

STOCHASTIC DYNAMICS OF TWO PICOPHYTOPLANKTON POPULATIONS IN A REAL MARINE ECOSYSTEM*

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A stochastic reaction-diffusion-taxis model is analyzed to get the stationary distribution along water column of two species of picophytoplankton, that is piceokaryotes and *Prochlorococcus*. The model is valid for weakly mixed waters, typical of the Mediterranean Sea. External random fluctuations are considered by adding a multiplicative Gaussian noise to the dynamical equation of the nutrient concentration. The statistical tests show that shape and magnitude of the theoretical concentration profile exhibit a good agreement with the experimental findings. Finally, we study the effects of seasonal variations on picophytoplankton groups, including an oscillating term in the auxiliary equation for the light intensity.

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1. Introduction

In an ecological context, the study of vertical distributions of the picophytoplankton communities is very important to predict and understand future changes in marine ecosystems, produced by global warming [1, 2]. In re-

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cent works, the spatio-temporal dynamics of phytoplankton in the Mediterranean Sea have been studied by using a stochastic approach to describe and simulate the experimental findings for the chlorophyll concentration [3, 4]. In order to analyze the dynamics of phytoplankton, it is worth underlining that marine ecosystems are complex systems, that is open systems characterized by nonlinear interactions between their parts and external perturbations, both deterministic and random, due to environmental variables [5–25]. As a consequence, the study of a marine ecosystem has to be performed by considering also the effects of random perturbations, which can be treated as environmental noise sources. The model is used to analyze the behaviour of two picophytoplankton groups, which are subject to a stochastic dynamics. In particular, in order to better simulate this nonlinear and noisy dynamics, we take into account in our model the presence of external random perturbations by including, in the equation that describes the dynamics of the nutrient concentration, a term of multiplicative noise [3, 4, 12].

In order to analyze the picophytoplankton dynamics, we consider an ecosystem which possesses the hydrological characteristics typical of the Strait of Sicily and is known to be a biologically rich area of the Mediterranean Sea with a key role in terms of fisheries [26–29]. Moreover, the two picophytoplankton groups considered belong to the smaller size fraction (less than 3 μm) of phytoplankton and are mainly represented by picoprokaryotes and picoeukaryotes. Specifically, the picoprokaryotes group is dominated in the Sicily Channel by a species of cyanobacteria, *i.e.* *Prochlorococcus*, while picoeukaryotes group is mainly represented by pelagophytes and prymnesiophytes [30]. In this work, we study the vertical profiles of picoeukaryotes and *Prochlorococcus*, which account for about 60% of total chlorophyll on average in the Mediterranean Sea.

The interaction of these populations with the environment occurs through two factors that limit the growth of this aquatic microorganisms: light intensity and nutrient. The light penetrates through the surface of the water and decreases exponentially along the water column. *Vice-versa*, nutrient concentrations along the water column are characterized by an increasing trend from the sea surface to the benthic layer. The opposite gradients of two limiting factors contribute to select different species, and sometimes groups, along the water column, and determine the biodiversity of the ecosystem [1, 2, 4].

The responses of each picophytoplankton group to environment solicitations depend strongly on the biological and physical parameters [31]. The values of the environmental parameters have been chosen to reproduce the marine ecosystem of the Sicily Channel in summer, characterized by oligotrophic water and high light intensity. Biological parameters depend on both the metabolism and cell structure of the specific microorganisms investigated. It is important noting that some biological parameters play a key

role on the position and magnitude of planktonic groups. In fact, the half-saturation constants locate the position of the production layer and depth of concentration peak for each picophytoplankton group. Moreover, the growth rates and nutrient uptake determine the balance of the primary production of phytoplankton in marine ecosystem [32], contributing to modify the composition and magnitude of the planktonic communities.

The first goal has been to obtain the spatio-temporal distributions of two groups of picophytoplankton, *i.e.* picoeukaryotes and *Prochlorococcus*, using a reaction-diffusion-taxis model [1, 2]. As a second step, we studied the spatio-temporal evolution of the biomass concentrations of both groups, obtaining the distributions of the total *chlorophyll a* (*chl a*) and *divinyl chlorophyll a* (*Dvchl a*) concentrations in stationary regime, and comparing the numerical results with experimental data collected in a site of the Sicily Channel. Finally, we analyzed the role of seasonal variations on picophytoplankton dynamics, including an oscillating term in the auxiliary equation for the light intensity.

