



UNIVERSITI PUTRA MALAYSIA

***IN VITRO AND IN VIVO EFFECT OF *Clinacanthus nutans* (BURM. F.)
LINDAU AQUEOUS EXTRACT ON IgE AND IgG-MEDIATED
ALLERGY PATHWAYS***

AUDREY KOW SIEW FOONG

FPSK(m) 2021 30



***IN VITRO AND IN VIVO* EFFECT OF *Clinacanthus nutans* (BURM. F.)
LINDAU AQUEOUS EXTRACT ON IgE AND IgG-MEDIATED ALLERGY
PATHWAYS**

By

AUDREY KOW SIEW FOONG

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of
Philosophy**

June 2021

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

IN VITRO AND IN VIVO EFFECT OF *Clinacanthus nutans* (BURM. F.) LINDAU AQUEOUS EXTRACT ON IgE AND IgG-MEDIATED ALLERGY PATHWAYS

By

AUDREY KOW SIEW FOONG

June 2021

Chair : Tham Chau Ling, PhD
Faculty : Medicine and Health Sciences

Allergy is a hypersensitive reaction against antigens which could be mediated through immunoglobulin (Ig) -E and IgG. About 30 – 40% of people globally are affected with allergy and it is projected to increase due to urbanisation. It has both personal and economic implications thus, requiring urgent attention. *Clinacanthus nutans* (Burm. f.) Lindau (*C. nutans*) is commonly found in Malaysia, Thailand and Indonesia. Traditionally used to treat snake and insect bites, skin rashes and others, it was evaluated for its anti-viral, anti-inflammatory and anti-cancer properties. Although used to treat skin rashes, its anti-allergy property was not evaluated. Hence, this study evaluated the anti-allergy property of *C. nutans* in *in vitro* and *in vivo* allergy models and its underlying mechanism. Meta-analysis was done to identify the commonly analysed soluble mediators in the less established IgG pathway (compared to IgE). The anti-allergy property via IgE pathway of 100% ethanolic, 70% ethanolic-aqueous, 50% ethanolic-aqueous and aqueous *C. nutans* extracts was assessed through *in vitro* IgE-induced mast cell degranulation model. The most active extract was then evaluated in IgG-induced macrophage activation model. The underlying mechanism was studied by analysing its effect on the mitogen-activated protein kinase (MAPK) and phosphoinositide-3-kinase (PI3K) pathways' proteins by Western blotting. The effect was then validated in rodents. Acute toxicity test that analysed the haematological, biochemical and histological profiles was done to determine its safety and safe doses to be used. The overall effect was analysed in ovalbumin-challenged active systemic anaphylaxis (OVA-ASA), whereby both IgE and IgG pathways were activated. The effect of CNAE on specific targeted pathway was analysed in IgE-

challenged passive systemic anaphylaxis (IgE-PSA) and IgG-PSA models. Soluble mediators were quantified by enzyme-linked immunosorbent assay (ELISA). The commonly studied soluble mediators of IgG pathway identified through meta-analysis were platelet activating factor (PAF), histamine, interleukins (IL)-6, -13 and tumour necrosis factor- α (TNF- α). The most active extract - CNAE significantly reduced histamine and β -hexosaminidase in IgE-induced mast cell degranulation model at 5 mg/mL and above. In the IgG-induced macrophage activation model, significant decrease of IL-6 and TNF- α were recorded at 1.75 mg/mL and this was due to the inhibition of the phosphorylation of ERK1/2 of the MAPK pathway. From the acute toxicity test, 5000 mg/kg of CNAE (single dose) was not toxic to the animals and the doses - 125, 500 and 2000 mg/kg were chosen for subsequent analyses. In OVA-ASA, CNAE (2000 mg/kg) inhibited IgG (89.5%), PAF (171.1%) and IL-6 (92.6%) but not IgE. There was no significant inhibition of histamine, IL-4 and leukotriene C₄ (LTC₄) in IgE-PSA even at 2000 mg/kg. However, at 2000 mg/kg, there was 128.2% reduction of PAF and 124.4% reduction of IL-6 in IgG-PSA. Significant reduction of histamine in *in vitro* IgE-induced mast cell degranulation that was not recorded in IgE-PSA could be due to the overall effect of different cells that were activated in IgE-PSA. In the *in vitro* model, Rat Basophilic Leukaemic (RBL-2H3) cells which mimicked mast cells were specifically induced while in IgE-PSA, other cells with high-affinity IgE receptor (Fc ϵ RI) such as mast cells, eosinophils and basophils were challenged giving an overall effect. In conclusion, the anti-allergy property of *C. nutans* was potentially in CNAE which targeted the IgG pathway with significant reductions of PAF, IL-6 and TNF- α by inhibiting ERK1/2 pathway. As anaphylaxis is a systemic allergy, the anti-allergy effect of CNAE could be further evaluated on localised models such as atopic eczema or allergic alveolitis.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN *IN VITRO* DAN *IN VIVO* EKSTRAK AKUEUS *Clinacanthus nutans*
(BURM. F.) LINDAU PADA LALUAN ALERGI YANG DIMEDIASI OLEH IgE
DAN IgG**

Oleh

AUDREY KOW SIEW FOONG

Jun 2021

Pengerusi : Tham Chau Ling, PhD
Fakulti : Perubatan dan Sains Kesihatan

Alergi adalah tindak balas hipersensitif terhadap antigen. Secara imunologi, alergi boleh dimediasi melalui imunoglobulin (Ig) -E dan IgG. Sebanyak 30-40% populasi dunia terjejas dengan alergi dan ini dianggarkan akan meningkat disebabkan oleh pembangunan. Alergi membawa impak negatif pada seseorang individu dan juga ekonomi negara. Oleh itu, ia memerlukan perhatian segera. *Clinacanthus nutans* (Burm. f.) Lindau (*C. nutans*) atau Belalai Gajah biasanya dijumpai di Malaysia, Thailand dan Indonesia. Secara tradisionalnya, ia digunakan untuk merawat gigitan ular dan serangga, ruam kulit dan lain-lain. Beberapa kajian saintifik telah membuktikan ia bersifat antivirus; antiradang dan antibarah. Walau bagaimanapun, sifat antialerginya belum dikaji walaupun ia digunakan untuk merawat ruam kulit. Oleh itu, kajian ini menilai sifat antialergi *C. nutans* dalam model alergi *in vitro* dan *in vivo* dan mekanismenya. Metaanalisis dilakukan untuk mengenal pasti mediator yang paling kerap dianalisis bagi alergi yang dimediasi oleh IgG; dimana ia kurang dikaji berbanding dengan laluan IgE. Sifat antialergi ekstrak 100% etanol (100% EtOH), 70% etanol-akueus (70% EtOH-Ak), 50% etanol-akueus (EtOH-Ak) dan ekstrak akueus (CNAE) *C. nutans* pertamanya dinilai menggunakan model degranulasi sel mast yang diinduksi oleh IgE *in vitro*. Kesan ekstrak paling aktif - CNAE kemudian dinilai dalam model pengaktifan makrofag yang disebabkan oleh IgG. Mekanisme antialergi CNAE kemudiannya dikaji dengan menganalisis kesannya pada laluan 'mitogen-activated protein kinase' (MAPK) dan fosfoinositida-3-kinase (PI3K) menggunakan kaedah 'Western blotting'. Seterusnya, kesan antialergi CNAE dinilai pada tikus. Kajian ketoksikan akut dilakukan untuk menilai keselamatannya dengan menilai profil hematologi,

biokimia dan histologi dan untuk menentukan dos yang selamat untuk analisis seterusnya. Kesan umum antialergi CNAE kemudian dianalisis dalam anafilaksis sistemik aktif yang dicabar oleh ovalbumin (OVA-ASA) di mana kedua-dua laluan IgE dan IgG diaktifkan. Untuk mengenal pasti laluan sasaran tertentu, kesan antialergi CNAE dianalisis lebih lanjut dalam model anafilaksis pasif IgE-PSA dan IgG-PSA. Mediator yang dihasilkan dinilai dengan pemeriksaan imunisorben berkait enzim (ELISA). Dari kajian metaanalisis, mediator laluan IgG yang paling kerap dikaji adalah faktor pengaktifan platelet (PAF), histamin, interleukin (IL) -6, -13 dan faktor nekrosis tumor- α (TNF- α). Ekstrak paling aktif - CNAE dapat mengurangkan tahap histamin dan β -heksosaminidase secara signifikan pada model degranulasi sel mast yang disebabkan oleh IgE pada kepekatan 5 mg/mL dan ke atas. Sebaliknya, dalam model pengaktifan makrofag yang disebabkan oleh IgG, penurunan IL-6 dan TNF- α yang ketara dicatatkan pada kepekatan 1.75 mg/mL. Pengurangan IL-6 dan TNF- α adalah hasil penghambatan fosforilasi ERK dalam laluan MAPK. Kajian ketoksikan akut menunjukkan bahawa satu dos CNAE pada 5000 mg/kg tidak toksik kepada haiwan. Dalam OVA-ASA, CNAE pada 2000 mg/kg mencatatkan perencatan IgG (89.5%), PAF (171.1%) dan IL-6 (92.6%) tanpa pengurangan tahap IgE. Tidak ada penurunan histamin, IL-4 dan leukotriena C₄ (LTC₄) direkodkan dalam IgE-PSA walaupun pada 2000 mg/kg. Walau bagaimanapun, pada 2000 mg/kg, terdapat penurunan PAF sebanyak 128.2% dan penurunan IL-6 (124.4%) dalam IgG-PSA. Penurunan histamin pada model degranulasi sel mast yang disebabkan oleh IgE tidak direkodkan dalam IgE-PSA berkemungkinan disebabkan perbezaan sel yang diaktifkan dalam kedua-dua model ini. Dalam model *in vitro*, sel basofilik yang menyerupai sel mast diaktifkan sebaliknya sel lain seperti sel mast, sel basofilik dan macrofaj juga diaktifkan dalam model *in vivo*. Secara keseluruhannya, sifat antialergi *C. nutans* berpotensi dijumpai dalam CNAE dan menasarkkan laluan alergi IgG dengan penurunan PAF, IL-6 dan TNF- α yang ketara yang menasarkkan laluan ERK1/2. Anafilaksis adalah model alergi sistemik, oleh itu sifat antialergi CNAE boleh dikaji ke atas model alergi setempat seperti ekzema atopi atau alergi alveolus.

ACKNOWLEDGEMENTS

All Glory to God, without Him this journey will not be possible. First of all, I would like to thank the Ministry of Agriculture Malaysia for funding this project under the NKEA Herbs Research Grant scheme (NRGS) NH1014D067. I would also like to thank my supervisory committee headed by Associate Professor Dr. Tham Chau Ling, Professor Dr. Faridah Abas, Professor Dr. Daud Ahmad and Associate Professor Dr. Lee Ming-Tatt for their guidance and support throughout this study. I would also like to thank Dr. Wan Mastura Shaik Mohamed Mossadeq, my very helpful attending veterinarian from the Department of Veterinary Preclinical Sciences, Faculty of Veterinary Medicine, Universiti Putra Malaysia; Dr. Fred D. Finkelman, Dr. John J. Ryan and Yves T. Falanga for their kind replies to my queries, and Professor Dr. Mohd Roslan Sulaiman and Dr. Enoch Kumar Perimal for allowing me to use the PowerLab program and equipment. I would also like to extend my gratitude to Dr. Khoo Leng Wei, Dr. Fong Lai Yen, Dr. Tan Ji Wei, Mr. Ali, Encik Ramli Suhaimi, Miss Heng Kai Yen, Miss Sarah Sapuan, Dr. Ong Hui Ming, Mr. Mohd. Azim Fikri Bin Abd Ghani and Miss Azirah Chik for their technical support towards this project.

