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Fungal Endophytes to Combat Biotic and Abiotic Stresses for Climate-Smart and Sustainable **Agriculture**

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The agricultural sustainability concept considers higher food production combating biotic and abiotic stresses, socio-economic well-being, and environmental conservation. On the contrary, global warming-led climatic changes have appalling consequences on agriculture, generating shifting rainfall patterns, high temperature, CO₂, drought, etc., prompting abiotic stress conditions for plants. Such stresses abandon the plants to thrive, demoting food productivity and ultimately hampering food security. Though environmental issues are natural and cannot be regulated, plants can still be enabled to endure these abnormal abiotic conditions, reinforcing the stress resilience in an eco-friendly fashion by incorporating fungal endophytes. Endophytic fungi are a group of subtle, non-pathogenic microorganisms establishing a mutualistic association with diverse plant species. Their varied association with the host plant under dynamic environments boosts the endogenic tolerance mechanism of the host plant against various stresses via overall modulations of local and systemic mechanisms accompanied by higher antioxidants secretion, ample enough to scavenge Reactive Oxygen Species (ROS) hence, coping over-expression of defensive redox regulatory system of host plant as an aversion to stressed condition. They are also reported to ameliorate plants toward biotic stress mitigation and elevate phytohormone levels forging them worthy enough to be used as biocontrol agents and as biofertilizers against various pathogens, promoting crop improvement and soil improvement, respectively. This review summarizes the present-day conception of the endophytic fungi, their diversity in various crops, and the molecular mechanism behind abiotic and biotic resistance prompting climate-resilient aided sustainable agriculture.

Keywords: abiotic stress, biotic stress, climate-resilience, drought, fungal endophytes, sustainable agriculture

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INTRODUCTION

Sustainable agriculture goals provide a new framework to consider climate change within multiple dimensions of sustainability, considering it as one of the major hindrances in achieving food security for an ever-increasing population. According to IPCC (2014) report, the global temperature may rise from 1.7 to 4.8°C, altering precipitation patterns by the twenty-first century. Abiotic stress driven by climatic changes has reduced global agricultural production by 1 to 5% during the last three decades elevating global food demand from 70 to 100% by 2050 (Newbery et al., 2016; Pais et al., 2020). Climate change has a risk on the ecosystem. Its adaptation is necessary as it imparts short and long-term effects on food production and concurrently displays a significant impact on agriculture, primarily through greenhouse gas emissions (Pais et al., 2020). According to IPCC (2018) report, cumulative emission of carbon dioxide (CO₂) and future non-CO₂ radiative forcing determine the probability of limiting global warming impacts of 1.5°C. The dire effect of climate change can be observed on agricultural productivity affecting overall sustainable agricultural systems. Various abiotic stress conditions, such as high carbon dioxide, drought, high temperature, salinity, etc., hamper food production (Keswani et al., 2019a). Generation of ROS during stress such as singlet oxygen (1O2), hydrogen peroxide (H2O2), superoxide radical (O₂), hydroxyl radical (OH⁻), etc., decreases crop yield (Keswani et al., 2019a). Thus sustainable agriculture needs to be climate-resilient agriculture at the first place. Various modern techniques like breeding, biotechnology, screening techniques, land reformation, etc., should be incorporated altogether, and new possibilities must be explored and given a chance.

On the other hand, over-reliance on synthetic pesticides in the era of the green revolution has anticipated adverse environmental impacts generating pesticide resistance, insect resurgence, effects on non-target organisms, environmental pollution, etc., demanding a lasting conservational approach toward plant protection (Meena et al., 2020; Jacquet et al., 2022). Crops are infested by various pests and pathogens (like fungi, bacteria, viruses, nematodes, etc.), which need a comprehensive antidote to overthrow food production and environmental conservation-related insecurities. Alternative tactics like biological control/biocontrol agents utilizing microorganisms for plant protection seem more reliable and ecosystem-friendly (Adeleke et al., 2022). Critical plant-associated microorganisms like endophytes can play a pivotal role in alleviating the response of crops toward climate change (Yan et al., 2019; Grabka et al., 2022). Fungal endophytes are most studied among other endophytes. Endophytic fungi are an ecological, non-pathogenic, polyphyletic group of highly diverse fungi of ascomycetes, basidiomycetes, and anamorphic fungi (Gupta et al., 2020). They may enter the tissue of their hosts, colonize them, and offer affirmative impacts (Ray et al., 2017). Fungal endophytic colonization induces physiological changes and modifies gene expression in the plants, thereby uplifting plant productivity via higher photosynthesis rate, promoting the shoots and roots growth, enhancing uptake and nutrient use efficiency conferring abiotic (drought, salinity, high temperature, high

CO₂, and metal toxicity) and biotic stress (pathogens and pests) tolerance (Harman and Uphoff, 2019; Grabka et al., 2022; Liu-Xu et al., 2022). Many fungal endophytes have been reported and extracted from healthy plants potent enough to be incorporated artificially as biofertilizers and biocontrol agents in crops (Busby et al., 2016; Bastami et al., 2021; Grabka et al., 2022). Vis-à-vis application of various biofertilizers are also reported to enhance fungal endophyte colonization (Kumar et al., 2021). Endophytic fungi have also been efficient in environmental remediation (agrochemical and metal pollutants solubilization/bio assimilation/mineralization) (Gavrilas et al., 2022). They can also be utilized as a nanosensor for detecting the contaminants, which, if further explored and incorporated, could help in environmental conservation (Sahoo et al., 2017; Khanam et al., 2020). However, environmental remediation is a broad department involving time-consuming processes and demands intensive research to resolve all associated risk factors. Henceforth, the present review aims to explore the role of symbiotic fungal endophytes, their interactions, and the mechanism involved in conferring adaptation to crops against abiotic and biotic stressed conditions, which may contribute toward climate resilience and aiding sustainable agriculture.

ENDOPHYTIC FUNGI: TREATY OF AMITY

Broad-spectrum endophytic fungi follow direct or indirect mechanisms for benefiting the plants. The direct beneficial mechanism involves phytohormone production, nitrogen fixation, phosphate solubilization, siderophore production, and anti-microbial metabolite production (El Enshasy et al., 2020). The indirect beneficial mechanism involves abiotic and biotic resistance (modifying the metabolism process), biocontrol, bioremediation, and phytoremediation (Dubey et al., 2020; Singh et al., 2021). Cosmopolitan fungal endophytes act as biostimulants to produce certain bioactive compounds, phytohormones, phosphate solubilization factors, etc., to enhance root growth, seed germination, and plant growth promotion (Rustamova et al., 2022). Fungal endophytes may potentially execute bounteous ubiquitous roles in the host plant via phytostimulation, phytoimmobilization, phytostabilization, phytotransformation, phytoremediation, biocontrol, etc. (Sahoo et al., 2017; Radziemska et al., 2021; Adeleke et al., 2022; Figure 1). They are reported to produce secondary metabolites which include bioactive anti-microbial siderophores which may execute defense against various pathogens (Srinivas et al., 2020). The endophytes associated with plants produce hydroxamate and carboxylate type siderophores of several classes which play a significant role in transporting iron and zinc to the plants. Siderophores solubilize an array of metals like cadmium, molybdenum, manganese, zinc, nickel, copper, and actinides and are found to mobilize them and make them available to the plants (Rana et al., 2020). To date, more than 500 siderophores have been documented from various fungi (Ahmed and Holmström, 2014; Chowdappa et al., 2020). Recently, many fungal endophytes have been identified which solubilize and mobilize phosphorus (Penicillium, Aspergillus, Curvularia,

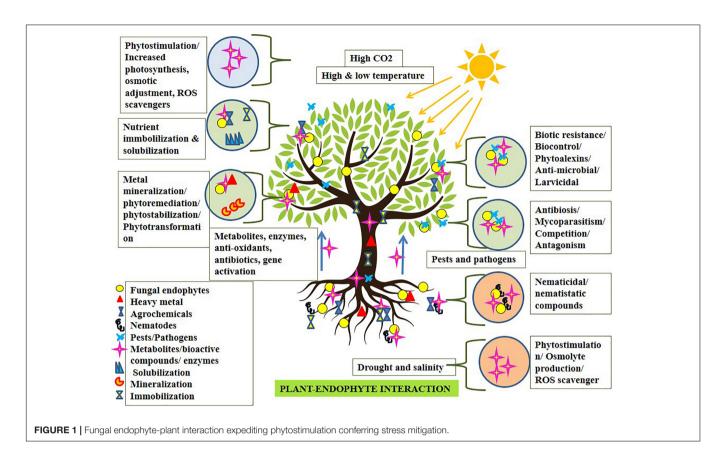
Trichoderma, Mesorhizobium, etc.) (Mehta et al., 2019), potassium (Aspergillus fumigatus and A. niger) (Haro and Benito, 2019), and zinc salts (Alternaria thlaspis and Metapochonia rubescens) (Yung et al., 2021), boosting plant metabolic activity, plant growth resulting in high crop production. They execute phytostimulation via lowering plant hormone ethylene levels by 1-aminocyclopropane-1-carboxylate deaminase (ACC), escalating plant growth (Nascimento et al., 2014; Singh et al., 2015); they may biodegrade biomass and recycle them in the environment, enhancing nitrogen availability, Zinc and phosphorus uptake for the host resulting in phytoimmobilization (Yung et al., 2021). Phytoimmobilization ultimately boosts plants to sustain abiotic stresses via immobilizing osmolytes and stabilizing membrane ion conductivity during stress conditions. Phytotransformation is the neutralization of soil pollutants, for example, pesticides, industrial by-products, xenobiotics, etc. Endophytic fungi like Piriformospora sp., Exophiala sp., Neotyphodium sp., etc., associated with many crop species, are reported to remediate soil from metal pollutants, avoiding phytotoxicity (Ningxiao et al., 2016; Sahoo et al., 2017). However, bioremediation utilizing fungal endophytes demands focused study in the near future incorporating endophyteled nanosensors to unveil its potential in environmental conservation (Khanam et al., 2020). Finally, biofertilizers for plant-growth promotion and biocontrol agents for mitigation of pests and diseases is its other major operation. In this context, culture-independent molecular techniques using metagenomics and multi-locus sequence typing can help isolate and characterize such endophytes (Gupta et al., 2020; Adeleke et al., 2022; Chouhan et al., 2022). Wherein species-specific fungal endophytes identification, isolation, characterization, inoculation, and interaction studies, unraveling their molecular signaling mechanism, and delivering abiotic and biotic stress mitigation, are highly expected. Furthermore, not only isolation but studies related to the shelf-life enhancing components for such fungal biocontrol agents and biofertilizers (Zope et al., 2019) are imperative.

