University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Insecta Mundi

Center for Systematic Entomology, Gainesville, Florida

6-26-2020

Identification of planthoppers (Hemiptera: Delphacidae) intercepted on aquarium plants in Florida and elucidation of a potential pathway for exotic aquatic and semiaquatic pests

Susan E. Halbert

Matthew R. Moore

Charles R. Bartlett

Jade S. Allen

Follow this and additional works at: https://digitalcommons.unl.edu/insectamundi



Part of the Ecology and Evolutionary Biology Commons, and the Entomology Commons

This Article is brought to you for free and open access by the Center for Systematic Entomology, Gainesville, Florida at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Insecta Mundi by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

INSECTA TINDI A Journal of World Insect Systematics

0775

Identification of planthoppers (Hemiptera: Delphacidae) intercepted on aquarium plants in Florida and elucidation of a potential pathway for exotic aquatic and semiaguatic pests

Susan E. Halbert

Florida Department of Agriculture and Consumer Services Division of Plant Industry Gainesville, FL

Matthew R. Moore

Florida Department of Agriculture and Consumer Services Division of Plant Industry Gainesville, FL

Charles R. Bartlett

University of Delaware Newark, DE

Jade S. Allen

Florida Department of Agriculture and Consumer Services Division of Plant Industry Gainesville, FL

Date of issue: June 26, 2020

Susan E. Halbert, Matthew R. Moore, Charles R. Bartlett and Jade S. Allen Identification of planthoppers (Hemiptera: Delphacidae) intercepted on aquarium plants in Florida and elucidation of a potential pathway for exotic aquatic and semiaquatic pests

Insecta Mundi 0775: 1-6

ZooBank Registered: urn:lsid:zoobank.org:pub:A6789BD6-1430-4C25-8BB3-E5DB94C3BA5A

Published in 2020 by

Center for Systematic Entomology, Inc. P.O. Box 141874 Gainesville, FL 32614-1874 USA http://centerforsystematicentomology.org/

Insecta Mundi is a journal primarily devoted to insect systematics, but articles can be published on any non-marine arthropod. Topics considered for publication include systematics, taxonomy, nomenclature, checklists, faunal works, and natural history. Insecta Mundi will not consider works in the applied sciences (i.e. medical entomology, pest control research, etc.), and no longer publishes book reviews or editorials. Insecta Mundi publishes original research or discoveries in an inexpensive and timely manner, distributing them free via open access on the internet on the date of publication.

Insecta Mundi is referenced or abstracted by several sources, including the Zoological Record and CAB Abstracts. Insecta Mundi is published irregularly throughout the year, with completed manuscripts assigned an individual number. Manuscripts must be peer reviewed prior to submission, after which they are reviewed by the editorial board to ensure quality. One author of each submitted manuscript must be a current member of the Center for Systematic Entomology.

Guidelines and requirements for the preparation of manuscripts are available on the Insecta Mundi website at http://centerforsystematicentomology.org/insectamundi/

Chief Editor: David Plotkin, insectamundi@gmail.com Assistant Editor: Paul E. Skelley, insectamundi@gmail.com

Head Layout Editor: Robert G. Forsyth **Editorial Board:** J. H. Frank, M. J. Paulsen

Founding Editors: Ross H. Arnett, Jr., Virendra Gupta, John B. Heppner, Lionel A. Stange, Michael C. Thomas,

Robert E. Woodruff

Review Editors: Listed on the Insecta Mundi webpage

Printed copies (ISSN 0749-6737) annually deposited in libraries

CSIRO, Canberra, ACT, Australia Museu de Zoologia, São Paulo, Brazil

Agriculture and Agri-Food Canada, Ottawa, ON, Canada

The Natural History Museum, London, UK

Muzeum i Instytut Zoologii PAN, Warsaw, Poland

National Taiwan University, Taipei, Taiwan

California Academy of Sciences, San Francisco, CA, USA

Florida Department of Agriculture and Consumer Services, Gainesville, FL, USA

Field Museum of Natural History, Chicago, IL, USA

National Museum of Natural History, Smithsonian Institution, Washington, DC, USA

Zoological Institute of Russian Academy of Sciences, Saint-Petersburg, Russia

Electronic copies (Online ISSN 1942-1354, CDROM ISSN 1942-1362) in PDF format

Printed CD or DVD mailed to all members at end of year. Archived digitally by Portico.

