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Pollen morphology of the Central European broomrapes (Orobanchaceae: Orobanche, Phelipanche and Orobanchella) and its taxonomical implications

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Abstract Pollen grains of 450 samples of 25 species of the genus Orobanche and Phelipanche occurring in Central Europe were investigated using light and scanning electron microscopy. Palynological data on 18 species are reported here for the first time. The usefulness of micromorphological studies on pollen of Orobanche and Phelipanche is demonstrated. Previous research showed the separation of Phelipanche from Orobanche, a finding which is also supported by pollen aperture type and exine ornamentation. The pollen of Phelipanche is tricolpate, while that of Orobanche is inaperturate. Our research showed that one of the species included so far within Orobanche, namely O. coerulescens, has tricolpate pollen with microreticulate sculpture. As a consequence of the finding of tricolpate pollen and intermediate macromorphological characteristics of this species between Orobanche and Phelipanche, it is proposed to transfer it to a new genus, Orobanchella, described in the paper. A comparison of the main characteristics distinguishing Orobanche, Phelipanche, Boulardia and Orobanchella is also presented and discussed.

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Introduction

The family Orobanchaceae contains 15 obligate parasitic genera with ca 250 species (Pusch and Günther 2009). The family has a worldwide distribution, but the main centres of distribution are the Mediterranean, western and central Asia, northern Africa and North America (Kreutz 1995). Orobanche s.l. is the largest genus and comprises more than 200 species, which lack chlorophyll and are holoparasites of the roots of other vascular plants (Uhlich et al. 1995; Pusch and Günther 2009). In Europe, these species usually grow in the warmest regions, with the highest species diversity in the Mediterranean countries. In central and northern parts of Europe the genus comprises about 30 species, which are mostly rare, endangered or declining (e.g. Zázvorka 1997, 2000; Pusch and Günther 2009; Halamski and Piwowarczyk 2008; Piwowarczyk and Przemyski 2009, 2010; Piwowarczyk et al. 2009, 2010, 2011; Piwowarczyk 2011, 2012a, b, c, d, e, f, g, h, 2013; Nobis et al. 2014).

The genus Orobanche has traditionally been divided into four sections: Trionychon, Orobanche, Gymnocaulis and Myzorrhiza (Beck von Mannagetta 1930). In the most recent taxonomic treatments, these sections have been recognised as separate genera: Aphyllon, Myzorrhiza, Phelipanche, Boulardia and Orobanche. However, the classification of Phelipanche (syn. O. sect. Trionychon) and Boulardia (syn. Orobanche latisquama) as separate genera is still being discussed (Holub 1977, 1990; Teryokhin et al. 1993; Schneeweiss 2001; Manen et al. 2004; Schneeweiss et al. 2004a, b; Weiss-Schneeweiss et al.

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2006; Bennett and Mathews 2006; Schneeweiss 2007; Park et al. 2007a, b; Pujadas Salvà 2007; Carlón et al. 2008). Some authors still treat the genus Orobanche s.l. in a wide sense (Greuter and Raab-Straube 2009). The separation of Phelipanche and Orobanche is confirmed by recent molecular phylogenetic studies (e.g. Schneeweiss et al. 2004a, b; Manen et al. 2004; Park et al. 2007a, b); morphological and carpological differences (Holub 1977; Teryokhin 1997; Joel 2009a, b); chemotaxonomic study (Velasco et al. 2000) and micromorphology of seeds (e.g. Abu Sbaih and Jury 1994; Plaza et al. 2004); and pollen grains (Zare et al. 2013). According to the authors of a recent study (Zare et al. 2013), verrucate exine ornamentation and inaperturate pollen type suggest a close relationship between Orobanche, Boulardia and Diphelypaea, whereas tricolpate pollen suggests a relationship between Cistanche and Phelipanche.

The flowers in Orobanche are always bisexual, with superior ovaries, more or less zygomorphic, with four didynamous stamens and two or three lobed stigma, characterised by cross-pollination or, rarely, by the absence of insects, i.e. self-pollination (Beck von Mannagetta 1930; Teryokhin 1997; Jones 1991). There are many adaptations of flowers for pollination: large or small flowers gathered in dense inflorescences; contrasting colouration and shine of the corolla and stigma; a wide range of often bright colours; the smell of the flowers, either pleasant (scent of cloves, e.g. O. alba, O. caryophyllacea, O. crenata, O. gracilis) or unpleasant (the smell of carrion, e.g. O. foetida, O. rigens, O. rapum-genistae, O. variegata) (the majority of species have a weak scent or none); the lower lip of the corolla serving as an alighting place for pollinators (two bright folds, coated with hairs); the production of nectar from coloured spots. Most species are pollinated by bumblebees and bees (Apidae), as well as, albeit less importantly, by species of the families Syrphidae, Vespidae, or Formicidae or carrion flies (Diptera) (Piwowarczyk, unpublished). Preliminary studies in relation to pollination and other shared interactions between parasitic plants and their hosts are presented by Ollerton et al. (2007).

Orobanchaceae is a eurypalynous family, with different types of pollen. Nevertheless, in many cases the biometric and morphological features of pollen grains possess a high diagnostic significance and are used in taxonomy. Pollen taxonomy can also explain the relationship and phylogenetic connections between particular genera of the family. Exine sculpture is one of the most important features and is a good tool in the recognition of genera or sections within a genus. Unfortunately, there are still not many papers dedicated to the pollen morphology of the family Orobanchaceae (e.g. Tiagi 1951; Rao 1963; Erdtman 1966; Polo 1987; Minkin and Eshbaugh 1989; Abu Sbaih et al. 1994; Teryokhin 1997; Shahi Shavvon and Saeidi Mehrvarz 2010; Zare and Dönmez 2013; Zare et al. 2013; Piwowarczyk et al. 2014) and further studies within the whole family are required.

The aim of the study was to present (1) the macro- and micromorphological variability of pollen in the Central European *Orobanche* and *Phelipanche* taxa; (2) the taxonomic implications of pollen morphology in the examined taxa.

Materials and methods

Plant material

This study is based on plant material deposited mainly in the KTC herbarium. In total, 25 taxa of three genera, *Orobanche, Phelipanche* and *Orobanchella*, were analysed. A list of voucher specimens used in the study is given in Table 1. Each sample corresponds to a single plant, but pollen grains originating from different flowers have not been distinguished.

