cluster4	cluster5
piR-hsa-108574	piR-hsa-1001021	piR-hsa-118348	piR-hsa-114666	piR-hsa-1233052
piR-hsa-132896	piR-hsa-1277994	piR-hsa-1263612	piR-hsa-1389062	piR-hsa-1870459
piR-hsa-137136	piR-hsa-147461	piR-hsa-1291516	piR-hsa-1463989	piR-hsa-1870588
piR-hsa-1399886	piR-hsa-151249	piR-hsa-1332287	piR-hsa-151136	piR-hsa-1872235
piR-hsa-1492262	piR-hsa-1872085_4	piR-hsa-1376916	piR-hsa-163695	piR-hsa-1890632
piR-hsa-1843231	piR-hsa-2482189	piR-hsa-1409954	piR-hsa-1647694	piR-hsa-1900529
piR-hsa-2090890	piR-hsa-2646470	piR-hsa-1416960	piR-hsa-169217	piR-hsa-1919455
piR-hsa-2172087	piR-hsa-611204	piR-hsa-1528884	piR-hsa-1901970	piR-hsa-1927965
piR-hsa-2333057	piR-hsa-76848	piR-hsa-1534285	piR-hsa-1988800	piR-hsa-1941780
piR-hsa-2536290	piR-hsa-1160813	piR-hsa-1550295	piR-hsa-20628	piR-hsa-2477264
piR-hsa-2856604	piR-hsa-1229611	piR-hsa-1555218	piR-hsa-214132	piR-hsa-2479371
piR-hsa-2882083	piR-hsa-1256360	piR-hsa-1593307	piR-hsa-2152778	piR-hsa-2481097
piR-hsa-298158	piR-hsa-1259933	piR-hsa-1593686	piR-hsa-2208850	piR-hsa-2494226
piR-hsa-3021684	piR-hsa-1302552	piR-hsa-1595580	piR-hsa-2286229	piR-hsa-2515454
piR-hsa-3119265	piR-hsa-1303811	piR-hsa-1607096	piR-hsa-2299252	piR-hsa-2530015
piR-hsa-3546008	piR-hsa-1340768	piR-hsa-1632961	piR-hsa-2346976	piR-hsa-2831593
piR-hsa-5077723	piR-hsa-1416366	piR-hsa-163499	piR-hsa-2780538	piR-hsa-2832647
piR-hsa-6913457	piR-hsa-1425899	piR-hsa-1686806	piR-hsa-2844156	piR-hsa-2840936
piR-hsa-721859_9	piR-hsa-1434629	piR-hsa-1726249	piR-hsa-3265318	piR-hsa-346271
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	piR-hsa-1530800	piR-hsa-1798104	piR-hsa-3817390	piR-hsa-368987
	piR-hsa-1531418	piR-hsa-1866304	piR-hsa-3875265	piR-hsa-3735787
	piR-hsa-1557538	piR-hsa-1875155	piR-hsa-3916570	piR-hsa-768321
	piR-hsa-1585146	piR-hsa-1929067	piR-hsa-4078407	piR-hsa-771714_3
	piR-hsa-1603046	piR-hsa-1998991	piR-hsa-4303719	piR-hsa-7892960
	piR-hsa-160969	piR-hsa-2027369	piR-hsa-5982715	
	, piR-hsa-161264 3	, piR-hsa-2030201	, piR-hsa-6245615	
	, piR-hsa-1678085	, piR-hsa-2069180	piR-hsa-783698	
	, piR-hsa-1688824	, piR-hsa-2090264	•	
	piR-hsa-1692624	piR-hsa-2137611		
	piR-hsa-1706026	piR-hsa-2148238		
	piR-hsa-1708978	piR-hsa-2165649		
	, piR-hsa-1756618	, piR-hsa-2213434		
	, piR-hsa-1883893	, piR-hsa-2230204		
	, piR-hsa-1905329	, piR-hsa-2240007		
	, piR-hsa-1909905	, piR-hsa-2248086		
	, piR-hsa-1921551	, piR-hsa-2268195		
	, piR-hsa-1922210	, piR-hsa-2308163		
	, piR-hsa-1933276	, piR-hsa-2319750		
	, piR-hsa-1939085	, piR-hsa-2351941		
	, piR-hsa-1941637	, piR-hsa-2353109		
	, piR-hsa-1943369	, piR-hsa-2398119		
	, piR-hsa-211123	, piR-hsa-2413094		
	piR-hsa-213829	piR-hsa-2426792		
	piR-hsa-2161668	piR-hsa-2427082		
	piR-hsa-21839	piR-hsa-2436454		
	piR-hsa-2209630	piR-hsa-2447480		
	piR-hsa-2220917	piR-hsa-2491463		
	piR-hsa-2253283	piR-hsa-2497478		
	piR-hsa-229786	piR-hsa-2502273		

