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1 **METHODS PAPER**

2 *Cartesian coordinates in two-dimensional bone histology images ~~are useful~~ in Quaternary bone*
3 *remodelling research*

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ABSTRACT

11 Palaeohistologists who work with well-preserved hominin cortical bone can examine two-dimensional
12 (2D) histology images for quantitative parameters of secondary osteons and Haversian canals to
13 reconstruct past bone remodelling. Standard techniques in this space ~~have included taking area~~
14 measurements and counts of histology components ~~recorded from an seen in an~~ image. The ‘point-
15 count’ technique involves counting all the items (e.g., secondary osteons, osteocyte lacunae) of interest
16 per image area. The open access image analysis software ImageJ/ FIJI facilitates this technique in a
17 user-friendly way. Raw data points are captured and can be saved in a spreadsheet. Aside from the total
18 number of counts, the software also issues Cartesian (XY) coordinates locating each counted point.
19 These XY coordinates are typically neglected within palaeohistological approaches due their assumed
20 irrelevance to research questions of bone remodelling significance. We provide a short evaluation of
21 XY coordinates captured by ImageJ/FIJI from 2D bone histology images, and a protocol for a simple
22 calculation of XY distances that follow the path of point counting. We focus on osteocyte lacunae which
23 serve as a proxy for osteoblast-osteocyte conversion in live bone by replicating the protocol on a bone
24 sample from a Medieval English individual. We discuss the potential of XY coordinates for
25 ~~reconstructing the proximity of efficient representation of how closely situated the counted~~ osteocyte
26 lacunae ~~points are,~~ and ~~related thus indicating localised~~ bone remodelling activity through exchange of
27 nutrients by neighbouring cells. We recommend palaeohistologists report XY coordinate data in their
28 results to ensure better hominin palaeobiology characterisation.

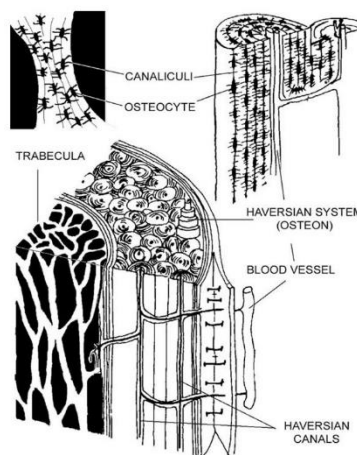
29 **Keywords:** hominins; image analysis; histomorphometry; bone remodelling; ImageJ

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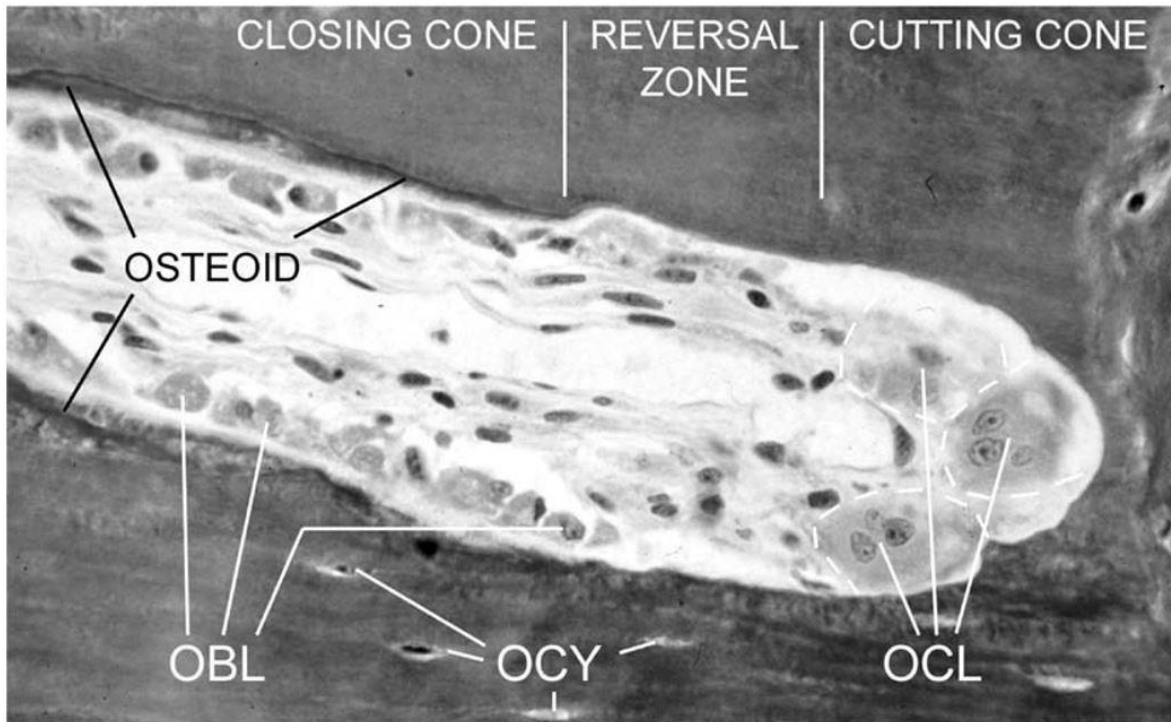
32 **1. INTRODUCTION**

