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1 **Investigating the critical variables of azithromycin oral absorption**
2 **using in vitro tests and PBPK Modeling**

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30 **Abstract**

31 Azithromycin is an antibiotic listed in the essential list of medicines for adults and pediatrics.
32 Conflicting evidence has been found regarding azithromycin classification according to the
33 Biopharmaceutics classification system (BCS). The purpose of this study was to identify the critical
34 variables that influence the oral absorption of azithromycin in adults and pediatrics.

35 Azithromycin solubility and dissolution studies (oral suspension) were performed in buffers and
36 biorelevant media simulating the fasted and fed gastrointestinal tract. A PBPK model was
37 developed for azithromycin for healthy adult volunteers and pediatrics (Simcyp® v18.2) informed
38 by *in vitro* solubility and dissolution studies to predict drug performance after administration of
39 azithromycin as an oral suspension.

40 The developed PBPK model predicted azithromycin plasma concentrations-time profiles after
41 administration of an oral suspension to adults and pediatrics. Sensitivity analysis of solubility vs
42 dose suggests that absorption is independent of solubility within the therapeutic dose range in both
43 adults and pediatrics. The developed PBPK model for adults and pediatrics was consistent with the
44 mechanism of permeation through the intestinal membrane (passive and active processes) being
45 the rate-limiting step of azithromycin's absorption.

46 The physiologically based approach proposed was shown to be useful to determine the factors
47 controlling drug absorption in adults and pediatrics.

48

49 **Key-words:** Pediatric; Absorption; Solubility; Permeability; Biopharmaceutics classification
50 system (BCS); Physiologically Based Pharmacokinetic (PBPK) modeling.

51

52 **Introduction**

53 The biopharmaceutics classification system (BCS) is a framework that describes the key
54 drivers of *in vivo* drug performance: dose, solubility and permeability ¹. The BCS is a vital part
55 of the drug development process, particularly in the establishment of BCS-based biowaivers ².
56 The need for a pediatric biopharmaceutical classification system (pBCS) has been emphasised
57 in the literature ^{2,3}. Although efforts have been made towards the establishment of a pBCS,
58 uncertainties in the current knowledge of pediatric physiology, heterogeneity of the pediatric
59 population, and other complicating factors still hinder the progress and development of such a
60 tool. Due to the lack of a pBCS, predictions of solubility and permeability classification in
61 pediatrics are often based on the adults' BCS criteria. Caution should be taken with this
62 approach as the pediatric population undergoes developmental changes in anatomy and
63 physiology (such as differences in gastric pH, gastrointestinal (GI) fluid composition, gastric
64 emptying (GE) times, small intestinal transit times (SITT), intestinal transporters, *etc.*), which
65 might alter the drug solubility and permeability classification criteria in the pediatric age groups
66 ^{3,4}. Consequently, drugs biopharmaceutical properties can be affected as a function of age, and
67 shifts in BCS classification between adults and pediatrics can occur. Unfavourable BCS class
68 shifts can negatively affect drug performance in the pediatric population when compared to
69 drug efficacy and safety in adults ^{3,5}. Therefore, an extensive evaluation should be performed
70 to assess the risk of BCS classification changes from extending adults' BCS criteria to
71 pediatrics ^{3,6,7}.

72 The BCS drug solubility criteria can be determined based on three main parameters: the drug's
73 highest dose the initial fasted gastric volume, and the drug solubility across a physiologically
74 relevant pH range. Drugs are classified as highly soluble if the highest therapeutic dose (EMA)
75 or highest strength (FDA) is soluble in 250 mL aqueous liquid at a relevant pH range ⁸⁻¹⁰.

76 Current methods for assessing the pediatric relevant luminal volumes are based on the
77 Bodyweight (BW) or Body Surface Area (BSA) extrapolation of adult luminal volumes ².
78 The permeability class of a drug is based on the extent of absorption. For a drug to be
79 considered highly permeable, the systemic bioavailability/extent of absorption needs to be
80 greater than 85–90% of the administered dose ^{9,10}. The preferred methods to determine the
81 permeability class of a drug are pharmacokinetic (PK) studies, such as mass balance or absolute
82 bioavailability studies. Alternative methods can be used, which do not involve human subjects,
83 such as *in vivo* or *in situ* intestinal perfusion in suitable animal models, or *in vitro* permeability
84 experiments such as cultured Caco-2 epithelial cell monolayers derived from human colon
85 adenocarcinoma cell line ^{9,10}. For pediatrics, permeability values have often been derived from
86 absolute bioavailability data in pediatric patients, however, due to the limited PK data in
87 pediatrics, alternative methods have been suggested. Calculated logP values guided the
88 provisional classification of the drugs included in the World Health Organization (WHO) list
89 of essential drugs for children ³. The applicability of this method to pediatric groups under 2
90 years of age remains unknown due to the rapid degree of maturation and growth changes that
91 take place until this age ². Recently, a mechanistic PBPK modeling approach has been proposed
92 to detect the sensitivity of the cumulative fraction absorbed to permeability changes in
93 pediatrics, as a method to evaluate the risk of BCS permeability classification changes between
94 adults and pediatrics ¹¹.

95 Pediatric biopharmaceutic risk assessment should be conducted since the early stages of drug
96 development, along with the adult product development program ⁷. A biopharmaceutical risk
97 assessment should involve the coupling of *in vitro* and *in silico* tools to predict the impact of
98 drug solubility, dissolution, and permeability on the drug systemic exposure. The best approach
99 to a successful risk assessment in pediatrics is to apply the current knowledge of pediatric
100 physiology in the development of age-appropriate *in vitro* tools (such as age-appropriate

101 biorelevant media composition and dissolution conditions) and *in silico* tools (such as age-
102 appropriate PBPK modeling focused on oral drug absorption). The coupling of age-appropriate
103 *in vitro* and *in silico* tools may be a valid strategy to transcend the existing knowledge gaps
104 surrounding each method on its own ².

105 In this study, azithromycin was selected as the model compound. Azithromycin is an antibiotic
106 commonly prescribed for respiratory tract infections in adults and pediatrics ¹². The pediatric
107 formulation available in the market is the Zithromax[®] immediate-release powder for
108 suspension which can be used in children under 45 Kg according to the drug product label ¹².

109 Conflicting evidence has been found regarding the BCS classification of azithromycin.
110 According to the FDA Zithromax[®] Clinical Pharmacology and Biopharmaceutics review,
111 azithromycin might be either a BCS class I or III compound ¹². On the other hand, WHO has
112 classified azithromycin as BCS class II (poor solubility) or IV (poor solubility and
113 permeability) compound ¹³. Additionally, three different studies have attempted to classify
114 azithromycin according to a provisional pBCS. delMoral-Sanchez *et al.* attributed a provisional
115 pBCS class for the oral drugs included in the Essential Medicines List by the World Health
116 Organization (WHO) and compared the pBCS classification with the adults BCS class ⁶. For
117 the solubility classification, the dose number was calculated, and a BW-extrapolation was used
118 to scale the pediatric luminal volumes from the reference adult volume ⁶. Permeability
119 classification was based on partition coefficient, logP (n-octanol/water partition coefficient),
120 and its relationship with human intestinal permeability. delMoral-Sanchez *et al.* considered
121 azithromycin as a BCS class II in adults, and favourable changes to BCS class I in newborns
122 were predicted based on Fried's Rule for the calculation of the pediatric dose (Fried's rule=[age
123 (months)/150] × adult dose) ⁶. Shawahna also proposed a provisional classification system
124 based on a BW-extrapolation to scale the pediatric luminal volumes from adult's initial gastric
125 volumes ¹⁴. Shawahna predicted that azithromycin was as a poorly water-soluble compound in

126 adults, with no solubility class changes predicted for pediatrics. Predictions for permeability
127 classification changes in pediatrics were not performed in this study. Finally, Gandhi and co-
128 workers used a potential pBCS, where BSA-extrapolation was used to scale pediatric reference
129 volumes from the adult reference volume⁷. Differences in azithromycin's BCS classification
130 between publications could be related to different methods used for the calculation of the dose
131 number (*i.e.* highest dosage strength vs highest single dose), but these were not always
132 reported. For assessment of permeability class changes from adults to pediatrics, the absolute
133 bioavailability in pediatrics was considered. Based on the study findings, azithromycin was
134 classified as BCS class III in adults, and no pediatric BCS class changes were expected for
135 pediatrics⁷.

136 This study aimed to use PBPK modeling informed by *in vitro* and *in silico* biopharmaceutical
137 tools to: (i). clarify the impact of *in vivo* solubility, dissolution, and permeability on the
138 absorption of azithromycin in adults and pediatrics; and (ii). assess the biopharmaceutical risk
139 of age-related changes on oral azithromycin performance. Solubility and dissolution studies
140 were performed in buffers and biorelevant media simulating the fasted and fed state
141 environment of the gastrointestinal tract. A PBPK model was developed for adults and
142 pediatrics in the Simcyp[®] v18.2 (Certara[®], US) simulator, informed by the results obtained in
143 the *in vitro* dissolution and solubility studies to predict drug performance after administration
144 of azithromycin as an oral suspension to adults and pediatrics. Parameter sensitivity analysis
145 (PSA) was performed to generate understanding of the role of critical variables on the oral
146 absorption of azithromycin in adults and pediatrics.

