



UNIVERSITI PUTRA MALAYSIA

**SUBCRITICAL WATER EXTRACTION OF LIPID CONTAINING OMEGA-3  
FROM MICROALGAE *Nannochloropsis gaditana***

BERNARD HO CHON HAN

FK 2021 24



**SUBCRITICAL WATER EXTRACTION OF LIPID CONTAINING OMEGA-3  
FROM MICROALGAE *Nannochloropsis gaditana***

By

**BERNARD HO CHON HAN**

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of  
Doctor of Philosophy

December 2020

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfillment of the requirement for the degree of Doctor of Philosophy

**SUBCRITICAL WATER EXTRACTION OF LIPID CONTAINING OMEGA-3  
FROM MICROALGAE *Nannochloropsis gaditana***

By

**BERNARD HO CHON HAN**

**December 2020**

**Chair : Mohd Razif Harun, PhD**  
**Faculty : Engineering**

Omega-3 fatty acids, mainly eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) are known to have many health benefits. Many researchers have found different algal species which have high omega-3 content. Conventional techniques such as to extract microalgal omega-3 have too many disadvantages. This hinders the application of the extracted products to be fully utilized for human consumption. Therefore, a need for green, fast, and robust approach to extract the lipids from microalgae is essential. This study investigates the effectiveness of subcritical water extraction (SWE) in extracting lipid and EPA from microalgae, *Nannochloropsis gaditana*.

The preliminary screening of SWE experiments were carried out to identify the suitable range of parameters. The highest yield of lipid for the preliminary screening was at a temperature of 210°C and reaction time of 10 min yielding 20.79 wt% of lipid. After screening, an optimization is done with the parameters set on extraction temperature (156.1-273.9°C), time (6.6-23.4 minutes), and biomass loading (33-117 g algae/L) that are further optimized for lipid yield and EPA composition using central composite design (CCD). All three parameters were found to be significant factors for the changes in lipid yield, but extraction time was not a significant factor for EPA composition change. It was found that the predicted optimum lipid yield and EPA composition at 236.54 °C, 13.95 minutes and 60.50 g algae/L was 18.278 wt% of total biomass and 14.036 wt% of total fatty acid methyl ester (FAME), respectively.

Furthermore, the separation of the lipid extracts was performed using a solid phase extraction (SPE) method, where the lipids were classified into polar lipid (POL), neutral lipid (NL) and free fatty acid (FFA) component. From the findings, the POL was more susceptible to hydrolysis than NL. The highest recoveries of NL and POL from the biomass were 81.16 wt% and 66.45 wt% of lipid as compared to B&D method, respectively.

A reaction pathway for SWE of *Nannochloropsis gaditana* was also developed and used to derive the kinetic equation. The highest rate constant and lowest activation energy was the pathway of algal EPA-POL to be converted into by-products showing further that EPA-POL have high rate of hydrolysis at higher temperature with activation energy of 37.56 kJ/mol. It was also found that the model successfully incorporated to both major and minor fatty acids present in the microalgae such as palmitoleic acid, linolenic acid, and arachidonic acid.

Overall, the outcome of this study contributes to a better utilization of microalgae as an available source of omega-3 fatty acids for food and pharmaceutical industry as well as achieving the green and fast extractions with high concentration of omega-3.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGEKSTRAKAN LEMAK MENGANDUNG OMEGA-3 DARIPADA  
MIKROALGA *Nannochloropsis gaditana* DENGAN MENGGUNAKAN  
KAEDAH AIR SUB-KRITIKAL**

Oleh

**BERNARD HO CHON HAN**

**Disember 2020**

**Pengerusi : Mohd Razif Harun, PhD**  
**Fakulti : Kejuruteraan**

Omega-3 asid lemak terutamanya asid eikosapentaenoik (EPA) dan asid dokosahexaenoik (DHA) mempunyai banyak faedah kesihatan seperti anti radang, mencegah penyakit kardiovaskular, meningkatkan kesihatan mental dan lain-lain. Banyak penyelidik telah menemui pelbagai spesis alga yang mempunyai kandungan omega-3 yang tinggi. Teknik lama untuk mengekstrak omega-3 daripada mikroalga mempunyai terlalu banyak kekurangan seperti penggunaan pelarut berbahaya, proses pengekstrakan yang panjang dan hasil produk yang rendah. Ini menghindarkan hasil pengekstrakan daripada dimanfaatkan untuk penggunaan oleh manusia. Oleh itu, proses hijau, cepat dan kukuh diperlukan untuk mengekstrak lemak daripada mikrolaga. Dengan itu, tujuan penyelidikan ini adalah untuk menganalisis keberkesanan kaedah air sub-kritikal (SWE) untuk mengekstrak EPA daripada *Nannochloropsis gaditana*.

Analisis awal bagi eksperimen SWE dilaksanakan untuk mengetahui jangkaan jarak. Hasil lemak tertinggi pada analisis awal boleh diperolehi dalam suhu 210 °C dan masa 10 minit yang berjumlah 20.79 wt% hasil lemak. Selepas analisis awal, suhu (156.1-273.9 °C), masa (6.6-23.4 minit), dan komposisi biomas (33-117 g alga/L) digunakan dalam analisis pengoptimuman untuk hasil lemak dan komposisi EPA dengan menggunakan central composite design (CCD). Jangkaan optimum hasil lemak dan komposisi EPA pada 236.54 °C, 13.95 minit dan 60.50 g alga/L adalah 18.278 wt% daripada jumlah biomas dan 14.036 wt% daripada jumlah ester asid lemak (FAME). Pengimbas mikroskop

elektron menunjukkan bahawa dinding sel telah dihancurkan dan komponen bioaktif dalam mikroalga telah berjaya diekstrak oleh kaedah SWE.

Penyifatan ekstrak lemak telah dilakukan dengan menggunakan kaedah pengekstrakan fasa pepejal (SPE) dengan mengklasifikasi lemak ke dalam komponen lemak berikutub (POL), lemak neutral (NL) dan asid lemak bebas (FFA). Daripada penemuan analisis, ekstrak lemak didapati mempunyai kandungan POL yang tinggi apabila dibandingkan dengan NL. Pendapatan NL dan POL daripada biomas adalah sebanyak 81.16 wt% dan 66.45 wt% daripada jumlah lemak.

Laluan tindak balas bagi SWE daripada *Nannochloropsis gaditana* juga telah dihasilkan dan digunakan untuk membentukkan penyamaan kinetik. Pemalar tindak balas yang tertinggi and tenaga pengaktifan yang paling rendah ialah laluan tindak balas bagi EPA-POL yang ditukarkan kepada produk yang tidak diingini dengan tenaga pengaktifan sebanyak 37.56 kJ/mol. Model tersebut berjaya digunakan untuk kedua-dua acid lemak utama dan acid lemak sebilangan kecil dalam mikroalga ini seperti asid palmitoleic, asid linolenic, dan asid arachidonic.

Kesimpulannya, penyelidikan ini akan memberi sumbangan kepada penggunaan mikroalga sebagai satu sumber omega-3 untuk makanan dan industry farmasi. Pada masa yang sama, ia akan menjayakan satu kaedah teknologi hijau dan pengekstrakan yang cepat dengan hasil kepekatan omega-3 yang tinggi.

## **ACKNOWLEDGEMENTS**

Thank God for His blessings and guidance upon my research. Because of His grace and protection, I can complete my research and thesis safely. Praise the Lord for His everlasting love for me.

I would like to first thank my supervisor, Associate Professor Ts. Dr. Mohd Razif Harun for the continuous support and guidance throughout this research. He had never failed to give me good advice and correcting my mistakes toward the completion of my research work. Despite his busy schedule, he would always find time to meet and discuss about my work regularly to check on my progress. His word of encouragement has motivated me to press on and finish this research to completion. I also would like to take this opportunity to express my gratitude to my co-supervisors, Associate Professor Dr. Norhafizah Abdullah, Associate Professor Dr. Ir. Shamsul Siajam Izhar, and Professor Dr. Ir. Siti Mazlina Mustapa Kamal for their support, advice, and guidance in certain areas I am lacking.

I also wish to extend my gratitude towards the staff in the laboratory, especially Pn. Shafizah Masuri, Mr. Norisham Abdul Wahab and Mr. Adli Nazri Mohd Kasim for allowing and teaching me to use the laboratory facilities. Their outstanding experience and intelligence in handling laboratory works had helped me throughout the research period. At the same time, I would like to express my appreciations toward my fellow labmates, Nur Hidayah Zainan, Anis Nurdhiani Rosdi, Quin Emparan and Selvakumar Thiruvenkadam for all their help in the laboratory and also sharing their knowledge.

Last but not least, I would like to thank my wife and my family members for their understanding and continuous support in every way possible. This journey was pleasant with their support and encouragement.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy . The members of the Supervisory Committee were as follows:

**Mohd Razif bin Harun, PhD**

Associate Professor, Ts.  
Faculty of Engineering  
Universiti Putra Malaysia  
(Chairman)

**Norhafizah binti Abdullah, PhD**

Associate Professor  
Faculty of Engineering  
Universiti Putra Malaysia  
(Member)

**Shamsul Izhar bin Siajam, PhD**

Associate Professor, Ir.  
Faculty of Engineering  
Universiti Putra Malaysia  
(Member)

**Siti Mazlina binti Mustapa Kamal, PhD**

Professor, Ir.  
Faculty of Engineering  
Universiti Putra Malaysia  
(Member)

---

**ZALILAH MOHD SHARIFF, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 06 May 2021

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Bernard Ho Chon Han (GS44619)

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman  
of Supervisory  
Committee:

\_\_\_\_\_

Associate Professor Ts. Dr. Mohd Razif Harun

Signature:

Name of Member of  
Supervisory  
Committee:

\_\_\_\_\_

Associate Professor Dr. Norhafizah Abdullah

Signature:

Name of Member of  
Supervisory  
Committee:

\_\_\_\_\_

Associate Professor Ir. Dr. Shamsul Izhar Siajam

Signature:

Name of Member of  
Supervisory  
Committee:

\_\_\_\_\_

Professor Ir. Dr. Siti Mazlina Mustapa Kamal

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xii
<b>LIST OF FIGURES</b>	xiii
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
<b>1      INTRODUCTION</b>	<b>1</b>
1.1     Background	1
1.2     Problem Statement	2
1.3     Research Objectives	3
1.4     Research Scope and Limitations	3
1.5     Significance of Research	4
<b>2      LITERATURE REVIEW</b>	<b>5</b>
2.1     Fatty Acid and Lipids	5
2.1.1    Benefits of Docosahexaenoic Acid (DHA) and Eicosapentaenoic Acid (EPA)	7
2.2     Sources of Omega-3	8
2.2.1    Fishes	8
2.2.2    Krills	12
2.2.3    Plants	13
2.2.4    Microalgae	13
2.3     Current Market of Microalgal Omega-3 Products	20
2.4     Conventional Lipid Extraction Techniques	21
2.4.1    Bligh and Dyer Extraction Method (B&D)	21
2.4.2    Direct Transesterification	22
2.4.3    Soxhlet	23
2.5     Green Lipid Extraction Method	24
2.5.1    Supercritical Fluid Extraction (SFE)	24
2.5.2    Bio-derived Solvent Extraction	26
2.5.3    Ionic Liquid (IL) Extraction	27
2.5.4    Subcritical Water Extraction (SWE)	28
2.6     Potential of <i>Nannochloropsis gaditana</i> in Omega-3 Extraction Using SWE	30
2.7     Optimization Study Using Central Composite Design	32
2.8     Kinetic Modelling of SWE	33
2.9     Summary	35

<b>3</b>	<b>MATERIALS AND METHODS</b>	36
3.1	Introduction	36
3.2	Raw Material	36
3.3	Characterization	36
3.3.1	Ultimate Analysis	36
3.3.2	Proximate Analysis	38
3.3.3	Scanning Electron Microscopy (SEM)	38
3.4	Extraction Method	38
3.4.1	Bligh and Dyer Extraction (B&D)	38
3.4.2	Subcritical Water Extraction (SWE)	39
3.5	Solid-Phase Extraction (SPE)	40
3.5.1	Thin Layer Chromatography (TLC)	40
3.6	Gas Chromatography (GC)	41
3.7	Experimental Design	42
3.8	Kinetic Modeling	43
3.8.1	Ordinary Differential Equation (ODE)	43
3.9	Statistical Analysis	45
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	46
4.1	Raw Biomass Characterization	46
4.2	Screening on the Effects of Parameters of SWE on Lipid Yield	47
4.2.1	Effect of Temperature	47
4.2.2	Effect of Extraction Time	48
4.3	CCD of Extraction Parameters	49
4.3.1	Analysis of Variance (ANOVA)	51
4.3.2	Response Surface Methodology (RSM)	55
4.3.3	Morphological Changes of Biomass	60
4.3.4	Optimization and Validation	61
4.4	Lipid Class Profiling	64
4.4.1	Effect of Temperature on Lipid Classes	64
4.4.2	Effect of Extraction Time on Lipid Classes	67
4.4.3	Fatty Acid Profiling	68
4.5	Kinetics of Eicosapentaenoic Acid (EPA) Extraction	70
4.5.1	Investigation for Reaction Pathway	71
4.5.2	Developing Reaction Network and Kinetic Model	75
4.5.3	Fitting of Data	76
4.5.4	Arrhenius Parameters	79
4.5.5	Model Prediction	79
<b>5</b>	<b>CONCLUSION</b>	82
5.1	Summary	82
5.2	Future Research and Recommendations	83
<b>REFERENCES</b>		85
<b>APPENDICES</b>		112
<b>BIODATA OF STUDENT</b>		127
<b>LIST OF PUBLICATION</b>		128

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	Lipid composition of different biomass with different extraction method	9
2.1	Lipid composition of different species of microalgae high in omega-3	16
2.3	Recent patents for the lipid extraction technologies for microalgal biomass	19
2.4	Comparison of different lipid extraction methods	25
3.1	Coded values and process conditions for CCD of SWE of <i>Nannochloropsis gaditana</i>	42
3.2	Experimental design matrix and responses of SWE of <i>Nannochloropsis gaditana</i>	43
4.1	Elemental composition, proximate analysis and energy value of <i>Nannochloropsis gaditana</i>	46
4.2	Experimental design and responses of SWE from <i>Nannochloropsis gaditana</i>	50
4.3	ANOVA of all factors and summary statistics of lipid yield	53
4.4	ANOVA of all factors and summary statistics of EPA composition	54
4.5	Optimization table for the specific condition and its desirability	63
4.6	Major fatty acid attached to NLs (mg/g lipid) extracted at 15 min	68
4.7	Major fatty acid attached to POLs (mg/g lipid) extracted at 15 min	69
4.8	Rate constants optimized through genetic algorithm at different temperature	77
4.9	Arrhenius parameter of the model for EPA extraction	79

