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Review

MAPK/ERK Signaling in Regulation of Renal Differentiation

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Abstract: Congenital anomalies of the kidney and urinary tract (CAKUT) are common birth defects derived from abnormalities in renal differentiation during embryogenesis. CAKUT is the major cause of end-stage renal disease and chronic kidney diseases in children, but its genetic causes remain largely unresolved. Here we discuss advances in the understanding of how mitogen-activated protein kinase/extracellular signal-regulated kinase (MAPK/ERK) activity contributes to the regulation of ureteric bud branching morphogenesis, which dictates the final size, shape, and nephron number of the kidney. Recent studies also demonstrate that the MAPK/ERK pathway is directly involved in nephrogenesis, regulating both the maintenance and differentiation of the nephrogenic mesenchyme. Interestingly, aberrant MAPK/ERK signaling is linked to many cancers, and recent studies suggest it also plays a role in the most common pediatric renal cancer, Wilms' tumor.

Keywords: extracellular signal-regulated kinase; MAPK/ERK signaling; intracellular signaling; kidney development; ureteric bud branching morphogenesis; nephrogenesis; progenitor cells; self-renewal; differentiation

1. Introduction

The developing kidneys undergo complex morphogenesis to become fully functional, non-regenerating organs at birth. Renal morphogenesis is guided by classical inductive interactions, which are now known to take place through cell-cell-induced signaling, leading to activation of cell-intrinsic intracellular cascades. It has become evident that receptor tyrosine kinase (RTK)-activated signaling is a key regulator of kidney development [1,2]. The major inducers of RTK signaling in the early developing kidney include glial cell line-derived neurotrophic factor (GDNF)-induced RET signaling and fibroblast growth factor (FGF) signaling. Binding of these growth factors to their cognate receptors causes receptor dimerization, autophosphorylation, and activation of intracellular signaling cascades, including the mitogen-activated protein kinase/extracellular signal-regulated kinase (MAPK/ERK) pathway [3,4]. The significant roles of the MAPK/ERK pathway in morphogenesis are emerging [5–8], and we recently reported the activation of MAPK/ERK patterns and functions in nephron progenitors (NPs) of the developing kidney [9]. However, the full spectrum of MAPK/ERK functions at the cellular level, and specifically during renal development, are only partially determined. This review focuses on the known roles of MAPK/ERK signaling in renal differentiation and discusses the missing pieces that are needed to complement our understanding of this important topic.

1.1. RTK Signaling in General

The RTKs form a family of cell-surface receptors possessing an extracellular domain that binds to signaling molecules such as growth factors, hormones, cytokines, and neurotrophic factors [10]. RTK signaling has important functions in regulating cellular processes such as cell cycle progression during proliferation, cell metabolism, migration, survival, and differentiation. Several growth factors (e.g., epidermal growth factor, insulin, and the insulin-like growth factor) bind to RTKs, upon which the tyrosine residues within the receptors are phosphorylated, leading to changes in either downstream target gene expression and/or cytosolic protein phosphorylation [11]. The intracellular pathways mediating extracellular stimuli into the cell interior include, for example, PI3K/AKT/mTOR, RAS/RAF/MEK/ERK, and PLC γ /PKC [12,13] (Figure 1).

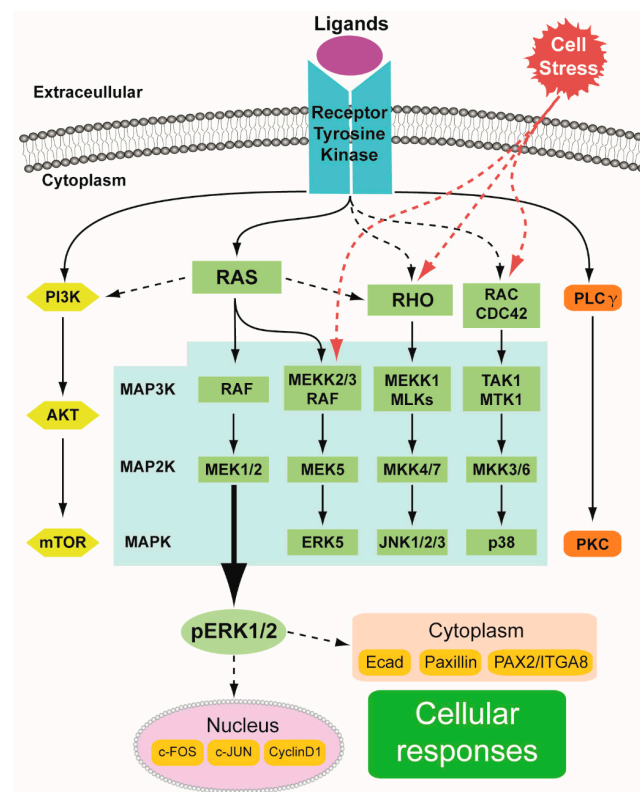


Figure 1. Schematic summary of the multiple MAPK pathways. MAPK pathways, highlighted in the light blue box, mediate extracellular information to the cell interior. Ligand (purple oval) binding to the dimeric transmembrane tyrosine kinase receptor (blue blocks) and/or cellular stress (inflammation, ischemia, hypoxia) activate a spectrum of downstream intracellular cascades such as PI3K/AKT/mTOR, RAS/RAF/MEK/ERK, and PLC γ /PKC. The last step of MAPK/ERK cascade activation results in phosphorylation of ERK1/2 leading to both transcriptional target regulation in the nucleus (light lilac oval, including c-FOS, c-JUN and Cyclin D1) and in the cytoplasm (peach-colored box, including those identified in the developing kidney: E-cadherin, Paxillin, and PAX2/ITGA8) followed by cell-type specific cellular responses (green box). Black lines indicate linear activation by phosphorylation, black dotted line indicates activation within the cascade and red dotted line indicates activation by cellular stress. Abbreviations: PI3K; Phosphoinositide 3-kinases, AKT; protein kinase B, mTOR; mammalian target of rapamycin, MAP3K; MAP kinase kinase kinase, MAP2K; MAP kinase kinase, MAPK; MAP kinase, MLKs; Mixed lineage kinases, MKK; Mitogen-activated protein kinase kinase, CDC42; Cell division control protein 42 homolog, TAK1; Mitogen-activated protein kinase kinase kinase 7, MTK1; Mitogen-activated protein kinase kinase kinase 4, PLC γ ; Phospholipase C gamma, PKC; Protein kinase C, pERK1/2; phosphorylated ERK1/2, Ecad; E-cadherin, PAX2; paired box protein 2, ITGA8; integrin alpha 8.

1.2. MAPK Pathway

Four distinct MAPK pathways execute RTK functions through the small G-protein RAS (Figure 1). The best characterized is the RAS/RAF/MEK/ERK pathway, which in mammals constitutes three RAF proteins (RAF1, A- and B-RAF), two MEK (MEK1 and -2), and two ERK (ERK1 and -2) proteins. Activated RAS generally stimulates RAF but may also phosphorylate MEKK2/3, thus inducing activation of another MAPK cascade consisting of MEKK2/3-MEK5-ERK5 [14]. Analogous to other MAPK pathways, sequential protein kinase phosphorylation via Rho-MEKK1(or MLKs)-MKK4/7 activates JNK1/2/3. Yet another parallel MAPK pathway consists of RAC (or Cdc42)-TAK1 (or MTK1)-MKK3/6 proteins to finally activate the effector protein p38 [15–21]. Highlighting the complex nature of cascade activation, RAS proteins not only induce the MAPK/ERK pathway but also activate the PI3K pathway [22,23]. The regulation and molecular events that determine which intracellular cascade gets activated upon RTK signal transduction are not fully known. Interestingly, high levels of ERK and p38 were detected by Western blotting and immunostaining in the rat embryonic kidney, while JNK was abundant in the adult kidney [24]. Moreover, in vitro experiments also suggest involvement of the JNK pathway during embryonic kidney development [25,26]. These observations imply a functional requirement of MAPK/ERK, p38, and JNK pathways for renal differentiation [27,28].

1.2.1. MAPK/ERK Cascade

The first identified MAPK pathway, the ERK1/2 cascade, is activated by various stimuli and contributes to the regulation of proliferation, differentiation, cell survival, learning, migration, apoptosis, morphology determination, and oncogenic transformation [29]. After RAS engagement, RAF protein kinases RAF1, B-RAF, and A-RAF are activated and phosphorylate their dual-specificity protein kinase substrates MEK1 and MEK2 (also known as MAP2K1 and MAP2K2) [30]. Subsequently, MEK1/2 phosphorylate their substrates, the best-known of which are ERK1/2, which have a wide range of both cytosolic and nuclear targets and functions [30,31].

Interestingly, a significant portion (50–70%) of ERK1/2 molecules can be found in the nucleus within 10–20 min of cellular stimulation, and this appears to be a key step for the quick regulation of cell cycle progression and DNA replication [32,33]. Recent work by Johnson and Toettcher demonstrated that not only signal duration but also total amplitude of ERK activity specifies the embryonic lineages in *Drosophila melanogaster* [34]. By combining optogenetic stimulation and genetic inactivation of ERK, they found that ectodermal cell types are induced with a fast ERK pulse, while differentiation of gut endoderm requires a longer ERK stimulus. Finally, they showed that incremental ERK activation leads to a switch in cell fates during endoderm/ectoderm specification thus indicating that total ERK load rather than amplitude or duration of ERK activity controls fate decisions. Other reports show that ERK signaling regulates pluripotency in mouse and human embryonic stem cells (ESCs). In ESCs, the MAPK/ERK pathway controls genomic stability, self-renewal, and differentiation [35,36]. Further highlighting the importance of the MAPK/ERK pathway in tissue homeostasis, mutations in *RAS*, *B-RAF*, *RAF1*, *MEK1*, or *MEK2* appear to drive uncontrolled cell proliferation and survival of cancer cells in a growth factor-independent manner [37,38]. Mutations in the negative regulators of MAPK/ERK pathway activation (such as *NF1*) significantly contribute to tumor biology and emphasize the importance of pathway control at multiple levels [39,40].

1.2.2. Other MAPK Cascades

Unlike the ERK pathway, which typically mediates RTK signaling, JNK and p38 pathways respond to a variety of stress signals [41]. For example, JNK and p38 pathways are activated by proinflammatory cytokines, such as tumor necrosis factor (TNF)- α and interleukin (IL)-1 β , and by cellular stresses [42]. JNK is involved in apoptosis, immune cell development, and autophagy [43,44]; while p38 can additionally control cell cycle, growth, and differentiation [45]. There is evidence that

both JNK and p38 pathways may participate in kidney development through the control of nephron differentiation [25,26].

Considerably less is known about ERK5 than the other MAPK pathways. It is activated by inflammatory cytokines, ischemia and hypoxia, and to a lesser extent by RTK signaling. The ERK5 pathway is linked to diverse cellular processes including proliferation, migration, survival, and angiogenesis [46]. In general, ERK5 is universally expressed in many tissues, but the highest level is detected in the brain with peak levels during early embryonic development [47]. The role of the MAPK/ERK5 pathway in kidney development has not been examined.

1.3. MAPK/ERK Pathway: Lessons from Mouse Models

Genetic experiments targeting multiple components of the RAF/MEK/ERK pathway have greatly facilitated the understanding of the consequences of MAPK/ERK activation *in vivo*. Typically, constitutive loss-of-function of MAPK/ERK pathway components (e.g., *B-raf*, *Raf-1*, *Erk2*) result in early embryonic lethality [48–52]. Therefore, gain-of-function mouse models together with conditional alleles are essential for the characterization of the tissue- and organ-specific requirements for MAPK/ERK activation [53–55]. Since this review focuses on renal differentiation, we will discuss in detail the genetic models of *Mek1* and -2, which have been utilized in studies of the developing kidney and compare those findings to what has been discovered in *Erk1* and -2 models.

Genetic inactivation of *Mek1* in mice demonstrated its indispensable function in embryogenesis [56]. Characterization of *Mek1* mutant mice revealed that the embryonic lethality is caused by defects in the placenta, which exhibited decreased amounts of endothelial cells in the labyrinthine region. Interestingly, the tetraploid complementation of mutant embryos rescues the placental phenotype and results in viable offspring, suggesting that MEK1 function is not required for the development of the embryo proper [57]. Also, *Mek2* deletion supports normal embryogenesis and results in viable offspring [58]. However, double heterozygosity of *Mek1* and -2 results in a phenotype very reminiscent of that observed in *Mek1*-null embryos, thus demonstrating that while MEK1 functions appear to predominate, both kinases are essential for normal placental development [59,60]. The work by Aoidi and colleagues further supports the redundant function of MEK1 and -2, which shows that genetic replacement of *Mek1* with *Mek2* under the control of endogenous *Mek1* regulatory elements fully supports normal development [61]. These experiments also clearly demonstrate an essential effect of gene dosage for functional ERK signaling.