147

148 **Materials and methods**

149 **Materials**

150 Azithromycin analytical standard (90%), dichloromethane, acetonitrile [High-performance
151 liquid chromatography (HPLC) grade], and methanol (HPLC grade) were obtained from Fisher
152 Scientific (UK). Sodium hydroxide, 37% hydrochloric acid, sodium chloride, potassium
153 dihydrogen phosphate, sodium acetate trihydrate were purchased from Fisher Scientific (UK).
154 Water was ultra-pure (Milli-Q) laboratory grade. Ultrafree filter units were obtained from
155 Merck Millipore, USA, and azithromycin dihydrate (98%) was purchased from VWR
156 Chemicals, UK. Zithromax[®] powder for the suspension (azithromycin dihydrate 209.64 mg/5
157 ml containing the equivalent of 200 mg azithromycin base per 5 ml, Pfizer Ltd., UK) was
158 obtained from UK pharmacies and Belgium pharmacies. Glass microfiber GF/F and GF/D
159 filters (0.7 and 2.7 μm , respectively) (Whatman[®], UK), pepsin (from porcine), were purchased
160 from Sigma Sigma-Aldrich Company Ltd. (UK). Sodium taurocholate (Prodotti Chimici
161 Alimentari S.P.A., Italy), egg lecithin – Lipoid EPCS (Lipoid GmbH, Germany), and Glycerol
162 monooleate (Biosynth Carbosynth[®], UK) were obtained from the specified sources. Ultra-high-
163 temperature treated whole cow's milk standardised to less than 4% fat was acquired from
164 Sainsbury's, UK. Polytetrafluoroethylene (PTFE) filters (13 mm, 0.45 μm) were used in the
165 solubility experiments and were purchased from Fisher Scientific (UK).

166

167 **Methods**

168 **Preparation of media**

169 USP simulated gastric fluid sine pepsin (SGF_{sp}) pH 1.2, acetate buffer pH 4.5, and phosphate
170 buffer pH 6.8 were prepared following the USP¹⁵. Adult and pediatric biorelevant media were
171 freshly prepared for each experiment, as described by Maharaj *et al.*¹⁶. Adult standard
172 biorelevant media used consisted of fasted state simulated gastric fluid (FaSSGF), fasted state

173 simulated intestinal fluid (FaSSIF-V2), fed state gastric simulated fluid (FeSSGF) or fed state
174 intestinal simulated fluid (FeSSIF-V2)¹⁶. The infant biorelevant media used was infant fasted
175 state simulated gastric fluid (Pi-FaSSGF) and infant fed state simulated intestinal fluid (Pi-
176 FeSSIF)¹⁶. For the two-stage dissolution testing (performed in the inForm platform), double
177 concentrated FaSSIF-V2 was prepared with an additional amount of sodium hydroxide to
178 achieve the final composition of FaSSIF-V2 (pH 6.5) after its addition to the gastric phase.

179

180 **Solubility studies**

181 **Solubility measurements**

182 Solubility experiments were performed using the shake flask method in a shaking water bath
183 (model Grant SS40-2, Grant Instruments, UK) (37 °C, 200 strokes/minute). All solubility
184 assessments were performed in triplicate. An excess of the solid drug was used to saturate the
185 volume of medium used. Experiments were conducted with 2 mL of medium in centrifuge
186 tubes. Aqueous-based samples were collected at 24 h, filtered and appropriately diluted in
187 methanol. Filter adsorption studies were performed prior to the experiment. No adsorption to
188 the filters used was observed. For samples containing milk-based products, the undissolved
189 drug was removed by centrifugation (Eppendorf Heraeus Fresco 17 centrifuge, Thermo
190 Electron LED GmbH, Germany) at 8000 rpm for 15 min at 4 °C. The supernatant was then
191 treated with methanol (1000 µL of methanol were added to 500 µL of the sample), vortexed
192 for 1 min, centrifuged (8000 rpm, 15 min, 4°C) and the resulting supernatant was filtered
193 through a 0.45 µm PTFE filter and injected into the HPLC.

194

195 **Drug Solubility Classification**

196 Azithromycin solubility classification was determined by the calculation of dose number for
197 pediatrics and adults. The dose number (D_0) was estimated across the different pediatric age
198 groups using the following equation:

199

$$D_0 = \frac{Dose}{S \times V_0}$$

Equation 1

200

201 where V_0 is the initial gastric volume and S is the drug (azithromycin) aqueous solubility. A
202 dose number > 1 indicates a low solubility compound, and a dose number < 1 indicates the
203 opposite^{3,6,7}.

204 For pediatrics, the initial gastric volume (V_0) was calculated by a BW-extrapolation or BSA-
205 extrapolation according to **Equations 1** and **2**, respectively.

206

$$V_0 = \frac{BW \times 0.4 \text{ mL/Kg}}{37.1 \text{ mL}} \times 250 \text{ mL}$$

Equation 2

207

208 In **Equation 2**, BW stands for the age-specific pediatric bodyweight. The BW values were the
209 50th percentile values in the Centers for Disease Control and Prevention (CDC) growth charts,
210 where for each age the average of boy and girl BW value was used¹⁷. The 0.4 mL/Kg and 37.1
211 mL are estimates of fasted gastric fluid volumes in pediatrics and adults, respectively, and 250
212 mL is the reference volume used in the BCS^{2,9,10,13}.

213

$$V_0 = \frac{BSA}{1.73 \text{ m}^2} \times 250 \text{ mL}$$

Equation 3

214

215 In **Equation 3**, BSA stands for the specific pediatric body surface area. BSA was calculated
216 according to Mosteller formula, where the average of boy and girl BW and body height (BH)
217 used for each specific age, corresponded to the mean of boys and girls 50 percentile values in
218 the CDC growth charts¹⁷. The adult BSA (1.73 m²) is the reference value for a 70 Kg fasted
219 adult⁷ and 250 mL is the standard adult reference volume used in the BCS^{2,9,10,13}.

220 The doses used in the calculations were the highest azithromycin dose for each age and its BW,
221 as indicated in the drug label ¹². For pediatric patients up to 15 Kg a 10 mg/Kg was used for
222 calculations. For children above 15 Kg the dose was (i). 200 mg dose for children 15 -25 Kg;
223 (ii). 300 mg for children 26 to 35 Kg; (iii). 400 mg for adolescents 35 to 45 Kg. The dose used
224 for adult dose number calculations was 500 mg.

225

226 **Dissolution studies**

227 **USP 4 apparatus dissolution studies**

228 Dissolution studies were conducted in an Erweka[®] flow-through dissolution tester (USP, 2009)
229 (DFZ720, Erweka GmbH, Germany) equipped with Ø 22.6 mm cells that were maintained at
230 37 ± 0.5 °C and connected to an Erweka[®] Piston Pump (model HKP720). A 5 mm – size glass
231 bead was positioned in the tip of the cell, and glass beads of 1 mm-size were added to the
232 conical part of each cell. On the top of the cell Whatman[®] glass fiber filters were used (a GF/F
233 (0.7 µm) followed by a GF/D (2.7 µm)). The azithromycin oral suspension (Zithromax[®]
234 immediate-release (IR) powder for oral suspension (200 mg/5mL)) was prepared by
235 reconstitution with the appropriate amount of water, as per the drug label ¹². During the
236 experiment, the exact volume of suspension was measured with a syringe and placed on top of
237 the glass beads. Residence times in the USP 4 apparatus were appropriately selected to mimic
238 the fasted and fed state gastrointestinal volumes and transit times in each subpopulation (**Table**
239 **1**) ². Flow-rates were selected to achieve a balance between the duration of the exposure of the
240 drug product to the various simulating media and total fluid volumes, taking into account the
241 lack of radial water flux *in vitro* ¹⁸. Experiments were run in open-mode (with sequential media
242 change from gastric to intestinal medium), with fresh media continuously passing through the
243 cell containing the dosage form ¹⁹. Fluid samples were collected in volumetric cylinders, which

244 were exchanged every 15 min after the start of the experiment. The drug content in the samples
245 was assayed by HPLC. Experiments were run in triplicate.

246

247 **inForm platform dissolution studies**

248 Dissolution studies were performed in the inForm platform (Pion[®] Inc., UK) using a two-stage
249 approach. The azithromycin oral suspension was prepared as in subsection 5.2.2.3.1. The
250 clinical dose (*i.e.* infants 130 mg, children 300 mg and adults 500 mg) was down-scaled
251 according to age-appropriate volumes (*i.e.* infants 150 mL, children 200 mL and adults 500
252 mL) (**Table 1**) according to a final dissolution volume of 80 mL. A two-stage approach was
253 followed: fasted gastric condition were simulated for 30 min (40 mL), followed by intestinal
254 simulated conditions (40 mL of two-fold concentrated FaSSIF-V2 was added leading to a final
255 volume of 80 mL). Sample collection (0.5 mL) took place at 5, 15, 30, 45, 60, 75, 90, 120, 180
256 and 240 min. After collection, samples were filtered through an ultrafree filtering unit 0.45 µm
257 PTFE and centrifuged (Eppendorf mini spin plus, Germany) at 8000 rpm for 2 min at room
258 temperature. The filtrate was diluted with methanol and injected into the HPLC. Experiments
259 were performed in triplicate.

260

261 *Please insert Table 1 here*

262

263 **Chromatographic conditions for drug quantification**

264 The chromatographic method used for quantification of azithromycin was a modification of
265 the method by Baxevanis *et al.*²⁰. Quantification of azithromycin was performed using HPLC
266 coupled with ultraviolet (UV) detector [Waters HPLC system (Saint-Quentin-en-Yvelines,
267 France) and Agilent HPLC system 1100/1200 series (Agilent Technologies, USA)] using a
268 C18 column (X-Bridge C18 column 150 × 4.6 3.5 µm) at 40 °C. The injection volume was 20
269 µL and the detection was carried out by UV at 210 nm. The analytical method used an isocratic

270 mobile phase composed of a mixture of 10 mM potassium phosphate buffer (pH 7.5) and
271 acetonitrile (45:55 v/v) at a flow rate of 1 mL/min. Quantification of montelukast in samples
272 was performed with calibration curves of freshly prepared standard solutions (calibration curve
273 range: 3–100 µg/mL). Standards were prepared in the medium of interest for each experiment
274 by appropriate dilution of a 1 mg/mL stock solution of azithromycin analytical standard in
275 methanol. The limit of detection (LOD) and the limit of quantification (LOQ) were 0.90 and
276 2.5 µg/mL, respectively.