33 Quantitative analysis of histology images (histomorphometry) is a gold standard in various biological
34 disciplines that collect data about tissue cells and other microscopic structures (e.g., Erben & Glösmann
35 2019; Revell 1983; O’Driscoll et al. 1999; Villa & Lynnerup 2010). Histology can also be applied to
36 bones that derive from palaeontological and archaeological contexts to reconstruct remodelling
37 processes that occur throughout an animal’s lifespan and respond to internal and environmental stimuli
38 (e.g., Bromage et al. 2009; Cook et al. 1998; Crowder & Stout 2011; de Buffrénil et al. 2021;
39 Miskiewicz 2015; Sawada et al. 2004; Stout & Stanley 1991). One useful analytical approach in bone
40 histology ~~examination~~ is quantifying geometric and density properties of secondary/ Haversian bone
41 tissue seen microscopically in thin sections taken in a transverse plane (Stout & Crowder 2012) (Figure
42 1). The histological appearance of Haversian bone reflects the activity of Bone Multicellular Units
43 (BMUs) which are three-dimensional (3D) cell structures that execute bone remodelling in live bone
44 by ‘digging’ longitudinal tunnels as bone is resorbed and deposited (Figure 2) (Lassen et al. 2017;
45 Maggiano et al. 2016; Sims & Martin 2014). Once quantified, cortical bone histology data can be
46 studied at an individual or large sample level representing past populations (e.g., Chan, Crowder &
47 Rogers 2007; Stout & Lueck 1995; Miskiewicz et al. 2020).



48
49 **Figure 1.** Sketch of the microscopic components of bone showing Haversian systems and osteocytes.
50 Reproduced from Smit et al. 2002 p. 2022, with permissions from John Wiley and Sons under licence
51 5297961008734.

52
53



54

55 **Figure 2.** An example of an active Bone Multicellular Unit (BMU) showing osteoclasts (OCL),
 56 osteoblasts (OBL), and osteocytes (OCY). Image is not to scale, but the width of the BMU would be
 57 approximately 200 μm . Reproduced from Smit et al. 2002 p. 2024, with permissions from John Wiley
 58 and Sons under licence 5297961008734.

59

60 All bone biologists who undertake histomorphometry are expected to follow recommended guidelines
 61 for unified nomenclature and techniques used in calculating parameters of bone remodelling (Dempster
 62 et al. 2013). Working with Haversian tissue, the relevant quantifiable histology variables include the
 63 area, diameter, density, and circumference of structures such as secondary osteons, Haversian canals,
 64 and osteocyte(s)/lacunae (osteocytes do not always preserve in post-mortem bone, so osteocyte lacunae
 65 indicate where the osteocytes had resided) (Crowder & Stout 2012; Hunter & Agnew 2016; Stout, Cole
 66 & Agnew, 2019; Bromage et al. 2009). The densities of secondary osteons or osteocyte lacunae are
 67 typically calculated by summing up the total counts seen in a specified field of view or a selected region
 68 of interest (ROI), and then dividing the raw count by the area examined (Crowder & Stout 2012;
 69 Crowder et al. 2022; Drew, Mahoney & Miskiewicz 2021). This counting approach is known as the
 70 'point count' technique (Stout & Stanley 1991). Secondary osteon density, referred to as osteon
 71 population density (OPD), data reflect the amount of remodelled bone per area (Frost 1987), whereas
 72 osteocyte density reflects osteoblast-osteocyte conversion (Bromage et al. 2016).

73 While these densities have been reported by multiple studies investigating topics that have ranged from
74 biomechanics (Drew, Mahoney & Miskiewicz, 2021), diet (Paine & Brenton 2006), to human
75 evolution (Sawada et al. 2004), a notable gap in analyses is that the proximities of individual counted
76 points are not considered. This is interesting because biologists regularly consider dispersal or proximity
77 analysis when analysing points against a specified area to assess the randomness, clustering, or
78 migration of cells (Cruz et al. 2005; Fernández, Lysakowski & Goldberg, 1995; Gorelik & Gautreau
79 2014; Wang et al. 2021). Only a handful of palaeontological, bioarchaeology, or anthropological studies
80 have considered spatial distribution relationships within bone histology sections. This has included
81 Rose et al. (2012), and Gocha and Agnew (2016), who brought to light the value of applying Geographic
82 Information Systems (GIS) software in understanding osteon population dispersal across complete
83 human metatarsal and femoral bone cross-section shafts, respectively. Further, Miskiewicz et al. (2020)
84 applied GIS to measure distances between neighbouring Haversian canals across thin sections from the
85 femoral midshaft in an archaeological specimen, spatially mapping cortical bone porosity. GIS can
86 require substantial training. An alternative, commonly used, user friendly, and self-intuitive, image
87 analysis software is the open access ImageJ, part of the FIJI package (Schindelin et al. 2015). In this
88 paper, we highlight that ImageJ/FIJI also registers XY coordinates alongside point counts and can be
89 used to efficiently calculate path distances between neighbouring histology structures. We focus on
90 hypothetical osteocyte lacunae in a sketched diagram, supply a manual protocol based on established
91 mathematics for the path distance calculations, and then replicate it on a secondary osteon image from
92 an archaeological (dated to Medieval England) human femur.

