



UNIVERSITI PUTRA MALAYSIA

**DECIPHERING THE PHARMACOLOGICAL EFFECTS OF  
ANDROGRAPHOLIDE ON ERYTHROCYTES AND  
*Plasmodium falciparum* 3D7 VIA METABOLIC CHANGES BY THE  
 $^1\text{H}$  NMR-BASED METABOLOMICS APPROACH**

**ASHRAF AHMAD ISSA ALAPID**

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**ASHRAF AHMAD ISSA ALAPID**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**March 2021**

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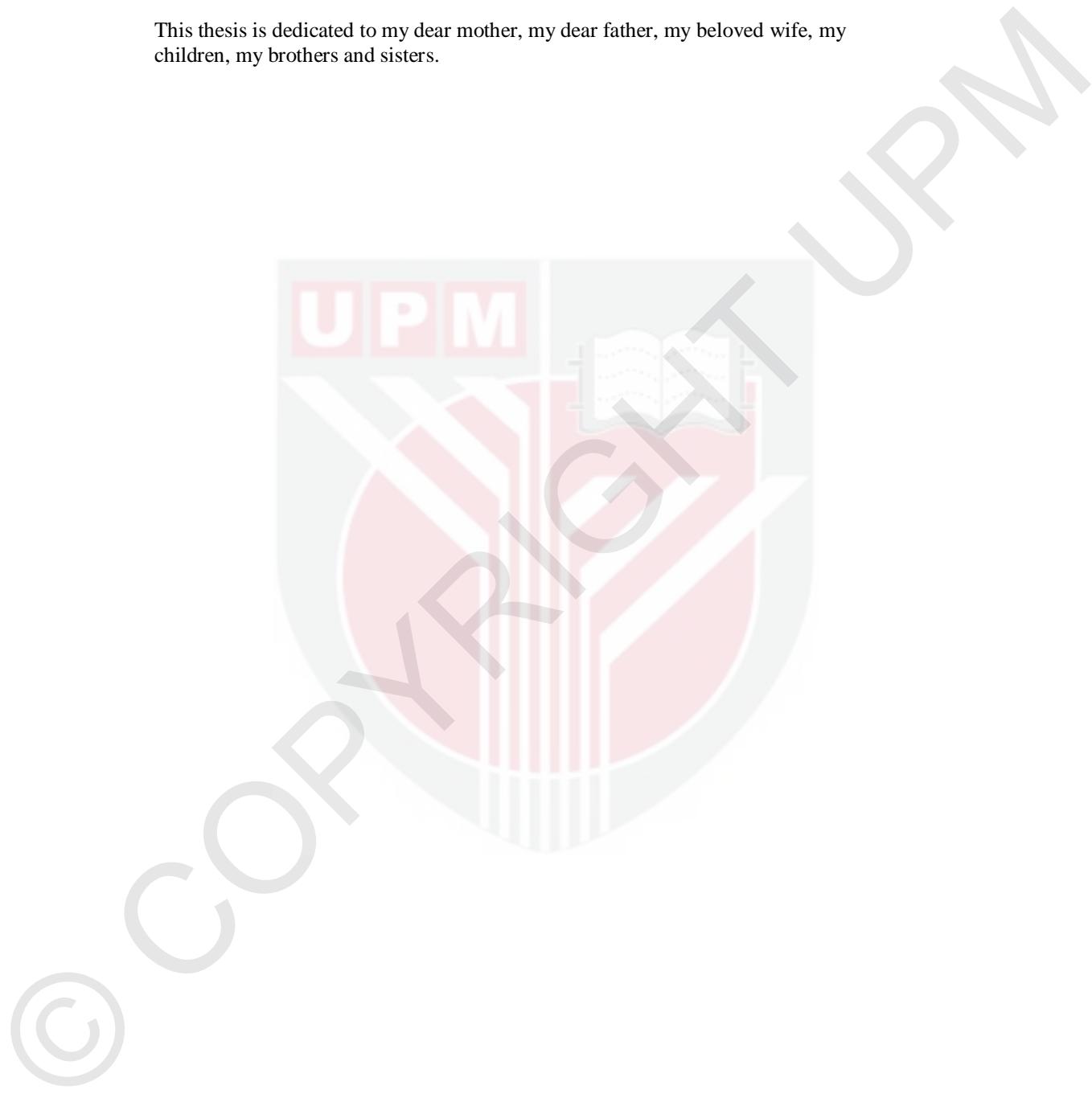
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## **DEDICATION**

This thesis is dedicated to my dear mother, my dear father, my beloved wife, my children, my brothers and sisters.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of  
the requirement for the degree of Doctor of Philosophy

**DECIPHERING THE PHARMACOLOGICAL EFFECTS OF  
ANDROGRAPHOLIDE ON ERYTHROCYTES AND *Plasmodium falciparum*  
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METABOLOMICS APPROACH**

By

**ASHRAF AHMAD ISSA ALAPID**

**March 2021**

**Chairman** : Associate Professor Rusliza Binti Basir, PhD  
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Malaria is a serious health problem associated with high morbidity and mortality rates, affecting millions of people across the world. The evolution of drug resistance among various strains of *Plasmodium falciparum* has thwarted the control efforts, thereby prompting scientists to seek for new and effective alternative therapeutic agents in order to forestall the menace caused by the parasite. This study was undertaken to evaluate and elucidate the pharmacological effects of andrographolide (AG) on *P. falciparum* 3D7 and erythrocytes using <sup>1</sup>H-NMR-based metabolomics approach. The first part of the study was aimed at investigating the anti-plasmodium effect of AG against *P. falciparum* 3D7, its time-dependent effect as well as its impact on the cellular morphology of various stages of plasmodium intra-erythrocytic cycle as compared to the conventional drug chloroquine (CQ). The malaria drug sensitivity assay was carried out using pLDH and Giemsa-stained thin blood smears to determine the differences and the morphological changes at different time intervals during the growth stages of the parasite.

In the second part of this study, the IC<sub>50</sub> and time-dependent of AG and CQ were used to determine the pharmacological effects of AG and CQ on the metabolic change of uninfected erythrocytes (uRBCs), infected erythrocytes (iRBCs) and the *P. falciparum* 3D7 parasite *in vitro*. The <sup>1</sup>H NMR-based metabolomics approach using Principal Component Analysis (PCA) and Orthogonal Partial Least Square discriminant analysis (OPLS-DA) were used. Overall, the results reveal that AG showed a good growth inhibitory effect (IC<sub>50</sub> = 4.14 μM) that was substantially lower than that of CQ (IC<sub>50</sub> = 20.19 nM). Unlike CQ, which showed its utmost activity within the first 12 hours of the cycle, the AG effect was more prominent during the second 12 hours interval of the cycle (early trophozoites stage). Although AG failed to produce any effect on the morphology of the ring stage, it produced a noticeable change in the morphological

appearance and the sizes of the mature trophozoites after 12 hours. In contrast, the rings and trophozoites stage of the parasites were fairly affected in the chloroquine-treated flasks within the first 12 hours and 24 hours of the cycle, respectively.

Based on unsupervised data analysis PCA, the effects of AG and CQ on the metabolic changes of uRBCs showed a clear separation between all uRBCs samples with a total variance of 89.10%. A total of 28 and 32 metabolites were identified as biomarkers in uRBCs-AG and uRBCs-CQ, respectively. In uRBCs-AG, ten metabolic pathways were determined as disturbed metabolic pathways, including riboflavin metabolic pathway, D-Glutamine and D-glutamate metabolism, phenylalanine metabolism, arginine and proline metabolism, glutathione metabolism, arginine biosynthesis, citrate cycle, pyruvate metabolism, alanine, aspartate and glutamate metabolism and glycolysis/gluconeogenesis. In contrast, in uRBCs-CQ, nine metabolic pathways have been determined as disturbed metabolic pathways similar to uRBCs-AG except for glutathione metabolism. These findings suggest an evident relationship between AG and CQ associated with metabolic perturbations in uRBCs.

The effects of AG and CQ on the metabolic changes of iRBCs, the PCA and OPLS-DA showed ideal differentiation between iRBCs samples treated and untreated. Thirty-five blood metabolites were identified from the <sup>1</sup>H-NMR spectra analysis of iRBCs samples. The outcome of PCA showed clear discrimination between AG and CQ. Both PC1 and PC2 show a total variance of 77.10%. A total of 23 and 24 metabolites were identified as biomarkers in iRBCs-AG and iRBCs-CQ, respectively. The metabolic pathways analysis revealed ten metabolic pathways were identified as disturbed in all groups. The iRBCs untreated group had a high number of disturbed metabolic pathways, including alanine, aspartate and glutamate metabolism, glutathione metabolism, arginine and proline metabolism, and riboflavin metabolism. In the group of iRBCs-CQ, the disturbed metabolic pathways identified as alanine, aspartate and glutamate metabolism, arginine and proline metabolism, and glutathione metabolism. Whereas in the iRBCs-AG, the disturbed metabolic pathways identified include glyoxylate and dicarboxylate metabolism, glycine, serine and threonine metabolism, and histidine metabolism.

The effects of AG and CQ on the metabolic changes of *P. falciparum* 3D7 *in-vitro* were identified. The results of multivariate data analysis show a clear discriminant between *P. falciparum* 3D7 samples treated and untreated. The model showed a total variance of 89.9% described by the PC1 and PC2. A total of 19 and 21 metabolites were identified as biomarkers in groups of *P. falciparum* 3D7-AG and *P. falciparum* 3D7- CQ, respectively. In *P. falciparum* 3D7-AG, very few metabolites biomarkers were observed, including threonine, ornithine, riboflavin, lactate and glutathione, compared to the group treated with CQ, which showed a high number of biomarkers. Analysis of the metabolic pathways reveals two metabolic pathways were significantly disturbed in *P. falciparum* 3D7-AG group; arginine and proline metabolism, and glutathione metabolism. In *P. falciparum* 3D7-CQ group, six disturbed metabolic pathways were identified: glyoxylate and dicarboxylate metabolism, glutathione metabolism, alanine, aspartate and glutamate metabolism, arginine biosynthesis, purine metabolism and citrate cycle.

In conclusion, the present study is the first to report on the antimalarial activity of AG utilizing the  $^1\text{H}$  NMR-based metabolomics approach. Results from this study suggest that the disturbed metabolic pathways identified could well serve as drug targets for future development of andrographolide-based therapeutic agents against *P. falciparum* 3D7.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai  
memenuhi keperluan untuk ijazah Doktor Falsafah

**MENGHURAIKAN KESAN FARMAKOLOGI ANDROGRAPHOLIDE KE  
ATAS ERITROSIT DAN *Plasmodium falciparum* 3D7 MELALUI PERUBAHAN  
METABOLIK MENGGUNAKAN PENDEKATAN METABOLOMIK  
BERASASKAN  $^1\text{H-NMR}$**

Oleh

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Malaria adalah masalah kesihatan yang serius yang dikaitkan dengan kadar mortaliti dan morbiditi yang tinggi melibatkan jutaan manusia di seluruh dunia. Evolusi kerintangan drug di kalangan pelbagai jenis *Plasmodium falciparum* telah membantutkan usaha mengawal, justeru mendesak para saintis mencari agen terapeutik baru dan berkesan bagi mengekang ancaman parasit ini. Kajian ini dijalankan bagi menilai dan menghuraikan kesan farmakologi andrographolide ke atas *P. falciparum* 3D7 dan eritrosit menggunakan pendekatan metabolomik berasaskan  $^1\text{H-NMR}$ . Bahagian pertama kajian ini bertujuan menyelidiki kesan anti-plasmodium AG terhadap *P. falciparum* 3D7, kesan kebergantungan masa dan juga impaknya ke atas morfologi sel pelbagai peringkat kitaran intra-eritrositik plasmodium berbanding drug chloroquine (CQ) konvensional. Asai sensitiviti drug malaria dilakukan menggunakan pLDH dan calitan darah nipis berlumur Giemsa bagi menentukan perbezaan dan perubahan morfologi pada selang masa berbeza semasa peringkat pertumbuhan parasit.