277

278 **PBPK model development**

279 **Pharmacokinetic Data Collection and Data treatment**

280 PK studies of azithromycin, administered intravenously or orally as an oral suspension, in
281 adults and pediatrics, were obtained from the literature. The plasma concentration-time
282 profiles, and trial design information were extracted from the published reports and used for
283 the PBPK model building and validation (**Table S1** and **Table S2**). One study reported single
284 dose administrations of IV infusion of azithromycin (1000 mg to 4000 mg doses) in adults ²¹.
285 Three studies reported single dose administration of azithromycin oral suspension (500 mg) to
286 adults in the fasted state ²²⁻²⁴ and one in the fed state ²². One pediatric study reported IV infusion
287 administration (10 mg/Kg) in infants (0.5 to 2 years), children (2 to 6 years), old children (6 to
288 12 years) and adolescents (12 to 16) ²⁵. Finally, two PK studies reported oral administration of
289 azithromycin suspension in a 5-day oral regimen (10 mg/Kg on the 1st day and 5 mg/Kg day 2
290 to 5) to pediatric patients: infants and young children (0.5 to 5 years) and old children and
291 adolescents (6 to 15 years) ^{26,27}. The observed PK profiles that were found in the literature were
292 digitalised with WebPlotDigitalizer[®] v4.1 software ²⁸. PK data analysis was performed with
293 PKSolver[®] add-in program for Microsoft Excel[®] ²⁹. Non-compartmental (NCA) analysis was

294 used to calculate the observed PK parameters, which are reported in the supplementary
295 materials (**Table S1** and **Table S2**).

296

297 **Adult PBPK model**

298 PBPK modeling and simulations were performed using the Simcyp® Simulator (V18.2;
299 Certara, UK). The PBPK modeling strategy followed the workflow presented in **Figure 1**. The
300 relevant input parameters for the development of the PBPK models and simulations performed
301 are summarised in **Table 2**. Azithromycin-specific information obtained from the literature
302 consisted of its physicochemical properties including molecular weight (MW), octanol: water
303 partition coefficient ($\log P_{o:w}$), fraction unbound in plasma (f_u) and blood to plasma ratio (B:P)
304 ³⁰⁻³².

305 The distribution model was described using a minimal PBPK model with a Single Adjusting
306 Compartment (SAC), a non-physiological compartment that represents a cluster of tissues
307 (excluding liver and portal vein). The parameters that defined the distribution were: first-order
308 rate constants [h^{-1}] of the drug into (k_{in}) and out (k_{out}) of the SAC, V_{SAC} (apparent volume
309 associated with the SAC) and V_{ss} (steady-state volume of distribution). The disposition
310 parameters were initially calculated based on a 2 compartmental fit to plasma concentration-
311 time profiles obtained after an IV infusion administration of 1000 mg of azithromycin ²¹. These
312 initial distribution parameters were optimised to accurately describe the IV infusion
313 administration of 1000 mg clinical data, and the simulations were externally validated for the
314 IV infusion administration of 2000 and 4000 mg doses ²¹. The intravenous clearance of
315 azithromycin ranges between 31 - 46.5 L/h in healthy adults ^{12,32-34}. Azithromycin is mainly
316 eliminated unchanged in the faeces via biliary excretion (more than 50%) with the rest being
317 attributed to renal clearance (less than 20% eliminated unchanged in urine) ^{21,32,33}.
318 Azithromycin PK profiles do not show evidence of enterohepatic recirculation, therefore this
319 process was not investigated (assumed 0% available for re-absorption in the elimination tab).

320 For the mechanistic absorption modeling, the advanced dissolution absorption and metabolism
321 (ADAM™) model was used³⁵. The ADAM™ model was used to investigate the impact of
322 passive and active permeability (*i.e.* Caco-2 vs MechPeff) and the impact of formulation input
323 (*i.e.* biorelevant solubility inputted into SimCYP® Diffusion Layer Model (DLM) vs direct
324 input of biorelevant *in vitro* dissolution) on azithromycin performance.

325 Azithromycin has been shown to be a P-gP substrate³⁶, efflux transport in the intestine was
326 implemented in the PBPK model in two ways. In option 1: the effect of active transport in
327 SimCYP® was assessed directly by using data from *in vitro* cell systems. By performing a
328 comparison of Caco-2 data that is treated/non-treated with P-gP inhibitors it is possible to
329 assess the limiting effects of P-gP on the passage of drug across the monolayer. Caco-2 data
330 treated with an inhibitor of P-gP were entered as 'Passive' and without inhibitor of P-gP were
331 inputted as 'Passive & Active'. In option 2: the influence of transporter was also investigated
332 in SimCYP® by activating the MechPeff model. Since information of regional absorption to
333 further validate the predicted regional permeability were not available, the predicted $P_{\text{eff,man}}$
334 was applied to all segments of the gastrointestinal tract to achieve a better description of the
335 PK profiles. The intestinal P-gP *in vitro* transporter intrinsic clearance $CL_{\text{int,T}}$ ($\mu\text{L}/\text{min}$) was
336 fitted to observed plasma concentration-time profiles after administration of azithromycin oral
337 suspension in the fasted and fed state in adults published by Foulds *et al*²².

338 Two levels of different activity of P-gP were assumed for fasted and fed state to simulate the
339 inhibition of P-gP substrate by food (**Table 2**). The interaction of drug transporters with food
340 has been discussed in the literature by several authors³⁷⁻³⁹, where a high-fat meal is suspected
341 to be able to inhibit P-gP³⁸.

342 The impact of azithromycin *in vitro* dissolution and solubility on drug absorption was evaluated
343 with the PBPK model following two approaches: (i). application of the diffusion layer model
344 (DLM), using the measured biorelevant solubility data as input; and (ii). direct entry of the *in*

345 *vitro* dissolution data [Dissolution Profile Model (DPM)] as discrete dissolution profiles. For
346 the fasted state DPM model, the impact of the dissolution on the performance of azithromycin
347 was evaluated by comparison of the observed and simulated plasma concentration-time profiles
348 when inputting the dissolution profiles from the USP 4 apparatus *vs* the inForm ones.
349 Precipitation was not explored, as the dissolution studies indicated no precipitation over 4 h.
350 In terms of physiology, all adult simulations were run using the healthy volunteer population
351 library of the Simcyp[®] simulator. Mean gastric residence time (MGRT) in the fasted state was
352 adjusted to 0.50 h to better describe the adult fasted *in vivo* data (adjusted against the Foulds *et*
353 *al.* 1996 dataset and validated with the remaining fasted state adult datasets)²²⁻²⁴. For the
354 remaining physiological parameters, default software values were assumed.

355

356 *Please insert Figure 1 here*

357

358 *Please insert Table 2 here*

359

360 **Pediatric PBPK model**

361 The Simcyp[®] pediatric population library was used. The population file gathers information on
362 pediatric demography (age, body height, bodyweight and body surface area) and
363 developmental physiology (liver size, renal function, liver blood flow *etc.*)⁴⁰. In pediatrics, the
364 adult SAC parameters, k_{in} (1/h), k_{out} (1/h), are allometrically scaled by the software according
365 to the pediatric BW and an adult BW of 70 Kg, with exponents of -0.25⁴¹. Since biliary
366 elimination appears to be reasonably well developed reaching adult levels at birth or in the first
367 few months of postnatal age no scaling was applied to biliary clearance³². Scaling of the renal
368 function was captured in the Simcyp[®] pediatric module as previously described⁴². For both the
369 fasted and fed state, the pediatric ADAM[™] model contains information of developmental

370 changes as a function of age on: size of the gastrointestinal tract, gastric emptying time,
371 gastrointestinal volumes, pH, *etc.* ^{40,43}. The fasted MGRT in pediatrics was assumed to be the
372 same as adults (*i.e.* 0.5h). Remaining physiological and anatomical values were maintained at
373 default values.

374

375 **Trial design information**

376 All simulations were performed with 10 trials of 10 subjects in each trial. The trial design was
377 performed using the ‘Virtual population’ option in Simcyp[®]. The maximum and minimum age,
378 as well as the proportion of females was adjusted according to the population of the PK study
379 used for the validation of the model. Study details are presented in the supplementary materials
380 (**Table S1** and **Table S2**)

381

382 **PBPK model validation**

383 The PBPK model was validated by comparison of the simulated azithromycin plasma
384 concentration-time profiles, and the relevant PK parameters, against the clinically observed
385 data. The mean predicted plasma concentration-time profiles were assessed by the average fold
386 error (AFE) and validated with the absolute average fold error (AAFE) (**Equations 5.4** and
387 **5.5**, respectively) ⁴⁴.

388

$$AFE = 10^{\frac{1}{n} \sum \log\left(\frac{predicted_i}{observed_i}\right)} \quad \text{Equation 5.4}$$

$$AAFE = 10^{\frac{1}{n} \sum \left| \log\left(\frac{predicted_i}{observed_i}\right) \right|} \quad \text{Equation 5.5}$$

389 where n denotes the number of observed sampling points, predicted_i and observed_i denote the
390 predicted and observed plasma concentration at the sampling time point i, respectively.