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	Structure of neutral lipid in the form of triglycerides (Oi et al., 2019)	6
2.2	Structures of polar lipids in the form of phospholipid and glycolipid. R' = variable polar head group; N' = variable sugar chain (Fang et al., 2019)	6
2.3	Graphical representation of water dielectric constant (Herrero et al., 2006)	29
2.4	Central composite design in the form of (a) circumscribed, (b) inscribed and (c) face-centered	33
2.5	SWE network incorporating biochemical content (Valdez et al., 2014)	34
3.1	Summary of methods	37
3.2	Schematic diagram of subcritical water extraction set up	39
3.3	Flowchart for MATLAB coding	44
4.1	Preliminary study of SWE of <i>Nannochloropsis gaditana</i> on lipid yield (n=2)	48
4.2	Box-Cox Plot for the response of (a) $Y_1$ and (b) $Y_2$ before transformation with 95% confidence	51
4.3	Plot of predicted values calculated from the model and the experimental yield of (a) $Y_1$ and (b) $Y_2$	55
4.4	The interaction of all the factors of SWE on lipid yield using (a) response surface plot and (b) contour plot with set factors of biomass loading = 75 g algae/L	56
4.5	The interaction of all the factors of SWE on lipid yield using (a) response surface plot and (b) contour plot with set factors of time = 15 min	57
4.6	The interaction of all the factors of SWE on lipid yield using (a) response surface plot and (b) contour plot with set factors of temperature = 215 °C	58

4.7	The interaction of significant factors of SWE on EPA composition using (a) response surface plot and (b) contour plot	59
4.8	SEM images of (a) untreated microalgae and extracted microalgae at 15 minutes and 75 g algae/L with different temperature of (b) 156.1 °C, (c) 215.0 °C and (d) 273.9 °C	62
4.9	NL yield from SWE of <i>Nannochloropsis gaditana</i> based on different temperature and extraction time	65
4.10	FFA yield from SWE of <i>Nannochloropsis gaditana</i> based on different temperature and extraction time	65
4.11	POL yield from SWE of <i>Nannochloropsis gaditana</i> based on different temperature and extraction time	66
4.12	Total lipid yield (Sum of NL, FFA and POL) from SWE of <i>Nannochloropsis gaditana</i> based on different temperature and extraction time	66
4.13	Yield of EPA-NL recovered	71
4.14	Yield of EPA-NL extracted	72
4.15	Yield of EPA-POL recovered	72
4.16	Yield of EPA-POL extracted	73
4.17	Yield of EPA by-product	73
4.18	Reaction network for SWE of EPA-NL and EPA-POL from <i>Nannochloropsis gaditana</i>	75
4.19	Yields of EPA-NL recovered (♦), EPA-POL recovered (■), EPA-NL extracted (▲), EPA-POL extracted (▼), and by-products (●), and corresponding correlation with the model from the reaction at a) 160°C, b) 170 °C, c) 180 °C, d) 190 °C, and e) 200 °C	78
4.20	Parity plot of experimental and model yield of EPA-NL recovered (♦), EPA-POL recovered (▲), EPA-NL extracted (●), EPA-POL extracted (+), and by-products (x)	80

4.21	Specific fatty acid yields of FA-NL recovered (♦), FA-POL recovered (■), FA-NL extracted (▲), FA-POL extracted (▼), and by-products (●) for a) palmitoleic acid, b) linolenic acid, and c) arachidonic acid at reaction temperature of 190 °C	81
A1	Product of samples after SWE at a temperature of (a) 156.1 °C, (b) 215 °C and (c) 273.9 °C	112
B1	Gas chromatography of 37 FAME mix standard and the retention time for each fatty acid component	113
B2	Gas chromatography of samples extracted using B&D method for (a) neutral lipid (NL) and (b) polar lipid (POL)	115
B3	Gas chromatography of extracted neutral lipid (NL) at 15 min for (a) 160 °C, (b) 170 °C, (c) 180 °C, (d), 190 °C and (e) 200 °C	116
B4	Gas chromatography of extracted polar lipid (POL) at 15 min for (a) 160 °C, (b) 170 °C, (c) 180 °C, (d), 190 °C and (e) 200 °C	118
C1	MATLAB coding for ODE solving	121
D1	Arrhenius parameters plot of ln k against 1/T for EPA extraction of (a) $k_1$ , (b) $k_2$ , (c) $k_3$ , (d) $k_4$ , (e) $k_5$ , (f) $k_6$ and (g) $k_7$	123

## LIST OF ABBREVIATIONS

ALA	$\alpha$ -linolenic acid
ANOVA	Analysis of variance
B&D	Bligh and Dyer extraction
CCD	Central composite design
CO <sub>2</sub>	Carbon dioxide
CVD	Cardiovascular diseases
DGDG	Digalactosyl-diacylglycerol
DHA	Docosahexaenoic acid
E <sub>a</sub>	Activation energy
EPA	Eicosapentaenoic acid
FAME	Fatty acid methyl ester
FFA	Free fatty acid
FID	Flame ionization detector
GC	Gas chromatography
GL	Glycolipid
GLA	$\gamma$ -linolenic acid
HDL-C	High density lipoprotein cholesterol
IL	Ionic liquid
<i>k<sub>i</sub></i>	Rate constant
K <sub>w</sub>	Ionic product of water
LDL-C	Low density lipoprotein cholesterol
MGDG	Monogalactosyl-diacylglycerol
M-SDE	Modified simultaneous distillation extraction

MUFA	Monounsaturated fatty acid
n-3	Omega-3
n-6	Omega-6
NL	Neutral lipid
PLRP2	Pancreatic lipase-related protein 2
POL	Polar lipid
PUFA	Polyunsaturated fatty acid
SCCO <sub>2</sub>	Supercritical carbon dioxide extraction
SDA	Stearidonic acid
SEM	Scanning electron microscopy
SFA	Saturated fatty acid
SFE	Supercritical fluid extraction
SPE	Solid phase extraction
SWE	Subcritical water extraction
TFA	Total fatty acid
TG	Triglyceride
TLC	Thin layer chromatography
$\sigma$	Standard deviation

# CHAPTER 1

## INTRODUCTION

### 1.1 Background

Cardiovascular diseases (CVDs) are at top of the chart for causing death globally. A study conducted among United States adults from 2007 to 2014 estimated that 56.9 million people in United States (25.9%) have high levels of triglyceride (TG) in a range of 150 mg/dL to 400 mg/dL (Fan et al., 2020). TG-rich remnant lipoproteins are considered to have atherogenic effect that will increase the risk of CVDs (Jacobson et al., 2007). Meanwhile, hypercholesterolemia is one of the most common inherited disorders to Malaysians as the prevalence increased from 35.1% to 38.1% in year 2011 until 2019 (Institute for Public Health Malaysia, 2019; Mat Rifin et al., 2018).

It has been reported that an increase in non-high density lipoprotein cholesterol (non-HDL-C) level may cause an elevated level of low density lipoprotein cholesterol (LDL-C) and TG-rich lipoprotein which is the main cause of hypercholesterolemia and CVD (Liu et al., 2006). The intake of 2 g/d to 4 g/d of omega-3 free fatty acid (FFA) could significantly reduce the non-HDL-C and TG level of patients suffering from hypertriglyceridemia (Kastelein et al., 2014; Maki et al., 2013). According to the study by Russell & Bürgin-Maunder (2012), most national and international health and government organisations recommend a significant dosage intake of omega-3 fatty acid daily ranging from 250 mg/d for a healthy adult and up to 4000 mg/d for an adult suffering from different diseases. For example, the American Heart Association suggested 1000 mg/day of omega-3 for chronic heart disease sufferers (Turchini et al., 2012). Besides reducing the risk of CVD which is the most significant benefits from omega-3, there are also other health benefits that omega-3 might offer such as curbing mental health problem (Dyall, 2015; Gow & Hibbeln, 2014; Sublette et al., 2011) and has anti-inflammatory effects (Buckley et al., 2014). With that, omega-3 supplements might be able to help 2.3% of adults suffering from depression and 7.9% of children below 15 years of age having some sort mental health problem in Malaysia as reported in 2019 (Institute for Public Health Malaysia, 2019).

The most commercial sources of omega-3 are found in aquatic organisms and plants. Likewise, microalgae are also considered as the potential producer of long-chain omega-3 fatty acid mainly EPA and DHA. Many researchers found different algal species that have high omega-3 content (Griffiths et al., 2012). Usage of microalgae in the production of omega-3 supplements commercially may have many benefits such as reducing of the possibilities of getting methylmercury poisoning from fishes, creating another alternative source of

long-chain omega-3 for vegan and non-fish eater, and becoming new source of antioxidants (Breivik, 2007; Freitas et al., 2012; Kris-Etherton et al., 2002).

Subcritical water extraction (SWE) has been used to extract various biomolecules, including omega-3. SWE is known to be a green extraction method as the process of extraction is performed without the usage of any harmful solvents. This method is able to extract high amount of nutraceuticals and lipids with just the changes in water properties at high temperature and high pressure conditions (Reddy et al., 2016).

Stakeholders such as Martek and Cellana who had lots of experience in omega-3 have already ventured into omega-3 production from microalgae hence proving its marketability and promising future (Adarme-Vega et al., 2012). In the meantime, Malaysia also currently has its own largest algae cultivation facility in Sarawak as the project is jointly organized by the Sarawak Biodiversity Centre and Mitsubishi Corporation (BERNAMA, 2019). Most of the research universities in Malaysia have developed interest in tapping into microalgae researches but their focus are more into the production of biodiesel instead of omega-3 extraction (Rajkumar & Sobri Takriff, 2016). However, some researchers believed in the potential of microalgae in global omega-3 market contributing nearly US\$350 million of sales in 2012 (Matos, 2017).

## 1.2 Problem Statement

Most conventional extraction techniques such as mechanical extraction, Soxhlet, and Bligh & Dyer (B&D) methods involve long processing time and usually use hazardous chemicals as solvents. This hinders the application of the extracted products to be fully utilized for human consumption. Meanwhile, there is also an increase in demands for sustainable omega-3 sources in the market but farmed fishes had dropped in qualities in terms of omega-3 content over the years from about 3 g of omega-3 per 100 g fish oil to less than 2 g of omega-3 per 100 g of oil (Tocher et al., 2019). Microalgae, *Nannochloropsis gaditana* has great omega-3 production potentials that are not only suitable for human consumption but also beneficial to the environment.

Lipid extracts constructed by complex neutral lipid (NL) and polar lipid (POL) mixtures commonly pose extraction and purification challenge. Therefore, a need for green, fast and robust approach to extract the lipids from *Nannochloropsis gaditana* is essential. Green method such as supercritical carbon dioxide ( $\text{CO}_2$ ) extraction still requires a need for co-solvent (hexane and ethanol) which is harmful for the environment for the extraction of both NL and POL (Patil et al., 2018). With the benefits offered by the SWE coupled with the correct choice of purification approach, the quality and the commercial viabilities of the omega-3 fatty acids produced can be ensured even without the usage of other harmful chemicals. The control in extraction temperature and

time will be a key factor in optimizing the extraction of EPA existing in both NL and POL (Qu et al., 2018).

Also, scaling up the green extraction method of algal oil via subcritical water extraction (SWE) is still a major challenge. Currently, there are no extensive study on the optimization and kinetics on the extraction of omega-3 fatty acids from *Nannochloropsis* species from SWE method even though this compound exists abundantly in this microalgae species. Basic understanding of the extraction and conversion of microalgae biomass to lipid, aqueous product, and gas is not adequate to fully utilize the lipid into nutraceuticals (Hietala et al., 2016). Optimization of reaction parameters and understanding the kinetic reaction of the process in SWE in terms of EPA products will effectively help in scaling up the process (Mathimani et al., 2019).

Meanwhile, the growth of this industry in producing omega-3 from microalgae should not be overlooked as it is currently one of the growing industries in the world (Milledge, 2011; Tocher et al., 2019). Thus, the aims of this study are to investigate the potential of SWE technology in extracting lipid from *Nannochloropsis gaditana* and to optimize its parameters for high quality lipid with high EPA content yield. The outcome of this study will contribute to a better utilization of microalgae as an available source of omega-3 fatty acids for food and pharmaceutical industry as well as achieving a green and fast extraction with high concentration of omega-3.

### **1.3 Research Objectives**

The main focus of the study is divided into the following specific objectives:

- a) To investigate and optimize the factors that affect the extraction yield of lipid and eicosapentaenoic acid (EPA) composition from *Nannochloropsis gaditana* via subcritical water extraction (SWE) method.
- b) To analyze the extracted lipid from SWE according to lipid composition and fatty acid profile of *Nannochloropsis gaditana*.
- c) To develop the reaction network and kinetic model of SWE for EPA extracted from *Nannochloropsis gaditana*.

### **1.4 Research Scopes and Limitations**

This study focuses on the extraction of lipid and EPA from *Nannochloropsis gaditana* using SWE method. The variables studied were the extraction temperature, time and biomass loading. The extraction volume of total biomass and water was limited to 32 mL due to the maximum size of the batch reactor. The extraction yield and quality of the lipids were analyzed using gas chromatography (GC) and optimized through central composite design (CCD).

Thin layer chromatography (TLC) and solid-phase extraction (SPE) method were incorporated into the study to separate the lipid into NL, POL and FFA. This separation of lipid classes clearly showed the content and quality of lipid extracted from *Nannochloropsis gaditana*. Further separations of NL and POL into sub-classes (acylglyceride, phospholipid, glycolipid (GL), etc.) were not done due to the small amount of lipid that can be extracted from the batch system of SWE per extraction cycle. Moreover, additional separation will cause more error in calculation and analysis. Hence, only the three major classes of lipid (NL, POL and FFA) were studied.

A reaction network of SWE of EPA from *Nannochloropsis gaditana* was also developed based on the literature. The reaction pathway will focus to the natural form EPA attached to NLs and POLs. With the reaction network developed, kinetic reaction equations can be formed. The Arrhenius parameters were solved and a mathematical model was established. The parity plot between experimental yield and model yield were also performed to validate the model.

### 1.5 Significance of Research

Various companies and research setups for algal cultivation and biofuels are coming up and established by major investors in Malaysia and abroad (BERNAMA, 2019). Algae have recently received a lot of attentions as a biomass source for the production of biofuels and bioproducts (Marrone et al., 2018). Some of the main characteristics which set microalgae apart from other biomass sources are that microalgae have high biomass yield per unit of light and area and high oil yield. Meanwhile, microalgae may not necessarily require agricultural land and fresh water for cultivation as the nutrients and CO<sub>2</sub> can be supplied by wastewater and flue gas, respectively (Hemaiswarya et al., 2011; Rosenberg et al., 2008). However, the commercialization of nutraceuticals from microalgae still has major hurdles to overcome. Therefore, this research is aiming to make the extraction of omega-3 from microalgae biomass more commercially viable and environmentally friendly by focusing on extracting microalgae compounds using SWE technique. To date, none of research has been reported in utilizing this technology for microalgae omega-3 extraction. With numerous advantages of this system, it is expected to be a good extraction method for producing omega-3 nutraceuticals from microalgal biomass.