Dalam bahagian kedua kajian ini,  $\text{IC}_{50}$  dan kebergantungan masa AG dan CQ digunakan bagi menentukan kesan farmakologi AG dan CQ ke atas perubahan metabolismik eritrosit yang tidak terjangkit (uRBCs), eritrosit terjangkit (IRBCs) dan parasit *P. falciparum* 3D7 *in vitro*. Pendekatan metabolomik berasaskan  $^1\text{H-NMR}$  mengguna pakai “Principal Component Analysis (PCA)” dan “Orthogonal Partial Least Square discriminant analysis (OPLS-DA)” telah digunakan. Secara keseluruhannya, keputusan menunjukkan bahawa AG memberikan kesan perencutan pertumbuhan yang baik ( $\text{IC}_{50} = 4.14 \mu\text{M}$ ), yang lebih rendah daripada CQ ( $\text{IC}_{50} = 20.19 \text{ nM}$ ). Tidak seperti CQ yang menunjukkan aktiviti paling tinggi dalam masa 12 jam pertama kitaran, kesan AG lebih ketara semasa selang 12 jam kedua kitaran (peringkat trofozoit awal). Walau pun AG gagal menghasilkan sebarang kesan ke atas morfologi peringkat gegelang semasa 12 jam pertama, ia menghasilkan perubahan yang nyata

dalam rupa bentuk morfologi dan saiz trofozoit matang selepas 12 jam. Sebaliknya, peringkat gegelang dan trofozoit parasit masing-masing hanya sedikit terkesan di dalam kelalang terawat CQ dalam masa 12 dan 24 jam pertama kitaran.

Berdasarkan data analisis PCA tanpa pengawasan, kesan AG dan CQ ke atas perubahan metabolismik uRBCs menunjukkan pemisahan yang ketara di antara kesemua sampel uRBCs dengan varians keseluruhan 89.10%. Sebanyak 28 dan 32 metabolit masing masing dikenalpasti sebagai biopenanda dalam uRBCs-AG dan uRBCs-CQ. Dalam uRBCs-AG, sepuluh laluan metabolismik telah ditentukan sebagai laluan metabolismik terganggu, termasuk laluan metabolismik riboflavin, metabolisme D-Glutamine and D-glutamate, metabolisme phenylalanine, metabolisme arginine and proline, metabolisme glutathione, biosintesis arginine, kitaran citrate, metabolisme pyruvate, metabolisme alanine, aspartate and glutamate, and glycolysis/gluconeogenesis. Sebaliknya, dalam uRBCs-CQ, sembilan laluan metabolismik telah ditentukan sebagai laluan metabolismik terganggu menyamai uRBCs-AQ kecuali metabolisme glutathione. Dapatkan ini mencadangkan hubungan jelas antara AG dan CQ yang berkait dengan gangguan metabolismik dalam uRBCs.

Kesan AG dan CQ ke atas perubahan metabolismik iRBCs, PCA dan OPLS-DA menunjukkan perbezaan yang ideal antara sample iRBCs terawat dan tidak terawat. Tiga puluh lima metabolit darah dikenal pasti dari analisis spektra  $^1\text{H-NMR}$  sampel iRBCs. Keputusan PCA menunjukkan diskriminasi yang jelas antara AG dan CQ. Kedua dua PC1 dan PC2 menunjukkan varians keseluruhan 77.10%. Sejumlah 23 dan 24 metabolit masing masing dikenal pasti sebagai biopenanda dalam iRBCs-AG dan iRBCs-CQ. Analisis laluan metabolismik mendedahkan sepuluh laluan metabolismik dikenal pasti sebagai terganggu dalam kesemua kumpulan. Kumpulan iRBCs tidak terawat mempunyai bilangan laluan metabolismik yang tinggi, termasuk metabolisme alanine, aspartate, glutamate, metabolisme glutathione, metabolisme arginine and proline, and metabolisme riboflavin. Dalam kumpulan iRBCs-CQ, laluan metabolismik terganggu yang dikenal pasti termasuk metabolisme alanine, aspartate and glutamate, metabolisme arginine and proline, and metabolisme glutathione. Sementara dalam iRBCs-AG, laluan metabolismik terganggu yang dikenal pasti termasuk metabolisme glyoxylate and dicarboxylate, metabolisme glycine, serine, threonine, and metabolisme histidine.

Kesan AG dan CQ ke atas perubahan metabolismik *P. falciparum* *in vitro* telah dikenalpasti. Keputusan analisa data multivariat menunjukkan perbezaan jelas antara sampel *P. falciparum* 3D7 terawat dan tidak terawat. Model menunjukkan varians keseluruhan 89.9% diterangkan oleh PC1 dan PC2. Sejumlah 19 dan 21 metabolit masing masing dikenal pasti sebagai biopenanda dalam kumpulan *P. falciparum* 3D7-AG dan *P. falciparum* 3D7-CQ. Dalam *P. falciparum* 3D7-AG, hanya sedikit biopenanda metabolismik diperhatikan termasuk threonine, ornithine, riboflavin, lactate dan glutathione, berbanding kumpulan yang dirawat dengan CQ, yang menunjukkan jumlah biopenanda yang tinggi. Analisa laluan metabolismik mendedahkan dua laluan metabolismik telah terganggu secara signifikan dalam kumpulan *P. falciparum* 3D7-AG; metabolisme arginine and proline and metabolisme glutathione. Dalam kumpulan *P. falciparum* 3D7-CQ, enam laluan metabolismik terganggu dikenal pasti sebagai metabolisme glyoxylate and dicarboxylate, metabolisme glutathione, metabolisme

alanine, aspartate dan glutamate, biosintesis arginine, metabolisme purine and kitaran citrate.

Kesimpulannya, kajian ini adalah yang pertama melaporkan tentang aktiviti antimalaria AG menggunakan pendekatan metabolik berasaskan  $^1\text{H}$  NMR. Keputusan dari kajian ini mencadangkan bahawa laluan metabolismik terganggu ini boleh menjadi sasaran drug bagi pengembangan agen terapeutik berasaskan andrographolide terhadap *P. falciparum* 3D7 di masa hadapan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## LIST OF SYMBOLS AND ABBREVIATIONS

|                    |  |
|--------------------|--|
| %PI                | Percentage of parasite inhibition              |
| ~                  | Approximately                                  |
| <                  | Less than                                      |
| =                  | Equals   |
| >                  | Greater than                                   |
| ±                  | Plus-minus sign                                |
| 1D                 | One-Dimensional                                |
| <sup>1</sup> H-NMR | Proton Nuclear Magnetic Resonance Spectroscopy |
| 2D                 | Two-Dimensional                                |
| A.A                | Amine acids                                    |
| Abs                | Absorbance                                     |
| AG                 | Andrographolide                                |
| ANOVA              | Analysis Of Variance                           |
| APAD               | 3-acetylpyridine adenine dinucleotide          |
| cMCM               | Complete Malaria Culture Medium                |
| CO <sub>2</sub>    | Carbon dioxide                                 |
| CPD                | Citrate Phosphate Dextrose                     |
| CQ                 | Chloroquine                                    |
| <i>d</i>           | Doublet  |
| D <sub>2</sub> O   | Deuterium Oxide                                |
| <i>dd</i>          | Doublet of doublet                             |
| DModx              | distance to the model in X-space               |
| DMSO               | Di-methylsulphoxide                            |

|                                 |  |
|---------------------------------|--|
| et al.,                         | alia: and others                                       |
| g                               | Gram   |
| g.m.wt                          | Gram Molecular Weight                                  |
| H <sub>2</sub> O <sub>2</sub>   | Hydrogen peroxide                                      |
| Hb                              | Haemoglobin  |
| HCA                             | Hierarchical Cluster Analysis                          |
| HCL                             | Hydrochloric acid                                      |
| Hct                             | Hemeatocrit  |
| HEPES                           | (4-(2-hydroxyethyl)-1-piperazine-ethan-sulphonic acid) |
| HMDB                            | Human metabolome database                              |
| Hz                              | Hertz  |
| i.e,                            | That is  |
| IC <sub>20</sub>                | inhibitory concentration at 20% of maximal growth      |
| IC <sub>50</sub>                | inhibitory concentration at 50% of maximal growth      |
| IC <sub>90</sub>                | inhibitory concentration at 90% of maximal growth      |
| iMCM                            | incomplete Malaria Culture Medium                      |
| iRBCs                           | infected red blood cells                               |
| <i>J</i>                        | Coupling constant in Hz                                |
| KCL                             | Potassium chloride                                     |
| KH <sub>2</sub> PO <sub>4</sub> | Dihydrogen phosphate                                   |
| L                               | Liter  |
| <i>m</i>                        | Multiplet  |
| MHz                             | MegaHertz  |
| Min                             | Minute   |

|                                  |  |
|----------------------------------|--|
| mL                               | Milliliter   |
| Mm                               | Milli Molar  |
| MVDA                             | multivariate data analysis                             |
| Na <sub>2</sub> HPO <sub>4</sub> | Disodium Hydrogen Phosphate                            |
| NaCl                             | Sodium Chloride  |
| NADPH                            | Nicotinamide adenine dinucleotide                      |
| NADPH                            | Nicotinamide adenine dinucleotide                      |
| NaOD                             | Sodium Deuterium Oxide                                 |
| NaOH                             | Sodium hydroxide                                       |
| NBT                              | Nitroblue tetrazolium chloride                         |
| nM                               | Nano Molar   |
| °C                               | Degree in Celsius                                      |
| P                                | Statistical significance level                         |
| P%                               | Parasitaemia percentage                                |
| PBS                              | Phosphate buffer saline                                |
| pLDH                             | Plasmodial lactic dehydrogenase enzyme                 |
| PC                               | Principal component                                    |
| PCA                              | Principal component analysis                           |
| PLS                              | Partial Least Squares                                  |
| OPLS-DA                          | Orthogonal Partial Least Squares–Discriminant Analysis |
| ppm                              | Part Per Million                                       |
| pRBCs                            | Parasitized red blood cells                            |
| <i>P. falciparum</i>             | <i>Plasmodium falciparum</i>                           |
| <i>q</i>                         | quartet  |

|               |  |
|---------------|--|
| $R^2$         | Coefficient of determination of linear regression              |
| RBCs          | Red Blood Cells  |
| RMSEE         | Root Mean Square Error of Estimation                           |
| RMSEP         | Root Mean Square Error of Prediction                           |
| rpm           | Revolution per minute  |
| $s$           | Singlet  |
| SEM           | standard error of the mean                                     |
| SIMCA         | Soft Independent Modeling of Class Analogy                     |
| TSP           | trimethylsilyl-2,2,3,3-tetradeuteropropionate acid sodium salt |
| uRBCs         | Uninfected red blood cells                                     |
| uRBCs-AG      | Uninfected red blood cells expose to Andrographolide           |
| uRBCs-CQ      | Uninfected red blood cells expose to chloroquine               |
| VIP           | variable Importance in the Projection                          |
| VP            | Viability of parasite  |
| Vs            | Versus   |
| WHO           | World Health Organization                                      |
| X             | Times  |
| $\delta$      | Chemical Shift in ppm  |
| $\mu\text{L}$ | Microliter   |

## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background**

For centuries, malaria has been one of the major devastating health issues to humanity. According to the World Health Organization (WHO) report in 2018, 228 million people were estimated to be suffering from the disease globally, causing about 405,000 deaths per annum (WHO, 2019). The majority of the recorded cases were from the African region with 93%, South East Asia with 3.4% and Eastern Mediterranean region had about 2.1% of the cases (WHO, 2019).

The partial success achieved by the eradication programs resulted in a sharp spatial concentration of malaria in the tropical areas. However, the number of malaria cases globally fell from an estimated 400 million in 2010 to 260 million in 2018. This translates to a decline of 18% globally, with the largest decline observed in the South-East Asia region, which had 50%, while the least decline was recorded in the African region (20%). Nevertheless, malaria remains a major killer of the world population, particularly in Sub-Saharan Africa (WHO, 2019).

Malaria is a life-threatening disease caused by one or a combination of five species of *Plasmodium* genus of the protozoan parasites. The *Plasmodium* parasite is transmitted to humans by infected female Anopheles mosquitoes, while taking a blood meal. The genus *Plasmodium* comprises of five species; *Plasmodium falciparum*, *Plasmodium ovale*, *Plasmodium vivax*, *Plasmodium malariae* and *Plasmodium knowlesi* (Cox-singh & Singh, 2008; Garnham 1966). *P. falciparum* is responsible for 99.7% of disease related deaths, especially in Sub-Saharan Africa (WHO, 2018).

#### **1.2 Problem statement**

Malaria is still one of the major public health concerns and persistent diseases worldwide. It is responsible for life-threatening infections in most endemic areas of the world. The Sub - Saharan Africa region carries a disproportionately more significant proportion of the global malaria burden (WHO, 2020). Ironically, nearly half of the world's population lives in endemic malaria areas (WHO, 2019). One of the significant problems of malaria infection is the development of drug resistance in malaria parasites that have become widespread, coupled with the limited number of effective antimalarial drugs available. Hence, disease management faces increasing difficulties to control or eradicate the disease (Mishra et al., 2016).