Lastly, this journey has been enriched with the help and support from laboratory staffs of Cell Signalling Laboratory and Histopathology Laboratory - Encik Zulkhairi Bin Zainol, Puan Nora Asyikin Binti Mohd Salim, Puan Juita Binti Chupri, Encik Abdul Rahman Hassan and fellow peers and my family.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Tham Chau Ling, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Faridah binti Abas, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Daud Ahmad bin Israf Ali, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Lee Ming-Tatt, PhD

Associate Professor
Faculty of Pharmaceutical Sciences
UCSI University
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 11 November 2021

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of Supervisory Committee: Associate Professor Dr. Tham Chau Ling

Signature: _____

Name of Member of Supervisory Committee: Professor Dr. Faridah binti Abas

Signature: _____

Name of Member of Supervisory Committee: Professor Dr. Daud Ahmad bin Israf Ali

Signature: _____

Name of Member of Supervisory Committee: Associate Professor Dr. Lee Ming-Tatt

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF APPENDICES	xxii
LIST OF ABBREVIATIONS	xxiii
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
1.2 Problem Statement	2
1.3 Direction of Study	2
1.4 Research Objectives	4
1.4.1 General Objective	4
1.4.2 Specific Objectives	4
1.5 Hypotheses	4
2 LITERATURE REVIEW	5
2.1 Allergy and Anaphylaxis	5
2.1.1 Allergy	5
2.2.1 Anaphylaxis	7
2.2 The Use of Traditional Herbal Medicines	17
2.3 <i>Clinacanthus nutans</i> (Burm. f.) Lindau	19
2.3.1 Botany	19
2.3.2 Toxicity	20
2.3.3 Ethnomedicinal Uses	21
2.3.4 Pharmacological Activities	21
2.4 Meta-Analysis	32
2.5 Study Overview	35
3 IDENTIFICATION OF SOLUBLE MEDIATORS IN IGG-MEDIATED ANAPHYLAXIS IN RODENTS BY META- ANALYSIS	36
3.1 Introduction	36
3.2 Objective	37
3.3 Methods	37

3.3.1	Search Strategy	37
3.3.2	Eligibility Criteria	37
3.3.3	Study Appraisal and Selection	37
3.3.4	Data Extraction and Organisation of Studies	38
3.3.5	Data Analysis	38
3.3.6	Study Outcome	38
3.4	Results	39
3.4.1	Literature Search	39
3.4.2	Data Extraction, Organisation and Meta-analysis	40
3.5	Discussion	53
3.6	Conclusion	57
4	EVALUATION OF THE ANTI-ALLERGY PROPERTY OF <i>CLINACANTHUS NUTANS</i> EXTRACT BY <i>IN VITRO</i> IGE-INDUCED MAST CELL DEGRANULATION AND IGG- INDUCED MACROPHAGE ACTIVATION	58
4.1	Introduction	58
4.2	Objectives	59
4.3	Materials	60
4.4	Methods	61
4.4.1	Preparation of <i>Clinacanthus nutans</i> (Burm. f.) Lindau (<i>C. nutans</i>) Extracts	61
4.4.2	Evaluation of the Anti-allergy Property of <i>C. nutans</i> Extracts by In Vitro IgE-induced Mast Cell Degranulation Model	62
4.4.3	Evaluation of the Anti-allergy Property of 100% Aqueous <i>C. nutans</i> Extract (CNAE) by In Vitro IgG-induced Macrophage Activation Model	64
4.4.4	Evaluation of the Anti-allergy Mode of Action of 100% Aqueous <i>C. nutans</i> Extract (CNAE) by Western Blot	66
4.4.5	Statistical Analysis	69

4.5	Results	69
4.5.1	Evaluation of the Anti-allergy Property of <i>C. nutans</i> Extracts by In Vitro IgE-induced Mast Cell Degranulation Model	69
4.5.2	Evaluation of the Anti-allergy Property of 100% Aqueous <i>C. nutans</i> Extract (CNAE) by In Vitro IgG-induced Macrophage Activation Model	83
4.5.3	Evaluation of the Anti-allergy Mode of Action of 100% Aqueous <i>C. nutans</i> Extract (CNAE) by Western Blot	85
4.6	Discussion	87
4.7	Conclusion	92
5	ACUTE TOXICITY OF 100% AQUEOUS <i>CLINACANTHUS NUTANS</i> EXTRACT (CNAE) ON FEMALE SPRAGUE-DAWLEY RATS	93
5.1	Introduction	93
5.2	Objective	94
5.3	Materials	94
5.3.1	Materials	94
5.3.2	Animals	95
5.4	Methods	95
5.4.1	Acute Oral Toxicity Study	95
5.4.2	Cage Side Observation	95
5.4.3	Body and Organs Weight Analysis	95
5.4.4	Food and Water Consumption	96
5.4.5	Haematology and Biochemical Analysis	96
5.4.6	Metabolomics Evaluation Using Advanced Proton Nuclear Magnetic Resonance (¹ H-NMR)	96
5.4.7	Histopathological Analysis by Haematoxylin and Eosin (H and E) Staining	97
5.5	Results	97
5.5.1	Physical and Behavioural Changes Observation	97

	5.5.2	Haematological and Biochemical Analysis	100
	5.5.3	Histopathological Analysis	102
	5.6	Discussion	108
	5.7	Conclusion	111
6		EVALUATION OF THE ANTI-ALLERGY PROPERTY OF 100% AQUEOUS <i>CLINACANTHUS NUTANS</i> EXTRACT (CNAE) AND IDENTIFICATION OF THE TARGETED PATHWAY IN <i>IN VIVO</i> MODELS OF SYSTEMIC ANAPHYLAXIS	112
	6.1	Introduction	112
	6.2	Objectives	113
	6.3	Materials	113
	6.3.1	Materials	113
	6.3.2	Animals	114
	6.4	Methods	114
	6.4.1	Ovalbumin-challenged Active Systemic Anaphylaxis (OVA-ASA)	115
	6.4.2	IgE-challenged Passive Systemic Anaphylaxis (IgE-PSA)	117
	6.4.3	IgG-challenged Passive Systemic Anaphylaxis (IgG-PSA)	119
	6.4.4	Statistical Analysis	121
	6.5	Results	121
	6.5.1	Effect of 100% Aqueous <i>C. nutans</i> Extract (CNAE) on OVA-ASA	121
	6.5.2	Effect of 100% Aqueous <i>C. nutans</i> Extract (CNAE) on IgE-PSA	126
	6.5.3	Effect of 100% Aqueous <i>C. nutans</i> Extract (CNAE) on IgG-PSA	130
	6.6	Discussion	132
	6.7	Conclusion	135
7		SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	136
	7.1	Summary	136
	7.2	Conclusion	137

7.3	Limitations	138
7.4	Future Research	139
REFERENCES		141
APPENDICES		163
BIODATA OF STUDENT		169
LIST OF PUBLICATIONS		170



LIST OF TABLES

Table		Page
1	Taxonomic classification and nomenclature of <i>Clinacanthus nutans</i> (Burm. f.) Lindau	19
2	Categorisation of all relevant articles based on the study model used and publication year	40
3	List of <i>in vivo</i> studies categorised based on the type of mediator(s) studied, inducer used	41
4	List of <i>ex vivo/in vitro</i> studies categorised based on the type of mediator(s) studied, inducer used	44
5	Compilation of the cumulative mean difference, <i>p-value</i> and 95% confidence level (CI) of each mediator studied in both <i>in vivo</i> and <i>ex vivo/in vitro</i> studies of IgG-mediated anaphylaxis between non-anaphylactic and anaphylactic groups	50
6	Summary and conclusion of publication bias assessment from asymmetry of funnel plot, Egger's regression test, Begg's test, Rosenthal's Fail-safe N and Trim and Fill (unbiased estimate) analysis	52
7	Experimental grouping for IgE-induced mast cell degranulation assay	64
8	Experimental grouping for IgG-induced macrophage activation assay	66
9	Haematology analysis on the acute oral toxicity effect of a single dose of 100% aqueous <i>Clinacanthus nutans</i> extract (CNAE) (5000 mg/kg) in Sprague Dawley rats	100
10	Biochemical analysis on the acute oral toxicity effect of a single dose of 100% aqueous <i>Clinacanthus nutans</i> extract (CNAE) (5000 mg/kg) in Sprague Dawley rats	101
11	Experimental grouping for ovalbumin-induced active systemic anaphylaxis (OVA-ASA) model	116

12	Experimental grouping for IgE-induced passive systemic anaphylaxis (IgE-PSA) model	118
13	Experimental grouping for IgG-induced passive systemic anaphylaxis (IgG-PSA) model	120
14	Anaphylaxis scoring criteria	166



LIST OF FIGURES

Figure		Page
1	Mechanism of IgE-mediated (left) and IgG-mediated (right) anaphylaxis	9
2	Schematic diagram depicting the events that happens after aggregation of antigen bound immunoglobulins to Fc receptors	11
3	Schematic diagram depicting the activation of PI3K pathway	13
4	Schematic diagram depicting the activation of ERK pathway	14
5	Schematic diagram depicting the activation of JNK pathway	15
6	Schematic diagram depicting the activation of p38 pathway	16
7	<i>Clinacanthus nutans</i> plant	20
8	An example of a forest plot of meta-analysis	33
9	Examples of A) A symmetrical funnel plot and B) An asymmetrical funnel plot of publication bias assessment	34
10	Study retrieval and selection criteria	39
11	Forest plot of histamine from <i>in vivo</i> studies	43
12	Forest plots of A) β -hexosaminidase, B) PAF, C) MIP-1 α , D) IL-6, E) IL-13 and F) TNF- α of <i>ex vivo/in vitro</i> studies	48
13	Funnel plot assessment of publication bias based on the effect size and standard error	51
14	Cytotoxicity profiles of <i>C. nutans</i> 100% ethanolic, 70% and 50% ethanolic-aqueous and 100% aqueous extracts by MTT assay	70

15	The effect of 100% ethanolic <i>C. nutans</i> extract on the percentage of β -hexosaminidase released (%) by RBL-2H3 cells	71
16	The effect of 70% ethanolic-aqueous <i>C. nutans</i> extract on the percentage of β -hexosaminidase released (%) by RBL-2H3 cells	72
17	The effect of 50% ethanolic-aqueous <i>C. nutans</i> extract on the percentage of β -hexosaminidase released (%) by RBL-2H3 cells	72
18	The effect of 100% aqueous <i>C. nutans</i> extract on the percentage of β -hexosaminidase released (%) by RBL-2H3 cells	73
19	The effect of 100% ethanolic <i>C. nutans</i> extract on the release of histamine (nM) by RBL-2H3 cells	74
20	The effect of 70% ethanolic-aqueous <i>C. nutans</i> extract on the release of histamine (nM) by RBL-2H3 cells	75
21	The effect of 50% ethanolic-aqueous <i>C. nutans</i> extract on the release of histamine (nM) by RBL-2H3 cells	75
22	The effect of 100% aqueous <i>C. nutans</i> extract on the release of histamine (nM) by RBL-2H3 cells	76
23	The effect of 100% ethanolic <i>C. nutans</i> extract on the release of IL-4 (pg/mL) by RBL-2H3 cells	77
24	The effect of 70% ethanolic-aqueous <i>C. nutans</i> extract on the release of IL-4 (pg/mL) by RBL-2H3 cells	78
25	The effect of 50% ethanolic-aqueous <i>C. nutans</i> extract on the release of IL-4 (pg/mL) by RBL-2H3 cells	78
26	The effect of 100% aqueous <i>C. nutans</i> extract on the release of IL-4 (pg/mL) by RBL-2H3 cells	79
27	The effect of 100% ethanolic <i>C. nutans</i> extract on the release of TNF- α (pg/mL) by RBL-2H3 cells	80

28	The effect of 70% ethanolic-aqueous and <i>C. nutans</i> extract on the release of TNF- α (pg/mL) by RBL-2H3 cells	81
29	The effect of 50% ethanolic-aqueous <i>C. nutans</i> extract on the release of TNF- α (pg/mL) by RBL-2H3 cells	81
30	The effect of 100% aqueous <i>C. nutans</i> extract on the release of TNF- α (pg/mL) by RBL-2H3 cells	82
31	Cytotoxicity analysis of 100% aqueous <i>C. nutans</i> extract (CNAE) on IC-21 cells by MTS assay	83
32	Effect of 100% aqueous <i>C. nutans</i> extract (CNAE) on the release of IL-6 (pg/mL) upon IgG-induced macrophage activation on IC-21 cells	84
33	Effect of 100% aqueous <i>C. nutans</i> extract (CNAE) on the release of TNF- α (pg/mL) upon IgG-induced macrophage activation on IC-21 cells	85
34	Effect of 100% aqueous <i>C. nutans</i> extract (CNAE) on ERK upon IgG-induced macrophage activation of IC-21 cells	86
35	Effect of 100% aqueous <i>C. nutans</i> extract (CNAE) on the p38 upon IgG-induced macrophage activation of IC-21 cells	87
36	Average food consumption of non-treated rats and rats fed with single dose of 5000 mg/kg of CNAE on Days 1, 6 and 10	98
37	Average water intakes of non-treated rats and rats fed with single dose of 5000 mg/kg of CNAE on Days 1, 6 and 10	98
38	Average body weight differences of non-treated rats and rats fed with single dose of 5000 mg/kg of CNAE after 14 days	99
39	Relative organ weight to body weight ratio of liver, spleen, kidneys, heart and lungs - of non-treated rats and rats fed with single dose of 5000 mg/kg of CNAE	102

40	Representative photographs of histological sections of liver tissues of rats fed with distilled water (left) and single dose of CNAE (5000 mg/kg) (right) (H & E X100 magnification; Inset: X400 magnification)	103
41	Representative photographs of histological sections of kidney tissues of rats fed with distilled water (left) and single dose of CNAE (5000 mg/kg) (right) (H & E X100 magnification; Inset: X400 magnification)	104
42	Representative photographs of histological sections of heart tissues of rats fed with distilled water (left) and single dose of CNAE (5000 mg/kg) (right) (H & E X100 magnification; Inset: X400 magnification)	105
43	Representative photographs of histological sections of lung tissues of rats fed with distilled water (left) and single dose of CNAE (5000 mg/kg) (right) (H & E X100 magnification; Inset: X400 magnification)	106
44	Representative photographs of histological sections of spleen tissues of rats fed with distilled water (left) and single dose of CNAE (5000 mg/kg) (right) (H & E X100 magnification; Inset: X400 magnification)	107
45	General overview of the evaluation of the anti-allergy effect of CNAE in a biological system	114
46	Graphical timeline of OVA-ASA model	116
47	Graphical timeline of IgE-PSA model	118
48	Graphical timeline of IgG-PSA model	120
49	Amount of IgE released (ng/mL) by Sprague-Dawley rats in OVA-ASA model	122
50	Amount of IgG released (% of control) by Sprague-Dawley rats in OVA-ASA model	123

