

Supplementary Information for

Blocking CHOP-dependent TXNIP shuttling to mitochondria attenuates albuminuria and mitigates kidney injury in nephrotic syndrome

Sun-Ji Park^a, Yeawon Kim^a, Chuang Li^a, Junwoo Suh^b, Jothilingam Sivapackiam^c, Tassia M. Goncalves^d, George Jarad^a, Guoyan Zhao^{d,e}, Fumihiko Urano^f, Vijay Sharma^{c,g,h*}, and Ying Maggie Chen^{a,i*}

^aDivision of Nephrology, Department of Medicine, Washington University School of Medicine, St. Louis, MO 63110

^bDepartment of Biomedical Engineering, Case Western Reserve University, Cleveland, OH, 44106

^cMallinckrodt Institute of Radiology, Washington University School of Medicine, St. Louis, MO 63110

^dDepartment of Neuroscience, Washington University School of Medicine, St. Louis, MO 63110

^eDepartment of Pathology and Immunology, Washington University School of Medicine, St. Louis, MO 63110

^fDivision of Endocrinology, Metabolism, and Lipid Research, Department of Medicine, Washington University School of Medicine, St. Louis, MO 63110

^gDepartment of Neurology, Washington University School of Medicine, St. Louis, MO 63110

^hDepartment of Biomedical Engineering, School of Engineering & Applied Science, Washington University, St. Louis, MO 63105

ⁱDepartment of Cell Biology & Physiology, Washington University School of Medicine, St. Louis, MO 63110

This PDF file includes:

Supplementary methods

Figs. S1 to S6

Table S1

Supplementary Methods

Adriamycin (ADR)-induced nephropathy

Male Balb/cJ mice (stock number 000651 from the Jackson Lab) at 10 weeks of age were injected with ADR in 0.9% saline at 12 mg/kg into the retro-orbital sinus to induce nephropathy.

Kidney mitochondrial fractionation

Kidneys were perfused with cold PBS first. Mitochondrial fractions were isolated from kidneys using Mitochondria Isolation Kit for Tissue (Thermo Fisher Scientific) according to the manufacturer's protocols.