Resistance to the commonly used drugs such as chloroquine (CQ) against the *P. falciparum* strains has been reported in the past (Wellems & Plowe, 2001). Recently, there is an increasing report of the development of resistance against the recommended first line antimalarial treatment including the artemisinin-based combination therapies (ACTs) at the Thai-Cambodian border (Dondorp et al., 2009; Ashley et al., 2014) and Thai-Myanmar border with Vietnam (Miotto et al., 2013). Similarly, in recent years, there has been an increasing interest in efforts to eradicate the disease through vaccine development, avoidance measures and drug therapies.

Undoubtedly, these efforts led to the fractional success recorded by the eradication programs, which has also resulted in the spatial concentration of malaria cases in the resource poor tropical areas (WHO, 2017). Nevertheless, malaria remains a major killer of the world population and the widespread drug-resistant strains of *Plasmodium* are continuously evolving (WHO, 2019). It has, therefore, become necessary to understand, identify and develop new antimalarial drugs with a novel mechanism of actions against these strains.

### **1.3 Significance of the study**

In most of the Asian, African and South American countries, approximately 90% of the people rely on traditional or alternative medicines that are based largely on different plants as a source of medicines (Fabricant & Farnsworth, 2001; Verpoorte et al., 2006; Appendino et al., 2010). One of the most commonly used medicinal plant is *Andrographis paniculata*. The plant is very popular among traditional medicine practitioners in China and other South East Asian countries (Kuroyanagi et al., 1987). The phytochemical analysis of *A. paniculata* reveals more than 55 ent-labdane diterpenoids, 30 flavonoids, 8 quinic acids, 4 xanthones, and 5 rare noriridoids (Hossain et al., 2014). The extracts from the aerial parts of the *A. paniculata* plant equally has many diterpenoids such as Andrographolide (AG), neoAndrographolide and dehydroAndrographolide.

Among these, the AG is considered one of the most essential bioactive compounds. It is a labdane diterpenoid derivative and it possesses plenty of medicinal and pharmaceutical properties, including antimicrobial, anti-inflammatory and antioxidant effects (Wasman et al., 2011). In addition, AG also possesses cardio-protective, hepato-protective, anti-HIV, anti-carcinogenic, anti-diabetic and anti-trypanosomal as well as antimalarial activity (Banerjee et al., 2017; Mishra et al., 2011; Hossain et al., 2014; Kandanur et al., 2019; Mishra et al., 2009; Misra et al., 1992; Niranjan et al., 2010; Sheeja & Kuttan, 2007; Sheeja et al., 2006; Widjyawaruyanti et al., 2017; Yoopan et al., 2007; Zaid et al., 2015; Zaridah et al., 2001). Despite the extensive properties of AG, studies exploring the potentials and mechanisms of action of the AG as an anti-malaria are still scarce. Hence, their activity on uninfected red blood cells (uninfected RBCs), infected red blood cells (infected RBCs) and *P. falciparum* 3D7 as antimalarial agent has not been investigated to a great extent. Therefore, more studies need to be conducted considering the determination of the mechanism of action of this

compound will certainly highlight a pathway for the development of a new and improved antimalarial drug with a novel mechanism of actions.

#### **1.4 Research hypothesis**

The chemotherapeutic impact of plenty of drugs depends on their ability to compromise parasite growth without having affected the host cells selectively. Besides, it might alter the ability of the parasite to resist the conventional antimalarials. Andrographolide is considered one of the essential phytochemical compounds in the *A. paniculata* plant. It is a labdane diterpenoid derivative and possesses plenty of medicinal and pharmaceutical properties including an antimalarial activity. Nonetheless, studies exploring the potentials and mechanisms of action of the AG and their activity on uninfected RBCs, infected RBCs and *P. falciparum* 3D7 as an antimalarial agent has not been extensively investigated. Hence, this research is based on the hypothesis that AG might have significant biological activity on the metabolic pathways of uninfected RBCs, infected RBCs with *Plasmodium* parasite and the *P. falciparum* 3D7.

#### **1.5 Objective**

##### **1.5.1 General objective**

The main objective of the study was to identify the effect of Andrographolide compound (AG) on the metabolic pathway of the uninfected RBCs, infected RBCs and the *P. falciparum* 3D7 by <sup>1</sup>H-NMR metabolomics approach.

##### **1.5.2 Specific objectives**

1. To determine the efficacy of AG and CQ ( $IC_{50}$ ) and the time dependency of antimalarial action on the intraerythrocytic cycle progression and morphology changes of *P. falciparum* 3D7 at a different time interval.
2. The determination of the expected metabolic derangement on uninfected RBCs following exposure to (AG) and (CQ) by <sup>1</sup>H-NMR based metabolomics approach.
3. The discovery of the expected metabolic derangement on infected RBCs following exposure to (AG) and (CQ) by <sup>1</sup>H-NMR based metabolomics approach.
4. To identify the biomarker(s) of the antimalarial activity of AG and CQ on *P. falciparum* 3D7 by <sup>1</sup>H-NMR based metabolomics approach.

## **1.6 Outline of the thesis**

The purpose of this study was to evaluate the pharmacological effects of AG on infected RBCs, uninfected RBCs and *P. falciparum* 3D7 using the  $^1\text{H}$  NMR-based metabolomics approach. The research is presented in a thesis report, which comprises of several chapters that provide and elaborate the details of the current study. Figure 1.1 exhibits the outline of the thesis chapters. Henceforth, a brief explanation of the specific objectives of the study is presented.

At the initial stage of the study, the efficacy  $\text{IC}_{50}$  (Inhibitory concentration to reduce the growth by 50%) of AG and CQ were determined. In addition, the time-dependency of antimalarial action on the intraerythrocytic cycle progression and morphological changes of *P. falciparum* 3D7 at the different time interval 12, 24 and 48 hours were determined (objective one). Then, the  $\text{IC}_{50}$  of AG and its time-action as well as  $\text{IC}_{50}$  of CQ and its time-action were used as a standard to achieving the other objectives. Subsequently, the metabolic variations on uninfected RBCs were determined using  $^1\text{H}$  NMR-based metabolomics approach after exposure to (AG) and (CQ) (objective two). Next, the metabolic variations on infected RBCs were determined using  $^1\text{H-NMR}$  after the exposure to (AG) and (CQ) (objective three). Finally, using the  $^1\text{H}$  NMR-based metabolomics approach a several biomarker(s) and disturbed metabolic pathways were identified on *P. falciparum* 3D7 after exposure to AG and CQ (objective four).

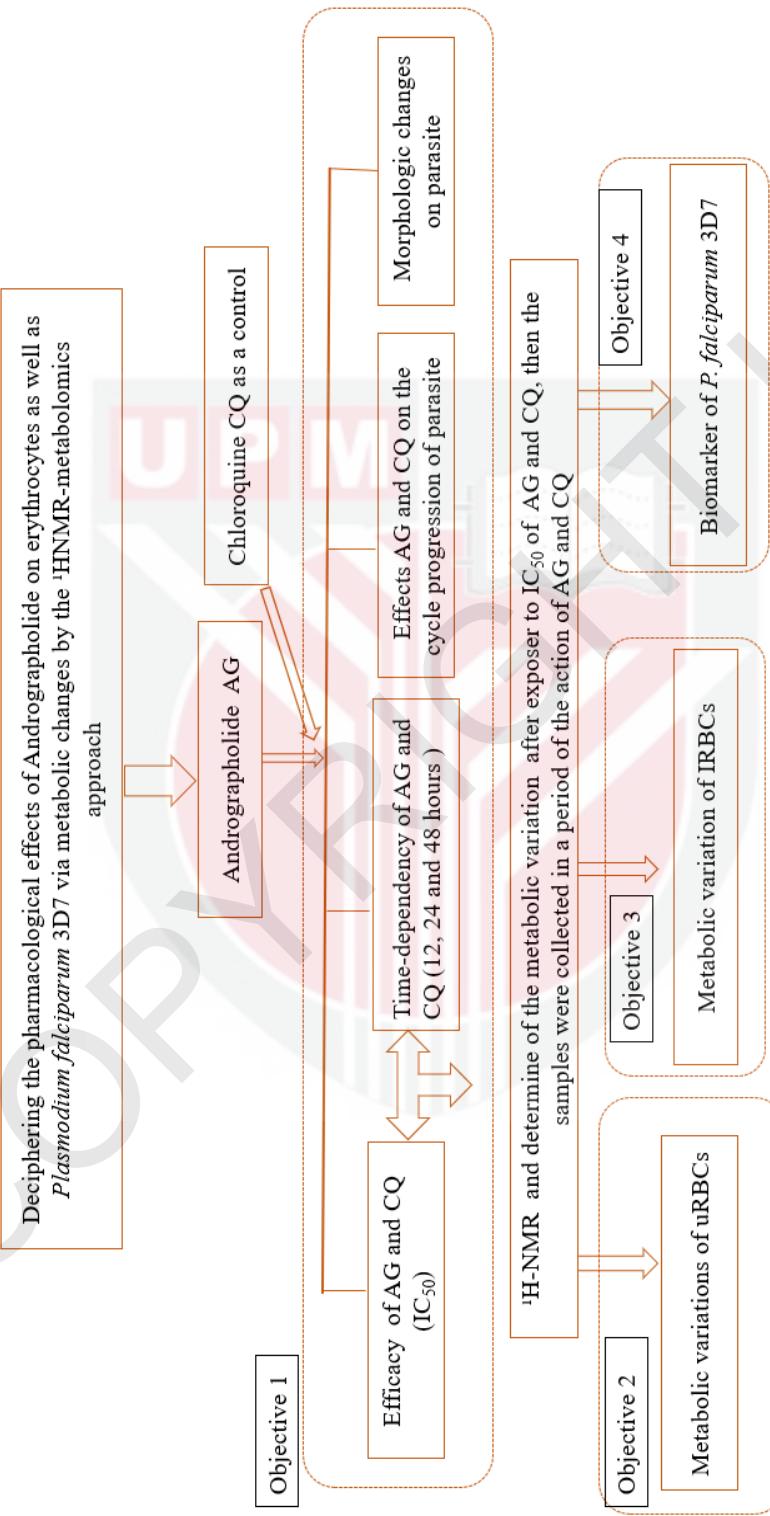


Figure 1.1 : Outline of the thesis objectives

## REFERENCES

- Akbar, S. (2011). *Andrographis paniculata*: a review of pharmacological activities and clinical effects. *Alternative Medicine Review*, 16(1), 66–77.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2002). *How cells obtain energy from food*. In *Molecular Biology of the Cell* (4th editio). Garland Science;<https://www.ncbi.nlm.nih.gov/books/NBK26882/>.
- Allen, R. J. W., & Kirk, K. (2004). The membrane potential of the intraerythrocytic malaria parasite *Plasmodium falciparum*. *Journal of Biological Chemistry*, 279(12), 11264–11272.
- Aminake, M., & Pradel, G. (2013). Antimalarial drugs resistance in *Plasmodium falciparum* and the current strategies to overcome them. *Microbial Pathogens and Strategies for Combating Them: Science, Technology and Education*, 1, 269–282.
- Antony, H. A., & Parija, S. C. (2016). Antimalarial drug resistance: An overview. *Tropical parasitology*, 6(1), 30.
- An, L., Shi, Q., & Feng, F. (2015). Metabolomics approach to identify therapeutically potential biomarkers of the Zhi-Zi-Da-Huang decoction effect on the hepatoprotective mechanism. *RSC Advances*, 5(102), 84048–84055. <https://doi.org/10.1039/c5ra16563f>
- Arnér, E. S. J., & Holmgren, A. (2000). Physiological functions of thioredoxin and thioredoxin reductase. *European Journal of Biochemistry*, 267(20), 6102–6109.
- Asahi, H., & Kanazawa, T. (1994). Continuous cultivation of intraerythrocytic *Plasmodium falciparum* in a serum-free medium with the use of a growth-promoting factor. *Parasitology*, 109(4), 397–401.
- Asahi, H., Kanazawa, T., Kajihara, Y., Takahashi, K., & Takahashi, T. (1996). Hypoxanthine: a low molecular weight factor essential for growth of erythrocytic *Plasmodium falciparum* in a serum-free medium. *Parasitology*, 113(1), 19–23.
- Ashley, E. A., Dhorda, M., Fairhurst, R. M., Amaratunga, C., Lim, P., Suon, S., ... Sam, B. (2014). Spread of artemisinin resistance in *Plasmodium falciparum* malaria. *New England Journal of Medicine*, 371(5), 411–423.
- Autino, B., Corbett, Y., Castelli, F., & Taramelli, D. (2012a). Pathogenesis of malaria in tissues and blood. *Mediterranean Journal of Hematology and Infectious Diseases*, 4(1). <https://doi.org/10.4084/MJHID.2012.061>