2. The model

The distributions of the two picophytoplankton groups in stationary regime are obtained by using a stochastic model with conditions typical of the Mediterranean Sea, where the vertical water columns are weakly mixed. The dynamics, competition and structuring of phytoplankton groups in marine ecosystems have been studied in a series of theoretical works based on deterministic models [1, 2, 33–35]. In particular, it was shown that biological and physical parameters determine the position of the maximum of the biomass concentration, that can be localized at the surface or, alternatively, in a deep layer. Moreover, these parameters have also to guarantee the coexistence of the two groups [2, 35], *i.e.* picoeukaryotes and *Prochlorococcus*, in the deep chlorophyll maximum (DCM), according to experimental findings. In order to take into account all these conditions and describe correctly the picophytoplankton dynamics, we devised a stochastic model by including, in a deterministic advection-reaction-diffusion model [1, 35], a source of the multiplicative Gaussian noise.

2.1. The stochastic model

In this paragraph, we use a stochastic model to study the dynamics of the two picophytoplanktonic populations [1, 2] distributed along a one-dimensional spatial domain (z -direction). In particular, we assume that the interaction of these microorganisms with the marine environment occurs through the two factors which limit the growth of the planktonic communities: light intensity (I) and nutrient (R), *i.e.* phosphorus. The stochastic

model has been obtained by modifying a deterministic model based on a system of three differential equations [1, 2]. The first two equations describe the dynamics of the picophytoplankton groups, *i.e.* picoeukaryotes ($b_1(z, t)$) and *Prochlorococcus* ($b_2(z, t)$). The third equation simulates the change of nutrient concentration ($R(z, t)$) along the water column.

It is worth noting that the magnitude of the picophytoplankton biomass at depth z depends on three processes: growth, loss, and movement. In our model, we consider these contributes in the differential equations for the dynamics of picophytoplankton groups. Moreover, we also take into account random fluctuations and their effect on the phytoplankton dynamics, by inserting in the equation for the nutrient dynamics a term of spatially uncorrelated noise. In particular, we use a source of multiplicative white Gaussian noise $\xi_R(z, t)$ with intensity σ_R and statistical properties: $\langle \xi_R(z, t) \rangle = 0$ and $\langle \xi_R(z, t) \xi_R(z', t') \rangle = \sigma_R \delta(z - z') \delta(t - t')$. Finally, the light intensity $I(z, t)$ is modeled by a function varying, along the water column, with the depth and biomass concentration. The stochastic model is defined by the following equations

$$\frac{\partial b_1(z, t)}{\partial t} = b_1 \min(f_{I_1}(I), f_{R_1}(R)) - m_1 b_1 + D \frac{\partial^2 b_1(z, t)}{\partial z^2} - v_1 \frac{\partial b_1(z, t)}{\partial z}, \quad (1)$$

$$\frac{\partial b_2(z, t)}{\partial t} = b_2 \min(f_{I_2}(I), f_{R_2}(R)) - m_2 b_2 + D \frac{\partial^2 b_2(z, t)}{\partial z^2} - v_2 \frac{\partial b_2(z, t)}{\partial z}, \quad (2)$$

$$\begin{aligned} \frac{\partial R(z, t)}{\partial t} = & - \sum \frac{b_i(z, t)}{Y_i} \times \min(f_{I_i}(I), f_{R_i}(R)) + D \frac{\partial^2 R(z, t)}{\partial z^2} \\ & + \sum \varepsilon_i m_i \frac{b_i(z, t)}{Y_i} + R \xi_R(z, t), \end{aligned} \quad (3)$$

$$I(z) = I_{\text{in}} \exp \left\{ - \int_0^z \left[\sum a_i b_i(Z) + a_{\text{bg}} \right] dZ \right\}, \quad (4)$$

where v_1 and v_2 are the buoyancy velocities of the two picophytoplankton groups, *i.e.* picoeukaryotes and *Prochlorococcus*, respectively; D is the vertical diffusion coefficient; ε_i , m_i and $1/Y_i$ are nutrient recycling coefficient, specific loss rate and nutrient content of the i th picophytoplankton group, respectively; a_1 and a_2 are the absorption coefficients of the two picophytoplankton groups, and a_{bg} is the background turbidity; I_{in} is the incident light intensity at the water surface. Furthermore, $f_{I_i}(I)$ and $f_{R_i}(R)$ are given by the Michaelis–Menten formulas

$$f_{I_i}(I) = r_i I / (I + K_{I_i}), \quad (5)$$

$$f_{R_i}(R) = r_i R / (R + K_{R_i}). \quad (6)$$

Here, r_i is the maximum growth rate, K_{I_i} and K_{R_i} are the half-saturation constants for light intensity and nutrient concentration, respectively, of the i th picophytoplankton group. These constants depend on the metabolism of the specific picophytoplankton groups considered. In particular, the half-saturation constants, K_{R_i} and K_{I_i} , contribute to determining the position along the water column of the biomass production layer for each group.