Light microscopy observations

The pollen grains were prepared with the standard method, Erdtman's acetolysis (Erdtman 1969; Faegri and Iversen 1989). Following acetolysis, the pollen grains were mounted in glycerine jelly and prepared for LM observation. Some characteristics, such as polar and equatorial axis and exine thickness, were examined by LM (Axio Imager 2, Zeiss) for 30 pollen grains under $1,000 \times$ magnifications with the help of the AxioVision computer program (version 4.8.1.0, Carl Zeiss Imaging Solutions). Measurements were taken under the light microscope only in accordance with changes in pollen shape and dimensions occurring during its preparation for SEM.

Scanning electron microscopy (SEM) observation

Using SEM observation, we analysed 450 samples of 25 taxa. For SEM observation, samples were coated with gold using a JFC-1100E Ion Sputter manufactured by JEOL. Micromorphological structures of pollen grains were observed and photographs taken by means of Hitachi S-4700 and Philips XL 20 scanning electron microscopes at various magnifications. The terminology was adopted from Punt et al. (2007) and Hesse et al. (2009).

Morphometric analysis

The statistical analyses are based on 1,170 fully-developed pollen grains from 25 taxa. Characteristics were measured in 30 grains of each specimen studied (Table 1). Finally, a

Table 1	Taxa	used	in	this	study	and	voucher	information
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Number	Taxon	Locality	Voucher	Host
	Orobanche			
1	O. alba subsp. alba	Poland	Bieszczady Mts, 14 Aug 2009, R. Piwowarczyk (KTC)	Thymus pulegioides
2a	O. alba subsp. major	Poland	Lasocin, 27 Jul 2006, R. Piwowarczyk (KTC)	Salvia verticillata
2b	O. alba subsp. major		Katy II, 15 Jul 2006, R. Piwowarczyk (KTC)	Salvia verticillata
3	O. alsatica	Poland	Broczówka, 8 Jun 2007, R. Piwowarczyk (KTC)	Peucedanum cervaria
4	O. bartlingii	Poland	Podzamcze, 4 Jun 2009, R. Piwowarczyk (KTC)	Libanotis pyrenaica
5a	O. caryophyllacea	Poland	Teresin, 8 Jun 2007, R. Piwowarczyk (KTC)	Galium mollugo
5b	O. caryophyllacea		Bogucice, 6 Jun 2007, R. Piwowarczyk (KTC)	Galium odoratum
5c	O. caryophyllacea		Pieniny Mts, Kozia Mt, 23 Jul 2010, R. Piwowarczyk (KTC)	Galium mollugo
6	O. crenata	Spain	Cordoba, 2 May 2012, R. Piwowarczyk (KTC)	Vicia sp.
7a	O. elatior	Poland	Baldram, 07 Jul 2010, R. Piwowarczyk (KTC)	Centaurea scabiosa
7b	O. elatior		Wólka Leszczańska, 08 Jun 2008, R. Piwowarczyk (KTC)	Centaurea scabiosa
8a	O. flava	Poland	Tatra Mts, Mała Łąka Valley, 22 Jun 2009, R. Piwowarczyk (KTC)	Petasites kablikianus
8b	O. flava	Slovakia	Nizkie Tatra Mts, 7 Aug 2011, R. Piwowarczyk (KTC)	Petasites kablikianus
9	O. gracilis	Austria	Hundsheim, 14 Jun 2012, R. Piwowarczyk (KTC)	Anthyllis vulneraria
10	O. hederae	Spain	Elx, palm garden, 28 Apr 2009, R. Piwowarczyk (KTC)	Hedera helix
11a	O. kochii	Poland	Pęczelice, 13 Jun 2010, R. Piwowarczyk (KTC)	Centaurea scabiosa
11b	O. kochii		Toporowice, 5 Jun 2008, R. Piwowarczyk (KTC)	Centaurea scabiosa
12	O. lucorum	Poland	Warsaw, botanical garden, 10 July 2009, R. Piwowarczyk (KTC)	Berberis vulgaris
13a	O. lutea	Poland	Wesołówka, 17 Jun 2006, R. Piwowarczyk (KTC)	Medicago falcata
13b	O. lutea		Chomentówek, 6 Jun 2007, R. Piwowarczyk (KTC)	Medicago sativa
13c	O. lutea		Młodzawy Duże, 2 Jun 2007, R. Piwowarczyk (KTC)	Medicago falcata
14	O. mayeri	Poland	Pieniny Mts, Trzy Korony, 30 Jun 2009, R. Piwowarczyk (KTC)	Laserpitium latifolium
15	O. minor	Poland	Żywiec, 19 Jun 2009, R. Piwowarczyk (KTC)	Trifolium repens
16a	O. pallidiflora	Poland	Bieszczady Mts, 14 Aug 2009, R. Piwowarczyk (KTC)	Carduus personata
16b	O. pallidiflora		Lubiatowo, 24 Jun 2009, R. Piwowarczyk (KTC)	Cirsium oleraceum
16c	O. pallidiflora		Ciepłe, 5 Jul 2010, R. Piwowarczyk (KTC)	Cirsium arvense
16d	O. pallidiflora		Potok Mały, 29 Jun 2009, R. Piwowarczyk (KTC)	Cirsium arvense
17a	O. picridis	Poland	Pasturka, 25 Jun 2007, R. Piwowarczyk (KTC)	Picris hieracioides
17b	O. picridis		Machnów Stary, 9 Jun 2007, R. Piwowarczyk (KTC)	Picris hieracioides
18	O. reticulata	Slovakia	Nizkie Tatra Mts, Puste, 5 Aug 2011, R. Piwowarczyk (KTC)	Carduus glaucus
19	O. teucrii	Austria	Hundsheim, 14 Jun 2012, R. Piwowarczyk (KTC)	Teucrium montanum
	Phelipanche			
20a	P. arenaria	Poland	Zawiercie-Bzów, 11 Jun 2010, R. Piwowarczyk (KTC)	Artemisia campestris
20b	P. arenaria		Pasturka, 25 Jun 2007, R. Piwowarczyk (KTC)	Artemisia campestris
21	P. bohemica	Poland	Zawiercie-Bzów, 11 Jun 2010, R. Piwowarczyk (KTC)	Artemisia campestris
22	P. caesia	Ukraine	Askania Nova, 16 Jun 2011, R. Piwowarczyk (KTC)	Artemisia austriaca
23	P. purpurea	Poland	Chrzanów, 18 Jun 2009, R. Piwowarczyk (KTC)	Achillea millefolium s.l.
24	P. ramosa	Poland	Zysławice, 7 Sep 2008, R. Piwowarczyk (KTC)	Nicotiana tabacum
	Orobanchella			
25a	O. coerulescens	Poland	Dobrowoda, 19 Jun 2010, R. Piwowarczyk (KTC)	Artemisia campestris
25b	O. coerulescens		Pasturka, 25 Jun 2007, R. Piwowarczyk (KTC)	Artemisia campestris

total of the seven most informative quantitative and qualitative morphological characteristics was chosen for the analyses: width of exine (mean μ m); polar axis (mean μ m); equatorial diameter (mean μ m); polar/equatorial ratio; number of furrows on grain; sculpture of pollen grains (1—granules look like a ball of wool, 2—granules consisted of a few smaller round elements knitted together into a smooth surface, 3—microreticulate); arrangement of granules on the pollen surface: 1—no areas without granules are present, 2—areas without granules are present, **Table 2** Characters used incluster analysis (comp.

