

# Red Tides and Phycotoxins: the Problems are in the Details

Chile is prone to HAB events, e.g. 2016



# Introduction

February/March 2016, Región de Los Lagos: massive bloom of  
*Pseudochattonella cf. verruculosa*



Informe Final, Comisión Marea Roja

[https://www.economia.gob.cl/wp-content/uploads/2016/11/InfoFinal\\_ComisionMareaRoja\\_24Nov2016-1.compressed.pdf](https://www.economia.gob.cl/wp-content/uploads/2016/11/InfoFinal_ComisionMareaRoja_24Nov2016-1.compressed.pdf)

# Introduction

April/May 2016, Región de Los Lagos: massive bloom event of  
*Alexandrium catenella*



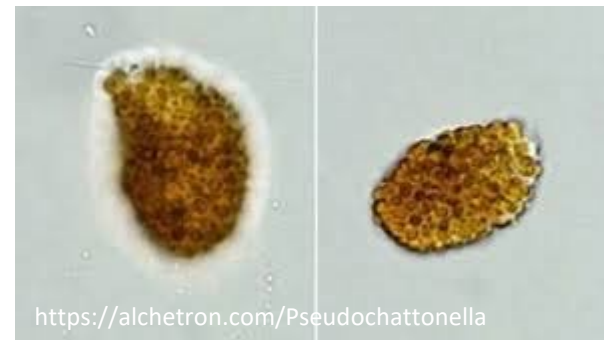
Informe Final, Comisión Marea Roja

[https://www.economia.gob.cl/wp-content/uploads/2016/11/InfoFinal\\_ComisionMareaRoja\\_24Nov2016-1.compressed.pdf](https://www.economia.gob.cl/wp-content/uploads/2016/11/InfoFinal_ComisionMareaRoja_24Nov2016-1.compressed.pdf)

Both blooms were almost coinciding in time and space and caused mass mortality of marine life, especially in salmon aquaculture

Mechanistically both blooms were different:

*Pseudochattonella* cf. *verriculosa*:  
Ichthyotoxic



*Alexandrium catenella*:  
PSP Toxin producer



## Variability

Amphidinols: 20+ known variants

Karlotoxins: 20+ known variants

Prymnesins: 100+ variants

### Other ichthyotoxic species:

*Alexandrium* spp.

*Chattonella* spp.

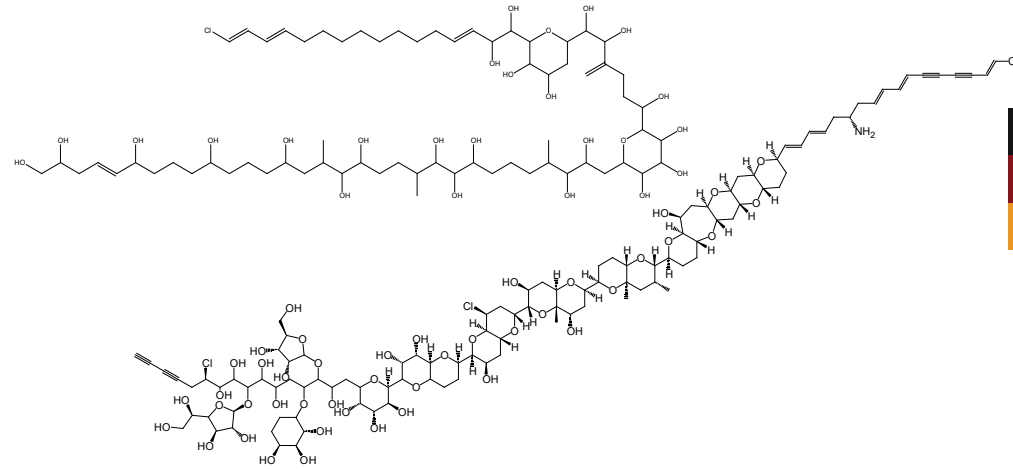
*Chrysocromulina* spp.

*Fibrocapsa japonica*

*Heterosigma akashiwo*

*Protoceratium reticulatum*

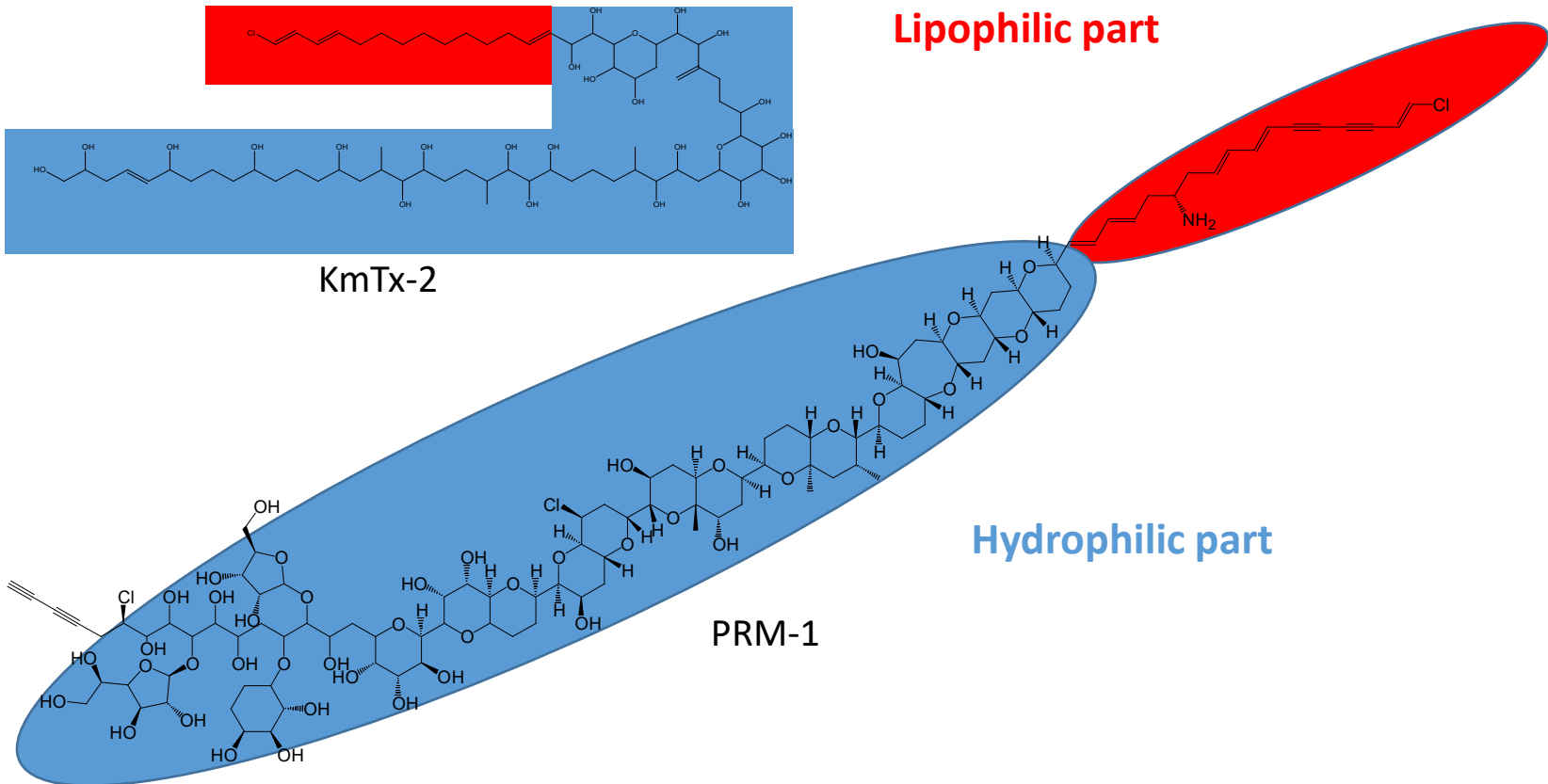
*Pseudochattonella* cf. *verruculosa*



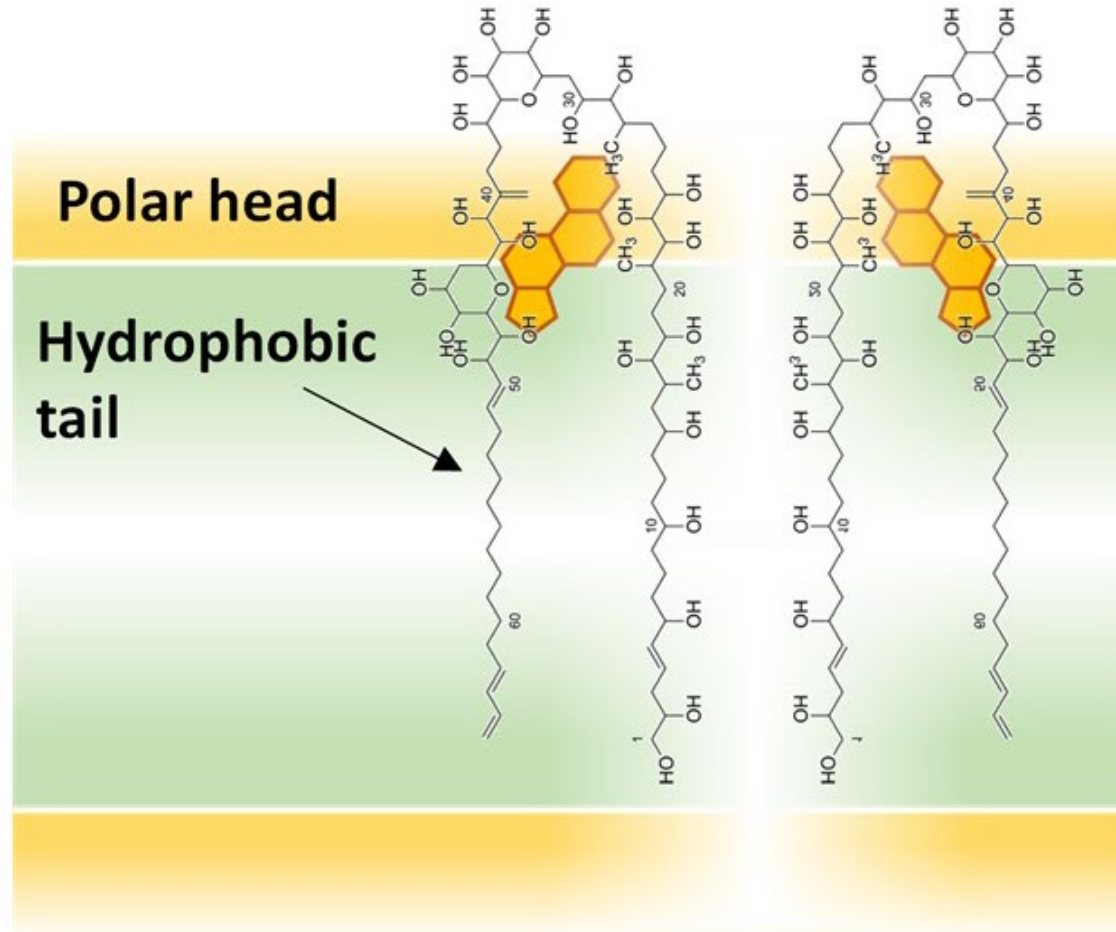
# Ichthyotoxins: Unknown !!

# Ichthyotoxins

## Known Ichthyotoxins



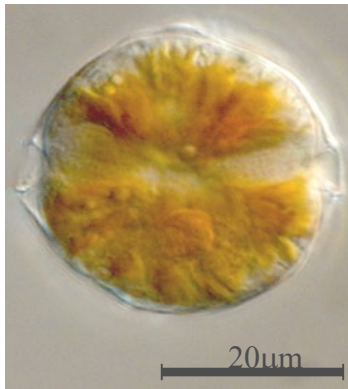
## Mode of action



Long et al. (2021) *Toxins* 13(12): 905.