The stochastic model also includes six equations for the boundary conditions. In particular, we can observe that phytoplankton biomass does not enter or leave the water column. Therefore, the boundary conditions for concentrations of picoeukaryotes and *Prochlorococcus* biomass account for the absence of flux through both surface layer $z = 0$ and seabed $z = z_b$,

$$\left[D \frac{\partial b_i}{\partial z} - v_i b_i \right] \Big|_{z=0} = \left[D \frac{\partial b_i}{\partial z} - v_i b_i \right] \Big|_{z=z_b} = 0. \quad (7)$$

Moreover, we fix the boundary conditions for nutrients, which do not come from the top of the water column but are provided from the bottom. In particular, nutrient concentration at the bottom of the water column, $R(z_b)$, is set at value R_{in} , which depends on the site investigated. These boundary conditions are described by the following equations

$$\frac{\partial R}{\partial z} \Big|_{z=0} = 0, \quad R(z_b) = R_{in}. \quad (8)$$

Equations (1)–(8) form the stochastic reaction-diffusion-taxis model used in this work. By solving them, we obtain the spatio-temporal dynamics of the biomass concentrations of picoeukaryotes and *Prochlorococcus*, nutrient concentration and light intensity.

2.2. Biological and environmental parameters

In order to reproduce the experimental profile of *chl a* and *Dvchl a* concentration, measured in a marine site close to the Libyan continental shelf (site L1129b in Fig. 1), we set the values of the environmental and biological parameters so that the presence of a deep chlorophyll maximum for both picophytoplankton groups is guaranteed [1, 2, 4]. According to our previous discussion, the values of the biological parameters have been fixed to simulate the behaviour of picoeukaryotes and *Prochlorococcus*. In particular, the half-saturation constants for each group are chosen to obtain suitable positions for both the production layers and the peak of biomass concentration.

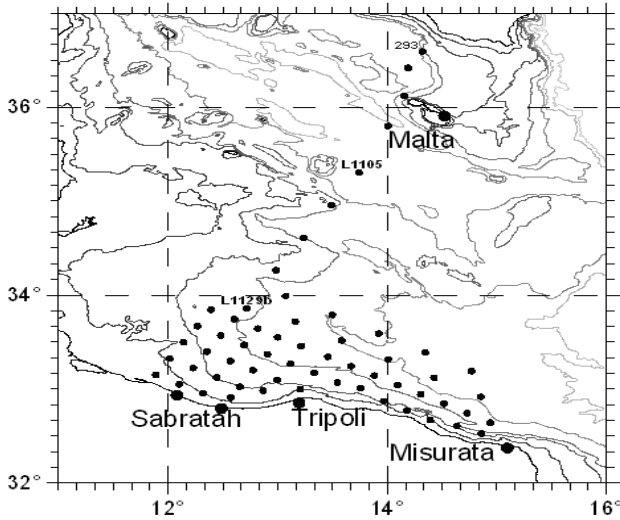


Fig.1. Locations of marine sites, where experimental data were collected in the period of the 12th–24th August 2006 in the Sicily Channel area, during the MedSudMed-06 Oceanographic Survey onboard the R/V Urania. Data used in this work, for comparison with theoretical results, are those acquired in site L1129b (close to the Libyan coast).

The parameters $1/Y_1$ and $1/Y_2$ contribute to determine the steady distributions of the picophytoplankton concentrations. In particular, experimental findings show that (i) the peak of biomass concentration of *Prochlorococcus* is shallower than that of picoeukaryotes and (ii) the average cell concentration of *Prochlorococcus* along the water column is much higher than that of picoeukaryotes [30, 37]. In these conditions, a smaller amount of nutrient is available for *Prochlorococcus* localized in the biomass peak. Therefore, in order to obtain, for the two picophytoplankton groups, cell concentrations in agreement with the real data, $1/Y_2$ is set at a value much smaller than $1/Y_1$ (see Table I). Moreover, the absorption coefficient of *Prochlorococcus*, fixed in our model, has to be very different from that of the picoeukaryotes. In fact, due to the higher average cell concentration of *Prochlorococcus* (5.2×10^4 cells ml^{-1}) with respect to that of picoeukaryotes (0.6×10^3 cells ml^{-1}), in order to simulate the same gradient of light intensity inside the production layers [35], we had to exploit an absorption coefficient for *Prochlorococcus* lower than that used for picoeukaryotes, setting $a_2 = 2.4 \times 10^{-15} \text{m}^2 \text{cell}^{-1}$.