Glomerular lysates

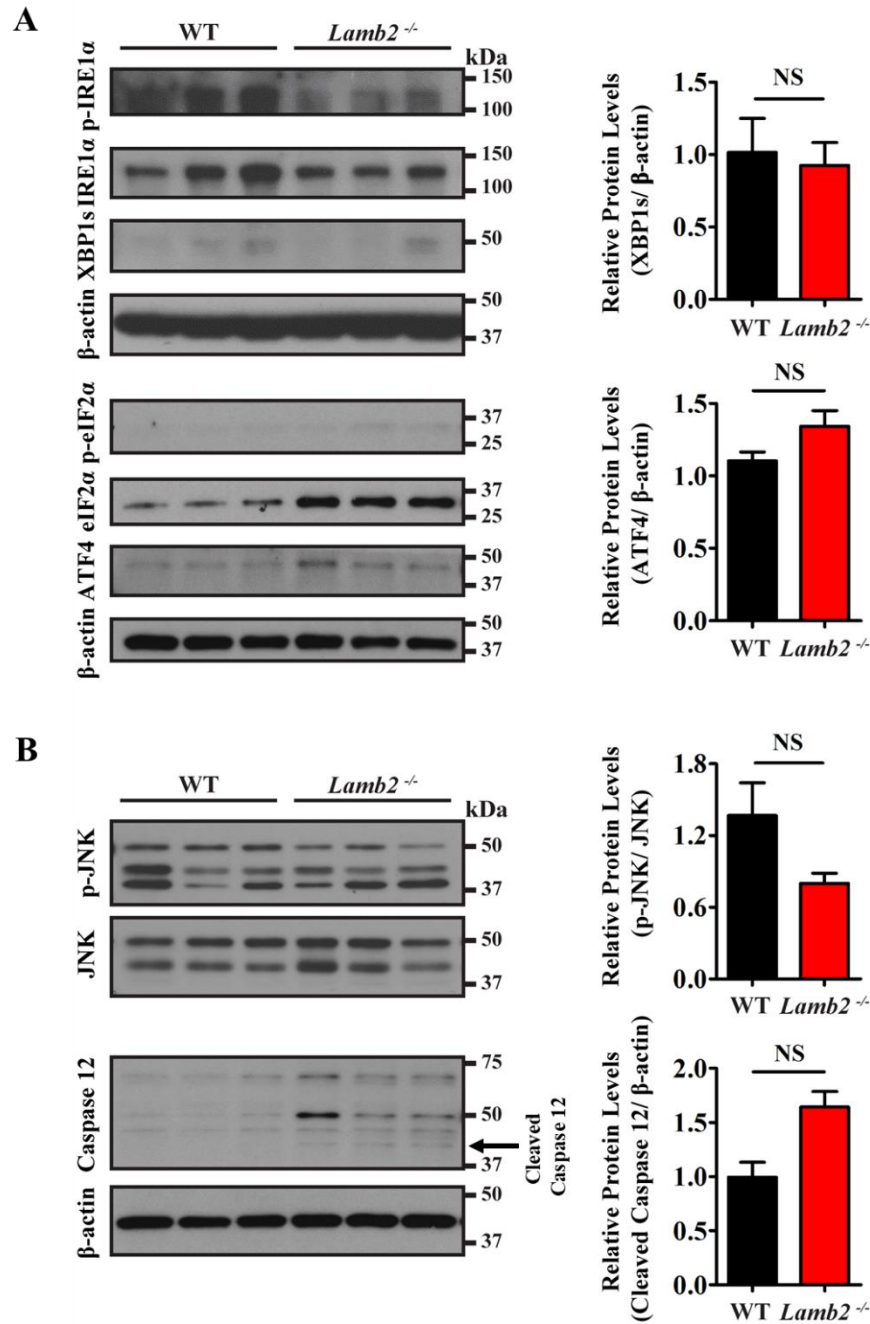


Fig. S1. IRE1 α /XBP1s and eIF2 α /ATF4 pathways, as well as JNK and caspase 12 pathways are not activated in *Lamb2*^{-/-} glomeruli. (A) WB analysis of isolated glomerular lysates from WT and *Lamb2*^{-/-} mice at P25 to detect p-IRE α , IRE1 α , XBP1s, p-eIF2 α , eIF2 α and ATF4. Densitometry analysis of XBP1s and ATF4 normalized to β -actin was performed. (B) WB analysis of glomerular lysates to detect p-JNK, JNK and cleaved caspase 12 (black arrow). Ratios of p-JNK to total JNK and cleaved caspase 12 to β -actin were quantified by densitometry analysis of three independent experiments. Mean \pm SD; NS: not significant by t-test.