## REFERENCES

- Adarme-Vega, T. C., Lim, D. K. Y., Timmins, M., Vernen, F., Li, Y., & Schenck, P. M. (2012). Microalgal biofactories: a promising approach towards sustainable omega-3 fatty acid production. *Microbial Cell Factories*, 11, 96. <https://doi.org/10.1186/1475-2859-11-96>
- Ait-Amir, B., Pougnet, P., & El Hami, A. (2015). Meta-model development. In *Embedded Mechatronic Systems* (Vol. 2, pp. 151–179). <https://doi.org/10.1016/b978-1-78548-014-0.50006-2>
- Ajibade, B. F., Nwosu, C. R., & Mbegbu, J. I. (2015). The distribution of the inverse square root transformed error component of the multiplicative time series model. *Journal of Modern Applied Statistical Methods*, 14(2), 172–200.
- Ajjawi, I., Verruto, J., Aqui, M., Soriaga, L. B., Coppersmith, J., Kwok, K., Peach, L., Orchard, E., Kalb, R., Xu, W., Carlson, T. J., Francis, K., Konigsfeld, K., Bartalis, J., Schultz, A., Lambert, W., Schwartz, A. S., Brown, R., & Moellering, E. R. (2017). Lipid production in *Nannochloropsis gaditana* is doubled by decreasing expression of a single transcriptional regulator. *Nature Biotechnology*, 35(7), 647–652. <https://doi.org/10.1038/nbt.3865>
- Akhtar, J., & Amin, N. A. S. (2011). A review on process conditions for optimum bio-oil yield in hydrothermal liquefaction of biomass. *Renewable and Sustainable Energy Reviews*, 15(3), 1615–1624. <https://doi.org/10.1016/j.rser.2010.11.054>
- Akiya, N., & Savage, P. E. (2002). Roles of water for chemical reactions in high-temperature water. *Chemical Reviews*. <https://doi.org/10.1021/cr000668w>
- Alenezi, R., Leeke, G. A., Santos, R. C. D., & Khan, A. R. (2009). Hydrolysis kinetics of sunflower oil under subcritical water conditions. *Chemical Engineering Research and Design*, 87(6), 867–873. <https://doi.org/10.1016/j.cherd.2008.12.009>
- Ali-Nehari, A., Kim, S. B., Lee, Y. B., Lee, H. Youn, & Chun, B. S. (2012). Characterization of oil including astaxanthin extracted from krill (*Euphausia superba*) using supercritical carbon dioxide and organic solvent as comparative method. *Korean Journal of Chemical Engineering*, 29(3), 329–336. <https://doi.org/10.1007/s11814-011-0186-2>
- Allnut, F. C. T. (2019). Promising future products from microalgae for commercial applications. In K. Gayen, T. K. Bhowmick, & S. K. Maity (Eds.), *Sustainable Downstream Processing of Microalgae for Industrial Application* (1st ed., pp. 40–59). CRC Press. <https://doi.org/https://doi.org/10.1201/9780429027970>
- Amara, S., Lafont, D., Fiorentino, B., Boullanger, P., Carrière, F., & De Caro, A.

(2009). Continuous measurement of galactolipid hydrolysis by pancreatic lipolytic enzymes using the pH-stat technique and a medium chain monogalactosyl diglyceride as substrate. *Biochimica et Biophysica Acta - Molecular and Cell Biology of Lipids*, 1791(10), 983–990. <https://doi.org/10.1016/j.bbali.2009.05.002>

Arterburn, L. M., Oken, H. A., Bailey Hall, E., Hamersley, J., Kuratko, C. N., & Hoffman, J. P. (2008). Algal-oil capsules and cooked salmon: Nutritionally equivalent sources of docosahexaenoic acid. *Journal of the American Dietetic Association*, 108(7), 1204–1209. <https://doi.org/10.1016/j.jada.2008.04.020>

Arterburn, L. M., Oken, H. A., Hoffman, J. P., Bailey-Hall, E., Chung, G., Rom, D., Hamersley, J., & McCarthy, D. (2007). Bioequivalence of docosahexaenoic acid from different algal oils in capsules and in a DHA-fortified food. *Lipids*, 42(11), 1011–1024. <https://doi.org/10.1007/s11745-007-3098-5>

Balasubramanian, R. K., Yen Doan, T. T., & Obbard, J. P. (2013). Factors affecting cellular lipid extraction from marine microalgae. *Chemical Engineering Journal*, 215–216, 929–936. <https://doi.org/10.1016/j.cej.2012.11.063>

Beacham, T. A., Bradley, C., White, D. A., Bond, P., & Ali, S. T. (2014). Lipid productivity and cell wall ultrastructure of six strains of *Nannochloropsis*: Implications for biofuel production and downstream processing. *Algal Research*, 6(PA), 64–69. <https://doi.org/10.1016/j.algal.2014.09.003>

Bechtel, P. J., & Oliveira, A. C. M. (2006). Chemical characterization of liver lipid and protein from cold-water fish species. *Journal of Food Science*, 71(6), 480–485. <https://doi.org/10.1111/j.1750-3841.2006.00076.x>

Benito, P., Caballero, J., Moreno, J., Gutiérrez-Alcántara, C., Muñoz, C., Rojo, G., García, S., & Soriguer, F. C. (2006). Effects of milk enriched with omega-3 fatty acid, oleic acid and folic acid in patients with metabolic syndrome. *Clinical Nutrition (Edinburgh, Scotland)*, 25(4), 581–587. <https://doi.org/10.1016/j.clnu.2005.12.006>

Berger, A. (2014). Polar Lipids — Phospholipids and Glycolipids — An Enhanced Omega-3 Structure Polar Lipids — Phospholipids and Glycolipids. In *Almega PL™ Qualitas Health* (Issue October, p. 11).

BERNAMA. (2019). *SBC-Mitsubishi algae cultivation facility to open next month*. <http://www.bernama.com/en/news.php?id=1743132>

Betenbaugh, M. J., Donohue, M. D., Oyler, G. A., & Rosenberg, J. N. (2012). Method for extraction and purification of oils from microalgal biomass using high-pressure CO<sub>2</sub> as a solute (Patent No. WO2012024340A3). In *United States Patent Application Publication* (WO2012024340A3).

Bhattacharjee, S., Sultana, A., Sazzad, M. H., Islam, M. A., Ahtashom, M. M., & Asaduzzaman. (2013). Analysis of the proximate composition and energy

- values of two varieties of onion (*Allium cepa* L.) bulbs of different origin: A comparative study. *International Journal of Nutrition and Food Sciences*, 2(5), 246–253. <https://doi.org/10.11648/j.ijnfs.20130205.16>
- Biller, P., Ross, A. B., Skill, S. C., Lea-Langton, A., Balasundaram, B., Hall, C., Riley, R., & Llewellyn, C. A. (2012). Nutrient recycling of aqueous phase for microalgae cultivation from the hydrothermal liquefaction process. *Algal Research*, 1(1), 70–76. <https://doi.org/10.1016/j.algal.2012.02.002>
- Biller, Patrick, Sharma, B. K., Kunwar, B., & Ross, A. B. (2015). Hydroprocessing of bio-crude from continuous hydrothermal liquefaction of microalgae. *Fuel*, 159, 197–205. <https://doi.org/10.1016/j.fuel.2015.06.077>
- Bligh, E. G., & Dyer, W. J. (1959a). A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*, 37(8), 911–917. <https://doi.org/10.1139/o59-099>
- Bligh, E. G., & Dyer, W. J. (1959b). A RAPID METHOD OF TOTAL LIPID EXTRACTION AND PURIFICATION. *Canadian Journal of Biochemistry and Physiology*, 37, 911–917.
- Bloch, M. H., & Qawasmi, A. (2011). Omega-3 fatty acid supplementation for the treatment of children with attention-deficit/hyperactivity disorder symptomatology: systematic review and meta-analysis. *Journal of the American Academy of Child and Adolescent Psychiatry*, 50(10), 991–1000. <https://doi.org/10.1016/j.jaac.2011.06.008>; 10.1016/j.jaac.2011.06.008
- Borowitzka, M. A. (2013). High-value products from microalgae-their development and commercialisation. *Journal of Applied Phycology*, 25(3), 743–756. <https://doi.org/10.1007/s10811-013-9983-9>
- Bradstreet, R. B. (1954). Kjeldahl Method for Organic Nitrogen. *Analytical Chemistry*. <https://doi.org/10.1021/ac60085a028>
- Breivik, H. (2007). Long-chain omega-3 specialty oils. In Harald Breivik (Ed.), *Long-Chain Omega-3 Specialty Oils*. Oily Press. <https://doi.org/10.1533/9780857097897.43>
- Brown, T. M., Duan, P., & Savage, P. E. (2010). Hydrothermal liquefaction and gasification of *Nannochloropsis* sp. *Energy and Fuels*, 24(6), 3639–3646. <https://doi.org/10.1021/ef100203u>
- Brunner, G. (2009). Near critical and supercritical water. Part I. Hydrolytic and hydrothermal processes. *The Journal of Supercritical Fluids*, 47(3), 373–381. <https://doi.org/10.1016/j.supflu.2008.09.002>
- Buckley, C. D., Gilroy, D. W., & Serhan, C. N. (2014). Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. *Immunity*, 40(3), 315–327. <https://doi.org/10.1016/j.immuni.2014.02.009>

- Burja, A. M., Armenta, R. E., Radianingtyas, H., & Barrow, C. J. (2007). Evaluation of fatty acid extraction methods for *Thraustochytrium* sp. ONC-T18. *Journal of Agricultural and Food Chemistry*, 55(12), 4795–4801. <https://doi.org/10.1021/jf070412s>
- Burns, J. L., Nakamura, M. T., & Ma, D. W. L. (2018). Differentiating the biological effects of linoleic acid from arachidonic acid in health and disease. *Prostaglandins Leukotrienes and Essential Fatty Acids*, 135, 1–4. <https://doi.org/10.1016/j.plefa.2018.05.004>
- Burri, L., Hoem, N., Banni, S., & Berge, K. (2012). Marine Omega-3 Phospholipids: Metabolism and Biological Activities. *International Journal of Molecular Sciences*, 13(12), 15401–15419. <https://doi.org/10.3390/ijms131115401>
- Camacho-Rodríguez, J., Cerón-García, M. C., Fernández-Sevilla, J. M., & Molina-Grima, E. (2015). Genetic algorithm for the medium optimization of the microalga *Nannochloropsis gaditana* cultured to aquaculture. *Bioresource Technology*, 177, 102–109. <https://doi.org/10.1016/j.biortech.2014.11.057>
- Casula, M., Soranna, D., Catapano, A. L., & Corrao, G. (2013). Long-term effect of high dose omega-3 fatty acid supplementation for secondary prevention of cardiovascular outcomes: A meta-analysis of randomized, double blind, placebo controlled trials. *Atherosclerosis Supplements*, 14(2), 243–251. [https://doi.org/10.1016/S1567-5688\(13\)70005-9](https://doi.org/10.1016/S1567-5688(13)70005-9)
- Cavonius, L. R., Carlsson, N. G., & Undeland, I. (2014). Quantification of total fatty acids in microalgae: Comparison of extraction and transesterification methods. *Analytical and Bioanalytical Chemistry*, 406(28), 7313–7322. <https://doi.org/10.1007/s00216-014-8155-3>
- Changi, S. M. (2012). Hydrothermal Reactions of Algae Model Compounds. In *Resources*. The University of Michigan.
- Changi, S. M., Faeth, J. L., Mo, N., & Savage, P. E. (2015). Hydrothermal Reactions of Biomolecules Relevant for Microalgae Liquefaction. *Industrial and Engineering Chemistry Research*, 54(47), 11733–11758. <https://doi.org/10.1021/acs.iecr.5b02771>
- Changi, S., Matzger, A. J., & Savage, P. E. (2012). Kinetics and pathways for an algal phospholipid (1,2-dioleoyl-sn-glycero-3-phosphocholine) in high-temperature (175–350 °C) water. *Green Chemistry*, 14(10), 2856. <https://doi.org/10.1039/c2gc35639b>
- Chaturvedi, R., & Fujita, Y. (2006). Isolation of enhanced eicosapentaenoic acid producing mutants of *Nannochloropsis oculata* ST-6 using ethyl methane sulfonate induced mutagenesis techniques and their characterization at mRNA transcript level. *Phycological Research*, 54(3), 208–219. <https://doi.org/10.1111/j.1440-1835.2006.00428.x>
- Chee, C. P., Gallaher, J. J., Djordjevic, D., Faraji, H., McClements, D. J.,

- Decker, E. a, Hollender, R., Peterson, D. G., Roberts, R. F., & Coupland, J. N. (2005). Chemical and sensory analysis of strawberry flavoured yogurt supplemented with an algae oil emulsion. *The Journal of Dairy Research*, 72(3), 311–316. <https://doi.org/10.1017/S0022029905001068>
- Chen, G. Q., Jiang, Y., & Chen, F. (2007). Fatty acid and lipid class composition of the eicosapentaenoic acid-producing microalga, *Nitzschia laevis*. *Food Chemistry*, 104(4), 1580–1585. <https://doi.org/10.1016/j.foodchem.2007.03.008>
- Chen, L., Liu, T., Zhang, W., Chen, X., & Wang, J. (2012). Biodiesel production from algae oil high in free fatty acids by two-step catalytic conversion. *Bioresource Technology*, 111, 208–214. <https://doi.org/10.1016/j.biortech.2012.02.033>
- Cheng, S.-S., Chua, M.-T., Chang, E.-H., Huang, C.-G., Chen, W.-J., & Chang, S.-T. (2009). Variations in insecticidal activity and chemical compositions of leaf essential oils from *Cryptomeria japonica* at different ages. *Bioresource Technology*, 100(1), 465–470. <https://doi.org/10.1016/j.biortech.2007.11.060>
- Chu, W.-L. (2012). Biotechnological applications of microalgae. *International E-Journal of Science, Medicine & Education*, 6(126), 24–37.
- Clayton, E. H., Gulliver, C. E., Piltz, J. W., Taylor, R. D., Blake, R. J., & Meyer, R. G. (2012). Improved extraction of saturated fatty acids but not omega-3 fatty acids from sheep red blood cells using a one-step extraction procedure. *Lipids*, 47(7), 719–727. <https://doi.org/10.1007/s11745-012-3674-1>
- Copat, C., Bella, F., Castaing, M., Fallico, R., Sciacca, S., & Ferrante, M. (2012). Heavy metals concentrations in fish from Sicily (Mediterranean Sea) and evaluation of possible health risks to consumers. *Bulletin of Environmental Contamination and Toxicology*, 88(1), 78–83. <https://doi.org/10.1007/s00128-011-0433-6>
- Da Porto, C., Decorti, D., & Tubaro, F. (2012). Fatty acid composition and oxidation stability of hemp (*Cannabis sativa L.*) seed oil extracted by supercritical carbon dioxide. *Industrial Crops and Products*, 36(1), 401–404. <https://doi.org/10.1016/j.indcrop.2011.09.015>
- Das, P., Aziz, S. S., & Obbard, J. P. (2011). Two phase microalgae growth in the open system for enhanced lipid productivity. *Renewable Energy*, 36(9), 2524–2528. <https://doi.org/10.1016/j.renene.2011.02.002>
- Davidson, M. H., Johnson, J., Rooney, M. W., Kyle, M. L., & Kling, D. F. (2012). A Novel Omega-3 Free Fatty Acid Formulation has Dramatically Improved Bioavailability During a Low-Fat Diet Compared with Omega-3-Acid Ethyl Esters: The ECLIPSE (Epanova® compared to Lovaza® in a pharmacokinetic single-dose evaluation) study. *Journal of Clinical Lipidology*, 6(6), 573–584. <https://doi.org/10.1016/j.jacl.2012.01.002>