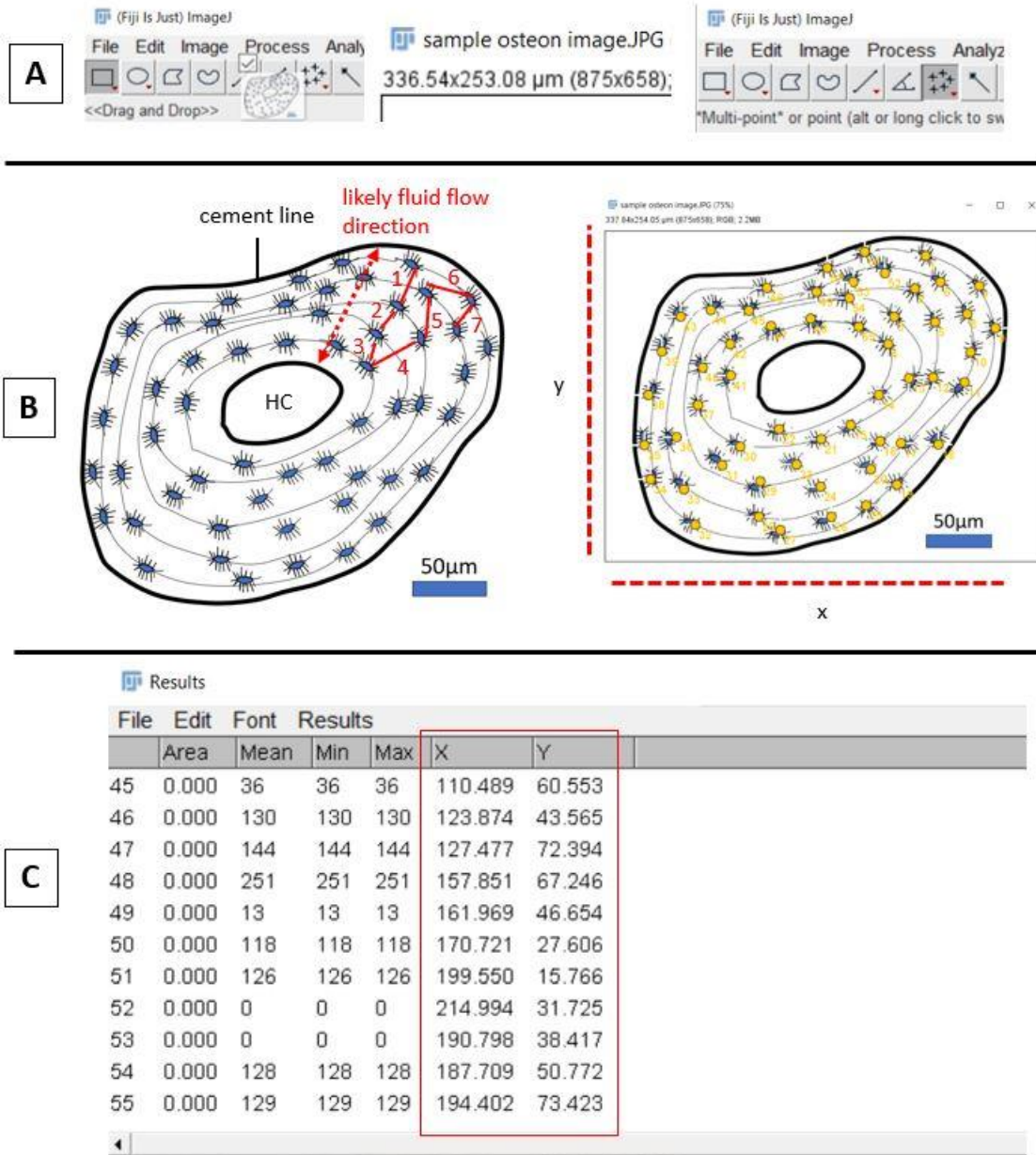
93 **2. METHODS: XY COORDINATES IN IMAGEJ/FIJI**