It has additionally been shown that MEK1 can negatively regulate MEK2-dependent activation of ERK signaling in cultured cells and MEK1-deficient embryos [62]. This suggests that MEK1 and -2 are not interchangeable, and as shown, loss of MEK1 may increase ERK1/2 phosphorylation. Despite this, combinational deletion of MEK1 alone or together with MEK2 has shown that MAPK signaling participates in the regulation of synaptic plasticity and development of excitatory neurons in the neocortex, functional skin barrier, and the respiratory tract [5,7,63,64].

Similar to MEK mouse models, genetic inactivation of *Erk2*, but not *Erk1*, results in embryonic lethality due to placental defects [52,65–67]. Although ERK1-deficient mice appear normal, ERK1 has been implicated in asthma (due to its requirement for T-helper cell differentiation), resistance to obesity and skin papillomas, long-term memory function, and control of hyperactivity [68–72]. Because homozygous deletion of *Erk2* in mice is embryonic lethal, hemizygote disruption (*Erk^{-/+}*) and hypomorphic mice have been utilized to reveal its functional requirements for development and organ functionality (e.g., for normal long-term memory [73]).

Tissue-specific roles of ERK1 and -2 and their potential physiologically distinct functions have been under intensive research during the last decade. Concluding from extensive experiments performed in both *in vitro* cell culture and *in vivo* mouse models, it appears that ERK1 and -2, again similar to their upstream kinases MEK1 and -2, have redundant functions [74,75]. Misinterpretation of some of these results may derive from the differences in expression levels of ERK1 and -2, as the latter is more abundantly expressed in the majority of tissues [75]. When this is taken into account, both proteins are

equally important (e.g., for normal proliferation [76]). This is also supported by genetic experiments that have revealed the essential involvement of ERK1 and -2 in craniofacial development, neurogenesis, hematopoietic stem cell maintenance, and placental differentiation [8,77–80].

2. Kidney Development

Kidneys develop via branching morphogenesis, similar to many other organs including the lungs, mammary glands, and seminal vesicles. Novel branch generation specific to each organ determines the characteristic growth and shape of these organs. As such, each organ has its own unique pattern of epithelial branching that is determined by signals from the surrounding nascent mesenchyme [81,82].

Mammalian renal development occurs in three spatially and temporally distinct stages: The pronephros, mesonephros, and metanephros. All three stages derive from the intermediate mesoderm [83,84]. On embryonic day 8.75 (E8.75) in mice (gestational day 28 in humans [85]), paired nephric ducts emerge and grow posteriorly until they contact and connect to the cloaca. During this time, the nephric ducts also induce the growth of the pro- and mesonephric tubules [86–88], which are transient structures in females. In males, the caudal mesonephric tubules and distal nephric duct contribute to the differentiation of the accessory organs of the reproductive system [89] (Figure 2).

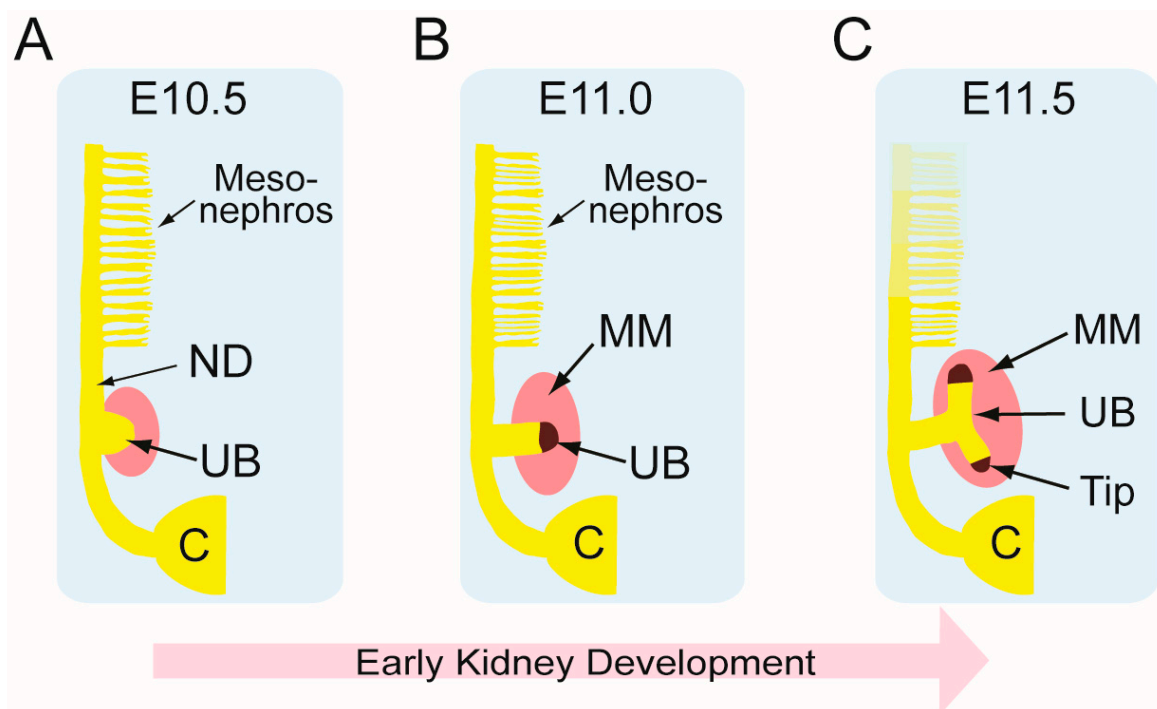


Figure 2. Induction of permanent kidney development in mice. (A) The most prominent kidney at embryonic day 10.5 (E10.5) is the mesonephros, which is a transient, embryonic kidney. At the initiation of permanent kidney development, the metanephric mesenchymal (MM, pink) cells interact with the ureteric bud (UB, yellow), and signal propagation between the MM and UB induces UB outgrowth from the nephric duct (ND) near the cloaca (C); (B) In the next step at E11.0, the MM secretes signals that attract the UB to grow towards it. The UB itself is divided into UB tip (dark brown) and trunk (yellow); (C) The first branching event takes place at E11.5 when reciprocal inductive interactions between the UB and MM lead to formation of a T-shaped UB. The previous kidney, mesonephros, begins to regress. C; cloaca, ND; nephric duct, MM; metanephric mesenchyme, UB; ureteric bud.

2.1. Kidney Induction—Formation of the Ureteric Bud and Branching Morphogenesis

The adult kidney, or metanephros, begins forming at E10.5 in mice and during weeks 4–5 of gestation in humans, when a mass of metanephric mesenchyme (MM) cells induces UB outgrowth from the nephric duct. Several transcriptional regulators are involved with the initial MM formation, including EYA1, SIX1/SIX4, and OSR1 [90–95]. Additionally, it has been suggested that PAX2/PAX8 function in guiding the initial MM establishment [93,96–99]. For a detailed review of kidney development, see Krause et al. [100].

Reciprocal inductive interactions between the MM and the nephric duct guide the initial formation of the UB. These interactions are mediated by secreted molecules and extracellular matrix components, such as GDNF, FGFs, and integrins, which channel molecular instructions to induce cellular responses. For UB outgrowth, MM cells secrete GDNF which binds to the RET tyrosine kinase receptor and co-receptor GFRA1, both expressed in the nephric duct [4,101,102]. All components of the GDNF/RET signaling pathway are required for normal UB outgrowth which takes place by an active movement of the RET^{high} cells within the nephric duct that clusters these cells and leads to UB formation [103–110]. These cellular movements are mediated by ETS transcription factors, ETV4 and ETV5 [111,112]. FGF signaling through receptors FGFR1/2 is likewise important for induction of UB formation, while FGF signaling in the MM, through an unknown mechanism, prevents ectopic UB formation [113,114].

Reciprocal inductive signaling continues to play significant roles in UB branching morphogenesis. Although the importance of GDNF/RET and FGF signaling for UB branching has been demonstrated in detail [3,104,115,116], downstream targets mediating their functions are still largely unclear. Genetically modified mouse models have shown that the balance in GDNF levels essentially dictates the morphology of the primary UB and its subsequent branching as not only reduction and loss of, but also endogenously increased or expanded GDNF, results in severe renal defects [117–122]. GDNF activates essential WNT functions that aid the progression of branching morphogenesis [123–127], while strong inhibitory effects of BMP4 on UB branching and GDNF functions are reported [98,128–130].

The entire collecting duct system is formed by repeated UB branching [131,132]. The UB is divided into two regions, known as tips and trunks, each having distinct cellular properties. Cells within the UB tips are immature, highly proliferative, and in direct contact with the MM, facilitating their interaction. The tip cells generate new branches and tips [133], and it has been convincingly shown that cells in the UB trunk derive from tip cells that differentiate to form the entire collecting duct system [134,135]. Therefore, tip cells are the progenitors of the entire collecting duct system. Although the specifics of the molecular signatures in UB tip cells and their regulation are largely unknown, RET signaling has been shown to maintain UB cells in the tip niche [110,111,135] (Figure 3).

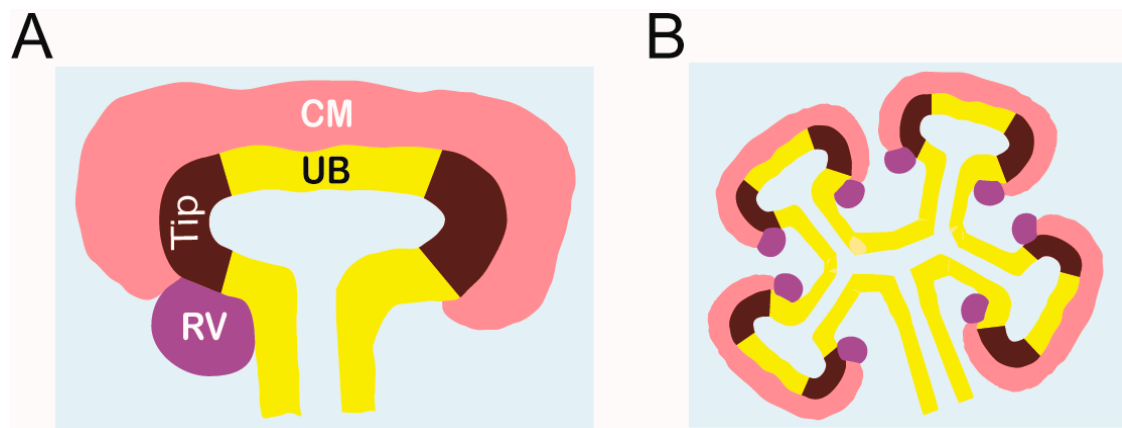


Figure 3. Ureteric bud (UB) branching morphogenesis. **(A)** Branching morphogenesis begins by the formation of a T-bud, which is composed of the UB trunk (yellow) and tips (dark brown). At this stage, the metanephric mesenchyme is condensed around the UB tips and forms cap mesenchyme (CM, light red). UB-derived signals induce CM cells to undergo mesenchyme-to-epithelial (MET) conversion and produce the renal vesicle (RV, purple) in the armpits of the T-bud; **(B)** By E12.5, the UB has undergone a series of iterative branching events under the influence of mesenchymal signals. Most of the UB branches form by stereotypic bifurcation of an existing tip, and each new tip maintains the surrounding CM. The UB undergoes a total of approximately 10 to 11 rounds of branching events during which it induces MET and thus nephrogenesis within each newly generated tip. CM; cap mesenchyme, UB; ureteric bud, Tip; UB tips, RV; renal vesicle.

2.2. Nephron Progenitors and Nephrogenesis

Once UB branching begins, the UB in turn reciprocally induces the condensation of the MM around the tips. This condensed mesenchyme surrounding each UB tip is referred to as the cap mesenchyme (CM) and acts as a source of nephron progenitors (NPs) [136]. Molecularly, NPs are defined by the expression of transcription factor SIX2 and transcriptional cofactor CITED1 [137,138]. NPs form their own niche, which is arranged by contacts between NP cells as well as contacts with the UB and stromal progenitors, but the mechanisms behind the niche arrangement are only about to be elucidated [9,139].

The multipotent, self-renewing NP population differentiates to give rise to the functional units of the kidney, the nephrons. Each newly formed UB tip induces the differentiation in subsets of NP cells which first cluster under the UB tip to form a pretubular aggregate. As nephrogenesis proceeds, the pretubular aggregate epithelializes to form the renal vesicle (RV), which grows and patterns into a comma-shaped and then S-shaped body. The distal S-shaped body differentiates to become the distal and connecting tubule, which plumb into the collecting duct system allowing fluid flow from the kidney [140]. The proximal and medial segments of the S-shaped body give rise to the glomerulus and proximal tubule/loop of Henle, respectively [81,141]. NP cells are gradually recruited into the epithelial nephron precursors, and the timing of the recruitment essentially predicts their future fate in the functional nephron [142].