The sinking velocity is set to a value typical of the picophytoplankton [1], while the maximum specific growth rates are in agreement with those experimentally observed by other authors [38]. The values of the environmental parameters have been chosen to reproduce marine ecosystem of the Sicily Channel in summer, *i.e.* oligotrophic water ($R_{in} < 10.0$ mmol nutrient m^{-3}) and high light intensity ($I_{in} > 1000$ $\mu\text{mol photons } m^{-2} s^{-1}$). Moreover, the vertical turbulent diffusivity is fixed at values typical of weakly mixed waters ($D = 1.0$ $cm^2 s^{-1}$). The numerical values of the parameters are reported in Table I.

TABLE I

Parameters used in the stochastic model. The values of the biological parameters are those typical of picophytoplankton groups.

Symbol	Interpretation	Units	Site L1129b
I_{in}	Incident light intensity	$\mu\text{mol photons } m^{-2} s^{-1}$	1404.44
a_{bg}	Background turbidity	m^{-1}	0.045
a_1	Absorption coefficient of picoeukaryotes	$m^2 \text{ cell}^{-1}$	6×10^{-10}
a_2	Absorption coefficient of <i>Prochlorococcus</i>	$m^2 \text{ cell}^{-1}$	2.4×10^{-15}
z_b	Depth of the water column	m	186
D	Vertical turbulent diffusivity	$cm^2 s^{-1}$	1.0
r_1	Maximum specific growth rate of picoeukaryotes	h^{-1}	0.08
r_2	Maximum specific growth rate of <i>Prochlorococcus</i>	h^{-1}	0.07
K_{I_1}	Half-saturation constant of light-limited growth of picoeukaryotes	$\mu\text{mol photons } m^{-2} s^{-1}$	20
K_{R_1}	Half-saturation constant of nutrient-limited growth of picoeukaryotes	mmol nutrient m^{-3}	0.0425
K_{I_2}	Half-saturation constant of light-limited growth of <i>Prochlorococcus</i>	$\mu\text{mol photons } m^{-2} s^{-1}$	98
K_{R_2}	Half-saturation constant of nutrient-limited growth of <i>Prochlorococcus</i>	mmol nutrient m^{-3}	0.0150
m_1	Specific loss rate of picoeukaryotes	h_1	0.01
m_2	Specific loss rate of <i>Prochlorococcus</i>	h_1	0.01
$1/Y_1$	Nutrient content of picoeukaryotes	mmol nutrient $cell^{-1}$	1×10^{-9}
$1/Y_2$	Nutrient content of <i>Prochlorococcus</i>	mmol nutrient $cell^{-1}$	4×10^{-15}
ϵ_1	Nutrient recycling coefficient of picoeukaryotes	dimensionless	0.5
ϵ_2	Nutrient recycling coefficient of <i>Prochlorococcus</i>	dimensionless	0.5
v_1	Buoyancy velocity of picoeukaryotes	$m h^{-1}$	-0.0042
v_2	Buoyancy velocity of <i>Prochlorococcus</i>	$m h^{-1}$	-0.0042
R_{in}	Nutrient concentration at z_b	mmol nutrient m^{-3}	5.0

2.3. Results of the stochastic model

In order to obtain the theoretical distributions of biomass concentrations for the two picophytoplankton populations, we solve numerically Eqs. (1)–(8) by using the Ito scheme, and averaging over 1000 realizations [12, 36].

The numerical method, whose computer implementation consists in a C++ program, is based on an explicit finite difference scheme with centered-in-space differencing for the diffusion term and upwind differencing for the taxis term. In order to get the steady spatial distributions, we integrate our system over a time interval long enough to obtain the stationary solution. In particular, we solve the equations fixing as a maximum time $t_{\max} = 4 \times 10^4$ h. As initial conditions, we set that the picoeukaryotes and *Prochlorococcus* biomass are concentrated in two layers close to the deep chlorophyll maximum experimentally observed. Moreover, we impose that the nutrient concentration remains approximately constant from the water surface up to the DCM, while increases linearly below the DCM up to the seabed.

We recall that the stochastic model provides the steady distributions of the picophytoplankton concentrations expressed in cell/m^3 , while the experimental data of *chl a* concentrations are expressed in $\mu\text{g}/\text{l}$. Therefore, in order to compare numerical results with the experimental profile, the theoretical cell concentrations of picoeukaryotes and *Prochlorococcus* are converted into *chl a* and *Dvchl a* concentrations, respectively, by using the curves of mean vertical profile (see Fig. 2) obtained by Brunet *et al.* [37]. In particular, these curves describe the exponential behaviour of the chlorophyll per cell ratio, as a function of depth, for both picophytoplankton groups.