"Materials and methods")

Species	Characte	er					
	Polar axis (mean µm)	Equatorial diameter (mean µm)	Width of exine (mean µm)	Polar/ equatorial ratio	Furrows number on grain	Sculpture of pollen grains	Arrangement of granules
Orobanchella coerulescens	20.4	21.9	1.4	0.934	3	3	3
Phelipanche arenaria	22.1	24.5	1.4	0.903	3	3	3
P. bohemica	22.9	24.2	1.7	0.944	3	3	3
P. caesia	25.0	23.8	1.3	1.053	3	3	3
P. purpurea	21.6	22.1	1.5	0.977	3	3	3
P. ramosa	19.3	21.3	1.4	0.905	3	3	3
Orobanche alba subsp. alba	22.6	19.8	0.9	1.142	0	1	2
O. alba subsp. major	22.5	18.9	0.9	1.191	0	1	2
O. alsatica	27.5	22.6	0.9	1.219	0	2	2
O. bartlingii	26.3	22.4	0.8	1.174	0	2	2
O. caryophyllacea	25.0	21.7	1.1	1.152	0	2	2
O. crenata	20.2	18.2	0.9	1.110	0	2	2
O. elatior	26.2	22.9	1.1	1.144	0	2	2
O. flava	27.5	23.7	1.1	1.163	0	2	1
O. gracilis	23.6	20.3	0.9	1.165	0	2	2
O. hederae	24.8	21.3	1.1	1.162	0	1	2
O. kochii	25.1	22.2	1.1	1.130	0	2	2
O. lucorum	27.4	23.7	1.0	1.157	0	2	1
O. lutea	23.8	20.8	0.8	1.144	0	2	2
O. mayeri	28.8	24.2	1.0	1.189	0	2	2
O. minor	23.3	19.6	0.8	1.188	0	2	1
O. pallidiflora	23.9	20.8	0.8	1.151	0	2	2
O. picridis	23.9	20.0	0.9	1.195	0	2	2
O. reticulata	27.0	23.2	0.9	1.165	0	2	2
O. teucrii	26.3	22.6	0.8	1.162	0	2	2

3—no granules are present (different type of pollen sculpture). The characteristics used are listed in Table 2. The mean value of the characteristics measured or scored from all samples of the taxon was analysed using cluster analysis to illustrate the general relationships and similarities between them. The analysis was performed in order to obtain results providing a general overview and to determine whether different morphological groups exist and are separated into distinct clusters. Similarity between samples was calculated on the basis of Gower's general similarity coefficient. The dendrogram was prepared using the UP-GMA method. Subsequently, principal coordinate analysis (PCoA) was performed on the basis of all quantitative and qualitative features. The goal of PCoA was the positioning of objects (individuals) in a space of reduced dimensionality while preserving their distance relationships. Data analyses were performed using MVSP 3.1 (Kovach 1999).

Results

LM and SEM study

Both LM and SEM observations enabled the team to distinguish, among *Orobanche* and *Phelipanche* pollen, two main groups with distinctly different features regarding pollen shape and exine micromorphology.

Group 1 contains pollen grains in monads, suboblate (*P. arenaria*), oblate spheroidal (*P. purpurea*, *P. bohemica*, *P. ramosa* and *Orobanche coerulescens*) (according to P/E

axon	Number	Polar a:	xis (µm)	_		Equator	ial dian	heter (µr	n)	P/E	Number of	Exine (mean,	Pollen shape (acc. to Erdtman	Exine
		Min	Max	Mean	±SD	Min	Max	Mean	±SD	ratio	colpi	(mm)	1966)	sculpture
helipanche arenaria	20a	19.85	24.95	22.24	1.37	21.46	28.22	25.11	1.66	0.89	3 (2)	1.32	Oblate spheroidal	Microreticulate
helipanche arenaria	20b	18.84	25.61	21.53	1.42	19.91	29.16	25.73	1.92	0.84	3	1.40	Suboblate	Microreticulate
helipanche bohemica	21	20.82	28.46	22.89	1.56	23.46	27.43	25.09	0.87	0.91	3 (2.4)	1.67	Oblate spheroidal	Microreticulate
helipanche caesia	22	21.01	28.89	25.01	1.34	22.00	26.24	23.75	1.21	1.05	3	1.28	Prolate spheroidal	Microreticulate
helipanche purpurea	23	19.35	24.49	21.60	1.20	20.31	25.77	23.15	1.31	0.93	3	1.46	Oblate spheroidal	Microreticulate
helipanche ramosa	24	17.10	22.68	19.29	1.14	19.22	23.87	21.87	1.33	0.89	3	1.38	Oblate spheroidal	Microreticulate
)robanchella coerulescens	25a	17.57	22.95	21.00	1.43	20.81	26.25	23.30	1.49	06.0	3 (2)	1.32	Oblate spheroidal	Microreticulate
)robanchella coerulescens	25b	16.49	24.82	20.22	2.03	17.66	24.52	22.19	1.73	0.91	3 (4)	1.38	Oblate spheroidal	Microreticulate

Fable 3 Characters of tricolpate *Phelipanche*-like type pollen morphology

ratio) and prolate spheroidal (*P. caesia*), and tricolpate (but some single grains with 2 or 4 colpi) forms. Some differences in colpi shape and distribution are visible (colpi were sometimes fused, wide at the equator and narrow near the poles) but these features appear in all members of group 1 (Tables 2, 3; Figs. 3, 5).

Group 2 contains pollen grains in monads, prolate spheroidal (*O. kochii* and *O. crenata*) to subprolate (other) and inaperturate forms, or with irregular colpi. The basic exine ornamentation type is granulate. Visible differences in the size of granules and their distribution are present between some species. Three types of granule surface were distinguished: (1) granules with almost smooth surfaces (*O. caryophyllacea*, *O. flava*, *O. elatior*, *O. lucorun*); (2) granules consisting of a few smaller round elements knitted together (*O. caryophyllacea*, *O. picridis*, *O. minor*, *O. alba* subsp. *major*, *O. kochii*, *O. lutea*, *O. reticulata*, *O. teucrii*, *O. gracilis*, *O. crenata*); and (3) granules consisting of elongated elements visible on granule surfaces (like balls of wool) (*O. alba* subsp. *alba*, *O. hederae*, *O. kochii*, *O. flava*) (Tables 2, 4; Figs. 1, 2, 4, 5).