Tubular lysates

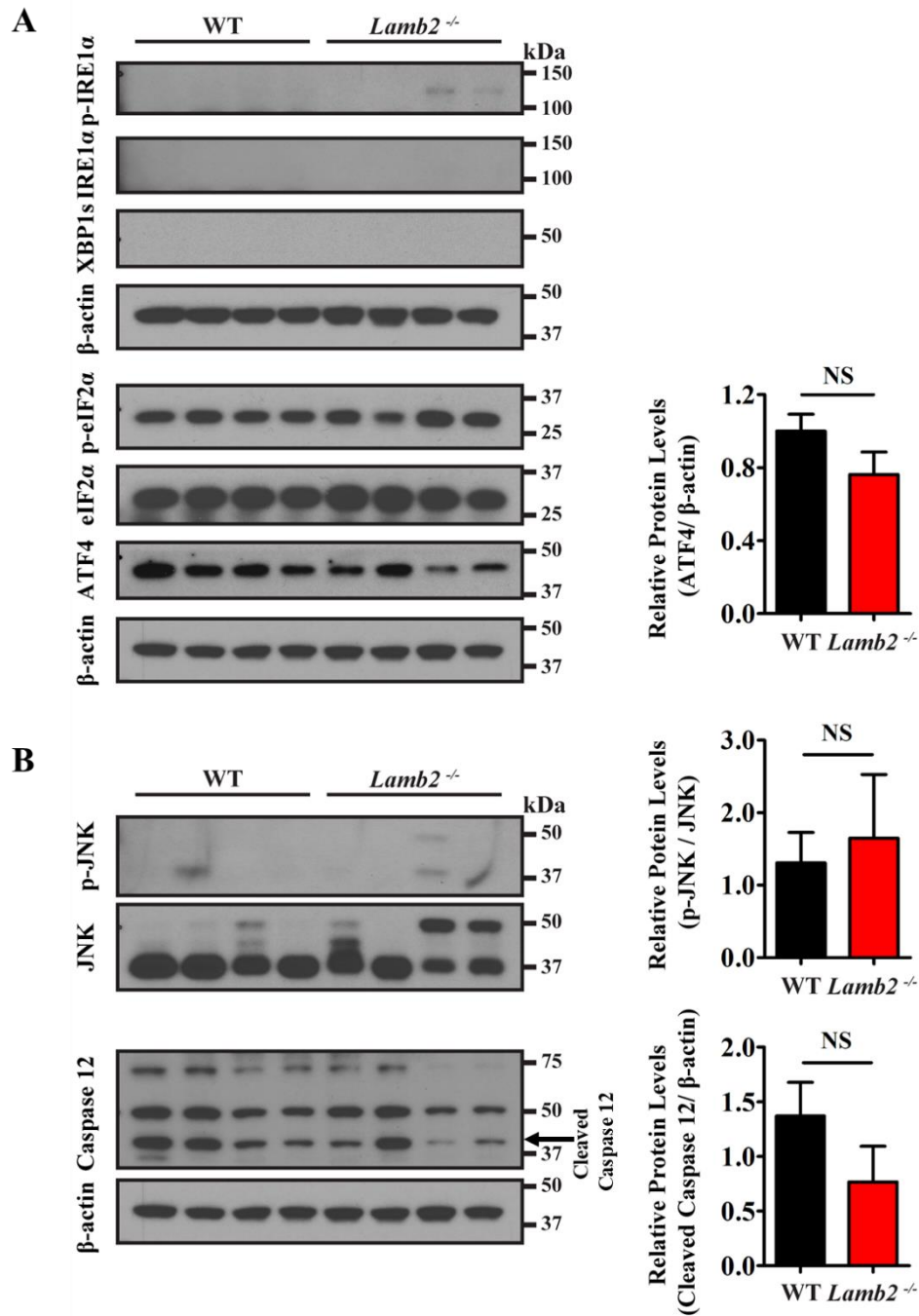


Fig. S2. IRE1 α /XBP1s and eIF2 α /ATF4 pathways, as well as JNK and caspase 12 pathways are not activated in *Lamb2*^{-/-} tubules. (A-B) Isolated tubular lysates from WT and *Lamb2*^{-/-} mice at P25 were analyzed by WB with the indicated antibodies. Ratios of ATF4 to β -actin, p-JNK to total JNK and cleaved caspase 12 (black arrow) to β -actin were quantified by densitometry analysis of three independent experiments. Mean \pm SD; NS: not significant by t-test.