Florida Virtual Campus: http://purl.fcla.edu/fcla/insectamundi

University of Nebraska-Lincoln, Digital Commons: http://digitalcommons.unl.edu/insectamundi/

Goethe-Universität, Frankfurt am Main: http://nbn-resolving.de/urn/resolver.pl?urn:nbn:de:hebis:30:3-135240

Copyright held by the author(s). This is an open access article distributed under the terms of the Creative Commons, Attribution Non-Commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited. http://creativecommons.org/licenses/by-nc/3.0/

Layout Editor for this article: Robert G. Forsyth

Identification of planthoppers (Hemiptera: Delphacidae) intercepted on aquarium plants in Florida and elucidation of a potential pathway for exotic aquatic and semiaquatic pests

Susan E. Halbert

Florida Department of Agriculture and Consumer Services Division of Plant Industry Gainesville, FL Susan.Halbert@FDACS.gov

Matthew R. Moore

Florida Department of Agriculture and Consumer Services Division of Plant Industry Gainesville, FL Matthew.Moore@FDACS.gov

Charles R. Bartlett

University of Delaware Newark, DE Bartlett@udel.edu

Jade S. Allen

Florida Department of Agriculture and Consumer Services Division of Plant Industry Gainesville, FL Jade.Allen@FDACS.gov

Abstract. Recent shipments of aquarium plants to pet stores in five Florida counties were found to be infested with an exotic delphacid planthopper. Rearing adult males allowed identification by morphological analysis. Molecular analysis confirmed that it was the same as authoritatively identified reference specimens of the planthopper, *Opiconsiva anacharsis* (Fennah) (new combination) (Hemiptera: Delphacidae), first reported from Florida in 1989 and known to be established only in Broward County. The host plants, *Echinodorus* spp. Rich. ex Engelm. (Alismatales: Alismataceae), originally from Thailand, were sold in enclosed plastic cylinders that provided a suitable environment for maintaining the planthoppers. Attempts to trace the shipment histories to these stores suggested a circuitous multi-state pathway leading to a Broward County, Florida, business that receives aquatic plants from Southeast Asia. While the infestation of these plants may have occurred in Florida, trade in semi-emergent aquatic plants is shown to be a potential pathway for introduction for insect pests.

Key words. *Harmalia anacharsis*, regulatory entomology.

Introduction

Florida has many adventive arthropods (Frank and McCoy 1992, 1995). Reasons include Florida's climate, ranging from temperate to subtropical, that allows insects to become established, high numbers of visitors and recent immigrants who may bring plants and plant products, high diversity of ornamental plants used in landscaping, and commerce in plant products to serve over 20 million residents. Florida is an international gateway providing plant products destined for the entire USA. One of the primary focuses of the Florida Department of Agriculture and Consumer Services, Division of Plant Industry (DPI) is to identify exotic pests and diseases early enough to prevent establishment in the state and potential spread to other locations. Knowledge of introduction pathways can be used to design science-based regulations to prevent future establishment of exotic pests.