## Lytic Effect



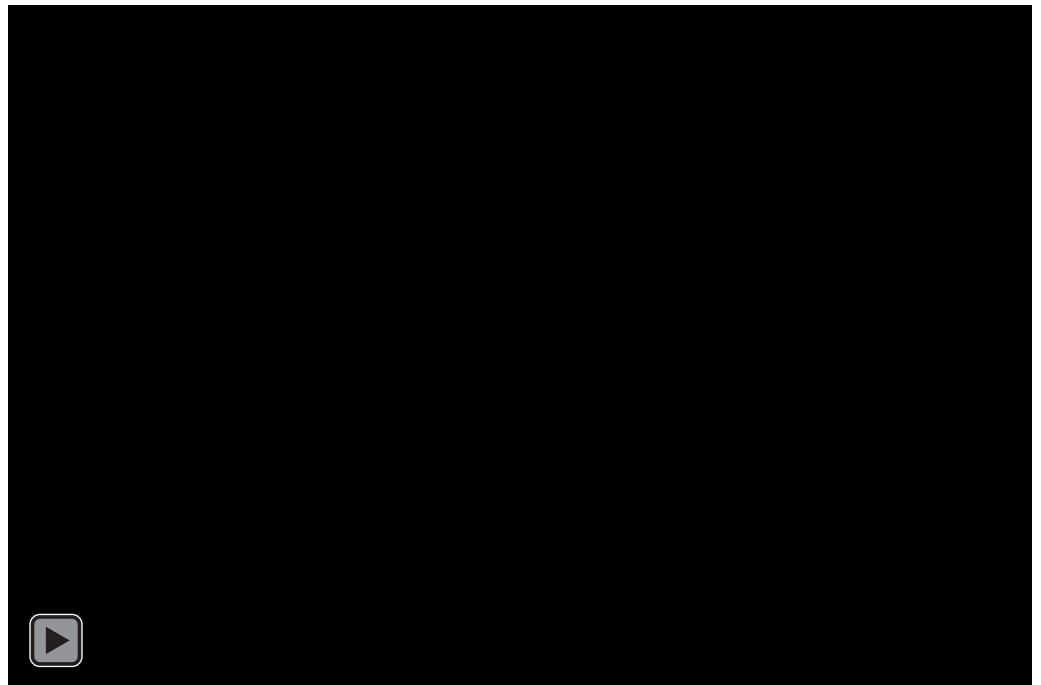
*Alexandrium catenella*  
strain 2 (Alex2)



*Rhodomonas salina*



*Rhodomonas salina* exposed to *A. catenella* supernatant (cell free)



Video: U. Tillmann

**Organism**

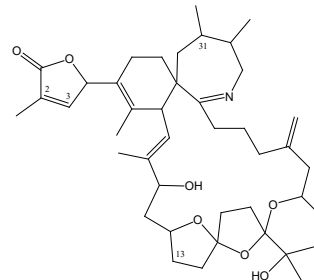
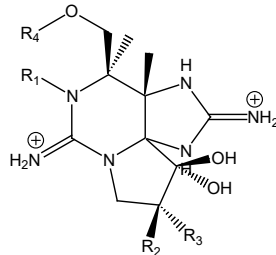
**Chemical compound**

**Ecological Function**

PSP-Toxins

Spirolides

*Alexandrium*  
spp.



?

*Pseudochattonella*  
cf. *verriculosa*

(*Alexandrium* spp.)

?

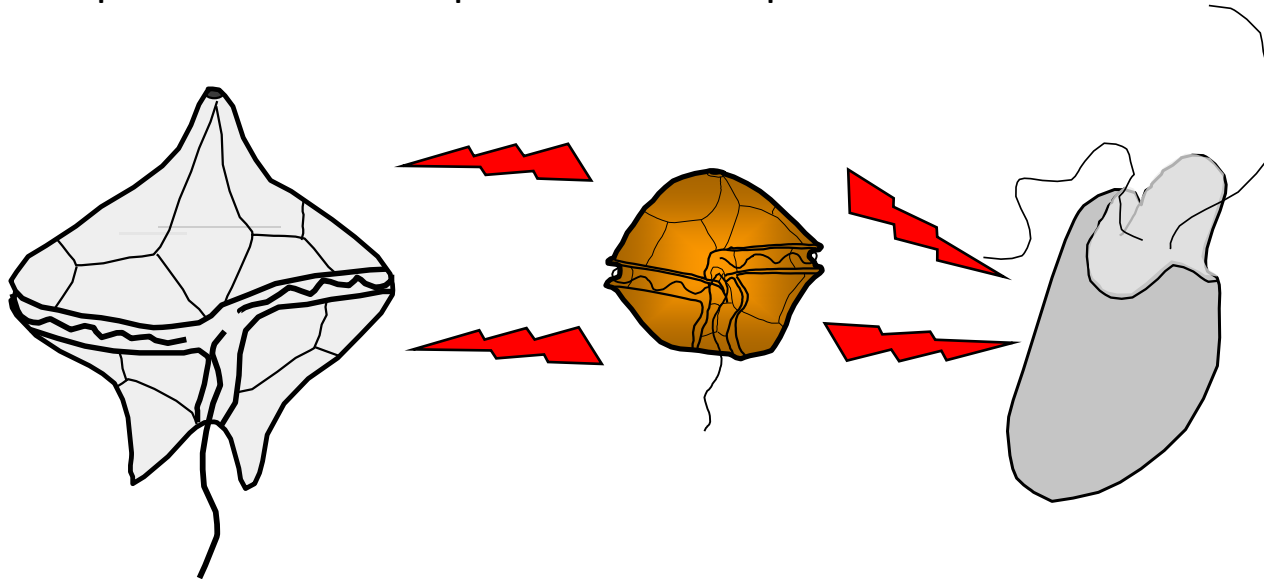
Defense against Predators

Elimination of Competitors

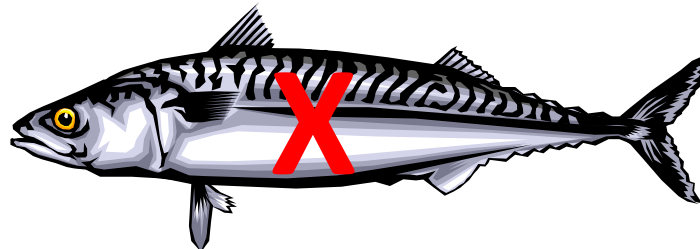
# Ichthyotoxins

Current Hypothesis:

Lytic compounds of marine protists are weapons of chemical warfare among protists



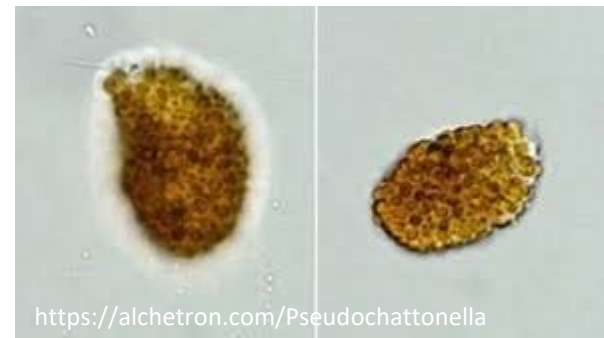
Ichthyotoxicity is a collateral damage of protistan allelochemistry



Both blooms were almost coinciding in time and space and caused mass mortality of marine life, especially in salmon aquaculture

Mechanistically both blooms were different:

*Pseudochattonella* cf. *verriculosa*:  
Ichthyotoxic

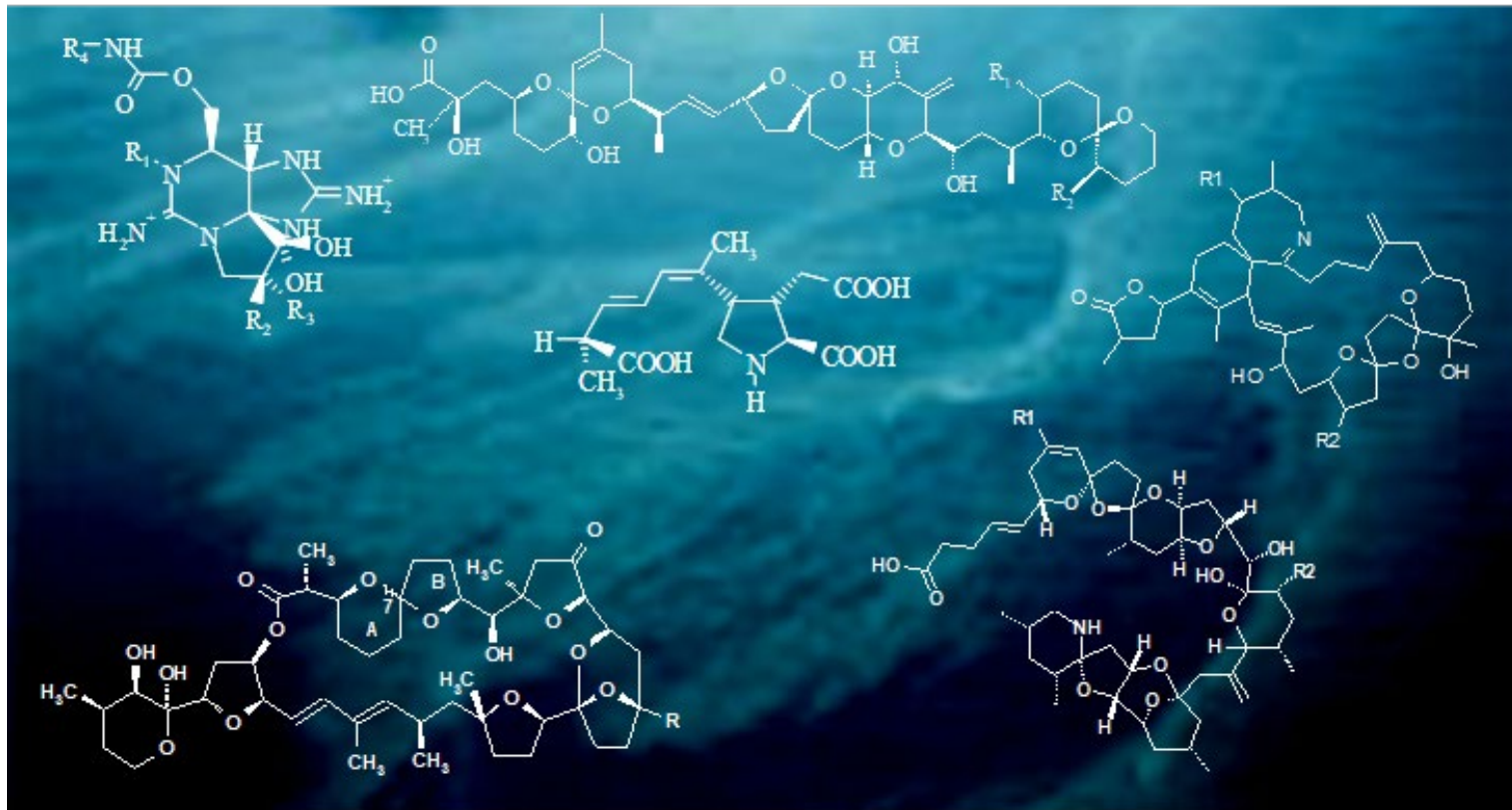


*Alexandrium catenella*:  
PSP Toxin producer



# Variability of phycotoxins

## High variability of phycotoxin classes



## Need of monitoring for seafood safety

# Variability of phycotoxins

Mouse Bioassay (MBA)

Liquid chromatography coupled to tandem mass spectrometry (LC-MS/MS)



MBA is banned in the United States and the European Union

# Variability of phycotoxins



## Problems of MBA:

- Poor reproducibility
- Low sensitivity
- Low specificity
- Ethical concerns



## Problems of LC-MS/MS:

- High cost
- Need of trained staff
- **Only targeted analysis**

# Variability of phycotoxins

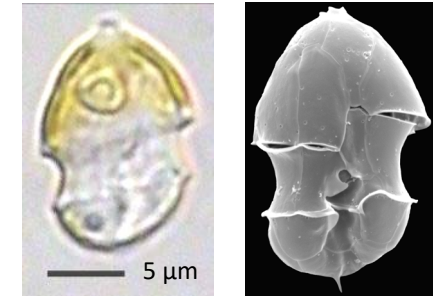
Toxin group	Analogue	TEF
OA-group toxins (OA-equivalents)	OA	1
	DTX1	1
	DTX2	0.6
AZA-group toxins (AZA-equivalents)	AZA1	1
	AZA2	1.8
	AZA3	1.4
YTX-group toxins (YTX-equivalents)	YTX	1
	1a-homoYTX	1
	45-hydroxyYTX	1
	45-hydroxy-1a-homoYTX	0.5
STX-group toxins (STX-equivalents)	STX	1
	NeoSTX	1
	GTX1	1
	GTX2	0.4
	GTX3	0.6
	GTX4	0.7
	GTX5	0.1
	GTX6	0.1
	C2	0.1
	C4	0.1
	dc-STX = 1	1
dc-NeoSTX	0.4	
dc GTX2	0.2	
dc GTX3	0.4	
PTX-group toxins (PTX2-equivalents)	PTX1	1
	PTX2	1
	PTX3	1
	PTX4	1
	PTX6	1
	PTX11	1
DA and its isomers	None established	-