To assure a sufficient final nephron number, the NP population must be maintained during active UB branching morphogenesis, as indicated by depletion experiments showing that nephron count critically depends on the NP population size [143]. Details of the molecules affecting NP cell commitment were recently reviewed [140]. Nephron progenitor maintenance is controlled by many transcriptional regulators, the most well-known being SIX2, which is required for self-renewal and preventing premature differentiation [144]. Recently, it was also determined that SALL1 has a similar role in NP maintenance as NP-specific deletion of *Sall1* results in premature NP depletion [145,146]. PAX2 maintains and promotes NP cell identity by repressing interstitial programs [147]. Further evidence suggests that PAX2 also specifies the initial kidney

progenitor populations in the intermediate mesoderm [148] and participates in the regulation of NP differentiation [96]. Interestingly, PAX2 may have different functions in humans, as suggested by recent experiments in PAX2-deficient iPS cells where the cells were competent to epithelialize into mature nephron cell types [149]. Other noteworthy regulators of NP maintenance include p53, OSR1, EYA1, and N- and c-MYC [150–153].

Additionally, growth factor signaling via FGF and bone morphogenetic protein (BMP) maintains nephron progenitor cells as a self-renewing population, and likely acts as an upstream regulator for the above-mentioned transcription factors, though the specifics of such regulation are only emerging. FGF signaling has additional functions in nephrogenesis, where it acts together with WNT and Notch signaling to induce the timely differentiation of progenitor cells into functional nephron segments [154–167]. Thus, FGF-induced signaling expands the NP population [168,169], which is especially relevant for experiments aiming to develop in vitro tools and methods mimicking in vivo kidney development [170,171].

Rapid UB branching morphogenesis continues until mid-gestation and slows down after E15.5 [131,132]. Nephron progenitors, however, are maintained until postnatal day three in mice by an unknown mechanism that is no longer directly linked to UB branching [132,172,173]. It is clear that nephrogenesis ceases when the NP cells lose their capacity for self-renewal and instead undergo accelerated differentiation during the final postnatal burst of nephrogenesis. While the exact mechanisms of nephrogenesis cessation remains unclear, it is likely linked to metabolic changes and increased blood flow and oxygen in the niche during this time [174,175]. Elevated mTOR and reduced FGF20 could also contribute to the exit of progenitors from the NP niche [157] while HAMARTIN (encoded by the *Tsc1* gene) was recently identified as a possible regulator of nephron progenitor aging [176].

3. MAPK/ERK Signaling Guides Ureteric Bud Branching

Our own work and that reported by others shows that several cell types of the developing kidney rely on MAPK signaling [9,27,177,178]. Experiments with in vitro kidney cultures originally pointed out the functional importance of MAPK signaling for the nephron numbers and UB branching morphogenesis by showing that inhibition of the MEK proteins led to a decrease in proliferation in the UB tip cells and abnormal branch formation [27,133,177].

During UB outgrowth, MAPK/ERK activity is polarized to the side of the nephric duct, giving rise to the UB, while ERK activity is completely lost in RET knockout ducts [110]. After the formation of the initial UB, prominent but heterogeneous MAPK/ERK activation is detected both in the epithelium and the surrounding MM. Upon initiation of branching morphogenesis, the signaling gets restricted to UB tip regions [9,178]. Perhaps reflecting the heterogeneity in the UB tips, ERK activation strength, when measured as fluorescent intensity in Förster resonance energy transfer (FRET)-based biosensor for ERK, is slightly lower than that detected in nephron progenitors and precursors. The UB trunk shows clearly the weakest activation in developing kidney.

The importance of the MAPK pathway for UB branching morphogenesis was first genetically recognized by disruption of the RET docking site (Y1062), which is involved in the downstream activation of MAPK and PI3K [116,179–182]. These experiments suggested that MAPK and PI3K activation downstream of RET is essential for kidney growth and patterning through UB branching and nephric duct connection to cloaca [4,183]. However, these experiments did not specifically address the functional requirement for MAPK/ERK signaling, and thus its role in collecting duct morphogenesis remained elusive.

Genetic experiments addressing UB-specific MAPK/ERK functions involved conditional inactivation of *Mek1* and conventional deletion of *Mek2*. Loss of *Mek1* alone in the UB resulted in normal renal morphogenesis, similarly to its deletion in the whole embryo and in epidermal keratinocytes [7,57]. Also, reduction of the *Mek1/2* gene dosages (double heterozygosity and $\frac{3}{4}$ loss of alleles) in any combinations supported normal UB branching and renal differentiation, thus suggesting

a high redundancy between these protein functions in the developing kidney as well. Importantly, deletion of both *Mek1* alleles in a *Mek2* knockout background resulted in normal UB outgrowth but demonstrated a requirement of MAPK signaling for UB branch formation. Abrogation of MAPK activity in the UB results in a major decrease in novel branch formation as well as an elongation-only phenotype [178].

Characterization of the cellular and molecular causes of the branching morphogenesis defect in MAPK-deficient UBs revealed two essential processes: The first is not so surprisingly involved in regulation of cell cycle progression while the second is a clearly less-known function involved in the control of cellular adhesion [178]. The proliferation defect in mutant UBs derives from an inability to increase cyclin D1 levels, which is required for G1-to-S phase progression. Subsequently, less UB cells are in mitosis, which causes a significant reduction in the total number of UB epithelial cells. Simultaneous to the cell cycle progression defects, increased and partially mislocalized E-cadherin is detected in plasma membranes of mutant UB cells, suggesting abnormalities in adherens junctions. Moreover, MAPK-deficient UB cells exhibit immature focal adhesions and accumulation of the focal adhesion proteins to adherens junctions [178]. Thus, MAPK/ERK activity does not simply regulate proliferation in the UB to achieve normal branching and growth, but it additionally regulates the dynamic cell adhesions during kidney development. Further studies are needed to determine how MAPK signaling regulates adhesions within UB cells and among the surrounding matrix.

As detected by FRET biosensor for ERK1/2 activation, the MAPK pathway shows very dynamic activation patterns in the UB [9]. This suggests that precise control of signal strength and duration is important in specifying cellular outcomes in the developing kidney. Indeed, excess endogenous GDNF results in an opposite phenotype of that seen with UB-specific loss of MAPK activity as the tips are abnormally expanded and accompanied by short UB trunks. Thus, GDNF positively regulates collecting ductal progenitor cell expansion, and this is mediated by MAPK activity as chemical MEK inhibition rescues both the tip morphology and trunk length [122]. Accurate control of MAPK activation is further supported by ectopic expression of *Sprouty2*, a negative regulator of RTK/MAPK pathway, in the UB, which shows changes in UB branching morphogenesis [184]. Deletion of another family member, *Sprouty1* (*Spry1*), confirmed the fundamental requirement of negative regulation in control of signal strength as *Spry1* knockout embryos exhibit multiple UBs along the nephric duct [117,118]. The roles of negative feedback mechanisms are further demonstrated by the findings that GDNF induces not only *Spry1* but also the expression of two other negative regulators of the MAPK/ERK pathway, *Spred2* and *Dusp6* [112,127]. Strikingly, genetic experiments inactivating *Gdnf* or *Ret* in a *Spry1* knockout background restore the normal UB branching, thus demonstrating how a fine synergistic signaling balance dictates in normal outcome of UB outgrowth and branching [118,185,186]. The exact roles of SPRED2 and DUSP6 in the feedback process remain to be explored.

4. The Function of MAPK/ERK in Nephrogenesis

Live-imaging of embryonic kidneys isolated from FRET based ERK biosensor mice revealed that MAPK/ERK activation is amazingly heterogeneous within the NP population and strong in RVs [9]. This suggests that in addition to UB epithelium, the MAPK pathway could participate in the regulation of nephrogenesis. The activation pattern further indicates that the pathway may have multiple functions in the guidance of nephron differentiation. Unlike in the UB where activation of MAPK pathway in a given cell appears to correlate with cell cycle phase and overlaps with expression of G1 and S phase markers [178], no clear link to proliferative or non-proliferative NP cells was found in the MM and differentiating nephrons.

We inactivated MAPK/ERK signaling specifically in the NPs to reveal that MAPK/ERK activity indeed is needed in two distinct processes of nephrogenesis: The first is its essential role in the maintenance of the NP population while the second faithfully adheres to its activation in differentiating nephron precursors and involves permissive functions for progression of nephron differentiation [9]. The renal phenotype of NP-specific MAPK inactivation resembles defects reported for FGF deficiency.

Similarity of phenotypes suggests that MAPK/ERK activation mediates the function of multiple FGF ligands (FGF8/9/20) in the maintenance of the NP population, while in differentiating nephron precursors it likely acts downstream of FGF8-induced signaling [154,160,161].

In the NP-specific absence of MAPK activation, the NP niche is disorganized and faces changes in the key molecular mediator of cell-to-matrix adhesion, integrin alpha 8 (ITGA8) [9]. This suggests that MAPK activity in NP cells is essential for their communication with the extracellular matrix. Loss of MAPK activity also causes cell-intrinsic changes, as levels of PAX2 are reduced specifically in mutant NPs. The molecular details of MAPK activity's regulation of PAX2 remain to be shown, but as suggested by previous experiments [147], we showed that PAX2 is required in NPs to maintain normal ITGA8 [9]. Likewise, FGF-activated FGFR1/2 signaling not only maintains self-renewal of NP cells but also PAX2 expression in this population [113,154]. In conclusion, MAPK activity regulates niche organization and communication with the extracellular matrix.

NP-specific MAPK inactivation results in an almost complete lack of nephrons in newborn mouse pups [9]. This is in line with previous findings where MEK was chemically inhibited in developing kidneys of rat embryos and caused reduced nephrogenesis [27]. In the NP-specific absence of MAPK activity, nephrogenesis proceeds normally up to the RV stage but then halts almost exclusively. Despite the normal-looking initial steps of nephrogenesis, accumulation of an undefined PAX2⁺ nephrogenic mass resembling pretubular aggregates is occasionally observed in kidneys lacking MAPK activity in NPs. As no increase in apoptotic cells specifically in this population nor a similar increase in RV amount was detected, consequences of this phenotype remain to be elucidated [9].

The nephrogenesis arrest at the RV stage correlates with the expression pattern of *Fgf8*, of which deletion also causes a similar nephron differentiation defect [160,161]. Molecularly, MAPK activity in RVs appears to be required for distal expression of WNT signaling mediator LEF1, cell cycle regulator cyclin D1, and NOTCH ligand JAG1. Moreover, mutant RVs fail to proximally upregulate WT1. Altogether, these results suggest that loss of MAPK/ERK activity indeed blocks the progression of nephron differentiation rather than specifies nephron segment identities. Interestingly, although a strong pERK1/2 signal is detected in the connecting segment of nephrons, which fuses with the collecting duct system, no obvious defects were observed in the connections [7]. In summary, MAPK/ERK activation in RVs primes future nephron cells to be receptive for signals from WNT and NOTCH pathways that are needed to propel further differentiation.

5. Aberrant ERK Signaling and Wilms' Tumorigenesis

Abnormal MAPK/ERK signaling is detected in many different types of cancers. The research in this field has focused on adult tissues and identified that the majority of the driver mutations causing malignancies in adults are detected in BRAF and RAF, while aberrations in MEK genes are far less frequent [187]. Considerably less is known about the contribution of MAPK/ERK in pediatric tumors, of which leukemia and brain and spinal cord tumors are the most frequent cancers in children. Like neuroblastoma, which affects nerves and originates during development, Wilms' tumor (WT) is a childhood tumor of kidneys that originates from deficiency in renal differentiation. While there are some indications of aberrant MAPK/ERK activation in renal cell carcinoma [188], due to the focus of this review, we restrict our discussion to MAPK/ERK pathway in pediatric kidney cancer.

WT is thought to arise from undifferentiated MM, which remains in the kidney after birth. The incidence of WT is increased in many syndromes, and mutations leading to WT are genetically heterogeneous [189]. Common mutations causing WT include aberrations of *WT1* expression [190] and biallelic expression of insulin-like growth factor 2 (IGF2) [191]. Recent studies revealed that inactivation of RNA degradation component DIS3L2 and microRNA processing genes increasing PLAG1 expression results in increased IGF2 expression through different mechanisms [192,193]. Thus, an increase in IGF2 appears to act as a common mechanism to maintain undifferentiated MM in postnatal kidneys, but its influence to intracellular pathways requires further characterization. On one hand, increased PLAG1 expression associates with increased mTOR signaling [193], while augmented pERK1/2 as a response

to increased IGF2 is observed in a mouse model of Wilm's tumor [194]. Furthermore, ERK signaling is linked to WT development as upregulation of pERK1/2 occurs in a subset of human tumors [195], but the relevance of activated MAPK/ERK signaling as a driving cause of WT remains to be studied.