- Dawczynski, C., Martin, L., Wagner, A., & Jahreis, G. (2010). N-3 LC-PUFA-enriched dairy products are able to reduce cardiovascular risk factors: A double-blind, cross-over study. *Clinical Nutrition*, 29(5), 592–599. <https://doi.org/10.1016/j.clnu.2010.02.008>
- de Lorgeril, M., & Salen, P. (2012). New insights into the health effects of dietary saturated and omega-6 and omega-3 polyunsaturated fatty acids. *BMC Medicine*, 10(1), 50. <https://doi.org/10.1186/1741-7015-10-50>
- Deckelbaum, R. J., & Torrejon, C. (2012). The omega-3 fatty acid nutritional landscape: health benefits and sources. *The Journal of Nutrition*, 142(3), 587S-591S. <https://doi.org/10.3945/jn.111.148080>
- Díez, S. (2008). Human Health Effects of Methylmercury. *Business*, 198, 18–26. <https://doi.org/10.1007/978-0-387-09646-9>
- Din, J. N., Harding, S. A., Valerio, C. J., Sarma, J., Lyall, K., Riemersma, R. A., Newby, D. E., & Flapan, A. D. (2008). Dietary intervention with oil rich fish reduces platelet-monocyte aggregation in man. *Atherosclerosis*, 197(1), 290–296. <https://doi.org/10.1016/j.atherosclerosis.2007.04.047>
- Doane-Weideman, T., & Liescheskii, P. (2004). Analytical supercritical fluid extraction for food applications. In L. D.L. (Ed.), *Oil Extraction and Analysis: Critical Issues and Competitive Studies* (Issue 4, pp. 69–99). AOCS Publishing. <https://doi.org/10.1201/9781439822340.ch5>
- Du, Z., Hu, B., Shi, A., Ma, X., Cheng, Y., Chen, P., Liu, Y., Lin, X., & Ruan, R. (2012). Cultivation of a microalga Chlorella vulgaris using recycled aqueous phase nutrients from hydrothermal carbonization process. *Bioresource Technology*, 126, 354–357. <https://doi.org/10.1016/j.biortech.2012.09.062>
- Dutka, M., Ditaranto, M., & Løvås, T. (2015). Application of a central composite design for the study of NOx emission performance of a low NOx burner. *Energies*, 8(5), 3606–3627. <https://doi.org/10.3390/en8053606>
- Dyall, S. C. (2015). Long-chain omega-3 fatty acids and the brain: a review of the independent and shared effects of EPA, DPA and DHA. *Frontiers in Aging Neuroscience*, 7(April), 1–15. <https://doi.org/10.3389/fnagi.2015.00052>
- Eckelberry, N., Green, M. P., & Fraser, S. A. (2013). Systems and methods for extracting non-polar lipids from an aqueous algae slurry and lipids produced therefrom (Patent No. US9085745B2). In *United States Patent Application Publication* (US9085745B2).
- Egli, T. (2015). Microbial growth and physiology: a call for better craftsmanship. *Frontiers in Microbiology*, 06. <https://doi.org/10.3389/fmicb.2015.00287>
- Fakirov, S. (2006). Modified soxhlet apparatus for high-temperature extraction. *Journal of Applied Polymer Science*, 102(2), 2013–2014. <https://doi.org/10.1002/app.23397>

- Fan, W., Philip, S., Granowitz, C., Toth, P. P., & Wong, N. D. (2020). Prevalence of US Adults with Triglycerides  $\geq$  150 mg/dl: NHANES 2007–2014. *Cardiology and Therapy*, 9(1), 207–213. <https://doi.org/10.1007/s40119-020-00170-x>
- Fang, J., Dasgupta, S., Zhang, L., & Zhao, W. (2019). Lipid Biomarkers in Geomicrobiology: Analytical Techniques and Applications. *Analytical Geomicrobiology*, July, 341–359. <https://doi.org/10.1017/9781107707399.014>
- Feller, R., Matos, Â. P., Mazzutti, S., Moecke, E. H. S., Tres, M. V., Derner, R. B., Oliveira, J. V., & Junior, A. F. (2017). Polyunsaturated  $\omega$ -3 and  $\omega$ -6 fatty acids, total carotenoids and antioxidant activity of three marine microalgae extracts obtained by supercritical CO<sub>2</sub> and subcritical n-butane. *The Journal of Supercritical Fluids*, 133(November 2017), 437–443. <https://doi.org/10.1016/j.supflu.2017.11.015>
- Fleischer, D., Jukic, M., Thompson, A., & Radaelli, G. (2014). Systems and methods for extracting lipids from and dehydrating wet algal biomass (Patent No. US8765983B2). In *United States Patent* (US8765983B2).
- Fonollá, J., López-Huertas, E., Machado, F. J., Molina, D., Álvarez, I., Márquez, E., Navas, M., Palacín, E., García-Valls, M. J., Remón, B., Boza, J. J., & Martí, J. L. (2009). Milk enriched with “healthy fatty acids” improves cardiovascular risk markers and nutritional status in human volunteers. *Nutrition*, 25(4), 408–414. <https://doi.org/10.1016/j.nut.2008.10.008>
- Freitas, A. C., Rodrigues, D., Rocha-Santos, T. A. P., Gomes, A. M. P., & Duarte, A. C. (2012). Marine biotechnology advances towards applications in new functional foods. *Biotechnology Advances*, 30(6), 1506–1515. <https://doi.org/10.1016/j.biotechadv.2012.03.006>
- Garcia Alba, L., Torri, C., Fabbri, D., Kersten, S. R. A., & Wim Brilman, D. W. F. (2013). Microalgae growth on the aqueous phase from Hydrothermal Liquefaction of the same microalgae. *Chemical Engineering Journal*, 228, 214–223. <https://doi.org/10.1016/j.cej.2013.04.097>
- Geppert, J., Kraft, V., Demmelmair, H., & Koletzko, B. (2006). Microalgal docosahexaenoic acid decreases plasma triacylglycerol in normolipidaemic vegetarians: a randomised trial. *The British Journal of Nutrition*, 95(4), 779–786. <https://doi.org/10.1079/BJN20051720>
- Ginsberg, G. L., & Toal, B. F. (2009). Quantitative approach for incorporating methylmercury risks and omega 3 fatty acid benefits in developing species specific fish consumption advice. *Environmental Health Perspectives*, 117, 267–275. <https://doi.org/10.1289/ehp.11368>
- Goris, K., Muylaert, K., Fraeye, I., Foubert, I., De Brabanter, J., & De Cooman, L. (2012). Antioxidant potential of microalgae in relation to their phenolic and carotenoid content. *Journal of Applied Phycology*, 24(6), 1477–1486. <https://doi.org/10.1007/s10811-012-9804-6>

- Gow, R. V., & Hibbeln, J. R. (2014). Omega-3 and treatment implications in Attention Deficit Hyperactivity Disorder (ADHD) and associated behavioral symptoms. *Lipid Technology*, 26(1), 7–10. <https://doi.org/10.1002/lite.201400002>
- Griffiths, M. J., Van Hille, R. P., & Harrison, S. T. L. (2010). Selection of direct transesterification as the preferred method for assay of fatty acid content of microalgae. *Lipids*, 45(11), 1053–1060. <https://doi.org/10.1007/s11745-010-3468-2>
- Griffiths, Melinda J., van Hille, R. P., & Harrison, S. T. L. (2012). Lipid productivity, settling potential and fatty acid profile of 11 microalgal species grown under nitrogen replete and limited conditions. *Journal of Applied Phycology*, 24(5), 989–1001. <https://doi.org/10.1007/s10811-011-9723-y>
- Guedes, A. C., Amaro, H. M., & Malcata, F. X. (2011). Microalgae as sources of carotenoids. *Marine Drugs*, 9(4), 625–644. <https://doi.org/10.3390/md9040625>
- Guihéneuf, F., Fouqueray, M., Mimouni, V., Ulmann, L., Jacquette, B., & Tremblin, G. (2010). Effect of UV stress on the fatty acid and lipid class composition in two marine microalgae *Pavlova lutheri* (Pavlovophyceae) and *Odontella aurita* (Bacillariophyceae). *Journal of Applied Phycology*, 22(5), 629–638. <https://doi.org/10.1007/s10811-010-9503-0>
- Guihéneuf, F., Mimouni, V., Ulmann, L., & Tremblin, G. (2009). Combined effects of irradiance level and carbon source on fatty acid and lipid class composition in the microalga *Pavlova lutheri* commonly used in mariculture. *Journal of Experimental Marine Biology and Ecology*, 369(2), 136–143. <https://doi.org/10.1016/j.jembe.2008.11.009>
- Halim, R., Gladman, B., Danquah, M. K., & Webley, P. a. (2011). Oil extraction from microalgae for biodiesel production. *Bioresource Technology*, 102(1), 178–185. <https://doi.org/10.1016/j.biortech.2010.06.136>
- Handayania, N. A., & Ariyantib, D. (2012). Potential Production of Polyunsaturated Fatty Acids from Microalgae. *Journal of Bioprocessing & Biotechniques*, 01(1), 13–16. <https://doi.org/10.4172/scientificreports.180>
- Harris, W. S., Pottala, J. V., Sands, S. A., & Jones, P. G. (2007). Comparison of the effects of fish and fish-oil capsules on the n – 3 fatty acid content of blood cells and plasma phospholipids 1 – 3. 1621–1625.
- Hemaiswarya, S., Raja, R., Kumar, R. R., Ganesan, V., & Anbazhagan, C. (2011). Microalgae: A sustainable feed source for aquaculture. *World Journal of Microbiology and Biotechnology*, 27(8), 1737–1746. <https://doi.org/10.1007/s11274-010-0632-z>
- Herrero, M., Cifuentes, A., & Ibanez, E. (2006). Sub- and supercritical fluid extraction of functional ingredients from different natural sources: Plants, food-by-products, algae and microalgaeA review. *Food Chemistry*, 98(1),

- 136–148. <https://doi.org/10.1016/j.foodchem.2005.05.058>
- Herrero, M., & Ibáñez, E. (2015). Green processes and sustainability: An overview on the extraction of high added-value products from seaweeds and microalgae. *Journal of Supercritical Fluids*, 96, 211–216. <https://doi.org/10.1016/j.supflu.2014.09.006>
- Hietala, D. C., Faeth, J. L., & Savage, P. E. (2016). A quantitative kinetic model for the fast and isothermal hydrothermal liquefaction of *Nannochloropsis* sp. *Bioresource Technology*, 214, 102–111. <https://doi.org/10.1016/j.biortech.2016.04.067>
- Ho, B. C. H., Kamal, S. M. M., Danquah, M. K., & Harun, R. (2018). Optimization of Subcritical Water Extraction (SWE) of Lipid and Eicosapentaenoic Acid (EPA) from *Nannochloropsis gaditana*. *BioMed Research International*, 2018. <https://doi.org/10.1155/2018/8273581>
- Hognon, C., Delrue, F., & Boissonnet, G. (2015). Energetic and economic evaluation of *Chlamydomonas reinhardtii* hydrothermal liquefaction and pyrolysis through thermochemical models. *Energy*, 93, 31–40. <https://doi.org/10.1016/j.energy.2015.09.021>
- Hu, W., Fitzgerald, M., Topp, B., Alam, M., & O'Hare, T. J. (2019). A review of biological functions, health benefits, and possible de novo biosynthetic pathway of palmitoleic acid in macadamia nuts. *Journal of Functional Foods*, 62(June), 103520. <https://doi.org/10.1016/j.jff.2019.103520>
- Hunter, S. E., & Savage, P. E. (2004). Recent advances in acid- and base-catalyzed organic synthesis in high-temperature liquid water. *Chemical Engineering Science*. <https://doi.org/10.1016/j.ces.2004.09.009>
- Institute for Public Health Malaysia. (2019). *National Health and Morbidity Survey 2019* (Vol. 1). <http://www.iku.gov.my/nhms-2019>
- Jacob, J. S., & Miller, K. R. (1986). The effects of galactolipid depletion on the structure of a photosynthetic membrane. *The Journal of Cell Biology*. <https://doi.org/10.1083/jcb.103.4.1337>
- Jacobson, T. A., Miller, M., & Schaefer, E. J. (2007). Hypertriglyceridemia and cardiovascular risk reduction. *Clinical Therapeutics*, 29(5), 763–777. <https://doi.org/http://dx.doi.org/10.1016/j.clinthera.2007.05.002>
- Jans, L. a W., Giltay, E. J., & Van der Does, a J. W. (2010). The efficacy of n-3 fatty acids DHA and EPA (fish oil) for perinatal depression. *The British Journal of Nutrition*, 104(11), 1577–1585. <https://doi.org/10.1017/S0007114510004125>
- Jeevan Kumar, S. P., Vijay Kumar, G., Dash, A., Scholz, P., & Banerjee, R. (2017). Sustainable green solvents and techniques for lipid extraction from microalgae: A review. *Algal Research*, 21, 138–147. <https://doi.org/10.1016/j.algal.2016.11.014>

- Kagan, M. L., Levy, A., & Leikin-Frenkel, A. (2015). Comparative study of tissue deposition of omega-3 fatty acids from polar-lipid rich oil of the microalgae *Nannochloropsis oculata* with krill oil in rats. *Food Funct.*, 6(1), 185–191. <https://doi.org/10.1039/C4FO00591K>
- Kagan, M. L., West, A. L., Zante, C., & Calder, P. C. (2013). Acute appearance of fatty acids in human plasma - A comparative study between polar-lipid rich oil from the microalgae *Nannochloropsis oculata* and krill oil in healthy young males. *Lipids in Health and Disease*, 12(1), 1. <https://doi.org/10.1186/1476-511X-12-102>
- Kale, A. (2012a). Nondisruptive methods of extracting algal components for production of carotenoids, omega-3 fatty acids and biofuels (Patent No. US8313647B2). In *United States Patent* (US8313647B2).
- Kale, A. (2012b). Products from step-wise extraction of algal biomasses (Patent No. US20120055079A1). In *United States Patent Application Publication* (US20120055079A1).
- Karagas, M. R., Choi, A. L., Oken, E., Horvat, M., Schoeny, R., Kamai, E., Cowell, W., Grandjean, P., & Korrick, S. (2012). Evidence on the Human Health Effects of Low-Level Methylmercury Exposure. *Environmental Health Perspectives*, 120(6), 799–806. <https://doi.org/10.1289/ehp.1104494>
- Kastelein, J. J. P., Maki, K. C., Susekov, A., Ezhov, M., Nordestgaard, B. G., Machielse, B. N., Kling, D., & Davidson, M. H. (2014). Omega-3 free fatty acids for the treatment of severe hypertriglyceridemia: The EpanoVa for Lowering Very high triglyceridEs (EVOLVE) trial. *Journal of Clinical Lipidology*, 8(1), 94–106. <https://doi.org/10.1016/j.jacl.2013.10.003>
- Kaye, Y., Grundman, O., Leu, S., Zarka, A., Zorin, B., Didi-Cohen, S., Khozin-Goldberg, I., & Boussiba, S. (2015). Metabolic engineering toward enhanced LC-PUFA biosynthesis in *Nannochloropsis oceanica*: Overexpression of endogenous Δ12 desaturase driven by stress-inducible promoter leads to enhanced deposition of polyunsaturated fatty acids in TAG. *Algal Research*. <https://doi.org/10.1016/j.algal.2015.05.003>
- Ke, M., Zhou, A., Song, Z., & Jiang, Q. (2007). Toxicity of ionic liquids. *Progress in Chemistry*. <https://doi.org/10.1002/cjen.200600015>
- Kelley, D. S., Siegel, D., Vemuri, M., Chung, G. H., & Mackey, B. E. (2008). Docosahexaenoic acid supplementation decreases remnant-like particle-cholesterol and increases the (n-3) index in hypertriglyceridemic men. *The Journal of Nutrition*, 138(1), 30–35. <http://www.ncbi.nlm.nih.gov/pubmed/18156400>
- Khadidja, R. (2017). Method for enriching protists with lipids rich in polyunsaturated fatty acids, more particularly of the omega 3 class, and implementation of same for the production of said lipids (Patent No. WO2017103421A1).

