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Selection-driven chicken phenome and phenomenon of pectoral angle variation across different chicken phenotypes

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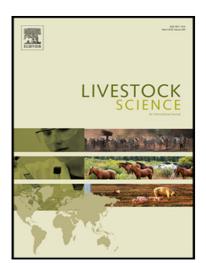


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| Genetics, Animal Genetic Resources and Breeding

Selection-driven chicken phenome and phenomenon of pectoral angle variation

across different chicken phenotypes

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Highlights

- Selection-driven chicken phenome embraces varied phenotypes including pectoral angle.
- We studied PA variation across divergently selected chicken breeds.
- Patterns of PA relationship with performance traits were analysed.
- We propose a promising PA-based auxiliary index for using in layer selection.

Abstract

An appreciation of the synergy between genome and phenome of poultry breed is essential for a complete understanding of their biology. Phenotypic traits are shaped under the influence of artificial, production-oriented, selection that often acts contrary to that which would occur during natural selection. In this comparative study, we analysed the phenotypic diversity of 39 chicken

breeds and populations that make up a significant part of the world gene pool. Grouping patterns of breeds found within the traditional, phenotypic models of their classification/clustering required indepth analysis using sophisticated mathematical approaches. As a result of studying performance and conformation phenotypes, a phenomenon of previously underestimated variability in pectoral angle (PA) was revealed. Moreover, patterns of PA relationship with productive traits were analysed. We propose using PA measurement as a promising new auxiliary index for selecting hens and roosters of breeding flocks in egg production improvement programs.

Keywords: phenome, phenotypic traits, pectoral angle, egg production, chicken breeds, gene pool

Introduction

The world poultry gene pool has been formed as a result of long-term processes of domestication and breeding for phenotypic traits. These include economically important traits that make up phenome – the sum total of all phenotypes expressed in a given species (Rexroad et al., 2019). This led to the creation of numerous breeds, often synthetic by origin, that represent valuable genetic resources for the development of contemporary poultry farming, thereby facilitating further selection (Romanov, Bondarenko, 1994a; Romanov et al., 1994; Weigend et al., 2004a; Tagirov et al., 2006; Paronyan, 2016). Poultry breeds and varieties can carry certain mutations that shape a breed (e.g., Moiseyeva et al., 2009a; Corti et al., 2010) or cause abnormal developmental deviations (Bondarenko et al., 1995, 1996); they are characterised by important traits of performance and resistance to diseases (e.g., Dementeva et al., 2017, 2018).

A prominent geneticist Aleksandr S. Serebrovsky (Moiseyeva et al., 2012a,b, 2013; Romanov et al., 2012) drew attention to the importance of studying the gene pools of animals including poultry and, along with Sergey G. Petrov (Moiseyeva et al., 2000) and others, performed studies on the phenogeography of domestic chicken (*Gallus gallus*) populations. Thereafter, collection flocks of poultry breeds were developed (e.g., Bondarenko et al., 1986), one of which was created at the Research Institute of Farm Animal Genetics and Breeding (RIFAGB) (Paronyan et al., 2014; Dementeva et al., 2017, 2018; Dementieva et al., 2018; Romanov et al., 2021). The preservation of

gene pools continues to be an urgent problem (Romanov et al., 1994; Wężyk et al., 1994; Wezyk et al., 1996; Romanov et al., 1995, 1999). Various approaches and methods have been used to assess the degree of diversity and structure of gene pool breeds and populations. Phenetic and phenogeographic studies can provide important primary information about known bird diversity, as has been shown for various gene pool breeds and populations, not only of chickens, but also geese and ducks (Romanov, 1992, 1993, 1994, 1995a,b). In contemporary breeding practices, molecular markers are widely used by geneticists for these purposes, e.g., RAPD, microsatellites, SNPs, etc. (Weigend, Romanov, 1999, 2002; Romanov et al., 1999; Romanov, Weigend, 2001; Weigend et al., 2004b; Dehghanzadeh et al., 2009; Dementeva et al., 2017; Lee et al., 2017; Dementieva et al., 2018).

Phenomics, an emerging area, covers the combined study of the phenome and big data associated with it, including the development of phenome databases, and genome-phenome correlations (e.g., Furbank et al., 2011; Rexroad et al., 2019; Amar et al., 2021; Matalonga et al., 2021; Silva et al., 2021). To some extent, this echoes the earlier discipline of phenetics (Romanov, 1992, 1993, 1994, 1995a,b), but phenomics is a more comprehensive concept and a newer, high throughput research field embodied in new projects such as the Agricultural Genome to Phenome Initiative (AG2PI, 2021). Less attention has been paid to the in-depth study of phenotypic traits in poultry, although their comparative characteristics can provide interesting and important information on the gene pool of various breeds (Larkina et al., 2021; Romanov et al., 2021).

The domestic gene pool of chickens is represented by the older and world–renowned breeds, e.g., the Orloff (Somes, 1988; Moiseyeva et al., 2016), Pavlov (Moiseyeva et al., 2009a; Corti et al., 2010), Yurlov Crower (YC) (Moiseyeva et al., 2007a, 2009b, 2011), Poltava Clay (PC) (Romanov, Bondarenko, 1994b; Moiseyeva et al., 2006, 2007b; Kulibaba et al., 2015) and others, as well as relatively younger breeds, e.g., Russian White (RWG) (Dementeva et al., 2017, 2018; Abdelmanova et al., 2021). Many of these breeds are synthetic, developed using different selection paths, and maintained in the RIFAGB bioresource collection (Paronyan et al., 2014), including the

Orloff Mille Fleur (OMF), two varieties of the restored Pavlov breed (Pavlov Spangled, PS; Pavlov White, PW), YC, PC, and others. They are targeted by conservation, breeding and selection efforts. The collection also contains imported foreign breeds that have specific, sometimes unusual, phenotypic and genetic characteristics, such as the French breed of Faverolles Salmon (FS) and the Chinese breed of Silkie White (SW) (Somes, 1988; Moiseyeva et al., 2009a; Corti et al., 2010). Various breeds and populations of the RIFAGB collection flock include poultry of all utility (purpose) and performance types according to the traditional classification model (TCM) that subdivides the entire chicken breed suite into egg-type breeds (ETB), meat-type breeds (MTB), dual purpose (egg-meat, EMB; and meat-egg breeds, MEB), game breeds (GB) and fancy breeds (FB) (Tables 1 and S1; Bogolyubsky, 1991). All these serve as a valuable resource for genetic and biological research (Paronyan, 2016).

In a previous study (Larkina et al., 2021), we examined models of evolutionarily determined subdivision of the world gene pool of chicken breeds as driven by artificial selection for performance (utility) and other phenotypic traits. The phenotypic clustering model (PCM) postulated by us enabled us to cluster, with sufficient plausibility, the existing RIFAGB collection chicken populations. PCM revealed certain differences from TCM. This facilitated amendment of the earlier concept of four evolutionary lineages in domestic chicken breeding postulated by Moiseyeva et al. (2003) and specified the attribution of the RIFAGB collection populations to one or another category of breeds (Tables S2 and S3). At the same time, we tested a number of phenotypic (exterior) and genotypic parameters that could be used when evaluating individuals in terms of their subsequent selection for increased egg or meat performance. As a result of this investigation (Larkina et al., 2021), it was not possible to identify any phenotypic (conformational) or genotypic indicator that would be directly related to the performance of a breeding flock. In this regard, we feel that it would be worthwhile to take a closer look at certain exterior features of breeder females and males, and their possible relationship with productivity traits.

The issue of the relationship between the observable characteristics of a bird (i.e., its phenome; Rexroad et al., 2019) and its performance is not new. Examination of the exterior of animals in order to determine their breeding value is always an important part of livestock breeding and selection. Examination of the external characteristics provides a general overview of assessing the appearance of a bird. The external characteristics include general conformation, outline and posture of the body and limbs, as well as features of external shape, and the development and structure of separate body elements, i.e., build, plumage, body weight (BW), skin colour, and other specifics of skin and feather covering. Performance traits and features of the exterior and conformation characteristic of birds of various poultry species, breeds and strains are inherited, but are subject to alteration under the influence of selection, feeding, growing conditions of young stock, and other factors (Landauer, 1941; Arzhankova, Ivanova, 2012). In this regard, a key significance of the external assessment is the most accurate understanding of the predisposition of a bird/breed to manifesting expected phenotypic traits. The latter include performance use, constitutional strength, health and fitness of an organism to specific environment conditions, and the overall utility type of an animal for which it is bred, raised and exploited. Thus, by examining the exterior, it is possible to judge, within certain limits, the performance potential of an animal/breed and important breed characteristics. These are correlated with the biological sustainability and adaptability to the environment in which the breed exists, produces and reproduces full-fledged offspring (Borisenko, 1966). Only birds purposefully assessed/selected and harmoniously conforming to each other and the environment are expected to be the most viable and productive.

Furthermore, in terms of the ETB exterior features that can be linked to a higher egg productivity, one should note that the head shape in these breeds is less massive than in MTB, the neck is longer and thinner, while they have a more pronounced comb, indicating a higher hormonal background (Su et al., 2014). Also, ETB have a more developed lower and posterior part of the trunk, where the reproductive organs and the digestive tract are located (Williams, Sharp, 1978), with a characteristic convex breast (Tyasi et al., 2020). In addition, ETB tend to have a more

developed tail, especially in roosters, as well as longer legs (He et al., 2019; Valentim et al., 2019). In addition, the following are most often used in ETB as exterior indicators: head shape, comb shape, length and breadth of the neck and back, abdomen volume, shape and plumage of the tail.

Determining the development of muscles in the breast area is of great importance both for breeding and for assessing marketable qualities of meat-type poultry, i.e., broilers, turkeys, geese and ducks (Kochish et al., 2004). These indicators also include breast angle, or pectoral angle (PA), which began to be used in the 1940s as an indicator of meat quality and a characteristic of the pectoral muscle development (Poley et al., 1940; Dolecek et al., 1941; Asmundson, Lerner, 1951). To date, PA continues to be used only in poultry meat production (e.g., Kochish et al., 2004; Miguel et al., 2008; Hou et al., 2013; Li et al., 2015; Mikryukova, 2020), as well as for assessing certain physiological characteristics in some other bird species (Millán et al., 2003; Mougeot et al., 2004; Villanúa et al., 2007; Irvine et al., 2007).

In the present study, we set a goal of assessing phenotypic traits and possible relationships between them, including indicators of egg production and BW, as well as various body measurements. The objects of the study were various chicken breeds and populations from the RIFAGB bioresource collection (Paronyan et al., 2014), phenomes of which have been created and shaped under selection for different breeding characteristics. Furthermore, since conformational indicators, including PA, may have a certain potential for a deeper analysis and subsequent selection, we have searched here for correlations between these indicators and the egg productivity in breed flocks of different performance, utility and phenotypes. Herewith, additional information has been obtained regarding PA as an indicator for discriminating the entire spectrum of chicken breeds and populations studied.