Kennedy et al. (2012) listed 128 delphacid planthoppers known in Florida. Of those, two are definitively adventive (Opiconsiva anacharsis (Fennah) and Perkinsiella saccharicida Kirkaldy), and two others (Sogatella kolophon (Kirkaldy) and Metadelphax propinqua (Fieber)) are probably adventive (Meagher et al. 1991; Wooten et al. 1993; Gonzon and Bartlett 2007; Hamilton 2010; Bartlett et al. 2014). On the other hand, Metadelphax propinqua, based on its widespread distribution (Bartlett et al. 2014), could be a circumtropical species (Stephen W. Wilson, University of Central Missouri, personal communication 2020). Since Bartlett et al. (2014), three additional adventive species (Tarophagus colocasiae (Matsumura), Opiconsiva tangira (Matsumura), and Pissonotus muiri Van Duzee) have been reported (Halbert and Bartlett 2015; Halbert 2016a, b), and an additional species was introduced as a biological control agent (Megamelus scutellaris Berg; Tipping et al. 2014). Harmalia anacharsis recently was transferred into Opiconsiva Distant by Huang et al. (2017) because the genus Harmalia Fennah was found to be a derived lineage nested within Opiconsiva, making Harmalia a junior subjective synonym. Because Huang et al. (2017) did not specify new combinations, this appears to be the first explicit use of the new combination Opiconsiva anacharsis (Fennah).

Opiconsiva anacharsis was discovered in Broward County, FL in 1989 (Wooten et al. 1993) and is considered to be established in the county. It is widely distributed across the Pacific islands and southeast Asia, where it feeds on aquatic plants. It is known from rice fields but is not considered a pest of the crop (Wooton et al. 1993). The only observed host plants in Florida, based on DPI records, are species in *Echinodorus* Rich. ex Engelm. (Alismatales: Alismataceae) and *Nymphaea* (L.) (Nymphaeales: Nymphaeaceae). So far, Florida records are restricted to Broward County, and recent finds in pet stores are interceptions. The original source of Florida's Broward County population of *O. anacharsis* is unknown. Here we report possible interceptions of *O. anacharsis* associated with aquatic plants from Thailand for use in aquariums.

Materials and Methods

DPI inspectors routinely inspect businesses that sell plants and plant products, including pet stores. Any live insects and infested plants found during these inspections are submitted to DPI Entomology staff for identification.

Several intercepted containers of host materials (Fig. 1A), all marked "Product of Thailand," with live reproducing colonies of *O. anacharsis* were held in the DPI Maximum Security Quarantine (BSL 2) for several days to obtain adult males from the colonies. Specimens were preserved in 70% or 95% ethanol, for morphological and molecular analysis, respectively.

Specimens of intercepted delphacids were identified using available literature (e.g. Fennah 1969), and by comparison with authoritatively identified reference specimens in the Florida State Collection of Arthropods, Gainesville, FL, USA (FSCA). For delphacids, adult males are necessary for confident identification. Abdomens of male specimens were removed, gently heated in 10% KOH, rinsed in water, and stored in glycerin in microvials. Each microvial was attached by a silicone plug to the pin that holds the rest of the specimen. Voucher specimens were deposited in the FSCA. After removing the planthoppers, all remaining material was autoclaved for disposal.

Recently intercepted and Florida established *O. anacharsis* specimens were DNA extracted using the Qiagen DNeasy Blood and Tissue Kit. PCRs were conducted using the standard COI barcoding primers LCO1490 and HCO2198 (Folmer et al. 1994). Purified PCR products were sequenced at DPI on an Applied Biosystems SeqStudio Genetic Analyzer using BigDye Terminator v3.1 chemistry. Sequence traces were trimmed and assembled into contigs in Sequencher 5.4.6 (Gene Codes Corporation). Newly generated sequences (GenBank accessions: MN922322–MN922327) were aligned in MEGA7 (Kumar et al. 2016) using the default settings of MUSCLE (Edgar 2004). Sequences were queried to BOLD (Ratnasingham and Hebert 2007) and GenBank using BLASTn (Altschul et al. 1990).

Results

Five samples of *O. anacharsis* were submitted from *Echinodorus* spp. plants sold in pet stores for use in aquariums (Fig. 1A). All the plants were marked "Product of Thailand." The samples came from stores in five different Florida counties, including Broward County, where the only known established population occurs. The plants were packaged in sealed clear plastic cylinders. Each cylinder had an inner container filled with gel and pearlite to sustain the roots. Four of the samples had live, reproducing colonies of *H. anacharsis* in at least one container. In one sample, the insects were dead, and the plant was in poor condition.