6. Future Outlook

The prevalence of chronic kidney disease, end-stage renal disease, and acute kidney injury is growing rapidly due to an aging population and an increased incidence of diseases like diabetes. Thus, kidney diseases, whether acquired or congenital, are not only a huge personal agony for patients but also their treatment is an increasing challenge for health care systems worldwide. Despite the clinical demands for the treatment of renal diseases, current therapies, dialysis and transplantation, have limitations in addressing these needs, and thus development of novel strategies to tackle kidney diseases are grievously sought after. High hopes were awoken a few years back when the first kidney organoids were generated from pluripotent stem cells [196–198]. The current state of the art, though successful in generation of kidney lineage from pluripotent stem cells, is inefficient in differentiating and propagating collecting duct and nephron progenitors, and thus gives rise to organoids of very small size. Limited knowledge of the molecular and cellular mechanisms regulating expansion of renal progenitor cells and their timely differentiation into functional structures has hampered the progress of taking organoids to the next level. Recent advances in understanding the functions of intracellular pathways, such as MAPK/ERK activation, ought to be next taken into account in organoid differentiation as well as in attempts aiming to reveal genetic causes of congenital kidney diseases. This is especially important in the current situation where organoid differentiation protocols heavily rely on protein supplementation rather than utilization of defined chemical compounds that would allow more precise activation–inactivation control over the given signaling pathway.

Surprisingly few studies have genetically addressed the function of intracellular pathways in developing kidney and thus limited information is available for evaluation of their specific functions. Systematic and tissue-specific inactivation of cascades induced downstream of RTK activation together with the generation of disease-specific mouse models bearing the known disease-causing mutations are the key next steps required for advancing therapeutic interventions in kidney diseases. Therefore, continuous basic research that generates novel information of kidney development together with systematic testing of the nephrogenic and diagnostic potential of these findings is needed for advancing kidney health in the future.

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Abbreviations

CAKUT	Congenital anomalies of the kidney and urinary tract
CM	Cap mesenchyme
EGF	Epidermal growth factor
ERK	Extracellular signal-regulated kinase
ESC	Embryonic stem cell
ETS	E26 transformation-specific
ETV	ETS translocation variant
EYA1	Eyes absent homolog 1
FGF	Fibroblast growth factor
FGFR	Fibroblast growth factor receptor
FRET	Förster resonance energy transfer
GDNF	Glial cell line-derived neurotrophic factor

GFRa1	GDNF family receptor alpha-1
IGF2	Insulin-like growth factor 2
IL-1 β	Interleukin 1 beta
ITGA8	Integrin alpha 8
JNK	c-Jun N-terminal kinase
MAPK	Mitogen-activated protein kinase
MEK	MAPK/ERK kinase
MM	Metanephric mesenchyme
mTOR	Mammalian target of rapamycin
NP	Nephron progenitor
ODD1	Odd-skipped related 1
PAX	Paired box gene
PI3K	Phosphoinositide 3-kinase
PLC γ /PKC	Phospholipase C gamma/protein kinase C
RTK	Receptor tyrosine kinase
RV	Renal vesicle
SIX	Sineoculis homeobox homolog
TNF α	Tumor necrosis factor alpha
UB	Ureteric bud
WT1	Wilms' tumor 1

References

1. Song, R.; El-Dahr, S.S.; Yosypiv, I.V. Receptor tyrosine kinases in kidney development. *J. Signal Transduct.* **2011**, *2011*, 869281. [[CrossRef](#)]
2. Kurtzeborn, K.; Cebrian, C.; Kuure, S. Regulation of Renal Differentiation by Trophic Factors. *Front. Physiol.* **2018**, *9*, 1588. [[CrossRef](#)] [[PubMed](#)]
3. Walker, K.A.; Sims-Lucas, S.; Bates, C.M. Fibroblast growth factor receptor signaling in kidney and lower urinary tract development. *Pediatric Nephrol.* **2016**, *31*, 885–895. [[CrossRef](#)]
4. Costantini, F. GDNF/Ret signaling and renal branching morphogenesis: From mesenchymal signals to epithelial cell behaviors. *Organogenesis* **2010**, *6*, 252–262. [[CrossRef](#)]
5. Boucherat, O.; Nadeau, V.; Berube-Simard, F.A.; Charron, J.; Jeannotte, L. Crucial requirement of ERK/MAPK signaling in respiratory tract development. *Development* **2014**, *141*, 3197–3211. [[CrossRef](#)]
6. Parada, C.; Han, D.; Grimaldi, A.; Sarrion, P.; Park, S.S.; Pelikan, R.; Sanchez-Lara, P.A.; Chai, Y. Disruption of the ERK/MAPK pathway in neural crest cells as a potential cause of Pierre Robin sequence. *Development* **2015**, *142*, 3734–3745. [[CrossRef](#)]
7. Scholl, F.A.; Dumesic, P.A.; Barragan, D.I.; Harada, K.; Bissonauth, V.; Charron, J.; Khavari, P.A. Mek1/2 MAPK kinases are essential for Mammalian development, homeostasis, and Raf-induced hyperplasia. *Dev. Cell* **2007**, *12*, 615–629. [[PubMed](#)]
8. Newbern, J.; Zhong, J.; Wickramasinghe, R.S.; Li, X.; Wu, Y.; Samuels, I.; Cherosky, N.; Karlo, J.C.; O'Loughlin, B.; Wikenheiser, J.; et al. Mouse and human phenotypes indicate a critical conserved role for ERK2 signaling in neural crest development. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17115–17120. [[CrossRef](#)]
9. Ihermann-Hella, A.; Hirashima, T.; Kupari, J.; Kurtzeborn, K.; Li, H.; Kwon, H.N.; Cebrian, C.; Soofi, A.; Dapkunas, A.; Miinalainen, I.; et al. Dynamic MAPK/ERK Activity Sustains Nephron Progenitors through Niche Regulation and Primes Precursors for Differentiation. *Stem Cell Rep.* **2018**, *11*, 912–928. [[CrossRef](#)] [[PubMed](#)]
10. Lemmon, M.A.; Schlessinger, J. Cell signaling by receptor tyrosine kinases. *Cell* **2010**, *141*, 1117–1134. [[CrossRef](#)] [[PubMed](#)]
11. Schlessinger, J. Receptor tyrosine kinases: Legacy of the first two decades. *Cold Spring Harb. Perspect. Biol.* **2014**, *6*, a008912. [[CrossRef](#)]
12. Regad, T. Targeting RTK Signaling Pathways in Cancer. *Cancers* **2015**, *7*, 1758–1784. [[CrossRef](#)] [[PubMed](#)]
13. Jiménez, G.; Shvartsman, S.Y.; Paroush, Z.E. The Capicua repressor—A general sensor of RTK signaling in development and disease. *J. Cell Sci.* **2012**, *125*, 1383–1391. [[CrossRef](#)]

14. Drew, B.A.; Burow, M.E.; Beckman, B.S. MEK5/ERK5 pathway: The first fifteen years. *Biochim. Biophys. Acta* **2012**, *1825*, 37–48. [[CrossRef](#)]
15. Zarubin, T.; Han, J. Activation and signaling of the p38 MAP kinase pathway. *Cell Res.* **2005**, *15*, 11–18. [[CrossRef](#)] [[PubMed](#)]
16. Chang, L.; Karin, M. Mammalian MAP kinase signalling cascades. *Nature* **2001**, *410*, 37–40. [[CrossRef](#)]
17. Nishida, E.; Gotoh, Y. The Map Kinase Cascade Is Essential for Diverse Signal Transduction Pathways. *Trends Biochem. Sci.* **1993**, *18*, 128–131. [[CrossRef](#)]
18. Robinson, M.J.; Cobb, M.H. Mitogen-activated protein kinase pathways. *Curr. Opin. Cell Biol.* **1997**, *9*, 180–186. [[CrossRef](#)]
19. Abe, J.; Kusuhara, M.; Ulevitch, R.J.; Berk, B.C.; Lee, J.D. Big mitogen-activated protein kinase 1 (BMK1) is a redox-sensitive kinase. *J. Biol. Chem.* **1996**, *271*, 16586–16590. [[CrossRef](#)] [[PubMed](#)]
20. Plotnikov, A.; Zehorai, E.; Procaccia, S.; Seger, R. The MAPK cascades: Signaling components, nuclear roles and mechanisms of nuclear translocation. *Biochim. Biophys. Acta* **2011**, *1813*, 1619–1633. [[CrossRef](#)] [[PubMed](#)]
21. Nishimoto, S.; Nishida, E. MAPK signalling: ERK5 versus ERK1/2. *EMBO Rep.* **2006**, *7*, 782–786. [[CrossRef](#)]
22. Johnson, L.; Greenbaum, D.; Cichowski, K.; Mercer, K.; Murphy, E.; Schmitt, E.; Bronson, R.T.; Umanoff, H.; Edelman, W.; Kucherlapati, R. K-ras is an essential gene in the mouse with partial functional overlap with N-ras. *Genes Dev.* **1997**, *11*, 2468–2481. [[CrossRef](#)]
23. Tan, P.B.; Kim, S.K. Signaling specificity: The RTK/RAS/MAP kinase pathway in metazoans. *Trends Genet. TIG* **1999**, *15*, 145–149. [[CrossRef](#)]
24. Omori, S.; Hida, M.; Ishikura, K.; Kuramochi, S.; Awazu, M. Expression of mitogen-activated protein kinase family in rat renal development. *Kidney Int.* **2000**, *58*, 27–37. [[CrossRef](#)]
25. Osafune, K.; Takasato, M.; Kispert, A.; Asashima, M.; Nishinakamura, R. Identification of multipotent progenitors in the embryonic mouse kidney by a novel colony-forming assay. *Development* **2006**, *133*, 151–161. [[CrossRef](#)]
26. Blank, U.; Brown, A.; Adams, D.C.; Karolak, M.J.; Oxburgh, L. BMP7 promotes proliferation of nephron progenitor cells via a JNK-dependent mechanism. *Development* **2009**, *136*, 3557–3566. [[CrossRef](#)]
27. Hida, M.; Omori, S.; Awazu, M. ERK and p38 MAP kinase are required for rat renal development. *Kidney Int.* **2002**, *61*, 1252–1262. [[CrossRef](#)]
28. Awazu, M.; Omori, S.; Hida, M. MAP kinase in renal development. *Nephrol. Dial. Transplant. Off. Publ. Eur. Dial. Transpl. Assoc.-Eur. Ren. Assoc.* **2002**, *17* (Suppl. 9), 5–7. [[CrossRef](#)]
29. McKay, M.M.; Morrison, D.K. Integrating signals from RTKs to ERK/MAPK. *Oncogene* **2007**, *26*, 3113–3121. [[CrossRef](#)]
30. Roskoski, R., Jr. MEK1/2 dual-specificity protein kinases: Structure and regulation. *Biochem. Biophys. Res. Commun.* **2012**, *417*, 5–10. [[CrossRef](#)]
31. Roskoski, R., Jr. ERK1/2 MAP kinases: Structure, function, and regulation. *Pharmacol. Res.* **2012**, *66*, 105–143. [[CrossRef](#)]
32. Pouysségur, J.; Lenormand, P. ERK1 and ERK2 map kinases: Specific roles or functional redundancy? *Front. Cell Dev. Biol.* **2016**, *4*, 53.
33. Brunet, A.; Roux, D.; Lenormand, P.; Dowd, S.; Keyse, S.; Pouysségur, J. Nuclear translocation of p42/p44 mitogen-activated protein kinase is required for growth factor-induced gene expression and cell cycle entry. *EMBO J.* **1999**, *18*, 664–674. [[CrossRef](#)]
34. Johnson, H.E.; Toettcher, J.E. Signaling Dynamics Control Cell Fate in the Early Drosophila Embryo. *Dev. Cell* **2019**, *48*, 361–370. [[CrossRef](#)]
35. Ma, X.; Chen, H.; Chen, L. A dual role of Erk signaling in embryonic stem cells. *Exp. Hematol.* **2016**, *44*, 151–156. [[CrossRef](#)]
36. Chen, H.; Guo, R.; Zhang, Q.; Guo, H.; Yang, M.; Wu, Z.; Gao, S.; Liu, L.; Chen, L. Erk signaling is indispensable for genomic stability and self-renewal of mouse embryonic stem cells. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, E5936–E5943. [[CrossRef](#)]
37. Caunt, C.J.; Sale, M.J.; Smith, P.D.; Cook, S.J. MEK1 and MEK2 inhibitors and cancer therapy: The long and winding road. *Nat. Rev. Cancer* **2015**, *15*, 577–592. [[CrossRef](#)]
38. Shrestha, G.; MacNeil, S.M.; McQuerry, J.A.; Jenkins, D.F.; Sharma, S.; Bild, A.H. The Value of Genomics in Dissecting the RAS-Network and in Guiding Therapeutics for RAS-Driven Cancers. In *Seminars in Cell & Developmental Biology*; Academic Press: Cambridge, MA, USA, 2016; pp. 108–117.