391 AFE values indicates whether the simulated data underpredicts ($AFE < 1$) or overpredicts (AFE
392 > 1) the observed plasma concentrations, while an AAFE value close to unity represents the
393 precision of the simulations. An $AAFE \leq 2$ indicates an acceptable prediction⁴⁵. The relative
394 accuracy of the mean predicted PK parameters describing drug exposure [area under the plasma
395 concentration-time curve (AUC), the maximum concentration (C_{max}), and time to reach the
396 maximum concentration (T_{max})] was assessed against the mean observed PK parameters using
397 the fold error (FE) (**Equation 5.6**). A FE within a 2-fold range (FE values between 0.5 and 2)
398 indicates an acceptable prediction.

$$FE = \frac{\textit{predicted}}{\textit{observed}}$$

Equation 5.6

400

401 **Parameter sensitivity analysis**

402 Parameter sensitivity analysis (PSA) was conducted to identify the limiting steps of absorption
403 and their impact on the *in vivo* drug performance (*e.g.* C_{max} , AUC). Sensitivity analysis was
404 performed for two types of parameters. The first type was related to drug properties and
405 included parameters such as drug solubility in the duodenum (range: 1 to 27 mg/mL) *vs* dose
406 (range for adults: 250 to 600 mg; infants and young children: 40 to 220 mg; children and
407 adolescents: 120 to 500 mg), effective permeability ($P_{eff,man}$ range: 0.1 to 3×10^{-4} cm/s) and
408 intrinsic clearance of intestinal P-gP transporter (P-gP $Cl_{int,T}$ range: 10 to 300 μ L/min). The
409 dose ranges were based on the maximum and minimum observed doses in PK studies in adults
410 and pediatrics. For the permeability, the range was based on the calculated P_{eff} from the Caco-
411 2 method and calculated P_{eff} with the MechPeff model (described in Adult PBPK section).
412 Finally, the investigated range of $Cl_{int,T}$ was based on the values used in the fasted and fed state
413 models (**Table 2**). The second type of parameters was related to physiology, with MGRT being
414 selected due to potential age-related differences in food types and feeding cycles between

415 adults and pediatrics. As the basis for the PSA, the simulations after oral administration of
416 azithromycin in the fasted state for adults and pediatrics were run with MechPeff as
417 permeability input and the DLM-based ADAM™ model. For the interpretation of the PSA
418 results, predicted PK parameters were compared to the values used in the developed PBPK
419 model (*i.e.* baseline simulation).
420

421 **Results**

422 **Solubility studies**

423 **Solubility measurements**

424 The mean azithromycin solubility in buffers and biorelevant media is presented in **Figure 2**.
425 Azithromycin solubility ranged from 2 mg/mL to 13 mg/mL in all media tested. Higher
426 solubility was observed in acidic media (pH 1.2 - 1.6) in comparison to more basic media (pH
427 6.5 - 6.8) as expected based on ionization properties of azithromycin (weak base, pKa = 8.6).
428 Comparison between azithromycin solubility in buffers and in biorelevant media showed that
429 drug solubility was sensitive to bile salts concentrations in the media. The azithromycin
430 solubility in the SGF_{sp} pH 1.2 was approximately 9 mg/mL and in the fasted gastric fluid
431 (FaSSGF) its solubility increases (slightly) by 1.4 fold. The highest solubility of azithromycin
432 was obtained in FaSSGF biorelevant media at pH 1.6. The solubility of azithromycin in infant
433 fasted gastric simulating fluid (Pi-FaSSGF) was lower than the one in the respective adult
434 FaSSGF. In intestinal simulated fluids, azithromycin presented the lowest solubility in the
435 fasted intestinal simulating fluid (FaSSIF-V2) and the highest solubility in the fed intestinal
436 simulating fluid (FeSSIF-V2), suggesting that buffer capacity and ionic strength affects its
437 solubility. Drug solubility was similar in the adult and the infant fed intestinal simulating fluid
438 (FeSSIF-V2 and Pi-FeSSIF, respectively).

439

440 *Please insert Figure 2 here*

441

442 **Drug Solubility Classification**

443 The dose numbers calculated for adults and pediatrics are presented in **Figure 3**. Azithromycin
444 displays dose number lower than 1 for adults based on the drug solubility measured in USP
445 buffers. For pediatrics, dose number was lower than 1 when calculated with USP buffers

446 solubility, whether pediatric V_0 was calculated through BW or BSA-based extrapolation, which
447 indicates that azithromycin is classified as a highly soluble compound in all age groups.
448 For pediatrics, dose numbers calculated based on azithromycin solubility in the fasted state
449 simulated intestinal fluid (FaSSIF-V2) and the BW-extrapolation method for the calculation of
450 pediatric V_0 show that azithromycin displays a dose number higher than 1 in the pediatric
451 population but not in adults. When the BSA-extrapolation method is used of extrapolation of
452 initial gastric volume, the calculated dose number was below or equal to one. The results
453 demonstrate that the BW-extrapolation leads to a more strict solubility classification, due to
454 the estimation of higher V_0 , as previously reported by delMoral Sanchez *et al.* ⁴⁶. The
455 differences between dose numbers calculated with BW- and BSA-based extrapolation reinforce
456 the need for a standardised pediatric Biopharmaceutics Classification System (pBCS).

457

458 *Please insert Figure 3 here*

459

460 **Dissolution studies**

461 **USP 4 apparatus studies**

462 *In vitro* dissolution studies of oral suspension in the USP 4 apparatus in the fasted and fed state
463 are presented in **Figure 4**.

464 In the fasted state the dissolution from azithromycin oral suspension was complete in all set-
465 ups. Dissolution was very rapid dissolution (> 85% within 15 min) in the adults' set-up. In the
466 children and infants dissolution set-ups, dissolution was rapid (> 85% within 30 min). All set-
467 ups tested more than 85% of azithromycin dissolved in the simulated gastric fluids, which is in
468 agreement with the azithromycin solubility in acidic conditions (**Figure 4**). Due to high
469 similarity of dissolution from azithromycin oral suspension between all age groups in the fasted
470 state, in the fed state, dissolution was only tested in the adults and infants set-ups. Under fed
471 state conditions, dissolution of azithromycin oral suspension with the adults set-up under the

472 fed state was very rapid (>85% within 5 min). A slightly slower azithromycin dissolution from
473 the oral suspension was observed in the infants set-up under the fed state conditions (>85%
474 within 45 min) compared to the one under the fasted state conditions. The (small) differences
475 observed in the azithromycin dissolution rate under the fasted and fed state between the
476 different age groups are attributed to the different *in vitro* hydrodynamics in the USP 4
477 apparatus (*i.e.* differences in flow-rates which reflect age-related changes in GI volumes and
478 GI fluids composition.

479

480 *Please insert Figure 4 here*

481

482 **inForm platform dissolution studies**

483 *In vitro* fasted state dissolution studies of Zithromax[®] oral suspension performed in the inForm
484 platform are presented in **Figure 4**. For all dissolution set-ups tested in the inForm platform,
485 the dissolution rate was slower in the gastric phase and increased once the intestinal medium
486 was added. Within one hour, approximately 80% azithromycin was dissolved in all the set-ups
487 tested. The dissolution rate appears to be slightly faster initially in the first 30 min in the
488 children set-up (when compared to the adult and infant dissolution profiles) and at the end of
489 the 4h a higher dissolution extent was observed in the adult dissolution set-up. A comparison
490 with the results of the USP 4 apparatus shows that the dissolution rate was slower in all age
491 groups set-ups when tested in the inForm platform. Dissolution profiles obtained in the inForm
492 platform present a high variability (coefficient of variation (CV) ranged 8 to 43 %). The inForm
493 platform can only run one replicate at a time, each replicate had to be performed on different
494 days, which appears to have contributed to the high CV % of the results. As azithromycin is a
495 weak base, the dissolution studies with this set-up were performed to investigate precipitation
496 *in vitro*. As expected from the results obtained in the solubility studies, azithromycin
497 precipitation was not observed during the dissolution experiments.

498

499 **Adult PBPK model**

500 **Intravenous Administration**

501 The results of the simulations of the IV administration of azithromycin in adults are presented
502 in the supplementary materials (**Figure S1**). The simulations after azithromycin IV
503 administration were verified against the observed PK studies after IV administration of 1000,
504 2000, and 4000 mg doses. The predicted plasma concentration-time profiles were in good
505 agreement with the observed plasma concentration-time profiles measured after the IV infusion
506 administration of a wide range of azithromycin doses with all AAFE values below 1.50 (**Table**
507 **S3**).

508

509 **Oral Administration**

510 Simulated plasma concentration-time profiles after oral administration of azithromycin
511 suspension to adults in the fasted and fed state are presented in **Figure 5**. The FE, AFE and
512 AAFE for all simulations are presented in the supplementary materials (**Table S4**).
513 Independently of the permeability or formulation input, the simulations of the administration
514 of azithromycin in the fasted state were able to meet the validation criteria with all AAFE
515 values ≤ 1.60 .

516 For the fasted state simulations, the simulation performance was very similar independently of
517 the permeability inputs tested (AAFE for Caco-2 and MechPeff + P-gP input ranged from 1.14
518 and 1.52). The simulations with Caco-2 data as input predicted better the mean observed T_{max}
519 (FE = 1.09) in the fasted state in comparison to the simulations with MechPeff + P-gP $Cl_{int,T}$ as
520 permeability input (FE = 0.72). The simulations with MechPeff + P-gP $Cl_{int,T}$ as permeability
521 input predicted a higher variability, reflected by larger 5th and 95th percentiles of the simulations
522 when compared to simulations from the simulations with Caco-2 data. As individual PK
523 profiles were not available in the literature, it is not possible to comment on which model is

524 more representative of the *in vivo* variability; the models (**Figure 5**) appear to overestimate the
525 *in vivo* variability (predicted C_{max} CV% ranged from 100-125% in comparison to an observed
526 C_{max} CV% from the Najib *et al* of approximately 38%; predicted AUC CV% was
527 approximately 40% in comparison to the observed AUC CV% from Najib *et al* of
528 approximately 32%)²⁴. When testing the model formulation input for the fasted state (the type
529 of dissolution or solubility data), the simulations performed very similarly on predicting the
530 observed data (AAFE ranged from 1.31 to 1.60). No differences were observed in the predicted
531 T_{max} (FE = 0.72) when comparing the simulation inputs of solubility (DLM-based PBPK
532 model) and dissolution (direct entry of dissolution profiles with USP 4 apparatus and inForm
533 platform).

534 Two different levels of P-gP Cl_{int,T} were set between fasted and fed state, in order to simulate
535 the inhibition of P-gP substrate by food (**Table 2**). For the fed state simulations with Caco-2
536 data, the *in vitro* Caco-2 data with inhibited P-gP was able to predict the observed data better
537 than if the Caco-2 data with non-inhibited P-gP (data not shown). In the PBPK model
538 developed with the MechPeff + P-gP Cl_{int,T} data as permeability input, by fitting a new intrinsic
539 clearance of P-gP (Cl_{int,T}) in the fed state, compared to the Cl_{int,T} used in the fasted state, we
540 assume a lower P-gP Cl_{int,T} in the fed state (due to inhibition by food). Simulation results of
541 oral administration of azithromycin suspension in the fed state show that both approaches for
542 adding the permeability input in the PBPK model predicted well the observed data; all the
543 predictions in the fed state met the validation criteria (AAFE values: 1.32 – 1.33).