Insects were identified morphologically using male genitalic characters (viz. Fennah 1969) (Fig. 1 B-D). Molecular sequences of the intercepted *O. anacharsis* were found to be a 100% match with a specimen from the established Florida population of *O. anacharsis*. BOLD contained only one close match (99.8%) to our new sequences, which was an unidentified delphacid from Bangladesh (GMBCH3433-15; BIN BOLD: ACZ2094). GenBank BLASTn searches did not yield any additional sequence matches.

Discussion

Our attempts to trace the source of the infested plants revealed a possible pathway that was long and circuitous. It led first to Arizona, then to California, and then back to a business in Broward County, Florida, that receives thousands of aquatic plants from southeastern Asia monthly. Although the plants were clearly marked "Product of Thailand," it is possible that the infestation came from the established population in Broward County, FL. A subsequent survey of the *Echinodorus* plants at the business revealed a single adult male *O. anacharsis*.

These interceptions indicate movement of aquarium plants could be a pathway of introduction for planthoppers (both from foreign sources and from other states). This pathway is the likely source of Florida's established population of *O. anacharsis*.

Other insects are suspected to have come to Florida on aquarium plants, including two species of Asian corixid bugs (Polhemus and Rutter 1997; Polhemus and Golia 2006). Our first experience with pet stores as a pathway for exotic insect interceptions came in 2016, when several species of Asian wood-boring beetles were found in sticks from China sold as chew toys for pet rodents (Skelley 2016). Subsequent inspections of aquatic plants sold for aquariums intercepted a soft scale, Coccus moestus De Lotto (Stocks 2016), a thrips, Copidothrips octarticulatus (Schmutz) (Williams 2016), and a moth, Paraponyx sp. (Hayden 2016). The scale, C. moestus, is recorded from several Caribbean, African, and Asian countries and, along with aquatic plants, infests several important fruit tree species and ornamental plants (García Morales et al. 2020). The thrips, C. octarticulatus, is an Asian species that has been moving globally, but is not known from Florida and is a potential agricultural pest. The moth, Paraponyx sp. is known from aquatic plants.

Southeast Asia has several serious delphacid pests, especially in rice (Wilson and O'Brien 1987; Wilson and Claridge 1991; Wilson 2005). It is possible that planthopper pests more serious than *O. anacharsis* could follow the aquarium plant pathway and be unintentionally introduced into Florida and other states.

Acknowledgments

We thank FDACS-DPI staff: inspectors Melanie Cain, Lisa Tyler, Mark Laurint, Samuel Hart, Justin Anto, and Kevin Williams for samples; David Davison and Nicole Casuso for arranging space in the DPI Maximum Security Quarantine; Cheryl Roberts and Lynn Combee for their assistance gathering the COI barcode sequence data; Cheryl Jones, for foreign shipment records; and Catherine White, for composing the figure. We thank Steven W. Wilson, University of Central Missouri, and MJ Paulsen, University of Nebraska, for reviewing the manuscript. We thank the Florida Department of Agriculture and Consumer Services, Division of Plant Industry for support of this work.