51	Amount of PAF released (% of control) by Sprague-Dawley rats in OVA-ASA model	124
52	Amount of IL-6 released (% of control) by Sprague-Dawley rats in OVA-ASA model	125
53	Amount of histamine released (nM) by Sprague-Dawley rats in IgE-PSA model	127
54	Amount of IL-4 released (pg/mL) by Sprague-Dawley rats in IgE-PSA model	128
55	Amount of LTC ₄ released (pg/mL) by Sprague-Dawley rats in IgE-PSA model	129
56	Amount of PAF released (ng/mL) by C57BL/6 mice in IgG-PSA model	131
57	Amount of IL-6 released (pg/mL) by C57BL/6 mice in IgG-PSA model	132
58	Summary of all findings	136
59	Induction time optimisation for ERK kinase	163
60	Induction time optimisation for p38 kinase	164
61	Rectal temperature change of C57BL/6 mice in IgG-PSA model	165
62	Anaphylaxis symptom score of C57BL/6 mice in IgG-PSA model	167

LIST OF APPENDICES

Appendix		Page
1	Induction time optimisation for ERK1/2 and p38 protein kinases	163
2	Drop in rectal temperature of C57BL/6 mice in IgG-induced passive systemic anaphylaxis model	165
3	Anaphylaxis symptoms scoring of C57BL/6 mice in IgG-induced passive systemic anaphylaxis model	166
4	IACUC Approval	168

LIST OF ABBREVIATIONS

1H-NMR	Proton nuclear magnetic resonance
AAPH	2,2'-azobis (2-amidinopropane) dihydrochloride
ANOVA	One-way analysis of variance
Btk	Bruton's tyrosine kinase
Ca ²⁺	Calcium
<i>C. nutans</i>	<i>Clinacanthus nutans</i>
CEFs	Chick embryo fibroblasts
cGMP	Cyclic guanosine monophosphate
CNAE	100% aqueous <i>Clinacanthus nutans</i>
cPLA ₂	Cytoplasmic phospholipase A ₂
DAG	Diacylglycerol
DGDG	Digalactosyl diglyceride
DNA	Deoxyribonucleic acid
DNP-BSA	2,4-dinitrophenol bovine serum albumin
DPPH	1,1-diphenyl-2-picrylhydrazyl
DV2	Dengue virus serotype 2
ELISA	Enzyme-linked immunosorbent assay
EtOH	Ethanol
EtOH-Aq	Ethanol-aqueous
EPP	Ethyl phenylpropionate
ERK	Extracellular signal-regulated kinases
FcεRI	High-affinity IgE receptor

FcγR	Low-affinity receptor
FRAP	Ferric reducing antioxidant power
fMLP/CB	N-formyl-L-methionyl-L-leucyl-L-phenylalanine/cytochalasin B
GAB2	GRB2-associated-binding protein 2
GDP	Guanosine diphosphate
GRB2	Growth factor receptor-bound protein 2
GTP	Guanosine triphosphate
HSV	Herpes Simplex virus
IC50	Half maximal inhibitory concentration
IFN	Interferon
Ig	Immunoglobulin
IgE-PSA	Immunoglobulin-E-challenged passive systemic anaphylaxis
IgG-PSA	Immunoglobulin-G-challenged passive systemic anaphylaxis
ITAMs	Immunoreceptor tyrosine-based activation motifs
IL	Interleukin
JNK	c-Jun N-terminal kinase
LAT	Linker for activation of T cells
LTC4	Leukotriene C4
MAPK	Mitogen-activated protein kinase
MAPKK	Mitogen-activated protein kinase kinase
MAPKKK	Mitogen-activated protein kinase kinase kinase
MEK	MAP/ERK kinase
MGDG	Monogalactosyl diglyceride
MIP-1α	Macrophage inflammatory protein-1α
MPO	Myeloperoxidase

NO	Nitric oxide
NTAL	Non-T-cell activation linker
OVA	Ovalbumin
OVA-ASA	Ovalbumin-challenged active systemic anaphylaxis
PA	Phosphatidic acid
PAF	Platelet activating factor
PDK1	PI3K-dependent protein kinase 1
PH	Pleckstrin homology
PI3K	Phosphoinositide-3-kinase
PIP3	Phosphatidylinositol 3,4,5-triphosphate
PKC	Protein kinase C
PLC γ	Phospholipase C γ
PLD	Phospholipase D
PLS	Partial least-square
PMA	Phorbol myristate acetate
Ras	Guanine nucleotide-binding protein
RBL-2H3	Rat Basophilic Leukaemic cells
RBP4	Retinol binding protein 4
RPM	Revolutions per minute
SOD	Superoxide dismutase
SOS	Son of Sevenless
TCM	Traditional and complementary medicine
TCR	T cell antigen receptor

TKBM	Traditional knowledge-based medicine
TNF- α	Tumour necrosis factor- α
VEGF	Vascular endothelial growth factor
VZV	Varicella-Zoster virus
WHO	World Health Organisation



CHAPTER 1

INTRODUCTION

1.1 Background

Allergy is a hypersensitivity reaction caused immunologically and it can be presented in many forms such as anaphylaxis, urticaria, asthma, atopic dermatitis and many others (Ring, 2014). The prevalence of allergic diseases has been on the rise in the last decade. It was estimated that globally, 30 – 40% of people will be affected by one form or another of allergy and the most vulnerable group comes from the younger population (Pawankar *et al.*, 2011). The rise of allergy prevalence puts an economic burden to the world and affected individuals usually have decreased quality of life as they experienced limitation to their activity and function (Mendis, 2014). Hence, it is a matter that requires urgent attention.

The use of natural products in the form of plant to address medical needs is a common practice among the locals of a certain country. It is usually a practice that was passed down from previous generations. Historical documents showed that medicinal plants have been in used in the past 60,000 years in Iraq and elsewhere (Pan *et al.*, 2014). Under the wide umbrella of traditional medicine, plant-based medicines have been gaining more attention lately. This is due to the side effects of modern medicine; the limitation of modern medicines in treating the root of the disease; microbial resistance and unparalleled spending on pharmaceutical research and development of modern medicine (Pan *et al.*, 2014). Undeniably there are many beneficial plants that are yet to be discovered in our backyard. However, the use of traditional plant-based medicine is usually shunned upon as it lacks proper scientific evaluation and proper regulation. Furthermore, there is no good documentation that could support the beneficial usage of these plants. Hence, it is of importance to scientifically evaluate and document the traditional medicinal claims of these plants so that they could be use in complement to modern medicine practice for a better outcome.

Clinacanthus nutans Burm. f. (Lindau) (*C. nutans*) is a perennial plant found mainly in Southeast Asia especially in Malaysia, Thailand and Indonesia. Known by the locals by many names – ‘Belalai Gajah’, ‘You Dun Cao’, ‘Payayor’ and ‘Daun Dandang Gendis’ - this shrub was used to treat many conditions in these countries (Khoo *et al.*, 2018b). These include snake and insect bites, skin rashes, eczema, Varicella Zoster lesions, diabetes, haematoma, gastrointestinal problems to name a few (Yahaya *et al.*, 2015; Zulkipli *et al.*, 2017). Based on these traditional claims, numerous pharmacological studies have been conducted such as anti-viral activities against Herpes Simplex virus (HSV), Varicella-Zoster virus (VZV) and dengue; anti-inflammatory activity, anti-venom, anti-nociceptive, anti-bacterial and anti-

fungal, anti-cancer, anti-diabetic, anti-oxidant, immunomodulating property and neuronal protection (Khoo *et al.*, 2018b). *C. nutans* have been used as a traditional remedy for skin rashes - a characteristic of an allergic reaction but it has not been pharmacologically evaluated for its anti-allergy property. Thus, this study was aimed to identify the potential anti-allergy property of this plant. This study will also expand the pharmacological documentation of *C. nutans* in the Malaysian Herbal Monograph (MHM) and it could also be beneficial to consumers and the economy of one country.

1.2 Problem Statement

The number of allergy cases worldwide is projected to rise as a result of urbanisation and the burden of allergy is not limited to only the affected individual. It is thus a matter of concern which requires urgent attention. *C. nutans* is a perennial shrub that has been used traditionally among locals of Southeast Asian countries to treat skin rashes, a symptom of allergy. However, there are no pharmacological studies thus far that has evaluated the anti-allergy property of this plant. Hence, in this study the anti-allergy property of *C. nutans* was evaluated both *in vitro* using cells and *in vivo* using rodents. Additionally, the mode of action was also studied.

1.3 Direction of Study

In this study, the anti-allergy property of *C. nutans* was analysed in both *in vitro* and *in vivo* models of allergy. Allergy can be mediated through two pathways known as the IgE-mediated pathway (classical) and IgG-mediated pathway (alternative). In comparison to the IgE-mediated pathway which is more commonly studied, the IgG-mediated pathway is relatively new and less studied. Until now there was no consensus on the commonly studied pro-inflammatory soluble mediators for this pathway unlike in the IgE-mediated pathway. Therefore, a meta-analysis study was conducted to identify the pro-inflammatory soluble mediators that were commonly evaluated in an IgG-mediated pathway of allergy. Meta-analysis is a statistical analysis of numerous individual studies with the aim to consolidate the outcomes. Results from previous studies were systematically assessed and quantitated in order to draw conclusions about the study (Haidich, 2010). Hence, in this meta-analysis, the most common pro-inflammatory soluble mediators that are released upon induction by IgG were being identified from a collection of previous studies conducted. Subsequently, these identified soluble mediators were evaluated in the following assays to determine the anti-allergy property of *C. nutans* in the IgG-mediated pathway.

C. nutans was extracted using two different solvents, namely ethanol and water in different compositions yielding four different extracts. These extracts – 100% ethanol; 70% ethanol: 30% water; 50% ethanol: 50% water and 100% water

(denoted as 100% ethanol, 70% ethanol-aqueous, 50% ethanol-aqueous and 100% aqueous respectively hereafter) were prepared and the most active extract was identified via the *in vitro* IgE-induced mast cell degranulation model. The most active extract among the four was determined by its ability to reduce the amount of pro-inflammatory soluble mediators released upon treatment. Through this model, the anti-allergy effect of the four extracts were also analysed. As allergy is mediated through two pathways, the anti-allergy property of the most active extract was then evaluated in the IgG-induced macrophage activation model. After which, the mode of action of this most active *C. nutans* extract was explored by evaluating its effect on a number of signaling proteins that governs the degranulation and activation of the immune cells to produce pro-inflammatory soluble mediators. This happened mainly through the activation of MAPK and Akt pathways. In these pathways, signaling proteins such as ERK, JNK and p38 of the MAPK pathway were evaluated while the Akt protein was evaluated in the Akt pathway. Evaluation was done by quantifying the upregulation or down regulation of these signaling proteins by Western blot analysis.

Next, the study of the anti-allergy property of *C. nutans* was conducted *in vivo* in rodents using anaphylaxis models. Anaphylaxis models were chosen as anaphylaxis is the acute form of allergy and it involves the whole body i.e., systemic reaction. The models were ovalbumin-challenged active systemic anaphylaxis (OVA-ASA), IgE-challenged passive systemic anaphylaxis (IgE-PSA) and IgG-challenged passive systemic anaphylaxis (IgG-PSA). Before evaluating the anti-allergy effect of the most active extract in a biological system, the toxicity of the extract was determined. This was to ensure that the extract does not cause any harm to the animals and also to determine the safe doses that could be used for the anti-allergy experiments. An acute toxicity study profile of the most active extract was done with the highest recommended dosage by the Organisation of Economic Co-operation and Development (OECD) i.e., at 5000 mg/kg. The toxicity effect was assessed based on physical and behavioural observations including haematological, biochemical and histopathological analyses.

After determining the safety of the extract, the anti-allergy effect was explored in the OVA-ASA model where both IgE and IgG pathways of allergy could be activated. As this model activates both pathways, it aimed to give a general evaluation on the effect of the extract. To further determine the targeted pathway of the extract, specific IgE and IgG passive systemic anaphylaxis (PSA) models were used next. In the IgE-PSA model, only the IgE-mediated pathway will be activated while only the IgG-mediated pathway will be activated in the IgG-PSA model. The anti-allergy effect of the most active extract was determined through the ability to suppress the amount of the pro-inflammatory soluble mediators that were released by ELISA.

1.4 Research Objectives

1.4.1 General objectives

In this study, the anti-allergy property of *C. nutans* (Burm. f.) Lindau was analysed both *in vitro* and *in vivo*. In addition, the mode of action was studied.