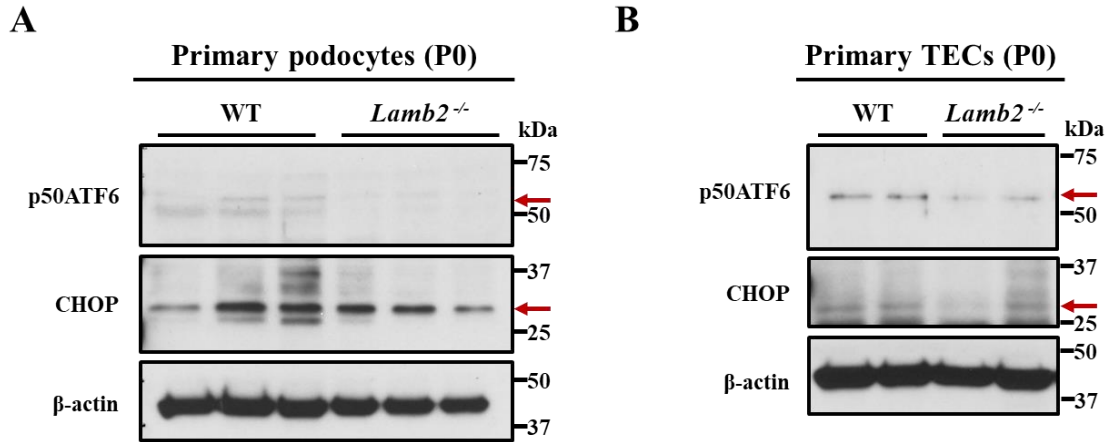


Fig. S3. The ATF6-CHOP signaling is not activated in both primary podocytes and primary TECs isolated from *Lamb2*^{-/-} mice, which are not treated with albumin. Mouse primary podocytes and TECs were isolated and cultured. Representative immunoblots of p50ATF6 (red arrows) and CHOP (red arrows) expression in primary podocytes (P0) (A) and primary TECs (B).