Biodiversity of Fungal Endophytes

The biodiversity of fungal endophytes is vast and reported from almost all plants/crops worldwide (Suryanarayanan et al., 2018; Table 1). According to Rodriguez et al. (2009), fungal endophytes can be classified into four groups based on the parts they colonize and their transmittance- (a) Class I are Clavicipitaceous fungi (resides in grasses with vertical transmittance), (b) Class- II-IV are non-clavicipitaceous fungi (Ascomycota and Basidiomycota), wherein Class-II endophytes inhabits roots, shoots, and rhizomes, and transmit through seeds or rhizomes (i.e., horizontally and vertically both), (c) Class-III inhabits leaves/shoots (transmitted horizontally) and (d) class-IV are typical ascomycetes, forming conidia and resides in roots (transmitted horizontally) of the plants. Class, I endophytes are free-living symbiotic species associated with grasses, rushes, and sedges. Their associations help plants in nematode resistance, anti-fungal compound production, and abiotic stress mitigation, including drought and metal tolerance (Epichloe festucae, Neotyphodium tembladerae, etc.). Class II endophytes increase plant root and shoot biomass and confer tolerance against disease, drought, desiccation heat, and salinity (for example, Mycophycia ascophylli, Phoma sp., Arthrobotrys sp., etc.). Class III includes hyper-diverse fungal endophytes mostly inhabiting above-ground tissues and isn't habitat-specific. They motivate the interaction among various microorganisms' populations (for example, Ustilago maydis, Phyllosticta sp., Colletotrichum sp., etc.) (Chitnis et al., 2020). Class IV endophytes are root-associated sterile ascomycetous fungi forming melanized structures prevalent in high-stress environments worldwide (for example, Chloridium paucisporum, Phialocephala dimorphosphora). Overall, most of the fungal endophytes reported in crops belong to Ascomycota and Basidiomycota (Rodriguez et al., 2009). These endophytes produce secondary metabolites that confer host plants resistance (Table 2). The secondary metabolites produced may be natural like phytohormones or any nutrients or alkaloids, terpenes, quinines, benzopyranones, chinones, phenolic acids, terpenoids, steroids, flavonoids, hydrocarbons, etc., in response to stressed condition (Tiwari, 2015; Bilal et al., 2018; Chouhan et al., 2022); depending upon fungal endophyte and host species (Grabka et al., 2022). Therefore, it is essential to gain in-depth knowledge of their host range, specificity, colonization, transmission, and host-endophyte interaction.

CLIMATE-SMART AGRICULTURE: FACTS AND FAITHS

Climate-smart agriculture (CSA) is directly linked to socioeconomic well-being, sustainable agroecosystem productivity, poverty alleviation, nutritional security, adaptability, and stress mitigation. CSA is anticipated by strategies such as increased resilience toward extreme stressed environments, transforming agriculture and food systems, policy formulations, and awareness promoting ecosystem services (Steenwerth et al., 2014; Fusco et al., 2020; Figure 2). Keeping the above points in mind, beneficial symbiotic fungal endophytes working at a micro-ecosystem level forming rhizosphere offer an immense scope of research for CSA benefaction. Their non-pathogenic nature, universal occurrence, triggered nutrient uptake, mineral solubilization, phytoremediation, enhanced vegetative and reproductive growth, stress tolerance resulting in optimum to high yield, etc., make them suitable for organizing full-fledged scientific research and attention (Galani et al., 2022). Fungal endophytes are reported to increase biomass and remove litter from the soil, increase mineral status and nutrient uptake which aids in achieving CSA concept indirectly (Datta et al., 2017; Santamaria et al., 2017). Whereby, litter is a soil organic matter with a complex structure and a recalcitrant nature that is difficult to degrade wherein, endophytic microbes can be helpful. Fungal endophyte-mediated crop improvement can bring sea changes. However, it suffers from some associated bottlenecks, for example, colonization of a new host plant with a suitable endophyte, their symbiotic association and native endobiome (their direct and indirect effect) (Keswani et al., 2019b). The association is further influenced by their genetics,



gene expression, molecular patterns, behavior alteration, etc. Thus, better insight into the behavior of an introduced endophyte and its beneficial/harmful association with host plant for long term if any, and to what extent is inevitable (Chitnis et al., 2020). Other barriers include economic barriers (high initial cost, high implementation cost, return uncertainty, etc.), institutional and organizational barriers (low support, lack of regulatory framework and policies, etc.), ease of conventional methods, and market barriers (lack of awareness, lack of management and poor market, etc.) impede its overall adoption (Fusco et al., 2020). Consequently, comprehensive and collaborative investments indulging long-term commitment from scientific and researcher communities, farmers, social communities, environmentalists, and policymakers are mandatory to fuel the prolonged CSA concept serving sustainability.

FUNGAL ENDOPHYTES FOR ABIOTIC STRESS MANAGEMENT

Plant growth is greatly hampered by abiotic stresses, and to tolerate it; the plant needs a proficient system or mechanism. Usually, several microbes are found to acquire nutrients from various plants, where some interactions are beneficial and some are harmful to the host. According to Ripa et al. (2019), microbes that inhabit the plant tissues devoid of doing substantive damage or acquiring remuneration other than securing their residency are considered 'endophytes.' Every plant harbors one or a

few endophytes that supply nutrients, enhance environment acclimatization, and confer biotic and abiotic stress tolerance in the host plant. An endophytic fungus lives in inter or intracellular spaces in the stem, petiole, and leaves of a plant for a part or whole of its life cycle. These highly efficient fungal endophytes can ensue for the entire life of the plant, which might also enable the plant to squelch the environmental constraints (Vandenkoornhuyse et al., 2015; Chouhan et al., 2022). Fungal endophytes are crucial for resilience and sustainability in today's agriculture. They offer diverse applications in the agricultural system by collaborating with host plants (Redman et al., 2021; Grabka et al., 2022), as depicted in Figure 3. They possess a vital ability to mobilize insoluble phosphate and provide nitrogen to their host plants boosting vegetative growth sequentially and strengthening associated plants to confer resistance (Hassan, 2017; Yadav and Yadav, 2017). During stress, the fungal endophytes secrete an alleviated amount of plant growth-promoting metabolites; for example, Penicillium sp. inhabiting Suaeda japonica secretes phytohormone Gibberellins. Likewise, Aspergillus sp., Cladosporium sp., Fusarium sp., Penicillium sp., Verticillium sp., and Ascomycete sp. inhabiting Panax ginseng secretes triterpenoid saponins and ginsenosides (enhances root growth and biotic and abiotic stress tolerance) (Sahoo et al., 2017). Keswani et al. (2022) reviewed biosynthesis of giberellins from various fungi imparting beneficial effects on the crop production. Similarly, Phoma sp. inhabiting Tinospora cordifolia secretes growth hormones; Trichoderma atroviride, T. polysporum, and T. harzianum inhabiting Phaseolus

TABLE 1 | Fungal endophytes recently reported in important agronomic and horticultural crops grown worldwide.

Crop	Botanical name	Naturally occurring fungal endophytes	References	
Wheat	Triticum aestivum	Alternaria, Acremonium, Aureobasidium, Cladosporium, Penicillium, Sarocladium, Anthracocystis, Cryptococcus, Sporobolomyces, Vishniacozyma	Rojas et al., 2020	
Apple	Malus domestica	Chaetomium, Epicoccum, Biscogniauxia, Penicillium, Diaporthe, Phlyctema	Liu et al., 2020	
Orchid	Vanda cristata	Fusarium sp.	Srivastava et al., 2018; Chand et al., 2020	
Crucifers	Brassica oleracea, B. rapa and Raphanus sativus	Colletotrichum, Fusarium, Cladosporium, Trichoderma (major) Aspergillus bombycis, Aspergillus nomius, Aspergillus penicillioides, Aspergillus tamarii, Aspergillus westerdijkiae, Cladosporium subuliforme, Cladosporium xanthochromaticum, Trichoderma hamatum, T. harzianum, and Zopfiella (stems) Alternaria alternata, Alternaria burnsii, Phialemoniopsis pluriloculosa, Aspergillus flavipes, Colletotrichum siamense, F. fujikuroi, Gilmaniella subornata, Macrophomina sp., Mycotribulus mirabilis and Penicillium citrinoviride (leaves)	Chen et al., 2020	
Onion	Allium cepa	Clonostachys rosea, Fusarium sp., Hypocrea lixii, Trichoderma asperellum, T. atroviride, T. harzianum	Muvea et al., 2014; Caruso et al., 2020	
Chicory	Cichorium intybus	Cladosporium, Epicoccum, Septoria, Plectosphaerella, Alternaria	Hatamzadeh et al., 2020	
Citrus	Citrus reticulata Alternaria alternate, Alternaria brassicicola, Alternaria carthami, Ascochyta medicaginicola, Aspergillus pallidofulvus, Aureobasidium melanogenum, Cladosporium cladosporioides, Colletotrichum, Diaporthe, Didymella, Fusarium, Meyerozyma, Myrothecium, Neocosmospora, Neosetophoma,		Sadeghi et al., 2019	
		Phaeoacremonium, Pseudozyma, Scedosporium, Talaromyces		
Chives	Allium schoenoprasum	Beauveria brassiana, Penicillum pinophilum	Espinoza et al., 2019	
Tomato	Solanum lycopersicum	Alternaria, Aspergillus, Chaetomium, Curvularia, Fusarium, Hypoxylon, Leptosphaerulina, Meyerozyma, Nigrospora, Penicillium, Periconia, Stemphyllium, Trichoderma	Bogner et al., 2016 Xia et al., 2019	
Chrysanthemum	Dendrobium sp.	Fusarium sp., Colletotrichum sp.	Shah et al., 2018	
Neem	Azadirachta indica	Chloridium, Colletotrichum, Curvularia, Fusarium, Trichoderma, Penicillium, Phyllostica, Phoma, Verticillium, Xylaria	Chutulo and Chalannavar, 2018	
Banana	Musa paradisica	Colletotrichum, Cochliobolus, Fusarium, Lasiodiplodia, Nigrospora, Pestalotiopsis, Phoma, Penicillium	Zakaria and Aziz, 2018	
Maize	Zea mays	Acremonium zeae, Cladosporium oxysporum, Colletotrichum boninense, Colletotrichum gloeosporioides, Coprinopsis cinerea, Curvularia lunata, Epicoccum sorghinum, Fusarium fujikuroi, Gibberella moniliformis, Nemania sp., Penicillium sp., Rigidoporus vinctus, Sarocladium zeae, Scopulariopsis gracilis.	Renuka and Ramanujam, 2016	
Onion	Allium longicuspis	Alternaria sp., A. terreus, Aspergillus ochraceus, Aspergillus versicolor, Aspergillus spectabilis, A. flavus, Fusarium sambucinum	Abdulmyanova et al., 2016	
Common bean	Phaseolus vulgaris	Aureo- Aureobasidium pullulans, Fusarium oxysporum, Xylaria sp., Cladosporium cladosporioides	Parsa et al., 2016	
Periwinkle	Catharanthus roseus	Colletotrichum, Fusarium, Macrophomina, Nigrospora	Ayob and Simarani, 2016	
Lemon grass	Cymbopogon citratus	Colletotrichum, Fusarium, Penicillium, Phoma	Chow and Ting, 2015	
Curry tree	Murraya Koenigii	Colletotrichum, Fusarium, Penicillium, Phoma	Chow and Ting, 2015	
Rapeseed	Brassica napus	Alternaria, Aspergillus, Botrytis, Cryptococcus, Epicoccum, Fusarium, Penicillium, Rhizoctonia, Phoma, Rhizopus, Sporobolomyces, etc.	Zhang et al., 2014	
Tea plant	Camellia sinensis	Colletotrichum gloeosporioides	Rabha et al., 2014	
Wood sorrel	Oxalis corniculata	Aspergillus, Chladosporium, Cunninghamella, Fusarium, Rhizopus	Kurandawad and Lakshman, 2014	

vulgaris secrete proteolytic enzymes, phosphate solubilizing factors, active volatile, and non-volatile metabolites, and *Mucor* sp. inhabiting *Brassica campestris* enhances Indole-3 amino acid (IAA), ACC deaminase and solubilize phosphate secretion resulting in plant growth promotion (Sahoo et al., 2017). Fungal endophytes perform phosphate solubilization,

siderophores production, antibiotic production, phytohormone production, nutrient mineralization, etc., functions for plant growth promotion during stress conditions (Yung et al., 2021). Due to abiotic stress stimuli, abscisic acid, jasmonic acid, and salicylic acid are produced and act as defense signaling substances. Most soil types are insufficient in phosphorus

TABLE 2 | Endophytic fungi and their wide range of bioactive compounds with their properties.