The EFSA Journal (2009) 1306, 1-23



# Variability of phycotoxins

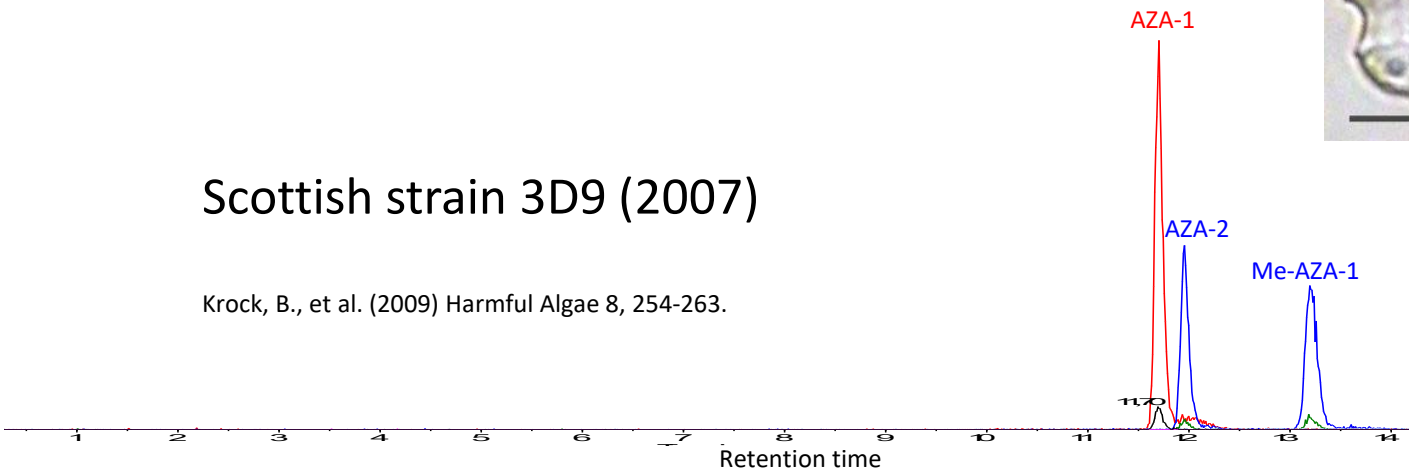
## *Azadinium* as AZA producer



*A. spinosum*

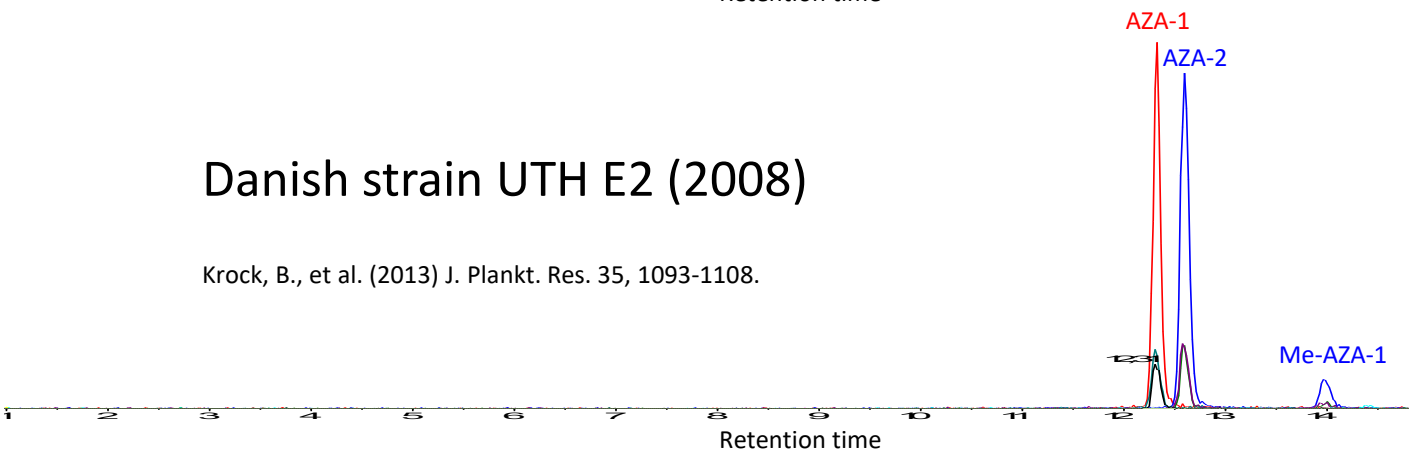
### Scottish strain 3D9 (2007)

Krock, B., et al. (2009) Harmful Algae 8, 254-263.



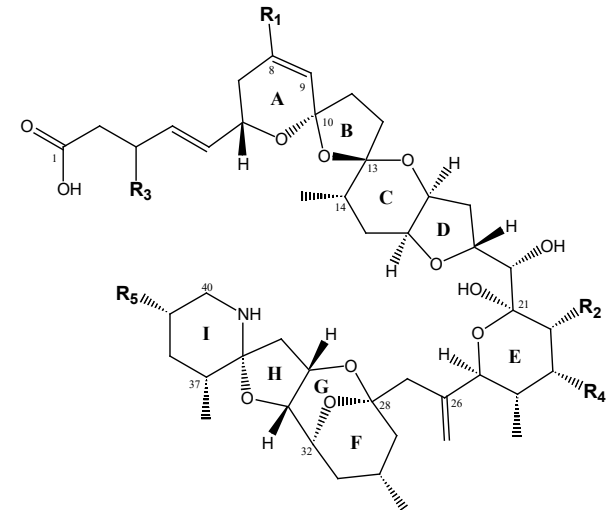
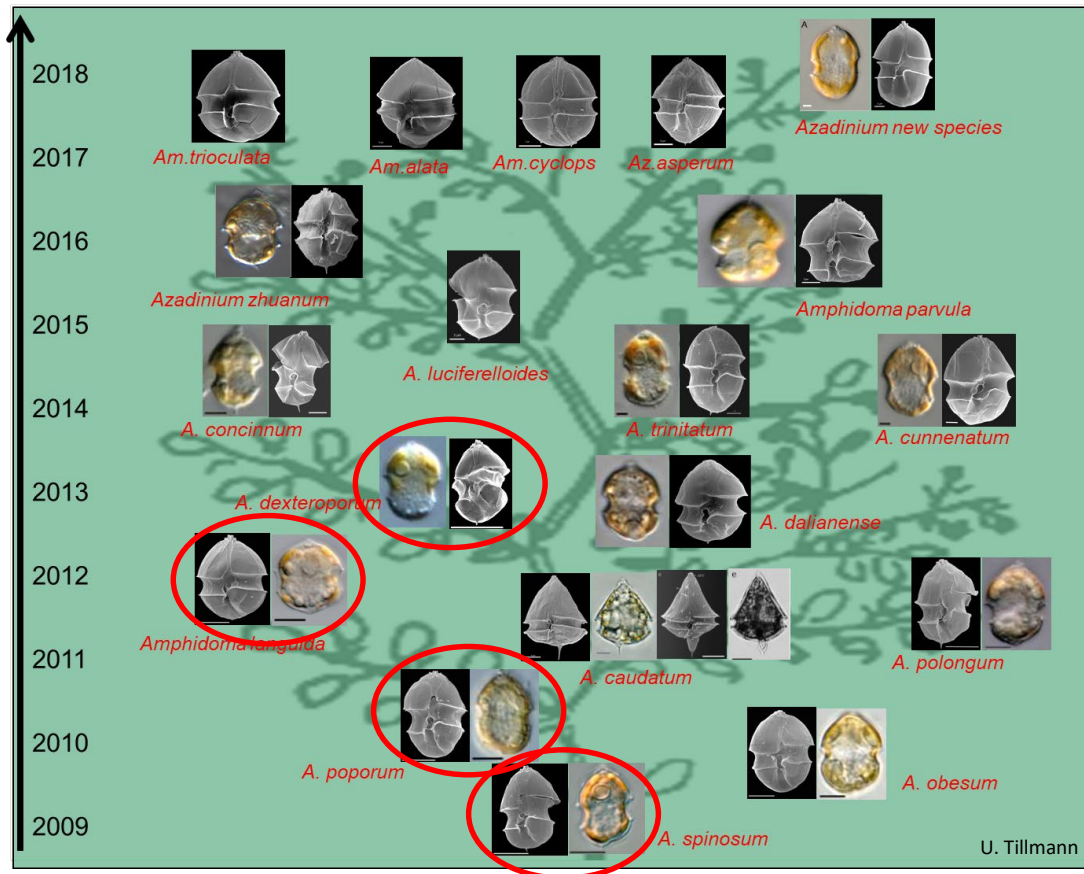
### Danish strain UTH E2 (2008)

Krock, B., et al. (2013) J. Plankt. Res. 35, 1093-1108.

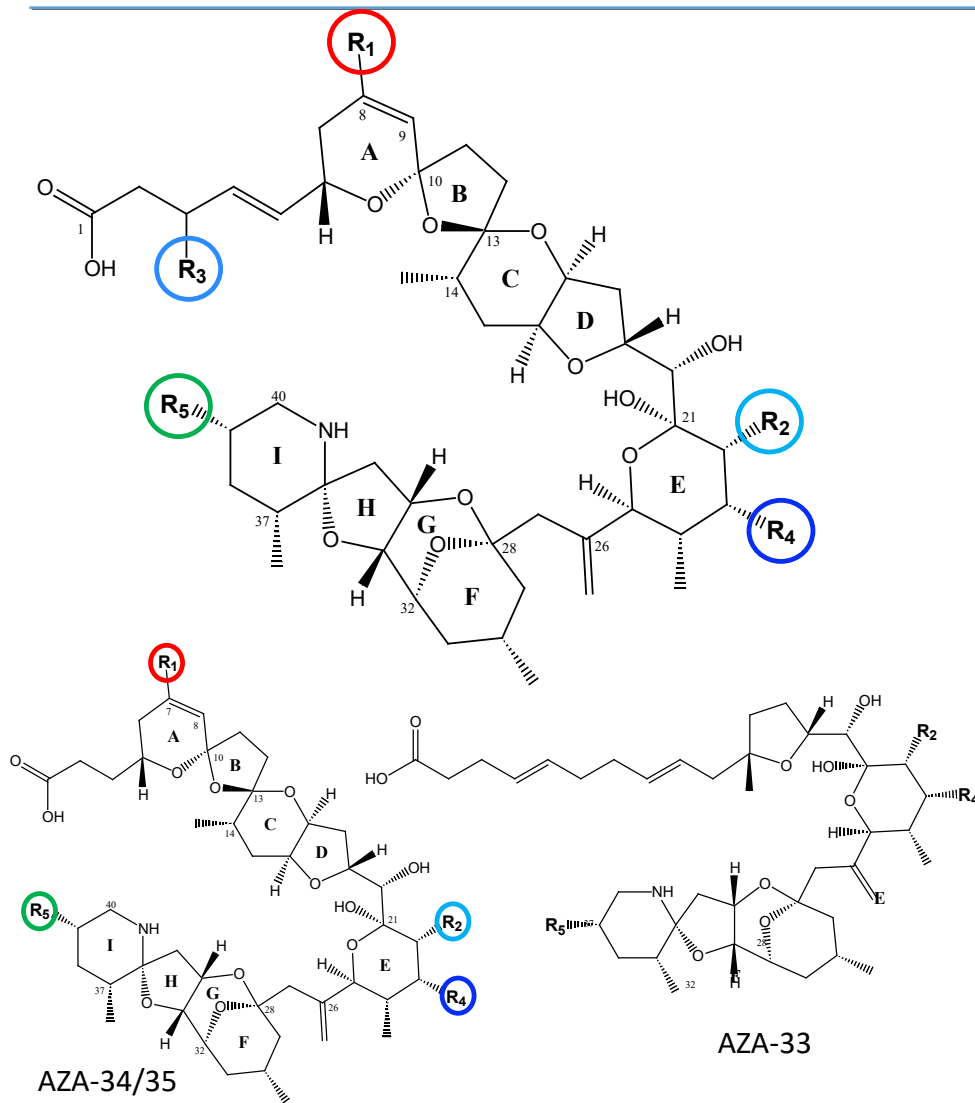


$m/z$  842>824  
 $m/z$  856>838

# Variability of phycotoxins



# Variability of phycotoxins



Toxin	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>	R <sub>5</sub>	Δ <sub>7,8</sub>	[M+H] <sup>+</sup>
AZA-1	H	CH <sub>3</sub>	H	H	CH <sub>3</sub>	✓	842
AZA-2	CH <sub>3</sub>	CH <sub>3</sub>	H	H	CH <sub>3</sub>	✓	856
AZA-3	H	H	H	H	CH <sub>3</sub>	✓	828
AZA-4	H	H	OH	H	CH <sub>3</sub>	✓	844
AZA-5	H	H	H	OH	CH <sub>3</sub>	✓	844
AZA-6	CH <sub>3</sub>	H	H	H	CH <sub>3</sub>	✓	842
AZA-7	H	CH <sub>3</sub>	OH	H	CH <sub>3</sub>	✓	858
AZA-8	H	CH <sub>3</sub>	H	OH	CH <sub>3</sub>	✓	858
AZA-9	CH <sub>3</sub>	H	OH	H	CH <sub>3</sub>	✓	858
AZA-10	CH <sub>3</sub>	H	H	OH	CH <sub>3</sub>	✓	858
AZA-11	CH <sub>3</sub>	CH <sub>3</sub>	OH	H	CH <sub>3</sub>	✓	872
AZA-33	-	CH <sub>3</sub>	H	H	CH <sub>3</sub>	-	716
AZA-34	H	CH <sub>3</sub>	-	H	CH <sub>3</sub>	✓	816
AZA-35	CH <sub>3</sub>	CH <sub>3</sub>	-	H	CH <sub>3</sub>	✓	830
AZA-36	CH <sub>3</sub>	CH <sub>3</sub>	OH	H	H	✓	858
AZA-37	H	CH <sub>3</sub>	OH	H	H	-	846
AZA-38	nd	nd	nd	nd	H	nd	830
AZA-39	nd	nd	nd	nd	H	nd	816
AZA-40	CH <sub>3</sub>	CH <sub>3</sub>	H	H	H	✓	842