niR-hsa-2365951	niR_hsa_2503702
pil - 113a - 2303331	piR-113a-2505702
pil-113a-237221	piR-115a-2515250
piR-115a-2400002	piR-115a-2519215
piR-nsa-2423519	piR-nsa-2538984
piR-hsa-2450089_2	piR-hsa-2/136/
piR-hsa-2464166	piR-hsa-2856544
piR-hsa-2489909	piR-hsa-2989729
piR-hsa-2505515	piR-hsa-3104125
piR-hsa-2525461	piR-hsa-3136454
piR-hsa-2537452	piR-hsa-3153428
piR-hsa-2589139	piR-hsa-315964
piR-hsa-2670375	piR-hsa-3177742
, piR-hsa-2853562	, piR-hsa-3231825
piR-hsa-2889978	piR-hsa-3263398
niR-hsa-307961	niR-hsa-3280518
niR-hsa-313568	niR-hsa-333507
piR-113a-315300 piP-bsa-3161050	piR-hsa-3357365
pil - 115a-5101050	pii - 115a - 3337 303
piR-115a-3343369	piR-115a-330024
piR-nsa-3365644	piR-nsa-3513154
piR-nsa-3383102	piR-nsa-3527815
piR-hsa-362913	piR-hsa-354004
piR-hsa-3634880	piR-hsa-3558751
piR-hsa-3693411	piR-hsa-3610092
piR-hsa-3694141	piR-hsa-3638679
piR-hsa-3710717	piR-hsa-3642754
piR-hsa-3714350	piR-hsa-3657078
piR-hsa-374600	piR-hsa-3674332
piR-hsa-3776081	piR-hsa-3732088
piR-hsa-3829948	piR-hsa-3807498
piR-hsa-3839126	piR-hsa-3818788
piR-hsa-3842249	piR-hsa-3944431
piR-hsa-3978322	piR-hsa-3974794
piR-hsa-4111185	piR-hsa-4020841
, piR-hsa-4120912	, piR-hsa-4030155
piR-hsa-4131663	piR-hsa-4053516
niR-hsa-4150185	piR-hsa-4091280
niR-hsa-4157592	piR-hsa-4100164
piR-hsa-/1752/1	niR-hea-/110708
piR 115a 4173241	piR 1150 4110700
piR-113a-4202001	piR-113a-4144200
piR-115a-42000	piR-115a-4175100
piR-fisa-427 1500	piR-fisa-4176299
piR-nsa-4378137_4	piR-nsa-4198101
piR-nsa-4388882	piR-nsa-4322932
piR-hsa-439/384	piR-hsa-4359698
piR-hsa-4398177	piR-hsa-4379982
piR-hsa-4408495	piR-hsa-4398796
piR-hsa-4416099_9	piR-hsa-4403577
piR-hsa-4450044	piR-hsa-4403628
piR-hsa-4467055_8	piR-hsa-508592
piR-hsa-623353	piR-hsa-5124632
piR-hsa-669874	piR-hsa-5411637

piR-hsa-7106256	piR-hsa-58291
piR-hsa-7308134	piR-hsa-5884872
piR-hsa-745484	piR-hsa-612387
piR-hsa-7544198	piR-hsa-645846
piR-hsa-771001	piR-hsa-6482184
piR-hsa-772699	piR-hsa-65029
piR-hsa-7760463	piR-hsa-67957
piR-hsa-778924	piR-hsa-727158
piR-hsa-7833890	piR-hsa-76694
piR-hsa-7893387	piR-hsa-77303
piR-hsa-848451	piR-hsa-8270846
piR-hsa-97458	piR-hsa-851842