According to differences in the size and distribution of granules, two main patterns of pollen surface ornamentation can be found: 1—granules distributed over the whole pollen grain, but small areas without granules are present; the granules are of different sizes but always smaller than 1 μ m; most granules are joined to others, but isolated granules are also present; 2—the surfaces are similar to those described above, but there are no areas without granules (Figs. 1, 2).

Statistical analysis

Cluster analysis (UPGMA) performed on the complete set of data (Table 2) resulted in the delimitation of two main subclusters. The first comprises a group of taxa with distinctly tricolpate pollen grains with reticulate surfaces, whereas the second comprises uncolpate grains with granulate surfaces (Figs. 3, 4, 5, 6). A similar pattern of results is shown by the ordination diagrams from the principal coordinate analysis (PCoA, not shown), also based on the complete quantitative and qualitative set of data. The first subcluster in UPGMA effectively separates the species P. ramosa, P. bohemica, P. arenaria, P. purpurea, P. caesia and Orobanchella coerulescens from all the rest of the specimens belonging to Orobanche placed in the second cluster. In the Phelipanche subcluster, all taxa are very similar in general habit, with tricolpate pollen grains. In cluster analysis, P. caesia differs slightly from the other members, with a higher polar/equatorial ratio and a thinner exine. However, pollen characteristics in this group of species are characterised by generally higher variance (Fig. 7). Although all the species located in the

Table 4 Characters of inaperturate Orobanche-like type pollen morphology

Taxon	Number	Polar axis (µm)			Equato	Equatorial diameter (µm)				Exine	Pollen shape (acc.	Exine	
		Min	Max	Mean	±SD	Min	Max	Mean	±SD	ratio	(mean, µm)	to Erdtman 1966)	sculpture
Orobanche alba subsp. alba	1	17.44	26.77	22.62	2.20	15.88	22.21	19.80	1.75	1.15	0.86	Subprolate	Granulate
Orobanche alba subsp. major	2a	18.72	29.79	23.10	2.86	16.24	23.86	19.11	1.92	1.21	0.89	Subprolate	Granulate
Orobanche alba subsp. major	2b	17.94	25.52	21.84	1.77	14.11	21.33	18.63	1.92	1.18	0.84	Subprolate	Granulate
Orobanche alsatica	3	21.25	33.45	27.50	3.12	15.46	29.08	22.57	2.86	1.23	0.87	Subprolate	Granulate
Orobanche bartlingii	4	21.32	30.84	26.33	2.51	16.72	28.30	22.43	2.71	1.18	0.77	Subprolate	Granulate
Orobanche caryophyllacea	5a	21.20	28.06	24.52	1.44	18.36	23.65	21.29	1.30	1.15	1.20	Subprolate	Granulate
Orobanche caryophyllacea	5b	19.68	30.21	25.44	2.55	17.24	27.79	22.10	2.65	1.16	1.04	Subprolate	Granulate
Orobanche caryophyllacea	5c	19.83	28.00	23.17	1.99	15.22	24.67	19.50	2.03	1.19	1.09	Subprolate	Granulate
Orobanche crenata	6	16.37	23.28	20.19	1.94	13.79	20.99	18.18	1.93	1.11	0.85	Prolate spheroidal	Granulate
Orobanche elatior	7a	21.52	30.83	25.91	2.44	17.71	27.52	22.67	2.48	1.15	1.12	Subprolate	Granulate
Orobanche elatior	7b	20.30	34.47	26.44	3.75	17.52	32.19	23.07	3.78	1.15	1.11	Subprolate	Granulate
Orobanche flava	8a	18.90	30.42	24.88	2.59	15.78	25.06	21.40	2.33	1.17	1.09	Subprolate	Granulate
Orobanche flava	8b	24.09	35.11	30.11	2.88	22.17	31.97	26.01	2.74	1.16	1.16	Subprolate	Granulate
Orobanche gracilis	9	17.38	27.39	23.59	2.14	13.82	24.16	20.25	2.09	1.17	0.86	Subprolate	Granulate
Orobanche hederae	10	21.59	30.67	24.77	2.40	17.21	25.30	21.31	2.02	1.17	1.11	Subprolate	Granulate
Orobanche kochii	11a	19.91	27.50	23.84	1.95	17.31	24.83	21.21	2.04	1.14	1.13	Subprolate	Granulate
Orobanche kochii	11b	22.17	32.22	26.26	2.81	18.56	29.72	23.15	2.46	1.14	1.11	Subprolate	Granulate
Orobanche lucorum	12	21.11	35.52	27.39	3.32	19.00	29.95	23.67	3.16	1.16	1.05	Subprolate	Granulate
Orobanche lutea	13a	18.36	26.78	22.26	2.36	15.54	22.33	19.31	1.71	1.15	0.75	Subprolate	Granulate
Orobanche lutea	13b	20.71	29.98	25.43	2.25	15.67	25.70	22.48	2.29	1.15	0.77	Subprolate	Granulate
Orobanche lutea	13c	22.19	39.21	29.16	3.39	16.97	31.17	24.86	3.12	1.18	1.02	Subprolate	Granulate
Orobanche mayeri	14	25.05	34.69	28.77	2.66	19.55	29.50	24.19	2.46	1.19	1.04	Subprolate	Granulate
Orobanche minor	15	18.08	27.80	23.28	1.93	15.60	24.30	19.59	2.37	1.20	0.78	Subprolate	Granulate
Orobanche pallidiflora	16a	18.75	27.16	22.23	2.15	16.37	23.90	19.43	2.03	1.15	0.70	Subprolate	Granulate
Orobanche pallidiflora	16b	19.95	30.96	26.11	2.50	17.61	27.19	23.08	2.14	1.14	0.84	Subprolate	Granulate
Orobanche pallidiflora	16c	19.52	31.36	24.08	2.39	16.85	25.60	21.07	2.15	1.16	0.84	Subprolate	Granulate
Orobanche pallidiflora	16d	18.03	28.42	23.02	3.10	14.71	24.19	19.55	2.49	1.18	0.70	Subprolate	Granulate
Orobanche picridis	17a	18.59	33.38	22.80	3.06	15.12	23.49	19.50	2.27	1.17	0.87	Subprolate	Granulate