Literature Cited

- Altschul, S. F., W. Gish, W. Miller, E. W. Myers, and D. J. Lipman. 1990. Basic local alignment search tool. Journal of Molecular Biology 215: 403–410.
- Bartlett, C. R., L. B. O'Brien, and S. W. Wilson. 2014. A review of the planthoppers (Hemiptera: Fulgoroidea) of the United States. Memoirs of the American Entomological Society 50: 1–287.
- **Edgar, R. C. 2004.** MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Research 32: 1792–1797.
- **Fennah, R. G. 1969.** Fulgoroidea (Homoptera) from New Caledonia and the Loyalty Islands. Pacific Insects Monograph 21: 1–116.
- **Folmer, O., M. Black, W. Hoeh, R. Lutz, and R. Vrijenhoek. 1994.** DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3: 294–299.
- Frank, J. H., and E. D. McCoy. 1992. The immigration of insects to Florida, with a tabulation of records published since 1970. Florida Entomologist 75: 1–28.
- Frank, J. H., and E. D. McCoy. 1995. Invasive adventive insects and other organisms in Florida. Florida Entomologist 78: 1–15.
- García Morales, M., B. D. Denno, D. R. Miller, G. L. Miller, Y. Ben-Dov, and N. B. Hardy. 2020. ScaleNet. *Coccus moestus*. Available at http://scalenet.info/catalogue/Coccus%20moestus/ (Last accessed May 2020.)
- Gonzon, A. T., and C. R. Bartlett. 2007. Systematics of Hadropygos n. g., Metadelphax Wagner and New World Toya Distant (Hemiptera: Delphacidae). Transactions of the American Entomological Society 133(3/4): 205–277.
- **Halbert, S. E. 2016a.** Entomology section, *Opiconsiva tangira*. p. 6, 10. *In*: P. J. Anderson and G. S Hodges (eds.). Tri-Ology 55(3): 1–16.
- **Halbert, S. E. 2016b.** Entomology section, *Pissonotus muiri*. p. 7, 13. *In*: P. J. Anderson and G. S Hodges (eds.). Tri-Ology 55(4): 1–21.
- Halbert, S. E., and C. R. Bartlett. 2015. The taro planthopper, *Tarophagus colocasiae* (Matsumura), a new delphacid planthopper in Florida. Florida Department of Agriculture & Consumer Services, Division of Plant Industry. Pest Alert, FDACS-P-02040. 3 p.
- **Hamilton, K. G. A. 2010.** Chapter 19."Short-horned" bugs (Homoptera) of the Atlantic Maritime Ecozone. p. 405–419. *In*: D. McAlpine (ed.). Assessment of species diversity in the Atlantic Maritime Ecozone. NRC Press Biodiversity Monograph Series; Ottawa. 785 p.
- **Hayden, J. E. 2016.** Entomology section, *Paraponyx* sp. p. 12. *In*: P. J. Anderson and G. S. Hodges (eds.). Tri-Ology 55(4): 1–21.
- Huang, Y.-X., L.-F. Zheng, C. R. Bartlett, and D.-Z. Qin. 2017. Resolving phylogenetic relationships of Delphacini and Tropidocephalini (Hemiptera: Delphacidae: Delphacinae) as inferred from four genetic loci. Scientific Reports 7(1): 3319. doi:10.1038/s41598-017-03624-w
- **Kennedy, A. C., C. R. Bartlett, and S. W. Wilson. 2012.** An annotated checklist of the delphacid planthoppers (Hemiptera: Delphacidae) of Florida with the description of three new species and the new genus, *Meristopsis*. Florida Entomologist 95: 395–421.
- Kumar, S., G. Stecher, and K. Tamura. 2016. MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular Biology and Evolution 33: 1870–1874.
- Meagher, R. L., Jr., S. W. Wilson, R. S. Pfannenstiel, and R. G. Breene. 1991. Documentation of two potential insect pests of south Texas sugarcane. Southwestern Entomologist 16(4): 365–366.
- **Polhemus, J. T., and V. Golia. 2006.** *Micronecta ludibunda* Breddin (Heteroptera: Corixidae: Micronectinae), the second Asian water bug introduced into Florida, U.S.A. Entomological News 117: 531–534.
- **Polhemus, J. T., and R. P. Rutter. 1997.** *Synaptonecta issa* (Heteroptera: Corixidae), first New World record of an Asian water bug in Florida. Entomological News 108 (4): 300–304.
- **Ratnasingham, S., and P. D. N. Hebert. 2007.** BOLD: The Barcode of Life Data System (http://www.barcodinglife.org). Molecular Ecology Notes 7: 355–364.
- **Skelley, P. E. 2016.** *Glaphyra* sp., *Plagionotus* sp., *Amamidytus* sp., and *Coccographis nigrorubra*. p. 3, 6–7. *In*: P. J. Anderson and G. S. Hodges (eds.). Tri-Ology 55(1): 1–17.