Materials and methods

Ethics statement

All experiments were conducted with an ethical approval of the RIFAGB – Branch of the L. K. Ernst Federal Research Center for Animal Husbandry (Protocol No. 2020-4 dated 3 March 2020). All procedures involving chickens implied no physical harm.

Characteristics of phenotypic features in the studied chicken breeds

To assess the phenotypic diversity, the core of the RIFAGB bioresource collection (Paronyan et al., 2014) was examined including 39 chicken breeds and populations, including synthetic ones, that are kept in floor pens, at a sex ratio of one male to eight females, and fed the complete compound feed PK1-1 (Gatchina Feed Mill). The same environmental conditions were used to maintain all chicken breed flocks. According to TCM, these breeds represent different types of productivity and utility of poultry breeds, i.e., ETB, MTB, dual purpose (EMB and MEB), GB and FB (Bogolyubsky, 1991; Larkina et al., 2021) (Tables 1 and S1). Some of them carry notable mutations at certain chicken genes, e.g., *GHR* in RWD (Burnside et al., 1992) and *Va* in NN (Desta et al., 2021).

Sampling of phenotypic traits

Values of morphometric parameters, including BW and 13 body measurements of 330-day-old hens and roosters, as well as egg number (EN) at 52 weeks of life and EW were collected in the studied breeds and populations in 2016 for all genotypes (Vakhrameev, 2021; Tables 1, S2 and S3). A descriptive and comparative assessment of the variation in these phenotypic traits was carried out using Microsoft Excel. Also, an integral indicator of egg mass yield (EMY) was used as an index of egg productivity, which estimated the total mean weight of eggs per population laid during the study period and, accordingly, was calculated as the product of EW and EN. Usage of the EMY index rather than EN is known to provide better comparisons of flocks and strains of birds in terms of layer production (TNAU, 2020).

The following individual body measurements were sampled with a compass and a tape measure: body length (cm), body slanting length (cm), body and neck length (cm), keel length (cm), chest girth (cm), chest depth (cm), PA (°), distance between shoulder joints (cm), distance between hip joints (cm), femur length (cm), tibia length (cm), shank length (cm), and shank girth (cm). The

techniques of taking these measurements, including PA, and their values in various breeds, have been most fully described in our monograph "Exterior Assessment of Chickens" (Vakhrameev et al., 2021). In particular, PA (in degrees) was measured with a special device called a protractor or goniometer (Kochish et al., 2004). This tool was applied perpendicular to the pectoral muscle at a distance of 1 cm from the edge of the sternum keel (Fig. 1; Erigina et al., 2015; Mikryukova, 2020).

When looking for the relationship between the obtained phenotypic parameters and performance data, we proceeded from the grouping of 39 breeds and chicken populations in accordance with TCM and PCM. For a more generalised comparison, we also combined data for those types of breeding flocks that were not subject to targeted selection for increased egg or meat production. These included chickens of FB, GB and Bantam type breeds (BTB).

Mathematical and statistical analyzes

Basic statistical parameters (mean, standard deviation, significance, and correlation coefficients) and graphical dependences for the experimental data obtained as a result of analytical measurements were calculated using standard MS Excel applications.

Mathematical approximation of graphical dependencies was performed using standard MS Excel applications as well as an advanced analytics software package STATISTICA 5.5 (StatSoft, Inc./TIBCO, Palo Alto, CA, USA).

Results and discussion

Overall characteristics of the main phenotypic (performance) traits. As a result of this study, the values of phenotypic traits were obtained for adult birds of both sexes (Tables 1, S2 and S3). Essential variation was observed for BW, 13 main body measurements, 52-week EN, and EW. Overall, the mean BW of females was 2.23 ± 0.80 kg, and that of males 2.89 ± 0.98 kg; the mean EN was 149.55 ± 27.67 eggs, and the mean EW was 57.14 ± 4.60 g (Tables 1, S2 and S3; see also Supporting Info (SI) S1–S4). The greatest linear correlation was found between EN and EW, i.e., at r = 0.57 (p < 0.001; SI S1). BW correlated significantly with EN, the Pearson correlation coefficient between EN and female BW being 0.31 (p < 0.001; SI S2) and that between EN and a sum of male

and female BW 0.30 (p < 0.001; SI S3), while there was no significant correlation between EN and male BW 0.28 (p > 0.05; SI S3). Linear correlation plots for pairs of compared traits EN – EW, EN – female BW, and EN – summed male and female BW are shown in Figs. S1a,b,c. Significant correlation was also observed between EW and BW (0.55–0.62; p < 0.001; SI S4).

It is noteworthy that the graphs in Figs. S1a,b,c showed some common patterns in the distribution of breeds and populations. In particular, there was an isolated single position of some populations, e.g., MTB chickens (population that consisted of three-way crossbreds based on WC, i.e., WC × (BL × SL), and bred *inter se*), RWD (a parental strain from a commercial broiler cross), SW, and CB that belong to BTB. On the other hand, formation of fairly stable groupings of closely localised populations can be noted, e.g., LLB, MB and Aurora Blue (AB); Poland White-crested Black (PWB) and two varieties of the newly restored Pavlov breed (PS and PW; Paronyan et al., 2014); Leningrad Golden-and-grey (LGG) and Leningrad Mille Fleur (LMF); Plymouth Rock Barred (PRB) and Pantsirevka Black (PB); RC and Pervomai (Pm); etc. The further characterization of the breeds and populations by phenotypic traits observed using more sophisticated mathematical approaches are discussed in more detail in the section below.

Body measurements with a focus on pectoral angle. We explored the data of conformation indicators (i.e., body measurements) for females and males, corresponding to different types of utility and performance when grouping the studied breeds and populations according to PCM, as presented in Tables S2 and S3, respectively. Since the population $WC \times (BL \times SL)$ used as an MTB representative in our investigation was an obvious outlier and stood out in terms of the characterised phenotypic traits, we further looked for possible interbreed differences in body measurements for the rest breed groups (i.e., other than MTB). Additionally, we combined FB, BTB and GB into a larger group of non-productive breeds, i.e., non-selected for egg or meat productivity, to see how phenomic features driven by selection could correlate with performance traits.

As shown by the analysis of correlations between egg productivity assessed by EMY, on the one hand, and the exterior traits, on the other, in females of ETB and EMB types, the closest

relationship was observed for PA (r = 0.974 and 0.909, respectively; p < 0.01). Because of that, we decided to focus on this featured indicator in further analysis, and the respective results of the correlation relationship were presented in the form of graphical dependencies (Fig. 2). Herein, we found out some decreasing tendency in the correlation coefficient for females of the MEB type (r = 0.303; p < 0.3) and breeds that have never been selected for the purpose of increased egg production (r = -0.406; p < 0.25). This suggested to take a closer look at the indicator of PA, and a similar tendency was observed in males using the same subdivision into productive types.

Since this conformational indicator was developed to assess the meat quality of poultry (Asmundson, Lerner, 1951), we considered it necessary to assess the relationship between PA and BW, both hens and roosters. The results of the assessment are presented in Fig. 3.

To assess the closeness of this relationship and its possible mathematical recalculation for the PA value depending on BW, the obtained linear dependences were approximated by the following formulae:

$$PA_{F} = 5.272BW_{F} + 65.805, \qquad (1)$$

$$r = 0.612, \qquad (1)$$

$$p < 0.01; \qquad (2)$$

$$r = 0.608, \qquad (2)$$

$$r = 0.608, \qquad (2)$$

where the respective subscript 'F' or 'M' was assigned to the characteristics of females and males.

The mathematical model that includes the dependencies (1) and (2) quite accurately describes the relationship between PA and BW of birds within the framework of studying various breeds. Herewith, the correlation estimates were practically the same for both females and males, suggesting that the resulting equations reflect a plausible approximate estimation of these parameters in a breed.

As evidenced by the obtained relationships (1) and (2), these two parameters, PA and BW, are rather closely interrelated with each other, and therefore, the dependencies shown in Fig. 2 could only be explained by the fact that individuals in breeds under egg production-oriented selection are heavier than others. Although the data in Tables S2 and S3 do not confirm this trend, we decided to reduce the influence of the bird's BW to zero, for which we introduced the specific PA indicator (i.e., PA/BW) into the analysis, for which the angle values were divided by the bird's BW.

A similar analysis showed the same trend (Fig. 4) that was obtained for PA alone (Fig. 2), although the values of the correlation coefficients decreased slightly.

Undoubtedly, the specific index PA/BW proposed by us deserved a further closer study, and therefore, we tried to assess variation of egg productivity indicator, EMY, within each of the analysed groups, depending on PA/BW value (Fig. 5).

Next, we also evaluated which indicator of egg production, i.e., mean EN or EW, is more influenced by the PA/BW value. The results by group are presented in Fig. 6.

Additionally, the intra-breed correlation between PA and BW was assessed in our study. The correlation coefficient for males was 0.1–0.2 higher than that for females, i.e., this correlation was at the level of 0.25–0.3 for hens and 0.3–0.4 for roosters. In productive breeds, the correlation was higher than in non- productive ones. In some productive breeds, a very high correlation was found. Particularly, in ZS chickens (43 head in 2017), the correlation between BW and PA was 0.624 (Vakhrameev, Makarova, 2021).

Discussion

Correlation between performance traits

It is well known that EN is negatively, and EW is positively, correlated with BW. However, it should be borne in mind that, as a rule, these correlation estimates are strongly influenced by the estimated population as well as the method used (Szwaczkowski, 2003). Thus, when comparing EN and BW in different populations of layers, the correlation coefficient can vary, e.g., from -0.10 to 0.01 (Anene et al., 2020) to 0.12 (Islam et al., 2013), although these values were insignificant in the

given examples. In our study, we found a significant correlation between these indicators at the level of ~0.30 (SI S2 and S3). At the same time, the correlation between BW and EW, according to other authors, can vary from 0.23–0.33 (Haq et al., 2011) to 0.28–0.36 (Anene et al., 2020) and up to 0.67 (Islam et al., 2013). In our current investigation, these indicators correlated within 0.55–0.62 (SI S4). The variation in the correlation between EN and EW can range from –0.03 (an insignificant value; Anene et al., 2020) to 0.42 (Islam et al., 2013), while we observed the correlation of these indicators equal to 0.57 among 39 most diverse chicken breeds and populations (SI S1).

Pectoral angle in breeding and selection

For the most part, in this study, we were interested in examining PA, which is an ambiguous phenotypic indicator. In poultry meat production, PA as measured (in degrees) with a protractor/goniometer is used for assessing the extent of pectoral muscle development in young poultry at slaughter age. This body measurement characterises meat productivity (Mikryukova, 2020), with the heritability coefficient for PA being high ($h_a^2 = 0.45-0.47$; Le Bihan-Duval et al., 1997; Szwaczkowski, 2003). However, there are few analytical articles among current publications regarding various chicken exterior measurements, and we have not come across detailed studies on the PA indicator. Recently, studies on body measurements in poultry were carried out at the Poultry Research and Technological Institute, Sergiyev Posad, Moscow Oblast (Egorova, 2016, 2017, 2018; Egorova et al., 2018: Grishina, Zharkova, 2020), but there were no published data on PA.