- Autino, B., Noris, A., Russo, R., & Castelli, F. (2012b). Epidemiology of malaria in endemic areas. *Mediterranean Journal of Hematology and Infectious Diseases*, 4(1). <https://doi.org/10.4084/MJHID.2012.060>
- Appendino, G., & Pollastro, F. (2010). Plants: revamping the oldest source of medicines with modern science. *Natural Product Chemistry for Drug Discovery*, 140-173.
- Avital Schurr (2017). Lactate, Not Pyruvate, Is the End Product of Glucose Metabolism via Glycolysis. In tech , 21–35.
- Avner Yayon, John A. Vande Waa, Malka Yayon, Timothy G. Geary, A. J. B. J. (1983). Stage-Dependent Effects of Chloroquine on *Plasmodium falciparum* in vitro. *J.Porozool.*, 30(4), 642–647.
- Banerjee, M., Chattopadhyay, S., Choudhuri, T., Bera, R., Kumar, S., Chakraborty, B., & Mukherjee, S. K. (2016). Cytotoxicity and cell cycle arrest induced by Andrographolide lead to programmed cell death of MDA-MB-231 breast cancer cell line. *Journal of Biomedical Science*, 23, 40. <https://doi.org/10.1186/s12929-016-0257-0>
- Banerjee, M., Parai, D., Dhar, P., Roy, M., Barik, R., Chattopadhyay, S., & Mukherjee, S. K. (2017). Andrographolide induces oxidative stress-dependent cell death in unicellular protozoan parasite *Trypanosoma brucei*. *Acta Tropica*, 176, 58–67. <https://doi.org/10.1016/j.actatropica.2017.07.023>
- Banerjee, R., Liu, J., Beatty, W., Pelosof, L., Klemba, M., & Goldberg, D. E. (2002). Four plasmepsins are active in the *Plasmodium falciparum* food vacuole, including a protease with an active-site histidine. *Proceedings of the National Academy of Sciences*, 99(2), 990-995.
- Barker, M., & Rayens, W. (2003). Partial least squares for discrimination. *Journal of Chemometrics: A Journal of the Chemometrics Society*, 17(3), 166–173.
- Barrand, M. A., Winterberg, M., Ng, F., Nguyen, M., Kirk, K., & Hladky, S. B. (2012). *Glutathione export from human erythrocytes and Plasmodium falciparum malaria parasites*. 400, 389–400. <https://doi.org/10.1042/BJ20121050>
- Barsoum, R. S. (2000). Malarial acute renal failure. *Journal of the American Society of Nephrology*, 11(11), 2147–2154.
- Bartoloni, A., & Zammarchi, L. (2012). Clinical aspects of uncomplicated and severe malaria. *Mediterranean Journal of Hematology and Infectious Diseases*, 4(1). <https://doi.org/10.4084/MJHID.2012.026>
- Basir, R., Rahiman, S. S. F., Hasballah, K., Chong, W. C., Talib, H., Yam, M. F., ... Moklas, M. A. M. (2012). *Plasmodium berghei* ANKA infection in ICR mice as a model of cerebral malaria. *Iranian Journal of Parasitology*, 7(4), 62.

- Becker, K., Rahlfs, S., Nickel, C., & Schirmer, R. H. (2003). Glutathione–functions and metabolism in the malarial parasite *Plasmodium falciparum*. *Biological Chemistry*, 384(4), 551–566.
- Beckonert, O., Keun, H. C., Ebbels, T. M. D., Bundy, J. G., Holmes, E., Lindon, J. C., & Nicholson, J. K. (2007). Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts. *Nature Protocols*, 2, 2692–2703. <https://doi.org/10.1038/nprot.2007.376>
- Belen Cassera, M., Zhang, Y., Z. Hazleton, K., & L. Schramm, V. (2011). Purine and Pyrimidine Pathways as Targets in *Plasmodium falciparum*. *Current Topics in Medicinal Chemistry*, 11(16), 2103–2115. <https://doi.org/10.2174/156802611796575948>
- Beri, D., Ramdani, G., Balan, B., Gadara, D., & Poojary, M. (2019). Insights into physiological roles of unique metabolites released from *Plasmodium* -infected RBCs and their potential as clinical biomarkers for malaria. *Scientific Reports*, 9(1), 1–11. <https://doi.org/10.1038/s41598-018-37816-9>
- Bianconi, E., Piovesan, A., Facchin, F., Beraudi, A., Casadei, R., Frabetti, F., ... Piva, F. (2013). An estimation of the number of cells in the human body. *Annals of Human Biology*, 40(6), 463–471.
- Blanco, A., & Blanco, G. (2017a). Amino Acid Metabolism, Medical Biochemistry. In *Jaypee Brothers Medical Publishers (P) Ltd* (pp. 367–399). [https://doi.org/https://doi.org/10.5005/jp/books/11450\\_11](https://doi.org/https://doi.org/10.5005/jp/books/11450_11)
- Blanco, A., & Blanco, G. (2017b). Carbohydrate Metabolism. In *Medical Biochemistry* In *Jaypee Brothers Medical Publishers (P) Ltd* (pp. 283–323). <https://doi.org/10.1016/B978-0-12-803550-4/00014-8>
- Booden, T., & Hull, R. W. (1973). Nucleic acid precursor synthesis by *Plasmodium lophurae* parasitizing chicken erythrocytes. *Experimental parasitology*, 34(2), 220–228.
- Bordbar, A., Jamshidi, N., & Palsson, B. O. (2011). iAB-RBC-283: A proteomically derived knowledge-base of erythrocyte metabolism that can be used to simulate its physiological and patho-physiological states. *BMC Systems Biology*, 5(1), 110.
- Brown, B. A., Hunter, R. C., O'Hare, A., & Erim, G. (1993). *Hematology: principles and procedures*. Lea & Febiger Philadelphia. pp 1-453.
- Bruce-chwatt. (1962). Classification of antimalarial drugs in relation to different stages in the life-cycle of the parasite: commentary on a diagram. *Bulletin of the World Health Organization*, 27(27), 287–290.
- Brunet, L. R. (2001). Nitric oxide in parasitic infections. *International Immunopharmacology*, 1(8), 1457–1467.

- Cabrera, M., Paguio, M. F., Xie, C., & Roepe, P. D. (2009). Reduced digestive vacuolar accumulation of chloroquine is not linked to resistance to chloroquine toxicity. *Biochemistry*, 48(47), 11152–11154. <https://doi.org/10.1021/bi901765v>
- Calabrese, C., Berman, S. H., Babish, J. G., Ma, X., Shinto, L., Dorr, M., ... Standish, L. J. (2000). A phase I trial of Andrographolide in HIV positive patients and normal volunteers. *Phytotherapy Research*, 14(5), 333–338.
- Chaleckis, R., Murakami, I., Takada, J., Kondoh, H., & Yanagida, M. (2016). Individual variability in human blood metabolites identifies age-related differences. *Proceedings of the National Academy of Sciences*, 113(16), 4252–4259.
- Chao, W.-W., & Lin, B.-F. (2010). Isolation and identification of bioactive compounds in *Andrographis paniculata* (Chuanxinlian). *Chinese Medicine*, 5(1), 17.
- Chesney, R. W., & Budreau, A. M. (1995). Chloroquine, a novel inhibitor of amino acid transport by rat renal brush border membrane vesicles. *Amino Acids*, 8, 141–158.
- Chong, J., Wishart, D. S., & Xia, J. (2019). Using MetaboAnalyst 4.0 for Comprehensive and Integrative Metabolomics Data Analysis. *Current Protocols in Bioinformatics*, 68(1), e86.
- Clendinen, C. S., Stupp, G. S., Ajredini, R., Lee-McMullen, B., Beecher, C., & Edison, A. S. (2015). An overview of methods using <sup>13</sup>C for improved compound identification in metabolomics and natural products. *Frontiers in Plant Science*, 6, 611.
- Cobbold, S. A., Llinás, M., & Kirk, K. (2016). Sequestration and metabolism of host cell arginine by the intraerythrocytic malaria parasite *Plasmodium falciparum*. *Cellular Microbiology*, 18(6), 820–830. <https://doi.org/10.1111/cmi.12552>
- Combrinck, J. M., Mabotha, T. E., Ncokazi, K. K., Ambele, M. A., Taylor, D., Smith, P. J., ... Egan, T. J. (2012). Insights into the role of heme in the mechanism of action of antimalarials. *ACS Chemical Biology*, 8(1), 133–137.
- Cooper, R. G., & Magwere, T. (2008). Chloroquine: novel uses & manifestations. *Indian Journal of Medical Research*, 127(4).
- Cowman, A. F., Berry, D., & Baum, J. (2012). The cellular and molecular basis for malaria parasite invasion of the human red blood cell. *Journal of Cell Biology*, 198(6), 961–971.
- Cox-singh, J., & Singh, B. (2008). *Knowlesi* malaria : newly emergent and of public health importance? *Trends in Parasitology*, 24(9), 406–410. <https://doi.org/10.1016/j.pt.2008.06.001>. *Knowlesi*

- Cox, F. E. (2010). History of the discovery of the malaria parasites and their vectors. *Parasites & Vectors*, 3, 1, 1–9. <https://doi.org/10.1186/1756-3305-3-5>
- Cragg, G. M., & Newman, D. J. (2013). Natural Products: a Continuing Source of Novel. *Biochimica et Biophysica Acta*, 1830(6), 3670–3695. <https://doi.org/10.1016/j.bbagen.2013.02.008.NATURAL>
- Creek, D. J., & Barrett, M. P. (2014). Determination of antiprotozoal drug mechanisms by metabolomics approaches. *Parasitology*, 141(1), 83–92. <https://doi.org/10.1017/S0031182013000814>
- Creek, D. J., Chua, H. H., Cobbold, S. A., Nijagal, B., Macrae, J. I., Dickerman, B. K., ... McConville, M. J. (2016). Metabolomics-based screening of the Malaria Box reveals both novel and established mechanisms of action. *Antimicrobial Agents and Chemotherapy*, 60(11), 6650–6663. <https://doi.org/10.1128/AAC.01226-16>
- Cuperlovic-Culf, M., & Culf, A. S. (2016). Applied metabolomics in drug discovery. *Expert Opinion on Drug Discovery*, 11(8), 759–770.
- D'Alessandro, A., & Zolla, L. (2013). Biochemistry of red cell aging *in vivo* and storage lesions. *Haematologica*, 7, 389–396.
- D'Alessandro, A., & Zolla, L. (2017). Proteomic analysis of red blood cells and the potential for the clinic: what have we learned so far? *Expert Review of Proteomics*, 14(3), 243–252.
- De Koning, H. P., Bridges, D. J., & Burchmore, R. J. S. (2005). Purine and pyrimidine transport in pathogenic protozoa: from biology to therapy. *FEMS Microbiology Reviews*, 29(5), 987–1020.
- Derbyshire, E. R., Mota, M. M., & Clardy, J. (2011). The next opportunity in anti-malaria drug discovery: The liver stage. *PLoS Pathogens*, 7(9). <https://doi.org/10.1371/journal.ppat.1002178>
- Dewick, P. M. (2002). *Medicinal natural products: a biosynthetic approach* (second edi). John Wiley & Sons.
- Diederich, L., Iv, T. C. S. K., Kuhn, V., & Kramer, C. M. (2017). Red Blood Cell Function and Dysfunction: Redox Regulation, Nitric Oxide Metabolism, Anemia. *Antioxidants & Redox Signaling*, 26(13), 718–742. <https://doi.org/10.1089/ars.2016.6954>
- Diggs, C., Joseph, K., Flemmings, B., Snodgrass, R., & Hines, F. (1975). Protein synthesis *in vitro* by cryopreserved *Plasmodium falciparum*. *The American Journal of Tropical Medicine and Hygiene*, 24(5), 760–763.
- Divino Filho, J. C., Barany, P., Stehle, P., Fürst, P., & Bergström, J. (1997). Free amino-acid levels simultaneously collected in plasma, muscle, and erythrocytes of uraemic patients. *Nephrology Dialysis Transplantation*, 12(11), 2339–2348.