544

545 *Please insert Figure 5 here*

546

547 **Pediatric PBPK model**

548 **Intravenous Administration**

549 Simulation results of IV infusion administration of azithromycin to pediatrics are presented in
550 the supplementary materials (**Figure S2**). Simulations were verified against observed PK
551 studies after IV infusion administration of azithromycin at a dose of 10 mg/Kg to infants (0.5
552 to 2 years), young children (2 to 5 years), old children (5 to 12 years) and adolescents (12 to
553 15 years). A good agreement between the simulated (obtained from the PBPK model) and the
554 observed plasma concentration-time profiles was observed with AAFE values ≤ 1.37 (**Table**
555 **S5**).

556

557 **Oral Administration**

558 Simulated and observed plasma concentration-time profiles in pediatrics after oral
559 administration of azithromycin suspension in the fasted state are presented in **Figure 6**. The
560 AAFE and AFE for all the simulations are presented in the supplementary materials [**Table S6**
561 (infants and young children) and **Table S7** (old children and adolescents)]. AAFE values for
562 all simulations in the investigated pediatric age groups are less than 1.49, indicating successful
563 predictions. The levels of expression of intestinal P-gP from birth to adulthood have been
564 reported to be similar to those in adults^{43,47}. Therefore, the permeability input of Caco-2 data
565 or MechPeff + P-gP $Cl_{int,T}$ data in the pediatric PBPK model were maintained at the same
566 values as in the adult PBPK models (**Table 2**). Both simulations with Caco-2 and MechPeff +
567 P-gP $Cl_{int,T}$ as permeability inputs successfully predicted the plasma concentration-time
568 profiles in both pediatric age groups (AAFE values ≤ 1.49). Similarly to what was observed in
569 the PBPK models for adults (section 3.3.2.), the best prediction of T_{max} was obtained when
570 using the Caco-2 data as permeability input in the model. For all simulation of oral
571 administration of azithromycin suspension, underprediction of the observed C_{max} in old

572 children and adolescents data was observed (FE ranged from 0.51 to 0.83). Overall, the
573 simulations with the MechPeff + P-gP $Cl_{int,T}$ as permeability input and the solubility input
574 (PBPK DLM-based model) provide the best prediction of the observed C_{max} in old children and
575 adolescents (FE = 0.83).

576

577 *Please insert Figure 6 here*

578

579 Since pediatric patients were fasted overnight before receiving the final dose on day 5, therefore
580 the observed PK data (obtained from the 5th day of the multiple-dose regimen) is representative
581 of the fasted state ^{26,27}. After validation of the fasted state PBPK models (comparison of
582 observed vs simulated PK data), prediction of the fed state was performed with the MechPeff
583 + P-gP $Cl_{int,T}$ based input and the solubility input (DLM-based PBPK model). The fed state
584 simulated results are presented in **Table 3**. The food effect was predicted based on the ratio of
585 the PK parameters (AUC_{0-inf} and C_{max}) of the fed state simulated plasma concentration-time
586 profile and the PK parameters (AUC_{0-inf} and C_{max}) of the observed fasted state data. Fed state
587 simulations in pediatrics show that the C_{max} and AUC_{0-inf} are within a 2-fold range when
588 compared to the fasted state observed ones, which indicates that azithromycin food-effect in
589 pediatrics is also not clinically significant, as it was observed in adults; a slight increase in
590 predicted C_{max} was observed but AUC_{0-inf} remained unchanged for both age groups.

591

592 *Please insert Table 3 here*

593

594 **Parameter sensitivity analysis**

595 The results of the PSA performed are presented in **Figure 7**. The investigated changes in gastric
596 residence time in the range of 0.10 to 2 h did not show a substantial impact on the predicted

597 AUC in all the investigated populations. An increase in MGRT from 0.50 to 2 h led to a
598 prolonged T_{max} of approximately 3 h and a decreased C_{max} by approximately 20% when
599 compared to simulation results with baseline values.

600 The results of the sensitivity analysis of the passive permeability ($P_{eff,man}$) and active transport
601 (P-gP $Cl_{int,T}$) are presented in **Figure 7a,b**. In adults, the reduction of $P_{eff,man}$ from 1.33×10^{-4}
602 cm/s (baseline value) to 0.10×10^{-4} cm/s resulted in a decrease of C_{max} by approximately 90%
603 and of AUC by 76%, when compared to the simulation results using the baseline values. An
604 increase in $P_{eff,man}$ from the baseline value (1.33×10^{-4} cm/s) to 3.00×10^{-4} cm/s led to an
605 increase in C_{max} by 79% and a 12% increase in AUC. A reduction in P-gP $Cl_{int,T}$ from 200
606 $\mu\text{L}/\text{min}$ (baseline value for the fasted state) to 10 $\mu\text{L}/\text{min}$ resulted in an increase of predicted
607 C_{max} by approximately 60% in adults. Similar trends are observed in both of the pediatric
608 groups investigated for $P_{eff,man}$ and P-gP $Cl_{int,T}$. Overall, the PSA results show that the AUC
609 and the C_{max} are influenced the most by permeation through the intestinal membrane, and both
610 passive and active transport control the absorption of azithromycin in both adults and
611 pediatrics.

612 The impact of solubility in intestinal fluids on azithromycin PK was further investigated
613 through PSA (**Figure 8** and **Figure S3**). Similar results were observed for PSA of solubility in
614 the gastric fluids and its impact of PK (data not shown). Based on the PSA results, it can be
615 concluded that azithromycin's solubility in the gastrointestinal fluids is not the rate-limiting
616 step for its absorption for the investigated therapeutic dose range for adults and pediatrics. This
617 finding sheds light on the BCS classification of azithromycin in terms of solubility, suggesting
618 that it is indeed a highly-soluble compound.

619

620 *Please insert Figure 7 here*

621

622 *Please insert Figure 8 here*

623

624 **Discussion**

625 PBPK absorption models have the potential to perform exploratory analysis to clarify the
626 critical parameters of drug absorption in the different pediatric age groups¹¹. In this study, the
627 impact of critical variables on azithromycin's oral absorption were evaluated by coupling *in*
628 *vitro* (age-biorelevant *in vitro* solubility and dissolution) and *in silico* methods. A general
629 schematic outlining the strategy followed in this manuscript for the biopharmaceutic risk
630 assessment of azithromycin critical parameters for absorption is presented in **Figure 9**.

631 Based on the solubility measurements a change in the solubility classification from adults to
632 pediatrics was seen when the dose number was calculated with biorelevant solubility in
633 FaSSIF-V2 and initial pediatric gastric volume calculated with the BW-extrapolation approach.
634 BW-based extrapolation and BSA-based extrapolation of initial gastric volumes can lead to
635 differences in the solubility classification of a drug. This demonstrates that standardization is
636 needed in the future on the best strategy to assess age-related changes of the solubility
637 classification of drugs. Although useful in theory, the establishment of a pBCS is not
638 straightforward since it is unlikely that the necessary scrutiny between the pediatric groups is
639 achieved while still maintaining the desirable simplistic character as observed for the adult
640 BCS. The need to take into account multiple doses for more than one age group can lead to
641 multiple classifications across the pediatric age range. The necessary differentiation of age
642 groups cannot be achieved without a certain degree of complexity, in which case, a strategy
643 coupling mechanistic PBPK absorption modeling with *in vitro* data can become a more
644 powerful tool. Therefore, to further clarify/assess the risk of age-related changes in the
645 solubility classification of azithromycin, an adult and pediatric PBPK model was built.

646 In adults, an increase of short duration (persists for less than 4 h) in C_{\max} (1.6-fold) in
647 comparison to the observed fasted state C_{\max} is observed²². In this study, we hypothesised that

648 this increase in C_{\max} after oral administration of azithromycin in the fed state could be attributed
649 to the inhibition of P-gP (an efflux transporter). As a P-gP substrate, azithromycin permeates
650 through the gut wall and is partially pumped back into the gastrointestinal lumen. If P-gP is
651 inhibited, an increase in the net permeability is seen and therefore an increase in the extent of
652 absorption would be observed ⁴⁸. The interaction of transporters with food has been previously
653 discussed in the literature ³⁷⁻³⁹, and a high-fat meal is suspected to inhibit P-gP ³⁸. Evidence to
654 support this scientific statement lays in several studies which showed that food components
655 present in gastrointestinal fluids as a result of the ingestion of high-fat meals affected the *in*
656 *vitro* transport of P-gP substrates ⁴⁹⁻⁵¹. Konishi and co-workers have shown that a variety of
657 monoglycerides and fatty acids were capable of inhibiting P-gP mediated efflux of rhodamine-
658 123 and daunomycin in Caco-2 cells ^{49,50}. The prediction of food effects that are caused by the
659 interaction of food components with intestinal transporters (influx or efflux) is complex due to
660 the inability to accurately measure the inhibitory effect caused by the food component ³⁷.

661

662 ***Please insert Figure 9 here***

663

664 Simulation results of oral administration of azithromycin suspension in the fed state showed
665 that both investigated permeability inputs [Caco-2 data (with P-gP inhibitor), or MechPeff +
666 P-gP $Cl_{\text{int,T}}$ permeability] in the PBPK model resulted in a good prediction of the small increase
667 in the *in vivo* observed C_{\max} in adults. When performing pediatric simulations, the ‘assumption’
668 of food-inhibited P-gP was also applied to the simulated pediatric age groups. The translation
669 of this mechanism to the pediatric simulations predicted a small increase in C_{\max} (below 2-fold
670 ratio when compared to the fasted state-observed C_{\max}) and AUC remained unchanged.
671 However, fed state data was not available in pediatric age groups to further validate these
672 predictions.

673 Our hypothesis that the increase in C_{max} after oral administration of azithromycin in the fed
674 state could be attributed to the inhibition of P-gP (an efflux transporter), was investigated by a
675 simple approach and the P-gp $CL_{in,T}$ was fitted in the model. It was beyond the scope of this
676 study to fully validate this hypothesis using DDI and/or polymorphism data. The contribution
677 of P-gP in the absorption of azithromycin should be further investigated in the future; since
678 azithromycin is a weak base with high lipophilicity ($\log P = 4^{30}$), some of its absorption features
679 could potentially be explained by other mechanisms such as lysosomal trapping.