# Variability of phycotoxins

## Currently Known AZA from Dinoflagellate (Algal) Origin

#	AZA	m/z [M+H] <sup>+</sup>	m/z group 4 fragment	m/z group 5 fragment	Producer	Reference
1	AZA-1	842	362	262	<i>A. spinosum</i>	Krock et al. 2009
2	AZA-2	856	362	262	<i>A. spinosum</i> <i>A. poporum</i> <i>Am. languida</i>	Krock et al. 2009 Krock et al. 2014 Tillmann et al. 2017
3	epi-AZA-7	858			<i>A. dexteroporum</i>	Rossi et al. 2017
4	AZA-11	872			<i>A. poporum</i>	Krock et al. 2014
5	AZA-33	716			<i>A. spinosum</i>	Kilcoyne et al. 2014
6	AZA-34	816			<i>A. spinosum</i>	{Kilcoyne et al. 2014
7	AZA-35	830	362	262	<i>A. spinosum</i> <i>A. dexteroporum</i>	Kilcoyne et al. 2014 Rossi et al. 2017
8	AZA-36	858			<i>A. poporum</i>	Krock et al. 2015
9	AZA-37	846			<i>A. poporum</i>	Krock et al. 2015
10	AZA-38	830			<i>Am. languida</i>	Krock et al. 2012
11	AZA-39	816			<i>Am. languida</i>	Krock et al. 2012
12	AZA-40	842			<i>A. poporum</i>	Krock et al. 2014
13	AZA-41	854	360	260	<i>A. poporum</i>	Krock et al. 2014
14	AZA-42	870	360	260	<i>A. poporum</i>	Krock et al. under review
15	AZA-43	828			<i>Am. languida</i>	Tillmann et al. 2017
16	AZA-50	842			<i>A. spinosum</i>	Tillmann et al. 2018
17	AZA-51	858			<i>A. spinosum</i>	Tillmann et al. 2018
18	AZA-52	830			<i>Am. languida</i>	Tillmann et al. 2018
19	AZA-53	830			<i>Am. languida</i>	Tillmann et al. 2018
20	AZA-54	870			<i>A. dexteroporum</i>	Rossi et al. 2017
21	AZA-55	868			<i>A. dexteroporum</i>	Rossi et al. 2017
22	AZA-56	884	362	262	<i>A. dexteroporum</i>	Rossi et al. 2017
23	AZA-57	844	362	262	<i>A. dexteroporum</i>	Rossi et al. 2017
24	AZA-58	828	362	262	<i>A. dexteroporum</i>	Rossi et al. 2017
25	AZA-59	860	362	262	<i>A. poporum</i>	Kim et al. 2017
26	AZA-62	870	362	262	<i>A. poporum</i>	Krock et al. 2019

26 AZAs from planktonic origin

Currently 62 published AZAs

And at least additional 10 known AZAs

## Azaspiracids - Variants

### AZAs produced by algae

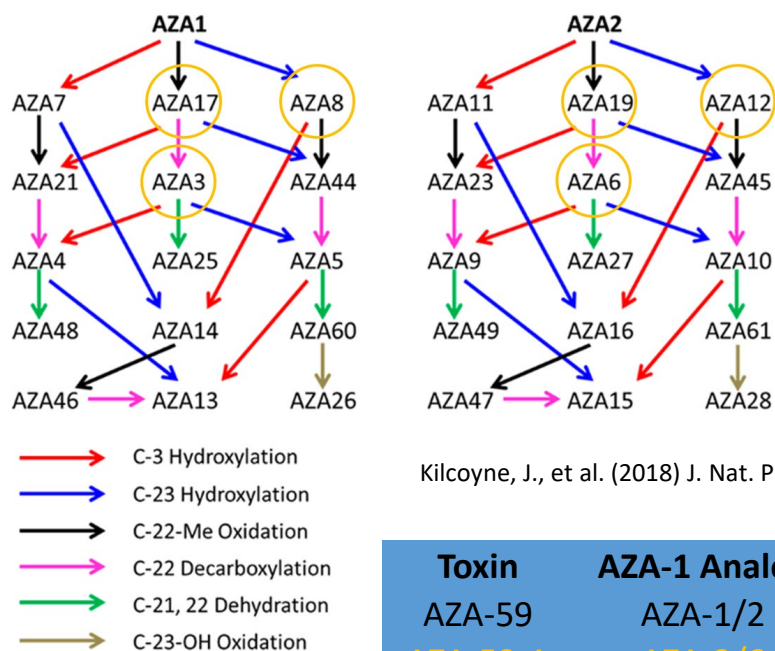
AZA-1	AZA-41
AZA-2	AZA-42
epi-AZA-7	AZA-43
AZA-11	AZA-50
AZA-33	AZA-51
AZA-34	AZA-52
AZA-35	AZA-53
AZA-36	AZA-54
AZA-37	AZA-55
AZA-38	AZA-56
AZA-39	AZA-57
AZA-40	AZA-58
AZA-59	AZA-62

### AZA shellfish metabolites of AZA-1 and -2

AZA-3	AZA-14	AZA-25	AZA-47
AZA-4	AZA-15	AZA-26	AZA-48
AZA-5	AZA-16	AZA-27	AZA-49
AZA-6	AZA-17	AZA-28	AZA-60
AZA-7	AZA-18	AZA-29	AZA-61
AZA-8	AZA-19	AZA-30	
AZA-9	AZA-20	AZA-31	
AZA-10	AZA-21	AZA-32	
AZA-11	AZA-22	AZA-44	
AZA-12	AZA-23	AZA-45	
AZA-13	AZA-24	AZA-46	

Two AZAs of phytoplankton origin result in 38 shellfish metabolites!

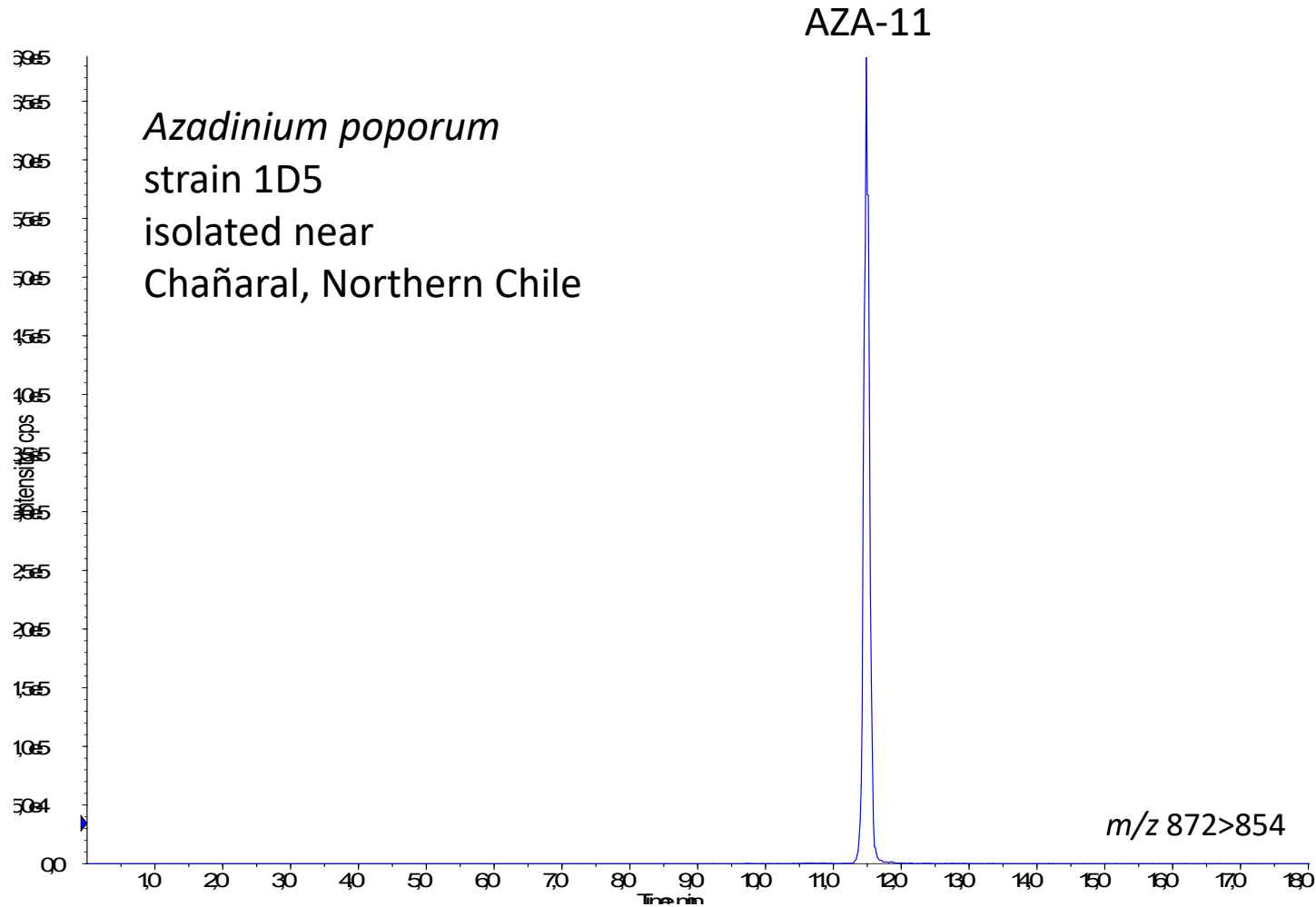
## Azaspiracids – Metabolism in Bivalves



Kilcoyne, J., et al. (2018) J. Nat. Prod. 81(4), 885-893.