Table 4 continued

Taxon	Number	Polar a	axis (µrr	ı)		Equato	orial diar	neter (µ	m)	P/E	Exine	Pollen shape (acc.	Exine
		Min	Max	Mean	±SD	Min	Max	Mean	±SD	ratio	(mean, µm)	to Erdtman 1966)	sculpture
Orobanche picridis	17b	18.92	33.61	25.01	3.49	13.34	25.45	20.52	2.70	1.23	0.88	Subprolate	Granulate
Orobanche reticulata	18	19.56	31.04	27.00	2.63	19.35	27.83	23.17	1.84	1.19	0.85	Subprolate	Granulate
Orobanche teucrii	19	22.67	31.29	26.31	1.86	16.09	25.75	22.63	2.12	1.17	0.83	Subprolate	Granulate



Fig. 1 Micromorphology of granules in selected species of Orobanche: a Orobanche hederae (granules like balls of wool), b O. reticulata (granules consisted of a few smaller round elements knitted together), c O. flava (granules glabrous or almost so)



Fig. 2 Arrangement of granules on the pollen surface: a areas without granules not present (Orobanche flava), b bald areas without granules present (O. reticulata)

Orobanche subclade are very similar according to the general habit of pollen grains, two smaller subgroups of taxa can be distinguished. They differ mainly in the size of grains and width of exines (cf Fig. 8). In the first group, which comprises Orobanche alba s. lato, O. crenata, O. gracilis, O. hederae, O. lutea, O. minor, O. pallidiflora and O. picridis, the mean polar axis of the pollen grains is <23.9, the equatorial mean is <20.8 and the exine is generally narrower (0.8–0.9 μ m). In all the remaining taxa from the subclade, the mean polar axis of pollen grains

varies from 23.3 to 28.8, the equatorial mean is 21.3-24.2 and the exine width is (0.8-) 0.9-1.1 µm (Table 2; Fig. 8).

In the *Phelipanche* group, a thinner exine has been noted in P. caesia, but at the same time, it has the highest value for equatorial and polar axis. The largest pollen grains in the Orobanche group were noted e.g. in O. elatior, O. flava or O. lucorum, the smallest in e.g. O. crenata and O. picridis. The thickest exines were recorded in O. caryophyllacea, O. elatior and O. flava, and the thinnest in O. lutea and O. bartlingii (Tables 2, 3, 4).



Fig. 3 Pollen grains and its micromorphology in: Orobanchella coerulescens (1), Phelipanche caesia (2), P. arenaria (3), P. ramosa (4); a general habit of pollen grain (magnification ×3,000), b, c microstructure of pollen grain (magnification ×10,000 and ×30,000)

Discussion

Analysis of the pollen grains of Central European representatives of *Orobanche* s.l. showed that they are characterised by high variability, both inter- and intra-specific, as well as in relation to oligo- and polyphagous species parasitic on various hosts. In addition, some taxa may produce heteromorphic pollen grains (Zare et al. 2013), but this issue requires further study. Surely pollen characteristics, i.e. the number of apertures and the ornamentation of exines, cannot be overlooked, because they bear valuable information to help analyse evolutionary relationships and classifications within the Orobanchaceae family.

The characteristics of pollen are important criteria for distinguishing genera or sections, but are less important in distinguishing species. However, it is worth mentioning the subtle yet noticeable differences in the size and sculpture of pollen of species treated in different taxonomic units, i.e. species of the problematic complex *O. alsatica* s.l., namely *O. alsatica*, *O. bartlingii* and *O. mayeri*, all three of which



Fig. 4 Pollen grains and its micromorphology in: Orobanche caryophyllacea (1), O. alba subsp. alba (2), O. lutea (3), O. flava (4); **a** general habit of pollen grain (magnification ×3,000), **b**, **c** microstructure of pollen grain (magnification ×10,000 and ×30,000)

are parasites of Apiaceae (Piwowarczyk 2011, 2012c; Piwowarczyk et al. 2014). In another example, *O. pallidiflora*, the species is treated differently in various studies: as a separate taxon, or in the rank of subspecies or variety within *Orobanche reticulata* (Beck von Mannagetta 1890, 1930; Zázvorka 2000; Pusch and Günther 2009). *O. reticulata* and *O. pallidiflora* differ in morphology, habitat and altitudinal ranges as well as host preferences (e.g. Kreutz 1995; Piwowarczyk et al. 2010). Our study showed that *O. pallidiflora* has thinner exines and smaller equatorial and polar axes than *O. reticulata*, along with differences in sculpture. *Orobanche bohemica* as a separate species was first described by Čelakovský (1879). It has been assigned to different taxonomic ranks, e.g., that of a variety, by Beck von Mannagetta (1930) and Hayek (1914); as a subspecies of *Phelipanche purpurea*, by e.g. Zázvorka (2000) and Carlón et al. (2008); or as a separate species, by Holub and Zázvorka (1999), Pusch (2006), Pusch and Günther (2009), and



Fig. 5 Pollen grains of Orobanchella coerulescens (a), Phelipanche caesia (b), P. arenaria (c), P. ramosa (d), Orobanche caryophyllacea (e), O. alba subsp. alba (f), O. lutea (g) and O. flava (h). Scale bar 5 µm



Fig. 6 Cluster analysis (UPGMA method of classification and Gower's general similarity coefficient) performed on seven quantitative and qualitative characters of pollen grains

Piwowarczyk (2012f, g). Palynological analysis also revealed auxiliary micromorphological differences: *P. bohemica* has a thicker exine and larger equatorial and polar axes than *P. purpurea*, along with differences in sculpture. A Central European species, parasitic mainly on *Centaurea*, considered until recently as *O. elatior* s.l. and now recognised as two distinct species, *O. elatior* and *O. kochii* (Zázvorka 2010), also demonstrates subtle differences in the size and level of sculpture of pollen grains (Tables 2, 3, 4).