680 After the PBPK model was built and validated in the populations of interest, PSA were
681 performed to identify the critical parameters influencing absorption of azithromycin.
682 Sensitivity analysis showed that azithromycin's solubility in the gastrointestinal fluids was not
683 the rate-limiting step for its absorption for adults and pediatrics in the investigated therapeutic
684 dose range. This finding sheds light on the BCS classification of azithromycin, which suggests
685 that azithromycin is indeed a highly-soluble compound in adults, and therefore belongs to
686 either BCS class I or III. Since sensitivity analysis in pediatrics also showed that azithromycin's
687 solubility in the gastrointestinal fluids was not the rate-limiting step for its absorption, there is
688 a low risk of changes in solubility class when extrapolating the solubility class from adults to
689 pediatrics. The results of the PSA showed that the C_{max} and the AUC were influenced by
690 permeation through the intestinal membrane, and both passive and active transport control the
691 absorption of azithromycin in adults and pediatrics. These results suggest that azithromycin
692 belongs to BCS class III in adults in the investigated dose range ⁵².

693 Consensus regarding the maturation of permeability in younger pediatric subgroups (especially
694 newborns) is still lacking ^{2,4,43}. As a result, it is still unclear at which age adult permeability
695 values are reached ^{43,47}. Permeability changes as a function of age could occur as a consequence
696 of the morphological development of villi and microvilli in the small intestine, which results
697 in a reduced surface area for absorption in younger pediatric patients ^{2,4,43}. Literature suggests

698 that passive permeability is developed by the age of 4 months, with general agreement of full
699 maturity by the age of 2 years old ^{2,4,43}. Although some contradictory information is available
700 on P-gP after birth, it is generally accepted that the levels of expression of intestinal P-gP from
701 birth to adulthood are likely to be similar to adults ^{2,4,43}. In the developed PBPK model,
702 azithromycin $P_{\text{eff,man}}$ or P-gP $Cl_{\text{int,T}}$ values were not changed when the adult model was
703 translated to the pediatrics. A similar degree of sensitivity to permeability-related parameters
704 was observed for the investigated pediatric age groups and adults. These results indicate that if
705 the adult permeability classification criterion is still applicable then no changes in permeability
706 classification criteria are expected between adults and pediatrics. Since a pBCS is not currently
707 available, a pediatrics biopharmaceutics classification of azithromycin is not feasible.
708 However, results of the PBPK model and PSA (of the impact of solubility and permeation
709 parameters on the PK of azithromycin) show that there is a low biopharmaceutics risk
710 associated with the extrapolation of the adult BCS class III of azithromycin to pediatrics in the
711 investigated dose range. Future studies could explore higher therapeutic doses in adults and
712 pediatrics, since adult doses for certain complications can be as high as 2 g and for pediatrics
713 doses higher than 10 mg/Kg are described in the British National Formulary for Children
714 (BNF-C). It should be noted that no clinical data was found for the administration of the higher
715 doses of azithromycin suspension in adults and/or pediatrics ⁵³.
716 Regulatory authorities have recognised the potential of PBPK modeling for predictions in
717 pediatrics ^{47,54,55}. PBPK modeling coupled with *in vitro* data (solubility/dissolution studies) has
718 a huge potential to support the development of pediatric medicines, therefore, continuous
719 improvement of these pediatric biopharmaceutic tools with high-quality physiological and
720 clinical data is essential.
721

722 **Conclusions**

723 In this study, PBPK modeling and *in vitro* tools (solubility and dissolution studies) were used
724 for identifying the critical variables affecting the oral absorption of azithromycin in adults and
725 pediatrics.

726 Differences were observed in the solubility classification of azithromycin when using buffers
727 and biorelevant media and according to two extrapolation methods of pediatric initial gastric
728 volume, which reinforces the need for a standardised strategy that can be used during pediatric
729 drug development to understand age-related changes in oral drug absorption. A mechanistic
730 investigation of the oral absorption of azithromycin with PBPK modeling was able to clarify
731 that azithromycin solubility is not a limiting step for adults and the investigated pediatric age
732 groups, for the therapeutic dose range used in each group. The PBPK modeling approach
733 revealed that permeation through the gut wall is the key driver of azithromycin oral absorption
734 in both adults and pediatrics. The absorption process is similar in the pediatric age groups
735 investigated in this study compared to adults as passive and transport mediated processes are
736 likely to be similar in these populations.

737 Overall, the PBPK modeling approach followed can be used to increase the understanding of
738 the critical parameters of absorption in pediatrics, especially when coupled with age-relevant
739 *in vitro* methods.

740

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911 10.1016/j.xphs.2018.10.033.

912 **List of Tables**913 **Table 1** Dissolution conditions used in the experiments performed in the USP 4 apparatus and the inForm platform.

USP 4 apparatus – fasted state									
Age groups	Clinical Dose (mg)	Gastric residence time (min)	Gastric flow-rate (ml/min)	Volume (mL)	Biorelevant media gastric compartment	Intestinal residence time (min)	Intestinal flow-rate (ml/min)	Volume (mL)	Biorelevant media intestinal compartment
Infants	120	0-30	4	120	Pi-FaSSGF	30-270	3	720	FaSSIF-V2
Children	120	0-30	8	240	FaSSGF	30-270	4	960	FaSSIF-V2
Adults	120	0-30	12	360	FaSSGF	30-270	4	960	FaSSIF-V2
USP 4 apparatus – fed state									
Age groups	Clinical Dose (mg)	Gastric residence time (min)	Gastric flow-rate (ml/min)	Volume (mL)	Biorelevant media gastric compartment	Intestinal residence time(min)	Intestinal flow-rate (ml/min)	Volume (mL)	Biorelevant media intestinal compartment
Infants	120	0-120	3	360	FeSSGF	120-360-	4	960	Pi-FeSSIF
Adults	120	0-90	8	720	FeSSGF	90-330	6	1440	FeSSIF-V2
inForm platform– fasted state									
Age groups	Clinical Dose (mg)	Down-scaled Dose (mg)	Gastric residence time (min)	Volume (mL)	Biorelevant media gastric compartment	Intestinal residence time (min)	Volume (mL)	Biorelevant media intestinal compartment	
Infants	130	69	0-30	40	Pi-FaSSGF	30-270	80	FaSSIF-V2	
Children	300	120	0-30	40	FaSSGF	30-270	80	FaSSIF-V2	
Adults	500	80	0-30	40	FaSSGF	30-270	80	FaSSIF-V2	

914

915 **Table 2.** Summary of azithromycin input parameters used in the Simcyp® simulator.

Input parameter	Value	Reference
Physicochemical Properties and Blood Binding		
Molecular weight (g/mol)	749	30
logP (experimental)	4	30
Compound type	Weak Base	30
pKa ₁	8.6	31
Blood: plasma ratio	1	32
f _{u,p}	0.69	32
Distribution		
Model	Minimal PBPK	
K _{in} (L/h)	0.93	21
K _{out} (L/h)	0.50	21
V _{sac} (L/Kg)	23	21
V _{ss} (L/Kg)	32	21
Elimination		
CL _{IV} (L/h)	46.5	32
Cl _{int} (bile) (μL/min/10 ⁶)	9.25	32
Cl _{renal} (L/h)	8.67	32
Absorption		
Model	ADAM™	
MechPeff+ P-gP Cl_{int,T} input		
P _{eff,man} [10 ⁻⁴ cm/s]	1.33	Predicted by Simcyp® Mech Peff model, used in all compartments [predicted P _{trans,0} = 1100]
Cl _{int,T} (μL/min) - Fasted state	200	Assumed - user selected after performing sensitivity analysis [against data from Foulds <i>et al</i> ²²]
Cl _{int,T} (μL/min) - Fed state	10	
Caco-2 input		
P _{eff,man} [10 ⁻⁴ cm/s], fasted state	0.29	Calculated from Caco-2 data (7.4:7.4) untreated with inhibitor, inputted as 'Passive & Active' (P _{app} 2.2 nm/s ³⁶)
P _{eff,man} [10 ⁻⁴ cm/s], fed state	0.71	Calculated from Caco-2 data (7.4:7.4) treated with a P-gP inhibitor, inputted as 'Passive' (P _{app} 16.7 nm/s ³⁶)
Formulation	azithromycin oral suspension	NA
Diffusion Layer Model (DLM)	Segmental solubility option with age-appropriate biorelevant solubility in fasted/fed state	NA
Solubility fasted state in adults, and children and adolescents (mg/mL)	13 (gastric); 2 (intestinal)	
Solubility fasted state in infants and young children (mg/mL)	12 (gastric); 2 (intestinal)	

Solubility fed state in all age-
groups (mg/mL)

10 (gastric); 6 (intestinal)

Dissolution Profile Model
(DPM)

Age-appropriate dissolution
profiles measured with USP 4
apparatus and inForm
platform, entered as discrete
profiles

NA

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917

918 **Table 3.** Simulated mean PK parameters of azithromycin in pediatrics in the fed and fasted state and
 919 predicted food effect (by comparison of predicted fed state PK parameters with fasted state-observed
 920 PK parameters).

Age group	PK Parameter	Observed Value*	Fasted state Predicted Value*	Fed state Predicted Value*	Food effect [Ratio (Pred/Obs)]
Older Children and Adolescents	T _{max} (h)	2.00	1.20	1.80	0.90
	C _{max} (ng/mL)	360	298	305	0.85
	AUC _{0-inf} (ng/mL.h)	4564	4218	4495	0.98
Infants and Young Children	T _{max} (h)	2.00	1.20	1.80	0.90
	C _{max} (ng/mL)	200	231	265	1.32
	AUC _{0-inf} (ng/mL.h)	3438	3485	3772	1.10

921

922 * after the administration of the last dose on the 5th day of a 5-day oral regimen (10 mg/Kg on the 1st day and 5 mg/Kg day 2
 923 to 5).