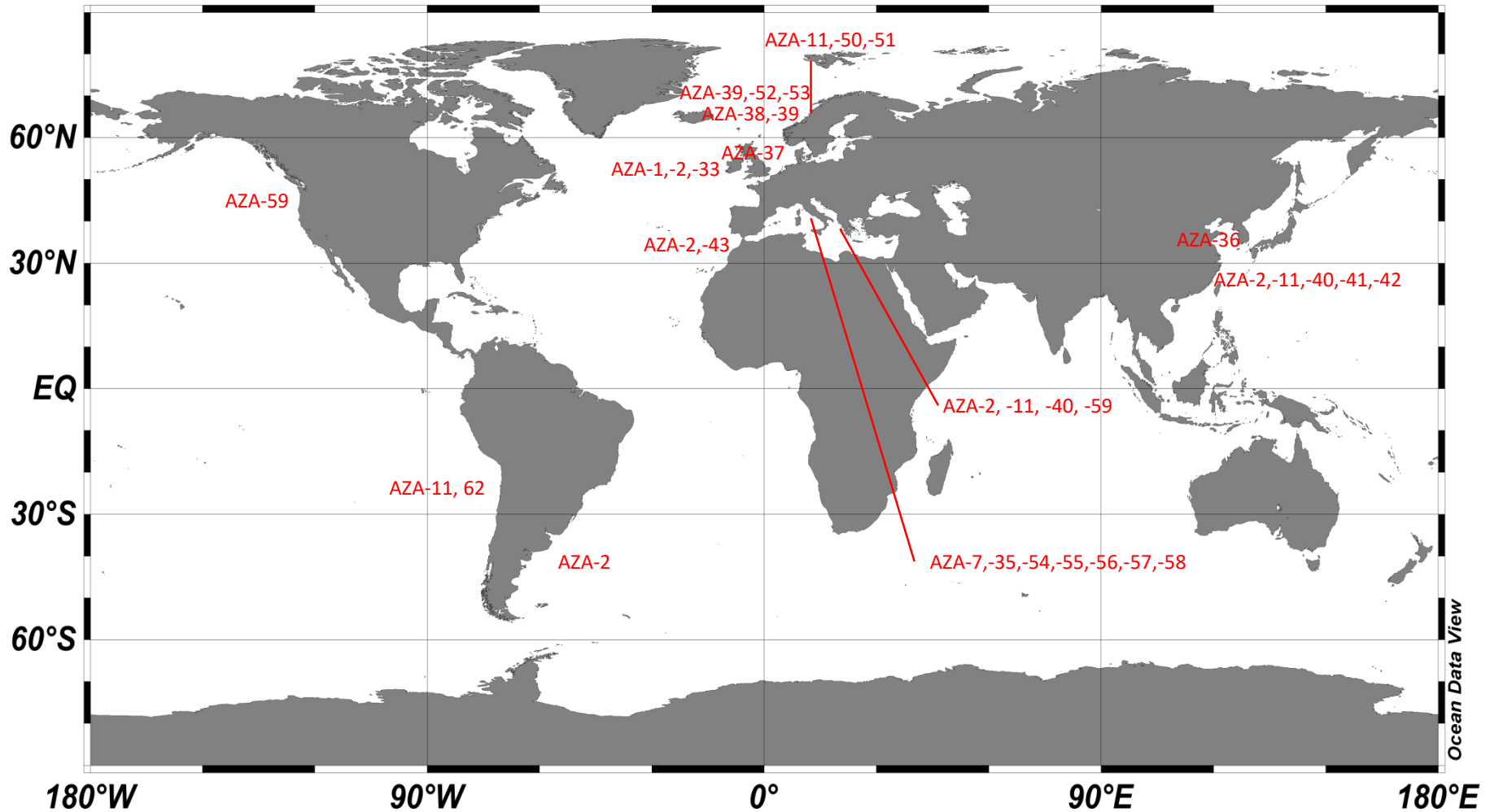
Toxin	AZA-1 Analog	m/z [M+H] <sup>+</sup>	m/z [M+H-H <sub>2</sub> O] <sup>+</sup>	Retentiontime [min]
AZA-59	AZA-1/2	860	842	10,6
AZA-59-A	AZA-3/6	846	828	10,1
AZA-59-B	AZA-8/12	876	858	9,6
AZA-59-C	AZA-17/19	890	872	9,9
AZA-59-D	?	890	872	10,3
AZA-59-E	No analog	892	874	9,0
AZA-59-F	No analog	878	860	9,65

# Variability of phycotoxins



Tillmann et al. (2017) J. Plankt. Res 39(2): 350-367)

## Azaspiracids – Geographic distribution





A wide-angle photograph of a massive glacier flowing into the sea. The glacier is a mix of white and light blue, with dark sediment lines visible. In the foreground, a dark blue research vessel with a white superstructure is on the water. The sky is overcast and grey.

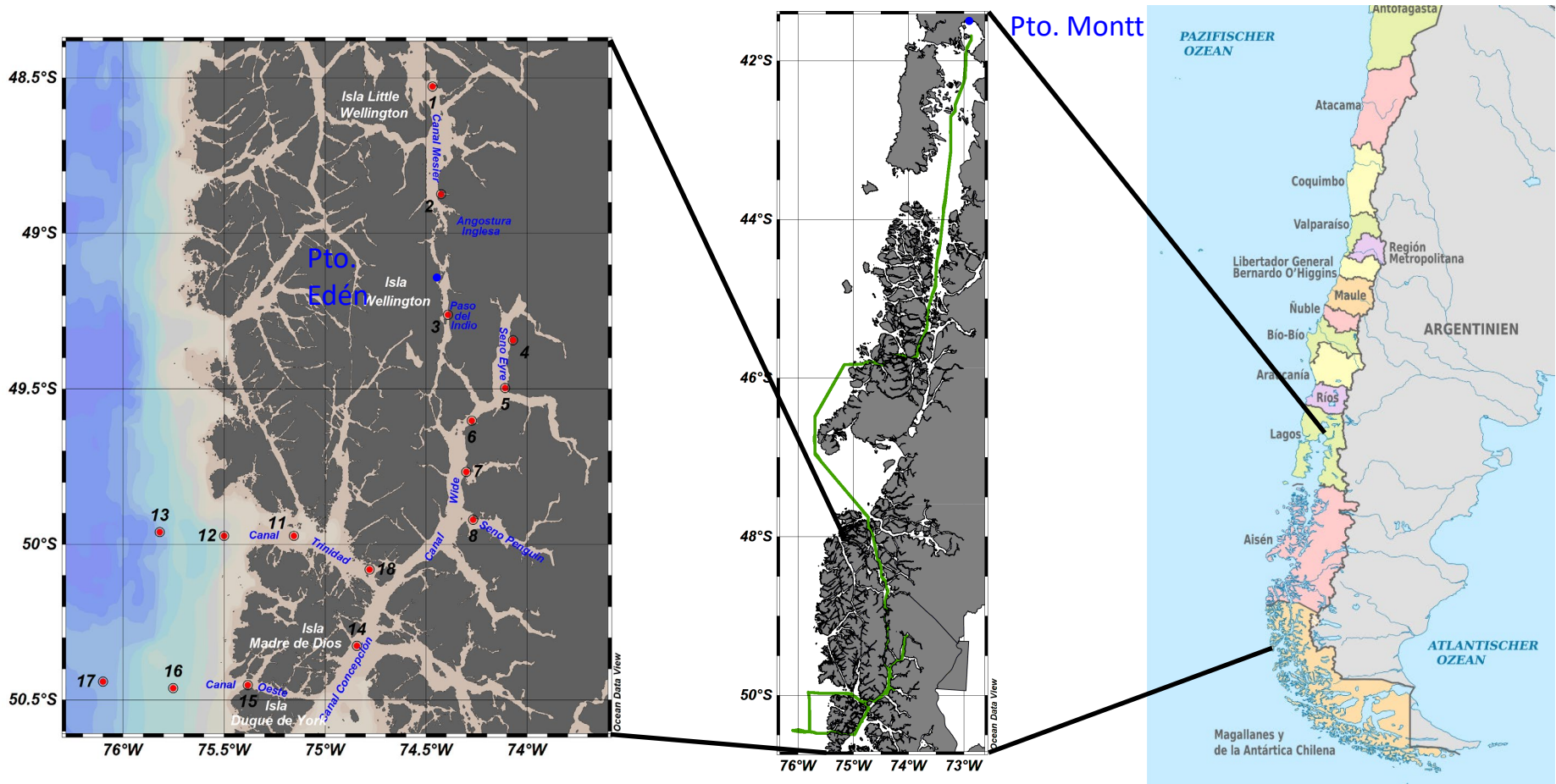
# Toxin distribution in the Última Esperanza Province (Southeast Pacific, Chile) during the PROFAN expedition November 2019



© Diego Nahuelhuén, IDEAL

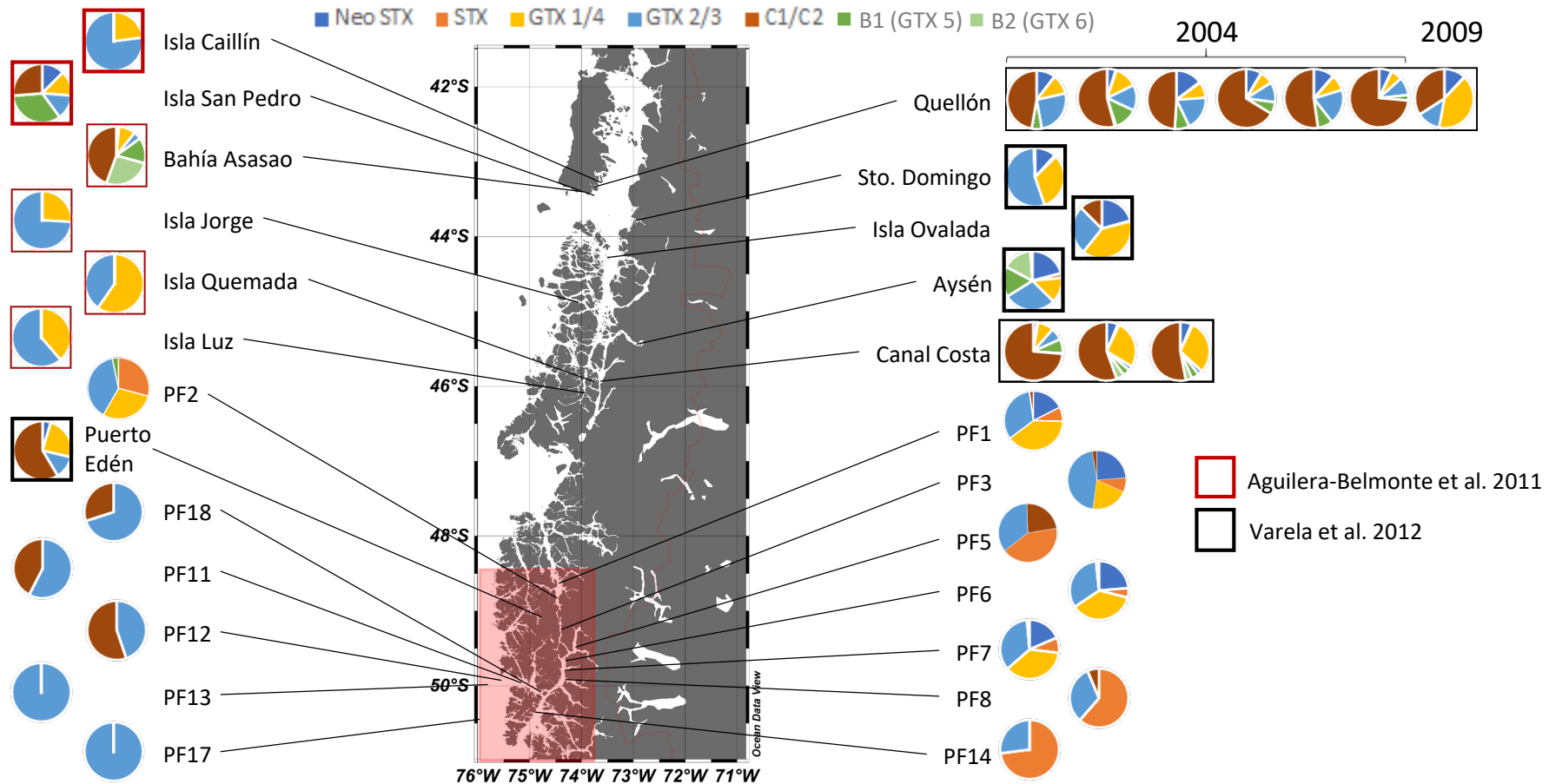
# PROFAN expedition (November 2019)