94 **2.1. The protocol and application to a hypothetical sample**

95 ImageJ/FIJI can be freely downloaded from the dedicated website (<https://imagej.net/software/fiji/>).
96 The preparation of bone histology slides will not be covered here. A ready image with a known scale
97 captured using a dedicated microscope camera is needed to apply the protocol we discuss. We note
98 ImageJ/FIJI provides macros and plug-ins that calculate various other aspects of XY coordinates (e.g.,
99 distance from centroid), but these require coding and script amendments. Distances between selected

100 points can also be measured manually one by one using the “Straight line” tool of ImageJ/FIJI, but this
101 approach takes more time than the presented XY coordinate analysis and has to be conducted in addition
102 to point counting. We provide a manual guide for path distance calculations from the XY coordinate
103 data recorded at the same time as point counts which is straightforward and efficient in its application.

104 The first step (Figure 3a) is to open ImageJ/FIJI (we used vol. 153o), import the image of interest, and
105 set the appropriate scale (“Analyze” > “Set scale”). The counts of the osteocyte lacunae can be
106 completed using the “Multi-point” function selected from the ImageJ/FIJI toolbar. For the distances to
107 be most meaningful biologically, the sequence of counts made would ideally be either conducted
108 starting at the Haversian canal and continuing outwards (towards the cement line), or *vice versa* (Figure
109 3b). That is because canaliculi, which connect successive osteocytes within a secondary osteon, are
110 largely arranged radially so as to maximise fluid flow (van Tol et al. 2020). By pressing “M”, the
111 measurements can be obtained. A “Results” window will appear where the following measurements are
112 recorded: “Area”, “Mean”, “Min”, “Max”, “X”, “Y” (Figure 3c). The “X” and “Y” are the coordinate
113 values that can be used to understand the proximity of these points. The output from the “Results”
114 window can be saved as an Excel spreadsheet, or the XY coordinates can be copied and pasted into a
115 new spreadsheet.



116

117 **Figure 3.** Series of initial steps of point counting in ImageJ/FIJI. A: importing of image (HC:
 118 Haversian canal), setting the scale, and selecting the “Multi-Point tool”. B: outline of secondary
 119 osteon diagram with osteocyte lacunae prior to (left), and after (right), counting of the lacunae. C:
 120 ImageJ/FIJI data window showing XY coordinates for each point (red box, left), transported into a
 121 spreadsheet (right).

122

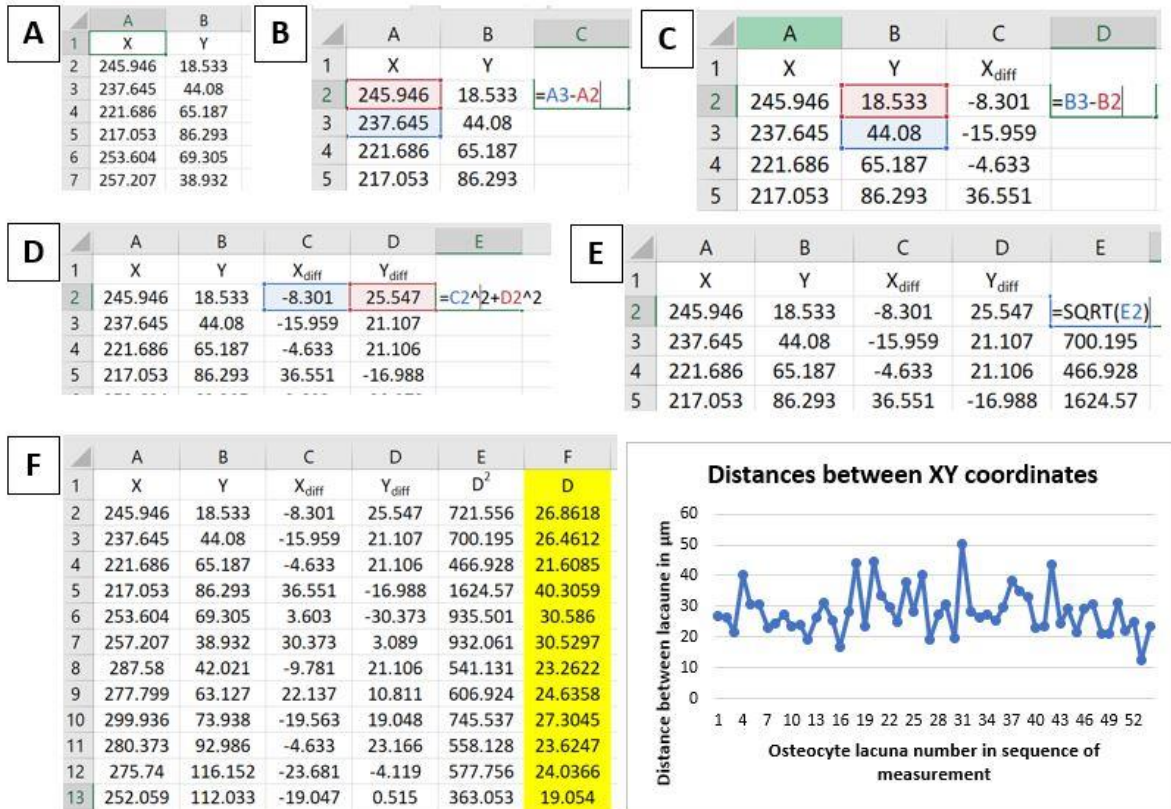
123 The next step is to calculate the 2D distance between XY coordinates, following the path of counting,
 124 using the formula