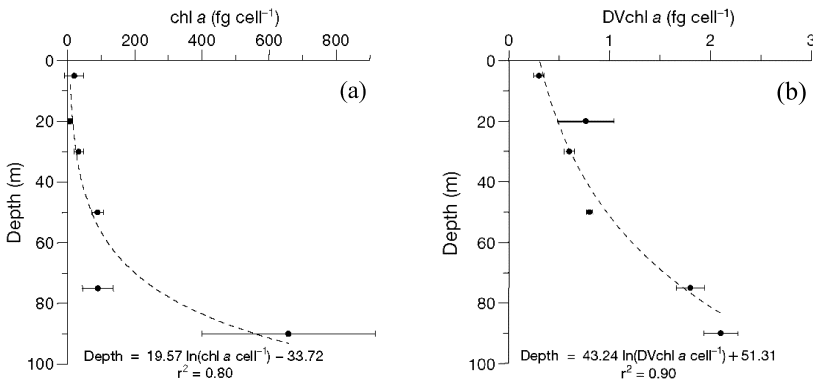


Fig. 2. Mean vertical profile of *chl a* per picoeukaryote cell (panel a) and *Dvchl a* per *Prochlorococcus* cell (panel b). Error bars are Standard Deviation. Equation and r^2 for the fit are reported on the plots. (Courtesy of Brunet *et al.* [37].)

Since the structure of the *chlorophyll a* molecule is almost identical to that of *divinil chlorophyll a*, we summed their concentrations to obtain the theoretical steady profile consistent with those obtained from the experimental data. Moreover, in the Mediterranean Sea about 43% of the total quantity of *chl a* and *divinil chl a* [1, 30] is due to nano- and micro-phytoplankton,

and *Synechococcus*. Therefore, we consider this fraction of the total biomass and divide it by depth, obtaining for each site the value $\Delta b_{(Dv)chl a}$, which represents a constant concentration of *chl a* and *Dvchl a* due to other phytoplankton species present in the water column. Finally, we add the theoretical concentrations with $\Delta b_{(Dv)chl a}$. By this way, we obtain the stationary distributions in deterministic regime and for three different values of the noise intensity. The results are shown in Fig. 3. Here, we observe that the noise causes a decrease and a deeper localization of the DCM. Position, shape and magnitude of the phytoplankton peak, obtained by the stochastic model, exhibit the best agreement with those of the experimental DCM for a noise intensity equal to 0.0040. This result is confirmed by the reduced

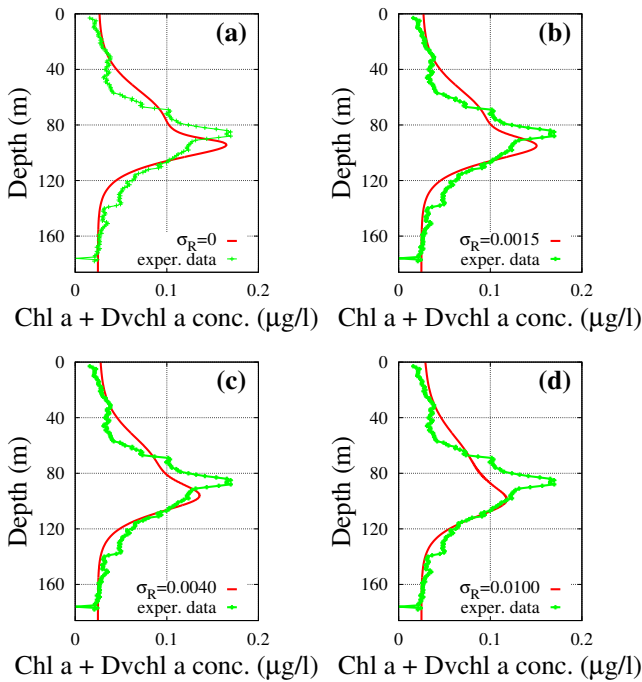


Fig. 3. Theoretical distributions (gray/red line) of the total *chl a* and *Dvchl a* concentration calculated for different values of σ_R by the stochastic model as a function of depth. The results are compared with the distributions of the total *chl a* and *Dvchl a* concentration measured (light gray/green line) in site L1129b. The theoretical values were obtained averaging over 1000 numerical realizations. The values of the parameters are those shown in Table I. The noise intensities are: (a) $\sigma_R = 0$ (deterministic case), (b) $\sigma_R = 0.0015$, (c) $\sigma_R = 0.0040$ and (d) $\sigma_R = 0.0100$.

χ^2 test that provides the best value, $\tilde{\chi}^2 = 0.0037$, for the same value of noise intensity (see Table II). In particular, the values of the reduced chi-square resulted to be much lower than the value previously obtained by the one-species model [4].

Other results (here not shown) reveal a rapid disappearance of phytoplankton biomass for $\sigma_R > 0.01$. This indicates that the stability of the nutrient concentration is a critical factor for both picophytoplankton populations studied in this paper. Finally, these results suggest that random fluctuations of the nutrient concentration could be the cause of the collapse of phytoplankton biomass in real marine ecosystems.