Our study of pollen morphology showed various characteristics of Orobanche coerulescens. O. coerulescens has a Eurasian distribution. It occurs in the Far East, Japan, China, the Caucasus, Latvia, Ukraine and Romania. Very rare in Central Europe, it is known from Slovakia, the Czech Republic, Austria, Germany and Poland. As a typical steppe plant, it has relict features of post-glacial Artemisia steppes in the western part of its range. The species is recognised as extinct in the majority of its localities at the western limit of its distribution and its populations are very scarce in Central Europe. It is mainly parasitic on Artemisia campestris (Zázvorka 2000; Pusch and Günther 2009; Piwowarczyk 2012d). Traditionally, the species was included in the section Orobanche, grex Coerulescentes Beck (Beck von Mannagetta 1890), or, according to the new classification, in the section Inflatae (Beck) Tzvel., subsection Coerulescentes (Beck) Teryokhin (Teryokhin et al. 1993). This subsection contains about 10 species, poorly known and often lacking a very clear taxonomic position, of mainly steppe parasites involving the genus Artemisia. They are mostly endemic, occurring mainly in eastern Asia and Russia (Teryokhin et al. 1993; Piwowarczyk 2012d). We have shown that the pattern of pollen grains of O. coerulescens clearly belongs to a Phelipanche tricolpate type of pollen with microreticulate

1,5 1,4

1.3



Fig. 7 Box and whisker plots of selected macromorphological characters of tricolpate *Phelipanche*-like pollen grains. *Points* indicate the mean value (*open square*), *boxes* represent 25 and 75 % percentile, and range (*whiskers*) represents 1 and 99 % percentile. *Numbers* indicated examined species (comp. Table 1)

sculpture (Figs. 3, 5). Previously, microreticulate sculpture of the exine was known only in *Cistanche* and *Phelipanche* (Zare et al. 2013). At the same time this contradicts the classification of the pollen of *O. coerulescens* by Zare et al.



Fig. 8 Box and whisker plots of selected macromorphological characters of inaperturate *Orobanche*-like pollen grains. *Points* indicate the mean value (*open square*), *boxes* represent 25 and 75 % percentile, and range (*whiskers*) represents 1 and 99 % percentile. *Numbers* indicated examined species (comp. Table 1)

(2013) into the inaperturate type. Macromorphological characteristics indicate its intermediate position between *Orobanche* and *Phelipanche*. *O. coerulescens* is quite easy to distinguish from related species because of its characteristic thick stem, especially near the inflorescence, and numerous tiny, geniculately bent, pale blue flowers. The entire plant, the stem in particular, is very hairy and, when dry, looks as if covered with mould (Piwowarczyk and Przemyski 2009). Features that place it within *Orobanche*

Character	Genus											
_	Orobanche	Phelipanche	Boulardia	Orobanchella								
Bracteoles	Absent	2 bracteoles under or attached the calyx	Absent	Absent								
Stem	Simple	Branched or simple	Simple	Simple								
Calyx	Divided into 2 lateral segments	Cylindrical or campanulate with 4–5 teeth	Divided into 2 lateral segments	Divided into 2 lateral segments								
Fruit	Style persists, lateral opening	Style falls off, top opening	Style persists, lateral opening	Style persists, lateral opening								
Colour of flowers	Yellow, red, brown	Usually blue, violet	Violet, purple, rarely white–pink, yellow	Light-blue, blue-violet								
Colour of stigma and anthers	Yellow, orange, purple, brown	Usually white, rare yellow–white	Usually white, rare yellow–white	Usually white, rare yellow-white								
Type of pollen												
Aperture	Inaperturate	Tricolpate	Inaperturate	Tricolpate								
Sculpture of the exine	Granulate	Microreticulate	Granulate	Microreticulate								
Type of seeds	Outer tangential wall pitted or rarely smooth	Wall with fibrillar ornamentation	Wall with fibrillar ornamentation	Periclinal wall granulate rugged or smooth, very rare weakly pitted								
Chromosomes	n = 19	n = 12	n = 38	n = 19								

Table 5 A comparison of the main characters distinguishing Orobanche, Phelipanche, Boulardia and Orobanchella

are: absence of bracteoles, calyx divided into 2 lateral segments, style persist, lateral opening, number of chromosomes n = 19; however, it is close to *Phelipanche* by virtue of such features as: blue or violet flowers, white stigma and anthers, and the presence of tricolpate pollen with microreticulate sculpture (Table 5). In the work of Manen et al. (2004), Schneeweiss et al. (2004b), and Weiss-Schneeweiss et al. (2006), it is shown for the first time to be a member of the Eurasian lineage *O. coerulescens*. Phylogenetic relationships also show a degree of separation of *O. coerulescens* from the rest of the species of the section *Orobanche* and place it in an intermediate position between *Orobanche*, *Boulardia* and *Phelipanche* (Schneeweiss et al. 2004a; Piednoël et al. 2012).

Summarising the above, we proposed to transferred *Orobanche coerulescens* to a new genus *Orobanchella*. A comparison of the main characters distinguishing *Orobanche, Phelipanche, Boulardia* and *Orobanchella* is presented in Table 5.

New genus and combination

Orobanchella R. Piwowarczyk, M. Nobis & J. Madeja gen. nov.

Type: Orobanchella caerulescens (Stephan in Willdenow) R. Piwowarczyk, M. Nobis & J. Madeja

Orobanchella coerulescens (Stephan in Willdenow) R. Piwowarczyk, M. Nobis & J. Madeja **comb. nov**.

Basionym: Orobanche coerulescens Stephan in Willdenow, Sp. Pl. 3(1): 349, 1800.

Type: "Habitat in Sibiria versus Mare Caspium". Willdenow, 1800 (p. 349). According to Novopokrovskij and Tzvelev, 1958 (p. 72), described from area in vicinity of lake Inder "iz raiona Inderskogo ozera" (W Kazakhstan) (type in LE).

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References

- Abu Sbaih HA, Jury SL (1994) Seed micromorphology and taxonomy in *Orobanche* (Orobanchaceae). Fl Medit 4:41–48
- Abu Sbaih HA, Keith-Lucas DM, Jury SL, Tubaileh AS (1994) Pollen morphology of genus *Orobanche L*. (Orobanchaceae). Bot J Linn Soc 116:305–313
- Beck von Mannagetta G (1890) Monographie der Gattung Orobanche. Bibl Bot 19. Theodor Fischer, Cassel
- Beck von Mannagetta G (1930) Orobanchaceae. In: Engler A (ed) Das Pflanzenreich. Regni Vegetabilis Conspectus. Wilhelm Engelmann, Leipzig, pp 1–348
- Bennett JR, Mathews S (2006) Phylogeny of the parasitic plant family Orobanchaceae inferred from phytochrome A. Amer J Bot 93(7):1039–1051