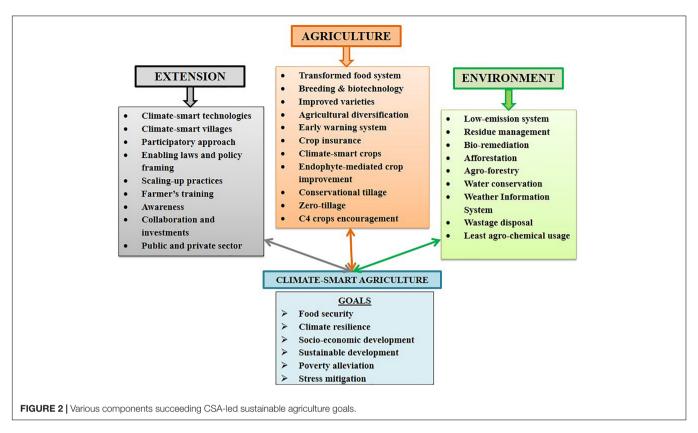
Endophytic fungus/fungi	Bioactive compounds/metabolites	Chemical classes/group	Properties	References	
Alternaria sp.	Altersolanol A (hydroxylated quinone); Altenusin	Quinones; alkaloids	Anti-bacterial	Mousa and Raizada, 2013; Anamika et al., 2018	
Aspergillus sp.	Cyclopeptides echinocandins; indoloditerpenes and asporyzin	Peptides; Sesquiterpenes (Diterpenes)	Anti-fungal; pesticidal activity	Mousa and Raizada, 2013; Anamika et al., 2018	
Pezicula sp.	(R)-mellein; Cyclopeptides echinocandins	Isocaumarin derivative; peptides	Anti-microbial; larvicidal	Anamika et al., 2018	
Neotyphodium sp.	Peramine; agroclavine, chanoclavine and elymoclavine; ergot alkaloids; tricin	Pyrrolopyrazine alkaloid; indole alkaloids; flavanoids	Pesticidal; anti-microbial; larvicidal	Mousa and Raizada, 2013; Anamika et al., 2018	
Penicillium sp.	Volatile organic compounds; Penicisteroid A	VOCs; Steroids	Promotes growth and starch accumulation; anti-fungal property	Mousa and Raizada, 2013; Anamika et al., 2018	
Phyllostica sp.	Hydroheptelidi, heptelidic acid; chlorinated metabolites	Sesquiterpenes; chlorinated compounds	Larvicidal	Anamika et al., 2018	
Phomopsis sp.	Phomopsichalasin; Cycloepoxylactone and cycloepoxytriol B	Amine and amide alkaloids; sesquiterpene	Anti-bacterial; pesticidal activity	Mousa and Raizada, 2013; Anamika et al., 2018	
Acremonium sp.	Leucinostatin A; Peramine (Ioline alkaloids); pyrrocidines A and B	Oligopeptide; amines and amides; polyketide amino acid	Phytotoxic; anti-fungal; anti-feedants; antibiotics	Mousa and Raizada, 2013; Anamika et al., 2018	
Cladosporium sp.	Methyl benzoate; 2-Methoxy-4-hydroxy-6- methoxymethylbenzaldehyde	Volatile compounds; phenols	Anti-microbial; anti-fungal	Anamika et al., 2018	
Colletotrichum sp.	6-isoprenylindole-3-carboxylic acid and steroids; Colletotric acid	Indole alkaloids and steroids; phenol	Anti-fungal, anti-bacterial, antagonistic and pesticidal	Anamika et al., 2018	
Trichoderma viride	Isobutyl alcohol, isopentyl alcohol, 3-methylbutanal, α-bergamotene, bicyclogermacrene, farnesene, geranylacetone, β-sesquiphellandrene, valencence, α-ylangene and zingiberene	Volatile organic compounds	Promotes plant growth, development and flowering	Anamika et al., 2018	
Fusarium sp.	Diterpene subglutinol; β-caryophyllene	Terpenoids (diterpenes); VOCS	Anti-microbial; growth promoter	Anamika et al., 2018	
Beauveria sp.	Chitinases, lipases and proteases; virulence factor	Enzymes and virulence factor	Pesticidal property	Anamika et al., 2018	
Phoma sp.	1-N-methylalbonoursin; Altersolanol A; 2-hydroxy-6-methylbenzoic acid; Phomadecalin C	Amine and amide alkaloids; Quinone; phenol; sesquiterpenes	Anti-microbial; antagonistic effect	Anamika et al., 2018	
Ampelomyces sp.	Altersonalol A	Tetrahydroanthraquinones	Biocontrol agent of parasitic fungus	Mousa and Raizada, 2013	
Trichoderma harzianum	Trichodermin	Sesquiterpenes	PGR synthesis, anti-fungal and inhibitory	Mousa and Raizada, 2013	
Taxomyces andreanae	Paclitaxel (Taxol)	Sesquiterpenes (Diterpenes)	Anti-fungal property	Mousa and Raizada, 2013	
<i>Xylaria</i> sp.	Eremophilane sesquiterpene; sordarin	Pheromones; Sesquiterpenes (Diterpenes)	Anti-fungal; phytotoxic	Mousa and Raizada, 2013	

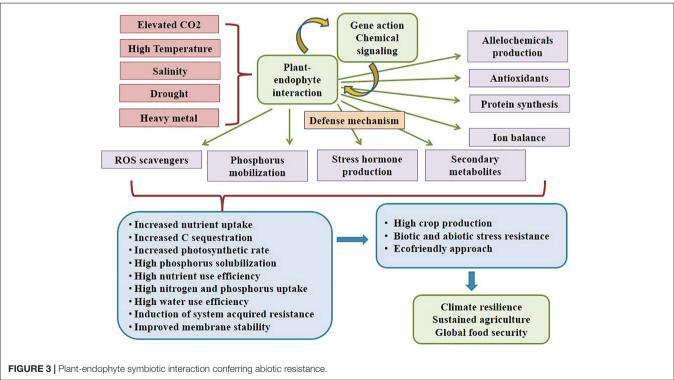
content, where P is considered an essential element for plant growth. They remain insoluble and unavailable to plant even when applied as an inorganic fertilizer to the soil; hence, aluminum and iron hydroxides commence their fixation in acidic soil. Iron having redox activity behaves as a cofactor of many enzymes and is crucial as a micronutrient but remains in insoluble (ferric hydroxide) form in the soil (Johnson, 2008). Fungal endophytes produce siderophores with low molecular weight and a high affinity for iron (Fe3+), thereby increasing

iron uptake and solubilization of phosphates and their uptake by the plants, promoting growth. Endophytes biodegrade the litter and enhance biomass production and nutrient uptake (Idbella et al., 2019).

High Carbon Di-Oxide

Endophytic fungi increase internal hyphal growth and colonization, carbon sequestration, biomass, and yield *via* increased photosynthesis and induced compositional changes





in elevated CO₂ (Galani et al., 2022). In this context, various researchers found symbiotic relationship beneficial as it escalates plant growth in crops viz. tomato, rice, beans, cereals,

potato, etc., secreting ascorbate and glutathione reducing Reactive Oxygen Species (ROS), following antioxidants and allelochemicals production (Yadav and Yadav, 2017). This

renders efficient nutrient uptake, photosynthetic rate, water use efficiency (Elhindi et al., 2016), elevating phytohormone production, amplifying gene expression, supplementing protein function, metabolite synthesis, signaling molecules secretion, and triggering stress-responsive pathways (Ma et al., 2019), for better survival without hampering the yield. However, some contrast results revealing decreased root and shoot growth exaggerated that endophytes vary in their effects on the same host plants subjected to the specific niche, preferentially under a particular growth condition (Gavito et al., 2000). Furthermore, colonization is also affected by the type of plants; for example, C3 plants consume additional carbon for biomass production and thus are less likely to be benefited from colonization, whilst C4 plants allocate more carbohydrates resulting in a profit from endophytic fungi association (Tang et al., 2009). Therefore, according to the present studies, the role of microbiota in biological and ecological processes remains a challenge (Rehman et al., 2016), especially for possible application in agriculture as endophytes adopt a molecular mechanism to induce stressresponsive genes, anti-stress metabolites, and Reactive Oxygen Species (ROS) scavenger molecules synthesis.

Drought

Drought is one of the significant stresses diminishing agricultural productivity and directly challenging food security attainment goals in developing countries. A range of physiological and biochemical responses are triggered in the plants to respond to drought conditions, such as stomatal closure, increased respiration, suppression of growth, and photosynthesis. According to researchers, two groups of drought stress-inducible genes function to confer plant tolerance, the first being the proteins and the second regulatory proteins (involved in signal transduction) (Shinozaki and Yamaguchi-Shinozaki, 2007). Fungal endophyte *Piriformospora indica* (family- Sebacinaceae) colonizes many plant roots, and its hyphae are present in the outer layer of the host's roots aiding in plant growth, higher chlorophyll content, and elevated yield under drought stress conditions; such fungal endophytes led drought stress tolerance in wheat at diverse temperature regimes has been studied by Ripa et al. (2019). The fungal genus studied for plant growth parameters and stress tolerance was Fusarium sp., Trichoderma sp., Penicillium sp., Aspergillus sp., Alternaria sp., etc. Most of them were found to confer drought resistance to wheat. Endophytic fungus-associated plants (rice, tomato, and dune grass) have used significantly less water and increased biomass than non-symbiotic plants. The drought tolerance phenomenon may be explained by an enhanced accumulation of solutes in tissues of endophyte-infected plants compared to non-infected plants, by reduced leaf conductance and a slowdown of the transpiration stream, or due to thicker cuticle formation (Ripa et al., 2019). Characterization of Trichoderma sp. inhabiting Theobroma cacao revealed changes in gene expression patterns, implying that Trichoderma sp. could induce tolerance to abiotic stresses, possibly including drought (Bailey et al., 2006). Drought, heat, and salt stress affect the plantwater relationship, triggering complex plant responses, including increased osmolytes production. According to Marquez et al.

(2007), the osmotic potential is determined primarily by solute potential and matrix potential; symbiotic fungi are likely to contribute to the matrix potential, which is particularly important in helping plants retain water and enhancing plant drought tolerance. Accordingly, the symbiotic fungi-associated plants consume significantly less water than non-symbiotic plants (Marquez et al., 2007).

Salinity

Salinity is another significant abiotic stress limiting the growth and productivity of plants worldwide. Tolerance of plants to salt stress is associated with the alleviation of antioxidant enzymes, i.e., ROS scavengers including glutathione, ascorbate, and tocopherol, and the enzymes superoxide dismutases (SOD), catalases (CAT), ascorbate- or thiol-dependent peroxidases (APX), glutathione reductases (GR), dehydroascorbate reductases (DHAR) and mono-dehydroascorbate reductases (MDHAR) (Rouhier et al., 2008). They are involved in removing ROS directly (SOD, CAT, and APX) or indirectly via regenerating ascorbate and glutathione in the cell. For instance, Piriformospora indica, a root fungal endophyte, upraised biomass production in salt stress conditions by intensifying antioxidative capacity, fatty acid composition, lipid peroxidase, dehydroascorbate reductase, catalase, and glutathione reductase enzymes accumulation in barley (Baltruschat et al., 2008). In another instance, the endophytic fungus Exopiala sp. LHL08 isolated from cucumber root appeared to confer tolerance against salinity and drought stress in rice (Murphy et al., 2018). Similarly, P. indica and S. vermifera showed salinity tolerance with increased growth and yield, and strong effects were observed by combining biological agents (Franken, 2012). Endophytic fungi Fusarium culmorum (FcRed1) was found to have a symbiotic relationship with rice seedlings under salinity and drought, resulting in increased plant height, root length, biomass, and higher osmolyte accumulation than the non-symbiotic rice seedlings. Nonetheless, there is a great need to unveil how fungal endophytes stimulate plant defense response against abiotic stress (Egamberdieva et al., 2017; Ma et al., 2019).