- Dondorp, A. M., Nosten, F., Yi, P., Das, D., Phyoe, A. P., Tarning, J., ... Lee, S. J. (2009). Artemisinin resistance in *Plasmodium falciparum* malaria. *New England Journal of Medicine*, 361(5), 455–467.
- Downie, M. J., Kirk, K., & Mamoun, C. Ben. (2008). Purine salvage pathways in the intraerythrocytic malaria parasite *Plasmodium falciparum*. *Eukaryotic Cell*, 7(8), 1231–1237.
- Dua, V. K., Ojha, V. P., Roy, R., Joshi, B. C., Valecha, N., Devi, C. U., ... Subbarao, S. K. (2004). Anti-malarial activity of some xanthones isolated from the roots of *Andrographis paniculata*. *Journal of Ethnopharmacology*, 95(2–3), 247–251. <https://doi.org/10.1016/j.jep.2004.07.008>
- Eggleson, K. K., Duffin, K. L., & Goldberg, D. E. (1999). Identification and characterization of falcilysin, a metallopeptidase involved in haemoglobin catabolism within the malaria parasite *Plasmodium falciparum*. *Journal of Biological Chemistry*, 274(45), 32411–32417. <https://doi.org/10.1074/jbc.274.45.32411>
- Elmore, S. (2007). Apoptosis: A Review of Programmed Cell Death. *Toxicol Pathol.*, 35(4), 495–516.
- Eriksson, L., Andersson, P. L., Johansson, E., & Tysklind, M. (2006). Megavariate analysis of environmental QSAR data. Part I—A basic framework founded on principal component analysis (PCA), partial least squares (PLS), and statistical molecular design (SMD). *Molecular Diversity*, 10(2), 169–186.
- Eriksson, L., Antti, H., Gottfries, J., Holmes, E., Johansson, E., Lindgren, F., ... Wold, S. (2004). Using chemometrics for navigating in the large data sets of genomics, proteomics, and metabolomics (gpm). *Analytical and Bioanalytical Chemistry*, 380(3), 419–429.
- Fabricant, D. S., & Farnsworth, N. R. (2001). The value of plants used in traditional medicine for drug discovery. *Environmental Health Perspectives*, 109(suppl 1), 69–75.
- Fairhurst, R. M., & Dondorp, A. M. (2016). Artemisinin-resistant *Plasmodium falciparum* malaria. *Emerging Infections*, 10, 409–429.
- Färber, P. M., Arscott, L. D., Williams, C. H., Becker, K., & Schirmer, R. H. (1998). Recombinant *Plasmodium falciparum* glutathione reductase is inhibited by the antimalarial dye methylene blue. *FEBS Letters*, 422(3), 311–314. [https://doi.org/10.1016/S0014-5793\(98\)00031-3](https://doi.org/10.1016/S0014-5793(98)00031-3)
- Fidock, D. A. (2016). Drug discovery: Chemical diversity targets malaria. *Nature*, 538(7625), 323–325. <https://doi.org/10.1038/nature19481>
- Forstermann, U., & Sessa, W. C. (2012). Nitric oxide synthases: regulation and function. *Eur Heart J*, 33(7), 829–837.

- Francis, S E, Sullivan, D. J., & Goldberg, D. E. (1997b). Hemoglobin metabolism in the malaria parasite *Plasmodium falciparum*. *Annual Review of Microbiology*, 51, 97–123. <https://doi.org/10.1146/annurev.micro.51.1.97>
- Francis, Susan E, Banerjee, R., & Goldberg, D. E. (1997a). Biosynthesis and Maturation of the Malaria Aspartic Hemoglobinases Plasmepsins I and II \*. *The Journal of Biological Chemistry*, 272(23), 14961–14968.
- Gamo, F.-J., Sanz, L. M., Vidal, J., de Cozar, C., Alvarez, E., Lavandera, J.-L., ... Hasan, S. (2010). Thousands of chemical starting points for antimarial lead identification. *Nature*, 465(7296), 305–310.
- Garnham, P. C. C. (1966). Malaria Parasites and other Haemosporidia Oxford. *Oxford Blackwell Scientific Publications.England; Davis, Philadelphia*, 1132.
- Gelband, H., Panosian, C. B., & Arrow, K. J. E. (2004). *Saving Lives, Buying Time: Economics of Malaria Drugs in an Age of Resistance*. Washington: the national academies press.
- Ghosh, K., & Ghosh, K. (2007). Pathogenesis of anemia in malaria: A concise review. *Parasitology Research*, 101(6), 1463–1469. <https://doi.org/10.1007/s00436-007-0742-1>
- Ginsburg, H., Krugliak, M., Eidelman, O., & Cabantchik, Z. I. (1983). New permeability pathways induced in membranes of *Plasmodium falciparum* infected erythrocytes. *Molecular and Biochemical Parasitology*, 8(2), 177–190.
- Gnanasekaran, G., & Murthy, G. V. S. (2012). Lectotypifications in *andrographis* (Acanthaceae). *Rheedea*, 22(2), 77–79.
- Goulart, H. R., Kimura, E. A., Peres, V. J., Couto, A. S., Duarte, F. A. A., & Katzin, A. M. (2004). Terpenes arrest parasite development and inhibit biosynthesis of isoprenoids in *Plasmodium falciparum*. *Antimicrobial Agents and Chemotherapy*, 48(7), 2502–2509.
- Graham H. Coombs, Daniel E. Goldberg, Michael Klemba, Colin Berry, John Kay, J. C. M. (2001). Aspartic proteases of *Plasmodium falciparum* and other parasitic protozoa as drug targets. *Trends in Parasitology*, 17(11), 532–537. [https://doi.org/10.1016/S1471-4922\(01\)02037-2](https://doi.org/10.1016/S1471-4922(01)02037-2)
- Greenwood, B. M., Fidock, D. A., Kyle, D. E., Kappe, S. H. I., Alonso, P. L., Collins, F. H., & Duffy, P. E. (2008). Malaria: progress, perils, and prospects for eradication. *The Journal of Clinical Investigation*, 118(4), 1266–1276.
- Guggisberg, A. M., Amthor, R. E., & Odom, A. R. (2014). Isoprenoid biosynthesis in *Plasmodium falciparum*. *Eukaryotic Cell*, 13(11), 1348–135. <https://doi.org/10.1128/EC.00160-14>

- Guiguemde, W. A., Shelat, A. A., Bouck, D., Duffy, S., Crowther, G. J., Smithson, D. C., ... Zhu, F. (2010). Chemical genetics of *Plasmodium falciparum*. *Nature*, 465(7296), 311–315.
- Halliwell, B., & Gutteridge, J. M. C. (2015). *Free radicals in biology and medicine*. Oxford University Press, USA.
- Harvey, A. L. (2008). Natural products in drug discovery. *Drug Discovery Today*, 13(19–20), 894–901.
- Hay, S. I., Guerra, C. A., Tatem, A. J., Noor, A. M., & Snow, R. W. (2004). The global distribution and population at risk of malaria: past, present, and future. *The Lancet Infectious Diseases*, 4(6), 327–336.
- Hempelmann, E. (2007). Hemozoin biocrystallization in *Plasmodium falciparum* and the antimalarial activity of crystallization inhibitors. *Parasitology Research*, 100(4), 671–676.
- Hendriks, M. M., van Eeuwijk, F. A., Jellema, R. H., Westerhuis, J. A., Reijmers, T. H., Hoefsloot, H. C. J., & Smilde, A. K. (2011). Data-processing strategies for metabolomics studies. *TrAC Trends in Analytical Chemistry*, 30(10), 1685–1698.
- Horgan, R. P., & Kenny, L. C. (2011). SAC review ‘Omic’ technologies: genomics, transcriptomics, proteomics and metabolomics. *The Obstetrician & Gynaecologist*, 13, 189–195. <https://doi.org/10.1576/toag.13.3.189.27672>
- Hossain, M. S., Urbi, Z., Sule, A., & Rahman, K. M. H. (2014). *Andrographis paniculata* (Burm. f.) Wall. ex Nees: A review of ethnobotany, phytochemistry, and pharmacology. *Scientific World Journal*, 2014. <https://doi.org/10.1155/2014/274905>
- House, M. (1985). Discovery of the hypnozoite and a new theory of malarial relapse. *ROYAL SOCIETY OF TROPICAL*, 1–11.
- Hyde, J. E. (2007). Targeting purine and pyrimidine metabolism in human apicomplexan parasites. *Current Drug Targets*, 8(1), 31–47.
- Huber, S. M., Uhlemann, A. C., Gamper, N. L., Duranton, C., Kremsner, P. G., & Lang, F. (2002). *Plasmodium falciparum* activates endogenous Cl<sup>-</sup> channels of human erythrocytes by membrane oxidation. *The EMBO journal*, 21(1-2), 22–30.
- Iruretagoyena, M. I., Tobar, J. A., González, P. A., Sepúlveda, S. E., Figueroa, C. A., Burgos, R. A., ... Kalergis, A. M. (2005). Andrographolide interferes with T cell activation and reduces experimental autoimmune encephalomyelitis in the mouse. *Journal of Pharmacology and Experimental Therapeutics*, 312(1), 366–372.

- Jager, M. M., Murk, J. L., Piqué, R. D., Hekker, T. A. M., & Vandenbroucke-Grauls, C. (2011). Five-minute Giemsa stain for rapid detection of malaria parasites in blood smears. *Tropical Doctor*, 41(1), 33–35.
- Jain, N. (2015). *A Study on the Application of Biophysical Techniques to Terminal Diseases Via the Metabolomic-Based Analyses of Cancers and the Analysis Of Alzheimer's Disease through the Action of Amyloid-Beta (PhD Thesis)*.
- Jarukamjorn, K., & Nemoto, N. (2008). Pharmacological Aspects of *Andrographis paniculata* on Health and Its Major Diterpenoid Constituent Andrographolide. *Journal of Health Science*, 54(4), 370–381. <https://doi.org/10.1248/jhs.54.370>
- Jan, Z., Khan, A., Sajjad, M., Muhammad, K., Rho, S., & Mehmood, I. (2018). A review on automated diagnosis of malaria parasite in microscopic blood smears images. *Multimedia Tools and Applications*, 77(8), 9801–9826
- Jones, D. P., Park, Y., & Ziegler, T. R. (2012). Nutritional metabolomics: progress in addressing complexity in diet and health. *Annu Rev Nutr*, 32, 183–202.
- Jongwutiwes, S., Putaporntip, C., Iwasaki, T., Sata, T., & Kanbara, H. (2004). Naturally acquired *Plasmodium knowlesi* malaria in human, Thailand. *Emerging Infectious Diseases*, 10(12), 2211.
- Jortzik, E., & Becker, K. (2012). Thioredoxin and glutathione systems in *Plasmodium falciparum*. *International Journal of Medical Microbiology*, 302(4–5), 187–194.
- Jortzik, E., Mailu, B. M., Preuss, J., Fischer, M., Bode, L., Rahlf, S., & Becker, K. (2011). Glucose-6-phosphate dehydrogenase–6-phosphogluconolactonase: A unique bifunctional enzyme from *Plasmodium falciparum*. *Biochemical Journal*, 436(3), 641–650.
- Joselin, J., & Jeeva, S. (2014). *Andrographis paniculata*: A Review of its Traditional Uses , Phytochemistry and Pharmacology. *Medicinal & Aromatic Plants*, 3(4), 169. <https://doi.org/10.4172/2167-0412.1000169>
- Kafsack, B. F., & Llinás, M. (2010). Eating at the Table of Another : Metabolomics of Host-Parasite Interactions. *Cell Host and Microbe*, 7(2), 90–99. <https://doi.org/10.1016/j.chom.2010.01.008>
- Kandanur, S. G. S., Tamang, N., Golakoti, N. R., & Nanduri, S. (2019). Andrographolide: A natural product template for the generation of structurally and biologically diverse diterpenes. *European Journal of Medicinal Chemistry*, 176, 513–533. <https://doi.org/10.1016/j.ejmech.2019.05.022>
- Kaur, K., Jain, M., Kaur, T., & Jain, R. (2009). Antimalarials from nature. *Bioorganic & Medicinal Chemistry*, 17(9), 3229–3256. <https://doi.org/10.1016/j.bmc.2009.02.050>