39. Kidger, A.M.; Siphthorp, J.; Cook, S.J. ERK1/2 inhibitors: New weapons to inhibit the RAS-regulated RAF-MEK1/2-ERK1/2 pathway. *Pharmacol. Ther.* **2018**, *187*, 45–60. [[CrossRef](#)]
40. Brown, N.A.; Furtado, L.V.; Betz, B.L.; Kiel, M.J.; Weigelin, H.C.; Lim, M.S.; Elenitoba-Johnson, K.S. High prevalence of somatic MAP2K1 mutations in BRAF V600E–Negative Langerhans cell histiocytosis. *Blood* **2014**, *124*, 1655–1658. [[CrossRef](#)]
41. Yang, S.H.; Sharrocks, A.D.; Whitmarsh, A.J. MAP kinase signalling cascades and transcriptional regulation. *Gene* **2013**, *513*, 1–13. [[CrossRef](#)]
42. Kim, E.K.; Choi, E.J. Pathological roles of MAPK signaling pathways in human diseases. *Biochim. Biophys. Acta* **2010**, *1802*, 396–405. [[CrossRef](#)]
43. Kyriakis, J.M.; Banerjee, P.; Nikolakaki, E.; Dai, T.; Rubie, E.A.; Ahmad, M.F.; Avruch, J.; Woodgett, J.R. The stress-activated protein kinase subfamily of c-Jun kinases. *Nature* **1994**, *369*, 156–160. [[CrossRef](#)]
44. Weston, C.R.; Davis, R.J. The JNK signal transduction pathway. *Curr. Opin. Genet. Dev.* **2002**, *12*, 14–21. [[CrossRef](#)]
45. Cuadrado, A.; Nebreda, A.R. Mechanisms and functions of p38 MAPK signalling. *Biochem. J.* **2010**, *429*, 403–417. [[CrossRef](#)]
46. Nithianandarajah-Jones, G.N.; Wilm, B.; Goldring, C.E.; Muller, J.; Cross, M.J. ERK5: Structure, regulation and function. *Cell. Signal.* **2012**, *24*, 2187–2196. [[CrossRef](#)]
47. Yan, L.; Carr, J.; Ashby, P.R.; Murry-Tait, V.; Thompson, C.; Arthur, J.S. Knockout of ERK5 causes multiple defects in placental and embryonic development. *BMC Dev. Biol.* **2003**, *3*, 11. [[CrossRef](#)]
48. Wojnowski, L.; Stancato, L.F.; Zimmer, A.M.; Hahn, H.; Beck, T.W.; Larner, A.C.; Rapp, U.R.; Zimmer, A. Craf-1 protein kinase is essential for mouse development. *Mech. Dev.* **1998**, *76*, 141–149. [[CrossRef](#)]
49. Wojnowski, L.; Zimmer, A.M.; Beck, T.W.; Hahn, H.; Bernal, R.; Rapp, U.R.; Zimmer, A. Endothelial apoptosis in Braf-deficient mice. *Nat. Genet.* **1997**, *16*, 293–297. [[CrossRef](#)]
50. Galabova-Kovacs, G.; Matzen, D.; Piazzolla, D.; Meissl, K.; Plyushch, T.; Chen, A.P.; Silva, A.; Baccarini, M. Essential role of B-Raf in ERK activation during extraembryonic development. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 1325–1330. [[CrossRef](#)]
51. Mikula, M.; Schreiber, M.; Husak, Z.; Kucerova, L.; R uth, J.; Wieser, R.; Zatloukal, K.; Beug, H.; Wagner, E.F.; Baccarini, M. Embryonic lethality and fetal liver apoptosis in mice lacking the c-raf-1 gene. *EMBO J.* **2001**, *20*, 1952–1962. [[CrossRef](#)]
52. Hatano, N.; Mori, Y.; Oh-hora, M.; Kosugi, A.; Fujikawa, T.; Nakai, N.; Niwa, H.; Miyazaki, J.-I.; Hamaoka, T.; Ogata, M. Essential role for ERK2 mitogen-activated protein kinase in placental development. *Genes Cells* **2003**, *8*, 847–856. [[CrossRef](#)] [[PubMed](#)]
53. Galabova-Kovacs, G.; Kolbus, A.; Matzen, D.; Meissl, K.; Piazzolla, D.; Rubiolo, C.; Steinitz, K.; Baccarini, M. ERK and beyond: Insights from B-Raf and Raf-1 conditional knockouts. *Cell Cycle* **2006**, *5*, 1514–1518. [[CrossRef](#)] [[PubMed](#)]
54. Bueno, O.F.; De Windt, L.J.; Tymitz, K.M.; Witt, S.A.; Kimball, T.R.; Kleivitsky, R.; Hewett, T.E.; Jones, S.P.; Lefer, D.J.; Peng, C.F. The MEK1–ERK1/2 signaling pathway promotes compensated cardiac hypertrophy in transgenic mice. *EMBO J.* **2000**, *19*, 6341–6350. [[CrossRef](#)]
55. Krens, S.G.; Spaink, H.P.; Snaar-Jagalska, B.E. Functions of the MAPK family in vertebrate-development. *FEBS Lett.* **2006**, *580*, 4984–4990. [[CrossRef](#)]
56. Giroux, S.; Tremblay, M.; Bernard, D.; Cadrin-Girard, J.F.; Aubry, S.; Larouche, L.; Rousseau, S.; Huot, J.; Landry, J.; Jeannotte, L.; et al. Embryonic death of Mek1-deficient mice reveals a role for this kinase in angiogenesis in the labyrinthine region of the placenta. *Curr. Biol.* **1999**, *9*, 369–372. [[CrossRef](#)]
57. Bissonauth, V.; Roy, S.; Gravel, M.; Guillemette, S.; Charron, J. Requirement for Map2k1 (Mek1) in extra-embryonic ectoderm during placentogenesis. *Development* **2006**, *133*, 3429–3440. [[CrossRef](#)] [[PubMed](#)]
58. Belanger, L.F.; Roy, S.; Tremblay, M.; Brott, B.; Steff, A.M.; Mourad, W.; Hugo, P.; Erikson, R.; Charron, J. Mek2 is dispensable for mouse growth and development. *Mol. Cell. Biol.* **2003**, *23*, 4778–4787. [[CrossRef](#)]
59. Nadeau, V.; Charron, J. Essential role of the ERK/MAPK pathway in blood-placental barrier formation. *Development* **2014**, *141*, 2825–2837. [[CrossRef](#)] [[PubMed](#)]
60. Nadeau, V.; Guillemette, S.; Belanger, L.F.; Jacob, O.; Roy, S.; Charron, J. Map2k1 and Map2k2 genes contribute to the normal development of syncytiotrophoblasts during placentation. *Development* **2009**, *136*, 1363–1374. [[CrossRef](#)] [[PubMed](#)]

61. Aoidi, R.; Maltais, A.; Charron, J. Functional redundancy of the kinases MEK1 and MEK2: Rescue of the Mek1 mutant phenotype by Mek2 knock-in reveals a protein threshold effect. *Sci. Signal.* **2016**, *9*, ra9. [[CrossRef](#)] [[PubMed](#)]
62. Catalanotti, F.; Reyes, G.; Jesenberger, V.; Galabova-Kovacs, G.; de Matos Simoes, R.; Carugo, O.; Baccarini, M. A Mek1–Mek2 heterodimer determines the strength and duration of the Erk signal. *Nat. Struct. Mol. Biol.* **2009**, *16*, 294. [[CrossRef](#)]
63. Kelleher, R.J., III; Govindarajan, A.; Jung, H.-Y.; Kang, H.; Tonegawa, S. Translational control by MAPK signaling in long-term synaptic plasticity and memory. *Cell* **2004**, *116*, 467–479. [[CrossRef](#)]
64. Xing, L.; Larsen, R.S.; Bjorklund, G.R.; Li, X.; Wu, Y.; Philpot, B.D.; Snider, W.D.; Newbern, J.M. Layer specific and general requirements for ERK/MAPK signaling in the developing neocortex. *eLife* **2016**, *5*, e11123. [[CrossRef](#)]
65. Saba-El-Leil, M.; Vella, F.D.; Vernay, B.; Voisin, L.; Chen, L.; Labrecque, N.; Ang, S.L.; Meloche, S. An essential function of the mitogen-activated protein kinase Erk2 in mouse trophoblast development. *EMBO Rep.* **2003**, *4*, 964–968. [[CrossRef](#)]
66. Pagès, G.; Guérin, S.; Grall, D.; Bonino, F.; Smith, A.; Anjuere, F.; Auberger, P.; Pouyssegur, J. Defective thymocyte maturation in p44 MAP kinase (Erk 1) knockout mice. *Science* **1999**, *286*, 1374–1377.
67. Yao, Y.; Li, W.; Wu, J.W.; Germann, U.A.; Su, M.S.S.; Kuida, K.; Boucher, D.M. Extracellular signal-regulated kinase 2 is necessary for mesoderm differentiation. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 12759–12764. [[CrossRef](#)]
68. Goplen, N.; Karim, Z.; Guo, L.; Zhuang, Y.H.; Huang, H.; Gorska, M.M.; Gelfand, E.; Pages, G.; Pouyssegur, J.; Alam, R. ERK1 is important for Th2 differentiation and development of experimental asthma. *FASEB J.* **2012**, *26*, 1934–1945. [[CrossRef](#)]
69. Bost, F.; Aouadi, M.; Caron, L.; Even, P.; Belmonte, N.; Prot, M.; Dani, C.; Hofman, P.; Pagès, G.; Pouyssegur, J. The extracellular signal-Regulated kinase isoform ERK1 is specifically required for in vitro and in vivo adipogenesis. *Diabetes* **2005**, *54*, 402–411. [[CrossRef](#)]
70. Bourcier, C.; Jacquelin, A.; Hess, J.; Peyrottes, I.; Angel, P.; Hofman, P.; Auberger, P.; Pouyssegur, J.; Pages, G. P44 mitogen-activated protein kinase (extracellular signal-regulated kinase 1)-dependent signaling contributes to epithelial skin carcinogenesis. *Cancer Res.* **2006**, *66*, 2700–2707. [[CrossRef](#)]
71. Mazzucchelli, C.; Vantaggiato, C.; Ciamei, A.; Fasano, S.; Pakhotin, P.; Krezel, W.; Welzl, H.; Wolfer, D.P.; Pages, G.; Valverde, O.; et al. Knockout of ERK1 MAP kinase enhances synaptic plasticity in the striatum and facilitates striatal-mediated learning and memory. *Neuron* **2002**, *34*, 807–820. [[CrossRef](#)]
72. Selcher, J.C.; Nekrasova, T.; Paylor, R.; Landreth, G.E.; Sweatt, J.D. Mice lacking the ERK1 isoform of MAP kinase are unimpaired in emotional learning. *Learn. Mem.* **2001**, *8*, 11–19. [[CrossRef](#)]
73. Satoh, Y.; Endo, S.; Ikeda, T.; Yamada, K.; Ito, M.; Kuroki, M.; Hiramoto, T.; Imamura, O.; Kobayashi, Y.; Watanabe, Y.; et al. Extracellular signal-regulated kinase 2 (ERK2) knockdown mice show deficits in long-term memory; ERK2 has a specific function in learning and memory. *J. Neurosci.* **2007**, *27*, 10765–10776. [[CrossRef](#)]
74. Saba-El-Leil, M.K.; Fremin, C.; Meloche, S. Redundancy in the World of MAP Kinases: All for One. *Front. Cell Dev. Biol.* **2016**, *4*, 67. [[CrossRef](#)]
75. Fremin, C.; Saba-El-Leil, M.K.; Levesque, K.; Ang, S.L.; Meloche, S. Functional Redundancy of ERK1 and ERK2 MAP Kinases during Development. *Cell Rep.* **2015**, *12*, 913–921. [[CrossRef](#)]
76. Lefloch, R.; Pouyssegur, J.; Lenormand, P. Single and combined silencing of ERK1 and ERK2 reveals their positive contribution to growth signaling depending on their expression levels. *Mol. Cell. Biol.* **2008**, *28*, 511–527. [[CrossRef](#)]
77. Fischer, A.M.; Katayama, C.D.; Pages, G.; Pouyssegur, J.; Hedrick, S.M. The role of erk1 and erk2 in multiple stages of T cell development. *Immunity* **2005**, *23*, 431–443. [[CrossRef](#)]
78. Satoh, Y.; Kobayashi, Y.; Takeuchi, A.; Pages, G.; Pouyssegur, J.; Kazama, T. Deletion of ERK1 and ERK2 in the CNS causes cortical abnormalities and neonatal lethality: Erk1 deficiency enhances the impairment of neurogenesis in Erk2-deficient mice. *J. Neurosci. Off. J. Soc. Neurosci.* **2011**, *31*, 1149–1155. [[CrossRef](#)]
79. Chan, G.; Gu, S.; Neel, B.G. Erk1 and Erk2 are required for maintenance of hematopoietic stem cells and adult hematopoiesis. *Blood* **2013**, *121*, 3594–3598. [[CrossRef](#)]
80. Staser, K.; Park, S.J.; Rhodes, S.D.; Zeng, Y.; He, Y.Z.; Shew, M.A.; Gehlhausen, J.R.; Cerabona, D.; Menon, K.; Chen, S.; et al. Normal hematopoiesis and neurofibromin-deficient myeloproliferative disease require Erk. *J. Clin. Investig.* **2013**, *123*, 329–334. [[CrossRef](#)]