It is clear that, in meat poultry production, the level of development of the pectoral muscles is almost the main significant trait for selection. In this regard, there was an interesting example of the breeding of Brwinów synthetic strain of meat-type chickens in Poland obtained as a result of crossing WC that had dominant and recessive white colour with White Leghorns and Rhode Island Reds and further selection for conformation, especially for PA (Somes, 1988). At the same time, it is difficult to deny that the development of the pectoral muscles is an indicator of not only meat content, but also the health of a bird. For instance, due to some diseases the birds can "dry up" and such sick individuals are called "rusks" in veterinary practice. When touching the keel of a bird with completely absent pectoral muscles, it is relatively easy to appreciate this term (Vakhrameev, 2021).

In 2007, at the Department of Poultry, St. Petersburg State Agrarian University, a small research project was conducted to assess chicken body measurements using appropriate tools. Among them, the goniometer was characterised by the department staff as a barely useful tool because there was no clear relationship between PA and productive traits. However, it should be noted that only a small number of birds were sampled in that investigation. This might explain why no clear pattern in various breeds was found (Vakhrameev, 2021). At RIFAGB since 2018, when it was necessary to measure a larger number of birds, only basic measurements (seven in total) and BW were measured; indeed PA was not measured in that study (Vakhrameev, 2021; Vakhrameev, Makarova, 2021). This indicator was included in further studies when a complete set of information on the examined birds was collected (Makarova, Vakhrameev, 2020; Vakhrameev, Makarova, 2021). The correlation of PA with BW studied in our own investigations showed quite predictable results, with a slightly higher correlation coefficient in males than females. This is understandable since roosters grow disproportionately in the breast area, while hens do so in the abdominal area (Vakhrameev, 2021; Vakhrameev, Makarova, 2021).

Thus, it can be stated that PA has been under-studied in terms of the methodological aspects of evaluating the phenotype of layer birds (e.g., Nyalala et al., 2021). However, our analytical assessment of the conformational performance of hens and roosters of breeding flocks and their relationship with egg productivity shows that such an omission is unwise (as discussed in the section below).

Pectoral angle implications in egg-type chickens

There is some benefit in exploring several traits jointly in genetic evaluations. In particular, based on sex-limited characteristics of egg production, information about selected layers is available only after the entire period of their production test, while for selection candidate males it is generally impracticable to obtain directly. Thereby, layer breeding programs may need to account for, and

use, indirect indicators from these selection candidates, such as conformation or PA (Besbes, Ducrocq, 2003).

In the general practice of layer breeding, PA, can be considered as an indirect indicator of development of egg producing hens. In other words, the underdevelopment of the pectoral muscles is inferior, but the overdeveloped pectoral muscles seem to detract from a part of the layer vitality. It is difficult to say how much specialists in layer breeding are guided by the PA value. Most often, scoring of hens is carried out by individual inspection of the abdomen, manual measuring of distance between the pubic bones, and subsequent scoring of a layer (Smith, 2015; Vakhrameev, 2021).

The results of our investigation, when grouping the studied breeds and populations (except one MTB population) into four major production-based groups, have shown the effectiveness of the PA trait, which suggests the possible selection of individuals of both sexes when breeding and/or culling in egg-type breeder flocks. Since our investigation was carried out on breeder flocks, we believe that the genetic contribution of males to productive traits is no less important than that of females (as also demonstrated in our previous study; Larkina et al., 2021). As evidenced by the data of the graphical dependencies in Fig. 2, the more the utility/purpose of breeder flocks approaches the egg type, the higher the relationship of PA with egg production.

Similar to MTB (Kochish et al., 2004; Miguel et al., 2008; Mikryukova, 2020), there is a closer correlation between the PA and BW values in both sexes in ETB and non-productive flocks (Fig. 3), which are described by linear dependences (Eqns1,2). In this regard, we suggest using a slightly modified specific indicator, the ratio of PA to BW (i.e., PA/BW). In our opinion, it might more adequately characterise an individual/strain in terms of its potential for the increased EMY. The graphical dependencies in Fig. 4 suggest the validity of this premise.

We obtained very similar data as in the graphs in Fig. 4 when analysing egg productivity (i.e., EMY, or the total weight of eggs laid during the estimated period) using the four types of breeding flocks we considered (Fig. 5). For the ETB and EMB types, there was a corresponding tendency to

increase EMY with an increase in PA/BW. However, for the MEB type and especially for nonproductive breeds, this tendency changed in the opposite direction. This reaffirms the notion that, in a flock where there has been no breeding for increased egg productivity, PA might not bear any positive information in terms of assessing an individual for its egg laying potential. Nonetheless, the PA usefulness could be more appropriate in flocks, like ETB, selected for egg productivity.

It should be noted that, in the case of chickens that are not classified as the ETB, EMB, MEB or MTB types, a strong selection bias towards ornamental or other special (fancy) traits could lead to an almost complete lack of attention to productive traits and associated body measurements, as suggested by the results of our study.

Our attempts to assess which particular indicator of egg production, EN or EW, depends to a greater extent on the PA/BW index (Fig. 6), demonstrated that in egg-oriented breeding flocks, the correlation with PA/BW is due precisely to the trend towards an increase in EN, not EW. Presumably, this observation can be explained by the fact that in this case the factor of a denser conformation with an overall small BW can play a role as some authors claim (OOO-VetCentr, 2021). This possible reason is also confirmed by other relevant studies, e.g., in managing indigenous chicken genetic resources (Dana et al., 2010) or breeding pheasants (Kırıkçı et al., 2004).

Phenomic studies in chickens

Based on the above studies, we feel that research on phenomics in chickens should be given more attention when implementing measures to preserve the gene pool of various breeds and strains (Romanov, Weigend, 2001; Weigend et al., 2004a). These should embrace those that have been created using different, often contrasting production-oriented, selection paths and include synthetic ones. Particularly, if there is a limited opportunity to apply high throughput molecular genetic methods, measurements of phenotypic traits become few that could be produced (Weigend et al., 2004a,b; Paronyan, 2016; Vakhrameev, 2021). At RIFAGB, a wealth of information has been accumulated regarding body measurements obtained in the bioresource collection in 2007 and 2017,

as well as in the last three years, when searching for the indicators of growth rate in the first four months of chicken life (Vakhrameev, 2021). The time has come to analyze the generated phenotypic data and, if possible, associate it with certain indicators of productivity and genotypic variability as undertaken in our recent studies (Larkina et al., 2021; Romanov et al., 2021) and the current research herein presented.

The unconditional interest in phenomic studies and assessment of the conformation in chickens will serve as a good incentive for further work in this direction. In general, view of the state of the art is that, with the advent of powerful techniques in molecular genetics, genomics and computational biology, the possibilities have increased dramatically for elucidating the patterns associated with measurements of the external characteristics and phenomics of chickens as a whole. Undoubtedly, the use of methods for collecting extensive phenotypic data, as well as improved statistical and computational methodologies for the integration of various phenotypic traits (including emerging ones) will contribute to the expansion of selection-driven phenomics research in poultry and the development of new breeding strategies (Silva et al., 2021). Analysis of such phenotypic traits, as undertaken in this study, will facilitate their use as indicators of new traits relevant to the genetic improvement of poultry and livestock populations. This is especially pertinent considering modern and pressing phenotypes of economic and societal significance (Silva et al., 2021).

Conclusions

Within the framework of the pilot phenomic investigation on chickens in previous (Larkina et al., 2021) and current work, we have started the process of evaluating the phenotypic diversity in a wide range of commercial, local and fancy breeds selected for productive and non-productive traits. Many of these breeds are synthetic, and together they constitute a large sample of the global gene pool, the sustainable conservation of which ensures the food security of countries of the world (Zubets et al., 2007; Mel'nyk et al., 2009; Tereshchenko et al., 2015). As a result of our research, we propose a hypothesis that the greater the vector of selection pressure toward egg production

among breeds varying from lower to high laying performance, the closer the relationship between their productivity and PA may be. Furthermore, we suggest the ratio of PA to BW as a promising new index for selecting females and males in breeding flocks in order to improve egg production. Models of relationships and clustering by phenotypic traits observed in different breeds and populations need further analysis using more sophisticated mathematical analytical tools. Phenotyping techniques, especially high-throughput ones, demonstrate new potential pathways for genomic dissection of complex traits and genomic improvement in poultry and livestock, with attention to previously known and new selected phenotypes (Silva et al., 2021). The present study exemplifies their potential application, given these promising findings.

Author Contributions

Conceptualization, V.G.N., M.N.R. and A.B.V.; methodology, V.G.N., M.N.R., A.B.V. and Y.S.S.; software, V.G.N. and M.N.R.; validation, A.B.V. and T.A L.; formal analysis, V.G.N., M.N.R. and A.B.V.; investigation, G.K.P., A.V.M., A.B.V., T.A.L. and O.Y.B.; re-sources, O.I.S., A.B.V., A.V.M. and G.K.P.; data curation, O.I.S., A.B.V., N.V.D., M.V.P., G.K.P. and T.A.L.; writing—original draft preparation, V.G.N., A.B.V., M.N.R., Y.V.B. and A.P.D.; writing—review and editing, V.G.N., M.N.R. and D.K.G.; visualization, V.G.N., M.N.R. and A.B.V.; supervision, T.A.L.; project administration, T.A.L.; funding acquisition, T.A.L. All authors have read and agreed to the published version of the manuscript.

Data statement

The data presented in this study are available in this article and supplementary material.

Supplementary Materials

The following are available online at https://, Fig. S1: Microsoft Excel plots of the correlation dependences of 52-week egg production on egg weight (a), female body weight (b), and summed male and female body weights (c) among the 39 phenotyped chicken breeds/populations. Population codes are given according to Table S1. Egg weight and egg number appear highly correlated (a), unlike a much lower correlation in the case of body weight by egg number (b, c);

Table S1: Thirty-nine chicken breeds, strains and crosses used in the study and listed in accordance with TCM; Table S2: Phenotypic traits in females among the 39 studied chicken gene pool breeds/populations listed in accordance with PCM; Table S3: Phenotypic traits in males among the 39 studied chicken gene pool breeds/populations listed in accordance with PCM; Supporting Information (SI) S1: Statistics and correlations for egg weight and egg number; SI S2: Statistics and correlations for female body weight and egg number; SI S3: Statistics and correlations for male/female body weight and egg number; SI S4: Statistics and correlations for body weight and egg weight.