## Study area



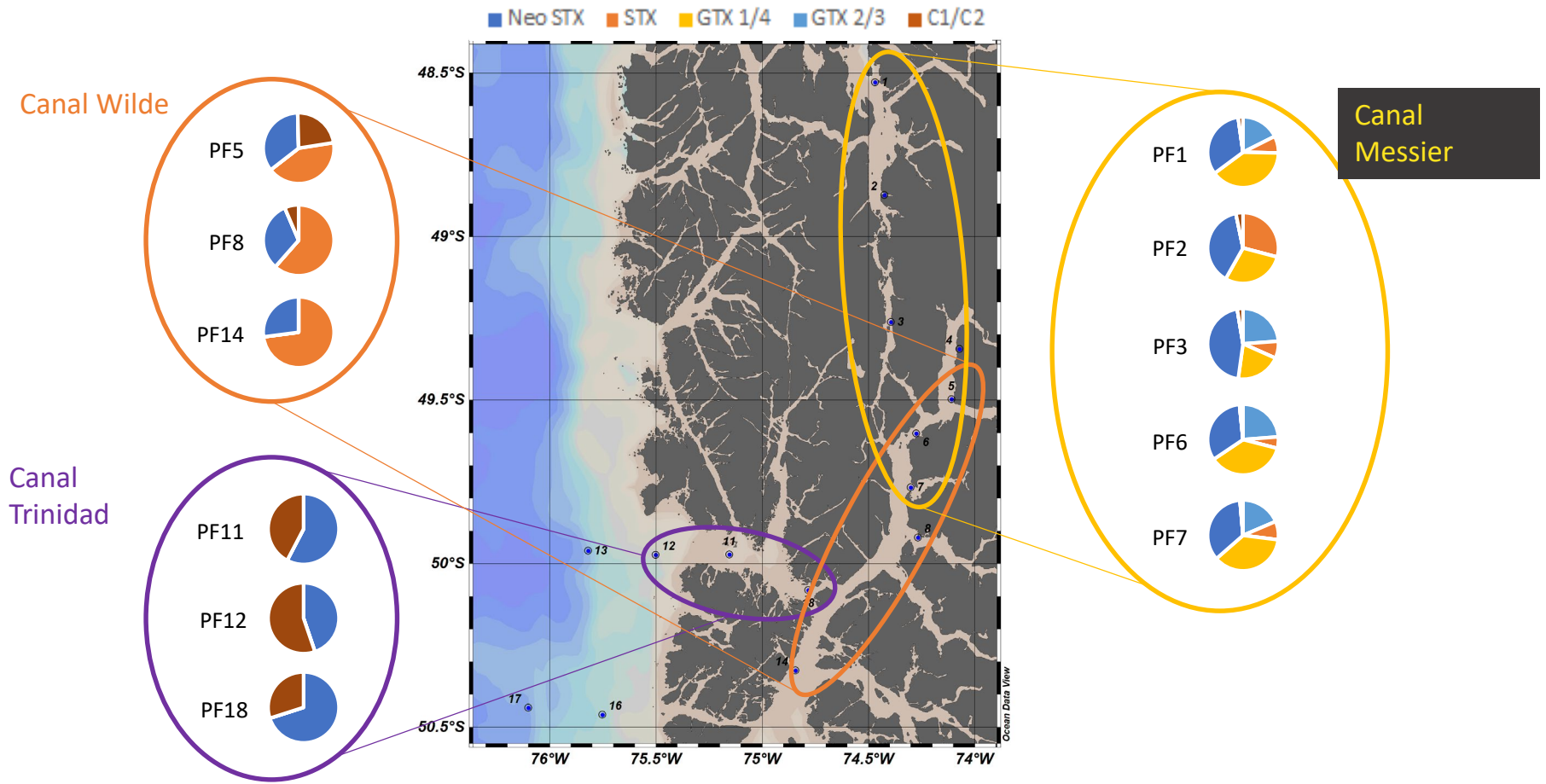
# PROFAN expedition (November 2019)

## 1. PSP-toxin – relative profiles



# PROFAN expedition (November 2019)

## 1. PSP-toxins – geographic distribution of profiles



## 1. PSP-toxins – species contributions?



*Alexandrium catenella*

*Alexandrium ostenfeldii*

Puerto  
Edén

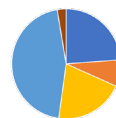
Varela et al. 2012



Cultured strains

Isla Vergara

Salgado et al. 2015



PF3

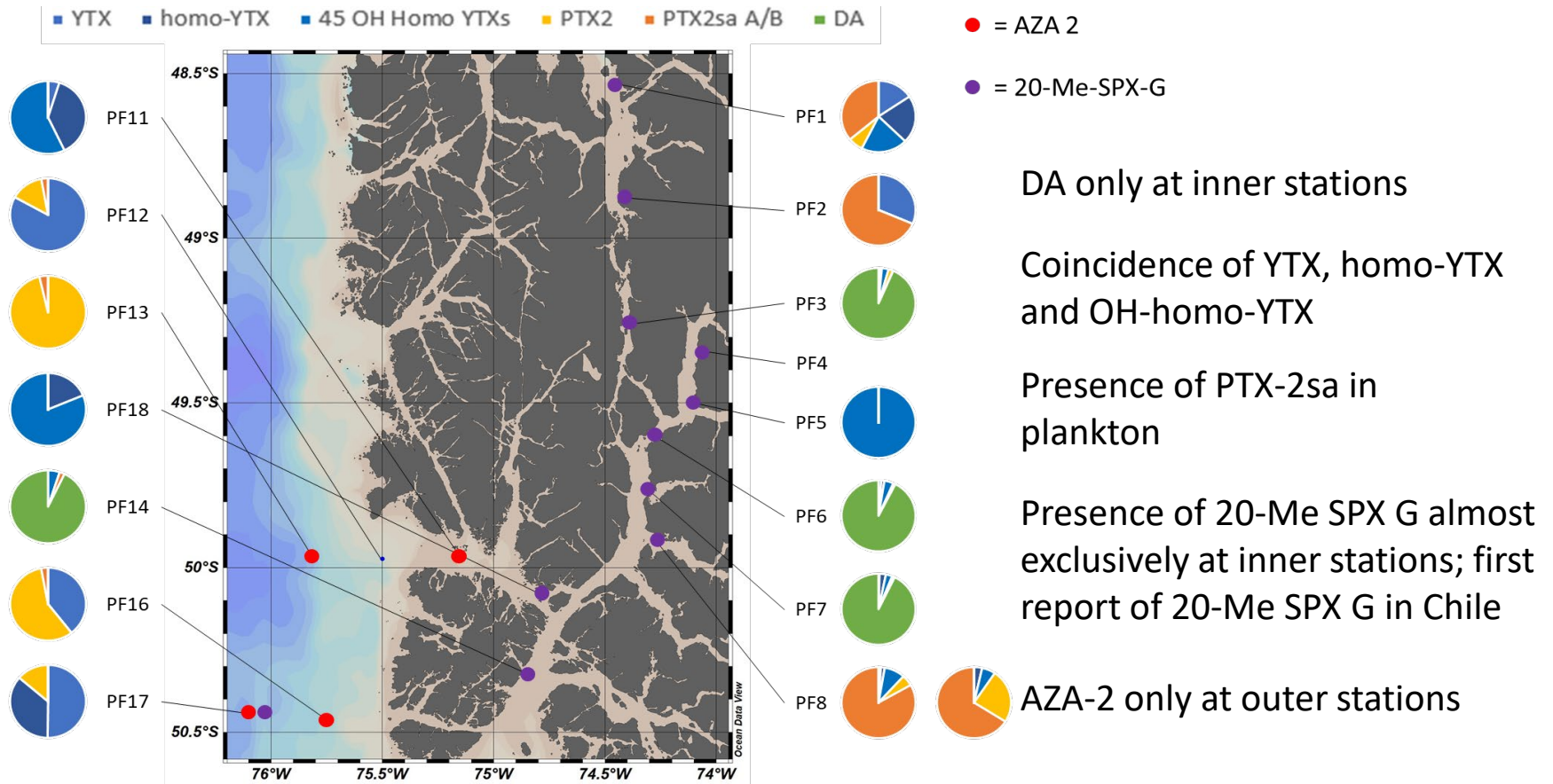


PF14

Planktonic field samples

# PROFAN expedition (November 2019)

## 2. Lipophilic toxins – qualitative profiles



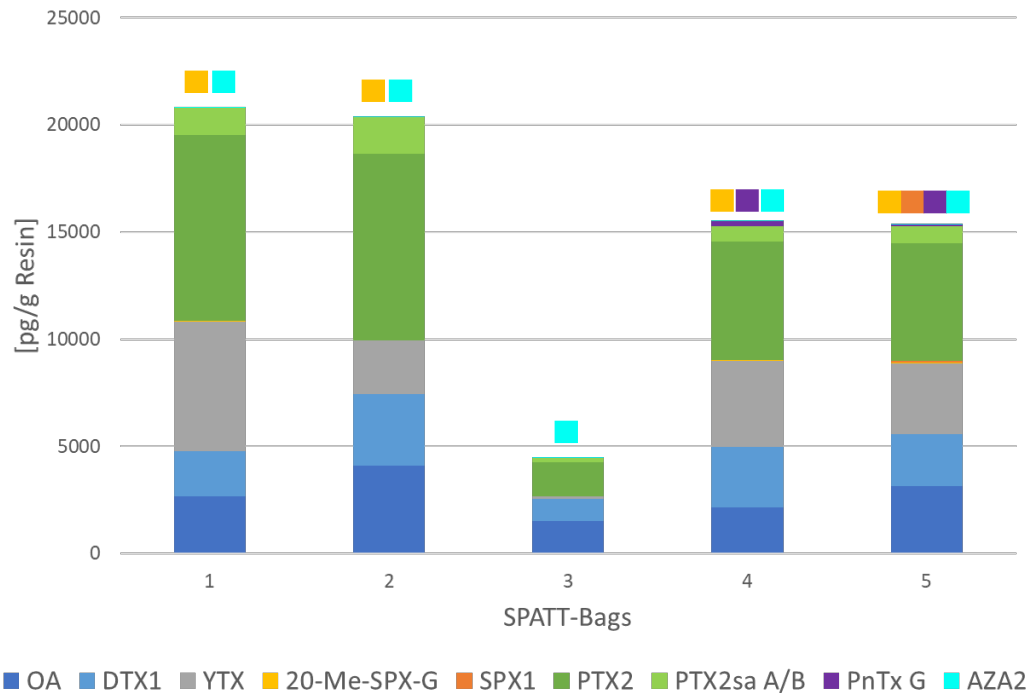
## 3. SPATT sampling

SPATT = Solid Phase Adsorption Toxin Tracking

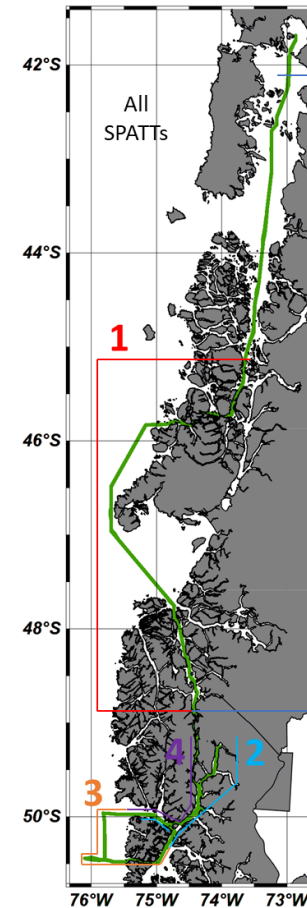
- Water is pumped through a porous synthetic resin  
→ passively adsorbing toxins
- Pros:
  - Time-integrating sampling  
→ Toxin dynamics
  - Low-cost; easy sampling→ Potential as an early warning system
- Cons:
  - Lack of calibration
  - Monitoring of dissolved toxins only  
→ no information on the current plankton presence



## 3. SPATT sampling



Möller et al. (2022) Prog. Oceanogr., in revision



**5** Toxin abundance higher in channels and fjords than in the open Pacific

OA/DTX-1 were detected during the entire track and were not restricted to Aysén Region

AZA-2 was present during the entire track at low abundances

First report of pinnatoxin G in Chile



## Main results

1. Phycotoxins were detected at all stations except for station 4 (Eyre Gulf, Pio XI Glaciar)
2. Geographically distinct distribution of PSP toxin profiles in Última Esperanza Province
3. DA, PTXs and YTXs were most abundant lipophilic toxins
4. Presence of homo-YTXs indicates the presence of other YTX-producing species than *Protoceratium reticulatum*
5. Even though no OA/DTX-1 were detected in plankton samples, these toxins were present in water samples during the entire cruise transect and indicate the presence of *Dinophysis acuta* also in the Magallanes Region
6. AZA-2 was detected in low concentrations in water samples during the entire cruise transect and seems to be the prevalent AZA variant in southern Chile
7. Pinnatoxin G (PnTx G) was detected for the first time in Chilean waters and strongly indicates the presence of *Vulcanodinium rugosum*