TABLE II

Results of χ^2 and reduced chi-square ($\tilde{\chi}^2$) goodness-of-fit test for site L1129b at different values of σ_R . The number of samples along the water column is $n = 176$.

R_{in}	σ_R	χ^2	$\tilde{\chi}^2$
5	0.0000	0.74	0.0042
5	0.0015	0.67	0.0039
5	0.0040	0.65	0.0037
5	0.0100	0.78	0.0045

3. Picophytoplankton dynamics and changes of DCM in the presence of periodical driving force

In this section, we study the time behaviour of the picophytoplankton groups in presence of seasonal variations of light intensity. In particular, in order to predict the seasonal variations of the primary production of phytoplankton in the Sicily Channel, we insert in our model an oscillating term in the auxiliary equation for the light intensity.

Recent theoretical works [3] indicate that the seasonal variations of light intensity have a strong effect on the distributions of phytoplankton populations along the water column. Therefore, in order to better reproduce spatio-temporal dynamics of picophytoplankton groups, we consider the parameter I_{in} as a periodical function of time. As a consequence, we replace in Eq. (4) the incident light intensity with the following equation

$$I_{\text{in}}(t) = I_{\text{in}}^{\text{aver}} + I_0 \cos \omega t, \quad (9)$$

where, $I_{\text{in}}^{\text{aver}}$ is the yearly weighted average of the light intensity on the sea surface taking into account only sunny days, *i.e.* in absence of cloud coverage, and I_0 is a multiplicative term that describes the magnitude of the seasonal oscillations of the incident light intensity.