- Kim, J. H., Kim, Y., Kim, Y. J., & Park, Y. (2016). Conjugated Linoleic Acid: Potential Health Benefits as a Functional Food Ingredient. *Annual Review of Food Science and Technology*, 7(1), 221–244. <https://doi.org/10.1146/annurev-food-041715-033028>
- Koba, K., & Yanagita, T. (2014). Health benefits of conjugated linoleic acid (CLA). *Obesity Research and Clinical Practice*, 8(6), e525–e532. <https://doi.org/10.1016/j.orcp.2013.10.001>
- Kobayashi, H., Hoshina, R., & Katagiri, K. (2018). *Method for producing dha-containing glyceride-containing composition* (Patent No. US20180051304A1). United States Patent Application Publication.
- Kolanowski, W., Jaworska, D., & Weißbrodt, J. (2007). Importance of instrumental and sensory analysis in the assessment of oxidative deterioration of omega-3 long-chain polyunsaturated fatty acid-rich foods. *Journal of the Science of Food and Agriculture*, 87(2), 181–191. <https://doi.org/10.1002/jsfa.2733>
- Krichnavaruk, S., Shotipruk, A., Goto, M., & Pavasant, P. (2008). Supercritical carbon dioxide extraction of astaxanthin from Haematococcus pluvialis with vegetable oils as co-solvent. *Bioresource Technology*, 99(13), 5556–5560. <https://doi.org/10.1016/j.biortech.2007.10.049>
- Kris-Etherton, P. M., Harris, W. S., & Appel, L. J. (2002). Fish consumption, fish oil, omega-3 fatty acids, and cardiovascular disease. *Circulation*, 106(21), 2747–2757. <https://doi.org/10.1161/01.CIR.0000038493.65177.94>
- Kuhnt, K., Degen, C., Jaudszus, A., & Jahreis, G. (2012). Searching for health beneficial n-3 and n-6 fatty acids in plant seeds. *European Journal of Lipid Science and Technology*, 114(2), 153–160. <https://doi.org/10.1002/ejlt.201100008>
- Kumari, P., Reddy, C. R. K., & Jha, B. (2011). Comparative evaluation and selection of a method for lipid and fatty acid extraction from macroalgae. *Analytical Biochemistry*, 415(2), 134–144. <https://doi.org/10.1016/j.ab.2011.04.010>
- Kunz, W., & Häckl, K. (2016). The hype with ionic liquids as solvents. *Chemical Physics Letters*, 661, 6–12. <https://doi.org/10.1016/j.cplett.2016.07.044>
- Kusdiana, D., & Saka, S. (2004). Two-Step Preparation for Catalyst-Free Biodiesel Fuel Production. In *Proceedings of the Twenty-Fifth Symposium on Biotechnology for Fuels and Chemicals Held May 4–7, 2003, in Breckenridge, CO*. [https://doi.org/10.1007/978-1-59259-837-3\\_63](https://doi.org/10.1007/978-1-59259-837-3_63)
- Kwantes, J. M., & Grundmann, O. (2015). A Brief Review of Krill Oil History, Research, and the Commercial Market. *Journal of Dietary Supplements*, 12(1), 23–35. <https://doi.org/10.3109/19390211.2014.902000>
- Lane, K., Derbyshire, E., Li, W., & Brennan, C. (2014). Bioavailability and potential uses of vegetarian sources of omega-3 fatty acids: A review of

- the literature. *Critical Reviews in Food Science and Nutrition*, 54(5), 572–579. <https://doi.org/10.1080/10408398.2011.596292>
- Lane, K. E., Li, W., Smith, C., & Derbyshire, E. (2014). The bioavailability of an omega-3-rich algal oil is improved by nanoemulsion technology using yogurt as a food vehicle. *International Journal of Food Science and Technology*. <https://doi.org/10.1111/ijfs.12455>
- Lee, J.-Y., Yoo, C., Jun, S.-Y., Ahn, C.-Y., & Oh, H.-M. (2010). Comparison of several methods for effective lipid extraction from microalgae. *Bioresource Technology*, 101(1), S75–S77. <https://doi.org/10.1016/j.biortech.2009.03.058>
- Lee, S. Y., Cho, J. M., Chang, Y. K., & Oh, Y. K. (2017). Cell disruption and lipid extraction for microalgal biorefineries: A review. *Bioresource Technology*, 244, 1317–1328. <https://doi.org/10.1016/j.biortech.2017.06.038>
- Lemke, S., Vicini, J., & Su, H. (2010). Dietary intake of stearidonic acid – enriched soybean oil increases the omega-3 index : randomized , double-blind clinical study of efficacy and safety 1 – 3. *Journal of Clinical*, 766–775. <https://doi.org/10.3945/ajcn.2009.29072.766>
- Lenihan-Geels, G., Bishop, K. S., & Ferguson, L. R. (2013). Alternative sources of omega-3 fats: Can we find a sustainable substitute for fish? *Nutrients*, 5(4), 1301–1315. <https://doi.org/10.3390/nu5041301>
- Lenti, M., Gentili, C., Pianezzi, A., Marcolongo, G., Lalli, A., Cancedda, R., & Cancedda, F. D. (2009). Monogalactosyldiacylglycerol anti-inflammatory activity on adult articular cartilage. *Natural Product Research*, 23(8), 754–762. <https://doi.org/10.1080/14786410802456956>
- Leow, S., Witter, J. R., Vardon, D. R., Sharma, B. K., Guest, J. S., & Strathmann, T. J. (2015). Prediction of microalgae hydrothermal liquefaction products from feedstock biochemical composition. *Green Chem.*, 17(6), 3584–3599. <https://doi.org/10.1039/C5GC00574D>
- Lepage, G., & Roy, C. C. (1984). Improved recovery of fatty acid through direct transesterification without prior extraction or purification. *Journal of Lipid Research*, 25(12), 1391–1396.
- Li, G., Sinclair, A. J., & Li, D. (2011). Comparison of lipid content and fatty acid composition in the edible meat of wild and cultured freshwater and marine fish and shrimps from China. *Journal of Agricultural and Food Chemistry*, 59(5), 1871–1881. <https://doi.org/10.1021/jf104154q>
- Li, P., Feng, X., & Qiu, G. (2010). Methylmercury exposure and health effects from rice and fish consumption: A review. *International Journal of Environmental Research and Public Health*, 7(6), 2666–2691. <https://doi.org/10.3390/ijerph7062666>
- Li, X., Liu, J., Chen, G., Zhang, J., Wang, C., & Liu, B. (2019). Extraction and

- purification of eicosapentaenoic acid and docosahexaenoic acid from microalgae: A critical review. *Algal Research*, 43(December 2018), 101619. <https://doi.org/10.1016/j.algal.2019.101619>
- Li, Z., Smith, K. H., & Stevens, G. W. (2016). The use of environmentally sustainable bio-derived solvents in solvent extraction applications - A review. *Chinese Journal of Chemical Engineering*, 24(2), 215–220. <https://doi.org/10.1016/j.cjche.2015.07.021>
- Liu, J., Sempos, C. T., Donahue, R. P., Dorn, J., Trevisan, M., & Grundy, S. M. (2006). Non-high-density lipoprotein and very-low-density lipoprotein cholesterol and their risk predictive values in coronary heart disease. *American Journal of Cardiology*, 98(10), 1363–1368. <https://doi.org/10.1016/j.amjcard.2006.06.032>
- Liu, T., Jiang, H., Gao, L., Zhang, W., & Chen, Y. (2010). Method for simultaneously extracting lipid and protein from microalgae (Patent No. CN101429467B). In *State Intellectual Property of the P.R.C* (No. CN101429467B).
- López Barreiro, D., Bauer, M., Hornung, U., Posten, C., Kruse, A., & Prins, W. (2015). Cultivation of microalgae with recovered nutrients after hydrothermal liquefaction. *Algal Research*, 9, 99–106. <https://doi.org/10.1016/j.algal.2015.03.007>
- López Barreiro, D., Zamalloa, C., Boon, N., Vyverman, W., Ronsse, F., Brilman, W., & Prins, W. (2013). Influence of strain-specific parameters on hydrothermal liquefaction of microalgae. *Bioresource Technology*, 146, 463–471. <https://doi.org/10.1016/j.biortech.2013.07.123>
- Lu, Y., Levine, R. B., & Savage, P. E. (2015). Fatty acids for nutraceuticals and biofuels from hydrothermal carbonization of microalgae. *Industrial & Engineering Chemistry Research*, 54(16), 4066–4071. <https://doi.org/10.1021/ie503448u>
- Luong, D., Sephton, M. A., & Watson, J. S. (2015). Subcritical water extraction of organic matter from sedimentary rocks. *Analytica Chimica Acta*, 879(April), 48–57. <https://doi.org/10.1016/j.aca.2015.04.027>
- Luque de Castro, M. D., & Priego-Capote, F. (2010). Soxhlet extraction: Past and present panacea. *Journal of Chromatography A*, 1217(16), 2383–2389. <https://doi.org/10.1016/j.chroma.2009.11.027>
- Ma, X. N., Chen, T. P., Yang, B., Liu, J., & Chen, F. (2016). Lipid production from *Nannochloropsis*. *Marine Drugs*, 14(4). <https://doi.org/10.3390/md14040061>
- Macías-Sánchez, M. D., Mantell, C., Rodríguez, M., Martínez de la Ossa, E., Lubián, L. M., & Montero, O. (2009). Comparison of supercritical fluid and ultrasound-assisted extraction of carotenoids and chlorophyll a from *Dunaliella salina*. *Talanta*, 77(3), 948–952. <https://doi.org/10.1016/j.talanta.2008.07.032>

- Mahaffey, K. R., Sunderland, E. M., Chan, H. M., Choi, A. L., Grandjean, P., Mariën, K., Oken, E., Sakamoto, M., Schoeny, R., Weihe, P., Yan, C. H., & Yasutake, A. (2011). Balancing the benefits of n-3 polyunsaturated fatty acids and the risks of methylmercury exposure from fish consumption. *Nutrition Reviews*, 69(9), 493–508. <https://doi.org/10.1111/j.1753-4887.2011.00415.x>
- Maity, J. P., Bundschuh, J., Chen, C. Y., & Bhattacharya, P. (2014). Microalgae for third generation biofuel production, mitigation of greenhouse gas emissions and wastewater treatment: Present and future perspectives - A mini review. *Energy*. <https://doi.org/10.1016/j.energy.2014.04.003>
- Maki, K. C., Orloff, D. G., Nicholls, S. J., Dunbar, R. L., Roth, E. M., Curcio, D., Johnson, J., Kling, D., & Davidson, M. H. (2013). A highly bioavailable omega-3 free fatty acid formulation improves the cardiovascular risk profile in high-risk, statin-treated patients with residual hypertriglyceridemia (the ESPRIT Trial). *Clinical Therapeutics*, 35(9), 1400-1411.e3. <https://doi.org/10.1016/j.clinthera.2013.07.420>
- Maki, K. C., Reeves, M. S., Farmer, M., Griniari, M., Berge, K., Vik, H., Hubacher, R., & Rains, T. M. (2009). Krill oil supplementation increases plasma concentrations of eicosapentaenoic and docosahexaenoic acids in overweight and obese men and women. *Nutrition Research*, 29(9), 609–615. <https://doi.org/10.1016/j.nutres.2009.09.004>
- Makni, M., Fetoui, H., Gargouri, N. K., Garoui, E. M., Jaber, H., Makni, J., Boudawara, T., & Zeghal, N. (2008). Hypolipidemic and hepatoprotective effects of flax and pumpkin seed mixture rich in ω-3 and ω-6 fatty acids in hypercholesterolemic rats. *Food and Chemical Toxicology*, 46(12), 3714–3720. <https://doi.org/10.1016/j.fct.2008.09.057>
- Makni, Mohamed, Fetoui, H., Garoui, E. M., Gargouri, N. K., Jaber, H., Makni, J., Boudawara, T., & Zeghal, N. (2010). Hypolipidemic and hepatoprotective seeds mixture diet rich in ω-3 and ω-6 fatty acids. *Food and Chemical Toxicology*, 48(8–9), 2239–2246. <https://doi.org/10.1016/j.fct.2010.05.055>
- Malaysia Healthcare Performance Unit. (2017). *Malaysian Mental Healthcare Performance: Technical report 2016* (Issue December).
- Mamidipally, P. K., & Liu, S. X. (2004). First approach on rice bran oil extraction using limonene. *European Journal of Lipid Science and Technology*, 106(2), 122–125. <https://doi.org/10.1002/ejlt.200300891>
- Mandal, C. C., Ghosh-Choudhury, T., Yoneda, T., Choudhury, G. G., & Ghosh-Choudhury, N. (2010). Fish oil prevents breast cancer cell metastasis to bone. *Biochemical and Biophysical Research Communications*, 402(4), 602–607. <https://doi.org/10.1016/j.bbrc.2010.10.063>
- Markou, G., & Georgakakis, D. (2011). Cultivation of filamentous cyanobacteria (blue-green algae) in agro-industrial wastes and wastewaters: A review. *Applied Energy*, 88(10), 3389–3401.

<https://doi.org/10.1016/j.apenergy.2010.12.042>

- Marrone, B. L., Lacey, R. E., Anderson, D. B., Bonner, J., Coons, J., Dale, T., Downes, C. M., Fernando, S., Fuller, C., Goodall, B., Holladay, J. E., Kadam, K., Kalb, D., Liu, W., Mott, J. B., Nikolov, Z., Ogden, K. L., Sayre, R. T., Trewyn, B. G., & Olivares, J. A. (2018). Review of the harvesting and extraction program within the National Alliance for Advanced Biofuels and Bioproducts. *Algal Research*, 33(June 2016), 470–485. <https://doi.org/10.1016/j.algal.2017.07.015>
- Martins, J. G. (2009). EPA but not DHA appears to be responsible for the efficacy of omega-3 long chain polyunsaturated fatty acid supplementation in depression: Evidence from a meta-analysis of randomized controlled trials. *Journal of the American College of Nutrition*. <https://doi.org/10.1080/07315724.2009.10719785>
- Mat Rifin, H., Robert Lourdes, T. G., Abdul Majid, N. L., Abd Hamid, H. A., Rodzlan Hasani, W. S., Ling, M. Y., Saminathan, T. A., Ismail, H., Mohd Yusoff, M. F., & Omar, M. A. (2018). Hypercholesterolemia Prevalence, Awareness, Treatment and Control among Adults in Malaysia: The 2015 National Health and Morbidity Survey, Malaysia. *Global Journal of Health Science*, 10(7). <https://doi.org/10.5539/gjhs.v10n7p11>
- Mathimani, T., Baldinelli, A., Rajendran, K., Prabakar, D., Matheswaran, M., Pieter van Leeuwen, R., & Pugazhendhi, A. (2019). Review on cultivation and thermochemical conversion of microalgae to fuels and chemicals: Process evaluation and knowledge gaps. *Journal of Cleaner Production*, 208, 1053–1064. <https://doi.org/10.1016/j.jclepro.2018.10.096>
- Mathimani, T., & Mallick, N. (2019). A review on the hydrothermal processing of microalgal biomass to bio-oil - Knowledge gaps and recent advances. *Journal of Cleaner Production*, 217, 69–84. <https://doi.org/10.1016/j.jclepro.2019.01.129>
- Matos, Â. P. (2017). The Impact of Microalgae in Food Science and Technology. *JAOCS, Journal of the American Oil Chemists' Society*, 94(11), 1333–1350. <https://doi.org/10.1007/s11746-017-3050-7>
- Matos, Â. P., Feller, R., Moecke, E. H. S., & Sant'Anna, E. S. (2015). Biomass, lipid productivities and fatty acids composition of marine *Nannochloropsis gaditana* cultured in desalination concentrate. *Bioresource Technology*, 197, 48–55. <https://doi.org/10.1016/j.biortech.2015.08.041>
- Mazereeuw, G., Herrmann, N., Andreazza, A. C., Scola, G., Ma, D. W. L., Oh, P. I., & Lanctôt, K. L. (2017). Oxidative stress predicts depressive symptom changes with omega-3 fatty acid treatment in coronary artery disease patients. *Brain, Behavior, and Immunity*. <https://doi.org/10.1016/j.bbi.2016.10.005>
- Memon, N. N., Talpur, F. N., Bhanger, M. I., & Balouch, A. (2011). Changes in fatty acid composition in muscle of three farmed carp fish species (*Labeo rohita*, *Cirrhinus mrigala*, *Catla catla*) raised under the same conditions.