Declaration of Competing Interest

The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

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References

- Abdelmanova, A.S., Dotsev, A.V., Romanov, M.N., Stanishevskaya, O.I., Gladyr, E.A., Rodionov,
 A.N., Vetokh, A.N., Volkova, N.A., Fedorova, E.S., Gusev, I.V., Griffin, D.K., Brem, G.,
 Zinovieva, N.A., 2021. Unveiling comparative genomic trajectories of selection and key
 candidate genes in egg-type Russian White and meat-type White Cornish chickens. Biology 10,
 876. https://doi.org/10.3390/biology10090876.
- AG2PI, 2021. Agricultural Genome to Phenome Initiative. https://web.archive.org/web/20210928205023/https://www.ag2pi.org/ (accessed 26 October 2021).

- Amar, D., Sinnott-Armstrong, N., Ashley, E.A., Rivas, M.A., 2021. Graphical analysis for phenome-wide causal discovery in genotyped population-scale biobanks. Nat. Commun. 12, 350. https://doi.org/10.1038/s41467-020-20516-2.
- Anene, D.O., Akter, Y., Thomson, P.C., Groves, P., O'Shea, C.J., 2020. Variation and association of hen performance and egg quality traits in individual early-laying ISA Brown pens. Animals 10, 1601. https://doi.org/10.3390/ani10091601.
- Arzhankova, Yu.V., Ivanova, T.N., 2012. [Exterior, Interior and Constitution of Poultry: Scientific Manual on the Discipline "Biology of Poultry"]. Publishing House of the Federal State Budgetary Educational Institution of Higher Professional Education "Velikie Luki State Agricultural Academy", Velikie Luki.
- Asmundson, V.S., Lerner, I.M., 1951. Breeding Chickens for Meat Production. Agricultural Experiment Station, Berkeley, CA, USA.
- Besbes, B., Ducrocq, V., 2003. Use of mixed model methodology in breeding strategies for layers, in: Muir, W.M., Aggrey, S.E. (Eds.), Poultry Genetics, Breeding and Biotechnology. CAB International, Wallingford, Oxon, UK; Cambridge, MA, USA, pp. 130–131. https://doi.org/10.1079/9780851996608.0165.
- Bogolyubsky, S.I., 1991. [Poultry Breeding]. Agropromizdat, Moscow.
- Bondarenko, Yu.V., Zharkova, I.P., Romanov, M.N., 1986. [Study of down colour genotype in the collection flock geese at the All-Union Poultry Research and Technological Institute]. Naučno-Tehničeskij Bûlleten Institut Pticevodstva. Poultry Research Institute, Ukrainian Academy of Agrarian Sciences, Kharkiv, Ukraine, No. 34, 3–7.
- Bondarenko, Yu.V., Sakhatsky, N.I., Kutnyuk, P.I., Romanov, M.N., 1995. Screening inherited abnormalities in poultry populations, in: Proceedings of the 11th International Symposium "Current Problems in Avian Genetics", Balice near Krakow, Poland, 29 May –1 June 1995. World's Poultry Science Association, Polish Branch, Balice near Krakow, Poland, 196–199.

- Bondarenko, Yu.V., Breslavets, V.A., Kuchmistov, V.A., Romanov, M.N., Ivanova, T.V., Kutnyuk,
 P.I., 1996. Homological variation of embryonic abnormalities in poultry, in: Proceedings of the XX World's Poultry Congress, Abstracts and Students' Papers Contributed in International Youth Programme, New Delhi, India, 2–5 September 1996. World's Poultry Science Association (India Branch), New Delhi, India, 4, 94.
- Borisenko, E.Ya., 1966. [Breeding of Farm Animals], 4th ed., rev. and add. Moscow: Kolos.
- Burnside, J., Liou, S.S., Zhong, C., Cogburn, L.A., 1992. Abnormal growth hormone receptor gene expression in the sex-linked dwarf chicken. Gen. Comp. Endocrinol. 88, 20–28. https://doi.org/10.1016/0016-6480(92)90190-u.
- Corti, E., Moiseyeva, I.G., Romanov, M.N., 2010. Five-toed chickens: their origin, genetics, geographical spreading and history. Izv. Timiryazev S.-Kh. Akad. [Proceedings of the Timiryazev Agricultural Academy] Special Issue 7, 156–170.
- Dana, N., van der Waaij, L.H., Dessie, T., van Arendonk, J.A., 2010. Production objectives and trait preferences of village poultry producers of Ethiopia: implications for designing breeding schemes utilizing indigenous chicken genetic resources. Trop. Anim. Health. Prod. 42, 1519–1529. https://doi.org/10.1007/s11250-010-9602-6.
- Dehghanzadeh, H., Mirhosseini, S.Z., Romanov, M.N., Ghorbani, A., 2009. Evaluation of genetic variability and distances among five Iranian native chicken populations using RAPD markers. Pak. J. Biol. Sci. 12, 866–871. https://doi.org/10.3923/pjbs.2009.866.871.
- Dementeva, N.V., Romanov, M.N., Kudinov, A.A., Mitrofanova, O.V., Stanishevskaya, O.I., Terletsky, V.P., Fedorova, E.S., Nikitkina, E.V., Plemyashov, K.V., 2017. Studying the structure of a gene pool population of the Russian White chicken breed by genome-wide SNP scan. Sel'skokhozyaistvennaya Biol. [Agric. Biol.] 52, 1166–1174. https://doi.org/10.15389/agrobiology.2017.6.1166eng.
- Dementeva, N.V., Kudinov, A.A., Mitrofanova, O.V., Stanishevskaya, O.I., Fedorova, E.S., Romanov, M.N., 2018. Genome-wide association study of reproductive traits in a gene pool

breed of the Russian White chickens. Reprod. Domest. Anim. 53(Suppl. 2), 123–124. https://doi.org/10.1111/rda.13300.

- Dementieva, N.V., Mitrofanova, O.V., Kudinov, A.A., Smaragdov, M.G., Yakovlev, A.F., Romanov, M.N., 2018. [Opportunities of SNP-genotyping for studying the features of the genetic architecture of chicken populations with different histories], in: [World and Russian Trends in Development of Poultry Industry: Realities and Challenges of Future]: Proceedings of the 19th International Conference, Sergiyev Posad, 15–17 May 2018. WPSA, Poultry Research Centre, Poultry Science and Technology Institute, Sergiyev Posad, pp. 80–81.
- Desta, T.T., 2021. The genetic basis and robustness of naked neck mutation in chicken. Trop. Anim. Health Prod. 53, 95. https://doi.org/10.1007/s11250-020-02505-1.
- Dolecek, R.L., Wilson, W.O., Poley, W.E., 1941. Physical measurements of carcass quality in roasters. Poult. Sci. 20,161–170. https://doi.org/10.3382/ps.0200161.
- Egorova, A.V., 2016. [Management of cocks in broiler parental flocks]. Pticevodstvo No. 3, 2-6.
- Egorova, A.V., 2017. [The principal directions in selection of broiler breeder females]. Pticevodstvo No. 3, 16–21.
- Egorova, A.V., 2018. [The exterior traits in broiler breeders]. Pticevodstvo No. 7, 9–11.
- Egorova, A.V., Emanuylova, Zh.V., Efimov, D.N., Tuchemsky, L.I., 2018. [The evaluation of broiler breeders of parental lines for growth rate]. Pticevodstvo No. 6, 8–13.
- Erigina, R.A., Popova, L.A., Komarchev, A.S., 2015. [Poultry: Workbook]. Publishing house of the RSAU Moscow Agricultural Academy named after K.A. Timiryazev, Moscow.
- Furbank, R.T., Tester, M., 2011. Phenomics technologies to relieve the phenotyping bottleneck. Trends Plant Sci. 16, 635–644. https://doi.org/10.1016/j.tplants.2011.09.005.
- Grishina, D.S., Zharkova, I.P., 2020. [Comparative assessment of geese of gene pool herd by their exterior]. Vladimirskij zemledelec [Vladimir Agricolist] No. 3(93), 64–68. https://doi.org/10.24411/2225-2584-2020-10136.

- Haq, R., Haq, E., Khan, M.F., 2011. Correlation between body weight and egg weight of Dokki and Fayoumi hen in Pakistan. J. Basic. Appl. Sci. 7, 165–168. https://doi.org/10.6000/1927-5129.2011.07.02.14.
- He, Y., Chang, L., Guan, X., Geng, L., Lu, C., Li, X., 2019. Comparison of nutritional components and egg quality between Bashang long tail laying hens and Hy-Line brown laying hens in cage. Chin. J. Anim. Nutr. 31, 3535–3542.
- Hou, X., Han, R., Tian, Y., Xie, W., Sun, G., Li, G., Jiang, R., Kang, X., 2013. Cloning of TPO gene and associations of polymorphisms with chicken growth and carcass traits. Mol. Biol. Rep. 40, 3437–3443. https://doi.org/10.1007/s11033-012-2421-2.
- Irvine, R.J., Leckie, F., Redpath, S.M., 2007. Cost of carrying radio transmitters: a test with racing pigeons Columba livia. Wildlife Biol. 13, 238–243. https://doi.org/10.2981/0909-6396(2007)13[238:COCRTA]2.0.CO;2.
- Islam, S.S., Paul, C., Sarker, B.C., 2013. A comparative study on the performances of layer hybrids in some selected areas of Khulna region. Bangladesh J. Anim. Sci. 42, 114–122. https://doi.org/10.3329/bjas.v42i2.18491.
- Kırıkçı, K., Çetin, O., Günlü, A., Garip, M., 2004. Effect of hen weight on egg production and some egg quality characteristics in pheasants (Phasianus colchicus). Asian Austral. J. Anim. Sci. 17, 684–687. https://doi.org/10.5713/ajas.2004.684.
- Kochish, I.I., Petrash, M.G., Smirnov, S.B., 2004. [Poultry Farming: Textbook]. KolosS, Moscow. ISBN 5-9532-0038-2.
- Kulibaba, R., Tereshchenko, A., 2015. Transforming growth factor β1, pituitary-specific transcriptional factor 1 and insulin-like growth factor I gene polymorphisms in the population of the Poltava clay chicken breed: association with productive traits. Agric. Sci. Pract. 2, 67–72. https://doi.org/10.15407/agrisp2.01.067.
- Landauer ,W., 1941. The hatchability of chicken eggs as influenced by environment and heredity. Bull. Storrs Agric. Exp. Stn. University of Connecticut Publication, Storrs, CT, USA, No. 124.