1.4.2 Specific objectives:

- I. To identify the common soluble mediators that are important in IgG-mediated allergy by meta-analysis
- II. To evaluate the anti-allergy property of *C. nutans* extracts by *in vitro* IgE-induced mast cell degranulation model and most active extract in IgG-induced macrophage activation model and to analyse its anti-allergy mode of action
- III. To evaluate the acute toxicity of the most active *C. nutans* extract in rodents
- IV. To validate the anti-allergy property of the most active *C. nutans* extract in rodents by OVA-ASA model and to identify the anti-allergy pathways inhibited by the most active *C. nutans* extract by IgE-PSA and IgG-PSA models

1.5 Hypotheses

It was hypothesized that *C. nutans* extract does possess anti-allergy property and it will exert its effect via IgE- or IgG-mediated pathway through regulation of certain signaling molecules in allergy.

REFERENCES

- Abd Hamid, H., & Yahaya, I. H. (2016). Cytotoxicity of Clinacanthus nutans Extracts on Human Hepatoma (HepG2) Cell Line. *International Journal of Pharmacy and Pharmaceutical Sciences*, 8(10), pp. 293–295.
- Abd Hamid, H., Yahya, I. H., Yusoff, M. M., & Zareen, S. (2016). Bioassay-guided Isolation and Antioxidant Activity of Sulfur-containing Compounds from Clinacanthus nutans. *Journal of the Chinese Chemical Society*, 63, pp. 1033–1037.
- Abdul Rahim, M. H., Zakaria, Z. A., Mohd Sani, M. H., Omar, M. H., Yakob, Y., Cheema, M. S., Siew, M. C., Ahmad, Z. & Abdul Kadir, A. (2016). Methanolic Extract of Clinacanthus nutans Exerts Antinociceptive Activity via the Opioid/nitric oxide-mediated, but cGMP-independent, Pathways. *Evidence-Based Complementary and Alternative Medicine*, 2016, pp. 1–11.
- Ai, W., Li, H., Song, N., Li, L., & Chen, H. (2013). Optimal Method to Stimulate Cytokine Production and Its Use in Immunotoxicity Assessment. *International Journal of Environmental Research and Public Health*, 10, pp. 3834–3842.
- Alam, R., & Gorska, M. M. (2011). Mitogen-activated Protein Kinase Signalling and ERK1/2 Bistability in Asthma. *Clinical and Experimental Allergy*, 41, pp. 149–159.
- Alam, A., Ferdosh, S., Ghafoor, K., Hakim, A., Juraimi, A. S., Khatib, A., & Sarker, Z. I. (2016). Clinacanthus nutans: A Review of the Medicinal Uses, Pharmacology and Phytochemistry. *Asian Pacific Journal of Tropical Medicine*, 9(4), pp. 402–409.
- Amin, K. (2012). The Role of Mast Cells in Allergic Inflammation. *Respiratory Medicine*, 106, pp. 9–14.
- Arome, D., & Chinedu, E. (2014). The Importance of Toxicity Testing. *Journal of Pharmaceutical and BioSciences*, 4(2013), pp. 146–148.
- Arrighi, J.-F., Rebsamen, M., Rousset, F., Kindler, V., & Hauser, C. (2001). A Critical Role for p38 Mitogen-activated Protein Kinase in the Maturation of Human Blood-derived Dendritic Cells Induced by Lipopolysaccharide, TNF- α , and Contact Sensitizers. *The Journal of Immunology*, 166, pp. 3837–3845.

- Arullappan, S., Rajamanickam, P., Thevar, N., & Kodimani, C. C. (2014). In Vitro Screening of Cytotoxic, Antimicrobial and Antioxidant Activities of *Clinacanthus nutans* (Acanthaceae) Leaf Extracts. *Tropical Journal of Pharmaceutical Research*, 13(9), pp. 1455–1461.
- Aslam, M. S., Ahmad, M. S., & Mamat, A. S. (2015). A Review on Phytochemical Constituents and Pharmacological Activities of *Clinacanthus nutans*. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(2), pp. 30–33.
- Barnes, T. C., Anderson, M. E., & Moots, R. J. (2011). The Many Faces of Interleukin-6: The Role of IL-6 in Inflammation, Vasculopathy, and Fibrosis in Systemic Sclerosis. *International Journal of Rheumatology*, 2011, pp. 1–6.
- Barros, J. B. S., da Silva Santos, R. & da Silva Reis, A. (2019). Implication of the MAPK Signalling Pathway in the Pathogenesis of Diabetic Nephropathy. *European Medical Journal of Diabetes*, 7(1), pp. 107–114.
- Ben-Shoshan, M., & Clarke, A. E. (2011). Anaphylaxis: Past, Present and Future. *Allergy*, 66, pp. 1–14.
- Beutier, H., Hechler, B., Godon, O., Wang, Y., Gillis, C. M., de Chaisemartin, L., Gouel-Chéron, A., Magnenat, S., Macdonald, L. E., Murphy, A. J., NASA study group, Chollet-Martin, S., Longrois, D., Gachet, C., Bruhns, P. & Jönsson, F. (2018). Platelets Expressing IgG Receptor FcγRIIA/CD32A Determine the Severity of Experimental Anaphylaxis. *Science Immunology*, 3(eaan5997), pp. 1–11.
- Bhavsar, I., Miller, C. S., & Al-Sabbagh, M. (2015). Macrophage Inflammatory Protein-1 Alpha (MIP-1 alpha)/CCL3: As a Biomarker. In V. R. Preedy & V. B. Patel (Eds.), *General Methods in Biomarker Research and Their Applications, Biomarkers in Disease: Methods, Discoveries and Applications* (pp. 223–249). Dordrecht: Springer Science+Business Media.
- Bieber, T., Leung, D., El Gamal, Y., Ivancevich, J.-C. (2013) 'Section 2.4 Atopic Eczema'. In Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F. and Blaiss, M. S. (Eds), *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (pp. 44-47). Wisconsin: World Allergy Organization.
- Borenstein, M., Hedges, L. V., Higgins, J. P. T., & Rothstein, H. R. (2009). *Introduction to Meta-Analysis* (1st Edition). New Jersey: John Wiley & Sons.

- Borenstein, M., Hedges, L. V., Higgins, J. P. T., & Rothstein, H. R. (2011). *Introduction to Meta-Analysis* (revised). New Jersey: John Wiley & Sons.
- Brzóška, M. M., Moniuszko-Jakoniuk, J., Pilat-Marcinkiewicz, B., & Sawicki, B. (2003). Liver and Kidney Function and Histology in Rats Exposed to Cadmium and Ethanol. *Alcohol and Alcoholism*, 38(1), pp. 2–10.
- Chalcraft, K. R., Kong, J., Wasserman, S., Jordana, M., & McCarry, B. E. (2014). Comprehensive Metabolomic Analysis of Peanut-induced Anaphylaxis in a Murine Model. *Metabolomics*, 10, pp. 452–460.
- Chareerntantanakul, W., & Kawaree, R. (2010). Effects of Medicinal Plants Extracts on Interleukin-10 and Tumor Necrosis Factor Alpha Gene Expressions in Porcine Peripheral Blood Mononuclear Cells. *Chiang Mai Veterinary Journal*, 8, pp. 93–103.
- Charuwichitratana, S., Wongrattanapasson, N., Timpatanapong, P., & Bunjob, M. (1996). Herpes zoster: Treatment with Clinacanthus nutans Cream. *International Journal of Dermatology*, 35(9), pp. 665–666.
- Chavalittumrong, P., Attawish, A., Rugsamon, P., & Chuntapet, P. (1995). Toxicological Study of Clinacanthus nutans (Burm.f.) Lindau. *Warasan Krom Witthayasat Kan Phaet*, 37(4), pp. 232-338.
- Chen, B. H., Hung, M. H., Chen, J. Y. F., Chang, H. W., Yu, M. L., Wan, L., Tsai, F. J., Wang, T.-P., Fu, T.-F. & Chiu, C. C. (2012). Anti-allergic Activity of Grapeseed Extract (GSE) on RBL-2H3 Mast Cells. *Food Chemistry*, 132(2), pp. 968–974.
- Cherdchu, C., Poopyruchpong, N., Adcharyasucha, R., & Ratanabanangkoon, K. (1977). The Absence of Antagonism between Extracts of Clinacanthus nutans Burm. and Naja naja siamensis Venom. *Southeast Asian Journal of Tropical Medicine Public Health*, 8(2), pp. 249–254.
- Choi, I., Kim, Y., Kim, D., Choi, J., Seo, K., Im, S., Kwon, K., Lee, M., Ha, T. & Lee, H. (2003). Platelet-activating Factor-mediated NF- κ B Dependency of a Late Anaphylactic Reaction. *The Journal of Experimental Medicine*, 198(1), pp. 145–151.
- Choi, Y. H., Yan, G. H., Chai, O. H., Lim, J. M., Sung, S. Y., Zhang, X., Kim, J.-H., Choi, S. H., Lee, M. S. Han, E.-H., Kim, H. T. & Song, C. H. (2006). Inhibition of Anaphylaxis-like Reaction and Mast Cell Activation by Water Extract from the Fruiting Body of Phellinus linteus. *Biological and Pharmaceutical Bulletin*, 29(7), pp. 1360–1365.
- Chomnawang, M. T., Surassmo, S., Nukoolkarn, V. S., & Gritsanapan, W. (2005). Antimicrobial Effects of Thai Medicinal Plants against Acne-inducing Bacteria. *Journal of Ethnopharmacology*, 101, pp. 330–333.

- Chomnawang, M. T., Surassmo, S., Wongsariya, K., & Bunyapraphatsara, N. (2009a). Antibacterial Activity of Thai Medicinal Plants against Methicillin-resistant *Staphylococcus aureus*. *Fitoterapia*, 80, pp. 102–104.
- Chomnawang, M. T., Trinapakul, C., & Gritsanapan, W. (2009b). In Vitro Antigonococcal Activity of *Coscinium fenestratum* Stem Extract. *Journal of Ethnopharmacology*, 122(3), pp. 445–449.
- Chowdhury Du, S., Islam, S., Akter, R., Islam, F., Mazumdar, S., Khaleda, L., Rahman, Z. & Al-Forkan, M. (2016). Protective Effect of *Spirodela polyrhiza* on Various Organs of Arsenic-induced Wistar Albino Rats. *Journal of Cytology & Histology*, 7(2), p. 410.
- Crowley, M. T., Costello, P. S., Fitzer-Attas, C. J., Turner, M., Meng, F., Lowell, C., Tybukewicz, V. L. J. & DeFranco, A. L. (1997). A Critical Role for Syk in Signal Transduction and Phagocytosis Mediated by Fcγ Receptors on Macrophages. *Journal of Experimental Medicine*, 186(7), pp. 1027–1039.
- Cruz, E. A., Da-Silva, S. A. G., Muzitano, M. F., Silva, P. M. R., Costa, S. S., & Rossi-Bergmann, B. (2008). Immunomodulatory Pretreatment with *Kalanchoe pinnata* Extract and Its Quercitrin Flavonoid Effectively Protects Mice against Fatal Anaphylactic Shock. *International Immunopharmacology*, 8(12), pp. 1616–1621.
- Daëron, M. (1997). Fc Receptor Biology. *Annual Review of Immunology*, 15, pp. 203–234.
- Dasgupta, A. (2003). Review of Abnormal Laboratory Test Results and Toxic Effects Due to Use of Herbal Medicines. *American Journal of Clinical Pathology*, 120, pp. 127–137.
- Direkbusarakom, S., Ezura, Y., Yoshimizu, M., & Herunsalee, A. (1998a). Efficacy of Thai Traditional Herb Extracts against Fish and Shrimp Pathogenic Bacteria. *Fish Pathology*, 33(4), pp. 437–441.
- Direkbusarakom, S., Ruangpan, L., Ezura, Y., & Yoshimizu, M. (1998b). Protective Efficacy of *Clinacanthus nutans* on Yellow-head Disease in Black Tiger Shrimp (*Penaeus monodon*). *Fish Pathology*, 33(4), pp. 401–404.
- Dollah, M. A., Parhizkar, S., Latiff, L. A., & Hassan, M. H. (2013). Toxicity Effect of *Nigella sativa* on the Liver Function of Rats. *Advanced Pharmaceutical Bulletin*, 3(1), pp. 97–102.
- Dombrowicz, D., Lin, S., Flamand, V., Brini, A. T., Koller, B. H., & Kinet, J. (1998). Allergy-associated FcRβ is a Molecular Amplifier of IgE- and IgG-mediated In Vivo Responses. *Immunity*, 8, pp. 517–529.