*Food Chemistry*, 126(2), 405–410.  
<https://doi.org/10.1016/j.foodchem.2010.10.107>

Meng, Y., Jiang, J., Wang, H., Cao, X., Xue, S., Yang, Q., & Wang, W. (2015). The characteristics of TAG and EPA accumulation in *Nannochloropsis oceanica* IMET1 under different nitrogen supply regimes. *Bioresource Technology*, 179(November 2015), 483–489. <https://doi.org/10.1016/j.biortech.2014.12.012>

Milledge, J. J. (2011). Commercial application of microalgae other than as biofuels: A brief review. *Reviews in Environmental Science and Biotechnology*, 10(1), 31–41. <https://doi.org/10.1007/s11157-010-9214-7>

Milledge, J. J., & Heaven, S. (2013). A review of the harvesting of micro-algae for biofuel production. *Reviews in Environmental Science and Bio/Technology*, 12(2), 165–178. <https://doi.org/10.1007/s11157-012-9301-z>

Milte, C. M., Coates, A. M., Buckley, J. D., Hill, A. M., & Howe, P. R. C. (2008). Dose-dependent effects of docosahexaenoic acid-rich fish oil on erythrocyte docosahexaenoic acid and blood lipid levels. *The British Journal of Nutrition*, 99(5), 1083–1088. <https://doi.org/10.1017/S000711450785344X>

Mocking, R. J. T., Harmsen, I., Assies, J., Koeter, M. W. J., Ruhé, H. G., & Schene, A. H. (2016). Meta-analysis and meta-regression of omega-3 polyunsaturated fatty acid supplementation for major depressive disorder. *Translational Psychiatry*. <https://doi.org/10.1038/tp.2016.29>

Molina Grima, E., Belarbi, E. H., Acién Fernández, F. G., Robles Medina, a., & Chisti, Y. (2003). Recovery of microalgal biomass and metabolites: Process options and economics. *Biotechnology Advances*, 20, 491–515. [https://doi.org/10.1016/S0734-9750\(02\)00050-2](https://doi.org/10.1016/S0734-9750(02)00050-2)

Molino, A., Mehariya, S., Di Sanzo, G., Larocca, V., Martino, M., Leone, G. P., Marino, T., Chianese, S., Balducchi, R., & Musmarra, D. (2020). Recent developments in supercritical fluid extraction of bioactive compounds from microalgae: Role of key parameters, technological achievements and challenges. *Journal of CO<sub>2</sub> Utilization*, 36(September 2019), 196–209. <https://doi.org/10.1016/j.jcou.2019.11.014>

Morse, N. (2015). Lipid-lowering and anti-inflammatory effects of palmitoleic acid: Evidence from human intervention studies. *Lipid Technology*, 27(7), 155–160. <https://doi.org/10.1002/lite.201500033>

Mun, S., Decker, E. A., & McClements, D. J. (2007). Influence of emulsifier type on in vitro digestibility of lipid droplets by pancreatic lipase. *Food Research International*, 40(6), 770–781. <https://doi.org/10.1016/j.foodres.2007.01.007>

Murphy, R. A., Mourtzakis, M., Chu, Q. S. C., Baracos, V. E., Reiman, T., & Mazurak, V. C. (2011). Nutritional intervention with fish oil provides a

- benefit over standard of care for weight and skeletal muscle mass in patients with nonsmall cell lung cancer receiving chemotherapy. *Cancer*, 117(8), 1775–1782. <https://doi.org/10.1002/cncr.25709>
- Mustafa, A., & Turner, C. (2011). Pressurized liquid extraction as a green approach in food and herbal plants extraction: A review. *Analytica Chimica Acta*, 703(1), 8–18. <https://doi.org/10.1016/j.aca.2011.07.018>
- Naidoo, T., Zulu, N., Maharajh, D., & Laloo, R. (2013). Value-added products from microalgae. In F. Bux (Ed.), *Biotechnological applications of microalgae: biodiesel and value-added products* (1st ed., pp. 137–156). CRC Press.
- Nash, S. M. B., Schlabach, M., & Nichols, P. D. (2014). A Nutritional-toxicological assessment of antarctic krill oil versus fish oil dietary supplements. *Nutrients*, 6(9), 3382–3402. <https://doi.org/10.3390/nu6093382>
- Neff, L., & Culiner, J. (2011). Algal docosahexaenoic acid affects plasma lipoprotein particle size distribution in overweight and obese adults. *The Journal of Nutrition*, 141(2), 207–213. <https://doi.org/10.3945/jn.110.130021.information>
- Nelder, J. A. (1998). The selection of terms in response-surface models—how strong is the weak-heredity principle? *American Statistician*, 52(4), 315–318. <https://doi.org/10.1080/00031305.1998.10480588>
- Nguyen, T. T., Zhang, W., Barber, A. R., Su, P., & He, S. (2015). Significant Enrichment of Polyunsaturated Fatty Acids (PUFAs) in the Lipids Extracted by Supercritical CO<sub>2</sub> from the Livers of Australian Rock Lobsters (*Jasus edwardsii*). *Journal of Agricultural and Food Chemistry*, 63(18), 4621–4628. <https://doi.org/10.1021/jf5059396>
- Njoroge, S. M., Koaze, H., Karanja, P. N., & Sawamura, M. (2005). Essential oil constituents of three varieties of Kenyan sweet oranges (*Citrus sinensis*). *Flavour and Fragrance Journal*, 20(1), 80–85. <https://doi.org/10.1002/ffj.1377>
- Oh, D. Y., Talukdar, S., Bae, E. J., Imamura, T., Morinaga, H., Fan, W., Li, P., Lu, W. J., Watkins, S. M., & Olefsky, J. M. (2010). GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects. *Cell*, 142(5), 687–698. <https://doi.org/10.1016/j.cell.2010.07.041>
- Oi, L. E., Choo, M. Y., Lee, H. V., Rahman, N. A., & Juan, J. C. (2019). Mesoporous and other types of catalysts for conversion of non-edible oil to biogasoline via deoxygenation. In *Sustainable Bioenergy: Advances and Impacts*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-817654-2.00009-5>
- Olagunju, A., Muhammad, A., Mada, S. B., Mohammed, A., Mohammed, H. A., & Mahmoud, K. T. (2012). Nutrient Composition of Tilapia zilli , Hemi-

*synodontis membranacea*, *Clupea harengus* and *Scomber scombrus* Consumed in Zaria. *World Journal Life Sciences and Medical Research*, 2, 16–19.

Olkiewicz, M., Caporgno, M. P., Font, J., Legrand, J., Lepine, O., Plechkova, N. V., Pruvost, J., Seddon, K. R., & Bengoa, C. (2015). A novel recovery process for lipids from microalgae for biodiesel production using a hydrated phosphonium ionic liquid. *Green Chemistry*. <https://doi.org/10.1039/c4gc02448f>

Olmstead, I. L. D., Hill, D. R. A., Dias, D. A., Jayasinghe, N. S., Callahan, D. L., Kentish, S. E., Scales, P. J., & Martin, G. J. O. (2013). A quantitative analysis of microalgal lipids for optimization of biodiesel and omega-3 production. *Biotechnology and Bioengineering*, 110(8), 2096–2104. <https://doi.org/10.1002/bit.24844>

Onofrejová, L., Vašíčková, J., Klejdus, B., Stratil, P., Mišurcová, L., Kráčmar, S., Kopecký, J., & Vacek, J. (2010). Bioactive phenols in algae: The application of pressurized-liquid and solid-phase extraction techniques. *Journal of Pharmaceutical and Biomedical Analysis*, 51(2), 464–470. <https://doi.org/10.1016/j.jpba.2009.03.027>

Orr, V. C. A., Plechkova, N. V., Seddon, K. R., & Rehmann, L. (2016). Disruption and Wet Extraction of the Microalgae *Chlorella vulgaris* Using Room-Temperature Ionic Liquids. *ACS Sustainable Chemistry and Engineering*, 4(2), 591–600. <https://doi.org/10.1021/acssuschemeng.5b00967>

Ozogul, Y., Şimşek, A., Ballıkçı, E., & Kenar, M. (2012). The effects of extraction methods on the contents of fatty acids, especially EPA and DHA in marine lipids. *International Journal of Food Sciences and Nutrition*, 63(3), 326–331. <https://doi.org/10.3109/09637486.2011.627844>

Pahl, S. L., Lewis, D. M., King, K. D., & Chen, F. (2012). Heterotrophic growth and nutritional aspects of the diatom *Cyclotella cryptica* (Bacillariophyceae): Effect of nitrogen source and concentration. *Journal of Applied Phycology*, 24(2), 301–307. <https://doi.org/10.1007/s10811-011-9680-5>

Patil, P. D., Dandamudi, K. P. R., Wang, J., Deng, Q., & Deng, S. (2018). Extraction of bio-oils from algae with supercritical carbon dioxide and co-solvents. *Journal of Supercritical Fluids*, 135(September 2017), 60–68. <https://doi.org/10.1016/j.supflu.2017.12.019>

Peoples, G. E., McLennan, P. L., Howe, P. R., & Groeller, H. (2008). Fish oil reduces heart rate and oxygen consumption during exercise. *Journal of Cardiovascular Pharmacology*, 52, 540–547. <https://doi.org/10.1097/FJC.0b013e3181911913>

Piccirilli, A. (2011). Method for obtaining algae extracts and use of these extracts (Patent No. US20110142875A1). In *United States Patent Application Publication* (US20110142875A1).

- Poenie, M., Jones, J., & Beach, J. (2012). Immobilized resins for algal oil extraction (Patent No. US8329449B2). In *United States Patent Application Publication* (US8329449B2).
- Pradhan, R. C., Meda, V., Rout, P. K., Naik, S., & Dalai, A. K. (2010). Supercritical CO<sub>2</sub> extraction of fatty oil from flaxseed and comparison with screw press expression and solvent extraction processes. *Journal of Food Engineering*, 98(4), 393–397. <https://doi.org/10.1016/j.jfoodeng.2009.11.021>
- Pretti, C., Chiappe, C., Pieraccini, D., Gregori, M., Abramo, F., Monni, G., & Intorre, L. (2006). Acute toxicity of ionic liquids to the zebrafish (*Danio rerio*). *Green Chemistry*. <https://doi.org/10.1039/b511554j>
- Qian, L., Wang, S., & Savage, P. E. (2020). Fast and isothermal hydrothermal liquefaction of sludge at different severities: Reaction products, pathways, and kinetics. *Applied Energy*, 260(June 2019), 114312. <https://doi.org/10.1016/j.apenergy.2019.114312>
- Qian, Y., Zuo, C., Tan, J., & He, J. (2007). Structural analysis of bio-oils from sub-and supercritical water liquefaction of woody biomass. *Energy*, 32(3), 196–202. <https://doi.org/10.1016/j.energy.2006.03.027>
- Qu, Z., Zeng, J., Zhang, Y., Liao, Q., Sharma, B. K., Fu, Q., Huang, Y., & Liu, Z. (2018). Hydrothermal cell disruption of *Nannochloropsis* sp. and its influence on lipid extraction. *Algal Research*, 35(September), 407–415. <https://doi.org/10.1016/j.algal.2018.09.015>
- Radaelli, G., & Fleischer, D. (2010). Systems and methods for extracting lipids from wet algal biomass (Patent No. US20100317088A1). In *United States Patent Application Publication* (US20100317088A1).
- Radakovits, R., Jinkerson, R. E., Fuerstenberg, S. I., Tae, H., Settlage, R. E., Boore, J. L., & Posewitz, M. C. (2012). Draft genome sequence and genetic transformation of the oleaginous alga *Nannochloropsis gaditana*. *Nature Communications*, 3, 686. <https://doi.org/10.1038/ncomms1688>
- Rajaram, S. (2014). Health benefits of plant-derived α-linolenic acid. *The American Journal of Clinical Nutrition*, 100(suppl\_1), 443S-448S. <https://doi.org/10.3945/ajcn.113.071514>
- Rajkumar, R., & Sobri Takriff, M. (2016). Prospects of Algae and their Environmental Applications in Malaysia: A Case Study. *Journal of Bioremediation & Biodegradation*, 07(01), 1–12. <https://doi.org/10.4172/2155-6199.1000321>
- Reddy, Harvind K., Muppaneni, T., Sun, Y., Li, Y., Ponnusamy, S., Patil, P. D., Dailey, P., Schaub, T., Holguin, F. O., Dungan, B., Cooke, P., Lammers, P., Voorhies, W., Lu, X., & Deng, S. (2014). Subcritical water extraction of lipids from wet algae for biodiesel production. *Fuel*, 133, 73–81. <https://doi.org/10.1016/j.fuel.2014.04.081>