924

925 **Figure Captions**

926 **Figure 1.** Schematic representation of the workflow describing the PBPK development for
927 azithromycin. Abbreviations: Parameter Estimation (PE), Parameter sensitivity analysis (PSA),
928 Dissolution Profile Model (DPM) and Diffusion Layer Model (DLM).

929 **Figure 2.** Mean solubility (\pm Standard Deviation (SD)) in buffers (white bars) and biorelevant media
930 (grey bars) at 37 °C for 24 h.

931 **Figure 3.** Adult (> 18 years) and Pediatric (0.1 to 14 years) dose numbers calculated for azithromycin
932 according to solubility in buffers and biorelevant media. In pediatrics, initial gastric volumes (V_0) were
933 extrapolated with a BW- or a BSA-based calculation. The horizontal line indicates a dose number = 1.
934 A dose number > 1 indicates a low solubility compound, and a dose number < 1 indicates the opposite.

935 **Figure 4.** Mean % azithromycin dissolved (\pm SD) from Zithromax[®] oral suspension. Straight vertical
936 lines represent the time for media change in the fasted state in all age groups set-ups ; dashed and dotted
937 vertical lines represent the time for media change in the fed state in adults and infants set-up,
938 respectively.

939 **Figure 5.** Simulated azithromycin plasma concentration-time profiles (solid line: population mean;
940 dashed lines: 5th and 95th percentile of the population) in healthy adult subjects after administration of
941 a single dose of 500 mg of azithromycin oral suspension in the fasted and the fed state against observed
942 data²²⁻²⁴ [simulations were performed with MechPeff + P-gP $Cl_{int,T}$ or Caco-2 data as permeability
943 input; for the formulation input, simulations were performed either with Diffusion Layer Model (DLM)
944 using solubility as input, or direct input of dissolution with the dissolution profile model (DPM) (input
945 of age-appropriate dissolution profiles that were obtained with USP 4 apparatus or inForm platform)].

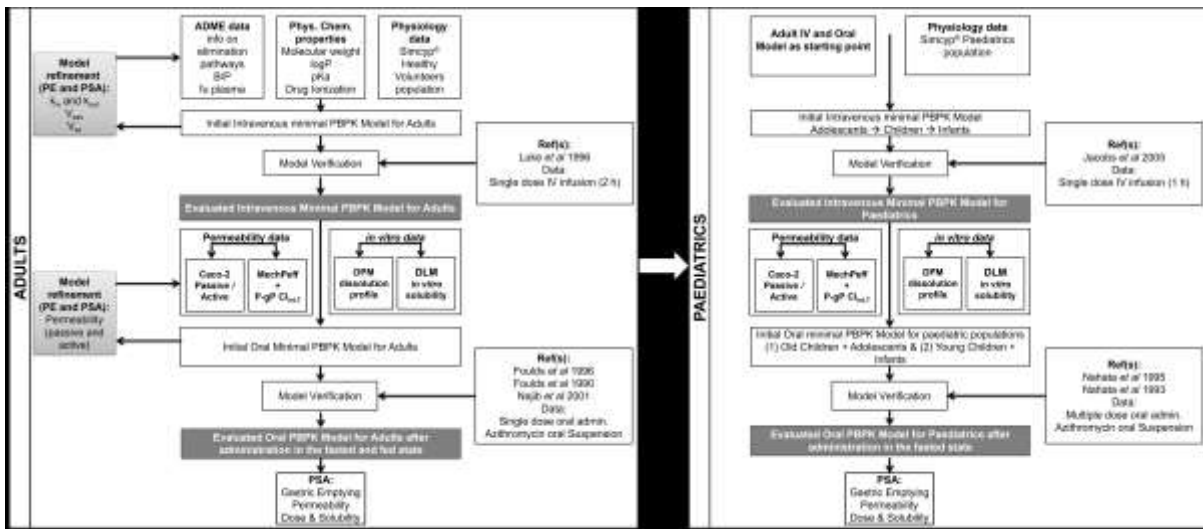
946 **Figure 6.** Simulated azithromycin plasma concentrations-time profiles (solid line: population mean;
947 dashed lines: 5th and 95th percentile of the population) on the 5th day after oral administration of
948 azithromycin as an oral suspension in a multiple-dose regimen of 10 mg/Kg (day 1) and 5 mg/Kg (day
949 2 to 5) to infants and young children (0.5 to 5 years) and old children and adolescents (6 to 15 years).
950 The final dose was administered after an overnight fast^{26,27} [simulations were performed with MechPeff
951 + P-gP $Cl_{int,T}$ or Caco-2 data as permeability input. For the formulation input, simulations were
952 performed either with Diffusion Layer Model (DLM) using solubility as input, or direct input of
953 dissolution with the dissolution profile model (DPM) (input of age-appropriate dissolution profiles that
954 were obtained with USP 4 apparatus or inForm platform)].

955 **Figure 7.** Parameter sensitivity analysis for azithromycin after administration of an oral suspension for
956 C_{max} and AUC [(a) and (b), respectively] as a function of effective permeability ($P_{eff,man}$) and P-gP $Cl_{int,T}$;
957 and (c) sensitivity analysis of C_{max} and AUC to mean gastric residence time (MGRT). Values used in
958 the PBPK model are shown in red (*i.e.* baseline simulation).

959 **Figure 8.** Parameter sensitivity analysis for the C_{max} and AUC obtained after administration of
960 azithromycin oral suspension to adults and infants and young children as a function of dose (adults –
961 250 to 600 mg; pediatrics – 40 to 220 mg) and duodenal solubility (range from 1 to 27 mg/mL).

962 **Figure 9.** Schematic summarizing the steps followed to perform the risk assessment investigation of
963 the critical variables of azithromycin's oral absorption.

964 **Figure 1**

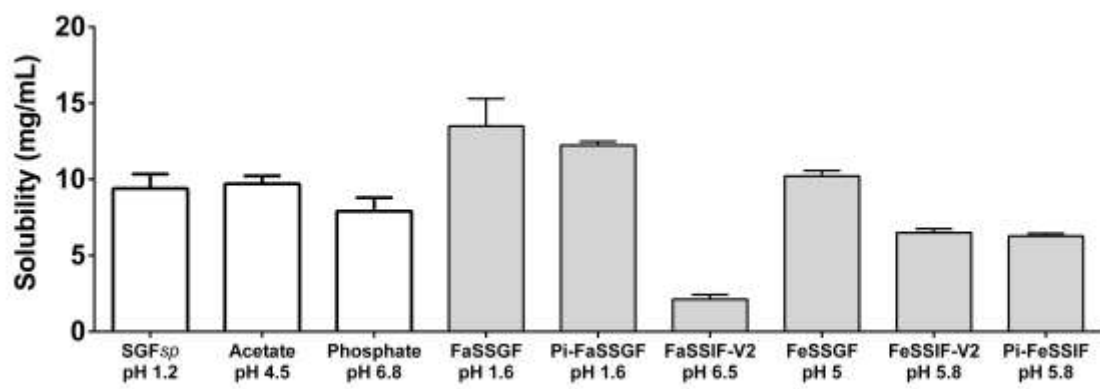


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967 **Figure 2**

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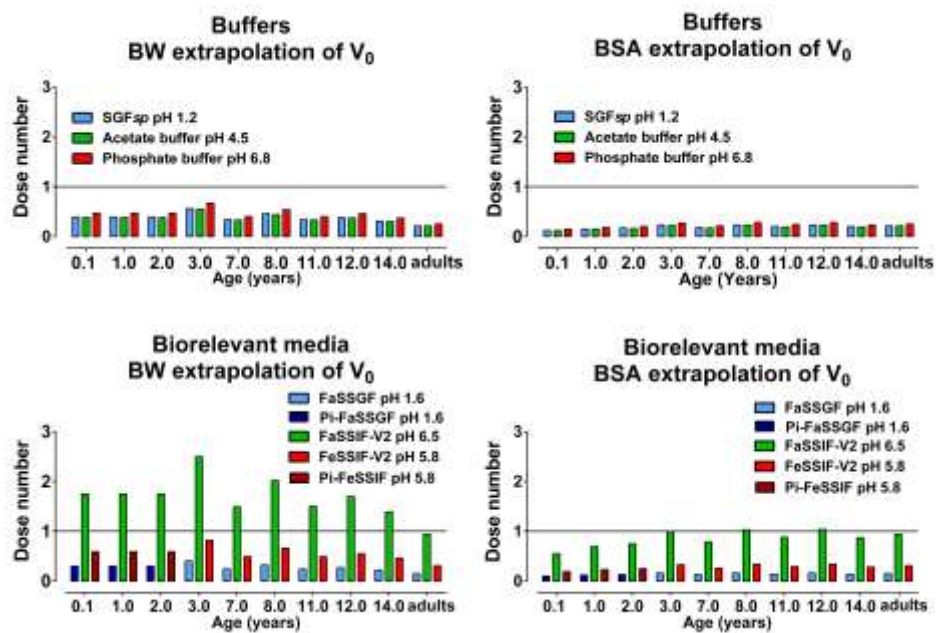
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972 **Figure 3**