81. Saxén, L.; Saxén, L. *Organogenesis of the Kidney*; Cambridge University Press: Cambridge, UK, 1987; Volume 19.
82. Lin, Y.; Zhang, S.; Rehn, M.; Itaranta, P.; Tuukkanen, J.; Heljasvaara, R.; Peltoketo, H.; Pihlajaniemi, T.; Vainio, S. Induced repatterning of type XVIII collagen expression in ureter bud from kidney to lung type: Association with sonic hedgehog and ectopic surfactant protein C. *Development* **2001**, *128*, 1573–1585.
83. Taguchi, A.; Kaku, Y.; Ohmori, T.; Sharmin, S.; Ogawa, M.; Sasaki, H.; Nishinakamura, R. Redefining the in vivo origin of metanephric nephron progenitors enables generation of complex kidney structures from pluripotent stem cells. *Cell Stem Cell* **2014**, *14*, 53–67. [[CrossRef](#)]
84. Takasato, M.; Little, M.H. The origin of the mammalian kidney: Implications for recreating the kidney in vitro. *Development* **2015**, *142*, 1937–1947. [[CrossRef](#)]
85. Cullen-McEwen, L.A.; Sutherland, M.R.; Black, M.J. The Human Kidney: Parallels in Structure, Spatial Development, and Timing of Nephrogenesis. In *Kidney Development, Disease, Repair and Regeneration*, 1st ed.; Little, M., Ed.; Academic Press: Cambridge, MA, USA, 2016; pp. 27–40.
86. Costantini, F.; Kopan, R. Patterning a complex organ: Branching morphogenesis and nephron segmentation in kidney development. *Dev. Cell* **2010**, *18*, 698–712. [[CrossRef](#)]
87. Little, M.H.; McMahon, A.P. Mammalian kidney development: Principles, progress, and projections. *Cold Spring Harb. Perspect. Biol.* **2012**, *4*, a008300. [[CrossRef](#)]
88. Combes, A.N.; Davies, J.A.; Little, M.H. Cell–cell interactions driving kidney morphogenesis. In *Current Topics in Developmental Biology*; Academic Press: Cambridge, MA, USA, 2015; Volume 112, pp. 467–508.
89. Jacob, M.; Yusuf, F.; Jacob, H.J. Development, differentiation and derivatives of the Wolffian and Müllerian ducts. In *The Human Embryo*; InTechOpen: London, UK, 2012.
90. Xu, P.X.; Adams, J.; Peters, H.; Brown, M.C.; Heaney, S.; Maas, R. Eya1-deficient mice lack ears and kidneys and show abnormal apoptosis of organ primordia. *Nat. Genet.* **1999**, *23*, 113–117. [[CrossRef](#)]
91. Sajithlal, G.; Zou, D.; Silvius, D.; Xu, P.X. Eya 1 acts as a critical regulator for specifying the metanephric mesenchyme. *Dev. Biol.* **2005**, *284*, 323–336. [[CrossRef](#)]
92. Li, X.; Ohgi, K.A.; Zhang, J.; Krones, A.; Bush, K.T.; Glass, C.K.; Nigam, S.K.; Aggarwal, A.K.; Maas, R.; Rose, D.W. Eya protein phosphatase activity regulates Six1–Dach–Eya transcriptional effects in mammalian organogenesis. *Nature* **2003**, *426*, 247–254. [[CrossRef](#)]
93. Kobayashi, H.; Kawakami, K.; Asashima, M.; Nishinakamura, R. Six1 and Six4 are essential for Gdnf expression in the metanephric mesenchyme and ureteric bud formation, while Six1 deficiency alone causes mesonephric-tubule defects. *Mech. Dev.* **2007**, *124*, 290–303. [[CrossRef](#)]
94. Xu, P.X.; Zheng, W.M.; Huang, L.; Maire, P.; Laclef, C.; Silvius, D. Six1 is required for the early organogenesis of mammalian kidney. *Development* **2003**, *130*, 3085–3094. [[CrossRef](#)]
95. Wang, Q.R.; Lan, Y.; Cho, E.S.; Maltby, K.M.; Jiang, R.L. Odd-skipped related 1 (Odd1) is an essential regulator of heart and urogenital development. *Dev. Biol.* **2005**, *288*, 582–594. [[CrossRef](#)]
96. Narlis, M.; Grote, D.; Gaitan, Y.; Boualia, S.K.; Bouchard, M. Pax2 and pax8 regulate branching morphogenesis and nephron differentiation in the developing kidney. *J. Am. Soc. Nephrol.* **2007**, *18*, 1121–1129. [[CrossRef](#)]
97. Dressler, G.R. Patterning and early cell lineage decisions in the developing kidney: The role of Pax genes. *Pediatric Nephrol.* **2011**, *26*, 1387–1394. [[CrossRef](#)]
98. Brophy, P.D.; Ostrom, L.; Lang, K.M.; Dressler, G.R. Regulation of ureteric bud outgrowth by Pax2-dependent activation of the glial derived neurotrophic factor gene. *Development* **2001**, *128*, 4747–4756.
99. Bouchard, M.; Souabni, A.; Mandler, M.; Neubuser, A.; Busslinger, M. Nephric lineage specification by Pax2 and Pax8. *Genes Dev.* **2002**, *16*, 2958–2970. [[CrossRef](#)]
100. Krause, M.; Rak-Raszewska, A.; Pietilä, I.; Quaggin, S.; Vainio, S. Signaling during kidney development. *Cells* **2015**, *4*, 112–132. [[CrossRef](#)]
101. Hellmich, H.L.; Kos, L.; Cho, E.S.; Mahon, K.A.; Zimmer, A. Embryonic expression of glial cell-line derived neurotrophic factor (GDNF) suggests multiple developmental roles in neural differentiation and epithelial-mesenchymal interactions. *Mech. Dev.* **1996**, *54*, 95–105. [[CrossRef](#)]
102. Golden, J.P.; DeMaro, J.A.; Osborne, P.A.; Milbrandt, J.; Johnson, E.M., Jr. Expression of neurturin, GDNF, and GDNF family-receptor mRNA in the developing and mature mouse. *Exp. Neurol.* **1999**, *158*, 504–528. [[CrossRef](#)]
103. Pachnis, V.; Mankoo, B.; Costantini, F. Expression of the c-ret proto-oncogene during mouse embryogenesis. *Development* **1993**, *119*, 1005–1017.

104. Sainio, K.; Suvanto, P.; Davies, J.; Wartiovaara, J.; Wartiovaara, K.; Saarma, M.; Arumae, U.; Meng, X.J.; Lindahl, M.; Pachnis, V.; et al. Glial-cell-line-derived neurotrophic factor is required for bud initiation from ureteric epithelium. *Development* **1997**, *124*, 4077–4087.
105. Moore, M.W.; Klein, R.D.; Farinas, I.; Sauer, H.; Armanini, M.; Phillips, H.; Reichardt, L.F.; Ryan, A.M.; Carver-Moore, K.; Rosenthal, A. Renal and neuronal abnormalities in mice lacking GDNF. *Nature* **1996**, *382*, 76–79. [[CrossRef](#)]
106. Pichel, J.G.; Shen, L.; Sheng, H.Z.; Granholm, A.C.; Drago, J.; Grinberg, A.; Lee, E.J.; Huang, S.P.; Saarma, M.; Hoffer, B.J.; et al. Defects in enteric innervation and kidney development in mice lacking GDNF. *Nature* **1996**, *382*, 73–76. [[CrossRef](#)]
107. Sanchez, M.P.; Silos-Santiago, I.; Frisen, J.; He, B.; Lira, S.A.; Barbacid, M. Renal agenesis and the absence of enteric neurons in mice lacking GDNF. *Nature* **1996**, *382*, 70–73. [[CrossRef](#)]
108. Schuchardt, A.; D'Agati, V.; Larsson-Blomberg, L.; Costantini, F.; Pachnis, V. Defects in the kidney and enteric nervous system of mice lacking the tyrosine kinase receptor Ret. *Nature* **1994**, *367*, 380–383. [[CrossRef](#)]
109. Cacalano, G.; Farinas, I.; Wang, L.C.; Hagler, K.; Forgie, A.; Moore, M.; Armanini, M.; Phillips, H.; Ryan, A.M.; Reichardt, L.F.; et al. GFRalpha1 is an essential receptor component for GDNF in the developing nervous system and kidney. *Neuron* **1998**, *21*, 53–62. [[CrossRef](#)]
110. Chi, X.; Michos, O.; Shakya, R.; Riccio, P.; Enomoto, H.; Licht, J.D.; Asai, N.; Takahashi, M.; Ohgami, N.; Kato, M.; et al. Ret-dependent cell rearrangements in the Wolffian duct epithelium initiate ureteric bud morphogenesis. *Dev. Cell* **2009**, *17*, 199–209. [[CrossRef](#)]
111. Kuure, S.; Chi, X.; Lu, B.; Costantini, F. The transcription factors Etv4 and Etv5 mediate formation of the ureteric bud tip domain during kidney development. *Development* **2010**, *137*, 1975–1979. [[CrossRef](#)]
112. Lu, B.C.; Cebrian, C.; Chi, X.; Kuure, S.; Kuo, R.; Bates, C.M.; Arber, S.; Hassell, J.; MacNeil, L.; Hoshi, M.; et al. Etv4 and Etv5 are required downstream of GDNF and Ret for kidney branching morphogenesis. *Nat. Genet.* **2009**, *41*, 1295–1302. [[CrossRef](#)]
113. Poladia, D.P.; Kish, K.; Kutay, B.; Hains, D.; Kegg, H.; Zhao, H.; Bates, C.M. Role of fibroblast growth factor receptors 1 and 2 in the metanephric mesenchyme. *Dev. Biol.* **2006**, *291*, 325–339. [[CrossRef](#)]
114. Zhao, H.; Kegg, H.; Grady, S.; Truong, H.T.; Robinson, M.L.; Baum, M.; Bates, C.M. Role of fibroblast growth factor receptors 1 and 2 in the ureteric bud. *Dev. Biol.* **2004**, *276*, 403–415. [[CrossRef](#)]
115. Pichel, J.; Shen, L.; Sheng, H.; Granholm, A.-C.; Drago, J.; Grinberg, A.; Lee, E.; Huang, S.; Saarma, M.; Hoffer, B. GDNF is required for kidney development and enteric innervation. In *Cold Spring Harbor Symposia on Quantitative Biology*; Cold Spring Harbor Laboratory Press: New York, NY, USA, 1996; pp. 445–457.
116. Jain, S.; Encinas, M.; Johnson, E.M., Jr.; Milbrandt, J. Critical and distinct roles for key RET tyrosine docking sites in renal development. *Genes Dev.* **2006**, *20*, 321–333. [[CrossRef](#)]
117. Basson, M.A.; Watson-Johnson, J.; Shakya, R.; Akbulut, S.; Hyink, D.; Costantini, F.D.; Wilson, P.D.; Mason, I.J.; Licht, J.D. Branching morphogenesis of the ureteric epithelium during kidney development is coordinated by the opposing functions of GDNF and Sprouty1. *Dev. Biol.* **2006**, *299*, 466–477. [[CrossRef](#)]
118. Basson, M.A.; Akbulut, S.; Watson-Johnson, J.; Simon, R.; Carroll, T.J.; Shakya, R.; Gross, I.; Martin, G.R.; Lufkin, T.; McMahon, A.P.; et al. Sprouty1 is a critical regulator of GDNF/RET-mediated kidney induction. *Dev. Cell* **2005**, *8*, 229–239. [[CrossRef](#)]
119. Kumar, A.; Kopra, J.; Varendi, K.; Porokuokka, L.L.; Panhelainen, A.; Kuure, S.; Marshall, P.; Karalija, N.; Harma, M.A.; Vilenius, C.; et al. GDNF Overexpression from the Native Locus Reveals its Role in the Nigrostriatal Dopaminergic System Function. *PLoS Genet.* **2015**, *11*, e1005710. [[CrossRef](#)]
120. Grieshammer, U.; Le, M.; Plump, A.S.; Wang, F.; Tessier-Lavigne, M.; Martin, G.R. SLIT2-mediated ROBO2 signaling restricts kidney induction to a single site. *Dev. Cell* **2004**, *6*, 709–717. [[CrossRef](#)]
121. Costantini, F.; Shakya, R. GDNF/Ret signaling and the development of the kidney. *Bioessays News Rev. Mol. Cell. Dev. Biol.* **2006**, *28*, 117–127. [[CrossRef](#)]
122. Li, H.; Jakobson, M.; Ola, R.; Gui, Y.; Kumar, A.; Sipilä, P.; Sariola, H.; Kuure, S. Development of the urogenital system is regulated via the 3'UTR of GDNF. *Sci. Rep.* **2019**, in press. [[CrossRef](#)]
123. Majumdar, A.; Vainio, S.; Kispert, A.; McMahon, J.; McMahon, A.P. Wnt11 and Ret/Gdnf pathways cooperate in regulating ureteric branching during metanephric kidney development. *Development* **2003**, *130*, 3175–3185. [[CrossRef](#)]