- Reddy, Harvind Kumar, Muppaneni, T., Ponnusamy, S., Sudasinghe, N., Pegallapati, A., Selvaratnam, T., Seger, M., Dungan, B., Nirmalakhandan, N., Schaub, T., Holguin, F. O., Lammers, P., Voorhies, W., & Deng, S. (2016). Temperature effect on hydrothermal liquefaction of *Nannochloropsis gaditana* and *Chlorella* sp. *Applied Energy*, *165*, 943–951. <https://doi.org/10.1016/j.apenergy.2015.11.067>
- Refsgaard, H. H. F., Brockhoff, P. M. B., & Jensen, B. (2000). Free polyunsaturated fatty acids cause taste deterioration of salmon during frozen storage. *Journal of Agricultural and Food Chemistry*, *48*(8), 3280–3285. <https://doi.org/10.1021/jf000021c>
- Ríos, S. D., Castañeda, J., Torras, C., Farriol, X., & Salvadó, J. (2013). Lipid extraction methods from microalgal biomass harvested by two different paths: Screening studies toward biodiesel production. *Bioresource Technology*, *133*, 378–388. <https://doi.org/10.1016/j.biortech.2013.01.093>
- Rizzi, L., Bochicchio, D., Bargellini, A., Parazza, P., & Simioli, M. (2009). Effects of dietary microalgae, other lipid sources, inorganic selenium and iodine on yolk n-3 fatty acid composition, selenium content and quality of eggs in laying hens. *Journal of the Science of Food and Agriculture*, *89*(10), 1775–1781. <https://doi.org/10.1002/jsfa.3655>
- Rosenberg, J. N., Oyler, G. A., Wilkinson, L., & Betenbaugh, M. J. (2008). A green light for engineered algae: redirecting metabolism to fuel a biotechnology revolution. *Current Opinion in Biotechnology*, *19*(5), 430–436. <https://doi.org/10.1016/j.copbio.2008.07.008>
- Roy, S. Sen, & Pal, R. (2015). Microalgae in Aquaculture: A Review with Special References to Nutritional Value and Fish Dietetics. *Proceedings of the Zoological Society*, *68*(1), 1–8. <https://doi.org/10.1007/s12595-013-0089-9>
- Rubio-Rodríguez, N., De Diego, S. M., Beltrán, S., Jaime, I., Sanz, M. T., & Rovira, J. (2012). Supercritical fluid extraction of fish oil from fish by-products: A comparison with other extraction methods. *Journal of Food Engineering*, *109*(2), 238–248. <https://doi.org/10.1016/j.jfoodeng.2011.10.011>
- Russell, F., & Bürgin-Maunder, C. (2012). Distinguishing Health Benefits of Eicosapentaenoic and Docosahexaenoic Acids. *Marine Drugs*, *10*(12), 2535–2559. <https://doi.org/10.3390/md10112535>
- Ryan, A., Keske, M., Hoffman, J., & Nelson, E. (2009). Clinical overview of algal-docosahexaenoic acid: effects on triglyceride levels and other cardiovascular risk factors. *American Journal of Therapeutics*, *16*(2), 183–192. <https://doi.org/10.1097/MJT.0b013e31817fe2be>
- Ryckebosch, E., Bruneel, C., Termote-Verhalle, R., Goiris, K., Muylaert, K., & Fouquet, I. (2014). Nutritional evaluation of microalgae oils rich in omega-3 long chain polyunsaturated fatty acids as an alternative for fish oil. *Food Chemistry*, *160*, 393–400. <https://doi.org/10.1016/j.foodchem.2014.03.087>

- Ryckebosch, E., Muylaert, K., & Foubert, I. (2012). Optimization of an analytical procedure for extraction of lipids from microalgae. *AOCS, Journal of the American Oil Chemists' Society*, 89(2), 189–198. <https://doi.org/10.1007/s11746-011-1903-z>
- Sahena, F., Zaidul, I. S. M., Jinap, S., Karim, A. A., Abbas, K. A., Norulaini, N. A. N., & Omar, A. K. M. (2009). Application of supercritical CO<sub>2</sub> in lipid extraction - A review. *Journal of Food Engineering*, 95(2), 240–253. <https://doi.org/10.1016/j.jfoodeng.2009.06.026>
- Sahena, F., Zaidul, I. S. M., Jinap, S., Saari, N., Jahurul, H. a., Abbas, K. a., & Norulaini, N. a. (2009). PUFAs in fish: Extraction, fractionation, importance in health. *Comprehensive Reviews in Food Science and Food Safety*, 8(2), 59–74. <https://doi.org/10.1111/j.1541-4337.2009.00069.x>
- Salvo, R. Di, Reich, A., Dykes, J. H. W. H., & Teixeira, R. (2013). Lipid extraction from microalgae using a single ionic liquid (Patent No. US8450111B2). In *United States Patent* (US8450111B2).
- Sánchez-Camargo, A. P., Martínez-Correa, H. A., Paviani, L. C., & Cabral, F. A. (2011). Supercritical CO<sub>2</sub> extraction of lipids and astaxanthin from Brazilian redspotted shrimp waste (*Farfantepenaeus paulensis*). *Journal of Supercritical Fluids*, 56(2), 164–173. <https://doi.org/10.1016/j.supflu.2010.12.009>
- Sanders, T. A. B., Gleason, K., Griffin, B., & Miller, G. J. (2006). Influence of an algal triacylglycerol containing docosahexaenoic acid (22:6n-3) and docosapentaenoic acid (22:5n-6) on cardiovascular risk factors in healthy men and women. *British Journal of Nutrition*, 95(03), 525. <https://doi.org/10.1079/BJN20051658>
- Sargi, S. C., Silva, B. C., Santos, H. M. C., Montanher, P. F., Boeing, J. S., Santos Júnior, O. O., Souza, N. E., & Visentainer, J. V. (2013). Antioxidant capacity and chemical composition in seeds rich in omega-3: chia, flax, and perilla. *Food Science and Technology (Campinas)*, 33(3), 541–548. <https://doi.org/10.1590/S0101-20612013005000057>
- Satyarthi, J. K., Srinivas, D., & Ratnasamy, P. (2011). Hydrolysis of vegetable oils and fats to fatty acids over solid acid catalysts. *Applied Catalysis A: General*, 391(1–2), 427–435. <https://doi.org/10.1016/j.apcata.2010.03.047>
- Scholz, M. J., Weiss, T. L., Jinkerson, R. E., Jing, J., Roth, R., Goodenough, U., Posewitz, M. C., & Gerken, H. G. (2014). Ultrastructure and Composition of the *Nannochloropsis gaditana* Cell Wall. *Eukaryotic Cell*, 13(11), 1450–1464. <https://doi.org/10.1128/EC.00183-14>
- Schuchardt, J.P., Neubronner, J., Kressel, G., Merkel, M., von Schacky, C., & Hahn, A. (2011). Moderate doses of EPA and DHA from re-esterified triacylglycerols but not from ethyl-esters lower fasting serum triacylglycerols in statin-treated dyslipidemic subjects: Results from a six month randomized controlled trial. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 85(6), 381–386.

<https://doi.org/10.1016/j.plefa.2011.07.006>

- Schuchardt, Jan Philipp, & Hahn, A. (2013). Bioavailability of long-chain omega-3 fatty acids. *Prostaglandins, Leukotrienes and Essential Fatty Acids (PLEFA)*, 89(1), 1–8. <https://doi.org/10.1016/j.plefa.2013.03.010>
- Schuchardt, Jan Philipp, Schneider, I., Meyer, H., Neubronner, J., von Schacky, C., & Hahn, A. (2011). Incorporation of EPA and DHA into plasma phospholipids in response to different omega-3 fatty acid formulations--a comparative bioavailability study of fish oil vs. krill oil. *Lipids in Health and Disease*, 10(1), 145. <https://doi.org/10.1186/1476-511X-10-145>
- Scrimgeour, C. (2005). Chemistry of Fatty Acids. In *Bailey's Industrial Oil and Fat Products*. <https://doi.org/10.1002/047167849X.bio005>
- Shahidi, F., & Ambigaipalan, P. (2018). Omega-3 Polyunsaturated Fatty Acids and Their Health Benefits. *Annual Review of Food Science and Technology*, 9(1), 345–381. <https://doi.org/10.1146/annurev-food-111317-095850>
- Shepherd, C. J., & Jackson, A. J. (2013). Global fishmeal and fish-oil supply: inputs, outputs and markets a. *Journal of Fish Biology*, 83(4), 1046–1066. <https://doi.org/10.1111/jfb.12224>
- Simionato, D., Block, M. A., La Rocca, N., Jouhet, J., Marechal, E., Finazzi, G., & Morosinotto, T. (2013). The Response of *Nannochloropsis gaditana* to Nitrogen Starvation Includes De Novo Biosynthesis of Triacylglycerols, a Decrease of Chloroplast Galactolipids, and Reorganization of the Photosynthetic Apparatus. *Eukaryotic Cell*, 12(5), 665–676. <https://doi.org/10.1128/EC.00363-12>
- Simopoulos, A. P. (2006). Evolutionary aspects of diet, the omega-6/omega-3 ratio and genetic variation: nutritional implications for chronic diseases. *Biomedicine & Pharmacotherapy*, 60(9), 502–507. <https://doi.org/10.1016/j.biopha.2006.07.080>
- Sitthithanaboon, W., Reddy, H. K., Muppaneni, T., Ponnusamy, S., Punsuvon, V., Holguim, F., Dungan, B., & Deng, S. (2015). Single-step conversion of wet *Nannochloropsis gaditana* to biodiesel under subcritical methanol conditions. *Fuel*, 147, 253–259. <https://doi.org/http://dx.doi.org/10.1016/j.fuel.2015.01.051>
- Solana, M., Rizza, C. S., & Bertucco, A. (2014). Exploiting microalgae as a source of essential fatty acids by supercritical fluid extraction of lipids: Comparison between *Scenedesmus obliquus*, *Chlorella protothecoides* and *Nannochloropsis salina*. *Journal of Supercritical Fluids*, 92, 311–318. <https://doi.org/10.1016/j.supflu.2014.06.013>
- Spolaore, P., Joannis-Cassan, C., Duran, E., & Isambert, A. (2006). Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*, 101(2), 87–96. <https://doi.org/10.1263/jbb.101.87>

- Sporring, S., Bøwadt, S., Svensmark, B., & Björklund, E. (2005). Comprehensive comparison of classic Soxhlet extraction with Soxtec extraction, ultrasonication extraction, supercritical fluid extraction, microwave assisted extraction and accelerated solvent extraction for the determination of polychlorinated biphenyls . *Journal of Chromatography A*, 1090(1–2), 1–9. <https://doi.org/10.1016/j.chroma.2005.07.008>
- Strigley, C. T., & Mossoba, M. M. (2017). Current Analytical Techniques for Food Lipids. In *Food Safety: Innovative Analytical Tools for Safety Assessment* (Issue 7, pp. 33–64).
- Sublette, M. E., Ellis, S. P., Geant, A. L., & Mann, J. J. (2011). Meta-analysis of the effects of eicosapentaenoic acid (EPA) in clinical trials in depression. *The Journal of Clinical Psychiatry*, 72(12), 1577–1584. <https://doi.org/10.4088/JCP.10m06634>
- Sudasinghe, N., Reddy, H., Csakan, N., Deng, S., Lammers, P., & Schaub, T. (2015). Temperature-Dependent Lipid Conversion and Nonlipid Composition of Microalgal Hydrothermal Liquefaction Oils Monitored by Fourier Transform Ion Cyclotron Resonance Mass Spectrometry. *BioEnergy Research*. <https://doi.org/10.1007/s12155-015-9635-9>
- Suman, K., Kiran, T., Devi, U. K., & Sarma, N. S. (2012). Culture medium optimization and lipid profiling of *Cylindrotheca*, a lipid- and polyunsaturated fatty acid-rich pennate diatom and potential source of eicosapentaenoic acid. *Botanica Marina*, 55(3), 289–299. <https://doi.org/10.1515/bot-2011-0076>
- Surette, M. E., Edens, M., Chilton, F. H., & Tramposch, K. M. (2004). Dietary echium oil increases plasma and neutrophil long-chain (n-3) fatty acids and lowers serum triacylglycerols in hypertriglyceridemic humans. *The Journal of Nutrition*, 134(6), 1406–1411.
- Swanson, D., Block, R., & Mousa, S. a. (2012). Omega-3 fatty acids EPA and DHA: Health benefits throughout life. *Advances in Nutrition: An International Review Journal*, 3(1), 1–7. <https://doi.org/10.3945/an.111.000893>
- Tallima, H., & El Ridi, R. (2018). Arachidonic acid: Physiological roles and potential health benefits – A review. *Journal of Advanced Research*, 11, 33–41. <https://doi.org/10.1016/j.jare.2017.11.004>
- Tang, D. Y. Y., Khoo, K. S., Chew, K. W., Tao, Y., Ho, S. H., & Show, P. L. (2020). Potential utilization of bioproducts from microalgae for the quality enhancement of natural products. *Bioresource Technology*, 304(February), 122997. <https://doi.org/10.1016/j.biortech.2020.122997>
- Tanzi, C. D., Vian, A. M., & Chemat, F. (2013). New procedure for extraction of algal lipids from wet biomass: A green clean and scalable process. *Bioresource Technology*, 134, 271–275. <https://doi.org/10.1016/j.biortech.2013.01.168>

- Taoka, Y., Nagano, N., Okita, Y., Izumida, H., Sugimoto, S., & Hayashi, M. (2011). Effect of Tween 80 on the growth, lipid accumulation and fatty acid composition of *Thraustochytrium aureum* ATCC 34304. *Journal of Bioscience and Bioengineering*, 111(4), 420–424. <https://doi.org/10.1016/j.jbiotec.2010.12.010>
- Taylor, C. G., Noto, A. D., Stringer, D. M., Froese, S., & Malcolmson, L. (2010). Dietary milled flaxseed and flaxseed oil improve N-3 fatty acid status and do not affect glycemic control in individuals with well-controlled type 2 diabetes. *Journal of the American College of Nutrition*, 29(1), 72–80. <https://doi.org/10.1080/07315724.2010.10719819>
- Tocher, D. R., Betancor, M. B., Sprague, M., Olsen, R. E., & Napier, J. A. (2019). Omega-3 long-chain polyunsaturated fatty acids, EPA and DHA: Bridging the gap between supply and demand. *Nutrients*, 11(1), 1–20. <https://doi.org/10.3390/nu11010089>
- Toor, S. S., Rosendahl, L., & Rudolf, A. (2011). Hydrothermal liquefaction of biomass: A review of subcritical water technologies. *Energy*, 36(5), 2328–2342. <https://doi.org/10.1016/j.energy.2011.03.013>
- Tur, J. A., Bibiloni, M. M., Sureda, A., & Pons, A. (2012). Dietary sources of omega 3 fatty acids: public health risks and benefits. *British Journal of Nutrition*, 107(S2), S23–S52. <https://doi.org/10.1017/S0007114512001456>
- Turchini, G. M., Nichols, P. D., Barrow, C., & Sinclair, A. J. (2012). Jumping on the omega-3 bandwagon: distinguishing the role of long-chain and short-chain omega-3 fatty acids. *Critical Reviews in Food Science and Nutrition*, 52(9), 795–803. <https://doi.org/10.1080/10408398.2010.509553>
- Ulivi, V., Lenti, M., Gentili, C., Marcolongo, G., Cancedda, R., & Descalzi Cancedda, F. (2011). Anti-inflammatory activity of monogalactosyldiacylglycerol in human articular cartilage in vitro: activation of an anti-inflammatory cyclooxygenase-2 (COX-2) pathway. *Arthritis Research & Therapy*, 13(3), R92. <https://doi.org/10.1186/ar3367>
- Usydus, Z., Szlinder-Richert, J., Adamczyk, M., & Szatkowska, U. (2011). Marine and farmed fish in the Polish market: Comparison of the nutritional value. *Food Chemistry*, 126(1), 78–84. <https://doi.org/10.1016/j.foodchem.2010.10.080>
- Valdez, P. J., & Savage, P. E. (2013). A reaction network for the hydrothermal liquefaction of *Nannochloropsis* sp. *Algal Research*, 2(4), 416–425. <https://doi.org/10.1016/j.algal.2013.08.002>
- Valdez, P. J., Tocco, V. J., & Savage, P. E. (2014). A general kinetic model for the hydrothermal liquefaction of microalgae. *Bioresource Technology*, 163, 123–127. <https://doi.org/10.1016/j.biortech.2014.04.013>
- van Beelen, V. A., Roeleveld, J., Mooibroek, H., Sijtsma, L., Bino, R. J., Bosch, D., Rietjens, I. M. C. M., & Alink, G. M. (2007). A comparative study on