High Temperature

Endophytic fungi may mediate the effect of high temperature by adjusting, regulating, or modifying plant physiological, biochemical, and metabolic activity depending on their biology, developmental stage, and cultivar of the host plant. Environmental conditions that affect the endophytic community may also escalate in the number and variety of endophytic populations influencing metabolite production. researchers have assayed several isolated fungal endophytes, their multifarious interactions beneficial for the host against high-temperature stress environments, and their precise roles in environment acclimatization and conferring tolerance (Lata et al., 2018). For instance, a study found Curvularia protuberate, a fungal endophyte, confer thermo-tolerance to tomato and panic grass. Paraphaeosphaeri aquadriseptata produces a heat shock protein 90 (HSP90) inhibitor conferring heat tolerance to Arabidopsis sp. When infected by a virus, as exemplified in a study of the potential of Stenotrophomonas maltophilia SBP-9 to promote the growth of wheat under salt stress plant, the plant may confer heat tolerance (Marquez et al., 2007; Yadav and Yadav, 2017). Thus, a critical evaluation of a more intriguing and inexplicable issue with many fungal endophytes to produce host metabolites needed to be harnessed for potential use in a diverse environment (Khare et al., 2018; Galani et al., 2022).

Heavy Metal Stress

Heavy metal pollution is of great concern in developing countries where industrial wastes and pollutants are directly dumped into rivers or left open on soil surfaces. Cultivable areas may get affected by the application of excessive agrochemicals and polluted water as the direct passage from waterways. Accumulation of such heavy metals in the plants through the soil and irrigated water, hampering their growth and giving direct entry into the food chain, may pose health hazards to humans and animals; hence, bioremediation is crucial. In this context, plant endophyte symbiotic interactions enhance antioxidant enzyme activity via uptake of heavy metal (cadmium) from soil followed by detoxification and transport led by three genes, namely ZYP, PCS, and MTP (Ma et al., 2016; Wang et al., 2016). Various fungal endophytes (Penicillium sp., Mucor sp., Alternaria sp., Microsphaeropsis sp., etc.) have ameliorated the adverse effects of heavy metal stress in plants. Fungal endophytes are found to be involved in the biosorption of cadmium (Xiao et al., 2010), copper (Li et al., 2012), and sodium chloride (Khan et al., 2013) by stabilizing the negative effect of heavy metal toxicity (Khan et al., 2013). Zahoor et al. (2017) isolated and screened some innate endophytes and cultured them on heavy metal (chromium, cobalt, copper, manganese, and zinc) contaminated soils and found Mucor sp. (strain MHR-7) to confer a multiple heavy metal toxicity tolerance. Recently, the endophyte mediated biosensor concept is also receiving much attention, promising a better bioremediation-controlled process (Khanam et al., 2020). Some endophytes induce phytohormone production, especially gibberellins, which contribute to fungus in eradicating of heavy metal toxicity, thus serving an ecofriendly approach to bioremediation benefiting a healthy diet environment.

Modulation of Plant Defense Mechanism Against Abiotic Stress

It has been hypothesized that fungal microbiota composition alters when plants experience environmental stress, which is determined by the plant genotype (Andreo-Jimenez et al., 2019). Consequently, it takes complementary microbial and plant genes together to establish a symbiotic genetic pathway as a consequence of molecules produced by endophytes that function as signaling molecules within plants, referred to collectively as "symbiont-associated molecular patterns" (SAMPs) (Harman and Uphoff, 2019). Earlier Schulz et al. (1999) proposed the "balanced antagonism" hypothesis in biotic stress resistance, which propounded a balance between plant defensive responsive genes, endophytes, and their toxic effects on each other. Since all the interactions between endophytes and plants are mediated *via* chemical signaling led by gene action, it is essential to understand the mechanisms of how these endophyte function

in plants and intensifies the defense system (Chagas et al., 2018; Etalo et al., 2018; Yan et al., 2019). As per the mechanism studied, the fungal spore germ tube initially is triggered by 5deoxy-strigol under stress, resulting in the branching of hyphae which later spreads and results in colonization accompanied by signaling molecules that lead to induction of plant defense genes triggering phytohormones accumulation. Chemoattractants like flavonoids are the metabolites that assist in the interaction of endophytes with root hair resulting in effective colonization (Khare et al., 2018). Researchers have found signaling molecules such as nod factor, strigolactone, arabinogalactan proteins, plant cell wall proteins, lower levels of ROS (Ca²⁺, NO₂, and NO), systemin and inositol phosphates, etc., to stimulate symbiotic signaling pathways and delayed induction of jasmonic acid, ethylene, and salicylic acid signaling pathway (Plett and Martin, 2018; Rozpadek et al., 2018). According to the studies, drought condition exerts osmotic stress, whereas salinity stress exerts both ionic and osmotic stress, which is counteracted by the accumulation of abscisic acid (Lata et al., 2018). Mitogenactivated protein kinase (MAPK) and heat shock proteins (HSP) play a crucial role in drought and heat stress, respectively (Lata et al., 2018). Modulation of the plant immune system by endophytes and how they target hormonal response pathways via miRNAs has been reviewed in detail by Khare et al. (2018). Plant triggers comparatively transient defense reaction to abiotic stresses generating ROS (Keswani et al., 2019a), and in turn, endophytes produce enzymes for their defense, aiding in modulating and intensifying the plant defense system (Mane et al., 2018). Intensive research on fungal endophytes and how exactly they get influenced by altered climatic conditions prompting the production of such allelochemicals that ultimately trigger the defense mechanism and how this affects plant performance is a requisite.

FUNGAL ENDOPHYTES FOR BIOTIC STRESS MANAGEMENT

Pesticides/agrochemicals resistance development in pests and pathogens demands more sustainable solutions, preferably pesticide environmental stewardship under biocontrol agents (Baron and Rigobelo, 2021). In this regard, non-pathogenic fungal endophytes isolates have demonstrated their ability to help other plants withstand subsequent pathogen attacks (Mane et al., 2018; Grabka et al., 2022). They usher upregulation of bioactive molecules, phenolic compounds, antioxidants, phytoalexins, phytohormone production, ecological occupancy, etc. They are responsible for safeguarding crops from biotic stress (Singh et al., 2021). Aspergillus sp., Colletotrichum sp., Gliocladium sp., Fusarium sp., Petriella sp., Piriformospora sp., Trichoderma sp., *Epicoccum* sp., *Epichloe* sp., etc., are a few of the major endophytes launching biotic stress resistance, while boosting growth and yield components of the host plant synchronously (Table 3). These major fungal endophytes can potentially accommodate crops in an eco-friendly manner worthy of being treated as biocontrol agents (Adeleke et al., 2022). Epichloe sp. offers biotic resistance to its host plant and controls herbivorous

TABLE 3 | Recently reported fungal endophytes mediated biotic stress management in agricultural crops worldwide.

Crop	Endophytic fungi/Biocontrol agent	Disease/Causal organism	Bioactive compounds	Activity	References
Soybean	Trichoderma longibrachiatum, T. asperellum, and T. atroviride	Root rot/Rhizoctonia solani	Pectinase, chitinase, siderophore, IAA	Disease severity controlled by 55–65%	Sallam et al., 2021
Musk melon	Trichoderma phayaoense	Gummy stem blight, Wilt/Stagonosporopsis cucurbitacearum and Fusarium equiseti	Parasitism and anti-fungal compound	81–91% mycelial growth inhibition	Nuangmek et al., 2021
Wheat	Sarocladium strictum, Anthracocystis floculossa, Anthracocystis floculossa, and Penicillium olsonii	Fusarium head blight (FHB)/ <i>F. graminearum</i>	Not studied	70% reduction in FHB	Rojas et al., 2020
Melons	Trichoderma harzianum, Trichoderma lentiforme, Epicoccum purpurascens	Rots, collapse, and Wilt/Macrophomina phaseolina, Monosporascus cannonballus, Fusarium sp.	Microbial antagonist, secondary compounds	42–93% inhibition in radial growth	Gonzalez et al., 2020
Tomato	Beauveria bassiana	Whitefly/Bemisia tabaci	Chitin and β-glucans	Non-prefernce of inocultaed plant	Wei et al., 2020
Cucumber	Penicillium sp. and Hypocrea sp.	Wilt/Fusarium oxysporum f. sp. cucumerinum	Anti-fungal agents	Antagonistic effect and 66% mycelial inhibition	Abro et al., 2019
Ginseng	Trichoderma citrinoviride	Botrytis cinerea, and Cylindrocarpon destructans	b-1,3- glucan-, b-1,4- glucan-, and b-glucoside-degrading enzymes	Mycoparasitism, antibiosis and inhibitory effects	Park et al., 2019
Cotton	Beauveria bassiana	Cotton leafworm larvae/Spodoptera littoralis	Beauvericin	57% larval death	Sánchez-Rodríguez et al., 2018
Tomato	Beauveria bassiana	Fusarium oxysporum f. sp. lycopersici race 3	Lytic enzyme	72% growth inhibition	Culebro-Ricaldi et al., 2017
Chinese Ginseng	Trichoderma gamsii	Scytalidium lignicola, Fusarium flocciferum, Phoma herbarum, and Epicocum nigrum	Volatile organic compound (dimethyl disulfide, dibenzofuran, methanethiol, ketones)	Hyperosmolar, mycoparasitism and antagonistic activity	Chen et al., 2016
Maize	Cladosporium oxysporum and Rigidoporus vinctus	Maize stem borer/Chilo partellus	mycotoxin (fumonisin)	10% mortality rate	Renuka and Ramanujam, 2016
Wheat	Trichoderma hamatum, Penicillium, and Paecilomyces lilacinus	Tan spot/ <i>Pyrenophora</i> tritici-repentis	Not studied	Growth suppression	Larran et al., 2016
Black pepper	Ceriporia lacerata Annulohypoxylon nitens Daldinia eschscholzii, Diaporthe sp., Phomopsis sp., Fusarium sp.	Rot fungus/Phytophthora capsici	Antibiosis, mycoparasitism and non-volatile metabolites	Mycoparasitic activity	Sreeja et al., 2016
Tomato	Beauveria bassiana	Fruit borer/Helicoverpa armigera	Not studied	Insecticidal activity	Qayyum et al., 2015

insects (Laihonen et al., 2022). Piriformospora indica, a root colonizing filamentous fungus belonging to the Basidiomycota class, reside in association with xerophytic plants specifically, found to have a broad host range including most angiosperms viz. wheat, rice, maize, barley, tomato, chili (Arora et al., 2022), etc. It counter-attacks pest/pathogen by accelerating the production of phytoalexins, anti-microbial compounds, activation of antioxidant defense enzyme (glutathione-ascorbate) system, phytohormone production, etc., resistance (Waller et al., 2005). Majorly three counteractions against pests/pathogens are reported, (a) antibiosis (antibiotic production), (b) competition for nutrients (c) mycoparasitism. Wilts, rots, mildews, leaf

mosaics, nematodes, blights, etc., can efficiently be controlled by *P. indica*; therefore, they are worthy of being considered a potential biocontrol agent (Waller et al., 2005; Ali et al., 2019). An anamorphic filamentous saprophytic fungus, *Trichoderma* sp. colonizes roots, twigs, and stems of host plants serving as potential biocontrol agents producing anti-fungal, anti-bacterial, and cytotoxic effects. Moreover, they grow naturally in different habitats in varied climatic conditions and easily adapt to edaphic conditions. *Trichoderma* sp. is a universal soil resident and inhabits roots and root hairs of various plants, where it secrets extracellular and intracellular metabolites with cytotoxic effects against fungal pathogens (Srivastava et al., 2012, 2014). Various