- Larkina, T.A., Barkova, O.Y., Peglivanyan, G.K., Mitrofanova, O.V., Dementieva, N.V., Stanishevskaya, O.I., Vakhrameev, A.B., Makarova, A.V., Shcherbakov, Y.S., Pozovnikova, M.V., Brazhnik, E.A., Griffin, D.K., Romanov, M.N., 2021. Evolutionary subdivision of domestic chickens: implications for local breeds as assessed by phenotype and genotype in comparison to commercial and fancy breeds. Agriculture 11, 914. https://doi.org/10.3390/agriculture11100914.
- Le Bihan-Duval, E., Beaumont, C., Colleau, J.J., 1997. Estimation of the genetic correlations between twisted legs and growth or conformation traits in broiler chickens. J. Anim. Breed Genet. 114, 239–259. https://doi.org/10.1111/j.1439-0388.1997.tb00510.x.
- Lee, M.O., Romanov, M.N., Plemyashov, K.V., Dementieva, N.V., Mitrofanova, O.V., Barkova, O.Y., Womack, J.E., 2017. Haplotype structure and copy number polymorphism of the betadefensin 7 genes in diverse chicken breeds. Anim. Genet. 48, 490–492. https://doi.org/10.1111/age.12552.
- Li, H., Wang, S., Yan, F., Liu, X., Jiang, R., Han, R., Li, Z., Li, G., Tian, Y., Kang, X., Sun, G., 2015. Effect of polymorphism within miRNA-1606 gene on growth and carcass traits in chicken. Gene 566, 8–12. https://doi.org/10.1016/j.gene.2015.03.037.
- Makarova, A.V., Vakhrameev, A.B., 2020. The development intensity and exterior effect on meat productivity of the TsS experience chicken population. Ptica i pticeprodukty [Poultry and Poultry Products] No. 2, 28–31.
- Matalonga, L., Hernández-Ferrer, C., Piscia, D., Solve-RD SNV-indel working group, Schüle, R., Synofzik, M., Töpf, A., Vissers, L.E.L.M., de Voer, R., Solve-RD DITF-GENTURIS, et al., 2021. Solving patients with rare diseases through programmatic reanalysis of genome-phenome data. Eur. J. Hum. Genet. 29, 1337–1347. https://doi.org/10.1038/s41431-021-00852-7.
- Mel'nyk, Y.F., Mykytyuk, D.M., Bilous, O.V., Kudryavs'ka, N.V., Burkat, V.P., Huzyev, I.V., Podoba, B.Y., Sharan, P.I., Kovtun, S.I., Platonova, N.P., et al., 2009. [Program of Preservation

of the Gene Pool of Main Types of Farm Animals in Ukraine for the Period Till 2015]. Aristey, Kyiv, Ukraine.

- Miguel, J.A., Ciria, J., Asenjo, B., Calvo, J.L., 2008. Effect of caponisation on growth and on carcass and meat characteristics in Castellana Negra native Spanish chickens. Animal 2, 305–311. https://doi.org/10.1017/S1751731107001127.
- Mikryukova, O.S., 2020. [Poultry Farming: Teaching Aid]. Federal State Budgetary Educational Institution of Higher Education "Perm Agrarian and Technological University named after Academician D.N. Pryanishnikov"; IPC "Prokrost", Perm. ISBN 978-5-94279-490-3.
- Millán, J., Gortázar, C., Buenestado, F.J., Rodríguez, P., Tortosa, S.F., Villafuerte, R., 2003. Effects of a fiber-rich diet on physiology and survival of farm-reared red-legged partridges (Alectoris rufa). Comp. Biochem. Physiol. A Mol. Integr. Physiol. 134, 85–91. https://doi.org/10.1016/s1095-6433(02)00189-7.
- Moiseyeva, I., Romanov, M., Pigaryev, N., 2000. Sergey Petrov Obituary. Worlds Poult. Sci. J. 56, 437–438.
- Moiseyeva, I.G., Romanov, M.N., Nikiforov, A.A., Sevastyanova, A.A., Semyenova, S.K., 2003. Evolutionary relationships of Red Jungle Fowl and chicken breeds. Genet. Sel. Evol. 35, 403– 423. https://doi.org/10.1186/1297-9686-35-5-403.
- Moiseyeva, I.G., Kovalenko, A.T., Mosyakina, T.V., Romanov, M.N., Bondarenko, Y.V., Kutnyuk, P.I., Podstreshny, A.P., Nikiforov, A.A., Tkachik, T.E., 2006. [Origin, history, genetics and economic traits of the Poltava chicken breed]. Elektronnyi zhurnal [Electronic Journal]. Laboratory of Animal Comparative Genetics, N.I. Vavilov Institute of General Genetics, Moscow, Issue 4. https://web.archive.org/web/20120205195904/http://www.lab-cga.ru/articles/Jornal04/Statia2.htm (accessed 26 October 2021).
- Moiseyeva, I.G., Nikiforov, A.A., Romanov, M.N., Aleksandrov, A.V., Moysyak, Y.V., Semyenova, S.K., 2007a. [Origin, history, genetics and economic traits of the Yurlov Crower chicken breed]. Elektronnyi zhurnal [Electronic Journal]. Laboratory of Animal Comparative

Genetics, N.I. Vavilov Institute of General Genetics, Moscow. https://web.archive.org/web/20120210170800/http://www.lab-

cga.ru/articles/Yurlovskaya/Yurlovskaya.htm (accessed 26 October 2021).

- Moiseyeva, I.G., Romanov, M.N., Kovalenko, A.T., Mosyakina, T.V., Bondarenko, Yu.V., Kutnyuk, P.I., Podstreshny, A.P., Nikiforov, A.A., 2007b. The Poltava chicken breed of Ukraine: its history, characterization and conservation. Anim. Genet. Resour. Inf. 40, 71–78. https://doi.org/10.1017/S1014233900002212.
- Moiseyeva, I.G., Corti, E., Romanov, M.N., 2009a. [Polydactyly in chickens], in: [Advances in Modern Poultry Science]: Proceedings of the 16th International Conference, Sergiyev Posad, 19–21 May 2009. WPSA, RAAS, Poultry Science and Technology Institute, Sergiyev Posad, 51–53.
- Moiseyeva, I.G., Romanov, M.N., Aleksandrov, A.V., Nikiforov, A.A., Sevastyanova, A.A., 2009b. [Evolution and genetic diversity of old domestic hen's breed–Yurlovskaya Golosistaya: system analysis of variability forms]. Izv. Timiryazev S.-Kh. Akad. [Proceedings of the Timiryazev Agricultural Academy] Issue 3, 132–147.
- Moiseyeva, I.G., Sevastyanova, A.A., Aleksandrov, A.V., Romanov, M.N., 2011. [Singing breeds of hens]. Priroda Issue 4(1148), 10–18.
- Moiseyeva, I.G., Avrutskaya, T.B., Romanov, M.N., 2012a. [A.S. Serebrovsky, founder of chicken genetics studies], in: [Innovation Developments and Their Implementation in Poultry Industry]: Proceedings of the 17th International Conference, Sergiyev Posad, 15–17 May 2012. WPSA, Poultry Research Centre, Sergiyev Posad, pp. 85–88.
- Moiseyeva, I.G., Romanov, M.N., Nikiforov, A.A., Avrutskaia, T.B., 2012b. [Studies in chicken genetics: commemorating the 120th anniversary of the outstanding Soviet geneticist A.S. Serebrovsky (1892–1948)]. Genetika 48, 1021–1038. PMID: 23113330.
- Moiseyeva, I.G., Avrutskaya, T.B., Romanov, M.N., 2013. [Studies in chicken genetics by A.S. Serebrovsky]. Dvorovaya zhivnost' i usad'ba Issue 1–2, 31–37.

- Moiseyeva, I.G., Sevastyanova, A.A., Aleksandrov, A.V., Vakhrameev, A.B., Romanov, M.N., Dmitriev, Y.I., Semenova, S.K., Sulimova, G.E., 2016. [Orloff chicken breed: history, current status and studies]. Izv. Timiryazev S.-Kh. Akad. [Proceedings of the Timiryazev Agricultural Academy] Issue 1, 78–96.
- Mougeot, F., Irvine, J.R., Seivwright, L., Redpath, S.M., Piertney, S., 2004. Testosterone, immunocompetence, and honest sexual signaling in male red grouse. Behav. Ecol. 15, 930-937. https://doi.org/10.1093/beheco/arh087.
- Nyalala, I., Okinda, C., Kunjie, C., Korohou, T., Nyalala, L., Chao, Q., 2021. Weight and volume estimation of poultry and products based on computer vision systems: a review. Poult. Sci. 100, 101072. https://doi.org/10.1016/j.psj.2021.101072.
- OOO-VetCentr,2021.OOO-VetCentr.ru,Moscow.https://web.archive.org/web/20211026174911/https://ooo-vetcentr.ru/kury/ot-chego-zavisit-produktivnost-kur-nesushek (accessed 26 October 2021).
- Paronyan, I.A., 2016. [Genetic Resources of Farm Animals]. Prospekt Nauki, St. Petersburg. ISBN 978-5-906109-41-5.
- Paronyan, I.A., Plemyashov, K.V., Segal, E.L., Yurchenko, O.P., Shabanova, S.A., Vakhrameev, A.B., Karpukhina, I.V., Makarova, A.V., Pervushina, A.T., 2014. [Breeds and Populations of Chickens Bred at the Germplasm Farm of the State Scientific Institution VNIIGRZh of the RAAS: Album]. GNU VNIIGRZh, St. Petersburg. https://web.archive.org/web/20180826232038/http://vniigen.ru/wp-content/uploads/2017/04/Katalog-Kur-1.pdf (accessed 26 October 2021).
- Poley, W.E., Moxon, A.L., Wilson, W.O., Dolecek, R.L., 1940. Effect of corn, wheat and barley in the diet on the physical and chemical composition of fryers and roasters. J. Agric. Res. 61, 161–178.
- Rexroad, C., Vallet, J., Matukumalli, L.K., Reecy, J., Bickhart, D., Blackburn, H., Boggess, M., Cheng, H., Clutter, A., Cockett, N., et al., 2019. Genome to phenome: improving animal health,

production, and well-being – a new USDA blueprint for animal genome research 2018–2027. Front. Genet. 10, 327. https://doi.org/10.3389/fgene.2019.00327.

- Romanov, M.N., 1992. [Population-morphological studies on domestic geese], in: Proceedings of the Ukrainian Conference of Young Scientists and Postgraduates on Poultry Problems, Borky, Kharkiv, Ukraine. Poultry Research Institute, UAAS, Borky, Kharkiv, Ukraine, pp. 8–9.
- Romanov, M.N., 1993. Phenetics of poultry populations, in: Abstracts of the 1st Ukrainian Poultry Conference, Borky, Simferopol, Ukraine. World's Poultry Science Association, Ukrainian Branch, Poultry Research Institute, UAAS, Borky, Simferopol, Ukraine, pp. 15–16, 123.
- Romanov, M.N., 1994. Using phenetic approaches for studying poultry populations under preservation and breeding, in: Gene Mapping, Polymorphisms, Disease Genetic Markers, Marker Assisted Selection, Gene Expression, Transgenes, Non-Conventional Animal Products, Conservation Genetics, Conservation of Domestic Animal Genetic Resources: Proceedings of the 5th World Congress on Genetics Applied to Livestock Production, Guelph, Canada, 7–12 August 1994. International Committee for World Congresses on Genetics applied to Livestock Production, University of Guelph, Guelph, Canada, 21, pp. 556–559.
- Romanov, M.N., 1995a. Genetic studies on geese of Ukraine and Russia. 3. Phenetic and phylogenetic approaches, in: Proceedings of the 10th International Symposium on Waterfowl, Halle (Saale), Germany, 26–31 March 1995. World's Poultry Science Association, Halle (Saale), Germany, pp. 429–432.
- Romanov, M.N., 1995b. Evolution of domestic geese. Reconstruction of microphylogenesis by methods of population phenetics, in: Gudina, A.N. (Ed.), New Investigations on Palearctic Geese. Zaporizhya Branch of the Ukrainian Ornithological Society, Zaporizhya State University, Zaporizhya, Ukraine, pp. 115–120.
- Romanov, M.N., Bondarenko, Yu.V., 1994a. Introducing Ukrainian indigenous poultry domestic ducks. Fancy Fowl 13(3), 13, 28.