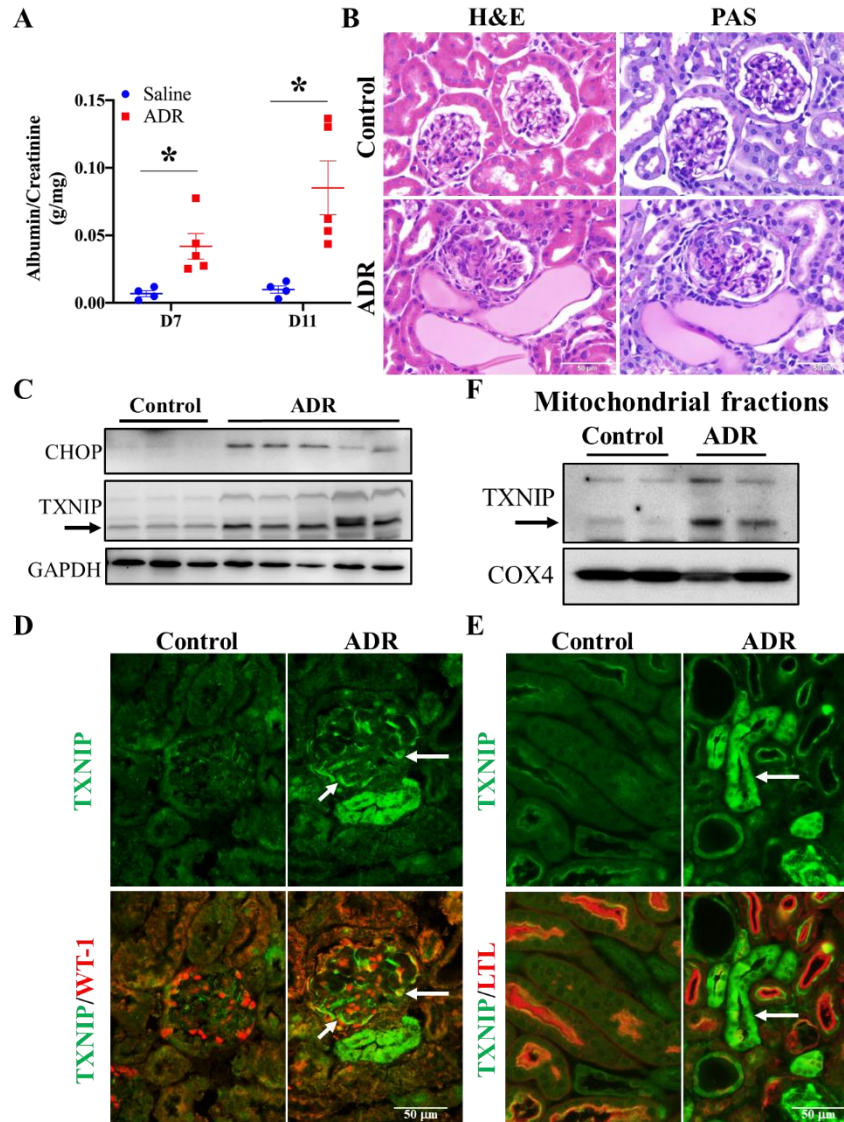


Fig. S4. ADR-induced nephropathy exhibited TXNIP upregulation in both podocytes and tubules, as well as increased mitochondrial accumulation. (A) Urinary albumin/creatinine ratios in saline- and ADR- treated male Balb/cJ mice on day 7 and 11 post-injection. Mean \pm SD (n=4-5 mice/group). * $P < 0.05$. (B) H&E and PAS staining of paraffin kidney sections from control and ADR-injected mice on day 11 post-injection. Scale bar: 50 μ m. (C) Immunoblot to detect CHOP and TXNIP (black arrow) proteins from whole kidney lysates of saline- and ADR-injected mice on day 11 after injection. n=4-5 mice/group. (D-E) IF images of frozen kidney sections stained for TXNIP (green) with WT-1 (red) (D), as well as paraffin sections stained for TXNIP (green) with LTL (red), from saline- and ADR-injected mice on day 11 post-injection. White arrows indicate TXNIP staining in podocytes (D) or in tubules (E). Scale bar: 50 μ m. (F) Representative immunoblots of TXNIP expression (black arrow) in the kidney mitochondrial fractions from control and ADR-injected mice on day 11 post-injection. COX IV was used as mitochondrial internal control.