fungal isolates like T. harzianum, T. koningii, T. atroviride, T. virens, T. reesei, and T. citrinoviride are reported, and their pathogenesis against pathogens has been demonstrated. Strong counter-attack activity against potent pests proves its suitable candidature for bio-control agents (Singh et al., 2021). Few of its strains are already being used as biofertilizers, reducing chemical fertilizer usage on farms. Additionally, Trichoderma sp. can produce phytohormones (indole-3-acetic acid (IAA) or auxin analogs, abscisic acid, and gibberellin), siderophores, and other plant growth-promoting substances (e.g., 6-pentylαpyrone, cyclonerodiol, harzianolide, and koninginins) enriching soil microflora and aiding soil fertility as well (El Enshasy et al., 2020; Keswani et al., 2020). A new strain, Trichoderma phayaoense, has recently been discovered with parasitic and anti-fungal properties against fungal pathogens with 91% mycelial reduction (Nuangmek et al., 2021). T. botryosum, T. caeruloviride, T. lentissimum, and T. pseudopyramidale isolated from wild coffee in Africa were reported to render biological control via mycoparasitism (Rodríguez et al., 2021). Highly efficient Trichoderma sp. have been recently cultured via various methods that produce high xylanase amounts (Ambehabati et al., 2020). Another soil-borne ascomycete fungal endophyte with antagonist property residing in many host plants is Myrothecium verrucaria. It is considered a natural mycoherbicide due to its high phytotoxin and lytic enzyme production (Clarke et al., 2007). Purpureocillium lilacinum (formerly Paecilomyces lilacinus) fungus can colonize root knot nematode (particularly Meloidogyne incognita) and destroys the females, cysts and eggs (Sharma et al., 2020). Similarly, Aureubasidium pullulans and Candida orthopsilosis isolated from teak plant can fight against Alternaria sp. Causing post harvest disease in citrus (Sukmawati et al., 2021). Generally, antagonistic activities of fungal endophytes are coupled with the synthesis of bioactive defense-related compounds as well as bioactive natural substances (Rustamova et al., 2022), such as antimicrobial, anti-fungal, and anti-viral metabolites, for instance, perfumoid, phomoenamide, dimethyl disulfide, dibenzofuran, methanethiol, ketones, b-1,3- glucan-, b-1,4- glucan-, and b-glucoside-degrading enzymes, Chitin, and β-glucans, β-1, 3 glucanases, chitinase, cellulose, protease, hydrolyzing enzymes, fumonisin, Beauvericin, etc., that contribute to control of pathogens (see Table 2). Many shreds of evidence strongly supporting the potentiality of such fungal endophytes with broad-spectrum pathogenesis and host adaptability mitigating biotic stress in many agriculturally important crops have already been published. Many of them can be utilized as safe, ecofriendly, and effective biocontrol agents for different crop species.

FUTURE PROSPECTS AND CHALLENGES

In place of escalated pollution, indiscriminate use of agrochemicals, population explosion, depleting fertile land, and food imbalances, adjusted researchers' attention toward an alternative eco-friendly and sustainable solution for food production. Endophytic fungi gained much interest in

agriculture, aiding plant growth processes under mutualistic relationships and mitigating various stresses. Various researchers are constantly studying the rhizosphere of diverse crops to understand the colonial pattern, interactions, and associated benefits. This has led to new fungal endophytes discovery with a broad range of host alliances in extreme environmental conditions or habitats. Applying endophytic fungi as biocontrol agents or biofertilizers, either alone or as a component of integrated pest management packages, may prove a remedial alternative against agrochemical application dependency. The establishment of consortia constituting potential strains of endophytes from local areas of agricultural farms producing complex biocontrol products can be the first farmer-friendly and eco-friendly step toward sustainability. Research and development associated with novel endophyte or endophytic consortiums associated with diverse crops and habitats can play a significant and lasting role in sustenance. Scientific research should focus on genetically modified endophytes by combining two endophytes performing different ecological roles. Optimization of microbial functions to enhance crop production and protection is also required. Detailed fungal community and endomicrobiome study utilizing biotechnological tools (omics) unveiling endophyte-host interaction and molecular patterns. Advance investigation on general and specific host fungal endophytes is required to develop a thriving bio-inoculum leading on the way to organic food crops for a better tomorrow. Hence, bioprospecting studies to isolate such endophytic symbiont fungi and screening them to be directly used in the field is of great importance. Their incorporation into cropping systems through seed treatments, foliar applications, or other means may improve agricultural efficiency and, at the same time, achieve positive environmental impacts (Harman and Uphoff, 2019; Omomowo and Babalola, 2019). The prime challenge is to comprehend its adverse effects on the host plant, such as the transmission of toxins in the food chain and if endophytes start acting as latent pathogens when inoculated artificially on another crop. What if they reach edible portions like fruits, tubers, storage roots, forage, etc.? Secondly, knowledge of the interplay between newly introduced endophyte and native plant endobiome. Its importance can be understood by the fact that endophytes produce antibiotics only in the presence of other endophytes and not in the pure culture (Schulz et al., 2015). Contrarily, some introduced endophytes might eliminate natural endophytes while colonization or form disease on the new host; this propels researchers to scrutinize endophytes and their pathogenesis both beforehand their release as biofertilizers or biocontrol agents (Keswani et al., 2019b). Lack of substantial information about field performance of introduced endophyte and alteration in functioning owing to climatic changes are yet other related aspects to work on. The potential barriers limiting large endophyte scale commercial incorporation are the poor understanding of general and specific plant-endophyte interactions, physio-chemical barriers suppressing the plant immune response stimulation, pathogen-plant immune defense system, gene action, and signaling pathway in agriculture (Ma et al., 2016, 2019). Finally, crop breeding programs have not considered endophytic biodiversity (a breeding objective).

There is no report of any marker designed/synthesized favoring endophyte association with crops. Finally, obtaining registration of the bio-formulated or bioagent products before their market placement is virtually a daunting task/challenge. The licensing laws and its regulatory bodies, framing rules and regulations to register and commercialize any product preceding release in the environment, are a time-consuming process. Nevertheless, such regulations ensure in-depth explorative and comprehensive research and understanding to steer clear of ecological concerns eventually. Eminent scientists from different fields are contributing and hopefully succeeding in generating novel solutions for fungal endophytes inoculations/formulations at a commercial scale, ultimately facilitating climate-smart agriculture-led sustainable agriculture.

CONCLUSION

It is concluded that crops naturally accommodate diverse communities of symbiotic fungal endophytes, which provide various benefits, including abiotic and biotic resistance leading to high food production. Fungal endophytes are a potentially vital source of metabolites or bioactive compounds for sustained agriculture with significant potential to combat crop pathogens

REFERENCES

- Abdulmyanova, L. I., Fayzieva, F. K., Ruzieva, D. M., Rasulova, G. A., Sattarova, R. S., and Gulyamova, T. G. (2016). Bioactivity of fungal endophytes associated with *Allium* plants growing in Uzbekistan. *Int. J. Curr. Microbiol. Appl. Sci.* 5, 769–778. doi: 10.20546/ijcmas.2016.509.088
- Abro, M. A., Sun, X., Li, X., Jatoi, G. H., and Guo, L. D. (2019). Biocontrol potential of fungal endophytes against *Fusarium oxysporum* f. sp. *cucumerinum* causing wilt in cucumber. *Plant Pathol. J.* 35, 598–608. doi: 10.5423/PPJ.OA.05.2019. 0129
- Adeleke, B. S., Ayilara, M. S., Akinola, S. A., et al. (2022). Biocontrol mechanisms of endophytic fungi. *Egypt J. Biol. Pest Control.* 32, :46. doi: 10.1186/s41938-022-00547.1
- Ahmed, E., and Holmström, S. J. (2014). Siderophores in environmental research: roles and applications. *Microbial biotechnology*. 7, 196–208. doi: 10.1111/1751-7915.12117
- Ali, A., Bilal, S., Khan, A. L., Mabood, F., Al-Harrasi, A., and Lee, I. J. (2019). Endophytic Aureobasidium pullulans BSS6 assisted developments in phytoremediation potentials of Cucumis sativus under Cd and Pb stress. Journal. of Plant Int. 14, 303–313. doi: 10.1080/17429145.2019.1633428
- Ambehabati, K. K., Hanapi, S. Z., El Baz, A. F., Sayyed, R. Z., Dailin, D. J., and El Enshasy, H. A. (2020). Isolation and identification studies on potential xylanaseproducing strain *Trichoderma* sp. WICC F46 Isolated from Trop. *Soil. J. Scient. Indust. Res.* 79, 153–159.
- Anamika, R., Joshi, S., Sahgal, M., Sahu, S., and Prakash, A. (2018). "Fungal endophytes and their secondary metabolites: role in sustainable agriculture," in Fungi and their Role in Sustainable Development: Current Perspectives, eds P. Gehlot and J. Singh (Singapore: Springer), doi: 10.1007/978-981-13-0393-7_8
- Andreo-Jimenez, B., Vandenkoornhuyse, P., Le Van, A., Heutinck, A., Duhamel, M., Kadam, N., et al. (2019). Plant host and drought shape the root associated fungal microbiota in rice. *Peer J.* 7, :7463. doi: 10.7717/peerj.7463
- Arora, H., Sharma, A., Poczai, P., Sharma, S., Haron, F. F., Gafur, A., et al. (2022). Plant-Derived protectants in combating soil-borne fungal infections in tomato and chilli. *J. Fungi.* 8, :213. jof8020213 doi: 10.3390/
- Ayob, F. W., and Simarani, K. (2016). Endophytic filamentous fungi from a Catharanthus roseus: identification and its hydrolytic enzymes. Saudi Pharm. J. 24, 273–278. doi: 10.1016/j.jsps.2016.04.019

that are becoming increasingly resistant to pesticides. Some of these endophytes can't be cultured and hence need a more advanced approach to harness the benefits *via* artificial inoculation/bioagents in cultivable crops. However, we need to discover efficient endophytes for climate-resilient and sustainable agriculture to understand their ecological roles and microbial interactions better. Overall fungal endophytes have vast potential to deal with changing climate and plant protection in an ecofriendly manner.

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HJ contributed to conceptualization and supervision. AV and NS contributed to writing. JP, ES, PP, and RS contributed to writing – review and editing. PP contributed to fund acquisition. All authors contributed to the article and approved the submitted version.