# M179

## FjordFlux



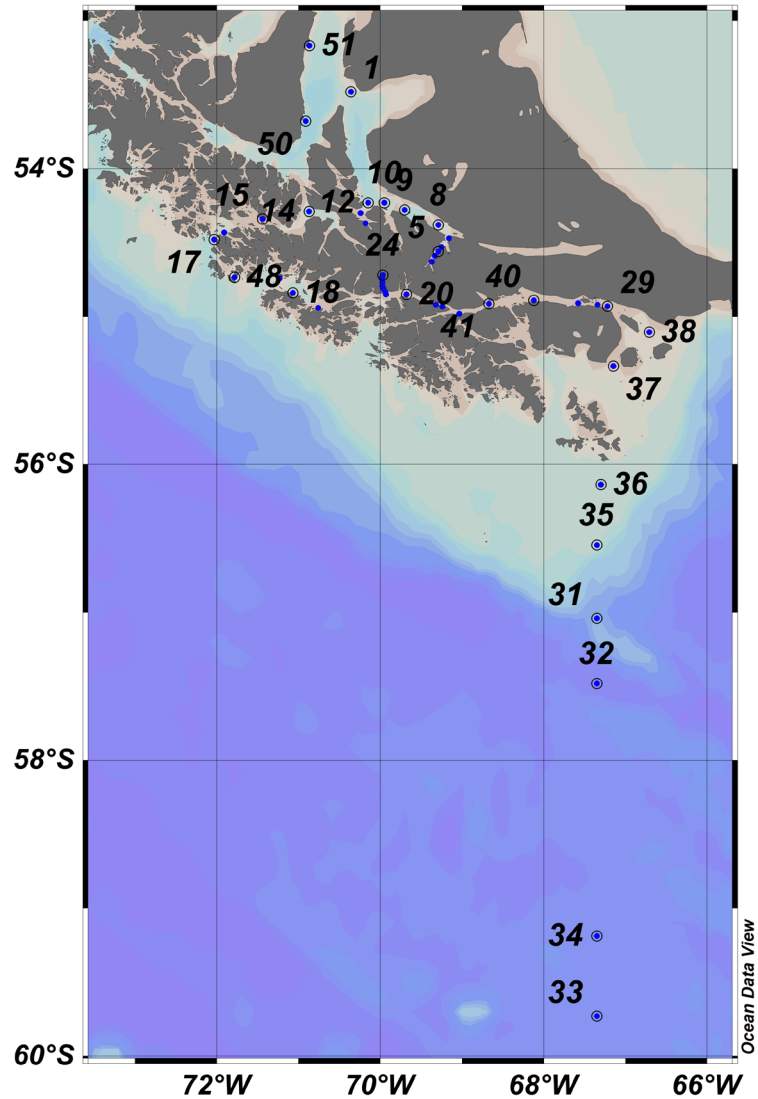
Further information:

<https://m.dw.com/es/una-ventana-al-futuro-desde-la-patagonia-la-campaña-trilateral-fjord-flux/a-60584473>

# FjordFlux expedition (January/February 2022)



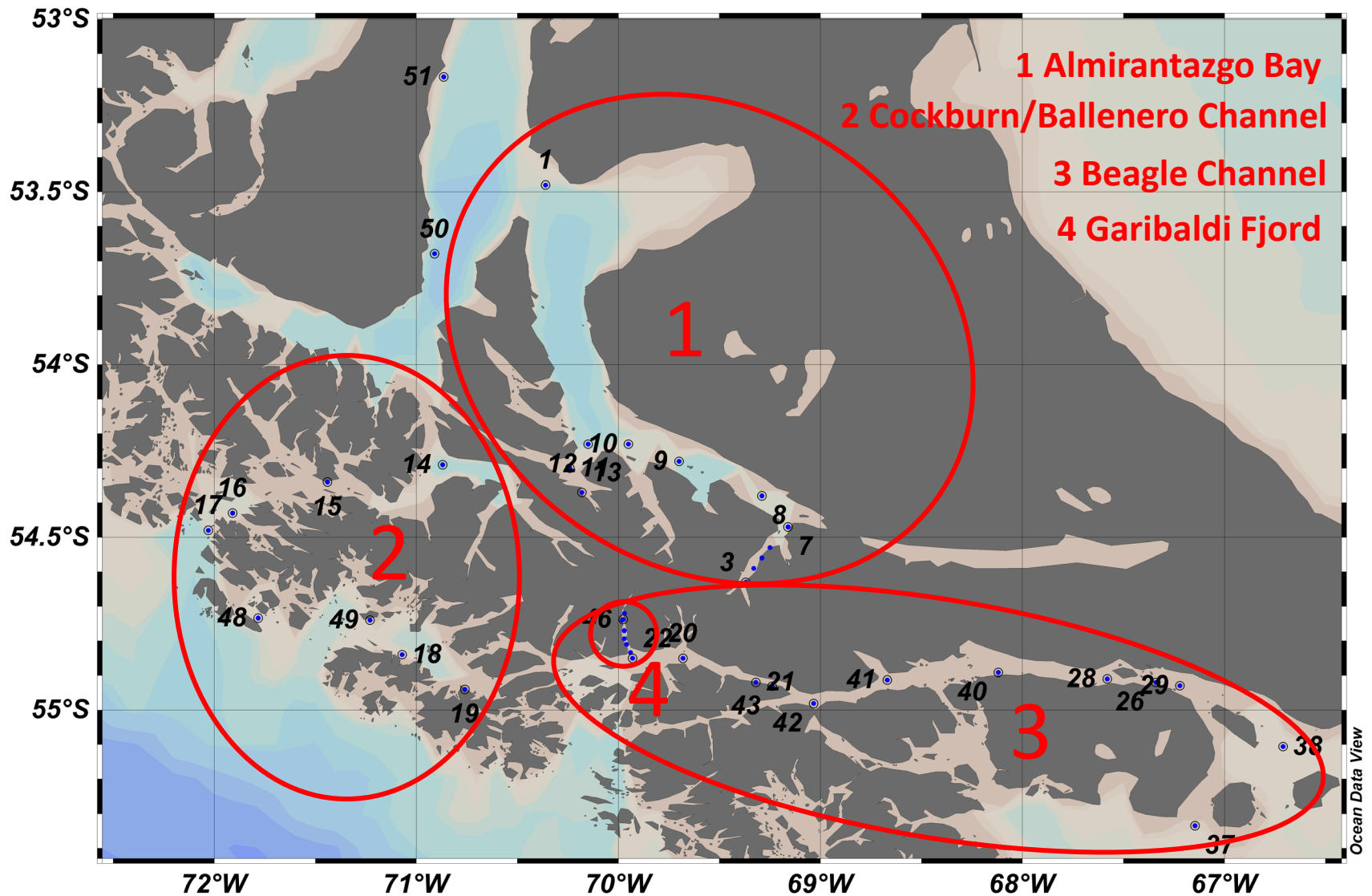
ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG



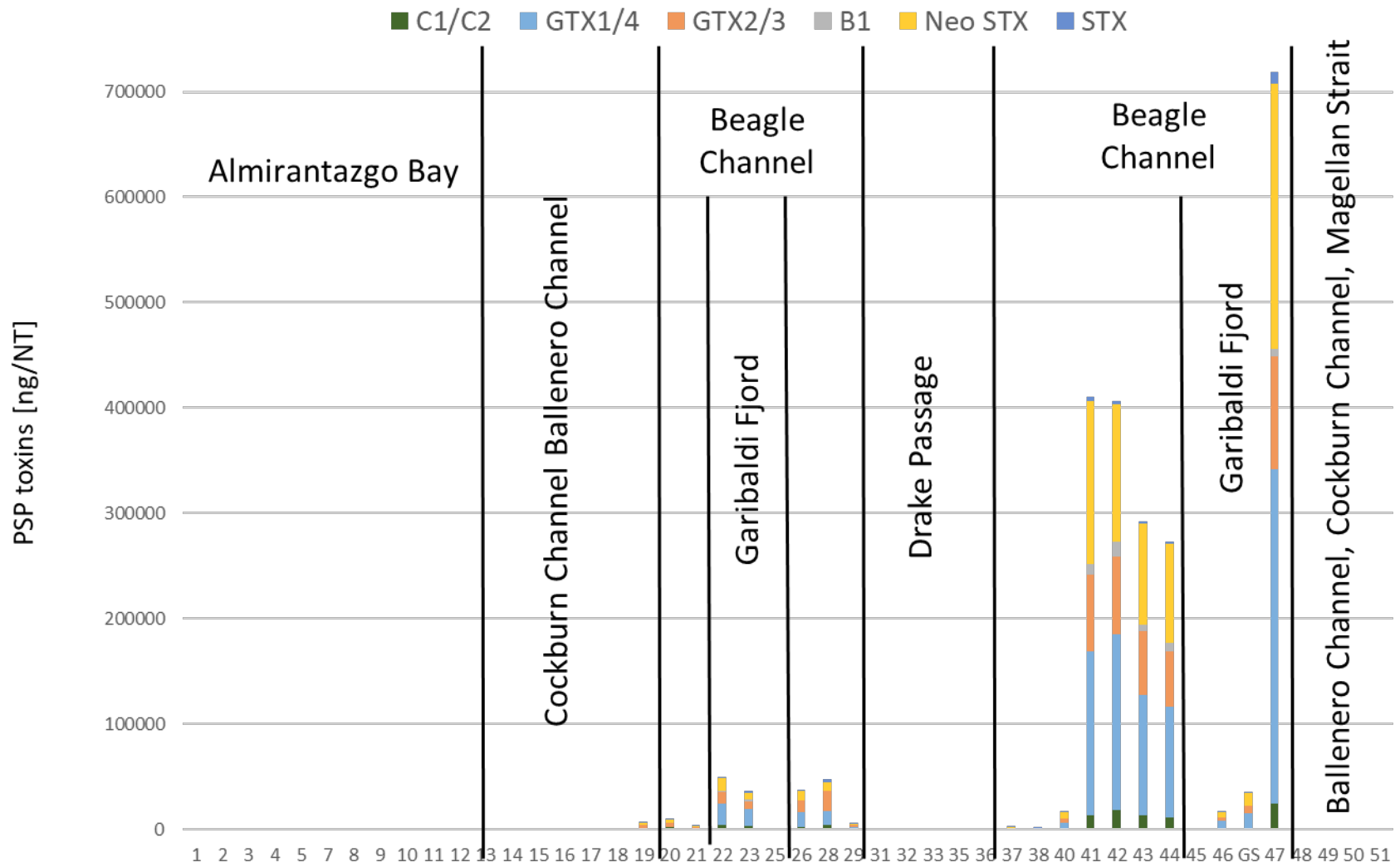
# FjordFlux expedition (January/February 2022)



ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG



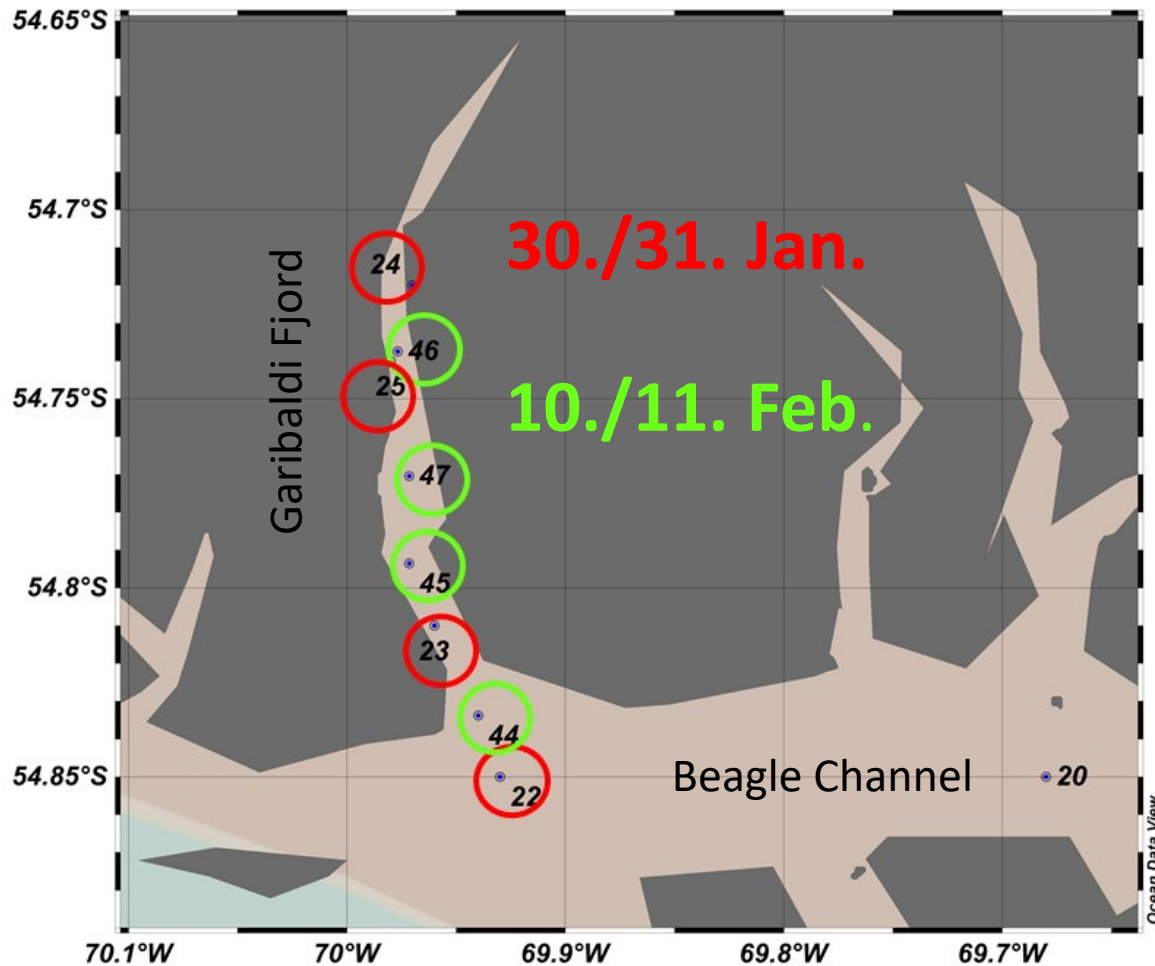
# FjordFlux expedition (January/February 2022) ALFRED-WEGENER-INSTITUT HELMHOLTZ-ZENTRUM FÜR POLAR- UND MEERESFORSCHUNG