cluster6	cluster7	cluster8
piR-hsa-1904126	piR-hsa-1205256	piR-hsa-151466
piR-hsa-1916259	piR-hsa-1429070	piR-hsa-1548068
piR-hsa-359160_2	piR-hsa-1690788	piR-hsa-1875212
piR-hsa-3732777	piR-hsa-1773241	piR-hsa-1876265
piR-hsa-1862211	piR-hsa-1903779	piR-hsa-1883972
piR-hsa-1872463	piR-hsa-2151268	piR-hsa-1908839
piR-hsa-6744266	piR-hsa-2252211	piR-hsa-1919272
	piR-hsa-2395910	piR-hsa-1920687
	piR-hsa-2398570	piR-hsa-1923208
	piR-hsa-2513278	piR-hsa-2490897
	piR-hsa-255984	piR-hsa-2521457
	piR-hsa-2565910	piR-hsa-2526525
	piR-hsa-2615134	piR-hsa-2539762
	piR-hsa-2863156	piR-hsa-316012
	piR-hsa-3232943	piR-hsa-343382
	piR-hsa-3352181	piR-hsa-3634065
	piR-hsa-3683883	piR-hsa-3658275
	piR-hsa-389007	piR-hsa-3658742
	piR-hsa-4028152	piR-hsa-3739406
	piR-hsa-4381848 2	piR-hsa-4460706
	piR-hsa-4472891	piR-hsa-1576285
	piR-hsa-724912	piR-hsa-785969
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	•	, piR-hsa-1242358
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		piR-hsa-144277_2
		piR-hsa-147696
		piR-hsa-1524373
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		piR-hsa-1894768
		piR-hsa-1905680
		piR-hsa-1912443
		piR-hsa-1917013
		piR-hsa-1917139
		piR-hsa-1921188
		piR-hsa-1927627
		piR-hsa-1938524
		piR-hsa-2042088
		•

piR-hsa-2072163 piR-hsa-2281305 piR-hsa-235205 piR-hsa-2369565 piR-hsa-2425220 piR-hsa-2478880\_2 piR-hsa-2484024 piR-hsa-2487038\_7 piR-hsa-2490287 piR-hsa-2490509 piR-hsa-2495779 piR-hsa-2499630 piR-hsa-2500917 piR-hsa-2501382 piR-hsa-2511205 piR-hsa-2518229 piR-hsa-2529368 piR-hsa-2534860 piR-hsa-2542835 piR-hsa-2592846 piR-hsa-2715002 piR-hsa-2742244 piR-hsa-2827579 piR-hsa-2829413 piR-hsa-2829712 piR-hsa-2832439 piR-hsa-2832794 piR-hsa-2839864 piR-hsa-2851130 piR-hsa-343616 piR-hsa-363100\_2 piR-hsa-368381 piR-hsa-3706918 piR-hsa-3718263\_3 piR-hsa-3741185 piR-hsa-4049552 piR-hsa-4135859 piR-hsa-4193743 piR-hsa-4346643 piR-hsa-4387218 piR-hsa-4391981\_9 piR-hsa-4402141 piR-hsa-4403262 piR-hsa-4424378 piR-hsa-4507261 piR-hsa-5039329\_2 piR-hsa-5062317 piR-hsa-5313694 piR-hsa-550050 piR-hsa-5568482 piR-hsa-5996985

piR-hsa-628215 piR-hsa-632933 piR-hsa-642866 piR-hsa-644441\_3 piR-hsa-665910 piR-hsa-6764147 piR-hsa-699024 piR-hsa-7098338 piR-hsa-728085 piR-hsa-7313613 piR-hsa-748358\_5 piR-hsa-753191 piR-hsa-7670531\_2 piR-hsa-778446 piR-hsa-7821967\_3 piR-hsa-8117137 piR-hsa-834074 piR-hsa-942735