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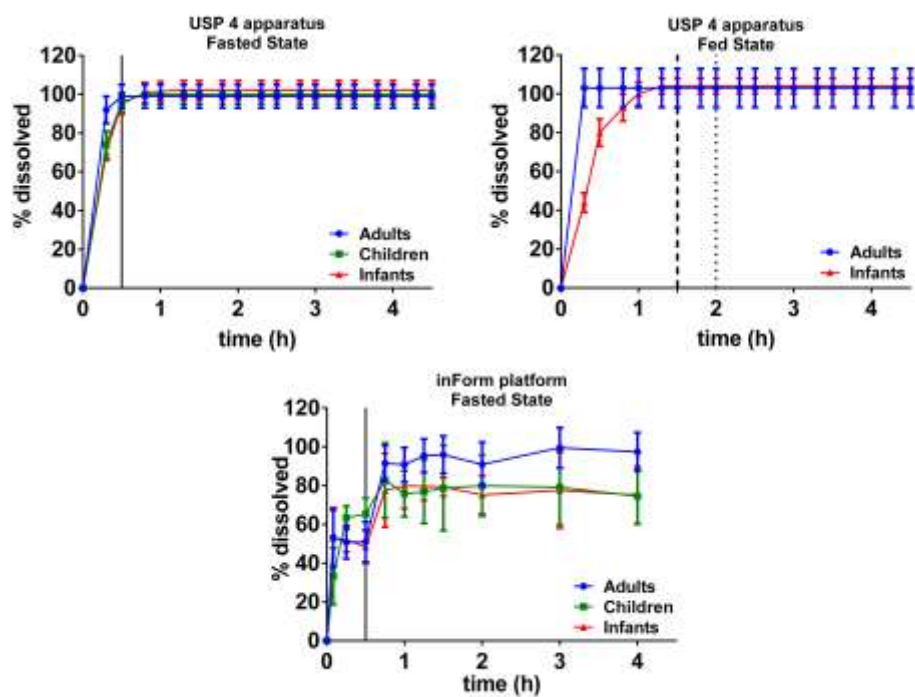
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977 **Figure 4**

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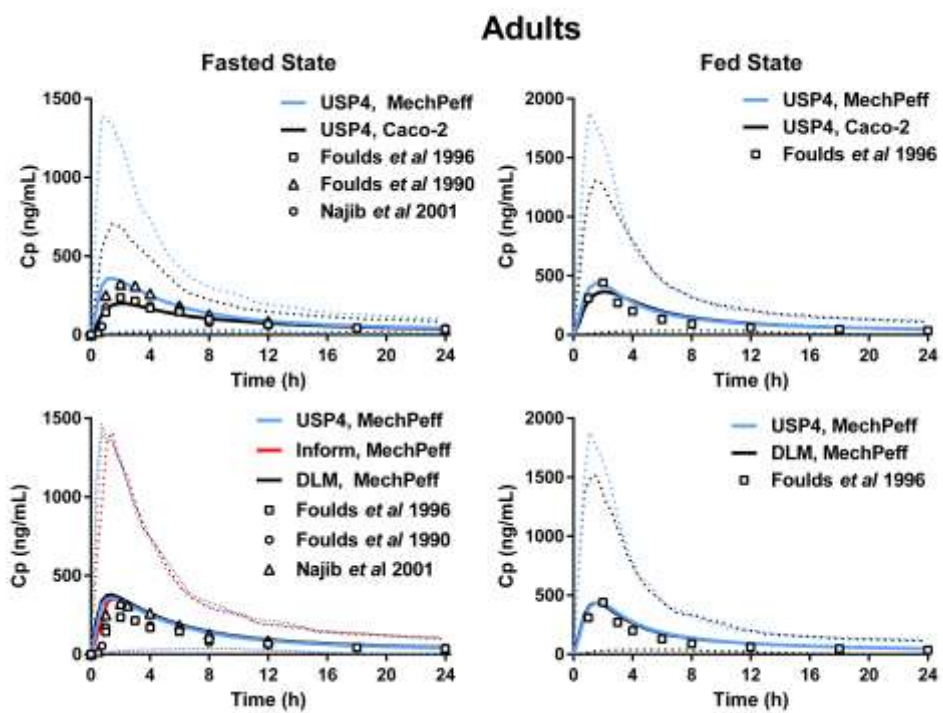


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981 Figure 5

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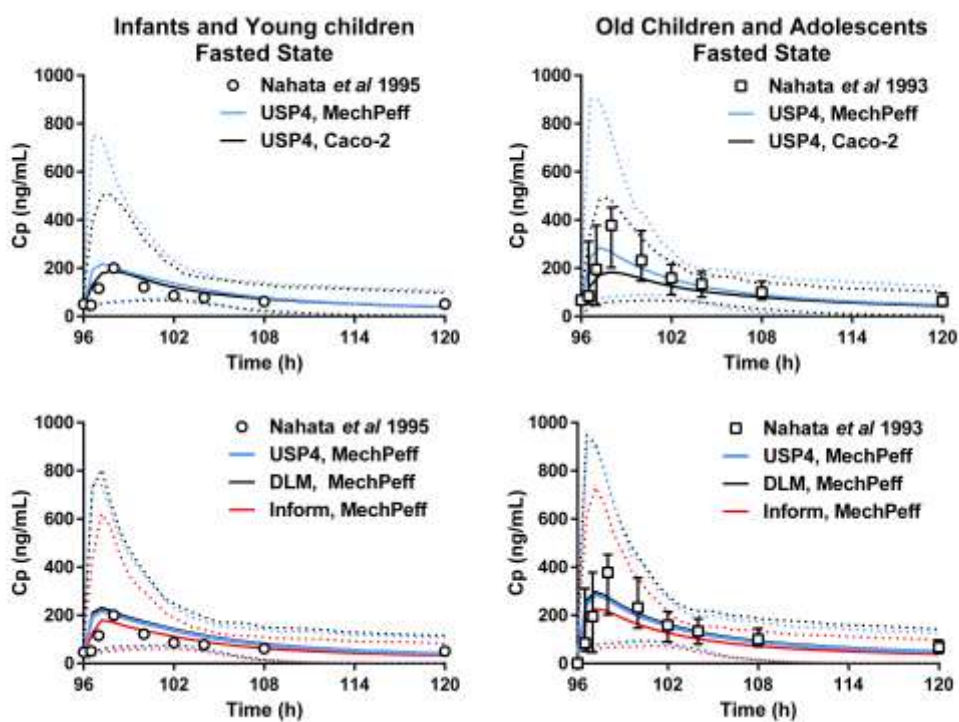


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985 **Figure 6**

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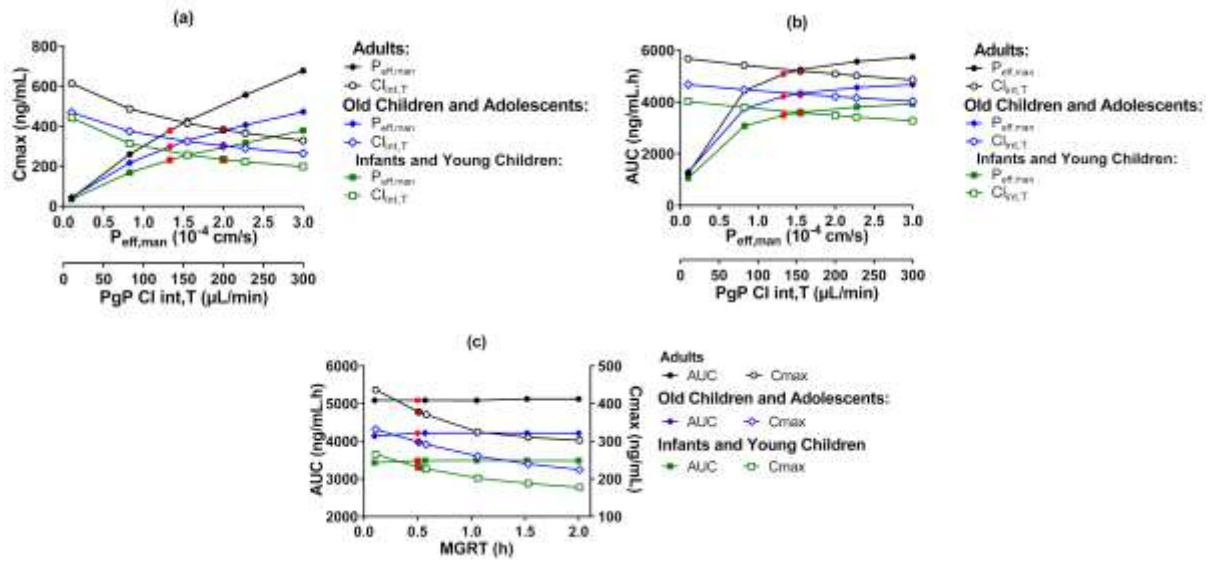


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989 **Figure 7**

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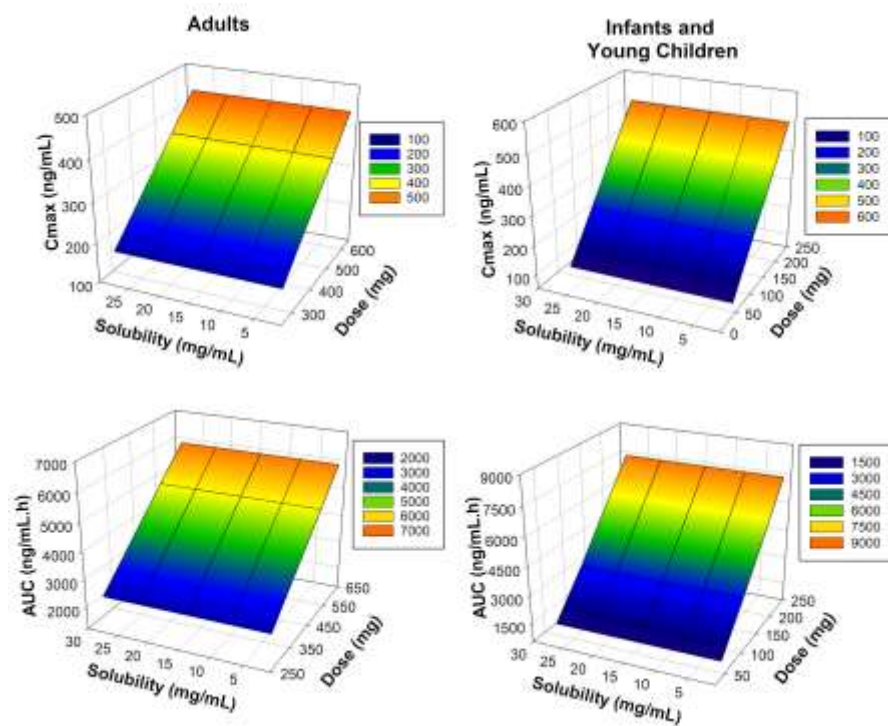
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994 **Figure 8**

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