- Doyle, E., Trosien, J., & Metz, M. (2013). Mouse Models of Allergic Disease: Methods and Protocols. In I. C. Allen (Ed.), *Methods in Molecular Biology*, 1032, pp. 133–138. Dordrecht: Springer Science+Business Media, LLC.
- eClinpath. *Veterinary Clinical Pathology*. (2013). Retrieved 13 October 2017, from <http://www.eclinpath.com/>
- Eloff, J. N. (1998). Which Extractant should be Used for the Screening and Isolation of Antimicrobial Components from Plants? *Journal of Ethnopharmacology*, 60, pp. 1–8.
- Ennis, M., Robinson, C., & Dollery, C. T. (1983). Action of 3-isobutyl-1-methylxanthine and Prostaglandins D2 and E1 on Histamine Release from Rat and Guinea Pig Mast Cells. *International Archives of Allergy and Applied Immunology*, 72, pp. 289–293.
- Esmaili, K., Alsuede, F., Abdalrahim, A., Shafaei, A., & Ismail, Z. (2015). Preliminary Phytochemical Analysis and Cytotoxicity Studies of Clinacanthus Nutans (Sabah Snake Grass). *The Open Conference Proceedings Journal*, 4(1), p. 187.
- Falanga, Y. T., Chaimowitz, N. S., Charles, N., Finkelman, F. D., Pullen, N. A., Barbour, S., Dholaria, K., Faber, T., Kolawole, M., Huang, B., Odom, S., Rivera, J., Carlyon, J., Conrad, D. H., Spiegel, S., Oskeritzian, C. A. & Ryan, J. J. (2012). Lyn but not Fyn Kinase Controls IgG-Mediated Systemic Anaphylaxis. *The Journal of Immunology*, 188, pp. 4360–4368.
- Fallon, P. G., Emson, C. L., Smith, P., & Mckenzie, A. N. J. (2001). IL-13 Overexpression Predisposes to Anaphylaxis Following Antigen Sensitization. *The Journal of Immunology*, 166, pp. 2712–2716.
- Faris, M., Kokot, N., Lee, L., & Nel, A. E. (1996). Regulation of Interleukin-2 Transcription by Inducible Stable Expression of Dominant Negative and Dominant Active Mitogen-activated Protein Kinase Kinase Kinase in Jurkat T cells: Evidence for the Importance of Ras in a Pathway that is Controlled by Dual. *Journal of Biological Chemistry*, 271(44), pp. 27366–27373.
- Farsi, E., Esmaili, K., Shafaei, A., Khaniabadi, P. M., Al Hindi, B., Ahamed, M. B. K., Sandai, D., Abdul Sattar, M., Ismail, Z., Abdul Majid, A. M. S. & Abdul Majid, A. S. (2016). Mutagenicity and Preclinical Safety Assessment of the Aqueous Extract of Clinacanthus nutans leaves. *Drug and Chemical Toxicology*, pp. 1–13.
- Filho, W. J., Lima, C. C., Paunksnis, M. R. R., Silva, A. A., Perilhão, M. S., Caldeira, M., Bocalini, D. & de Souza, R. R. (2018). Reference Database of Hematological Parameters for Growing and Aging Rats. *Aging Male*, 21(2), pp. 145–148.

- Finkelman, F. D. (2007). Anaphylaxis: Lessons from Mouse Models. *Journal of Allergy and Clinical Immunology*, 120(3), pp. 506–515.
- Finkelman, F. D., Rothenberg, M. E., Brandt, E. B., Morris, S. C., & Strait, R. T. (2005). Molecular Mechanisms of Anaphylaxis: Lessons from Studies with Murine Models. *Journal of Allergy and Clinical Immunology*, 115, pp. 449–457.
- Fiocchi, A., Sampson, H. A., Bahna, S. L. & Lack, G. (2013) 'Section 2.6 Food Allergy'. In Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F. and Blaiss, M. S. (Eds), *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (pp. 54-59). Wisconsin: World Allergy Organization.
- Fong, S. Y., Piva, T., Dekiwadia, C., Urban, S., & Huynh, T. (2016). Comparison of Cytotoxicity between Extracts of *Clinacanthus nutans* (Burm. f.) Lindau Leaves from Different Locations and the Induction of Apoptosis by the Crude Methanol Leaf Extract in D24 Human Melanoma Cells. *BMC Complementary and Alternative Medicine*, 16(368), pp. 1–12.
- Francis, A., Bosio, E., Stone, S. F., Fatovich, D. M., Arendts, G., Macdonald, S. P. J., Burrows, S. & Brown, S. G. A. (2019). Markers Involved in Innate Immunity and Neutrophil Activation are Elevated During Acute Human Anaphylaxis: Validation of A Microarray Study. *Journal of Innate Immunity*, 11, pp. 63–73.
- Fukuishi, N., Murakami, S., Ohno, A., Yamanaka, N., Matsui, N., Fukutsuji, K., Yamada, S., Itoh, K. & Akagi, M. (2014). Does β -Hexosaminidase Function Only as a Degranulation Indicator in Mast Cells? The Primary Role of β -Hexosaminidase in Mast Cell Granules. *The Journal of Immunology*, 193, pp. 1886–1894.
- Gado, A. M., & Aldahmash, B. A. (2013). Protective Effect Of L-carnitine against Amiodarone-induced Lung Toxicity in Rats. *The Internet Journal of Toxicology*, 10(1), pp. 1–9.
- Ghasemzadeh, A., Nasiri, A., Jaafar, H. Z. E., Baghdadi, A., & Ahmad, I. (2014). Changes in Phytochemical Synthesis, Chalcone Synthase Activity and Pharmaceutical Qualities of Sabah Snake Grass (*Clinacanthus nutans* L.) in Relation to Plant Age. *Molecules*, 19, pp. 17632–17648.
- Gilfillan, A. M., & Tkaczyk, C. (2006). Integrated Signalling Pathways for Mast-Cell Activation. *Nature Reviews Immunology*, 6(March), pp. 218–231.
- Gill, P., Jindal, N. L., Jagdis, A., & Vadas, P. (2015). Platelets in the Immune Response: Revisiting Platelet-activating Factor in Anaphylaxis. *Journal of Allergy and Clinical Immunology*, 135(6), pp. 1424–1432.

- Globinmed: Global Informational Hub on Integrated Medicine. *Malaysian Herbal Monograph, Medicinal Herbs and Plants Database*. (2018). Retrieved 21 December 2020, from https://globinmed.com/index.php?option=com_content&view=article&id=105729:clinacanthus-nutans-burm-f-lindau&catid=286&Itemid=357
- Goonasakaran, S. (2013). *Preliminary antimicrobial and phytochemical analysis of Clinacanthus nutans and Azadirachta indica* (Masters). Universiti Teknologi Malaysia.
- Greaves, P. (2012). *Histopathology of preclinical toxicity studies: Interpretation and relevance in drug safety studies* (4th ed.). Amsterdam: Elsevier.
- Grisanti, K., & Grayson, M. H. (2018). The Allergy Epidemic. In D. Stukus (Ed.), *Allergies and Adolescents*. Cham: Springer
- Guo, Y., Hedqvist, P., & Gustafsson, L. E. (2001). Absence of Mast Cell Involvement in Active Systemic Anaphylaxis in Rats. *European Journal of Pharmacology*, 430(2–3), pp. 305–310.
- Gupta, V., Rathore, D. S., Kansara, N. P., & Badiger, A. M. (2013). In Vivo Antioxidant Activity of Topical Cream of Cassia tora L. Leaves Extract . *Dataset Papers in Pharmacology, 2013*, pp. 1–5.
- Haidich, A. (2010). Meta-analysis in Medical Research. *Hippokratia*, 14(Suppl 1), pp. 29–37.
- Handlogten, M. W., Serezani, A. P., Sinn, A. L., Pollok, K. E., Kaplan, M. H., & Bilgicer, B. (2014). A Heterobivalent Ligand Inhibits Mast Cell Degranulation via Selective Inhibition of Allergen–IgE Interactions In Vivo. *The Journal of Immunology*, 192, pp. 2035–2041.
- Higgins, J. P. T., & Thompson, S. G. (2002). Quantifying Heterogeneity in a Meta-analysis. *Statistics in Medicine*, 21(11), pp. 1539–1558.
- Ho, S. Y., Tiew, W. P., Madhavan, P., Abdul Shukkoor, M. S., & Akowuah, G. A. (2013). Phytochemical Analysis and Antibacterial Activity of Methanolic Extract of Clinacanthus nutans Leaf. *International Journal of Drug Development & Research*, 5(3), pp. 349–355.
- Holgate, S. T., Canonica, G. W., Baena-Cagnani, C. E., Casale, T. B., Zitt, M., Nelson, H. & Vichyanond, P. (2013) 'Section 2.2 Asthma'. In Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F. and Blaiss, M. S. (Eds), *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (pp. 34-38). Wisconsin: World Allergy Organization.

- Huang, D., Guo, W., Gao, J., Chen, J., & Olatunji, J. O. (2015). Clinacanthus nutans (Burm. f.) Lindau Ethanol Extract Inhibits Hepatoma in Mice through Upregulation of the Immune Response. *Molecules*, 20, pp. 17405–17428.
- Huang, F. M., Chang, Y. C., Lee, S. S., Yang, M. L., & Kuan, Y. H. (2019). Expression of Pro-inflammatory Cytokines and Mediators Induced by Bisphenol A via ERK-NF κ B and JAK1/2-STAT3 Pathways in Macrophages. *Environmental Toxicology*, 34(4), pp. 486–494.
- Huang, L., Pi, J., Wu, J., Zhou, H., Cai, J., Li, T., & Liu, L. (2016). A Rapid and Sensitive Assay Based on Particle Analysis for Cell Degranulation Detection in Basophils and Mast Cells. *Pharmacological Research*, 111, pp. 374–383.
- Hussein, R. R. S., Soliman, R. H., Abdelhaleem Ali, A. M., Tawfeik, M. H., & Abdelrahim, M. E. A. (2013). Effect of Antiepileptic Drugs on Liver Enzymes. *Beni-Suef University Journal of Basic and Applied Sciences*, 2(1), pp. 14–19.
- Hwang, S. L., Lu, Y., Li, X., Kim, Y. D., Cho, Y. S., Jahng, Y., Son, J-K., Lee, Y. J., Kang, W., Taketomi, Y., Murakami, M., Moon, T. C. & Chang, H. W. (2014). ERK1/2 Antagonize AMPK-dependent Regulation of Fc ϵ RI-mediated Mast Cell Activation and Anaphylaxis. *The Journal of Allergy and Clinical Immunology*, 134(3), pp. 714-721.e7.
- Ishii, S., Kuwaki, T., Nagase, T., Maki, K., Tashiro, F., Sunaga, S., Cao, W-H., Kume, K., Fukuchi, Y., Ikuta, K., Miyazaki, J. Kumada, M. & Shimizu, T. (1998). Impaired Anaphylactic Responses with Intact Sensitivity to Endotoxin in Mice Lacking a Platelet-activating Factor Receptor. *The Journal of Experimental Medicine*, 187(11), pp. 1779–1788.
- Israel, H., & Richter, R. R. (2011). A Guide to Understanding Meta-analysis. *Journal of Orthopaedic and Sports Physical Therapy*, 41(7), pp. 496–504.
- Jantan, I. (2006). The Scientific Values of Malaysian Herbal Products. *Jurnal Sains Kesihatan Malaysia*, 4(1), pp. 59–70.
- Jeong, H., Koo, H., Na, H., Kim, M., Hong, S., Eom, J., Kim, K., Shin, T. & Kim, H. (2002). Inhibition of TNF- α and IL-6 Production by Aucubin through Blockade of NF- κ B Activation in RBL-2H3 Mast Cells. *Cytokine*, 18(5), pp. 252–259.
- Jimenez-Rodriguez, T. W., Garcia-Neuer, M., Alenazy, L. A., & Castells, M. (2018). Anaphylaxis in the 21st Century: Phenotypes, Endotypes, and Biomarkers. *Journal of Asthma and Allergy*, 11, pp. 121–142.