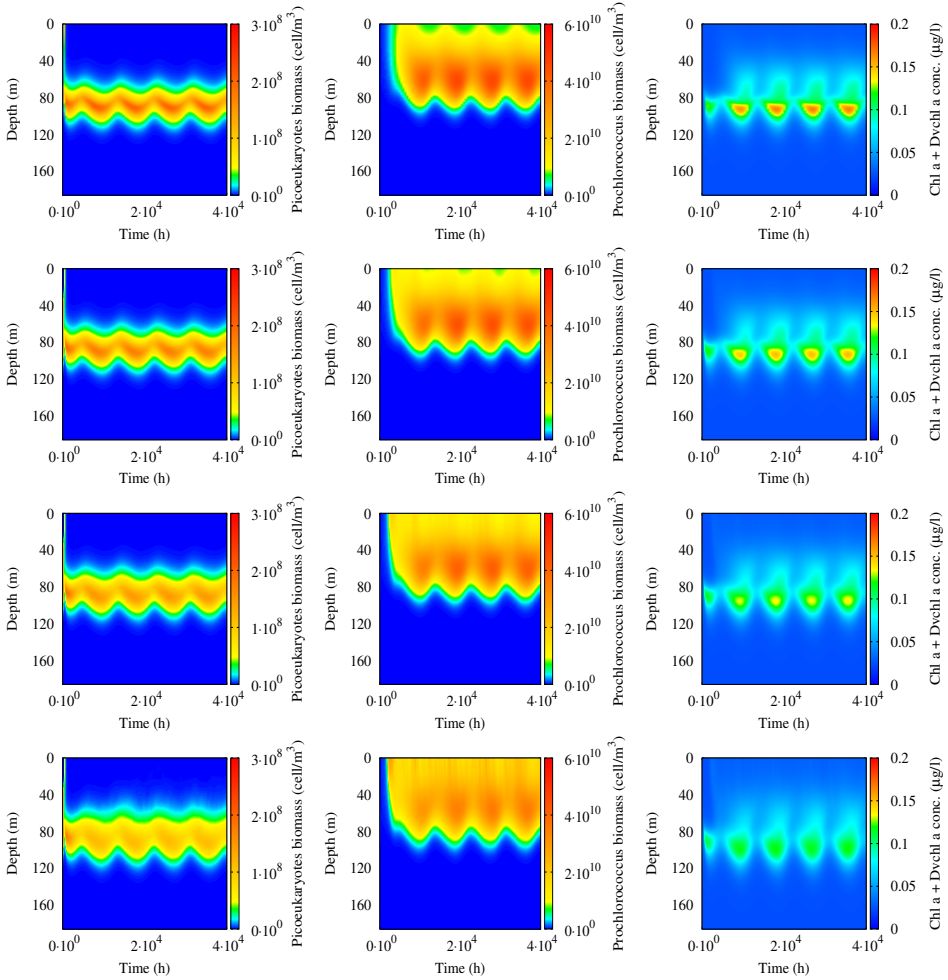


Fig. 4. Spatio-temporal behaviour of picoeukaryotes biomass, *Prochlorococcus* biomass and total *chl a* and *Dvchl a* concentration (from left to right). Contour maps were obtained in the presence of periodical (seasonal) behaviour of the light intensity $I_{in}(t)$ according to Eq. (9), with the nutrient concentration subject to random fluctuations (see Eq. (3)). The results refers to different values of the noise intensity: $\sigma_R = 0$ (deterministic regime), $\sigma_R = 0.0015$, $\sigma_R = 0.0040$ and $\sigma_R = 0.0100$ (from top to bottom). All contour maps were obtained averaging over 1000 numerical realizations. The average value of the incident light intensity is $I_{in}^{aver} = 1068.58 \mu\text{mol photons m}^{-2} \text{s}^{-1}$. The values of the other parameters are those shown in Table I.

We analyze the ecosystem dynamics in the presence of oscillating light intensity and random fluctuations of environmental variables. In particular, as a first step, we obtain the spatio-temporal dynamics of picoeukaryotes and *Prochlorococcus* by solving equation system (1)–(8). Second, as in previous section, we convert the theoretical cell concentrations of the picophytoplankton groups into *chl a* and *Dvchl a* concentrations, by using the curves of mean vertical profile shown in Fig. 2. The spatio-temporal dynamics of the two picophytoplankton groups and total *chl a* and *Dvchl a* concentration is shown in Fig. 4. Here, we observe a decreasing magnitude of the picophytoplankton concentrations during autumn, followed by an increase of biomass concentrations in spring. Moreover, in winter, we obtain a width of the DCM less than that of the summer period. Finally, we note that the position of picophytoplankton concentration peaks changes during the whole solar year, due to the seasonal variations of incident light intensity.

We recall that the position, magnitude and shape of phytoplankton distributions depend on three environmental parameters: the incident light intensity at the sea surface, the nutrient concentration at the seabed and the vertical turbulent diffusivity along water column. In particular, since the seasonal variations influence the values of the incident light intensity, the dynamics of picophytoplankton groups can pass from *deep chlorophyll maximum* to *upper chlorophyll maximum* (UCM) stability and *vice-versa* [2]. This behaviour occurs in marine ecosystems, such that analyzed in this work, close to a border between eutrophic and oligotrophic waters. Moreover, the vertical turbulent diffusivity could change due to random modifications of the environmental variables, *i.e.* velocity field, salinity and temperature, inducing a faster transition from one chlorophyll concentration profile to another [2, 35]. In our study, we observe also that the *Prochlorococcus* concentration reaches higher values close to the water surface, as the noise intensity increases, with the system approaching the UCM configuration (see Fig. 4). A stochastic model, where seasonal variations are considered, allows therefore to take into account more possible scenarios for the spatio-temporal dynamics of phytoplankton biomass in real marine ecosystems.

4. Conclusions

In this work, we studied a stochastic model for the spatio-temporal dynamics of two picophytoplankton populations, *i.e.* picoeukaryotes and *Prochlorococcus*, which tend to occupy different zones of the water column. In particular, the model was devised to investigate the picophytoplankton dynamics in a site of the Sicily Channel, where the waters are prevalently oligotrophic and the climatic parameters are those typical of a temperate region. In order to better reproduce the experimental profile of the total *chl a*

and *Dvchl a* concentration, we took in account the effects of the random perturbations due to environmental variables.

The numerical results showed that the presence of a noise source, which acts directly on the dynamics of the nutrient, allows to simulate the average stationary profile of the total *chl a* and *Dvchl a* concentration, obtaining a better agreement with the experimental findings respect to the deterministic case. In particular, the χ^2 goodness-of-fit test indicated that the position and magnitude of the DCM are coherent with those of the real data. In addition, the results obtained agree with experimental data reported in Refs. [30, 37]. Finally, we observed that the external random perturbations can give rise to two interesting effects on phytoplankton dynamics: (i) “shift” of the peak of biomass concentrations towards a greater depth; (ii) “disappearance” of picoeukaryotes and *Prochlorococcus* for higher noise intensity.

We completed our study by analyzing the role of the seasonal oscillations of the light intensity, simulating the spatio-temporal dynamics of the picophytoplankton concentrations during the solar year. The numerical results obtained in the presence of periodical driving force could be useful to predict and understand future changes in the phytoplankton profiles in the Mediterranean Sea, contributing to prevent in marine ecosystem the decline of the phytoplankton production and the consequent decrease of fish species [2, 39–41].