- Romanov, M.N., Bondarenko, Yu.V., 1994b. Introducing the Ukrainian indigenous poultry the Poltava chickens. Fancy Fowl 14(2), 8–9.
- Romanov, M.N., Weigend, S., 1999. Genetic diversity in chicken populations based on microsatellite markers, in: Dekkers, J.C.M., Lamont, S.J., Rothschild, M.F. (Eds.), Proceedings of the Conference "From Jay Lush to Genomics: Visions for Animal Breeding and Genetics", Ames, IA, USA, 16–18 May 1999. Iowa State University, Department of Animal Science, Ames, IA, USA, p. 174.
- Romanov, M.N., Weigend, S., 2001. Using RAPD markers for assessment of genetic diversity in chickens. Arch. Geflugelkd. 65, 145–148.
- Romanov, M.N., Bondarenko, Yu.V., Veremeyenko, R.P., 1994. Conservation of Ukrainian domestic duck genetic resources, in: Abstracts of the 13th Ukrainian Conference of Young Poultry Scientists and Postgraduates, Borky, Ukraine, 12–14 October 1994. World's Poultry Science Association, Ukrainian Branch, Poultry Research Institute, UAAS, Borky, Ukraine, pp. 24–26.
- Romanov, M.N. Veremeyenko, R.P., Bondarenko, Yu.V., 1995. Conservation of waterfowl germplasm in Ukraine, in: Proceedings of the 10th International Symposium on Waterfowl, Halle (Saale), Germany, 26–31 March 1995. World's Poultry Science Association, Halle (Saale), Germany, pp. 401–414.
- Romanov, M.N., Weigend, S., Bondarenko, Yu.V., Podstreshny, A.P., Kutnyuk, P.I., Sakhatsky, N.I., 1999. Studies on poultry germplasm diversity and conservation in Ukraine, in: Proceedings of the Poultry Genetics Symposium, Mariensee, Germany, 6–8 October 1999. Working Group 3 of WPSA, Mariensee, Germany, p. 140.
- Romanov, M.N., Avrutskaya, T.B., Moiseyeva, I.G., 2012. [Life given to science. To 120th anniversary of A.S. Serebrovsky]. Priroda Issue 2(1158), 63–70.
- Romanov, M.N., Larkina, T.A., Barkova, O.Yu., Peglivanyan, G.K., Mitrofanova, O.V., Dementieva, N.V., Stanishevskaya, O.I., Vakhrameev, A.B., Makarova, A.V., Shcherbakov,

Y.S., Pozovnikova, M.V., Griffin, D.K., 2021. [Comparative Analysis of Phenotypic Traits in Various Breeds Representing the World Poultry Gene Pool], in: [Materials of the 3rd International Scientific and Practical Conference on Molecular Genetic Technologies for Analysis of Gene Expression Related to Animal Productivity and Disease Resistance], Moscow, 29 September 2021. Sel'skokhozyaistvennye tekhnologii, Moscow, pp. 52–63. https://doi.org/10.18720/SPBPU/2/z21-43.

- Silva, F.F., Morota, G., Rosa, G.J.M., 2021. Editorial: High-throughput phenotyping in the genomic improvement of livestock. Front. Genet. 12, 707343.
- Smith, T.W., 2015. Culling Hens. Extension Service of Mississippi State University, cooperating with U.S. Department of Agriculture, Mississippi State, MS, USA, Publication 358. https://web.archive.org/web/20191201204653/http://extension.msstate.edu/content/culling-hens (accessed 26 October 2021).
- Somes, R.G., Jr., 1988. International registry of poultry genetic stocks. Bull Storrs Agric Exp Stn. University of Connecticut Publication, Storrs, CT, USA, No. 476.
- Su, Y.J., Shu, J.T., Zhang, M., Zhang, X.Y., Shan, Y.J., Li, G.H., Yin, J.M., Song, W.T., Li, H.F., Zhao, G.P., 2014. Association of chicken growth hormone polymorphisms with egg production. Genet. Mol. Res. 13, 4893–4903. https://doi.org/10.4238/2014.July.4.3.
- Szwaczkowski, T. 2003. Use of mixed model methodology in poultry breeding: estimation of genetic parameters, in: Muir, W.M., Aggrey, S.E. (Eds.), Poultry Genetics, Breeding and Biotechnology. CAB International, Wallingford, Oxon, UK; Cambridge, MA, USA, pp. 187– 188. https://doi.org/10.1079/9780851996608.0165.
- Tagirov, M.T., Tereshchenko, L.V., Tereshchenko, A.V., 2006. [Substantiation of the possibility of using primary germ cells as material for the preservation of poultry genetic resources]. Ptakhivnytstvo 58, 464–473.

- Tagirov, M., Artemenko, A., Tereshchenko, A., 2010. [Preservation of the poultry gene pool by cryoconservation]. Agrarnoe reshenie [Agrar. Solution] No. 10.
- Tereshchenko, O.V., Katerinich, O.O., Pankova, S.M., Borodai, V.P., 2015. [Formation of genetic resources of domestic breeds of poultry in the context of food security of the state]. Sučasne Ptahìvnictvo No. 7–8, 19–21.
- TNAU, 2020. Layer Production Indices. Expert System for Poultry, Tamil Nadu Agricultural University, Coimbatore, India. https://web.archive.org/web/20200728085318/http://www.agritech.tnau.ac.in/expert_system/po ultry/Layer%20Production%20Indices.html (accessed 26 October 2021).
- Tyasi, T.L., Makgowo, K.M., Mokoena, K., Rashijane, L.T., Mathapo, M.C., Danguru, L.W., Molabe, K.M., Bopape, P.M., Mathye, N.D., Maluleke, D., Gunya, B., Gxasheka, M., 2020.
 Classification and regression tree (CRT) analysis to predict body weight of Potchefstroom Koekoek laying hens. Adv. Anim. Vet. Sci. 8, 354–359. https://doi.org/10.17582/journal.aavs/2020/8.4.354.359.
- Vakhrameev, A.B., 2021. Research Institute of Farm Animal Genetics and Breeding Branch of the L.K. Ernst Federal Research Center for Animal Husbandry, Pushkin, St. Petersburg. Unpublished work.
- Vakhrameev, A.B., Makarova, A.V., 2021. [Exterior Assessment of Chickens: Monograph], Electronic resource (CD-R). Research Institute of Farm Animal Genetics and Breeding – Branch of the L.K. Ernst Federal Research Center for Animal Husbandry; Publishing House of FSBSI FRC VIZh named after L.K. Ernst, Dubrovitsy.
- Valentim, J.K., Lima, H.J.D.Á., Bittencourt, T.M., Barros, F.K.Q., Braga, J.D.D.C., Antunes, H.C.F., 2019. Performance and welfare of different genetic groups of laying hen. Acta Sci. 41, e42904. https://doi.org/10.4025/actascianimsci.v41i1.42904.

- Villanúa, D., Pérez-Rodríguez, L., Rodríguez, O., Viñuela, J., Gortazar, C., 2007. How effective is pre-release nematode control in farm-reared red-legged partridges Alectoris rufa? J. Helminthol. 81, 101–103. https://doi.org/10.1017/S0022149X07251211.
- Weigend, S., Romanov, M.N., 1999. Molekulare Charakterisierung genetischer Vielfalt beim Geflügel – Molecular characterization of genetic diversity in chicken, in: Jahresbericht 1998.
 Bundesforschungsanstalt für Landwirtschaft (FAL), Braunschweig, Germany, p. 66.
- Weigend, S., Romanov, M.N., 2002. Genetische Diversitätsanalysen bei Hühnern mit Hilfe molekularer Marker – Assessment of genetic diversity in chickens using molecular markers, in: Jahresbericht 2001. Bundesforschungsanstalt für Landwirtschaft (FAL), Braunschweig, Germany, p. 67.
- Weigend, S., Romanov, M.N., Rath, D., 2004a. Methodologies to identify, evaluate and conserve poultry genetic resources, in: XXII World's Poultry Congress & Exhibition, Participant List & Full Text CD + Book of Abstracts, Istanbul, Turkey, 8–13 June 2004. WPSA – Turkish Branch, Istanbul, Turkey, p. 84.
- Weigend, S., Romanov, M.N., Ben-Ari, G., Hillel, J., 2004b. Overview on the use of molecular markers to characterize genetic diversity in chickens, in: XXII World's Poultry Congress & Exhibition, Participant List & Full Text CD + Book of Abstracts, Istanbul, Turkey, 8–13 June 2004. WPSA – Turkish Branch, Istanbul, Turkey, p. 192.
- Wężyk, S., Cywa-Benko, K., Romanov, M.N., 1994. Ochrona przed zagładą rodzimych ras drobiu w krajach Wschodniej Europy / Protection against the extinction of the native breeds of poultry in the countries of Eastern Europe, in: Proceedings of the International Symposium "Conservation Measures for Rare Farm Animal Breeds", Balice, Poland, 17–19 May 1994. National Research Institute of Animal Production, Polish Society of Animal Production, Balice, Poland, pp. 25–26.
- Wezyk, S., Cywa-Benko, K., Romanov, M.N., 1994. Protection against the extinction of the native breeds of poultry in the countries of Eastern Europe, in: Abstracts of the International

Symposium "Conservation Measures for Rare Farm Animal Breeds", Balice, Poland, 17–19 May 1994. National Research Institute of Animal Production, Polish Society of Animal Production, Kraków, Poland, p. 20 + Suppl.

- Wezyk, S., Cywa-Benko, K., Romanov, M.N., 1996. Protection against the extinction of the native breeds of poultry in Eastern European countries, in: Proceedings of the International Symposium "Conservation Measures for Rare Farm Animal Breeds", Balice, Poland. National Research Institute of Animal Production, Polish Society of Animal Production, Kraków, Poland, pp. 89–101.
- Williams, J.B., Sharp, P.J., 1978. Ovarian morphology and rates of ovarian follicular development in laying broiler breeders and commercial egg-producing hens. Br. Poult. Sci. 19, 387–395. https://doi.org/10.1080/00071667808416490.
- Zubets, M.V., Burkat, V.P., Melnyk, Yu.F., Guzyev, I.V., Yefimenko, M.Ya., Podoba, B.Ye., Behma, L.O., Biryukova, O.D., Boroday, I.S., Kovtun, S.I., et al., 2007. [Methodological Aspects of Farm Animal Gene Pool Conservation]. Ahrarna nauka, Kyiv, Ukraine.