- Johansson, S. G. O., O'B Hourihane, J., Bousquet, J., Bruijnzeel-Koomen, C., Dreborg, S., Haahtela, T., Kowalski, M. L., Mygind, N., Ring, J., van Cauwenberge, P., van Hage-Hamsten, M. & Wuthrich, B. (2001). A Revised Nomenclature for Allergy. *Allergy*, 56, pp. 813–824.
- Junqueira, L., & Carneiro, J. (2003). *Basic Histology, Text and Atlas* (10th ed.). New York: Lange Medical Books McGraw-Hill.
- Jutel, M., Fukuda, T., Frew, A., Bonadonna, P. & Lockey, R. F. (2013) 'Section 2.9 Insect Allergy'. In Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F. and Blaiss, M. S. (Eds), *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (pp. 69-71). Wisconsin: World Allergy Organization.
- Kang, N. I., Kim, H. K., Ko, H. M., Kim, J. H., You, H. J., Choi, I. W., Im, S. Y. & Lee, H. K. (2008). Tumor Necrosis Factor- α Develops Late Anaphylactic Reaction through Cytosolic Phospholipase A2 Activation. *International Archives of Allergy and Immunology*, 147, pp. 315–322.
- Kemp, S. F., & Lockey, R. F. (2009). Mechanisms of Anaphylaxis. In R. Pawankar, S. Holgate, & L. J. Rosenwasser (Eds.), *Allergy Frontiers: Clinical Manifestations*, (1st ed., pp. 367–377). Japan: Springer.
- Khan, B. Q., & Kemp, S. F. (2011). Pathophysiology of Anaphylaxis. *Current Opinion in Allergy and Clinical Immunology*, 11, pp. 319–325.
- Khodoun, M. V, Strait, R., Armstrong, L., Yanase, N., & Finkelman, F. D. (2011). Identification of Markers that Distinguish IgE- from IgG-mediated Anaphylaxis. *Proceedings of the National Academy of Sciences of the United States of America*, 108(30), pp. 12413–12418.
- Khoo, L. W., Mediani, A., Zolkefee, N. K. Z., Leong, S. W., Ismail, I. S., Khatib, A., Shaari, K. & Abas, F. (2015). Phytochemical Diversity of *Clinacanthus nutans* Extracts and Their Bioactivity Correlations Elucidated by NMR Based Metabolomics. *Phytochemistry Letters*, 14, pp. 123–133.
- Khoo, L. W., Kow, A. S. F., Maulidiani, M., Ang, M. Y., Chew, W. Y., Lee, M. T., Tan, C. P., Shaari, K., Tham, C. L. & Abas, F. (2018a). ¹H-NMR Metabolomics for Evaluating the Protective Effect of *Clinacanthus nutans* (Burm. f) Lindau Water Extract against Nitric Oxide Production in LPS-IFN- γ Activated RAW 264.7 Macrophages. *Phytochemical Analysis*, pp. 1–16.
- Khoo, L. W., Kow, A. S. F., S., Lee, M. T., Tan, C. P., Shaari, K., Tham, C. L., & Abas, F. (2018b). A Comprehensive Review on Phytochemistry and Pharmacological Activities of *Clinacanthus nutans* (Burm.f.) Lindau. *Evidence-Based Complementary and Alternative Medicine*, 2018.

- Khoo, L. W., Kow, A. S. F., Maulidiani, M., Lee, M. T., Tan, C. P., Shaari, K., Tham, C. L. & Abas, F. (2018c). Hematological, Biochemical, Histopathological and ¹H-NMR Metabolomics Application in Acute Toxicity Evaluation of Clinacanthus nutans Water Leaf Extract. *Molecules*, 23(9), pp. 2172-2189.
- Khoo, L. W., Kow, A. S. F., Maulidiani, M., Lee, M. T., Tan, C. P., Shaari, K., Tham, C. L. & Abas, F. (2018d). Plasma and Urine Metabolite Profiling Reveals the Protective Effect of Clinacanthus nutans in an Ovalbumin-induced Anaphylaxis Model: ¹H-NMR Metabolomics Approach. *Journal of Pharmaceutical and Biomedical Analysis*, 158, pp. 438–450.
- Kim, N. S., Lee, S. E., Choi, D. J., Choi, E. J., Lee, J. H., Park, S., Lee, J. H., Park, S., Lee, Y. S., Lee, J. W., Lee, D. Y., Kim, G. S. & Lee, S. E. (2018). Screening and Evaluation of the Anti-allergic Effect of Korean Medicinal Plant Extracts. *Korean Journal of Medicinal Crop Science*, 26(1), pp. 42–54.
- Kimata, M., Inagaki, N., Kato, T., Miura, T., Serizawa, I., & Nagai, H. (2000). Roles of Mitogen-activated Protein Kinase Pathways for Mediator Release from Human Cultured Mast Cells. *Biochemical Pharmacology*, 60(4), pp. 589–594.
- Kongkaew, C., & Chaiyakunapruk, N. (2011). Efficacy of Clinacanthus nutans Extracts in Patients with Herpes Infection: Systematic Review and Meta-analysis of Randomised Clinical Trials. *Complementary Therapies in Medicine*, 19, pp. 47–53.
- Kowalski, M. L., Demoly, P., Pichler, W. J. & Sanchez-Borges, M. (2013) 'Section 2.8 Hypersensitivity to Drugs and Biological Agents'. In Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F. and Blaiss, M. S. (Eds), *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (pp. 64-68). Wisconsin: World Allergy Organization.
- Kumar, S., Boehm, J., & Lee, J. C. (2003). P38 MAP Kinases: Key Signalling Molecules as Therapeutic Targets for Inflammatory Diseases. *Nature Reviews Drug Discovery*, 2, pp. 717–726.
- Kunsorn, P., Ruangrunsi, N., Lipipun, V., Khanboon, A., & Rungsihirunrat, K. (2013). The Identities and Anti-herpes Simplex Virus Activity of Clinacanthus nutans and Clinacanthus siamensis. *Asian Pacific Journal of Tropical Biomedicine*, 3(4), pp. 284–290.
- Kwan, Y. P., Darah, I., Chen, Y., Sreeramanan, S., & Sasidharan, S. (2013). Acute and Subchronic Toxicity Study of Euphorbia hirta L. Methanol Extract in Rats. *BioMed Research International*, 2013, pp. 1–14.

- Laffargue, M., Calvez, R., Finan, P., Trifilieff, A., Barbier, M., Altruda, F., Hirsch, E. & Wymann, M. P. (2002). Phosphoinositide 3-Kinase γ is an Essential Amplifier of Mast Cell Function. *Immunity*, 16, pp. 441–451.
- Lau, K. W., Lee, S. K. & Chin, J. H. (2014). Effect of the Methanol Leaves Extract of *Clinacanthus nutans* on the Activity of Acetylcholinesterase in Male Mice. *Journal of Acute Disease*, pp. 22–25.
- Lee, S. Y., Mediani, A., Nur Ashikin, A. H., Azliana, A. B. S., & Abas, F. (2014). Antioxidant and α -glucosidase Inhibitory Activities of the Leaf and Stem of Selected Traditional Medicinal Plants. *International Food Research Journal*, 21(1), pp. 165–172.
- Lemonnier, N., Zhou, G.-B., Prasher, B., Mukerji, M., Chen, Z., Brahmachari, S. K., Noble, D., Auffray, C. & Sagner, M. (2017). Traditional Knowledge-based Medicine: A Review of History, Principles and Relevance in the Present Context of P4 Systems Medicine. *Progress in Preventive Medicine*, 2(7), p. e0011.
- Levey, H. A. (1969). Toxicity of the Venom of the Sea-snake, *Laticauda colubrina*, with Observations on a Malay 'Folk Cure.' *Toxicon*, 6(4), pp. 269–276.
- Li, Y. C., Yeh, C. H., Yang, M. L., & Kuan, Y. H. (2012). Luteolin Suppresses Inflammatory Mediator Expression by Blocking the Akt/NF κ B Pathway in Acute Lung Injury Induced by Lipopolysaccharide in Mice. *Evidence-Based Complementary and Alternative Medicine*, 2012, p. 383608.
- Liew, S. Y., Stanbridge, E. J., Yusoff, K., & Shafee, N. (2012). Hypoxia Affects Cellular Responses to Plant Extracts. *Journal of Ethnopharmacology*, 144, pp. 453–456.
- Liju, V. B., Jeena, K., & Kuttan, R. (2013). Acute and Subchronic Toxicity as well as Mutagenic Evaluation of Essential Oil from Turmeric (*Curcuma longa* L). *Food and Chemical Toxicology*, 53, pp. 52–61.
- Lin, Y. T., Wu, C. T., Huang, J. L., Cheng, J. H., & Yeh, K. W. (2016). Correlation of Ovalbumin of Egg White Components with Allergic Diseases in Children. *Journal of Microbiology, Immunology and Infection*, 49(1), pp. 112–118.
- Lionetto, M. G., Caricato, R., Calisi, A., Giordano, M. E., & Schettino, T. (2013). Acetylcholinesterase as a Biomarker in Environmental and Occupational Medicine: New Insights and Future Perspectives. *BioMed Research International*, 2013, pp. 1–8.

- Liu, Q. M., Xie, C. L., Gao, Y. Y., Liu, B., Lin, W. X., Liu, H., Cao, M. J., Su, W. J., Yang, X. W. & Liu, G. M. (2018). Deep-sea-derived Butyrolactone I Suppresses Ovalbumin-induced Anaphylaxis by Regulating Mast Cell Function in a Murine Model. *Journal of Agricultural and Food Chemistry*, 66, pp. 5581–5592.
- Lorke, D. (1983). A New Approach to Practical Acute Toxicity Testing. *Archives of Toxicology*, 54, pp. 275–287.
- Lu, Y., Yang, J. H., Li, X., Hwangbo, K., Hwang, S. L., Taketomi, Y., Murakami, M., Chang, Y. C., Kim, C. H., Son, J. K. & Chang, H. W. (2011). Emodin, a Naturally Occurring Anthraquinone Derivative, Suppresses IgE-mediated Anaphylactic Reaction and Mast Cell Activation. *Biochemical Pharmacology*, 82(11), pp. 1700–1708.
- Lusia Berek, M., Hasmadi, M., Zaleha, A. Z., & Mohd Fadzelly, A. B. (2015). Effect of Different Drying Methods on Phytochemicals and Antioxidant Properties of Unfermented and Fermented Teas from Sabah Snake Grass (*Clinacanthus nutans* Lind.) Leaves. *International Food Research Journal*, 22(2), pp. 661–670.
- Maamar, M. S., Al-Griw, M. A., Al-Ghazeer, R. O., Al-Azreg, S. A., Salama, N. M., & Bennour, E. M. (2016). Oxidative Stress Mediated Cytotoxicity of Trichloroethane in a Model of Murine Splenic Injury. *American Journal of Bioscience*, 4(1), pp. 1–8.
- Mahendran, S., & Abdul Rashid, N. (2016). Formulation, Evaluation and Antibacterial Properties of Herbal Ointment Containing Methanolic Extract of *Clinacanthus nutans* Leaves. *International Journal of Pharmaceutical and Clinical Research*, 8(8), pp. 1170–1174.
- Mai, C. W., Yap, K. S. I., Kho, M. T., Ismail, N. H., Yusoff, K., Shaari, K., Chin, S. Y. & Lim, E. S. H. (2016). Mechanisms Underlying the Anti-inflammatory Effects of *Clinacanthus nutans* Lindau Extracts: Inhibition of Cytokine Production and Toll-like Receptor-4 Activation. *Frontiers in Pharmacology*, 7(FEB), pp. 1–11.
- Mannhalter, J. W., Neychev, H. O., Zlabinger, G. J., Ahmad, R., & Eibl, M. M. (1985). Modulation of the Human Immune Response by the Non-toxic and Non-pyrogenic Adjuvant Aluminium Hydroxide: Effect on Antigen Uptake and Antigen Presentation. *Clinical and Experimental Immunology*, 61(1), pp. 143–151.
- Md Said, N. & Abiola., O. (2014). Haematological Profile Shows that Inbred Sprague Dawley Rats have Exceptional Promise for Use in Biomedical and Pharmacological Studies. *Asian Journal of Biomedical and Pharmaceutical Sciences*, 4(37), pp. 33–37.

- Mendis, S. (2014) 'Contribution of Allergy to the Burden of Non-communicable Diseases'. In Akdis, C. A. and Agache, I. (Eds), *Global Atlas of Allergy*. (pp. 336-337). Zurich: European Academy of Allergy and Clinical Immunology.
- Meulenbroek, A. (2008). *Useful Diagnostic Markers for Immunocompetence* (Third Edit). Amsterdam: Sanquin.
- Meyer, F., Lizana, J. N., Dziedricki, L. F., & Bleggi-Torres, L. F. (2010). Histologic Alterations of Rat Kidneys Perfused with a Euro-Collins Diltiazem Solution. *Acta Cirurgica Brasileira*, 25(6), pp. 496–500.
- Minai-Fleminger, Y. & Levi-Schaffer, F. (2009). Mast Cells and Eosinophils: The Two Key Effector Cells in Allergic Inflammation. *Inflammation Research*, 58, pp. 631–638.
- Miyazaki, D., Nakamura, T., Toda, M., Cheung-Chau, K.-W., Richardson, R. M., & Ono, S. J. (2005). Macrophage Inflammatory Protein-1 α as a Costimulatory Signal for Mast Cell-mediated Immediate Hypersensitivity Reactions. *The Journal of Clinical Investigation*, 115(2), pp. 434–442.
- Moon, T. C., Befus, A. D., & Kulka, M. (2014). Mast Cell Mediators: Their Differential Release and the Secretory Pathways Involved. *Frontiers in Immunology*, 5, pp. 1–18.
- Muñoz-Cano, R., Picado, C., Valero, A., & Bartra, J. (2016). Mechanisms of Anaphylaxis Beyond IgE. *Journal of Investigational Allergology and Clinical Immunology*, 26(2), pp. 73–82.
- Muñoz-Cano, R., Pascal, M., Araujo, G., Goikoetxea, M. J., Valero, A. L., Picado, C., & Bartra, J. (2017). Mechanisms, Cofactors, and Augmenting Factors Involved in Anaphylaxis. *Frontiers in Immunology*, 8, pp. 1–7.
- Musio, S., Pedotti, P., Mantegazza, R., Ohtsu, H., Boon, L., Steinman, L., Galli, S. J. & Pedotti, R. (2009). Anaphylaxis to a Self-peptide in the Absence of Mast Cells or Histamine. *Laboratory Investigation*, 89, pp. 398–405.
- Naik, S. R., Bhagat, S., Shah, P. D., Tare, A. A., Ingawale, D., & Wadekar, R. R. (2013). Evaluation of Anti-allergic and Anti-anaphylactic Activity of Ethanolic Extract of Zizyphus jujuba Fruits in Rodents. *Revista Brasileira de Farmacognosia*, 23, pp. 811–818.
- Neveu, W. A., Allard, J. B., Dienz, O., Wargo, M. J., Ciliberto, G., Whittaker, L. A., & Rincon, M. (2009). IL-6 is Required for Airway Mucus Production Induced by Inhaled Fungal Allergens. *Journal of Immunology*, 183(3), pp. 1732–1738.