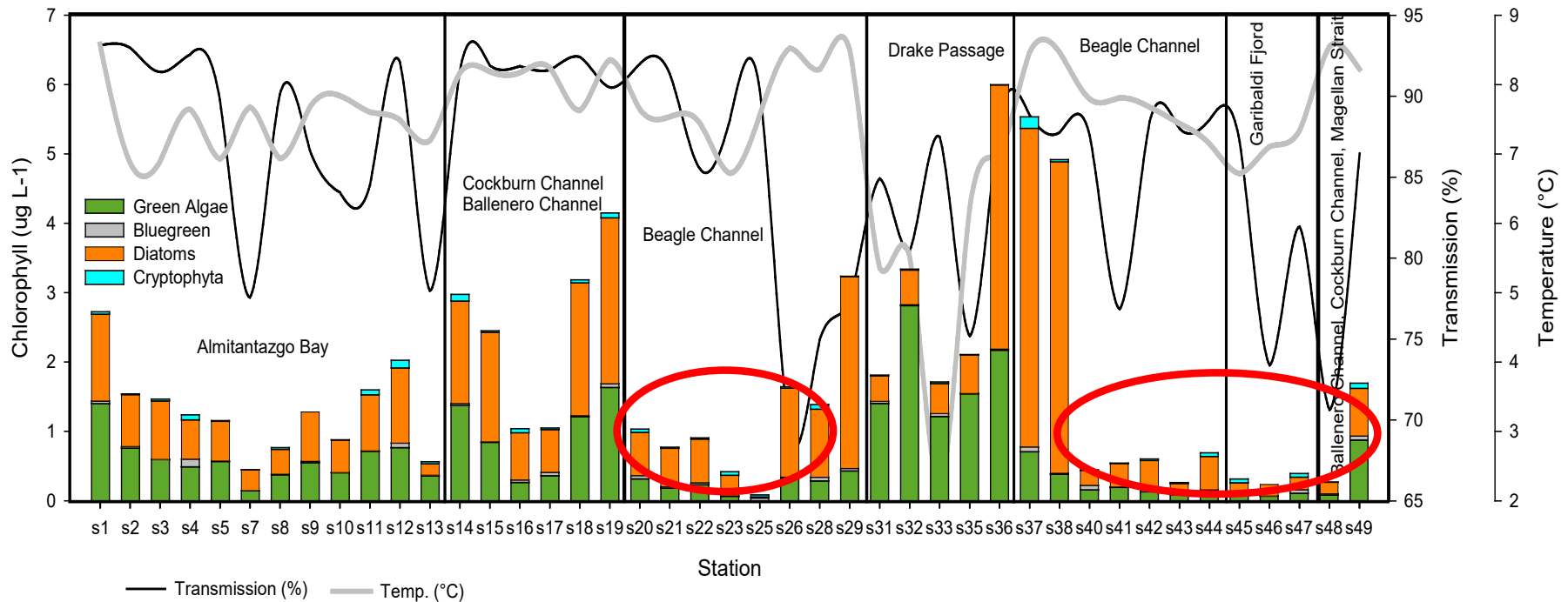
# FjordFlux expedition (January/February 2022)



ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG



## Functional plankton groups

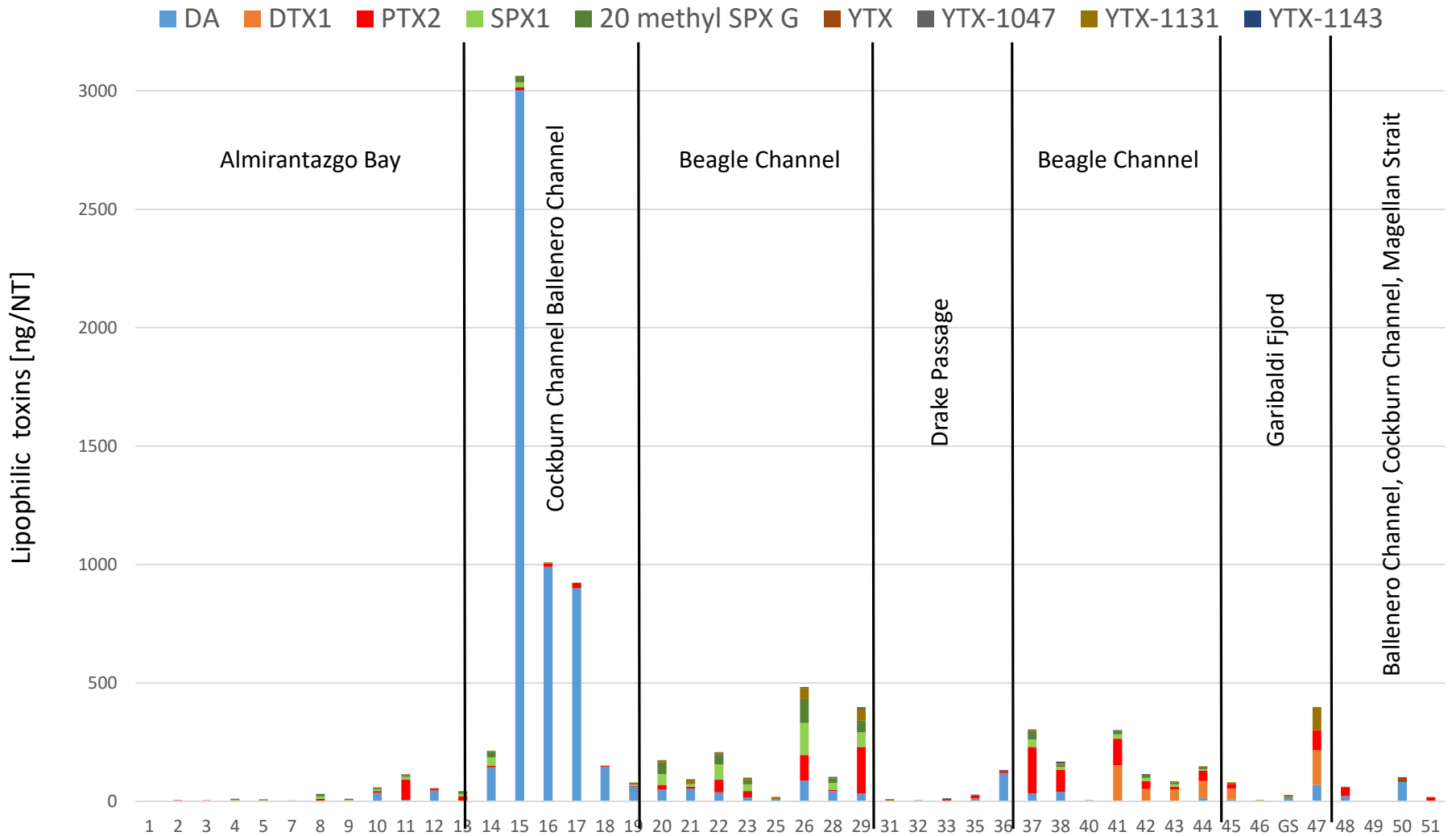


Data: Ricardo Giesecke, IDEAL

# FjordFlux expedition (January/February 2022)



ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG

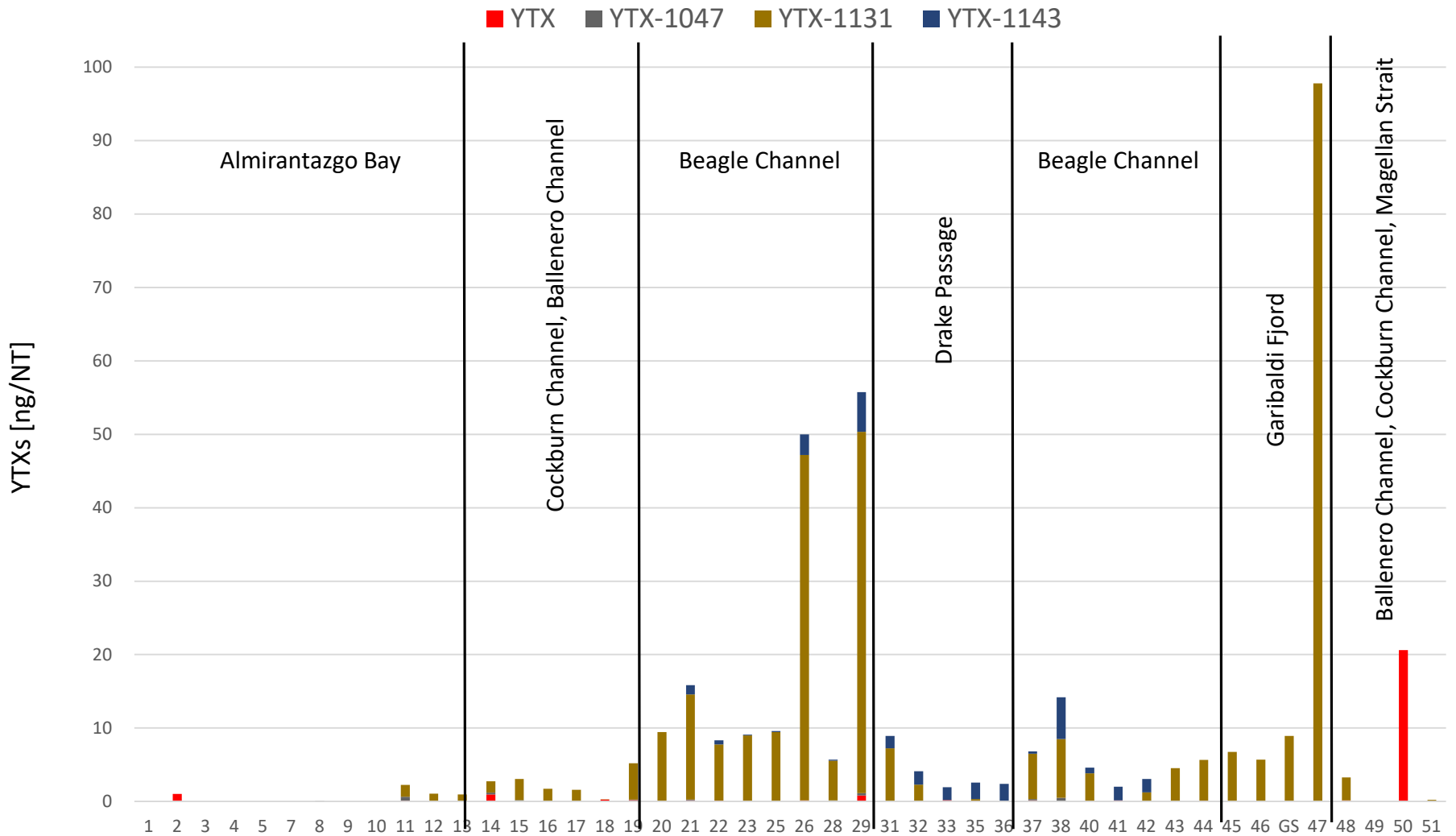




# FjordFlux expedition (January/February 2022)



ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG



1. Ichthyotoxins are rarely characterized, but pose an increasing threat to increasing aquacultural activities
2. Phycotoxin variability of known toxin classes is high and yet not fully explored, especially in hard to access coastal areas in Southern Chile
3. Phycotoxin profiles tend to be geographically relatively stable
4. Locally occurring phycotoxin variants should be included in chemical monitoring efforts in addition to officially regulated toxins



Any  
Questions?

Thanks for  
Your Attention!