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- Bailey, B. A., Bae, H., Strem, M. D., Roberts, D. P., Thomas, S. E., Samuels, G. J., et al. (2006). Fungal and plant gene expression during the colonization of cacao seedlings by endophytic isolates of four *Trichoderma* species. *Planta* 224, 1449–1464. doi: 10.2135/cropsci2000.404923x
- Baltruschat, H., Fodor, J., Harrach, B. D., Niemczyk, E., Barna, B., Gullner, G., et al. (2008). Salt tolerance of barley induced by root endophyte *Piriformospora indica* is associated with a strong increase in antioxidants. *New Phytol.* 180, 501–510. doi: 10.1111/j.1469-8137.2008.02583.x
- Baron, N. C., and Rigobelo, E. C. (2021). Endophytic fungi: a tool for plant growth promotion and sustainable agriculture. *MycologyMycol.* 13, 39–55. doi: 10.1080/ 21501203.2021.1945699
- Bastami, A., Amirnia, R., Sayyed, R. Z., and Enshasy, H. A. E. (2021). The effect of mycorrhizal fungi and organic fertilizers on quantitative and qualitative traits of two important *Satureja Species*. *Agronomy*. 11, :1285. doi: 10.3390/ agronomy11071285
- Bilal, L., Asaf, S., Hamayun, M., Gul, H., Iqbal, A., Ullah, I., et al. (2018). Plant growth-promoting endophytic fungi Aspergillus fumigatus TS1 and Fusarium proliferatum BRL1 produce gibberellins and regulate plant endogenous hormones. Symbiosis 97, 117–1271–11. doi: 10.1007/s13199-018-0545-4
- Bogner, C. W., Kariuki, G. M., Elashry, A., Sichtermann, G., Buch, A.-K., Mishra, B., et al. (2016). Fungal root endophytes of tomato from Kenya and their nematode biocontrol potential. *Mycol. Progress.* 15:30. doi: 10.1007/s11557-016-1169-9
- Busby, P. E., Ridout, M., and Newcombe, G. (2016). Fungal endophytes, modifiers of plant disease. *Plant Mol. Biol.* 90, 645–655. doi: 10.1007/s11103-015-0412-0
- Caruso, G., Golubkina, N., Tallarita, A., Abdelhamid, M. T., and Sekara, A. (2020). Biodiversity, ecology, and secondary metabolites production of endophytic fungi associated with Amaryllidaceae crops. *Agriculture* 10, :533. doi: 10.3390/ agriculture10110533
- Chagas, F. O., Pessotti, R. D., Rodriguez, A. M. C., and Pupo, M. T. (2018). Chemical signaling is involved in plant-microbe interactions. *Chem. Soc. Rev.* 47, 1652–1704. doi: 10.1039/c7cs00343a
- Chand, K., Shah, S., Sharma, J., Paudel, M. R., and Pant, B. (2020). Isolation, characterization, and plant growth-promoting activities of endophytic fungi from a wild orchid *Vanda cristata*. *Plant Signal Behav*. 15,: 1744294. doi: 10. 1080/15592324.2020.1744294
- Chen, J. L., Sun, S. Z., Miao, C. P., Wu, K., Chen, Y. W., Xu, L. H., et al. (2016). Endophytic *Trichoderma gamsii* YIM PH30019: A promising biocontrol agent

- with hyperosmolar, mycoparasitism, and antagonistic activities of induced volatile organic compounds on root-rot pathogenic fungi of *Panax notoginseng*. *J. Ginseng Res.* 40, 315–324. doi: 10.1016/j.jgr.2015.09.006
- Chen, J., Akutse, K. S., Saqib, H. S. A., Wu, X., Yang, F., Xia, X., et al. (2020). Fungal endophyte communities of crucifer crops are seasonally dynamic and structured by plant identity, plant tissue and environmental factors. Front. Microbiol. 11:1519. doi: 10.3389/fmicb.2020.01519
- Chitnis, V. R., Suryanarayanan, T. S., Nataraja, K. N., Prasad, S. R., Oelmüller, R., and Shaanker, R. U. (2020). Fungal endophyte-mediated crop improvement: The way ahead. Front. Plant Sci. 11:561007. doi: 10.3389/fpls.2020.561007
- Chouhan, S., Agrawal, L., and Prakash, A. (2022). Amelioration in traditional farming system by exploring the different plant growth-promoting attributes of endophytes for sustainable agriculture. Arch. Microbiol. 204, :15. doi: 10.1007/ s00203-021-02637-4
- Chow, Y., and Ting, A. S. (2015). Endophytic L-asparaginase-producing fungi from plants are associated with anticancer properties. J. Adv. Res. 6, 869–876. doi: 10.1016/j.jare.2014.07.005
- Chowdappa, S., Jagannath, S., Konappa, N., Udayashankar, A. C., and Jogaiah, S. (2020). Detection and Characterization of Antibacterial Siderophores Secreted by Endophytic Fungi from *Cymbidium aloifolium*. *Biomolecules*. 10,:1412. doi: 10.3390/biom10101412
- Chutulo, E. C., and Chalannavar, R. K. (2018). Endophytic mycoflora and their bioactive compounds from *Azadirachta indica*: A comprehensive review. *J. Fungi.* 4, :42. doi: 10.3390/jof4020042
- Clarke, T. C., Shetty, K. G., Jayachandran, K., and Norland, M. R. (2007). Myrothecium verrucaria– a potential biological control agent for the invasive 'old world climbing fern' (*Lygodium microphyllum*). BioControl 52, 349–411.
- Culebro-Ricaldi, J. M., Ruíz-Valdiviezo, V. M., Rodríguez-Mendiola, M. A., Ávila-Miranda, M. E., Gutiérrez-Miceli, F. A., et al. (2017). Antifungal properties of *Beauveria bassiana* strains against *Fusarium oxysporum* f. sp. *lycopersic*i race 3 in tomato crop. *J. Environ. Biol.* 38, 821–827.
- Datta, R., Kelkar, A., Baraniya, D., Molaei, A., Moulick, A., et al. (2017). Enzymatic degradation of lignin in soil: A review. Sustainability. 9, :1163. doi: 10.3390/ su9071163
- Dubey, A., Malla, M. A., Kumar, A., Dayanandan, S., and Khan, M. L. (2020). Plants endophytes: unveiling hidden agenda for bioprospecting toward sustainable agriculture. Crit. Rev. Biotechnol. 40, 1210–1231. doi: 10.1080/07388551.2020. 1808584
- Egamberdieva, D., Wirth, S., Alqarawi, A., Abd Allah, E. F., and Hashem, A. (2017).
 Phytohormones and beneficial microbes. Essential components for plants to balance stress and fitness. Front. Microbio. 8, :2104. doi: 10.3389/fmicb.2017.
 02104
- El Enshasy, H., Ambehabati, K., El Baz, A., Ramchuran, S. O., Sayyed, R., et al. (2020). "Trichoderma: biocontrol agents for promoting plant growth and soil health," in Agriculturally Important Fungi for Sustainable Agriculture. Functional Annotation for Crop Protection, Vol. 2, eds A. Yadav, S. Mishra, D. Kour, N. Yadav, and A. Kumar (Cham: Springer), doi: 10.1007/978-3-030-48474-3.8
- Elhindi, K., Hendawy, S. E., and Salam, E. M. A. (2016). Foliar application of potassium nitrate affects the growth and photosynthesis in coriander (*Coriander sativum* L.) plants under salinity. *Prog. Nutr.* 18, 63–73. https://mattiolihealth.com/progress-in-nutrition/.
- Espinoza, F., Vidal, S., Rautenbach, F., Lewu, F., and Nchu, F. (2019). Effects of *Beauveria bassiana* (Hypocreales) on plant growth and secondary metabolites of extracts of hydroponically cultivated chive (*Allium schoenoprasum L.* [Amaryllidaceae]). *Heliyon* 5, :12. doi: 10.1016/j.heliyon.2019.e03038
- Etalo, D., Jeon, J. S., and Raaijmakers, J. M. (2018). Modulation of plant chemistry by beneficial root microbiota. *Nat. Prod. Rep.* 35, 398–409. doi: 10.1039/ c7np00057i
- Franken, P. (2012). The plant strengthening root endophyte *Piriformospora indica*, potential application, and the biology behind it. *Appl. Microbiol. Biotech.* 96, 1455–1464. doi: 10.1007/s00253-012-4506-1
- Fusco, G., Melgiovanni, M., Porrini, D., and Ricciardo, T. M. (2020). How to Improve the Diffusion of Climate-Smart Agriculture: What the Literature Tells us. SustainabilitySustain. 12,:5168. doi: 10.3390/su12125168
- Galani, Y. J. H., Hansen, E. M. O., Droutsas, I., et al. (2022). Effects of combined abiotic stresses on nutrient content of European wheat and implications for

- nutritional security under climate change. Scientific. Reports.. 12, :5700. doi: 10.1038/s41598-022-09538-6
- Gavito, M. E., Curtis, P. S., Mikkelsen, T. N., and Jakobsen, I. (2000). Atmospheric CO₂ and mycorrhiza effects on biomass allocation and nutrient uptake of nodulated pea (*Pisum sativum L.*) plants. *J. Exp. Bot.* 51, 1931–1938. doi: 10. 1093/jexbot/51.352.1931
- Gavrilaş, S., Ursachi, C. Ş, Perţa-Crişan, S., and Munteanu, F.-D. (2022). Recent trends in biosensors for environmental quality monitoring. Sensors 22, :1513. doi: 10.3390/s22041513
- Gonzalez, V., Armijos, E., and Garcés-Claver, A. (2020). Fungal endophytes as biocontrol agents against the main soil-borne diseases of melon and watermelon in spain. AgronomyAgron. 10,:820. doi: 10.3390/agronomy10060820
- Grabka, R., d'Entremont, T. W., Adams, S. J., Walker, A. K., Tanney, J. B., Abbasi, P. A., et al. (2022). Fungal endophytes and their role in agricultural plant protection against pests and pathogens. *Plants* 11, :384. doi: 10.3390/ plants11030384
- Gupta, S., Chaturvedi, P., Kulkarni, M. G., and Staden, J. V. (2020). A critical review on exploiting the pharmaceutical potential of plant endophytic fungi. *Biotechnol. Adv.* 39, :107462. doi: 10.1016/j.biotechadv.2019.107462
- Harman, G. E., and Uphoff, N. (2019). Symbiotic root-endophytic soil microbes improve crop productivity and provide environmental benefits. *Scientifica* 201925, 1–25. doi: 10.1155/2019/9106395
- Haro, R., and Benito, B. (2019). The Role of Soil Fungi in K⁺ Plant Nutrition. *International. Journal. of Molecular. Sciences.*. 20,:3169. doi: 10.3390/ iims20133169
- Hassan, S. E. D. (2017). Plant growth-promoting activities for bacterial and fungal endophytes isolated from medicinal plant of *Teucrium polium L. J. Adv. Res.* 8, 687–695. doi: 10.1016/j.jare.2017.09.001
- Hatamzadeh, S., Rahnama, K., Nasrollahnejad, S., Fotouhifar, K. B., Hemmati, K., White, J. F., et al. (2020). Isolation and identification of L-asparaginase-producing endophytic fungi from the Asteraceae family plant species of Iran. *Peer. J.* 8, :e8309. doi: 10.7717/peerj.8309
- Idbella, M., Zotti, M., Cesarano, G., Fechtali, T., Mazzoleni, S., and Bonanomi, G. (2019). Fungal endophytes affect plant response to leaf litter with contrasting chemical traits. *Community. Ecology.*. 20, 205–213. doi: 10.1556/168.2019.20.2.
- IPCC (2014). "Climate change 2014: mitigation of climate change," in Contribution of Working Group III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change, eds O. Edenhofer, R. Pichs-Madruga, Y. Sokona, E. Farahani, S. Kadner, K. Seyboth, et al. (Cambridge: Cambridge University Press).
- IPCC (2018). "Global warming of 1.5°C," in An IPCC Special Report on the Impacts of Global Warming of 1.5°C Above Pre-industrial Levels and Related Global Greenhouse Gas Emission Pathways, in the Context of Strengthening the Global Response to the Threat of Climate Change, Sustainable Development, and Efforts to Eradicate Poverty, eds V. Masson-Delmotte, P. Zhai, H.-O. Pörtner, D. Roberts, J. Skea, P. R. Shukla, et al. (Geneva: Intergovernmental Panel on Climate Change).
- Jacquet, F., Jeuffroy, M. H., Jouan, J., et al. (2022). Pesticide-free agriculture as a new paradigm for research. Agronomy. Sustainability. Development. 42, :8. doi: 10.1007/s13593-021-00742-8
- Johnson, L. J. (2008). Iron and siderophores in fungal-host interactions. Mycological. Research.. 112, 170–183. doi: 10.1016/j.mycres.2007.11.012
- Keswani, C., Dilnashin, H., Birla, H., and Singh, S. (2019a). Unravelling efficient applications of agriculturally important microorganisms for alleviation of induced inter-cellular oxidative stress in crops. Acta Agriculturae. Slovenica 114:121. doi: 10.14720/aas.2019.114.1.14
- Keswani, C., Prakash, O., Bharti, N., Vílchez, J. I., et al. (2019b). Re-addressing the biosafety issues of plant growth promoting rhizobacteria. Science. of Total Environment.. 690, 841–852. doi: 10.1016/j.scitotenv.2019.07.046
- Keswani, C., Singh, S. P., Cueto, L., et al. (2020). Auxins of microbial origin and their use in agriculture. Applied. Microbiology. and Biotechnology. 104, 8549–8565. doi: 10.1007/s00253-020-10890-8
- Keswani, C., Singh, S. P., García-Estrada, C., Mezaache-Aichour, S., Glare, T. R., Borriss, R., et al. (2022). Biosynthesis and beneficial effects of microbial gibberellins on crops for sustainable agriculture. *Journal. of Applied. Microbiology.*, 132, 1597–1615. doi: 10.1111/jam.15348