126
$$D = \sqrt{(x_2 - x_1)^2 + (y_2 - y_1)^2}$$

125 (derived from the Pythagorean theorem) (Figure 4).

- 127 1. In the same or a new spreadsheet, new columns aligned next to each other can be created and
128 named: X_{diff} (this will be the difference between successive X coordinates), Y_{diff} (this will be
129 the difference between successive Y coordinates), D^2 (squared distance between X_{diff} and Y_{diff}),
130 D (square root of D^2).
- 131 2. The difference between successive X and successive Y coordinates is then calculated
132 individually (i.e., per axis). This can be achieved in Microsoft Excel by typing in “=cell with
133 raw X value-preceding cell with raw X value” - the example in Figure 4 would be “=A3-A2”.
134 This then can be repeated on the Y coordinates. The last cell should not be considered because
135 it does not have a successive coordinate. The cell function can be dragged down the entire
136 column.
- 137 3. D^2 , defined through the formula: $D^2 = X_{diff}^2 + Y_{diff}^2$, is then calculated. This can be achieved in
138 Excel by typing “=cell with X_{diff}^2 +cell with Y_{diff}^2 ”. The example shown in Figure 4 would be
139 “=C2^2+D2^2”. The cell function can be dragged down the entire column.
- 140 4. Finally, the square root of D^2 is calculated which can be done in Excel by typing “=SQRT(cell
141 with D^2 value). In Figure 4, the example would be “=SQRT(E2)”. The cell function can be
142 dragged down the entire column.

143 Raw data resulting from this hypothetical image analysis can be found in our Supplemental File. The
144 resulting D values are the distances in the unit appropriate to the image between the XY coordinates
145 registered along the pathway of counting. They can be analysed across a whole osteon, summarised as
146 averages per osteon, a full image, individual, larger sample – whatever the desired level of analysis as
147 per one’s research question. In the presented example, the average distance across this one hypothetical
148 secondary osteon was 28.25 μm (see Supplemental File).



149

150 **Figure 4.** Screenshots of successive steps in XY distance coordinate calculations in Microsoft Excel.
 151 A: XY coordinates from the ImageJ/FIJI results output, B-C: calculating differences between successive
 152 X and Y coordinates, D: calculating the squared distance (D^2) between XY differences, E: calculating
 153 the square root of D^2 , F: final distance values of XY coordinates (D) in μm (highlighted in yellow),
 154 which can be quickly visualised using a simple line graph.