- Ng, C. T., Fong, L. Y., Tan, J. J., Rajab, N. F., Abas, F., Shaari, K., Chan, K. M., Juliana, F. & Yong, Y. K. (2018). Water Extract of *Clinacanthus nutans* Leaves Exhibits In Vitro, Ex Vivo and In Vivo Anti-angiogenic Activities in Endothelial Cell via Suppression of Cell Proliferation. *BMC Complementary and Alternative Medicine*, 18, p. 210.
- Nimmerjahn, F. (2014). Molecular and Cellular Pathways of Immunoglobulin G Activity In Vivo. *ISRN Immunology*, 2014, pp. 1–13.
- Nirmal, S. A., Vaykole, A. M., Khadse, G. B., Pal, S. C., & Mandal, S. C. (2013). Mast Cell Degranulation: A Target for Bioactive Natural Products. *Phytotherapy Research*, 27(3), pp. 575–597.
- Nordmann, A. J., Kasenda, B., & Briel, M. (2012). Meta-analyses: What they can and cannot do. *Swiss Medical Weekly*, 142, p. w13518.
- Oettgen, H. C., Martin, T. R., Wynshaw-Boris, A., Deng, C., Drazen, J. M., & Leder, P. (1994). Active Anaphylaxis in IgE-deficient Mice. *Nature*, 370, pp. 367–370.
- Okamoto, T., Iwata, S., Ohnuma, K., Dang, N. H., & Morimoto, C. (2009). Histamine H1-Receptor Antagonists with Immunomodulating Activities: Potential Use for Modulating T Helper Type 1 (Th1)/Th2 Cytokine Imbalance and Inflammatory Responses in Allergic Diseases. *The Journal of Translational Immunology*, 157, pp. 27–34.
- Olivera, A., Dillahunt, S. E., & Rivera, J. (2013). Interrogation of Sphingosine-1-phosphate Receptor 2 Function In Vivo Reveals a Prominent Role in the Recovery from IgE and IgG-mediated Anaphylaxis with Minimal Effect on its Onset. *Immunology Letters*, 150, pp. 89–96.
- Ong, S. L., Paneerchelvan, S., Lai, H. Y., & Rao, N. K. (2014). In Vitro Lipase Inhibitory Effect of Thirty-Two Selected Plants in Malaysia. *Asian Journal of Pharmaceutical and Clinical Research*, 7(SUPPL. 2), pp. 19–24.
- Organisation for Economic Co-operation and Development (2001). *OECD Guideline for Testing of Chemicals*. Paris: Organisation for Economic Co-operation and Development.
- P'ng, X. W., Akowuah, G. A., & Chin, J. H. (2012). Acute Oral Toxicity Study of *Clinacanthus nutans* in Mice. *International Journal of Pharmaceutical Sciences and Research*, 3(11), pp. 4202–4205.
- Pan, S. Y., Litscher, G., Gao, S. H., Zhou, S. F., Yu, Z. L., Chen, H. Q., Zhang, S. F., Tang, M. K., Sun, J. N. & Ko, K. M. (2014). Historical Perspective of Traditional Indigenous Medical Practices: The Current Renaissance and Conservation of Herbal Resources. *Evidence-Based Complementary and Alternative Medicine*, 2014.

- Pannangpetch, P., Laupattarakasem, P., Kukongviriyapan, V., Kukongviriyapan, U., Kongyingoes, B., & Aromdee, C. (2007). Antioxidant Activity and Protective Effect against Oxidative Hemolysis of *Clinacanthus nutans* (Burm. f) Lindau. *Songklanakarin Journal of Science and Technology*, 29(Suppl. 1), pp. 1–9.
- Parameswaran, N., & Patial, S. (2010). Tumor Necrosis Factor- α Signaling in Macrophages. *Critical Reviews in Eukaryotic Gene Expression*, 20(2), pp. 87–103.
- Passante, E. (2014). Mast Cell and Basophil Cell Lines: A Compendium. In F. H. Gibbs, Bernhard F. and Falcone (Ed.), *Basophils and Mast Cells: Methods and Protocols, Methods in Molecular Biology* (1192nd ed., pp. 101–113). New York: Springer Science+Business Media, LLC.
- Pawankar, R., Canonica, G. W., Holgate, S. T., & Lockey, R. F. (2011). *WAO White Book on Allergy 2011-2012: Executive Summary*. Wisconsin: World Allergy Organization.
- Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F., & Blaiss, M. S. (2013). *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (R. Pawankar, G. W. Canonica, S. T. Holgate, R. F. Lockey, & M. S. Blaiss, Eds.). Wisconsin: World Allergy Organization.
- Peavy, R. D., & Metcalfe, D. D. (2008). Understanding the Mechanisms of Anaphylaxis. *Current Opinion in Allergy and Clinical Immunology*, 8(4), pp. 310–315.
- Petterino, C., & Argentino-Storino, A. (2006). Clinical Chemistry and Haematology Historical Data in Control Sprague-Dawley Rats from Pre-clinical Toxicity Studies. *Experimental and Toxicologic Pathology*, 57, pp. 213–219.
- Pongmuangmul, S., Phumiamorn, S., Sanguansermisri, P., Wongkattiya, N., Fraser, I. H., & Sanguansermisri, D. (2016). Anti-herpes Simplex Virus Activities of Monogalactosyl Diglyceride and Digalactosyl Diglyceride from *Clinacanthus nutans*, a Traditional Thai Herbal Medicine. *Asian Pacific Journal of Tropical Biomedicine*, 6(3), pp. 192–197.
- Pongphasuk, N., Khunkitti, W., & Chicharoenthum, M. (2005). Anti-Inflammatory and Analgesic Activities of the Extract from *Garcinia mangostana* Linn. In Palaniswamy, U. R., Craker, L. E. and Gardner, Z. E. (Eds.), *WOCMAP III, Vol. 6: Traditional Medicine and Nutraceuticals* (pp. 125–130).

- Punnonen, J., Yssel, H., & De Vries, J. E. (1997). The Relative Contribution of IL-4 and IL-13 to Human IgE Synthesis Induced by Activated CD4+ or CD8+ T cells. *Journal of Allergy and Clinical Immunology*, 100(6 1), pp. 792–801.
- Putwatana, P., Sanmanowong, P., Oonprasertpong, L., Junda, T., Pitiporn, S., & Narkwong, L. (2009). Relief of Radiation-induced Oral Mucositis in Head and Neck Cancer. *Cancer Nursing*, 32(1), pp. 82–87.
- Rathnasamy, S., Mohamed, K., Sulaiman, S. F., & Akinboro, A. (2013). Evaluation of Cytotoxic, Mutagenic and Antimutagenic Potential of Leaf Extracts of Three Medicinal Plants using *Allium cepa* Chromosome Assay. *International Current Pharmaceutical Journal*, 2(8), pp. 131–140.
- Raya, K. B., Ahmad, S. H., Farhana, S. F., Mohammad, M., Tajidin, N. E., & Parvez, A. (2015). Changes in Phytochemical Contents in Different Parts of *Clinacanthus nutans* (Burm. f.) Lindau due to Storage Duration. *Bragantia, Campinas*, 74(4), pp. 445–452.
- Reber, L. L., Hernandez, J. D., & Galli, S. J. (2017). The Pathophysiology of Anaphylaxis. *The Journal of Allergy and Clinical Immunology*, 140(2), pp. 335–348.
- Reed, G. L., Fitzgerald, M. L., & Polgár, J. (2000). Molecular Mechanisms of Platelet Exocytosis: Insights into the “Secrete” Life of Thrombocytes. *Blood*, 96(10), pp. 3334–3342.
- Ring, J. (2014). What is Allergy. In C. A. Akdis & I. Agache (Eds.), *Global Atlas of Allergy* (pp. 2–3). Zurich: European Academy of Allergy and Clinical Immunology.
- Rivera, J., & Gilfillan, A. M. (2006). Molecular Regulation of Mast Cell Activation. *Journal of Allergy and Clinical Immunology*, 117(6), pp. 1214–1225.
- Roeslan, M. O., Ayudhya, T. D. N., Yingyongnarongkul, B. & Koontongkaew, S. (2019). Anti-biofilm, Nitric Oxide Inhibition and Wound Healing Potential of Purpurin-18 Phytyl Ester Isolated from *Clinacanthus nutans* Leaves. *Biomedicine and Pharmacotherapy*, 113, pp. 1–13.
- Rosales, C., & Uribe-Querol, E. (2013). Fc Receptors: Cell Activators of Antibody Functions. *Advances in Bioscience and Biotechnology*, 4, pp. 21–33.
- Sag, D., Carling, D., Stout, R. D., & Suttles, J. (2008). Adenosine 5'-monophosphate-activated Protein Kinase Promotes Macrophage Polarization to an Anti-inflammatory Functional Phenotype. *The Journal of Immunology*, 181(12), pp. 8633–8641.

- Sakdarat, S., Sittiso, S., Ekalaksananan, T., Pientong, C., Charoensri, N., & Kongyingoes, B. (2017). Study on Effects of Compounds from *Clinacanthus nutans* on Dengue Virus Type 2 Infection. *SSRN Electronic Journal*, 25, pp. 272–275.
- Sangkitporn, S., Chaiwat, S., Balachandra, K., Dechatiwongse Na-Ayudhaya, T., Bunjob, M., & Jayavas, C. (1995). Treatment of Herpes Zoster with *Clinacanthus nutans* (Bi Phaya Yaw) Extract. *Journal of the Medical Association of Thailand*, 78(11), pp. 624–627.
- Sarega, N., Imam, M. U., Esa, N. M., Zawawi, N., & Ismail, M. (2016a). Effects of Phenolic-rich Extracts of *Clinacanthus nutans* on High Fat and High Cholesterol Diet-induced Insulin Resistance. *BMC Complementary and Alternative Medicine*, 16, pp. 88-98.
- Sarega, N., Imam, M. U., Ooi, D. J., Chan, K. W., Md Esa, N., Zawawi, N., & Ismail, M. (2016b). Phenolic Rich Extract from *Clinacanthus nutans* Attenuates Hyperlipidemia-associated Oxidative Stress in Rats. *Oxidative Medicine and Cellular Longevity*, 2016, pp. 1–16.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., & Yoga Latha, L. (2011). Extraction, Isolation and Characterization of Bioactive Compounds from Plants' Extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(1), pp. 1–10.
- Schaper, F., & Rose-John, S. (2015). Interleukin-6: Biology, Signaling and Strategies of Blockade. *Cytokine and Growth Factor Reviews*, 26, pp. 475–487.
- Scholey, J. M., & Harrison, J. E. (2003). Publication Bias: Raising Awareness of a Potential Problem in Dental Research. *British Dental Journal*, 194(5), pp. 235–237.
- Simons, F. E. R., Ebisawa, M., Sanchez-Borges, M., Thong, B. Y., Worm, M., Tanno, L. K., Lockey, R. F., El-Gamal, Y. M., Brown, S. G. A., Park, H. S. & Sheikh, A. (2015). 2015 Update of the Evidence Base: World Allergy Organization Anaphylaxis Guidelines. *World Allergy Organization Journal*, 8(32), pp. 32-47.
- Singh, M., McKenzie, K., & Ma, X. (2017). Effect of Dimethyl Sulfoxide on In Vitro Proliferation of Skin Fibroblast Cells. *Journal of Biotech Research*, 8, pp. 78–82.
- Sriwanthana, B., Chavalittumrong, P., & Chompuk, L. (1996). Effect of *Clinacanthus nutans* on Human Cell-mediated Immune Response In Vitro. *Thai Journal of Pharmaceutical Science*, 20(4), pp. 261–267.