- **Stocks, I. C. 2016.** Entomology section. *Coccus moestus*. p. 8. *In*: P. J. Anderson and G. S. Hodges (eds.). Tri-Ology 55(3): 1–17.
- Tipping, P. W., A. Sosa, E. N. Pokorny, J. Foley, D. C. Schmitz, J. S. Lane, L. Rodgers, L. Mccloud, P. Livingston-Way, M. S. Cole, and G. Nichols. 2014. Release and establishment of *Megamelus scutellaris* (Hemiptera: Delphacidae) on waterhyacinth in Florida. Florida Entomologist 97(2): 804–806.
- Williams, K. A. 2016. *Copidothrips octarticulatus*. p. 8. *In*: P. J. Anderson and G. S. Hodges (eds.). Tri-Ology 55(31): 1–17.
- Wilson, M. R., and M. F. Claridge. 1991. Handbook for the identification of leafhoppers and plant-hoppers of rice. CAB, Wallingford, UK. p. i–x, 1–142.
- Wilson, S. W. 2005. Keys to the families of Fulgoromorpha with emphasis on planthoppers of potential economic importance in the southeastern United States (Hemiptera: Auchenorrhyncha). Florida Entomologist 88(4): 464–481.
- Wilson, S. W., and L. B. O'Brien. 1987. A survey of planthopper pests of economically important plants (Homoptera: Fulgoroidea). p. 343–360. *In*: Proceedings of the second International Workshop on Leafhoppers and Planthoppers of Economic Importance, Provo, Utah, USA, 28 July–1 August 1986.
- Wooten, C. M., S. W. Wilson, and J. H. Tsai. 1993. Descriptions of nymphs of the planthopper *Harmalia anacharsis* Fennah, a species new to the United States (Homoptera: Delphacidae). Journal of the New York Entomological Society 101: 567–573.

Received May 16, 2020; accepted May 19, 2020. Review editor Joe Eger.

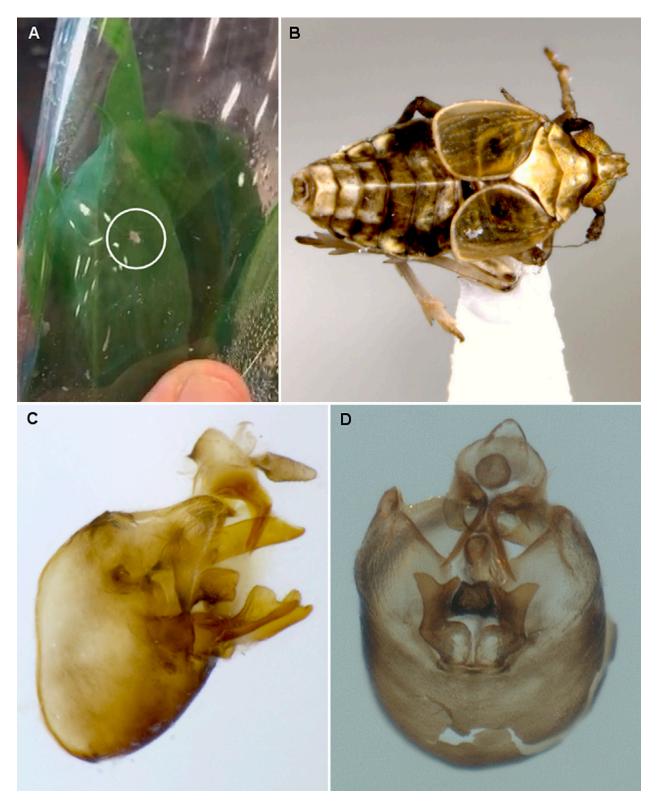


Figure 1. Opiconsiva anacharsis (Fennah). **A)** Opiconsiva anacharsis on Echinodorus sp. plant as sold in stores. Photograph by Melanie Cain, DPI. **B)** Adult female dorsal habitus. Photograph by Jade S. Allen, DPI. **C)** Male genital capsule, lateral view. Photograph by Jade S. Allen, DPI. **D)** Male genital capsule, posterior view. Photograph by Susan E. Halbert, DPI.