- Khan, A. L., Waqas, M., Khan, A. R., Hussain, J., Kang, S. M., et al. (2013). Fungal endophyte *Penicillium janthinellum* LK5 improves growth of ABA-deficient tomato under salinity. *World J. Microbiol. Biotechnol.* 29, 2133–2144. doi: 10.1007/s11274-013-1378-1
- Khanam, Z., Gupta, S., and Verma, A. (2020). Endophytic fungi-based biosensors for environmental contaminants- A perspective. S. Afr. J. Bot. 132, 401–406. doi: 10.1016/j.sajb.2020.08.007
- Khare, E., Mishra, J., and Arora, N. K. (2018). Multifaceted interactions between endophytes and plants, development and prospects. Front. Microbiol. 9:2732. doi: 10.3389/fmicb.2018.02732
- Kumar, S., Meena, R. S., Singh, R. K., Munir, T. M., Datta, R., et al. (2021). Soil microbial and nutrient dynamics under different sowings environment of Indian mustard (*Brassica juncea* L.) in rice based cropping system. *Scientific. Reports.* 11, :5289. doi: 10.1038/s41598-021-84742-4
- Kurandawad, J. M., and Lakshman, H. C. (2014). Diversity of the endophytic fungi isolated from *Oxalis Corniculata* Linn. *Int. J. Scient. Res.* 3, 393–397.
- Laihonen, M., Saikkonen, K., Helander, M., Vázquez, de Aldana, B. R., et al. (2022). Epichloë Endophyte-Promoted Seed Pathogen Increases Host Grass Resistance Against Insect Herbivory. Frontiers. in Microbiology.. 12:786619. doi: 10.3389/ fmicb.2021.786619
- Larran, S., Simon, M. R., Moreno, M. V., Siurana, M. P. S., and Perello, A. (2016). Endophytes from wheat as biocontrol agents against tan spot disease. *Biol. Cont.* 92, 17–23. doi: 10.1016/j.biocontrol.2015.09.002
- Lata, R., Chowdhury, S., Gond, S., and White, J. F. (2018). Induction of abiotic stress tolerance in plants by endophytic microbes. *Lett. Appl. Microbiol.* 66, 268–276. doi: 10.1111/lam.12855
- Li, H. Y., Da-Qiao, W., Mi, S., and Zuo-Ping, Z. (2012). Endophytes and their role in phytoremediation. Fungal. Div. 54, 11–18. doi: 10.1007/s13225-012-0165-x
- Liu, J., Ridgway, H. J., and Jones, E. E. (2020). Apple endophyte community is shaped by tissue type, cultivar and site and has members with biocontrol potential against *Neonectria ditissima*. J. Appl. Microbiol. 128, 1735–1753. doi: 10.1111/jam.14587
- Liu-Xu, L., Vicedo, B., García-Agustín, P., and Llorens, E. (2022). Advances in endophytic fungi research: a data analysis of 25 years of achievements and challenges. J. Plant Interact. 17, 244–266. doi: 10.1080/17429145.2022.2032429
- Ma, Y., Miroslav, V., and Freitas, H. (2019). Beneficial microbes alleviate climatic stresses in plants. Front. Plant Sci. 10,: 595. doi: 10.3389/fpls.2019.00595
- Ma, Y., Rajkumar, M., Zhang, C., and Freitas, H. (2016). Beneficial role of bacterial endophytes in heavy metal phytoremediation. *J. Environ. Manag.* 174, 14–25. doi: 10.1016/j.jenvman.2016.02.047
- Mane, R. S., Paarakh, P. B., and Vedamurthy, A. B. (2018). Brief review on fungal endophytes. Int. J. Second. Metab. 5, 288–303. doi: 10.21448/ijsm.482798
- Marquez, L. M., Redman, R. S., Rodriguez, R. J., and Roossinck, M. J. (2007).
 A virus in a fungus in a plant three-way symbiosis required for thermal tolerance. *ScienceSci.* 315, 513–515. doi: 10.1126/science.1136237
- Meena, R. S., Kumar, S., Datta, R., Lal, R., Vijayakumar, V., et al. (2020). Impact of Agrochemicals on Soil Microbiota and Management: A Review. *Land.* 9, :34. doi: 10.3390/land9020034
- Mehta, P., Sharma, R., Putatunda, C., and Walia, A. (2019). "Endophytic fungi: role in phosphate solubilization," in Advances in Endophytic Fungal Research, Fungal Biology, ed. B. P. Singh (Berlin: Springer Nature Switzerland AG), doi: 10.1007/978-3-030-03589-1_9
- Mousa, W. K., and Raizada, M. N. (2013). The diversity of anti-microbial secondary metabolites produced by fungal endophytes: an interdisciplinary perspective. *Front. Microbiol.* 4:65. doi: 10.3389/fmicb.2013.00065
- Murphy, B., Fiona, R., Doohan, M., Trevor, R., and Hodkinson. (2018). From concept to commerce, developing a successful fungal endophyte inoculant for agricultural crops. J. Fungi. 4, :24. doi: 10.3390/jof4010024
- Muvea, A. M., Meyhöfer, R., Subramanian, S., Poehling, H. M., Ekesi, S., and Maniana, N. K. (2014). Colonization of onions by endophytic fungi and their impacts on the biology of *Thrips tabaci. PLoS One* 9:e108242. doi: 10.1371/journal.pone.0108242
- Nascimento, F. X., Rossi, M. J., Soares, C. R., McConkey, B. J., and Glick, B. R. (2014). New insights into 1-aminocyclopropane-1carboxylate (ACC) deaminase phylogeny, evolution and ecological significance. *PLoS One* 9:e99168. doi: 10.1371/journal.pone.009 9168

- Newbery, F., Qi, A., and Fitt, B. D. L. (2016). Modelling impacts of climate change on arable crop diseases, progress, challenges and applications. *Curr. Opinion. Plant Biol.* 32, 101–109. doi: 10.1016/j.pbi.2016.07.002
- Ningxiao, L. I., Alfiky, A., Vaughan, M. M., and Kang, S. (2016). Stop and smell the fungi: Fungal volatile metabolites are overlooked signals involved in fungal interaction with plants. *Fungal. Biol. Rev.* 30, 134–144. doi: 10.1016/j.fbr.2016. 06.004
- Nuangmek, W., Aiduang, W., Kumla, J., Lumyong, S., and Suwannarach, N. (2021). Evaluation of a newly identified endophytic fungus, *Trichoderma phayaoense* for plant growth promotion and biological control of gummy stem blight and wilt of muskmelon. *Front. Microbiol.* 12:634772. doi: 10.3389/fmicb.2021. 634772
- Omomowo, O. I., and Babalola, O. O. (2019). Bacterial and fungal endophytes, tiny giants with immense beneficial potential for plant growth and sustainable agricultural productivity. *Microorganisms Microorg.* 7, :481. doi: 10.3390/ microorganisms7110481
- Pais, I. P., Roboredo, F. H., Ramalho, J. C., Pessoa, M. F., Lidon, F. C., and Silva, M. M. (2020). Potential impact of climate change on agriculture- A review. Emir. J. Food Agr. 32, 397–407. doi: 10.9755/ejfa.2020.v32.i6.2111
- Park, Y. H., Mishra, R. C., Yoon, S., Kim, H., Park, C., et al. (2019). Endophytic Trichoderma citrinoviride isolated from mountain-cultivated ginseng (Panax ginseng) has great potential as a biocontrol agent against ginseng pathogens. J. Ginseng Res. 43, 408–420. doi: 10.1016/j.jgr.2018.03.002
- Parsa, S., García-Lemos, A. M., Castillo, K., Ortiz, V., López-Lavalle, L. A. B., Braun, J., et al. (2016). Fungal endophytes in germinated seeds of the common bean. Phaseolus vulgaris. Fungal. Biol. 120, 783–790. doi: 10.1016/j.funbio.2016.01. 017
- Plett, J. M., and Martin, F. M. (2018). Know your enemy, embrace your friend, using omics to understand how plants respond differently to pathogenic and mutualistic microorganisms. *Plant J.* 93, 729–746. doi: 10.1111/tpj.13802
- Qayyum, M. A., Wakil, W., Arif, M. J., Sahi, S. T., and Dunlap, C. A. (2015). Infection of *Helicoverpa armigera* by endophytic *Beauveria bassiana* colonizing tomato plants. *Biol. Cont.* 90, 200–207.
- Rabha, A. J., Naglot, A., Sharma, G. D., Gogoi, H. K., and Veer, V. (2014). In vitro evaluation of antagonism of endophytic *Colletotrichum gloeosporioides* against potent fungal pathogens of *Camellia sinensis*. *Ind. J. Microbiol.* 54, 302–309. doi: 10.1007/s12088-014-0458-8
- Radziemska, M., Gusiatin, A., Cydzik-Kwiatkowska, A., Cerda, V., and Pecina, A. (2021). Insight into metal immobilization and microbial community structure in soil from a steel disposal dump phytostabilized with composted, pyrolyzed or gasified wastes. *Chemosphere* 272,:129576. doi: 10.1016/j.chemosphere.2021. 129576
- Rana, K. L., Kour, D., Kaur, T., Devi, R., Yadav, A. N., Yadav, N., et al. (2020). Endophytic microbes: biodiversity, plant growth-promoting mechanisms and potential applications for agricultural sustainability. *Antonie Van Leeuwenhoek International Journal of General and Molecular Microbiology*. 113, 1075–1107. doi: 10.1007/s10482-020-01429-y
- Ray, S., Singh, V., Bisen, K., Keswani, C., Singh, S., and Singh, H. B. (2017). "Endophytomicrobiont: A Multifaceted Beneficial Interaction," in *Advances in PGPR Research*, eds H. B. Singh, B. K. Sarma, and C. Keswani (Wallingford, UK: CABI), 218–233.
- Redman, R. S., Kim, Y. O., Cho, S., Mercer, M., Rienstra, M., Manglona, R., et al. (2021). A Symbiotic Approach to Generating Stress Tolerant Crops. Microorganisms Microorg. 9, :920. doi: 10.3390/microorganisms9050920
- Rehman, S., Akhtar, M., and Abdullah, S. N. A. (2016). *Plant, Soil and Microbes*. Cham: Springer, doi: 978-3-319-29573-2
- Renuka, S., and Ramanujam, B. (2016). Fungal endophytes from maize (*Zea mays* L.): isolation, identification and screening against maize stem borer, *Chilo partellus* (Swinhoe). *J. Pure Appl. Microbiol.* 10, 523–529. doi: 10.1007/s12275-013-2356-x
- Ripa, F. A., Cao, W., Tong, S., and Sun, J. G. (2019). Assessment of plant growth promoting and abiotic stress tolerance properties of wheat endophytic fungi. *Biomed. Res. Int.* 2019, 1–12. doi: 10.1155/2019/6105865
- Rodríguez, M. C. H., Evans, H. C., de Abreu, L. M., Macedo, M., Miraine, K., Bekele, K. B., et al. (2021). New species and records of *Trichoderma* isolated as mycoparasites and endophytes from cultivated and wild coffee in Africa. *Sci. Rep.* 11:5671. doi: 10.1038/s41598-021-84111-1