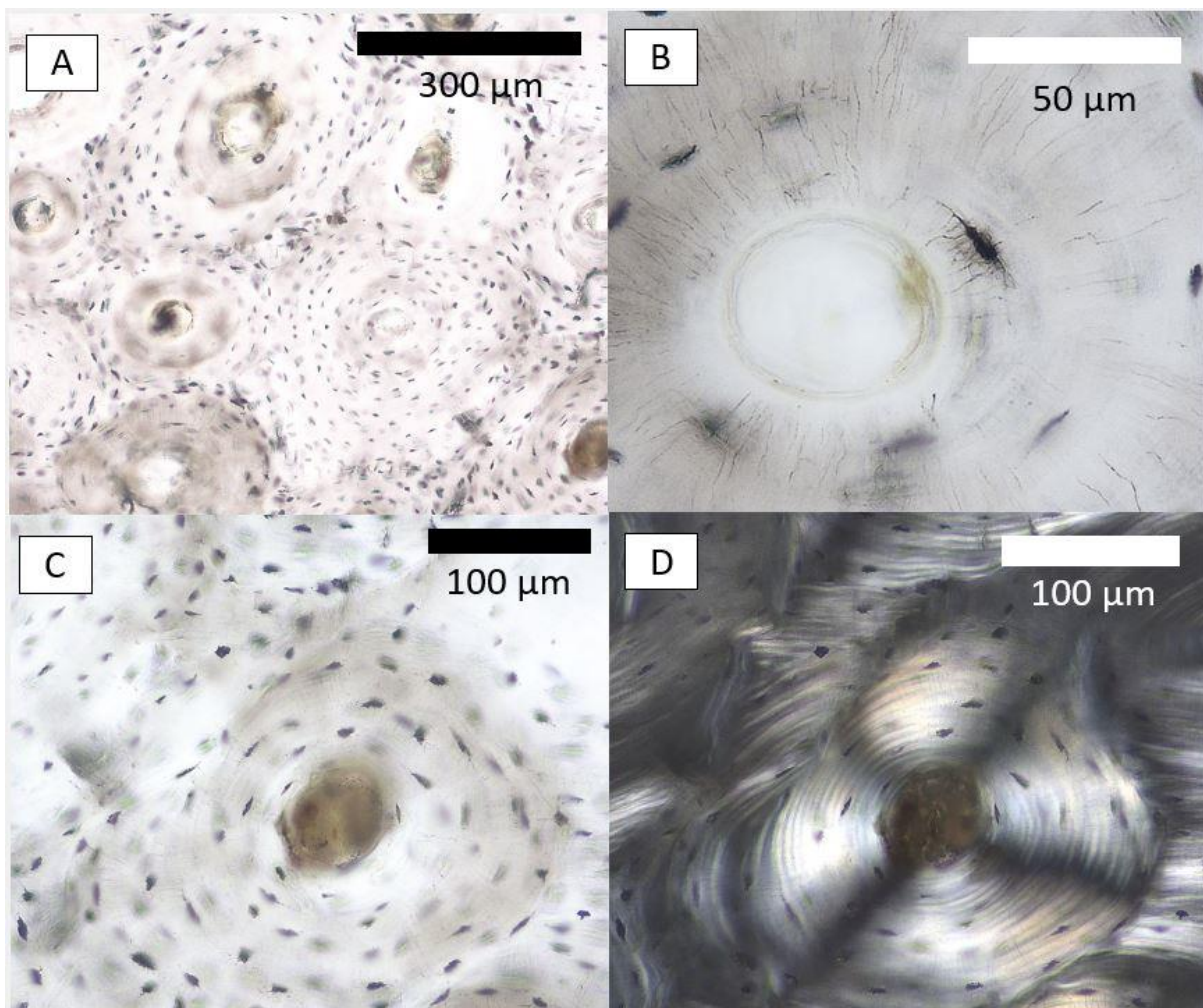
155

156 **2.2. Replication on an authentic sample**

157 To validate this short protocol, we applied it step-by-step to a real case – using a cortical bone histology
 158 image showing a well-preserved secondary osteon in the femur of a Medieval English individual (ID:
 159 NGB89SK2). The thin section is a 100- μm thick slice of midshaft cortex extracted from the anterior
 160 aspect of the femur. This individual was estimated to be a middle-aged (35-50 years old) male
 161 (Miszkievicz, 2014), and the sample is part of a larger collection of thin sections representing a
 162 Medieval English human osteological assemblage curated at the University of Kent, UK. These thin
 163 sections had been prepared following standard histology methods applied to archaeological human
 164 remains (Miszkievicz, 2014). A full ethics board review was not required as this is an archaeological
 165 collection, but a local (School of Anthropology and Conservation) ethics application form and check
 166 list was filed on 11 November 2010 and approved by the 2010 Ethics Advisory Group at the University

167 of Kent, UK. In addition to curatorial permissions, all analyses on these sections follow codes of ethics
168 and good practice as stipulated by the international biological anthropology bodies (British Association
169 for Biological Anthropology and Osteoarchaeology; American Association of Biological
170 Anthropologists). The anterior femur sample presented here was also previously included in research
171 by Drew, Mahoney and Miskiewicz et al. (2021), Miskiewicz and Mahoney (2012), and Miskiewicz
172 (2014). The XY coordinate distance data are calculated here for the first time.