- Stone, S. F., & Brown, S. G. A. (2012). Mediators Released during Human Anaphylaxis. *Current Allergy and Asthma Reports*, 12, pp. 33–41.
- Stone, S. F., Cotterell, C., Isbister, G. K., Holdgate, A., & Brown, S. G. A. (2009). Elevated Serum Cytokines during Human Anaphylaxis: Identification of Potential Mediators of Acute Allergic Reactions. *Journal of Allergy and Clinical Immunology*, 124(4), pp. 786-792.e4.
- Strait, R. T., Morris, S. C., Yang, M., Qu, X. W., & Finkelman, F. D. (2002). Pathways of Anaphylaxis in the Mouse. *Journal of Allergy and Clinical Immunology*, 109(4), pp. 658–668.
- Strait, R. T., Morris, S. C., Smiley, K., Urban, J. F. J., & Finkelman, F. D. (2003). IL-4 Exacerbates Anaphylaxis. *The Journal of Immunology*, 170, pp. 3835–3842.
- Sun, J., Arias, K., Alvarez, D., Fattouh, R., Walker, T., Goncharova, S., Kim, B., Wasserman, S., Reed, J., Coyle, A. J. & Jordana, M. (2007). Impact of CD40 Ligand, B Cells, and Mast Cells in Peanut-induced Anaphylactic Responses. *The Journal of Immunology*, 179, pp. 6696–6703.
- Syrbu, S. I., Waterman, W. H., Molski, T. F. P., Nagarkatti, D., Hajjar, J. J., & Sha'afi, R. I. (1999). Phosphorylation of Cytosolic Phospholipase A2 and the Release of Arachidonic Acid in Human Neutrophils. *The Journal of Immunology*, 162, pp. 2334–2340.
- Tan, C. S. H., Ho, C. F. Y., Heng, S. S., Wu, J. S., Tan, B. K. H., Ng, Y. K., Sun, G. Y., Lin, T. N. & Ong, W. Y. (2016). Clinacanthus nutans Extracts Modulate Epigenetic Link to Cytosolic Phospholipase A2 Expression in SH-SY5Y Cells and Primary Cortical Neurons. *NeuroMolecular Medicine*, 18, pp. 441–452.
- Tan, J. W., Israf, D. A., Harith, H. H., Md Hashim, N. F., Ng, C. H., Shaari, K., & Tham, C. L. (2017). Anti-allergic Activity of 2,4,6-trihydroxy-3-geranylacetophenone (tHGA) via Attenuation of IgE-mediated Mast Cell Activation and Inhibition of Passive Systemic Anaphylaxis. *Toxicology and Applied Pharmacology*, 319, pp. 47–58.
- Tan, T. Y. C., Lee, J. C., Mohd Yusof, N. A., Teh, B. P., & Syed Mohamed, A. F. (2020). Malaysian Herbal Monograph Development and Challenges. *Journal of Herbal Medicine*, 23(June 2020), p. 100380.
- Teoh, P. L., Cheng, A. Y. F., Liau, M., Lem, F. F., Kaling, G. P., Chua, F. N., & Cheong, B. E. (2017). Chemical Composition and Cytotoxic Properties of Clinacanthus nutans Root Extracts. *Pharmaceutical Biology*, 55(1), pp. 394–401.

- Thompson, S. G., & Sharp, S. J. (1999). Explaining Heterogeneity in Meta-analysis: A Comparison of Methods. *Statistics in Medicine*, 18(20), pp. 2693–2708.
- Thongchai, S., Ekalaksananan, T., Pientong, C., Aromdee, C., Kongyingyoes, B., & Seubsasana, S. (2008). Anti-herpes Simplex Virus Type 1 Activity of Crude Ethyl Acetate Extract of *Clinacanthus nutans*. *Journal of Science and Technology Mahasarakham University*, 27(4), pp. 318–326.
- Thongharb, C. & Tejasen, P. (1977). The Effect of Slaed Pang Porn (*Clinacanthus nutans*) on Thailand Cobra Venom (*Naja naja siamensis*). *Thai Journal of Pharmaceutical Sciences*, 2, pp. 1057-1063.
- Thongrakard, V., & Tencomnao, T. (2010). Modulatory Effects of Thai Medicinal Plant Extract on Proinflammatory Cytokines-induced Apoptosis in Human Keratinocyte HaCat Cells. *African Journal of Biotechnology*, 9(31), pp. 4999–5003.
- Tiew, W. P., Wen, P. X., Han, C. J., & Akowuah, G. A. (2014). Effect of Methanol Extract of *Clinacanthus nutans* on Serum Biochemical Parameters in Rats. *Journal of Applied Pharmacy*, 6(1), pp. 77–86.
- Tilburt, J. C., & Kaptchuk, T. J. (2008). Herbal Medicine Research and Global Health: An Ethical Analysis. *Bulletin of the World Health Organization*, 86, pp. 594–599.
- Timpawat, S., & Vajrabhaya, L. (1994). Clinical Evaluation of *Clinacanthus nutans* Lindau in Orabase in the Treatment of Recurrent Aphthous Stomatitis. *Mahidol Dental Journal*, 14(1), pp. 10–16.
- Tkaczyk, C., Okayama, Y., Woolhiser, M. R., Hagaman, D. D., Gilfillan, A. M., & Metcalfe, D. D. (2001). Activation of Human Mast Cells through the High Affinity IgG Receptor. *Molecular Immunology*, 38, pp. 1289–1293.
- Togias, A. (2004). Systemic Effects of Local Allergic Disease. *Journal of Allergy and Clinical Immunology*, 113(1), pp. S8-14.
- Tsai, H. Da, Wu, J. S., Kao, M. H., Chen, J. J., Sun, G. Y., Ong, W. Y., & Lin, T. N. (2016). *Clinacanthus nutans* Protects Cortical Neurons against Hypoxia-induced Toxicity by Downregulating HDAC1/6. *NeuroMolecular Medicine*, 18, pp. 274–282.
- Tsujimura, Y., Obata, K., Mukai, K., Shindou, H., Yoshida, M., Nishikado, H., Kawano, Y., Minegishi, Y., Shimizu, T. & Karasuyama, H. (2008). Basophils Play a Pivotal Role in Immunoglobulin-G-mediated but not Immunoglobulin-E-mediated Systemic Anaphylaxis. *Immunity*, 28(April), pp. 581–589.

- Tu, S.-F., Liu, R. H., Cheng, Y.-B., Hsu, Y.-M., Du, Y.-C., El-Shazly, M., Wu, Y.-C. & Chang, F.-R. (2014). Chemical Constituents and Bioactivities of *Clinacanthus nutans* Aerial Parts. *Molecules*, 19, pp. 20382–20390.
- Turner, R. M., Bird, S. M., & Higgins, J. P. T. (2013). The Impact of Study Size on Meta-analyses: Examination of Underpowered Studies in Cochrane Reviews. *PLoS ONE*, 8(3), p. e59202.
- Uawonggul, N., Chaveerach, A., Thammasirirak, S., Arkaravichien, T., Chuachan, C., & Daduang, S. (2006). Screening of Plants Acting against *Heterometrus laoticus* Scorpion Venom Activity on Fibroblast Cell Lysis. *Journal of Ethnopharmacology*, 103, pp. 201–207.
- Uawonggul, N., Thammasirirak, S., Chaveerach, A., Chuachan, C., Daduang, J., & Daduang, S. (2011). Plant Extract Activities against the Fibroblast Cell Lysis by Honey Bee Venom. *Journal of Medicinal Plants Research*, 5(10), pp. 1978–1986.
- US Department of Health and Human Services (2005). *Guidance for Industry: Estimating the Maximum Safe Starting Dose in Initial Clinical Trials for Therapeutics in Adult Healthy Volunteers*. Rockville, MD: Centre for Drug Evaluation and Research, Food and Drug Administration
- Vadas, P., Gold, M., Perelman, B., Liss, G. M., Lack, G., Blyth, T., Simons, F. E. R., Simons, K. J., Cass, D. & Yeung, J. (2008). Platelet-activating Factor, PAF Acetylhydrolase, and Severe Anaphylaxis. *The New England Journal of Medicine*, 358(1), pp. 28–35.
- Vandenplas, O., Worm, M., Cullinan, P., Park, H. S. & van Wijk, R. G. (2013) 'Section 2.10 Occupational Allergy' In Pawankar, R., Canonica, G. W., Holgate, S. T., Lockey, R. F. and Blaiss, M. S. (Eds) *World Allergy Organization (WAO) White Book on Allergy: Update 2013* (pp. 72-76). Wisconsin: World Allergy Organization.
- Wakayama, H., Hasegawa, Y., Kawabe, T. Saito, H., Kikutani, H. & Shimokata, K. (1998). IgG-mediated Anaphylaxis via Fcγ Receptor in CD40-deficient Mice. *Clinical and Experimental Immunology*, 114, pp. 154-160.
- Wanikiat, P., Panthong, A., Sujayanon, P., Yoosook, C., Rossi, A. G. & Reutrakul, V. (2008). The Anti-inflammatory Effects and the Inhibition of Neutrophil Responsiveness by *Barleria lupulina* and *Clinacanthus nutans* Extracts. *Journal of Ethnopharmacology*, 116, pp. 234–244.
- Wendeler, M. & Sandhoff, K. (2009). Hexosaminidase Assays. *Glycoconjugate Journal*, 26, pp. 945–952.

- Wong, F-C., Yong, A-L., Ong, H-C. & Chai, T-T. (2013). Evaluation of the Antibacterial Activities of Selected Medicinal Plants and Determination of their Phenolic Constituents. *ScienceAsia*, 39, pp. 591–595.
- Wong, F-C., Yong, A-L., Ting, E. P-S., Khoo, S-C., Ong, H-C. & Chai, T-T. (2014). Antioxidant, Metal Chelating, Anti-glucosidase Activities and Phytochemical Analysis of Selected Tropical Medicinal Plants. *Iranian Journal of Pharmaceutical Research*, 13(4), pp. 1409–1415.
- World Health Organization (2019). *WHO Global Report on Traditional and Complementary Medicine 2019*. Geneva: World Health Organization.
- Wu, J. S., Kao, M-H., Tsai, H-D., Cheung, W-M., Chen, J-J., Ong, W-Y., Sun, G. Y. & Lin, T-N. (2018). Clinacanthus nutans Mitigates Neuronal Apoptosis and Ischemic Brain Damage through Augmenting the C/EBP β -driven PPAR- γ Transcription. *Molecular Neurobiology*, 55, pp. 5425–5438.
- Xing, Y., Liu, Z., Yang, G., Gao, D. & Niu, X. (2015). MicroRNA Expression Profiles in Rats with Selenium Deficiency and the Possible Role of the Wnt/ β -catenin Signaling Pathway in Cardiac Dysfunction. *International Journal of Molecular Medicine*, 35, pp. 143–152.
- Xu, W., Tamura, T. & Takatsu, K. (2008). CpG ODN Mediated Prevention from Ovalbumin-induced Anaphylaxis in Mouse through B Cell Pathway. *International Immunopharmacology*, 8(2), pp. 351-361.
- Yahaya, R., Dash, G. K., Abdullah, M. S. & Mathews, A. (2015). Clinacanthus nutans (Burm. f.) Lindau: An Useful Medicinal Plant of South-East Asia. *International Journal of Pharmacognosy and Phytochemical Research*, 7(6), pp. 1244–1250.
- Yalavarthi, C. & Thiruvengadarajan, V. S. (2013). A Review on Identification Strategy of Phyto Constituents Present in Herbal Plants. *International Journal of Research in Pharmaceutical Sciences*, 4(2), pp. 123–140.
- Yimin & Kohanawa, M. (2006). A Regulatory Effect of the Balance between TNF- α and IL-6 in the Granulomatous and Inflammatory Response to Rhodococcus aurantiacus Infection in Mice. *The Journal of Immunology*, 177, pp. 642–650.
- Yong, Y. K., Tan, J. J., Teh, S. S., Mah, S. H., Cheng, G. L. E., Chiong, H. S. & Ahmad, Z. (2013). Clinacanthus nutans Extracts are Antioxidant with Antiproliferative Effect on Cultured Human Cancer Cell Lines. *Evidence-based Complementary and Alternative Medicine*, 2013, pp. 1–8.

- Yoosook, C., Panpisutchai, Y., Chaichana, S., Santisuk, T. & Reutrakul, V. (1999). Evaluation of Anti-HSV-2 Activities of *Barleria lupulina* and *Clinacanthus nutans*. *Journal of Ethnopharmacology*, 67, pp. 179–187.
- Yuann, J-M. P., Wang, J-S., Jian, H-L., Lin, C-C. & Liang, J-Y. (2012). Effects of *Clinacanthus nutans* (Burm. f) Lindau Leaf Extracts on Protection of Plasmid DNA from Riboflavin Photoreaction. *MC-Transaction on Biotechnology*, 4(1), p. e5.
- Yuasa, T., Ono, M., Watanabe, T. & Takai, T. (2001). Lyn is Essential for Fc γ receptor III-mediated Systemic Anaphylaxis but not for the Arthus reaction. *Journal of Experimental Medicine*, 193(5), pp. 563–571.
- Zhang, K., Zhang, L., Zhu, D., Bae, D., Nel, A. & Saxon, A. (2002). CD40-mediated p38 Mitogen-activated Protein Kinase Activation is Required for Immunoglobulin Class Switch Recombination to IgE. *Journal of Allergy and Clinical Immunology*, 110(3), pp. 421-428.
- Zulkipli, I. N., Rajabalaya, R., Idris, A., Sulaiman, N. A. & David, S. R. (2017). *Clinacanthus nutans*: A review on Ethnomedicinal Uses, Chemical Constituents and Pharmacological Properties. *Pharmaceutical Biology*, 55(1), pp. 1093–1113.