- Kell, D. B., Brown, M., Davey, H. M., Dunn, W. B., Spasic, I., & Oliver, S. G. (2005). Metabolic footprinting and systems biology: the medium is the message. *Nature Reviews Microbiology*, 3(7), 557–565. <https://doi.org/10.1038/nrmicro1177>
- Kemsley, E. K., Le Gall, G., Dainty, J. R., Watson, A. D., Harvey, L. J., Tapp, H. S., & Colquhoun, I. J. (2007). Multivariate techniques and their application in nutrition: a metabolomics case study. *British Journal of Nutrition*, 98(1), 1–14.
- Kicska, G. A., Tyler, P. C., Evans, G. B., Furneaux, R. H., Schramm, V. L., & Kim, K. (2002). Purine-less Death in *Plasmodium falciparum* Induced by Immucillin-H, a Transition State Analogue of Purine Nucleoside Phosphorylase. *Journal of Biological Chemistry*, 277(5), 3226–3231.
- Kirsten Moll, Inger Ljungström, Hedvig Perlmann, Artur Scherf, M. W. (2008). *Methods in malaria research*.
- Kiszewski, A. E., & Tekleghaimanot, A. (2004). A review of the clinical and epidemiological burdens of epidemic malaria. *American Journal of Tropical Medical Hygiene*, 71(Suppl 2), 128–135. Retrieved from [http://www.ajtmh.org/content/71/2\\_suppl/128.full.pdf](http://www.ajtmh.org/content/71/2_suppl/128.full.pdf)
- Kohl, S. M., Klein, M. S., Hochrein, J., Oefner, P. J., Spang, R., & Gronwald, W. (2012). State-of-the art data normalization methods improve NMR-based metabolomic analysis. *Metabolomics*, 8(1), 146–160.
- Kourti, T., & MacGregor, J. F. (1995). Process analysis, monitoring and diagnosis, using multivariate projection methods. *Chemometrics and Intelligent Laboratory Systems*, 28(1), 3–21.
- Krampa, F., Aniweh, Y., Awandare, G., & Kanyong, P. (2017). Recent Progress in the Development of Diagnostic Tests for Malaria. *Diagnostics*, 7(3), 54. <https://doi.org/10.3390/diagnostics7030054>
- Krithika, R., Verma, R. J., & Shrivastav, P. S. (2013). Antioxidative and cytoprotective effects of Andrographolide against CCl<sub>4</sub>-induced hepatotoxicity in HepG2 cells. *Human & Experimental Toxicology*, 32(5), 530–543.
- Kropf, P., Fuentes, J. M., Fähnrich, E., Arpa, L., Herath, S., Weber, V., ... Müller, I. (2005). Arginase and polyamine synthesis are key factors in the regulation of experimental leishmaniasis *in vivo*. *The FASEB Journal*, 19(8), 1000–1002.
- Krugliak, M., Zhang, J., & Ginsburg, H. (2002). Intraerythrocytic *Plasmodium falciparum* utilizes only a fraction of the amino acids derived from the digestion of host cell cytosol for the biosynthesis of its proteins. *Molecular and Biochemical Parasitology*, 119(2), 249–256.
- Kuroyanagi, M., Sato, M., Ueno, A., & Nishi, K. (1987). Flavonoids from *Andrographis paniculata*. *Chemical and Pharmaceutical Bulletin*, 35(11), 4429–4435.

- Lambros, C., & Vanderberg, J. P. (1979). Synchronization of *Plasmodium falciparum* Erythrocytic Stages in Culture. *The Journal of Parasitology*, 65(3), 418–420.
- Landau, I., & Chabaud, A. G. (1994). Latency of *Plasmodium* merozoites and drug-resistance. A review. *Parasite*, 1(2), 105–114.
- Laveran, A. (1881). Un nouveau parasite trouvé dans le sang des malades atteints de fièvre palustre: origine parasitaire des accidents de l'impaludisme.
- Lehane, A. M., Hayward, R., Saliba, K. J., & Kirk, K. (2008). A verapamil-sensitive chloroquine-associated H<sup>+</sup> leak from the digestive vacuole in chloroquine-resistant malaria parasites. *Journal of Cell Science*, 121(10), 1624–1632.
- Le Guennec, A., Tayyari, F., Edison, A.S. (2017). Alternatives to nuclear overhauser enhancement spectroscopy presat and Carr-Purcell-Meiboom-Gill presat for NMR-based metabolomics. *Analytical Chemistry*, 89(17), 8582–8588.
- Lelliott, P. M., McMorran, B. J., Foote, S. J., & Burgio, G. (2015). In vivo assessment of rodent *Plasmodium* parasitaemia and merozoite invasion by flow cytometry. *JoVE (Journal of Visualized Experiments)*, (98), e52736.
- Lewis, I. A., Wacker, M., Olszewski, K. L., Cobbold, S. A., Baska, K. S., Tan, A., ... Llina, M. (2014). Metabolic QTL analysis links chloroquine resistance in *Plasmodium falciparum* to impaired hemoglobin catabolism. *PLoS Genetics*, 10(1). <https://doi.org/10.1371/journal.pgen.1004085>.
- Lew, V. L., Macdonald, L., Ginsburg, H., Krugliak, M., & Tiffert, T. (2004). Excess haemoglobin digestion by malaria parasites: a strategy to prevent premature host cell lysis. *Blood Cells, Molecules, and Diseases*, 32(3), 353–359.
- Li, J. W.-H., & Vedera, J. C. (2009). Drug discovery and natural products: end of an era or an endless frontier? *Science*, 325(5937), 161–165.
- Li Xiayan Cristina Legido-quigley. (2008). Advances in separation science applied to metabolomics. *Electrophoresis*, 29(18), 3724–3736. <https://doi.org/10.1002/elps.200700851>
- Lim, J. C. W., Chan, T. K., Ng, D. S., Sagineedu, S. R., Stanslas, J., & Wong, W. F. (2012). Andrographolide and its analogues: Versatile bioactive molecules for combating inflammation and cancer. *Clinical and Experimental Pharmacology and Physiology*, 39(3), 300–310. <https://doi.org/10.1111/j.1440-1681.2011.05633.x>
- Lin, F. L., Wu, S. J., Lee, S. C., & Ng, L. T. (2009). Antioxidant, antioedema and analgesic activities of *Andrographis paniculata* extracts and their active constituent Andrographolide. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 23(7), 958–964.

- Lindon, J. C., Holmes, E., & Nicholson, J. K. (2006). Metabonomics techniques and applications to pharmaceutical research & development. *Pharmaceutical Research*, 23(6), 0175-1088.
- Liu, J., Istvan, E. S., Gluzman, I. Y., Gross, J., & Goldberg, D. E. (2006). *Plasmodium falciparum* ensures its amino acid supply with multiple acquisition pathways and redundant proteolytic enzyme systems. *Proceedings of the National Academy of Sciences.*, 103(23), 8840-8845.
- López, M. L., Blair, S., Sáez, J., & Segura, C. (2009). Effect of Solanum nudum steroids on uninfected and *Plasmodium falciparum*-infected erythrocytes. *Memorias do Instituto Oswaldo Cruz*, 104(5), 683-688.
- Luo, W., Liu, Y., Zhang, J. U. N., Luo, X., Lin, C., & Guo, J. (2013). Andrographolide inhibits the activation of NF -  $\kappa$  B and MMP - 9 activity in H3255 lung cancer cells. *Experimental and Therapeutic Medicine*, 6(3), 743-746. <https://doi.org/10.3892/etm.2013.1196>
- Macallum, W. G. (1897). On the flagellated form of the malarial parasite. *The Lancet*, 150(3872), 1240-1241.
- MacCallum, W. G. (1898). On the haematozoan infections of birds. *The Journal of Experimental Medicine*, 3(1), 117.
- Magallón-Tejada, A., Machevo, S., Cisteró, P., Lavstsen, T., Aide, P., Rubio, M., ... Gupta, H. (2016). Cytoadhesion to gC1qR through *Plasmodium falciparum* erythrocyte membrane protein 1 in severe malaria. *PLoS Pathogens*, 12(11), e1006011.
- Makler, M. T., & Hinrichs, D. J. (1993). Measurement of the lactate dehydrogenase activity of *Plasmodium falciparum* as an assessment of parasitaemia. *American Journal of Tropical Medicine and Hygiene*, 48(2), 205-210.
- Makler MT, Ries JM, Williams JA, Bancroft JE, Piper RC, Gibbins BL, H. D. (1993). Parasite lactate dehydrogenase as an assay for *Plasmodium falciparum* drug sensitivity. *Am J Trop Med Hyg*, 48(6), 739-741.
- Mathema, V. B., & Na-Bangchang, K. (2015). A brief review on biomarkers and proteomic approach for malaria research. *Asian Pacific Journal of Tropical Medicine*, 8(4), 253-262. [https://doi.org/10.1016/S1995-7645\(14\)60327-8](https://doi.org/10.1016/S1995-7645(14)60327-8)
- Martin, R. E., & Kirk, K. (2007). Transport of the essential nutrient isoleucine in human erythrocytes infected with the malaria parasite *Plasmodium falciparum*. *Blood*, 109(5), 2217-2224.
- Mathew, L., Rajasekaran, A., Arivukkarasu, R., & Mathew, L. (2016). A systematic comprehensive review on therapeutic potential of *Andrographis paniculata* (Burm. f.) Wall. ex Nees. *Journal of Pharmacognosy and Phytochemistry*, 5(5), 189-199.

- Matsuda, T., Kuroyanagi, M., Sugiyama, S., Umehara, K., Ueno, A., & Nishi, K. (1994). Cell differentiation-inducing diterpenes from *Andrographis paniculata* Nees. *Chemical and Pharmaceutical Bulletin*, 42(6), 1216–1225.
- Megantara, S., Levita, J., & Ibrahim, S. (2015). *In Silico* Study of Andrographolide as Protease Inhibitors for Antimalarial Drug Discovery. In *3rd International Conference on Computation for Science and Technology (ICCST-3)*. Atlantis Press., 36–39.
- Meierjohann, S., Walter, R. D., & Müller, S. (2002). Regulation of intracellular glutathione levels in erythrocytes infected with chloroquine-sensitive and chloroquine-resistant *Plasmodium falciparum*. *Biochemical Journal*, 368(3), 761–768.
- Messana, I., Ferroni, L., Misiti, F., Girelli, G., Pupella, S., Castagnola, M., ... Giardina, B. (2000). Blood bank conditions and RBCs: the progressive loss of metabolic modulation. *Transfusion*, 40(3), 353–360.
- Miller, L. H., Ackerman, H. C., Su, X., & Wellem, T. E. (2013). Malaria biology and disease pathogenesis: insights for new treatments. *Nature Medicine*, 19(2), 156.
- Miotto, O., Almagro-Garcia, J., Manske, M., MacInnis, B., Campino, S., Rockett, K. A., ... Kwiatkowski, D. P. (2013). Multiple populations of artemisinin-resistant *Plasmodium falciparum* in Cambodia. *Nature Genetics*, 45(6), 648–655. <https://doi.org/10.1038/ng.2624>
- Mishra, K., Dash, A. P., & Dey, N. (2011). Andrographolide: A novel antimalarial diterpene lactone compound from *Andrographis paniculata* and its interaction with curcumin and artesunate. *Journal of Tropical Medicine*, 2011, 1–6. <https://doi.org/10.1155/2011/579518>
- Mishra, K., Dash, A. P., Swain, B. K., & Dey, N. (2009). Anti-malarial activities of *Andrographis paniculata* and *Hedyotis corymbosa* extracts and their combination with curcumin. *Malaria Journal*, 8(1), 26. <https://doi.org/10.1186/1475-2875-8-26>
- Mishra, M., Mishra, V. K., Kashaw, V., Iyer, A. K., & Kashaw, S. K. (2016). Comprehensive review on various strategies for antimalarial drug discovery. *European Journal of Medicinal Chemistry*, 125, 1300–1320. <https://doi.org/10.1016/j.ejmech.2016.11.025>
- Misra, P., Pal, N. L., Guru, P. Y., Katiyar, J. C., Srivastava, V., & Tandon, J. S. (1992). Antimalarial activity of *Andrographis paniculata* (Kalmegh) against *Plasmodium berghei* NK 65 in Mastomys natalensis. *International Journal of Pharmacognosy* 274–263, (4)30, <https://doi.org/10.3109/13880209209054010>

- Moore, L. R., Fujioka, H., Williams, P. S., Chalmers, J. J., Grimberg, B., Zimmerman, P. A., & Zborowski, M. (2006). Hemoglobin degradation in malaria-infected erythrocytes determined from live cell magnetophoresis. *The FASEB Journal*, 20(6), 747–749.
- Moura, I. C., Wunderlich, G., Uhrig, M. L., Couto, A. S., Peres, V. J., Katzin, A. M., & Kimura, E. A. (2001). Limonene arrests parasite development and inhibits isoprenylation of proteins in *Plasmodium falciparum*. *Antimicrobial Agents and Chemotherapy*, 45(9), 2553–2558. <https://doi.org/10.1128/AAC.45.9.2553-2558.2001>
- Müller, S. (2004). Redox and antioxidant systems of the malaria parasite *Plasmodium falciparum*. *Molecular Microbiology*, 53(5), 1291–1305. <https://doi.org/10.1111/j.1365-2958.2004.04257.x>
- Müller, S. (2015). Role and regulation of glutathione metabolism in *plasmodium falciparum*. *Molecules*, 20(6), 10511–10534. <https://doi.org/10.3390/molecules200610511>
- Murithi, J. M., Owen, E. S., Istvan, E. S., Llina, M., Fidock, D. A., Vanaerschot, M., ... Chibale, K. (2020). Combining stage specificity and metabolomic profiling to advance antimalarial drug discovery. *Cell Chemical Biology*, 27(2), 158–171. <https://doi.org/10.1016/j.chembiol.2019.11.009>
- Mussard, E., Cesaro, A., Lespessailles, E., Legrain, B., Berteina-Raboin, S., & Toumi, H. (2019). Andrographolide, a natural antioxidant: An update. *Antioxidants*, 8(12), 571. <https://doi.org/10.3390/antiox8120571>
- Mustacich, D., & Powis, G. (2000). Thioredoxin reductase. *Biochemical Journal*, 346(1), 1–8.
- Nagana Gowda, G. A., Gowda, Y. N., & Raftery, D. (2015). Expanding the limits of human blood metabolite quantitation using NMR spectroscopy. *Analytical Chemistry*, 87(1), 706–715. <https://doi.org/10.1021/ac503651e>
- Nagana Gowda, G. A., & Raftery, D. (2017). Whole Blood Metabolomics by <sup>1</sup>H NMR Spectroscopy Provides a New Opportunity to Evaluate Coenzymes and Antioxidants. *Analytical Chemistry*, 89(8), 4620–4627. <https://doi.org/10.1021/acs.analchem.7b00171>
- Newman, D. J., & Cragg, G. M. (2007). Natural products as sources of new drugs over the last 25 years. *Journal of Natural Products*, 70(3), 461–477.
- Niranjan, A., Tewari, S. K., & Lehri, A. (2010). Biological activities of Kalmegh (*Andrographis paniculata* Nees) and its active principles-A review. *Indian Journal of Natural Products and Resources*, 1(2), 125–135.