- Rodriguez, R. J., White, J. F. Jr., Arnold, A. E., and Redman, R. S. (2009). Fungal endophytes: diversity and functional roles. *New Phytologist.* 182, 314–330. doi: 10.1111/j.1469-8137.2009.02773.x
- Rojas, E., Birgit, J., Hans, J., Lyngs, J., Meike, L., Pilar, E., et al. (2020). Selection of fungal endophytes with biocontrol potential against Fusarium head blight in wheat. *Biol. Cont.* 144,: 104222. doi: 10.1016/j.biocontrol.2020.104222
- Rouhier, N., Koh, C. S., Gelhaye, E., Corbier, C., Favier, F., Didierjean, C., et al. (2008). Redox based antioxidant systems in plants: biochemical and structural analyses. *Biochimica. Biophysica. Acta.* 1780, 1249–1260.
- Rozpadek, P., Domka, A. M., Nosek, M., Wazny, R., Jedrzejczyk, R. J., Wiciarz, M., et al. (2018). The role of strigolactone in the crosstalk between *Arabidopsis thaliana* and the endophytic fungus *Mucor* sp. *Front. Microbiol.* 9:441. doi: 10.3389/fmicb.2018.00441
- Rustamova, N., Litao, N., Bozorov, K., Sayyed, R., Aisa, H. A., and Yili, A. (2022). Plant-associated endophytic fungi: A source of structurally diverse and bioactive natural products. *Plant Cell Biotechnol. Mol. Biol.* 23, 1–19. https://www.ikppress.org/index.php/PCBMB/article/view/7454
- Sadeghi, F., Samsampour, D., Seyahooei, M. A., Bagheri, A., and Soltani, J. (2019). Diversity and spatiotemporal distribution of fungal endophytes associated with Citrus reticulata cv. Siyahoo. Curr. Microbiol. 76, 279–289. doi: 10.1007/s00284-019-01632-9
- Sahoo, S., Sarangi, S., and Kerry, R. G. (2017). "Bioprospecting of endophytes for agricultural and environmental sustainability (Chapter-19)," in *Microbial Biotechnology*, ed. J. K. Patra –429–458. doi: 10.1007/978-981-10-6847-8_19
- Sallam, N., Ali, E. F., Seleim, M. A. A., et al. (2021). Endophytic fungi associated with soybean plants and their antagonistic activity against *Rhizoctonia solani*. *Egypt. J. Biol. Pest Cont.* 31, :54. doi: 10.1186/s41938-021-00402-9
- Sánchez-Rodríguez, A. R., Raya-Díaz, S., Zamarreño, A. M., García-Mina, J. M., del Campillo, M. C., and Quesada-Moraga, E. (2018). An endophytic Beauveria bassiana strain increases spike production in bread and durum wheat plants and effectively controls cotton leafworm (Spodoptera littoralis) larvae. Biol. Control. 116, 90–102. doi: 10.1016/j.biocontrol.2017.01.012
- Santamaria, O., Lledó, S., Rodrigo, S., and Poblaciones, M. J. (2017). Effect of fungal endophytes on biomass yield, nutritive value and accumulation of minerals in ornithopus compressus. *Microbial Ecology.*. 74, 841–852. doi: 10.1007/s00248-017-1001-3
- Schulz, B., Haas, S., Corina, J., Nicole, A. B., and Max, S. (2015). Fungal endophytes are involved in multiple balanced antagonisms. *Curr. Sci.* 109, 39–45.
- Schulz, B., Römmert, A. K., Dammann, U., Aust, H. J., and Strack, D. (1999). The endophyte-host interaction, a balanced antagonism? *Mycol. Res.* 103, 1275–1283. doi: 10.1017/S0953756299008540
- Shah, S., Shrestha, R., Maharjan, S., Selosse, M. A., and Pant, B. (2018). Isolation and characterization of plant growth-promoting endophytic fungi from the roots of *Dendrobium moniliforme*. *Plants (Basel)*. 8, :5. doi: 10.3390/ plants8010005
- Sharma, A., Gupta, A., Dalela, M., Sharma, S., Sayyed, R. Z., Enshasy, H. A. E., et al. (2020). Linking organic metabolites as produced by *Purpureocillium lilacinum* 6029 cultured on karanja deoiled cake medium for the sustainable management of root-knot nematodes. *Sustainability*. 12, :8276. doi: 10.3390/su12198276
- Shinozaki, K., and Yamaguchi-Shinozaki, K. (2007). Gene networks involved in drought stress response and tolerance. *J. Exp. Bot.* 58, 221–227. doi: 10.1093/jxb/erl164
- Singh, N., Singh, A., and Dahiya, P. (2021). "Plant growth-promoting endophytic fungi from different habitats and their potential applications in agriculture," in *Recent trends in Mycological Research. Fungal Biology*, ed. A. N. Yadav (Cham: Springer), doi: 10.1007/978-3-030-60659-6_3
- Singh, R. P., Shelke, G. M., Kumar, A., and Jha, P. N. (2015). Biochemistry and genetics of ACC deaminase: a weapon to "stress ethylene" produced in plants. Frontiers. in Microbiology.. 6:937. doi: 10.3389/fmicb.2015. 00937
- Sreeja, K., Anandaraj, M., and Bhai, R. S. (2016). In vitro evaluation of fungal endophytes of black pepper against *Phytophthora capsici* and *Radopholussimilis*. *J. Spices Aromat. Crops* 25, 113–122.
- Srinivas, C., Shubha, S., Narasimhamurthy, K., Arakere, U., and Sudisha, J. (2020). Detection and Characterization of Antibacterial Siderophores Secreted by Endophytic Fungi from Cymbidium aloifolium. *Biomolecules* 10:1412. 10.3390/biom10101412 doi: 10

- Srivastava, A., Srivastava, N., and Sayyed, R. Z. (2012). In vitro bio-control activity of *Trichoderma* species against phytopathogenic A. brassicae. Int. J. Biotech. BioSci. 2, 6–9.
- Srivastava, A., Srivastava, N., and Sayyed, R. Z. (2014). Bio-control Potential of Trichoderma Species Against Alternaria brassicae. Recent advances in biofertilizers & biofungicides (PGPR) for sustainable Agriculture. London, UK: Cambridge Scholars Press, 393–400.
- Srivastava, S., Kadooka, C., and Uchida, J. Y. (2018). Fusarium species as pathogen on orchids. Microbiol. Res. 207, 188–195. doi: 10.1016/j.micres.2017.12.002
- Steenwerth, K. L., Hodson, A. K., Bloom, A. J., Carter, M. R., Cattaneo, A., et al. (2014). Climate-smart agriculture global research agenda: scientific basis for action. Agric. Food Sec. 3, :11. doi: 10.1186/2048-7010-3-11
- Sukmawati, D., Family, N., Hidayat, I., Sayyed, R. Z., Elsayed, E. A., Dailin, D. J., et al. (2021). Biocontrol activity of Aureubasidium pullulans and Candida orthopsilosis isolated from tectona grandis l. phylloplane against aspergillus sp. in post-harvested citrus fruit. Sustainability. 13,:7479. doi: 10.3390/su13137479
- Suryanarayanan, T. S., Devarajan, P. T., Girivasan, K. P., Govindarajulu, M. B., Kumaresan, V., et al. (2018). The host range of multi-host endophytic fungi. Curr. Sci. 115,:25. doi: 10.18520/cs/v115/i10/1963-1969
- Tang, J., Xu, L., Chen, X., and Hu, S. (2009). Interaction between C4 barnyard grass and C3 upland rice under elevated CO₂, impact of mycorrhizae. *Acta Oecol.* 35, 227–235. doi: 10.1016/j.actao.2008.10.005
- Tiwari, K. (2015). The future products, Endophytic fungal metabolites. *J. Biodiv. Bioprosp. Develop.* 2, 2–7. doi: 10.4172/2376-0214.1000145
- Vandenkoornhuyse, P., Quaiser, A., Duhamel, M., Van, A. L., and Dufresne, A. (2015). The importance of the microbiome of the plant holobiont. *New Phytol.* 206, 1196–1206. doi: 10.1111/nph.13312
- Waller, F., Achatz, B., Baltruschat, H., Fodor, J., Becker, K., Fischer, M., et al. (2005).
 The endophytic fungus *Piriformospora indica* reprograms barley to salt-stress tolerance, disease resistance, and higher yield. *Proc. Natl. Acad. Sci. U.S.A.* 102, 13386–13391. doi: 10.1073/pnas.0504423102
- Wang, J., Li, T., Liu, G. Y., et al. (2016). Unraveling the role of dark septate endophyte (DSE) colonizing maize (*Zea mays*) under cadmium stress: physiological, cytological and genic aspects. *Sci. Rep.* 6, :22028. doi: 10.1038/ srep22028
- Wei, Q. Y., Li, Y. Y., Xu, C., Wu, Y. X., Zhang, Y. R., and Liu, H. (2020). Endophytic colonization by Beauveria bassiana increases the resistance of tomatoes against *Bemisia tabaci. Arthropod. Plant Interact.* 14, 289–300.
- Xia, Y., Sahib, M. R., Amna, A., et al. (2019). Culturable endophytic fungal communities associated with plants in organic and conventional farming systems and their effects on plant growth. Sci. Rep. 9, :1669. doi: 10.1038/ s41598-018-38230-x
- Xiao, X., Luo, S. L., Zeng, G. M., Wei, W. Z., Wan, Y., et al. (2010). Biosorption of cadmium by endophytic fungus (EF) *Microsphaeropsis* sp. LSE10 isolated from cadmium hyperaccumulator *Solanum nigrum L. Bioresour. Technol.* 101, 1668–1674. doi: 10.1016/j.biortech.2009.09.083
- Yadav, A., and Yadav, K. (2017). Exploring the potential of endophytes in agriculture, a mini review. Adv. Plant Agricul. Res. 6, 102-106. doi: 10.15406/ apar.2017.06.00221
- Yan, L., Zhu, J., Zhao, X., Shi, J., Jiang, C., and Shao, D. (2019). Beneficial effects of endophytic fungi colonization on plants. Appl. Microbio. Biotechnol. 103, 3327–3340. doi: 10.1007/s00253-019-09713-2
- Yung, L., Sirguey, C., Azou-Barré, A., and Blaudez, D. (2021). Natural fungal endophytes from *Noccaea caerulescens* mediate neutral to positive effects on plant biomass, mineral nutrition and Zn Phytoextraction. *Frontiers. in Microbiology.*, 12:689367. doi: 10.3389/fmicb.2021.689367
- Zahoor, M., Muhammad, I., Hazir, R., Muhammad, Q., Sahib, A., et al. (2017).
 Alleviation of heavy metal toxicity and phytostimulation of *Brassica campestris*L. by endophytic Mucor sp. MHR-7. *Ecotoxicol. Environ. Safety.* 142, 139–149. doi: 10.1016/j.ecoenv.2017.04.005
- Zakaria, L., and Aziz, W. N. W. (2018). Molecular Identification of Endophytic Fungi from Banana Leaves (*Musa* spp.). Trop. Life Sci. Res. 29, 201–211. doi: 10.21315/tlsr2018.29.2.14
- Zhang, Q., Zhang, J., Yang, L., Zhang, L., Jiang, D., Chen, W., et al. (2014). Diversity and biocontrol potential of endophytic fungi in Brassica napus. Biol. Control. 72, 98–108. doi: 10.1016/j.biocontrol.2014. 02.018

Zope, V. P., Jadhav, H. P., and Sayyed, R. Z. (2019). Neem cake carrier prolongs shelf life of biocontrol fungus *Trichoderma viridae*. *Ind. J. Expl. Biol.* 57, 372–375.

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