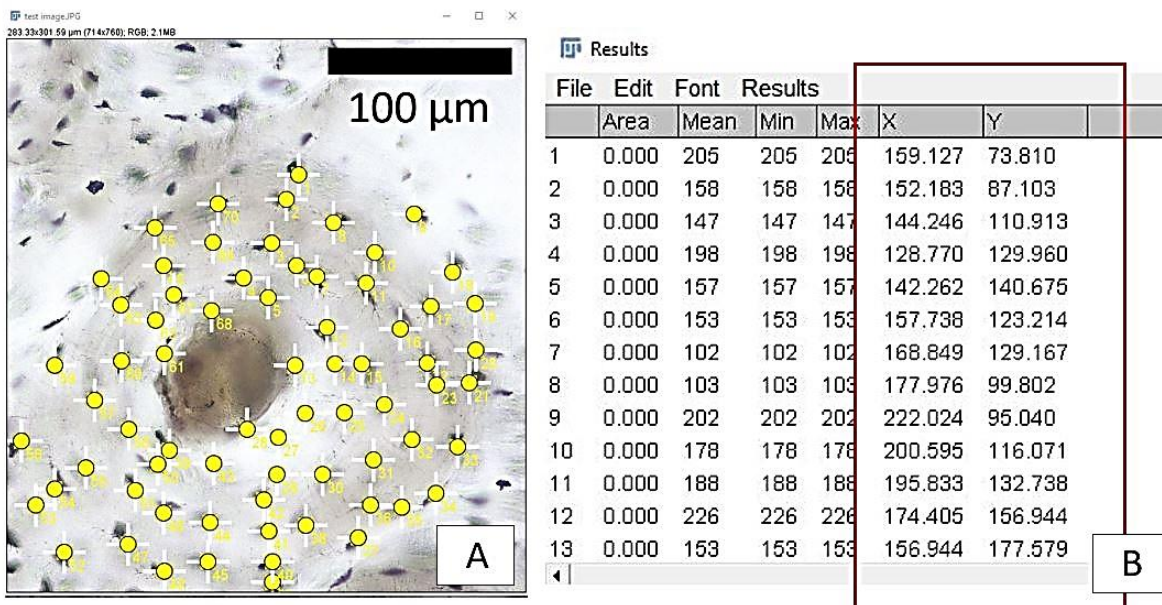
173 Using an Olympus BX60 high powered microscope equipped with a MIchrome 5 Pro SS-936 camera
174 and operated using Capture v2.1 software, one secondary osteon showing clear cement lines (Figure
175 5A) and well-preserved osteocyte lacunae (Figure 5B) was selected from intra-cortical bone. It was
176 viewed under linearly polarised light to check good preservation, i.e., ensuring that individual
177 concentric lamellar rings could be seen clearly (Figure 5CD).



178

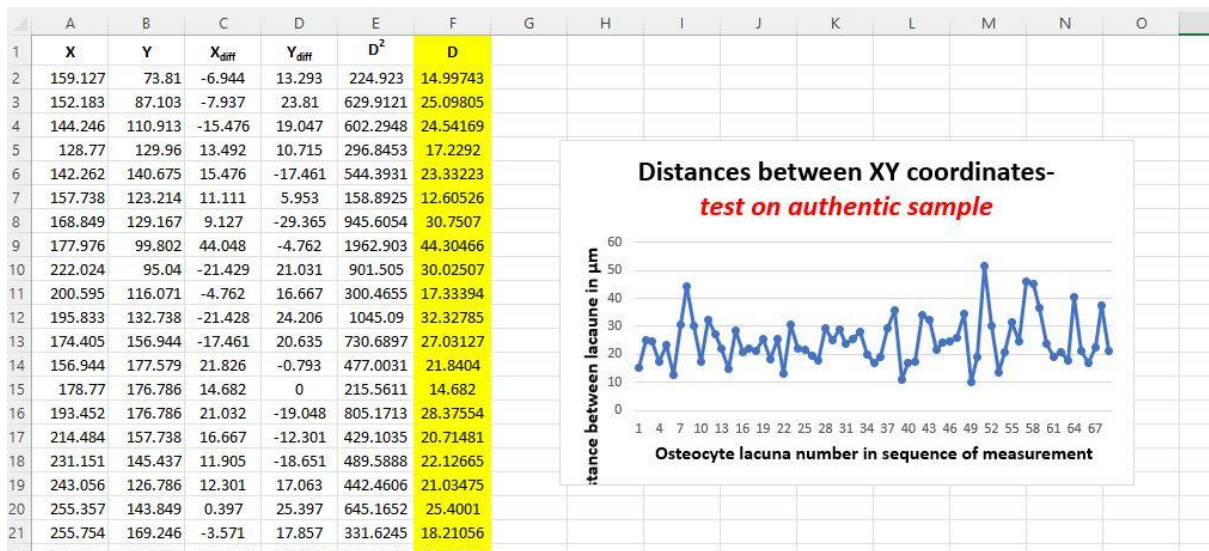
179 **Figure 5.** Cortical bone histology images in a sample from a Medieval English male (ID: NGB89SK2,
 180 St. Gregory's Priory and Cemetery collection, University of Kent, UK) taken from the anterior midshaft.
 181 Image A shows a cluster of secondary osteons with a range of osteocyte lacunae preservation. Image B
 182 shows a closeup of an osteocyte lacuna as an example of good preservation in the secondary osteon
 183 selected for our replication analysis. Images C and D show the secondary osteon under polarised
 184 transmitted (C) and linearly polarised light (D).

185 The secondary osteon image captured using transmitted light (so that each osteocyte lacuna could be
 186 identified easily (black lacunae against white bone)) was examined following our protocol above. The
 187 image was imported into ImageJ/FIJI (vol. 153o). Osteocyte lacunae were counted starting at the cement
 188 line and continuing towards the Haversian canal in a linear fashion (Figure 6). The resulting XY
 189 coordinates were used for calculating distances between the neighbouring osteocyte lacuna, the average
 190 of which for the entire secondary osteon was 24.84 μm .