Fig. 1. Measurement of pectoral angle in birds of the studied breeds and populations using a goniometer.

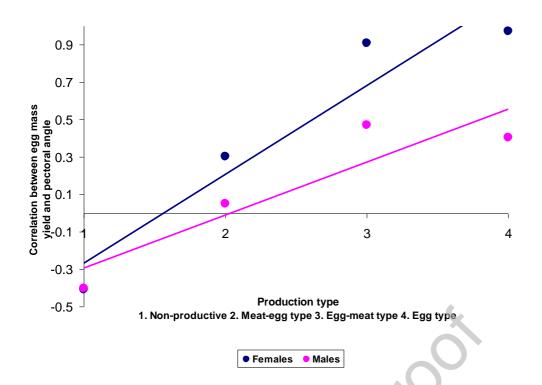


Fig. 2. Correlation relationship between the egg productivity of the breeding flock and pectoral angle in females and males when grouping the studied breeds and populations(except one MTB population as explained in the text) into four major production-based groups.

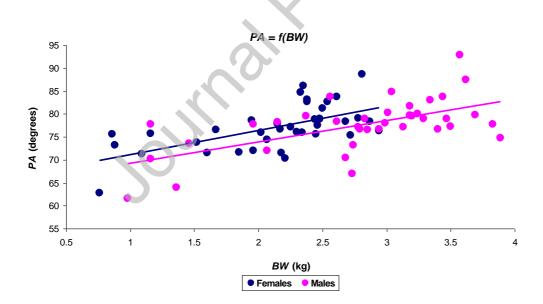


Fig. 3. Dependence of mean pectoral angle (PA) on mean body weight (BW) in 38 studied breeds and chicken populations (except one MTB population).

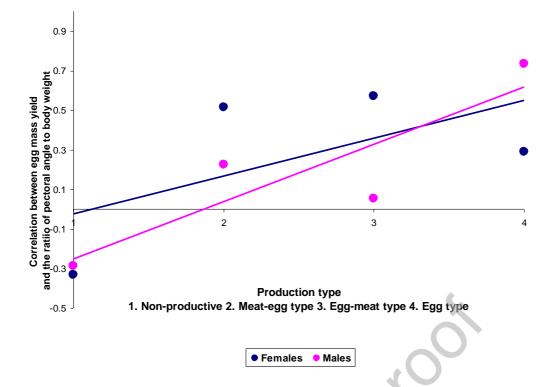
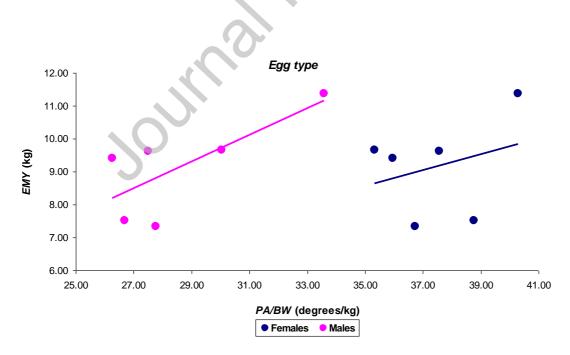
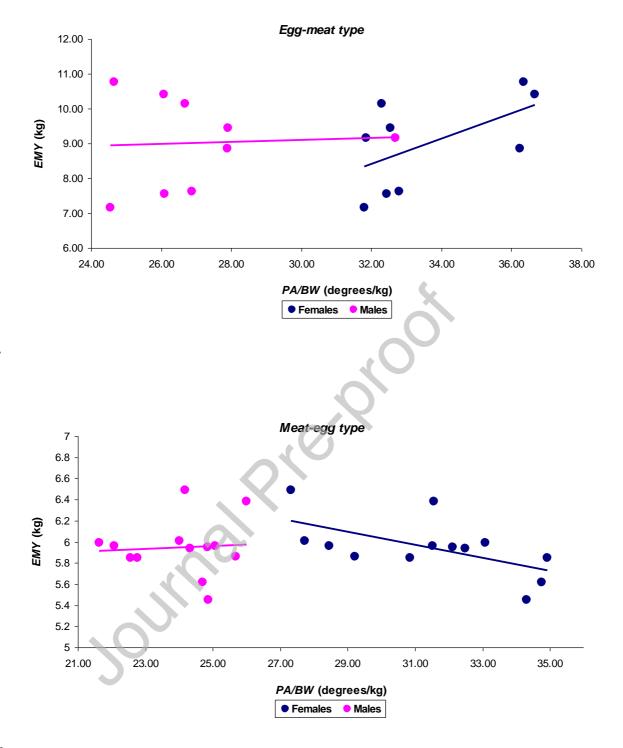


Fig. 4. Correlation relationship between the egg productivity of the breeding flock and the specific pectoral angle (PA/BW) in hens and roosters when grouping the studied breeds and populations (except one MTB population) into four major production-based groups.

a



b



С



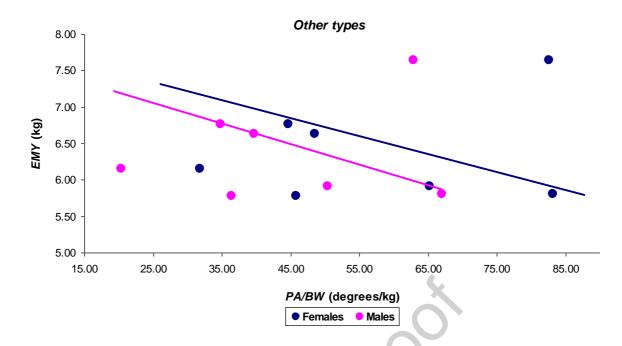
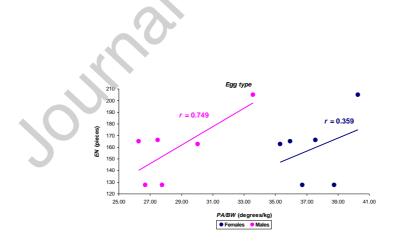


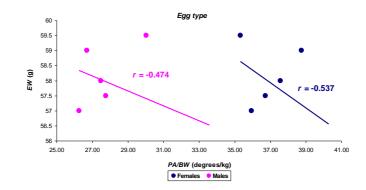
Fig. 5. Dependence of egg productivity assessed by egg mass yield (EMY) on the specific pectoral angle (PA/BW) for ETB (**a**); EMB (**b**); MEB (**c**); and other breed types non-selected for performance (**d**) when grouping the studied breeds and populations (except one MTB population) into four major production-based groups.

a

b

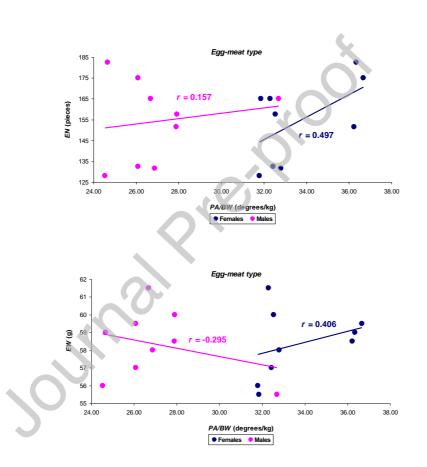


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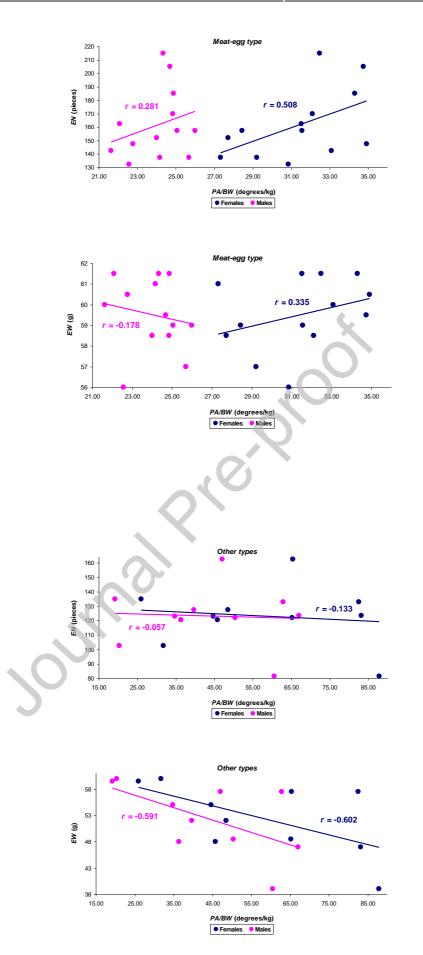
d



e

f

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g h

Fig. 6. Dependences of the mean number of eggs laid (EN) and their mean weight (EW) on the specific pectoral angle (PA/BW) for ETB (**a**, **b**); EMB (**c**, **d**); MEB (**e**, **f**); and other breed types non-selected for performance (**g**, **h**) when grouping the studied breeds and populations (except one MTB population) into four major production-based groups.

Table 1. Performance traits (mean \pm standard deviation) in the 39 studied chicken breeds (populations) when grouped according to the traditional classification model (TCM; Larkina *et al.*, 2021; Romanov *et al.*, 2021)

Classificati	Breed (population)	Code	n	Egg	Egg	Body weight, kg	
on type				number	weight, g	females	males
	Leghorn Light			166.0±4.	58.0±0.	2.02±0.	2.79±0.1
	Brown (or Italian	LLB	43	0	3	05	6
ð	Partridge)		5				-
Egg-type	Russian White	RWG	266	205.0±0.	55.5±0.	1.95±0.	2.37±0.0
Egg	Russian wine		200	5	5	07	8
	Minorca Black	МВ	135	165.0±5.	55.5±1.	2.48±0.	2.56±0.1
	Minorea Black			0	5	13	5
e	White Cornish \times	WC \times (BL \times SL)	119	157.5±2.	59.5±0.	5.63±0.	6.63±0.4
	(Brahma Light ×			5	5	19	2
Meat-type	Sussex Light)						
Me	Red White-tailed	RWD	49	162.5±2.	57.5±0.	1.09±0.	1.36±0.0
	Dwarf			5	5	07	5
Dual purpose (egg- meat)	Zagorsk Salmon ZS	ZS	108	170.0±10	58.5±0.	2.61±0.	3.34±0.1
		20	100	.0	5	10	0
	Pushkin	Pu 362	362	215.0±5.	61.5±0.	2.50±0.	3.44±0.1
			502	0	5	08	1