REFERENCES

- [1] J. Huisman, N.N. Pham Thi, D.M. Karl, B. Sommeijer, *Nature* **439**, 322 (2006).
- [2] A.B. Ryabov, L. Rudolf, B. Blasius, *J. Theor. Biol.* **263**, 120 (2010).
- [3] D. Valenti *et al.*, *Acta Phys. Pol. B* **43**, 1227 (2012).
- [4] G. Denaro *et al.*, *Ecol. Complex.* **13**, 21 (2013).
- [5] B.T. Grenfell *et al.*, *Nature* **394**, 674 (1998).
- [6] C. Zimmer, *Science* **284**, 83 (1999).
- [7] O.N. Bjørnstad, B.T. Grenfell, *Science* **293**, 638 (2001).
- [8] B. Spagnolo, M. Cirone, A. La Barbera, F. de Pasquale, *J. Phys. Condens. Mat.* **14**, 2247 (2002).
- [9] A. La Barbera, B. Spagnolo, *Physica A* **314**, 120 (2002).
- [10] B. Spagnolo, A. La Barbera, *Physica A* **315**, 114 (2002).
- [11] B. Spagnolo, A. Fiasconaro, D. Valenti, *Fluct. Noise Lett.* **3**, L177 (2003).
- [12] B. Spagnolo, D. Valenti, A. Fiasconaro, *Math. Biosci. Eng.* **1**, 185 (2004).
- [13] B. Spagnolo, D. Valenti, A. Fiasconaro, *Prog. Theor. Phys. Supp.* **157**, 312 (2005).
- [14] A. Caruso *et al.*, *Fluct. Noise Lett.* **5**, L349 (2005).

- [15] O. Chichigina, D. Valenti, B. Spagnolo, *Fluct. Noise Lett.* **5**, L243 (2005).
- [16] A. Fiasconaro, D. Valenti, B. Spagnolo, *Eur. Phys. J.* **B50**, 189 (2006).
- [17] D. Valenti, L. Schimansky-Geier, X. Sailer, B. Spagnolo, *Eur. Phys. J.* **B50**, 199 (2006).
- [18] O.A. Chichigina, *Eur. Phys. J.* **B65**, 347 (2008).
- [19] A. La Cognata, D. Valenti, A. Dubkov, B. Spagnolo, *Phys. Rev.* **E81**, 011121 (2010).
- [20] O.A. Chichigina, A. Dubkov, D. Valenti, B. Spagnolo, *Phys. Rev.* **E84**, 021134 (2011).
- [21] D. Valenti, A. Fiasconaro, B. Spagnolo, *Acta Phys. Pol. B* **35**, 1481 (2004).
- [22] A. Fiasconaro, D. Valenti, B. Spagnolo, *Acta Phys. Pol. B* **35**, 1491 (2004).
- [23] G. Bonanno, D. Valenti, B. Spagnolo, *Eur. Phys. J.* **B53**, 405 (2006).
- [24] D. Valenti, B. Spagnolo, G. Bonanno, *Physica A* **382**, 311 (2007).
- [25] D. Valenti *et al.*, *Ecol. Model.* **213**, 449 (2008).
- [26] J. Garcia Lafuente *et al.*, *Fish Oceanogr.* **11**, 31 (2002).
- [27] A. Cuttitta *et al.*, *Hydrobiologia* **503**, 117 (2003).
- [28] M. Ribera d'Alcalà, G. Civitarese, F. Conversano, R. Lavezza, *J. Geophys. Res.* **108**, 8106 (2003).
- [29] B. Patti *et al.*, *Sci. Mar.* **74**, 577 (2010).
- [30] C. Brunet *et al.*, *Aquat. Microb. Ecol.* **44**, 127 (2006).
- [31] J. Norberg, *Limnol. Oceanogr.* **49**, 1269 (2004).
- [32] B.B. Prézelin, M.M. Tilzer, O. Schofield, C. Haese, *Aquat. Sci.* **53**, 136 (1991).
- [33] J. Huisman, F.J. Weissing, *Am. Nat.* **146**, 536 (1995).
- [34] C.A. Klausmeier, E. Litchman, S.A. Levin, *J. Theor. Biol.* **246**, 278 (2007).
- [35] A.B. Ryabov, *Theor. Ecol.* **5**, 373 (2012).
- [36] A. Giuffrida *et al.*, *Eur. Food Res. Technol.* **228**, 767 (2009).
- [37] C. Brunet, R. Casotti, V. Vantrepotte, F. Conversano, *Mar. Ecol. Prog. Ser.* **346**, 15 (2007).
- [38] C. Dimier, C. Brunet, R. Geider, J. Raven, *Limnol. Oceanogr.* **54**, 823 (2009).
- [39] L. Bopp *et al.*, *Global Biogeochem. Cy.* **15**, 81 (2001).
- [40] J.L. Sarmiento *et al.*, *Global Biogeochem. Cy.* **18**, GB3003 (2004).
- [41] A. Schmittner, *Nature* **434**, 628 (2005).