124. Michos, O.; Goncalves, A.; Lopez-Rios, J.; Tiecke, E.; Naillat, F.; Beier, K.; Galli, A.; Vainio, S.; Zeller, R. Reduction of BMP4 activity by gremlin 1 enables ureteric bud outgrowth and GDNF/WNT11 feedback signalling during kidney branching morphogenesis. *Development* **2007**, *134*, 2397–2405. [[CrossRef](#)]
125. Bridgewater, D.; Cox, B.; Cain, J.; Lau, A.; Athaide, V.; Gill, P.S.; Kuure, S.; Sainio, K.; Rosenblum, N.D. Canonical WNT/ β -catenin signaling is required for ureteric branching. *Dev. Biol.* **2008**, *317*, 83–94. [[CrossRef](#)]
126. Iglesias, D.M.; Hueber, P.A.; Chu, L.; Campbell, R.; Patenaude, A.M.; Dziarmaga, A.J.; Quinlan, J.; Mohamed, O.; Dufort, D.; Goodyer, P.R. Canonical WNT signaling during kidney development. *Am. J. Physiol. Ren. Physiol.* **2007**, *293*, F494–F500. [[CrossRef](#)]
127. Ola, R.; Jakobson, M.; Kvist, J.; Perala, N.; Kuure, S.; Braunewell, K.H.; Bridgewater, D.; Rosenblum, N.D.; Chilov, D.; Immonen, T.; et al. The GDNF target Vsn11 marks the ureteric tip. *J. Am. Soc. Nephrol. JASN* **2011**, *22*, 274–284. [[CrossRef](#)]
128. Miyazaki, Y.; Oshima, K.; Fogo, A.; Hogan, B.L.; Ichikawa, I. Bone morphogenetic protein 4 regulates the budding site and elongation of the mouse ureter. *J. Clin. Investig.* **2000**, *105*, 863–873. [[CrossRef](#)]
129. Miyazaki, Y.; Oshima, K.; Fogo, A.; Ichikawa, I. Evidence that bone morphogenetic protein 4 has multiple biological functions during kidney and urinary tract development. *Kidney Int.* **2003**, *63*, 835–844. [[CrossRef](#)]
130. Cain, J.E.; Nion, T.; Jeulin, D.; Bertram, J.F. Exogenous BMP-4 amplifies asymmetric ureteric branching in the developing mouse kidney in vitro. *Kidney Int.* **2005**, *67*, 420–431. [[CrossRef](#)]
131. Cebrián, C.; Borodo, K.; Charles, N.; Herzlinger, D.A. Morphometric index of the developing murine kidney. *Dev. Dyn. Off. Publ. Am. Assoc. Anat.* **2004**, *231*, 601–608. [[CrossRef](#)]
132. Short, K.M.; Combes, A.N.; Lefevre, J.; Ju, A.L.; Georgas, K.M.; Lamberton, T.; Cairncross, O.; Rumballe, B.A.; McMahon, A.P.; Hamilton, N.A.; et al. Global quantification of tissue dynamics in the developing mouse kidney. *Dev. Cell* **2014**, *29*, 188–202. [[CrossRef](#)]
133. Watanabe, T.; Costantini, F. Real-time analysis of ureteric bud branching morphogenesis in vitro. *Dev. Biol.* **2004**, *271*, 98–108. [[CrossRef](#)]
134. Michael, L.; Davies, J.A. Pattern and regulation of cell proliferation during murine ureteric bud development. *J. Anat.* **2004**, *204*, 241–255. [[CrossRef](#)]
135. Shakya, R.; Watanabe, T.; Costantini, F. The role of GDNF/Ret signaling in ureteric bud cell fate and branching morphogenesis. *Dev. Cell* **2005**, *8*, 65–74. [[CrossRef](#)]
136. Sariola, H. Nephron induction revisited: From caps to condensates. *Curr. Opin. Nephrol. Hypertens.* **2002**, *11*, 17–21. [[CrossRef](#)]
137. Kobayashi, A.; Valerius, M.T.; Mugford, J.W.; Carroll, T.J.; Self, M.; Oliver, G.; McMahon, A.P. Six2 defines and regulates a multipotent self-renewing nephron progenitor population throughout mammalian kidney development. *Cell Stem Cell* **2008**, *3*, 169–181. [[CrossRef](#)]
138. Boyle, S.; Misfeldt, A.; Chandler, K.J.; Deal, K.K.; Southard-Smith, E.M.; Mortlock, D.P.; Baldwin, H.S.; de Caestecker, M. Fate mapping using Cited1-CreERT2 mice demonstrates that the cap mesenchyme contains self-renewing progenitor cells and gives rise exclusively to nephronic epithelia. *Dev. Biol.* **2008**, *313*, 234–245. [[CrossRef](#)]
139. Uchiyama, Y.; Sakaguchi, M.; Terabayashi, T.; Inenaga, T.; Inoue, S.; Kobayashi, C.; Oshima, N.; Kiyonari, H.; Nakagata, N.; Sato, Y.; et al. Kif26b, a kinesin family gene, regulates adhesion of the embryonic kidney mesenchyme. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 9240–9245. [[CrossRef](#)]
140. O'Brien, L.L. Nephron progenitor cell commitment: Striking the right balance. *Semin. Cell Dev. Biol.* **2018**. [[CrossRef](#)]
141. Georgas, K.; Rumballe, B.; Valerius, M.T.; Chiu, H.S.; Thiagarajan, R.D.; Lesieur, E.; Aronow, B.J.; Brunskill, E.W.; Combes, A.N.; Tang, D.; et al. Analysis of early nephron patterning reveals a role for distal RV proliferation in fusion to the ureteric tip via a cap mesenchyme-derived connecting segment. *Dev. Biol.* **2009**, *332*, 273–286. [[CrossRef](#)]
142. Lindström, N.O.; Brandine, G.D.S.; Tran, T.; Ransick, A.; Suh, G.; Guo, J.; Kim, A.D.; Parvez, R.K.; Ruffins, S.W.; Rutledge, E.A. Progressive recruitment of mesenchymal progenitors reveals a time-dependent process of cell fate acquisition in mouse and human nephrogenesis. *Dev. Cell* **2018**, *45*, 651–660.e4. [[CrossRef](#)]
143. Cebrian, C.; Asai, N.; D'Agati, V.; Costantini, F. The number of fetal nephron progenitor cells limits ureteric branching and adult nephron endowment. *Cell Rep.* **2014**, *7*, 127–137. [[CrossRef](#)]

144. Self, M.; Lagutin, O.V.; Bowling, B.; Hendrix, J.; Cai, Y.; Dressler, G.R.; Oliver, G. Six2 is required for suppression of nephrogenesis and progenitor renewal in the developing kidney. *EMBO J.* **2006**, *25*, 5214–5228. [[CrossRef](#)]
145. Kanda, S.; Tanigawa, S.; Ohmori, T.; Taguchi, A.; Kudo, K.; Suzuki, Y.; Sato, Y.; Hino, S.; Sander, M.; Perantoni, A.O.; et al. Sall1 maintains nephron progenitors and nascent nephrons by acting as both an activator and a repressor. *J. Am. Soc. Nephrol. JASN* **2014**, *25*, 2584–2595. [[CrossRef](#)]
146. Basta, J.M.; Robbins, L.; Kiefer, S.M.; Dorsett, D.; Rauchman, M. Sall1 balances self-renewal and differentiation of renal progenitor cells. *Development* **2014**, *141*, 1047–1058. [[CrossRef](#)]
147. Naiman, N.; Fujioka, K.; Fujino, M.; Valerius, M.T.; Potter, S.S.; McMahon, A.P.; Kobayashi, A. Repression of Interstitial Identity in Nephron Progenitor Cells by Pax2 Establishes the Nephron-Interstitium Boundary during Kidney Development. *Dev. Cell* **2017**, *41*, 349–365.e3. [[CrossRef](#)]
148. Ranghini, E.J.; Dressler, G.R. Evidence for intermediate mesoderm and kidney progenitor cell specification by Pax2 and PTIP dependent mechanisms. *Dev. Biol.* **2015**, *399*, 296–305. [[CrossRef](#)]
149. Kaku, Y.; Taguchi, A.; Tanigawa, S.; Haque, F.; Sakuma, T.; Yamamoto, T.; Nishinakamura, R. PAX2 is dispensable for in vitro nephron formation from human induced pluripotent stem cells. *Sci. Rep.* **2017**, *7*, 4554. [[CrossRef](#)]
150. Li, Y.; Liu, J.; Li, W.; Brown, A.; Baddoo, M.; Li, M.; Carroll, T.; Oxburgh, L.; Feng, Y.; Saifudeen, Z. p53 Enables metabolic fitness and self-renewal of nephron progenitor cells. *Development* **2015**, *142*, 1228–1241. [[CrossRef](#)]
151. Xu, J.; Liu, H.; Park, J.S.; Lan, Y.; Jiang, R. Osr1 acts downstream of and interacts synergistically with Six2 to maintain nephron progenitor cells during kidney organogenesis. *Development* **2014**, *141*, 1442–1452. [[CrossRef](#)]
152. Xu, J.; Wong, E.Y.; Cheng, C.; Li, J.; Sharkar, M.T.; Xu, C.Y.; Chen, B.; Sun, J.; Jing, D.; Xu, P.X. Eya1 interacts with Six2 and Myc to regulate expansion of the nephron progenitor pool during nephrogenesis. *Dev. Cell* **2014**, *31*, 434–447. [[CrossRef](#)]
153. Pan, X.; Karner, C.M.; Carroll, T.J. Myc cooperates with β -catenin to drive gene expression in nephron progenitor cells. *Development* **2017**, *144*, 4173–4182. [[CrossRef](#)]
154. Barak, H.; Huh, S.H.; Chen, S.; Jeanpierre, C.; Martinovic, J.; Parisot, M.; Bole-Feysot, C.; Nitschke, P.; Salomon, R.; Antignac, C.; et al. FGF9 and FGF20 maintain the stemness of nephron progenitors in mice and man. *Dev. Cell* **2012**, *22*, 1191–1207. [[CrossRef](#)]
155. Ahn, S.Y.; Kim, Y.; Kim, S.T.; Swat, W.; Miner, J.H. Scaffolding proteins DLG1 and CASK cooperate to maintain the nephron progenitor population during kidney development. *J. Am. Soc. Nephrol. JASN* **2013**, *24*, 1127–1138. [[CrossRef](#)]
156. Motamedi, F.J.; Badro, D.A.; Clarkson, M.; Lecca, M.R.; Bradford, S.T.; Buske, F.A.; Saar, K.; Hubner, N.; Brandli, A.W.; Schedl, A. WT1 controls antagonistic FGF and BMP-pSMAD pathways in early renal progenitors. *Nat. Commun.* **2014**, *5*, 4444. [[CrossRef](#)]
157. Chen, S.; Brunskill, E.W.; Potter, S.S.; Dexheimer, P.J.; Salomonis, N.; Aronow, B.J.; Hong, C.I.; Zhang, T.; Kopan, R. Intrinsic Age-Dependent Changes and Cell-Cell Contacts Regulate Nephron Progenitor Lifespan. *Dev. Cell* **2015**, *35*, 49–62. [[CrossRef](#)]
158. Qiao, J.; Uzzo, R.; Obara-Ishihara, T.; Degenstein, L.; Fuchs, E.; Herzlinger, D. FGF-7 modulates ureteric bud growth and nephron number in the developing kidney. *Development* **1999**, *126*, 547–554.
159. Ohuchi, H.; Hori, Y.; Yamasaki, M.; Harada, H.; Sekine, K.; Kato, S.; Itoh, N. FGF10 acts as a major ligand for FGF receptor 2 IIIb in mouse multi-organ development. *Biochem. Biophys. Res. Commun.* **2000**, *277*, 643–649. [[CrossRef](#)]
160. Grieshammer, U.; Cebrian, C.; Ilagan, R.; Meyers, E.; Herzlinger, D.; Martin, G.R. FGF8 is required for cell survival at distinct stages of nephrogenesis and for regulation of gene expression in nascent nephrons. *Development* **2005**, *132*, 3847–3857. [[CrossRef](#)]
161. Perantoni, A.O.; Timofeeva, O.; Naillat, F.; Richman, C.; Pajni-Underwood, S.; Wilson, C.; Vainio, S.; Dove, L.F.; Lewandoski, M. Inactivation of FGF8 in early mesoderm reveals an essential role in kidney development. *Development* **2005**, *132*, 3859–3871. [[CrossRef](#)]
162. Brown, A.C.; Muthukrishnan, S.D.; Guay, J.A.; Adams, D.C.; Schafer, D.A.; Fetting, J.L.; Oxburgh, L. Role for compartmentalization in nephron progenitor differentiation. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 4640–4645. [[CrossRef](#)]