- Carlón L, Gómez Casares G, Laínz M, Moreno Moral G, Sánchez Pedraja Ó, Schneeweiss GM (2008) Más, a propósito de algunas *Phelipanche* Pomel, *Boulardia* F. W. Schultz y *Orobanche* L. (Orobanchaceae) del oeste del Paleártico. Documentos Jard Bot Atlántico (Gijón) 6:1–128
- Čelakovský L (1879) Ueber eine neue oder verkannte Orobanche. Österr Bot Zeitschr 29:361–364
- Erdtman G (1966) Pollen morphology and plant taxonomy, angiosperms. Hafner Publishing Co, New York
- Erdtman G (1969) Handbook of palynology. An introduction to study of pollen grains and spores. Munksgaard, Copenhagen
- Faegri K, Iversen I (1989) Textbook of pollen analysis, 4th edn. In: Faegri K, Kaland PE, Krzywinski K (eds). Wiley, New York
- Greuter W, Raab-Straube E (2009) Euro + Med Notulae, 4. Willdenowia 39:327–333
- Halamski AT, Piwowarczyk R (2008) Graines d'Orobanches comme critere taxonomique-information sur les travaux en cours. Bull Mens Soc Linn Lyon 77(3–4):37–40
- Hayek A (1914) Orobanchaceae. In: Hegi G (ed) Illustrierte Flora von Mitteleuropa, vol 6. München, pp 132–155
- Hesse M, Halbritter H, Zetter R, Weber M, Buchner R, Frosch-Radivo A, Ulrich S (2009) Pollen terminology. An illustrated handbook. Springer Wien, New York
- Holub J (1977) New names in Phanerogamae 6. Folia Geobot Phytotax 12:417–432
- Holub J (1990) Some taxonomic and nomenclatural changes within *Orobanche* s. l. (Orobanchaceae). Preslia 62:193–198
- Holub J, Zázvorka J (1999) Phelipanche bohemica. In: Cerovsky J et al (eds) Cervena kniha ohrozenych a vzacnych druhu rostlin a zivocichu CR a SR, vol 5. Bratislava, p 279
- Joel DM (2009a) Taxonomic and evolutionary justifications for considering *Phelipanche* as a separate genus. In: Ministry of Agriculture and Rural Affairs and Ege University (eds). Proceedings of the International parasitic plant society (IPPS), 10th World Congress of parasitic plants, 8–12 June 2009. Kusadasi, Turkey, pp 152
- Joel DM (2009b) The new nomenclature of *Orobanche* and *Phelipanche*. Weed Res 49(Suppl. 1):6–7
- Jones M (1991) Studies into the pollination of *Orobanche* species in the British Isles. Prog Orobanche Res Tubingen 1:6–17
- Kovach WL (1999) MVSP—a multivariate statistical package for windows, ver. 3.1. Kovach Computing Services, Pentraeth
- Kreutz CAJ (1995) *Orobanche*. The European broomrape species. Central and northern Europe. Natuurhistorisch Genootschap, Limburg
- Manen JF, Habashi C, Jeanmonod D, Park JM, Schneeweiss GM (2004) Phylogeny and intraspecific variability of holoparasitic *Orobanche* (Orobanchaceae) inferred from plastid rbcL sequences. Mol Phylogen Evol 33(2):482–500
- Minkin PJ, Eshbaugh WH (1989) Pollen morphology of the Orobanchaceae and Rhinanthoid Scrophulariaceae. Grana 28:1–18
- Nobis M, Nowak A, Nobis A, Paszko B, Piwowarczyk R, Nowak S, Plášek V (2014) Contribution to the flora of Asian and European countries: new national and regional vascular plants records. Acta Bot Gal 160(1):81–89
- Novopokrovskij IV, Tzvelev NN (1958) Orobanchaceae. In: Shishkin BK (ed) Flora Unionis Republicarum Socialisticarum Sovieticarum (Flora URSS), 23:19–117. Institutum Botanicum nomine V. L. Komarovii Academiae Scientiarum URSS, Mosqua, Leningrad, pp 685–687
- Ollerton J, Stott A, Allnutt E, Shove S, Taylor C, Lamborn E (2007) Pollination niche overlap between a parasitic plant and its host. Oecologia 151(3):473–485
- Park J, Manen J, Schneeweiss G (2007a) Horizontal gene transfer of a plastid gene in the non-photosynthetic flowering plants

Orobanche and *Phelipanche* (Orobanchaceae). Mol Phylogen Evol 43(3):974–985

- Park J, Schneeweiss G, Weiss-Schneeweiss H (2007b) Diversity and evolution of Ty1-copia and Ty3-gypsy retroelements in the nonphotosynthetic flowering plants *Orobanche* and *Phelipanche* (Orobanchaceae). Gene 387(1–2):75–86
- Piednoël M, Aberer AJ, Schneeweiss GM, Macas J, Novak P, Gundlach H, Temsch EM, Renner SS (2012) Next-generation sequencing reveals the impact of repetitive DNA across phylogenetically closely related genomes of *Orobanchaceae*. Mol Biol Evol 29(11):3601–3611. doi:10.1093/molbev/mss168
- Piwowarczyk R (2011) Orobanche mayeri (Suess. & Ronniger) Bertsch & F. Bertsch—the new species to Poland. Acta Soc Bot Pol 80(3):179–183
- Piwowarczyk R (2012a) The genus Orobanche L. (Orobanchaceae) in the Wyżyna Małopolska upland (S Poland): distribution, habitat, host preferences and taxonomic problems. Biodivers Res Conserv 26:3–22
- Piwowarczyk R (2012b) Orobanche alba subsp. alba and subsp. major (Orobanchaceae) in Poland: current distribution, taxonomy, plant communities, hosts, and seed micromorphology. Biodivers Res Conserv 26:23–38
- Piwowarczyk R (2012c) Revised distribution and plant communities of Orobanche alsatica and notes on the Orobanchaceae series Alsaticae in Poland. Biodivers Res Conserv 26:39–51
- Piwowarczyk R (2012d) Revised distribution and phytosociological data of *Orobanche coerulescens* Stephan in Willd. (Orobanchaceae): Poland in relation to Central Europe. Biodivers Res Conserv 26:61–72
- Piwowarczyk R (2012e) A revision of distribution and the ecological description of *Orobanche picridis* (Orobanchaceae) at the NE limit of its geographical range from Poland and Ukraine. Acta Agrobot 65(1):91–106
- Piwowarczyk R (2012f) Orobanche bohemica Čelak. (Orobanchaceae) at the eastern limit of its geographical range: new data on its distribution in Poland. Biodivers Res Conserv 26:53–59
- Piwowarczyk R (2012g) Orobanche purpurea (Orobanchaceae) in Poland: current distribution, taxonomy, plant communities, and preferred hosts. Biodivers Res Conserv 26:73–81
- Piwowarczyk R (2012h) A revision of distribution and historical analysis of preferred hosts of *Orobanche ramosa* (Orobanchaceae) in Poland. Acta Agrobot 65(1):53–62
- Piwowarczyk R (2013) Seed productivity in relation to other schoot features for endangered parasitic plant *Orobanche picridis* F.W. Schultz (Orobanchaceae). Polish J Ecol 61(1):55–64
- Piwowarczyk R, Jankowska-Blaszczuk M (2014) Intra-specific diversity of seed productivity and morphological features in parasitic species *Orobanche bartlingii* Griseb. (Orobanchaceae). Polish J Ecol (in press)
- Piwowarczyk R, Przemyski A (2009) New locality of Orobanche coerulescens Stephan ex Willd. (Orobanchaceae) at the NW limit of its geographical range. Acta Soc Bot Pol 78(4):291–295
- Piwowarczyk R, Przemyski A (2010) The distribution and habitat preferences of the declining species Orobanche arenaria (Orobanchaceae) at the northern limit of its geographical range. Acta Soc Bot Pol 79(1):43–50
- Piwowarczyk R, Nobis M, Przemyski A (2009) Orobanche bartlingii Griseb. (Orobanchaceae) in Poland: taxonomical position, distribution and habitat requirements. Biodivers Res Conserv 13:3–8
- Piwowarczyk R, Chmielewski P, Gierczyk B, Piwowarski B, Stachyra P (2010) Orobanche pallidiflora Wimm. & Grab. in Poland: distribution, habitat and host preferences. Acta Soc Bot Pol 79(3):197–205
- Piwowarczyk R, Chmielewski P, Cwener A (2011) The distribution and habitat requirements of the genus *Orobanche* L. (Orobanchaceae) in SE Poland. Acta Soc Bot Pol 80(1):37–48