- the effect of algal and fish oil on viability and cell proliferation of Caco-2 cells. *Food and Chemical Toxicology*, 45(5), 716–724. <https://doi.org/10.1016/j.fct.2006.10.017>
- Van Den Hende, S., Vervaeren, H., Desmet, S., & Boon, N. (2011). Bioflocculation of microalgae and bacteria combined with flue gas to improve sewage treatment. *New Biotechnology*, 29(1), 23–31. <https://doi.org/10.1016/j.nbt.2011.04.009>
- Van Vooren, G., Le Grand, F., Legrand, J., Cuiné, S., Peltier, G., & Pruvost, J. (2012). Investigation of fatty acids accumulation in *Nannochloropsis oculata* for biodiesel application. *Bioresource Technology*, 124, 421–432. <https://doi.org/10.1016/j.biortech.2012.08.009>
- Vanthoor-Koopmans, M., Wijffels, R. H., Barbosa, M. J., & Eppink, M. H. M. (2013). Biorefinery of microalgae for food and fuel. *Bioresource Technology*, 135, 142–149. <https://doi.org/10.1016/j.biortech.2012.10.135>
- Vendramini-Costa, D. B., & Carvalho, J. E. (2012). Molecular link mechanisms between inflammation and cancer. *Current Pharmaceutical Design*, 18(26), 3831–3852. <https://doi.org/10.2174/138161212802083707>
- Virot, M., Tomao, V., Ginies, C., Visinoni, F., & Chemat, F. (2008). Green procedure with a green solvent for fats and oils' determination. Microwave-integrated Soxhlet using limonene followed by microwave Clevenger distillation. *Journal of Chromatography A*, 1196–1197(1–2), 147–152. <https://doi.org/10.1016/j.chroma.2008.04.035>
- Vo, T. K., Lee, O. K., Lee, E. Y., Kim, C. H., Seo, J., Kim, J., & Kim, S. (2016). Kinetics study of the hydrothermal liquefaction of the microalga *Aurantiochytrium* sp. KRS101. *Chemical Engineering Journal*, 306, 763–771. <https://doi.org/10.1016/j.cej.2016.07.104>
- Wahlen, B. D., Willis, R. M., & Seefeldt, L. C. (2011). Biodiesel production by simultaneous extraction and conversion of total lipids from microalgae, cyanobacteria, and wild mixed-cultures. *Bioresource Technology*, 102(3), 2724–2730. <https://doi.org/10.1016/j.biortech.2010.11.026>
- Wall, R., Ross, R. P., Fitzgerald, G. F., & Stanton, C. (2010). Fatty acids from fish: The anti-inflammatory potential of long-chain omega-3 fatty acids. *Nutrition Reviews*, 68(5), 280–289. <https://doi.org/10.1111/j.1753-4887.2010.00287.x>
- Wan, J. B., Huang, L. L., Rong, R., Tan, R., Wang, J., & Kang, J. X. (2010). Endogenously decreasing tissue n-6/n-3 fatty acid ratio reduces atherosclerotic lesions in apolipoprotein E-deficient mice by inhibiting systemic and vascular inflammation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 30(12), 2487–2494. <https://doi.org/10.1161/ATVBAHA.110.210054>
- Wanasundara, U. N., & Shahidi, F. (1999). Concentration of omega 3-polyunsaturated fatty acids of seal blubber oil by urea complexation:

Optimization of reaction conditions. *Food Chemistry*, 65(1), 41–49.  
[https://doi.org/10.1016/S0308-8146\(98\)00153-8](https://doi.org/10.1016/S0308-8146(98)00153-8)

Wang, G., & Wang, T. (2012). Characterization of Lipid Components in Two Microalgae for Biofuel Application. *Journal of the American Oil Chemists' Society*, 89(1), 135–143. <https://doi.org/10.1007/s11746-011-1879-8>

Watson, E. (2011). *Algal omega - 3 market heats up as new players bid for a slice of the action*. <http://www.nutraingredients-usa.com/Suppliers2/Algal-omega-3-market-heats-up-as-new-players-bid-for-a-slice-of-the-action>

Weylandt, K. H., Chiu, C.-Y., Gomolka, B., Waechter, S. F., & Wiedenmann, B. (2012). Omega-3 fatty acids and their lipid mediators: Towards an understanding of resolvin and protectin formation. *Prostaglandins & Other Lipid Mediators*, 97(3–4), 73–82. <https://doi.org/10.1016/j.prostaglandins.2012.01.005>

World Health Organization. (2017). *Depression and Other Common Mental Disorders: Global Health Estimates*.

Wu, W. H., Lu, S. C., Wang, T. F., Jou, H. J., & Wang, T. a. (2006). Effects of docosahexaenoic acid supplementation on blood lipids, estrogen metabolism, and in vivo oxidative stress in postmenopausal vegetarian women. *European Journal of Clinical Nutrition*, 60(3), 386–392. <https://doi.org/10.1038/sj.ejcn.1602328>

Xiao, L., Mjøs, S. A., & Haugsgjerd, B. O. (2012). Efficiencies of three common lipid extraction methods evaluated by calculating mass balances of the fatty acids. *Journal of Food Composition and Analysis*, 25(2), 198–207. <https://doi.org/10.1016/j.jfca.2011.08.003>

Yang, B., Chen, H., Stanton, C., Ross, R. P., Zhang, H., Chen, Y. Q., & Chen, W. (2015). Review of the roles of conjugated linoleic acid in health and disease. *Journal of Functional Foods*, 15, 314–325. <https://doi.org/10.1016/j.jff.2015.03.050>

Yang, J., Xu, M., Zhang, X., Hu, Q., Sommerfeld, M., & Chen, Y. (2011). Life-cycle analysis on biodiesel production from microalgae: Water footprint and nutrients balance. *Bioresource Technology*, 102(1), 159–165. <https://doi.org/10.1016/j.biortech.2010.07.017>

Yen, H., Yang, S., Chen, C., Jesisca, & Chang, J. (2015). Supercritical fluid extraction of valuable compounds from microalgal biomass. *Bioresource Technology*, 184, 291–296. <https://doi.org/10.1016/j.biortech.2014.10.030>

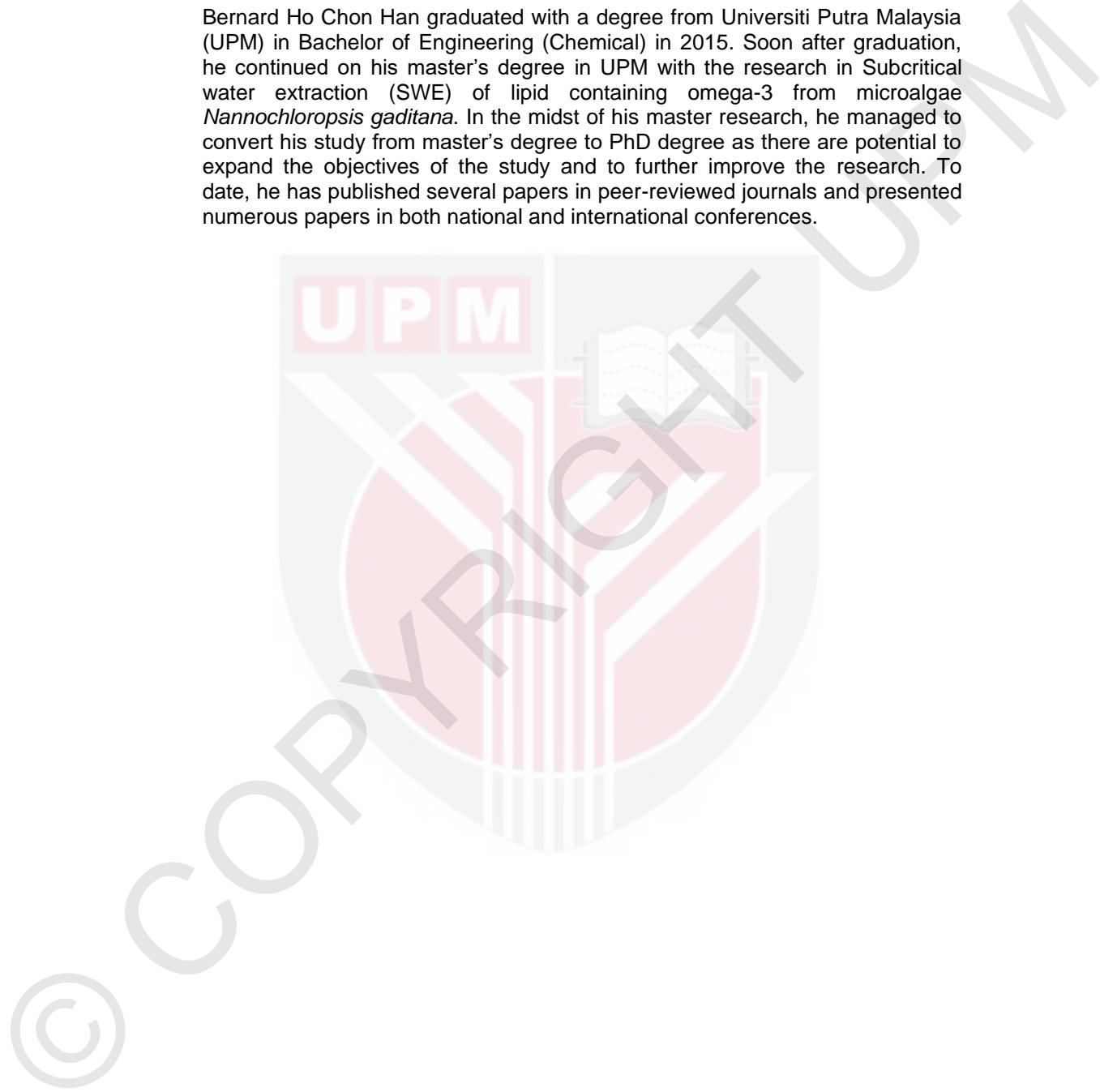
Yoo, G., Park, M. S., Yang, J., & Choi, M. (2015). Lipid content in microalgae determines the quality of biocrude and Energy Return On Investment of hydrothermal liquefaction. *Applied Energy*, 156, 354–361. <https://doi.org/10.1016/j.apenergy.2015.07.020>

Yueming, L., Jianchun, X., Xiuluan, X., Xia, L., Daohui, W., Huibin, S., & Shuijian, Y. (2016). Method for extracting EPA from marine microalgae

- (Patent No. CN105646189A). In *State Intellectual Property of the P.R.C* (No. CN105646189A).
- Zakaria, S. M., & Kamal, S. M. M. (2016). Subcritical water extraction of bioactive compounds from plants and algae: Applications in pharmaceutical and food ingredients. *Food Engineering Reviews*, 8(1), 23–34. <https://doi.org/10.1007/s12393-015-9119-x>
- Zanqui, A. B., De Morais, D. R., Da Silva, C. M., Santos, J. M., Gomes, S. T. M., Visentainer, J. V., Eberlin, M. N., Cardozo-Filho, L., & Matsushita, M. (2015). Subcritical extraction of flaxseed oil with n-propane: Composition and purity. *Food Chemistry*, 188, 452–458. <https://doi.org/10.1016/j.foodchem.2015.05.033>
- Zhang, Y., Ward, V., Dennis, D., Plechkova, N. V., Armenta, R., & Rehmann, L. (2018). Efficient extraction of a docosahexaenoic acid (DHA)-rich lipid fraction from *Thraustochytrium* sp. using ionic liquids. *Materials*, 11(10), 1–11. <https://doi.org/10.3390/ma11101986>
- Zhou, W., Min, M., Li, Y., Hu, B., Ma, X., Cheng, Y., Liu, Y., Chen, P., & Ruan, R. (2012). A hetero-photoautotrophic two-stage cultivation process to improve wastewater nutrient removal and enhance algal lipid accumulation. *Bioresource Technology*, 110, 448–455. <https://doi.org/10.1016/j.biortech.2012.01.063>

## **BIODATA OF STUDENT**

Bernard Ho Chon Han graduated with a degree from Universiti Putra Malaysia (UPM) in Bachelor of Engineering (Chemical) in 2015. Soon after graduation, he continued on his master's degree in UPM with the research in Subcritical water extraction (SWE) of lipid containing omega-3 from microalgae *Nannochloropsis gaditana*. In the midst of his master research, he managed to convert his study from master's degree to PhD degree as there are potential to expand the objectives of the study and to further improve the research. To date, he has published several papers in peer-reviewed journals and presented numerous papers in both national and international conferences.



## LIST OF PUBLICATIONS

### Journal

- Ho, B. C. H.**, & Harun, R. (2017). Extraction of bioactive compounds from *Nannochloropsis gaditana* via sub-critical water extraction (SWE). *FEII International Journal of Engineering and Technology*, 14(1), 19–24.
- Ho, B. C. H.**, Kamal, S. M. M., & Harun, M. R. (2018). Extraction of eicosapentaenoic acid from *Nannochloropsis gaditana* using sub-critical water extraction. *Malaysian Journal of Analytical Sciences*, 22(4), 619–625. <https://doi.org/10.17576/mjas-2018-2204-07>
- Ho, B. C. H.**, Kamal, S. M. M., Danquah, M. K., & Harun, R. (2018). Optimization of subcritical water extraction (SWE) of lipid and eicosapentaenoic acid (EPA) from *Nannochloropsis gaditana*. *BioMed Research International*, 2018. <https://doi.org/10.1155/2018/8273581>

### Proceedings

- Ho, B. C. H.** and Harun, R. (2016) *Extraction of bioactive compounds from Nannochloropsis gaditana via sub-critical water extraction (SWE)*. In: World Research & Innovation Convention on Engineering & Technology WRICET2016, 24-25 October 2016, Langkawi, Kedah, Malaysia.
- Ho, B. C. H.** and Harun, R. (2017) *Extraction of eicosapentaenoic acid (EPA) from Nannochloropsis gaditana using sub-critical water extraction (SWE) method*. In: The 2<sup>nd</sup> International Conference on Separation Technology 2017 (ICoST2017), 15-16 April 2017, Centre of Lipids Engineering and Applied Research (CLEAR), Universiti Teknologi Malaysia. (p. F45-F47)
- Ho, B. C. H.** and Harun, R. (2017) *Recovery of aqueous phase of sub-critical water extraction (SWE) from Nannochloropsis gaditana*. In: 5th International Symposium on Applied Engineering and Sciences (SAES2017), 14-15 November 2017, Universiti Putra Malaysia. (p. 23).



## UNIVERSITI PUTRA MALAYSIA

### STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : Second Semester 2020/2021

#### TITLE OF THESIS / PROJECT REPORT :

SUBCRITICAL WATER EXTRACTION OF LIPID CONTAINING OMEGA-3 FROM  
MICROALGAE *Nannochloropsis gaditana*

NAME OF STUDENT: BERNARD HO CHON HAN

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (v )

**CONFIDENTIAL**

(Contain confidential information under Official Secret Act 1972).

**RESTRICTED**

(Contains restricted information as specified by the organization/institution where research was done).

**OPEN ACCESS**

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

**PATENT**

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

**Approved by:**

(Signature of Student)  
New IC No/ Passport No.:

(Signature of Chairman of Supervisory Committee)  
Name:

Date :

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentiality or restricted.]