163. Dudley, A.T.; Lyons, K.M.; Robertson, E.J. A requirement for bone morphogenetic protein-7 during development of the mammalian kidney and eye. *Genes Dev.* **1995**, *9*, 2795–2807. [[CrossRef](#)]
164. Dudley, A.T.; Godin, R.E.; Robertson, E.J. Interaction between FGF and BMP signaling pathways regulates development of metanephric mesenchyme. *Genes Dev.* **1999**, *13*, 1601–1613. [[CrossRef](#)]
165. Luo, G.; Hofmann, C.; Bronckers, A.L.J.J.; Sohocki, M.; Bradley, A.; Karsenty, G. Bmp-7 Is an Inducer of Nephrogenesis, and Is Also Required for Eye Development and Skeletal Patterning. *Genes Dev.* **1995**, *9*, 2808–2820. [[CrossRef](#)]
166. Tomita, M.; Asada, M.; Asada, N.; Nakamura, J.; Oguchi, A.; Higashi, A.Y.; Endo, S.; Robertson, E.; Kimura, T.; Kita, T.; et al. Bmp7 maintains undifferentiated kidney progenitor population and determines nephron numbers at birth. *PLoS ONE* **2013**, *8*, e73554. [[CrossRef](#)]
167. O'Brien, L.L.; McMahon, A.P. Induction and patterning of the metanephric nephron. *Semin. Cell Dev. Biol.* **2014**, *36*, 31–38. [[CrossRef](#)]
168. Brown, A.C.; Adams, D.; de Caestecker, M.; Yang, X.; Friesel, R.; Oxburgh, L. FGF/EGF signaling regulates the renewal of early nephron progenitors during embryonic development. *Development* **2011**, *138*, 5099–5112. [[CrossRef](#)]
169. Brown, A.C.; Muthukrishnan, S.D.; Oxburgh, L. A synthetic niche for nephron progenitor cells. *Dev. Cell* **2015**, *34*, 229–241. [[CrossRef](#)]
170. Murray, P.A.; Woolf, A.S. Using stem and progenitor cells to recapitulate kidney development and restore renal function. *Curr. Opin. Organ Transplant.* **2014**, *19*, 140–144. [[CrossRef](#)]
171. Kimber, S.J.; Woolf, A.S. From human pluripotent stem cells to functional kidney organoids and models of renal disease. *Stem Cell Investig.* **2018**, *5*, 20. [[CrossRef](#)]
172. Hartman, H.A.; Lai, H.L.; Patterson, L.T. Cessation of renal morphogenesis in mice. *Dev. Biol.* **2007**, *310*, 379–387. [[CrossRef](#)]
173. Rumballe, B.A.; Georgas, K.M.; Combes, A.N.; Ju, A.L.; Gilbert, T.; Little, M.H. Nephron formation adopts a novel spatial topology at cessation of nephrogenesis. *Dev. Biol.* **2011**, *360*, 110–122. [[CrossRef](#)]
174. Brunskill, E.W.; Lai, H.L.; Jamison, D.C.; Potter, S.S.; Patterson, L.T. Microarrays and RNA-Seq identify molecular mechanisms driving the end of nephron production. *BMC Dev. Biol.* **2011**, *11*, 15. [[CrossRef](#)]
175. Rymer, C.; Paredes, J.; Halt, K.; Schaefer, C.; Wiersch, J.; Zhang, G.; Potoka, D.; Vainio, S.; Gittes, G.K.; Bates, C.M.; et al. Renal blood flow and oxygenation drive nephron progenitor differentiation. *Am. J. Physiol. Ren. Physiol.* **2014**, *307*, F337–F345. [[CrossRef](#)]
176. Volovelsky, O.; Nguyen, T.; Jarmas, A.E.; Combes, A.N.; Wilson, S.B.; Little, M.H.; Witte, D.P.; Brunskill, E.W.; Kopan, R. Hamartin regulates cessation of mouse nephrogenesis independently of Mtor. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 5998–6003. [[CrossRef](#)]
177. Fisher, C.E.; Michael, L.; Barnett, M.W.; Davies, J.A. Erk MAP kinase regulates branching morphogenesis in the developing mouse kidney. *Development* **2001**, *128*, 4329–4338.
178. Ihermann-Hella, A.; Lume, M.; Miinalainen, I.J.; Pirttiniemi, A.; Gui, Y.; Peranen, J.; Charron, J.; Saarma, M.; Costantini, F.; Kuure, S. Mitogen-activated protein kinase (MAPK) pathway regulates branching by remodeling epithelial cell adhesion. *PLoS Genet.* **2014**, *10*, e1004193. [[CrossRef](#)]
179. De Graaff, E.; Srinivas, S.; Kilkenny, C.; D'Agati, V.; Mankoo, B.S.; Costantini, F.; Pachnis, V. Differential activities of the RET tyrosine kinase receptor isoforms during mammalian embryogenesis. *Genes Dev.* **2001**, *15*, 2433–2444. [[CrossRef](#)]
180. Wong, A.; Bogni, S.; Kotka, P.; de Graaff, E.; D'Agati, V.; Costantini, F.; Pachnis, V. Phosphotyrosine 1062 is critical for the in vivo activity of the Ret9 receptor tyrosine kinase isoform. *Mol. Cell. Biol.* **2005**, *25*, 9661–9673. [[CrossRef](#)]
181. Jijiwa, M.; Fukuda, T.; Kawai, K.; Nakamura, A.; Kurokawa, K.; Murakumo, Y.; Ichihara, M.; Takahashi, M. A targeting mutation of tyrosine 1062 in Ret causes a marked decrease of enteric neurons and renal hypoplasia. *Mol. Cell. Biol.* **2004**, *24*, 8026–8036. [[CrossRef](#)]
182. Jain, S.; Knoten, A.; Hoshi, M.; Wang, H.; Vohra, B.; Heuckeroth, R.O.; Milbrandt, J. Organotypic specificity of key RET adaptor-docking sites in the pathogenesis of neurocristopathies and renal malformations in mice. *J. Clin. Investig.* **2010**, *120*, 778–790. [[CrossRef](#)]
183. Hoshi, M.; Batourina, E.; Mendelsohn, C.; Jain, S. Novel mechanisms of early upper and lower urinary tract patterning regulated by RetY1015 docking tyrosine in mice. *Development* **2012**, *139*, 2405–2415. [[CrossRef](#)]

184. Chi, L.; Zhang, S.; Lin, Y.; Prunskaitė-Hyyryläinen, R.; Vuolteenaho, R.; Itaranta, P.; Vainio, S. Sprouty proteins regulate ureteric branching by coordinating reciprocal epithelial Wnt11, mesenchymal Gdnf and stromal Fgf7 signalling during kidney development. *Development* **2004**, *131*, 3345–3356. [[CrossRef](#)]
185. Rozen, E.J.; Schmidt, H.; Dolcet, X.; Basson, M.A.; Jain, S.; Encinas, M. Loss of Sprouty1 rescues renal agenesis caused by Ret mutation. *J. Am. Soc. Nephrol. JASN* **2009**, *20*, 255–259. [[CrossRef](#)]
186. Michos, O.; Cebrian, C.; Hyink, D.; Grieshammer, U.; Williams, L.; D'Agati, V.; Licht, J.D.; Martin, G.R.; Costantini, F. Kidney development in the absence of Gdnf and Spry1 requires Fgf10. *PLoS Genet.* **2010**, *6*, e1000809. [[CrossRef](#)]
187. Imperial, R.; Toor, O.M.; Hussain, A.; Subramanian, J.; Masood, A. Comprehensive pancancer genomic analysis reveals (RTK)-RAS-RAF-MEK as a key dysregulated pathway in cancer: Its clinical implications. In *Seminars in Cancer Biology*; Academic Press: Cambridge, MA, USA, 2017.
188. Oka, H.; Chatani, Y.; Hoshino, R.; Ogawa, O.; Kakehi, Y.; Terachi, T.; Okada, Y.; Kawaichi, M.; Kohno, M.; Yoshida, O. Constitutive activation of mitogen-activated protein (MAP) kinases in human renal cell carcinoma. *Cancer Res.* **1995**, *55*, 4182–4187.
189. Hastie, N.D. Wilms' tumour 1 (WT1) in development, homeostasis and disease. *Development* **2017**, *144*, 2862–2872. [[CrossRef](#)]
190. Kreidberg, J.A.; Sariola, H.; Loring, J.M.; Maeda, M.; Pelletier, J.; Housman, D.; Jaenisch, R. Wt-1 Is Required for Early Kidney Development. *Cell* **1993**, *74*, 679–691. [[CrossRef](#)]
191. Ogawa, O.; Eccles, M.R.; Szeto, J.; McNoe, L.A.; Yun, K.; Maw, M.A.; Smith, P.J.; Reeve, A.E. Relaxation of insulin-like growth factor II gene imprinting implicated in Wilms' tumour. *Nature* **1993**, *362*, 749–751. [[CrossRef](#)]
192. Hunter, R.W.; Liu, Y.; Manjunath, H.; Acharya, A.; Jones, B.T.; Zhang, H.; Chen, B.; Ramalingam, H.; Hammer, R.E.; Xie, Y. Loss of Dis3l2 partially phenocopies Perlman syndrome in mice and results in up-regulation of Igf2 in nephron progenitor cells. *Genes Dev.* **2018**, *32*, 903–908. [[CrossRef](#)]
193. Chen, K.S.; Stroup, E.K.; Budhipramono, A.; Rakheja, D.; Nichols-Vinueza, D.; Xu, L.; Stuart, S.H.; Shukla, A.A.; Fraire, C.; Mendell, J.T. Mutations in microRNA processing genes in Wilms tumors derepress the IGF2 regulator PLAG1. *Genes Dev.* **2018**, *32*, 996–1007. [[CrossRef](#)]
194. Hu, Q.; Gao, F.; Tian, W.; Ruteshouser, E.C.; Wang, Y.; Lazar, A.; Stewart, J.; Strong, L.C.; Behringer, R.R.; Huff, V. Wt1 ablation and Igf2 upregulation in mice result in Wilms tumors with elevated ERK1/2 phosphorylation. *J. Clin. Investig.* **2011**, *121*, 174–183. [[CrossRef](#)]
195. Subbiah, V.; Brown, R.E.; Jiang, Y.; Buryanek, J.; Hayes-Jordan, A.; Kurzrock, R.; Anderson, P.M. Morphoproteomic profiling of the mammalian target of rapamycin (mTOR) signaling pathway in desmoplastic small round cell tumor (EWS/WT1), Ewing's sarcoma (EWS/FLI1) and Wilms' tumor (WT1). *PLoS ONE* **2013**, *8*, e68985. [[CrossRef](#)]
196. Mae, S.I.; Shono, A.; Shiota, F.; Yasuno, T.; Kajiwara, M.; Gotoda-Nishimura, N.; Arai, S.; Sato-Otubo, A.; Toyoda, T.; Takahashi, K.; et al. Monitoring and robust induction of nephrogenic intermediate mesoderm from human pluripotent stem cells. *Nat. Commun.* **2013**, *4*, 1367. [[CrossRef](#)]
197. Takasato, M.; Er, P.X.; Becroft, M.; Vanslambrouck, J.M.; Stanley, E.G.; Elefanty, A.G.; Little, M.H. Directing human embryonic stem cell differentiation towards a renal lineage generates a self-organizing kidney. *Nat. Cell Biol.* **2014**, *16*, 118–126. [[CrossRef](#)]
198. Morizane, R.; Lam, A.Q.; Freedman, B.S.; Kishi, S.; Valerius, M.T.; Bonventre, J.V. Nephron organoids derived from human pluripotent stem cells model kidney development and injury. *Nat. Biotechnol.* **2015**, *33*, 1193–1200. [[CrossRef](#)]