- Noedl, H., Se, Y., Schaecher, K., Smith, B. L., Socheat, D., & Fukuda, M. M. (2008). Evidence of artemisinin-resistant malaria in western Cambodia. *New England Journal of Medicine*, 359(24), 2619–2620.
- Nussenblatt, V., & Semba, R. D. (2002). Micronutrient malnutrition and the pathogenesis of malarial anemia. *Acta Tropica*, 82(3), 321–337. [https://doi.org/10.1016/S0001-706X\(02\)00049-9](https://doi.org/10.1016/S0001-706X(02)00049-9)
- Olszewski, K. L., & Llinás, M. (2011). Central carbon metabolism of *Plasmodium* parasites. *Molecular and Biochemical Parasitology*, 175(2), 95–103. <https://doi.org/10.1016/j.molbiopara.2010.09.001>
- Olszewski, K. L., Morrisey, J. M., Wilinski, D., Burns, J. M., Vaidya, A. B., Rabinowitz, J. D., & Llinás, M. (2009). Host-Parasite Interactions Revealed by *Plasmodium falciparum* Metabolomics. *Cell Host & Microbe*, 5(2), 191–199. <https://doi.org/10.1016/J.CHOM.2009.01.004>
- Olszewski K.L.; Llinás M. (2012). Extraction of Hydrophilic Metabolites from *Plasmodium falciparum*-Infected Erythrocytes for Metabolomic Analysis. In In: Ménard R. (eds) *Malaria. Methods in Molecular Biology (Methods and Protocols)*; vol 923. pp. 259–266. Humana Press; Totowa; NJ. <https://doi.org/10.1007/978-1-62703-026-7>
- Pan, Z., & Raftery, D. (2007). Comparing and combining NMR spectroscopy and mass spectrometry in metabolomics. *Analytical and Bioanalytical Chemistry*, 387(2), 525–527. <https://doi.org/10.1007/s00216-006-0687-8>
- Panossian, A., Hovhannisyan, A., Mamikonyan, G., Abrahamian, H., Hambardzumyan, E., Gabrielian, E., ... Wagner, H. (2000). Pharmacokinetic and oral bioavailability of Andrographolide from *Andrographis paniculata* fixed combination Kan Jang in rats and human. *Phytomedicine*, 7(5), 351–364. [https://doi.org/10.1016/S0944-7113\(00\)80054-9](https://doi.org/10.1016/S0944-7113(00)80054-9)
- Park, Y. H., Shi, Y. P., Liang, B., Medriano, C. A. D., Jeon, Y. H., Torres, E., ... Jones, D. P. (2015). High-resolution metabolomics to discover potential parasite-specific biomarkers in a *Plasmodium falciparum* erythrocytic stage culture system. *Malaria Journal*, 14(1), 122. <https://doi.org/10.1186/s12936-015-0651-1>
- Patel, S. D., Ahoudi, A. D., Bei, A. K., Dieye, T. N., Mboup, S., Harrison, S. C., & Duraisingh, M. T. (2013). *Plasmodium falciparum* merozoite surface antigen, PfRH5, elicits detectable levels of invasion-inhibiting antibodies in humans. *The Journal of Infectious Diseases*, 208(10), 1679–1687.
- Perkins, D. J., Were, T., Davenport, G. C., Kempaiah, P., Hittner, J. B., & Ong'echa, J. M. (2011). Severe malarial anemia: Innate immunity and pathogenesis. *International Journal of Biological Sciences*, 7(9), 1427–1442. <https://doi.org/10.7150/ijbs.7.1427>

- Petersen, I., Eastman, R., & Lanzer, M. (2011). Drug-resistant malaria: Molecular mechanisms and implications for public health. *FEBS Letters*, 585(11), 1551–1562. <https://doi.org/10.1016/j.febslet.2011.04.042>
- Phompradit, P., Chaijaroenkul, W., & Na-bangchang, K. (2017). Cellular mechanisms of action and resistance of *Plasmodium falciparum* to artemisinin. *Parasitology Research*, 116(12), 3331–3339.
- Phyo, A. P., Nkhoma, S., Stepniewska, K., Ashley, E. A., Nair, S., Mcgready, R., ... Al-saai, S. (2012). Emergence of artemisinin-resistant malaria on the western border of Thailand: a longitudinal study. *The Lancet*, 379(9830), 1960–1966. [https://doi.org/10.1016/S0140-6736\(12\)60484-X](https://doi.org/10.1016/S0140-6736(12)60484-X)
- Phompradit, Papichaya, Wanna Chaijaroenkul, and Kesara Na-bangchang. (2017). Cellular Mechanisms of Action and Resistance of *Plasmodium falciparum* to Artemisinin. *Parasitology research* 116(12): 3331–39
- Picot, S., & Burnod, J. (1997). Apoptosis related to chloroquine *Plasmodium falciparum* sensitivity of the human malaria parasite. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 91(5), 590–591.
- Pinder, J., Fowler, R., Bannister, L., Dluzewski, A., & Mitchell, G. (2000). Motile systems in malaria merozoites: how is the red blood cell invaded? *Parasitology Today*, 16(6), 240–245.
- Plouffe, D., Brinker, A., McNamara, C., Henson, K., Kato, N., Kuhen, K., ... Anderson, P. (2008). *In silico* activity profiling reveals the mechanism of action of antimalarials discovered in a high-throughput screen. *Proceedings of the National Academy of Sciences*, 105(26), 9059–9064.
- Pompella, A., Visvikis, A., Paolicchi, A., De Tata, V., & Casini, A. F. (2003). The changing faces of glutathione, a cellular protagonist. *Biochemical Pharmacology*, 66(8), 1499–1503. [https://doi.org/10.1016/S0006-2952\(03\)00504-5](https://doi.org/10.1016/S0006-2952(03)00504-5)
- Putri, S. P., Yamamoto, S., Tsugawa, H., & Fukusaki, E. (2013). Current metabolomics: Technological advances. *Journal of Bioscience and Bioengineering*, 116(1), 9–16. <https://doi.org/10.1016/j.jbiosc.2013.01.004>
- Rabenstein, D. L. (1984). <sup>1</sup>H NMR methods for the noninvasive study of metabolism and other processes involving small molecules in intact erythrocytes. *Journal of Biochemical and Biophysical Methods*, 9(4), 277–306.
- Raftos, J. E., Whillier, S., & Kuchel, P. W. (2010). Glutathione synthesis and turnover in the human erythrocyte: Alignment of a model based on detailed enzyme kinetics with experimental data. *Journal of Biological Chemistry*, 285(31), 23557–23567. <https://doi.org/10.1074/jbc.M109.067017>

- Rasoanaivo, P., Wright, C. W., Willcox, M. L., & Gilbert, B. (2011). Whole plant extracts versus single compounds for the treatment of malaria: synergy and positive interactions. *Malaria Journal*, 10(1), S4. <https://doi.org/10.1186/1475-2875-10-S1-S4>
- Reglinski, J., Smith, W. E., Brzeski, M., Marabani, M., & Sturrock, R. D. (1992). Clinical analysis by proton spin-echo NMR. 2. Oxidation of intracellular glutathione as a consequence of penicillamine therapy in rheumatoid arthritis. *Journal of Medicinal Chemistry*, 35(11), 2134–2137.
- Reglinski, J., Smith, W. E., Wilson, R., Buchanan, L. M., McKillop, J. H., Thomson, J. A., ... Sturrock, R. D. (1991). Clinical analysis in intact erythrocytes using  $^1\text{H}$  spin echo NMR. *Clinica Chimica Acta*, 201(1–2), 45–57.
- Ribacke, U., Moll, K., Albrecht, L., Ismail, H. A., Normark, J., Flaberg, E., ... Egwang, T. G. (2013). Improved *in vitro* culture of *Plasmodium falciparum* permits establishment of clinical isolates with preserved multiplication, invasion and rosetting phenotypes. *PloS One*, 8(7), e69781.
- Ringwald, P., & Vestergaard, L. S. (2007). Responding to the Challenge of Antimalarial Drug Resistance by Routine Monitoring to Update National Malaria Treatment Policies. *The American Journal of Tropical Medicine and Hygiene*, 77(6\_Suppl), 153–159. <https://doi.org/10.4269/ajtmh.2007.77.153>
- Rochfort, S. (2005). Metabolomics reviewed: a new “omics” platform technology for systems biology and implications for natural products research. *Journal of Natural Products*, 68(12), 1813–1820.
- Rockett, K. A., Awburn, M. M., Cowden, W. B., & Clark, I. A. (1991). Killing of *Plasmodium falciparum* *in vitro* by nitric oxide derivatives. *Infection and Immunity*, 59(9), 3280–3283.
- Ross R. (1898). The role of the mosquito in the evolution of the malaria parasite. *Lancet*, 152(3912), 488–490. <https://doi.org/DOI: 10.1017/S0031182000083864>
- Rossi, R., Milzani, A., Dalle-Donne, I., Giustarini, D., Lusini, L., Colombo, R., & Di Simplicio, P. (2002). Blood glutathione disulfide: *in vivo* factor or *in vitro* artifact? *Clinical Chemistry*, 48(5), 742–753.
- Rowe, J. A., Claessens, A., & Corrigan, R. A. (2009). Adhesion of *Plasmodium falciparum*-infected erythrocytes to human cells: molecular mechanisms and therapeutic implications. *Expert Reviews in Molecular Medicine*, 11(E 16), 1–29. <https://doi.org/10.1017/S1462399409001082>.
- Rosenthal, P. J. (2002). Hydrolysis of erythrocyte proteins by proteases of malaria parasites. *Current opinion in hematatology*, 9(2), 140–145.