Rhode Island Red RIR 125 0 5 09 9 Leningrad Mille LMF 131 185.0±5. 61.5±0. 2.25±0. 3.20±0.1 Fleur LMF 131 0 5 06 4 Leningrad Golden- LGG 240 182.5±2. 59.0±1. 2.33±0. 3.13±0.1 and-gray PB 165 165.0±5. 61.5±0. 2.44±0. 3.01±0.0 Pantsirevka Black PB 165 10 5 9.0±1. 2.78±0. 3.18±0.1 Speckled ABS 166 5 0 08 4 Aurora Blue AB 86 165.0±5. 57.0±1. 2.07±0. 2.68±0.1 Australorp Black Ao8 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Naked Neck NN 109 5 5 11 0 <td< th=""><th rowspan="6"></th><th rowspan="2">Rhode Island Red</th><th rowspan="2">RIR</th><th rowspan="2">125</th><th>175.0±5.</th><th>59.5±0.</th><th>2.35±0.</th><th>2.99±0.0</th></td<>		Rhode Island Red	RIR	125	175.0±5.	59.5±0.	2.35±0.	2.99±0.0
$ \left(\begin{array}{cccccccccccccccccccccccccccccccccccc$					0	5	09	9
Fleur Image: constraint of the state of the		Leningrad Mille	LMF	131	185.0±5.	61.5±0.	2.25±0.	3.20±0.1
Image: state of the s		Fleur			0	5	06	4
and-gray Image: second se		Leningrad Golden-	LGG	240	182.5±2.	59.0±1.	2.33±0.	3.13±0.1
Pantsirevka Black PB 165 0 5 13 8 Australorp Black ABS 166 5 90.10 2.78±0. 3.18±0.1 Speckled ABS 166 5 0 08 4 Aurora Blue AB 86 165.0±5. 57.0±1. 2.07±0. 2.68±0.1 Australorp Black AB 86 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black AOE 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black AOE 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Amrock Ar 88 162.5±2. 57.5±0. 1.96±0. 2.78±0.0 Naked Neck NN 109 127.5±2. 57.5±0. 1.96±0. 2.78±0.0 New Hampshire NH 84 0 5 06 4 <t< td=""><td>and-gray</td><td>5</td><td>0</td><td>07</td><td>2</td></t<>		and-gray			5	0	07	2
Image: specified Australorp Black ABS 166 157.5±2. 59.0±1. 2.78±0. 3.18±0.1 Speckled ABS 166 5 0 08 4 Aurora Blue AB 86 165.0±5. 57.0±1. 2.07±0. 2.68±0.1 Aurora Blue AB 86 165.0±5. 57.0±1. 2.07±0. 2.68±0.1 Australorp Black AB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black AoB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Australorp Black Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Marock NN 109 127.5±2. 57.5±0. 1.96±0. 2.78±0.0 New Hampshire NH 84 0 5 12 7 New Hampshire Pr 114 0 5 14 </td <td></td> <td rowspan="2">Pantsirevka Black</td> <td rowspan="2">РВ</td> <td rowspan="2">165</td> <td>165.0±5.</td> <td>61.5±0.</td> <td>2.44±0.</td> <td>3.01±0.0</td>		Pantsirevka Black	РВ	165	165.0±5.	61.5±0.	2.44±0.	3.01±0.0
Normal Speckled ABS 166 5 0 08 4 Aurora Blue AB 86 165.0±5. 57.0±1. 2.07±0. 2.68±0.1 Aurora Blue AB 86 0 0 07 0 Australorp Black AoB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Naked Neck NN 109 127.5±2. 57.5±0. 1.96±0. 2.78±0.0 New Hampshire NH 84 205.0±5. 59.5±0. 2.38±0. 3.24±0.2 New Hampshire NH 84 0 5 06 4 Pervomai Pm 114 152.0±3. 58.5±0. 2.72±0. 3.29±0.2 Plymouth Rock PRB 48 162.5±2. 61.5±0. 2.46±0. 3.50±0.0 B					0	5	13	8
Speckled 5 0 08 4 Aurora Blue AB 86 65.0+5. 57.0+1. 2.07±0. 2.68±0.1 Aurora Blue AB 86 0 0 07 0 Australorp Black AoB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black AoB 47 5 0 07 9 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Naked Neck NN 109 127.5±2. 57.5±0. 1.96±0. 2.78±0.0 Naked Neck NN 109 5 5 12 7 New Hampshire NH 84 205.0±5. 59.5±0. 2.38±0. 3.24±0.2 Pervomai Pm 114 0 5 14 2 Plymouth Rock PRB 48 162.5±2. 61.5±0. 2.46±0. 3.50±0.0 Barred PC 120 142.5±2. 6		Australorp Black		166	157.5±2.	59.0±1.	2.78±0.	3.18±0.1
Aurora Blue AB 86 0 0 07 0 Australorp Black AoB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black AoB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Naked Neck NN 109 127.5±2. 57.5±0. 1.96±0. 2.78±0.0 New Hampshire NH 84 205.0±5. 59.5±0. 2.38±0. 3.24±0.2 New Hampshire NH 84 205.0±5. 59.5±0. 2.38±0. 3.24±0.2 Pervomai Pm 114 0 5 14 2 Plymouth Rock PRB 48 5 5 09 7 Barred PC 120 142.5±2. 60.0±1. 2.30±0. 3.69±0.3		Speckled	ABS		5	0	08	4
Opposite 0 0 07 0 Australorp Black AoB 47 157.5±2. 59.0±1. 2.81±0. 3.57±0.0 Australorp Black AoB 47 5 0 07 9 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Amrock Ar 88 5 5 11 0 Naked Neck NN 109 5 5 12 7 New Hampshire NH 84 205.0±5. 59.5±0. 2.38±0. 3.24±0.2 Pervomai Pm 114 152.0±3. 58.5±0. 2.72±0. 3.29±0.2 Plymouth Rock Pm 114 0 5 14 2 Plymouth Rock PRB 48 162.5±2. 61.5±0. 2.46±0. 3.50±0.0 Barred PC 120 142.5±2. 60.0±1. 2.30±0. 3.69±0.3		Aurora Blue	٨B	86	165.0±5.	57.0±1.	2.07±0.	2.68±0.1
Australorp Black AoB 47 5 0 07 9 Amrock Ar 88 162.5±2. 59.5±0. 2.17±0. 2.61±0.1 Amrock Ar 88 5 5 11 0 Naked Neck NN 109 127.5±2. 57.5±0. 1.96±0. 2.78±0.0 New Hampshire NH 84 0 5 12 7 New Hampshire NH 84 0 5 06 4 Pervomai Pm 114 152.0±3. 58.5±0. 2.72±0. 3.29±0.2 Plymouth Rock PRB 48 162.5±2. 61.5±0. 2.46±0. 3.50±0.0 Barred PRB 48 5 5 09 7 Poltava Clay PC 120 142.5±2. 60.0±1. 2.30±0. 3.69±0.3		Aurora Blue	АВ	80	0	0	07	0
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$		Australorp Black	AoB	47	157.5±2.	59.0±1.	2.81±0.	3.57±0.0
$ \begin{array}{ c c c c c c c } \hline \mbox{Mirrock} & \mbox{Ar} & \mbox{88} & \mbox{5} & \mbox{5} & \mbox{11} & \mbox{0} & \mbox{0} & \mbox{11} & \mbox{0} & \mbox{11} & \mbox{0} & \mbox{11} & \mbox{0} & \mbox{11} & \mbox{11} & \mbox{0} & \mbox{11} & \mbox{11} & \mbox{11} & \mbox{12} & \mbox{11} & \mbox{12} & \mbox{11} & \mbox{11} & \mbox{12} & \mbox{12} & \mbox{11} & \mbox{12} & \mbox{12}$					5	0	07	9
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$		Amrock	Ar	88	162.5±2.	59.5±0.	2.17±0.	2.61±0.1
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$					5	5	11	0
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	(mea	Naked Neck	NN	109	127.5±2.	57.5±0.	1.96±0.	2.78±0.0
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	Irpose				5	5	12	7
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	ual p	New Hampshire	NH	84	205.0±5.	59.5±0.	2.38±0.	3.24±0.2
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Ď				0	5	06	4
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$		Pervomai	Pm	114	152.0±3.	58.5±0.	2.72±0.	3.29±0.2
PRB 48 5 5 09 7 Poltava Clay PC 120 142.5±2. 60.0±1. 2.30±0. 3.69±0.3					0	5	14	2
Barred 5 5 09 7 Poltava Clay PC 120 142.5±2. 60.0±1. 2.30±0. 3.69±0.3		Plymouth Rock	PRB	48	162.5±2.	61.5±0.	2.46±0.	3.50±0.0
Poltava Clay PC 120		Barred			5	5	09	7
		Poltava Clay PC	DC	120	142.5±2.	60.0±1.	2.30±0.	3.69±0.3
			rC		5	0	06	5

157.5±2. 60.0±1.		
	2.54±0.	2.83±0.1
Sussex LightSL10350	10	5
	2.34±0.	2.94±0.1
Faverolles SalmonFS9850	11	3
	2.38±0.	3.47±0.0
Tsarskoye SeloTs32255	04	9
	2.87±0.	3.62±0.1
Yurlov Crower YC 161 5 0	10	9
	2.45±0.	3.40±0.1
Orloff Mille Fleur OMF 84 5 0	11	8
⁰ 135.0+5. 59.5±1.	2.94±0.	3.89±0.1
Moscow GameMG9705	09	7
	2.48±0.	3.83±0.2
(Kulangi) UG 93 5 0	10	2
	2.15±0.	3.04±0.0
Russian Crested RC 77 5 5	07	9
	2.68±0.	3.18±0.1
Ukrainian Muffed UM 103 5 0	10	4
	0.88±0.	1.16±0.0
Bantam Mille Fleur BMF 100 5 0	03	3
	2.21±0.	2.73±0.1
Brahma Buff BB 57 0 1.0	12	1
	2.18±0.	2.85±0.1
Brahma Light BL 62 5 0	07	1
	1.16±0.	1.46±0.0
Silver Spangled HBSS 25 0 5	04	8

		Journa	l Pre	-proof			
	Poland White-crested	DUID		123.0±3.	55.0±1.	1.60±0.	2.07±0.0
	Black	PWB	63	0	0	08	5
	Silkie White	SW	91	81.5±1.5	39.0±1.	0.86±0.	1.16±0.0
			91	01.J±1.J	0	03	7
	Cochin Bantam	СВ	38	133.0±3.	57.5±0.	0.76±0.	0.98±0.0
	(Pekin Bantam)			0	5	02	4
	Frizzle	F	87	127.5±2.	59.0±1.	1.85±0.	2.74±0.0
			07	5	0	03	2
	Pavlov Spangled	PS	107	127.5±2.	52.0±1.	1.52±0.	1.96±0.0
			107	5	0	07	8
	Pavlov White	PW	32	120.5±2.	48.0±1.	1.67±0.	2.15±0.0
			32	5	0	05	9
	Average		5	149.6±27	57.1±4.	2.23±0.	2.89±0.9
Average		2		.7	6	80	8

Johna