- Piwowarczyk R, Halamski AT, Durska E (2014) Seed and pollen morphology in the Orobanche alsatica complex (Orobanchaceae) and its taxonomic significance. Austral Syst Bot (in press)
- Plaza L, Fernández I, Juan R, Pastor J, Pujadas A (2004) Micromorphological studies on seeds of *Orobanche* species from the Iberian Peninsula and the Balearic Islands, and their systematic significance. Ann Bot (Oxford) 94:167–178
- Polo JM (1987) Orobanchaceae. In: Valdes B, Dicz MJ, Fernandez I (eds) Atlas Polinico de Andalucia Occidental. Inst. De Desarollo Regional, #43. de la Universidad de Sevilla, Sevilla
- Pujadas Salvà AJ (2007) Novedades taxonómicas y nomenclaturalers en el género Orobanche L. (Orobanchaceae). Acta Bot Malacitana 32:1–3
- Punt W, Hoen PP, Blackmore S, Nilsson S, Le Thomas A (2007) Glossary of pollen and spore terminology. Rev Palaeobot Palyn 143:1–81
- Pusch J (2006) Die Böhmische Sommerwurz (Orobanche bohemica Čelak.)—Ein Beitrag zur Abgrenzung. Verbreitung und Gefährdung dieses zentraleuropäischen Endemiten Veröffentlichungen Naturkundemuseum Erfurt 25:127–148
- Pusch J, Günther KF (2009) Orobanchaceae (Sommerwurzgewächse).In: Hegi G (ed) Illustrierte Flora von Mitteleuropa Bd. 6/1A, Lieferung 1. Weissdorn, Jena, pp 1–99
- Rao TS (1963) Pollen morphology of two species of Orobanchaceae. Curr Sci 32:557–558
- Schneeweiss GM (2001) Relationships within Orobanche sect. Trionychon: insights from its sequences markers. In: Fer A, Thalouarn P, Joel DM, Musselman LJ, Parker C, Verkleij JAC (eds) Proceeding of the 7th International Parasitic Weed Symposium. Nantes, France, pp 49–52
- Schneeweiss GM (2007) Correlated evolution of life history and host range in the nonphotosynthetic parasitic flowering plants Orobanche and Phelipanche (Orobanchaceae). J Evol Biol 20(2):471–478
- Schneeweiss GM, Colwell A, Park JM, Jang CG, Stuessy TF (2004a) Phylogeny of holoparasitic *Orobanche* (Orobanchaceae) inferred from nuclear ITS sequences. Mol Phylogen Evol 30(2):465–478

- Schneeweiss GM, Palomeque T, Colwell A, Weiss-Schneeweiss H (2004b) Chromosome numbers and karyotype evolution of holoparasitic *Orobanche* (Orobanchaceae) and related genera. Amer J Bot 91(13):439–448
- Shahi Shavvon R, Saeidi Mehrvarz S (2010) Pollen and seed morphology of the genus *Cistanche* (Orobanchaceae) in Iran. Biologia (Bratislava) 65(4):615–620
- Teryokhin ES (1997) Weed Broomrapes—systematics, ontogenesis, biology, evolution. Aufstieg, Germany
- Teryokhin ES, Shibakina GV, Serafimovich NB, Kravtsova T (1993) Determinator of broomrapes of the USSR flora. Nauka, Leningrad
- Tiagi B (1951) Studies in the family Orobanchaceae. III. A contribution to the embryology of *Orobanche cernua* Loeffl. and *O. aegyptiaca* Pers. Phytomorph 1:158–169
- Uhlich H, Pusch J, Barthel KJ (1995) Die Sommerwurzarten Europa: Gattung Orobanche. Westarp-Wiss, Magdeburg
- Velasco L, Goffman FG, Pujadas A (2000) Fatty acids and tocochromanols in seeds of *Orobanche*. Phytochemistry 54:295–300
- Weiss-Schneeweiss H, Greilhuber J, Schneeweiss GM (2006) Genome size evolution in holoparasitic *Orobanche* (Orobanchaceae) and related genera. Amer J Bot 93(1):148–156
- Willdenow CL (1800) Caroli a Linné Species Plantarum. Tomus 3. Berolini
- Zare G, Dönmez AA (2013) Two new records of the genus Orobanche (Orobanchaceae) from Turkey. Turk J Bot 37:1–7
- Zare G, Dönmez AA, Dönmez EO (2013) Pollen morphology and evolution in the genus *Orobanche L.* s.l. and its allied genera (Orobancheae/Orobanchaceae) in Turkey. Pl Syst Evol. doi:10. 1007/s00606-013-0919-2
- Zázvorka J (1997) Orobanchaceae Vent. Zárazovité. In: Goliášová K (ed) Flóra Slovenska, vol 5(2). VEDA, Bratislava, pp 460–529
- Zázvorka J (2000) Orobanchaceae—zarazovite. In: Slavik B (ed) Kvetena Česke Republiky 6, Academia Praha
- Zázvorka J (2010) Orobanche kochii and O. elatior (Orobanchaceae) in central Europe. Acta Mus Morav Sci Biol (Brno) 95(2):77–119