P35

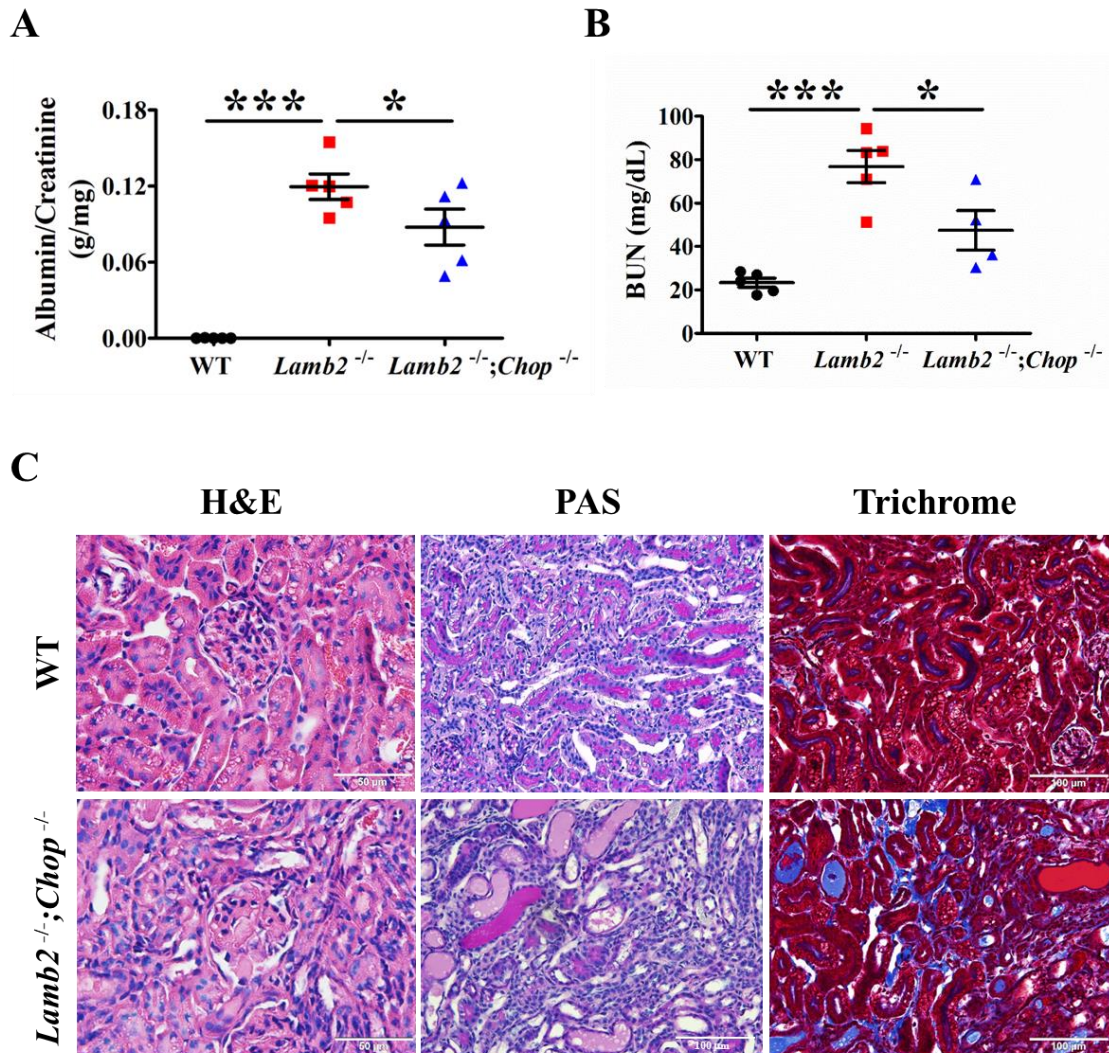


Fig. S5. Characterization of disease phenotype and renal histology in *Lamb2*^{-/-};*Chop*^{-/-} mice at a later stage of the disease (P35). (A) Urinary albumin/creatinine ratios in WT, *Lamb2*^{-/-}, and *Lamb2*^{-/-};*Chop*^{-/-} mice at P35. Mean ± SD (n=5 mice/genotype). **P* < 0.05, ****P* < 0.001. (B) BUN levels from mice of the indicated genotypes at P35. Mean ± SD (n=4-5 mice/genotype). **P* < 0.05, ****P* < 0.001. (C) H&E, PAS, and Gomori's Trichrome staining of paraffin kidney sections from WT and *Lamb2*^{-/-};*Chop*^{-/-} mice at P35. Scale bar: 50 μm for H&E or 100 μm for PAS and Trichrome staining.