191

192 **Figure 6.** Screenshots of image analysis steps replicating our XY coordinate calculation protocol. A:
 193 secondary osteon with osteocyte lacunae (yellow dots). B: raw data output window in ImageJ/FIJI
 194 showing resulting XY coordinates (red rectangle).



195

196 **Figure 7.** Screenshot of final calculations of distances between XY coordinates yielded using the
 197 Medieval English histology sample. Column F, highlighted in yellow, shows the D values in μm . The
 198 line graph illustrates variation in D values from across the entire secondary osteon.

199

200 **3. DISUSSION: INTERPRETATIVE VALUE**

201 We present here a simple and effective way to obtain an insight into the distances between neighbouring
 202 points [along a path of counting](#) in a 2D histology image. Those who regularly work with osteocyte
 203 lacunae or OPD data might already have raw data files from ImageJ/FIJI stored where the XY
 204 coordinates are saved. Certainly, whenever possible these should be provided alongside other
 205 commonly reported variables, either in the main body of an article or the online supplementary
 206 information. Re-analyses of data as outlined here may provide new insights into new or old research
 207 questions, although it is also vital to provide the sequence in which the points are counted. For example,
 208 very wide distances between some coordinates may simply reflect random and widespread counting
 209 across an image and thus have no meaning for understanding distances between neighbouring bone
 210 histology elements.

211 The interpretative value of the distances between osteocyte lacunae on a path of counting is that it would
 212 inform how well inter-connected osteocytes were in a given sample. This provides information about
 213 bone remodelling processes and how actively bone was maintained (Andreas, Colloca & Iacoviello
 214 2014; Bromage et al. 2009). This is because the inter-connectedness of neighbouring lacunae facilitates

215 healthy exchange of nutrients, bone biological signals, and fluid flow (Kollmannsberger et al. 2017;
216 van Tol et al. 2020). For example, it has been suggested that, in humans, osteocyte-to-osteocyte distance
217 of 20-30 μm is enough to facilitate effective dissemination of molecules (Chen et al. 2018; Wang et al.
218 Sugawara et al. 2005). In our Medieval example, the resulting D values fell into this distance range,
219 which could be used to infer that osteocyte communication was not disrupted in this region of bone.
220 Canè et al. (2000) noted changes in osteocyte distribution depending on the long bone region (e.g.,
221 cortical vs trabecular bone, diaphysis vs metaphysis vs epiphysis). Thus, future research into osteocyte
222 lacunae distance data applied across hominin bone could be placed within the wider bone biology
223 knowledge and help further refine interpretations made from bone histology density and other geometric
224 variables alone.

225 The XY coordinate distance measurements may not be as useful in understanding secondary osteon
226 distribution as they might be when applied to osteocyte lacunae, simply because osteons can remodel
227 both stochastically and in a targeted fashion (Martin 2007), so it is almost impossible to determine the
228 timing (sequence of secondary osteon appearance) of remodelling from post-mortem, archaeological,
229 or fossilised bone samples. Identifying the timing of remodelling would also be difficult to achieve
230 when dealing with bone from older individuals that show several generations of remodelled secondary
231 osteons, possibly reflecting an OPD asymptote (Frost 1987). However, the technique could be
232 applicable to counts of intact osteons (see Crowder et al. 2022) across a relatively larger image area (or
233 a shaft cross-section) to determine whether BMU products were operating in a clustered or dispersed
234 manner, alongside the osteon density data.

235 The key limitation of using the XY coordinate distance in bone histology examination is, of course, the
236 2D level of perspective which does not capture the 3D nature of osteocytes or osteonal structures in a
237 bone sample (Andronowski & Cole 2021). Another limitation is that the outlined calculations need to
238 be processed manually, in the sense that they have to be analysed outside of ImageJ/FIJI. However,
239 once the main cell functions are set up, the XY coordinates from the ImageJ/FIJI output can be simply
240 copied and pasted into a spreadsheet (a template is supplied in our Supplemental File). The advantage
241 of using ImageJ/FIJI lies in having access to a forum of supporting scholars who are continually

242 developing macros and plug-ins, and assist with coding, that can be used to calculate other XY
243 coordinate distances that might be of interest to palaeohistologists (e.g. distance from centroid). Our
244 protocol does not require significant training or access to expensive software and so would be applicable
245 across a wide range of institutions, particularly where there is limited infrastructure (see Miskiewicz
246 2020). While other disciplines can employ automated approaches to cell counting and measuring from
247 2D or 3D images (e.g., O’Driscoll et al. 1999), histologists working with archaeological or fossilised
248 samples are hindered by issues with preservation, meaning that careful manual approaches to histology
249 image analysis will continue to be needed. Being able to conduct manual XY coordinate counts could
250 be a useful addition to the technical toolkit.

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253 **Competing interests**

254 We have no competing interests to declare.

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