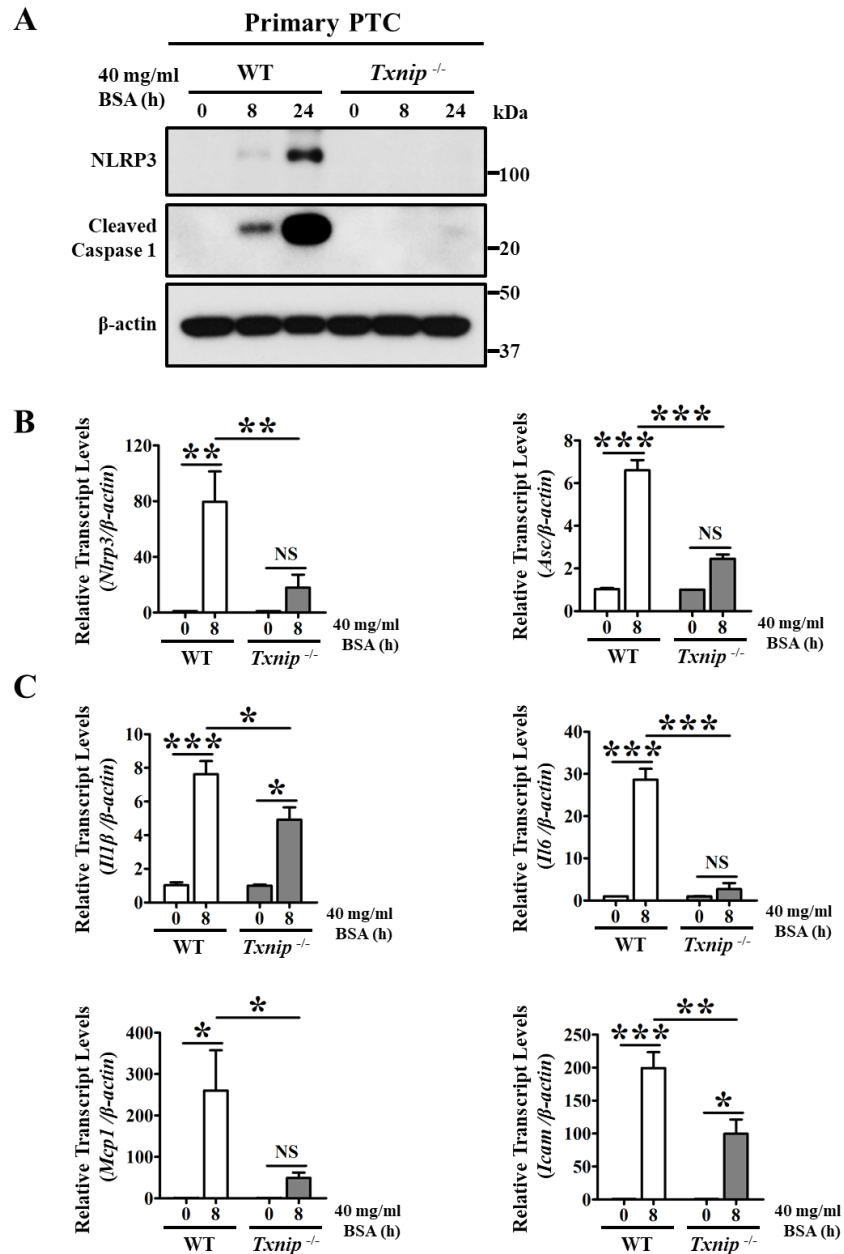


Fig. S6. TXNIP deletion suppresses activation of NLRP3 inflammasome and downstream proinflammatory cytokines in albuminuria. Primary PTCs were isolated and cultured from WT and *Txnip*^{-/-} mice. Cultured PTCs were starved for 16h and then treated with 40 mg/ml BSA for 8 or 24 hours. (A) Representative WBs of NLRP3, cleaved caspase 1 and β -actin in whole cell lysates of primary PTCs in the absence or presence of 40 mg/ml BSA for 8 and 24 hours. The WB image shown is representative of at least three independent experiments. (B-C) Quantitative PCR analysis of primary PTCs treated without or with 40 mg/ml BSA for 8 hours for mRNA levels of NLRP3, ASC (B) and inflammatory cytokines IL1 β , IL6, MCP-1 and ICAM (C). Gene expression was normalized to β -actin. Mean \pm SD from three independent experiments. NS, not significant; * P < 0.05; ** P < 0.01; *** P < 0.001 by ANOVA.

Table S1 List of q-PCR primers

Target gene	Species	Direction	Sequence
<i>Chop</i>	Mouse	Forward	CTGGAAGCCTGGTATGAGGAT
		Reverse	CAGGGTCAAGAGTAGTGAAGGT
<i>Txnip</i>	Mouse	Forward	TATGTACGCCCCTGAGTTCC
		Reverse	GCTCACTGCACGTTGTTGTT
<i>Nlrp3</i>	Mouse	Forward	CCTTCCAGGATCCTCTTCCT
		Reverse	CTTGGGCAGCAGTTTCTTTC
<i>Asc</i>	Mouse	Forward	TGAGCAGCTGCAAACGACTA
		Reverse	ACTTCTGTGACCCTGGCAAT
<i>Il1β</i>	Mouse	Forward	GCACTACAGGCTCCGAGATGAAC
		Reverse	TTGTCGTTGCTTGGTTCTCCTTGT
<i>Il6</i>	Mouse	Forward	TAGTCCTTCCTACCCCAATTTC
		Reverse	TTGGTCCTTAGCCACTCCTTC
<i>Mcp1</i>	Mouse	Forward	TAAAAACCTGGATCGGAACCAA
		Reverse	GCATTAGCTTCAGATTTACGGGT
<i>Icam</i>	Mouse	Forward	AGCACCTCCCCACCTACTTT
		Reverse	AGCTTGCACGACCCTTCTAA
<i>Tgfb</i>	Mouse	Forward	TGGAGCAACATGTGGAACTC
		Reverse	CAGCAGCCGGTTACCAAG