- Sachanonta, N., Chotivanich, K., Chaisri, U., Turner, G. D. H., Ferguson, D. J. P., Day, N. P. J., & Pongponratn, E. (2011). Ultrastructural and Real-time Microscopic Changes in *P. falciparum*-infected Red Blood Cells Following Treatment with Antimalarial Drugs. *Ultrastructural Pathology*, 35(5), 214–225. <https://doi.org/10.3109/01913123.2011.601405>
- Saliba, K. J., Horner, H. A., & Kirk, K. (1998). Transport and Metabolism of the Essential Vitamin Pantothenic Acid in Human Erythrocytes Infected with the Malaria Parasite *Plasmodium falciparum*. *Journal of Biological Chemistry*, 273(17), 10190–10195.
- Salinas, J. L., Kissinger, J. C., Jones, D. P., & Galinski, M. R. (2014). Metabolomics in the fight against malaria. *Memorias Do Instituto Oswaldo Cruz*, 109(5), 589–597. <https://doi.org/10.1590/0074-0276140043>
- Sana, T. R., Gordon, D. B., Fischer, S. M., Tichy, S. E., Kitagawa, N., Lai, C., ... Chang, S. P. (2013). Global Mass Spectrometry Based Metabolomics Profiling of Erythrocytes Infected with *Plasmodium falciparum*. *PLoS ONE*, 8(4), e60840. <https://doi.org/doi:10.1371/journal.pone.0060840>
- Sana, T. R., Waddell, K., & Fischer, S. M. (2008). A sample extraction and chromatographic strategy for increasing LC/MS detection coverage of the erythrocyte metabolome. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 871(2), 314–321. <https://doi.org/10.1016/j.jchromb.2008.04.030>
- Segre, D., Vitkup, D., & Church, G. M. (2002). Analysis of optimality in natural and perturbed metabolic networks. *Proceedings of the National Academy of Sciences*, 99(23), 15112–15117.
- Serkova, N. J., Zhang, Y., Coatney, J. L., Hunter, L., Michael, E., Niemann, C. U., & Mandell, M. S. (2007). Early detection of graft failure using the blood metabolic profile of a liver recipient. *Transplantation*, 83(4), 517–521. <https://doi.org/10.1097/01.tp.0000251649.01148.f8.Early>
- Sheeja, K., & Kuttan, G. (2007). Activation of Cytotoxic T Lymphocyte Responses and Attenuation of Tumor Growth *in vivo* by *Andrographis paniculata* Extract and Andrographolide. *Immunopharmacology and Immunotoxicology*, 29(1), 81–93. <https://doi.org/10.1080/08923970701282726>
- Sheeja, K., Shihab, P. K., & Kuttan, G. (2006). Antioxidant and Anti-Inflammatory Activities of the Plant *Andrographis*. *Immunopharmacology and Immunotoxicology*, 28(4), 129–140. <https://doi.org/10.1080/08923970600626007>
- Shen, Y., Chen, C., & Chiou, W. (2002). Andrographolide prevents oxygen radical production by human neutrophils: possible mechanism (s) involved in its anti- inflammatory effect. *British Journal of Pharmacology*, 135(2), 399–406.

- Shortt, H. E., & Garnham, P. C. C. (1948). Pre-erythrocytic stage in mammalian malaria parasites. *Nature*, 161(4082), 126.
- Sidhu, A. B. S., Uhlemann, A.-C., Valderramos, S. G., Valderramos, J.-C., Krishna, S., & Fidock, D. A. (2006). Decreasing *pfmndr1* copy number in *Plasmodium falciparum* malaria heightens susceptibility to mefloquine, lumefantrine, halofantrine, quinine, and artemisinin. *The Journal of Infectious Diseases*, 194(4), 528–535.
- Sivananthan, M., & Elamaraan, M. (2013). Medicinal and pharmacological properties of *Andrographis paniculata*. *International Journal of Biomolecules and Biomedicine*, 3(2), 1–12.
- Spangenberg, T., Burrows, J. N., Kowalczyk, P., McDonald, S., Wells, T. N. C., & Willis, P. (2013). The Open Access Malaria Box: A Drug Discovery Catalyst for Neglected Diseases. *PLOS ONE*, 8(6), e62906. <https://doi.org/10.1371/journal.pone.0062906>
- Sparrow, R. L. (2012). Time to revisit red blood cell additive solutions and storage conditions: A role for “omics” analyses. *Blood Transfusion*, 10(SUPPL. 2), s7–s11. <https://doi.org/10.2450/2012.003S>
- Steuer, A. E., Brockbals, L., & Kraemer, T. (2019). Metabolomic strategies in biomarker research—New approach for indirect identification of drug consumption and sample manipulation in clinical and forensic toxicology? *Frontiers in Chemistry*, 7, 319.
- Surolia, N., & Padmanaban, G. (1991). Chloroquine inhibits heme-dependent protein synthesis in *Plasmodium falciparum*. *Proceedings of the National Academy of Sciences*, 88(11), 4786–4790.
- Tachado, S. D., Gerold, P., McConville, M. J., Baldwin, T., Quilici, D., Schwarz, R. T., & Schofield, L. (1996). Glycosylphosphatidylinositol toxin of *Plasmodium* induces nitric oxide synthase expression in macrophages and vascular endothelial cells by a protein tyrosine kinase-dependent and protein kinase C-dependent signaling pathway. *The Journal of Immunology*, 156(5), 1897–1907.
- Teng, R., Junankar, P. R., Bubb, W. A., Rae, C., Mercier, P., & Kirk, K. (2009). Metabolite profiling of the intraerythrocytic malaria parasite *Plasmodium falciparum* by <sup>1</sup>H NMR spectroscopy. *NMR in Biomedicine: An International Journal Devoted to the Development and Application of Magnetic Resonance In Vivo*, 22(3), 292–302. <https://doi.org/10.1002/nbm.1323>
- Teng, R., Lehane, A. M., Winterberg, M., Shafik, S. H., Summers, R. L., Martin, R. E., ... Kirk, K. (2014). <sup>1</sup>H-NMR metabolite profiles of different strains of *Plasmodium falciparum*. *Bioscience Reports*, 34(6), 685–699. <https://doi.org/10.1042/BSR20140134>

- Tewari, S. G., Swift, R. P., Reifman, J., Prigge, S. T., & Wallqvist, A. (2020). Metabolic alterations in the erythrocyte during blood - stage development of the malaria parasite. *Malaria Journal*, 19(1), 1–18. <https://doi.org/10.1186/s12936-020-03174-z>
- Tiwari, N. K., Reynolds, P. J., & Calderón, A. I. (2016). Preliminary LC-MS Based Screening for Inhibitors of *Plasmodium falciparum* Thioredoxin Reductase (PfTrxR) among a Set of Antimalarials from the Malaria Box. *Molecules*, 21(4), 424. <https://doi.org/10.3390/molecules21040424>
- Trager, W., & Jensen, J. B. (1976). Human malaria parasites in continuous culture. *Science*, 193(4254), 673–675. <https://doi.org/10.1126/science.781840>
- Kalra, B. S., Chawla, S., Gupta, P., & Valecha, N. (2006). Screening of antimalarial drugs: An overview. *Indian journal of pharmacology*, 38(1), 5.
- Tripathy, S., Chattopadhyay, S., Dash, S. K., Chowdhuri, A. R., Das, S., Sahu, S. K., Roy, S. (2015). Chitosan conjugated chloroquine: Proficient to protect the induction of liver apoptosis during malaria. *International Journal of Biological Macromolecules*, 74, 585–600.
- Tritten, L., Keiser, J., Godejohann, M., Utzinger, J., Vargas, M., Beckonert, O., ... Saric, J. (2013). Metabolic profiling framework for discovery of candidate diagnostic markers of malaria. *Scientific Reports*, 3. <https://doi.org/10.1038/srep02769>
- Trivedi, N. P., Rawal, U. M., & Patel, B. P. (2007). Hepatoprotective effect of Andrographolide against hexachlorocyclohexane-induced oxidative injury. *Integrative Cancer Therapies*, 6(3), 271–280.
- Trung, H. D., Van Bortel, W., Sochantha, T., Keokenchanh, K., Quang, N. T., Cong, L. D., & Coosemans, M. (2004). Malaria transmission and major malaria vectors in different geographical areas of Southeast Asia. *Tropical Medicine & International Health*, 9(2), 230–237.
- Tu, Y. (2011). The discovery of artemisinin (qinghaosu) and gifts from Chinese medicineo Title. *Nature Medicine*, 17(10), 1217–1220.
- Van den Berg, R. A., Hoefsloot, H. C. J., Westerhuis, J. A., Smilde, A. K., & van der Werf, M. J. (2006). Centering, scaling, and transformations: improving the biological information content of metabolomics data. *BMC Genomics*, 7(1), 142.
- Van Der Werf, M. J., Jellema, R. H., & Hankemeier, T. (2005). Microbial metabolomics: replacing trial-and-error by the unbiased selection and ranking of targets. *Journal of Industrial Microbiology and Biotechnology*, 32(6), 234–252.
- Van Dooren, G. G., Stimmmer, L. M., & McFadden, G. I. (2006). Metabolic maps and functions of the *Plasmodium* mitochondrion. *FEMS Microbiology Reviews*, 30(4), 596–630. <https://doi.org/10.1111/j.1574-6976.2006.00027.x>

- Venkatesan, M., Gadalla, N. B., Stepniewska, K., Dahal, P., Nsanzabana, C., Moriera, C., ... Dorsey, G. (2014). Polymorphisms in *Plasmodium falciparum* chloroquine resistance transporter and multidrug resistance 1 genes: parasite risk factors that affect treatment outcomes for *P. falciparum* malaria after artemether-lumefantrine and artesunate-amodiaquine. *The American Journal of Tropical Medicine and Hygiene*, 91(4), 833–843.
- Verma, N., & Vinayak, M. (2008). Antioxidant action of *Andrographis paniculata* on lymphoma. *Molecular Biology Reports*, 35(4), 535–540.
- Verpoorte, R., Kim, H. K., & Choi, Y. H. (2006). Plants as source for medicines: New perspectives. *Frontis*, 261-273.
- Vogetseder, A., Ospelt, C., Reindl, M., Schober, M., & Schmutzhard, E. (2004). Time course of coagulation parameters, cytokines and adhesion molecules in *Plasmodium falciparum* malaria. *Tropical Medicine and International Health*, 9(7), 767–773. <https://doi.org/10.1111/j.1365-3156.2004.01265.x>
- Vossen, M. G., Pferschy, S., Chiba, P., & Noedl, H. (2010). The SYBR green I malaria drug sensitivity assay: Performance in low parasitaemia samples. *American Journal of Tropical Medicine and Hygiene*, 82(3), 398–401. <https://doi.org/10.4269/ajtmh.2010.09-0417>
- Waller, R. F., & McFadden, G. I. (2005). The Apicoplast: A Review of the Derived Plastid of Apicomplexan Parasites. *Current Issues in Molecular Biology*, 7, 57–80.
- Warhurst, D. C. (1987). Antimalarial Drugs. *Drugs*, 33(1), 50–65. <https://doi.org/10.2165/00003495-198733010-00003>
- Warrell, D. A. (1997). Cerebral malaria: clinical features, pathophysiology and treatment. *Annals of Tropical Medicine & Parasitology*, 91(7), 875–884.
- Wasman, S. Q., Mahmood, A. A., Chua, L. S., Alshawsh, M. A., & Hamdan, S. (2011). Antioxidant and gastroprotective activities of *Andrographis paniculata*(Hempedu Bumi) in Sprague Dawley rats. *Indian Journal of Experimental Biology*, 49(10), 767–772.
- Wellems, T. E., & Plowe, C. V. (2001). Chloroquine-Resistant Malaria. *The Journal of Infectious Diseases*, 184.(6), 770–776.
- Wells, N. T. (2012). New drugs for the control and elimination of malaria: A snapshot of the pipeline. *Malaria Journal*, 11(S1), 031. <https://doi.org/10.1186/1475-2875-11-S1-O31>
- White, N. J. (2004). Antimalarial drug resistance. *The Journal of Clinical Investigation*, 113(8), 1084–1092.

- White, N. J. (2011). Determinants of relapse periodicity in *Plasmodium vivax* malaria. *Malaria Journal*, 10(1), 297.
- World Health Organization. (2010). Basic malaria microscopy. *Parasitology Today*, 2nd Learne(Part1.). [https://doi.org/10.1016/0169-4758\(92\)90107-D](https://doi.org/10.1016/0169-4758(92)90107-D)
- World Health Organization . (2015). World Malaria Report. <https://www.who.int/malaria>. (Accessed 5 Aug. 2016).
- World Health Organization . (2017). World Malaria Report. <https://www.who.int/malaria>. (Accessed 3 Dec. 2018).
- World Health Organization. (2018). World Malaria Report. <https://www.who.int/malaria>. (Accessed 23 Sep. 2019).
- World Health Organization. (2020). World Malaria Report. <https://www.who.int/malaria>. (Accessed 8 Apr. 2021).
- Widyawaruyanti, A., Astrianto, D., Ilmi, H., Tumewu, L., Setyawan, D., Widiaستuti, E., ... Hafid, A. F. (2017). Antimalarial activity and survival time of *Andrographis paniculata* fraction (AS202-01) on *Plasmodium berghei* infected mice. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 8(1S), 49–54.
- Wiklund, S. (2008). Multivariate data analysis for Omics. *Umea: Umetrics*.
- Wilson, M. C., Trakarnsanga, K., Heesom, K. J., Cogan, N., Green, C., Toye, A. M., ... Frayne, J. (2016). Comparison of the proteome of adult and cord erythroid cells, and changes in the proteome following reticulocyte maturation. *Molecular & Cellular Proteomics*, 15(6), 1938–1946.
- Wishart, D. S. . (2010). Computational approaches to metabolomics. In. *Bioinformatics Methods in Clinical Research*.Humana Press., 283–313.
- Wishart, D. S., Feunang, Y. D., Marcu, A., Guo, A. C., Liang, K., Vázquez-Fresno, R., ... Karu, N. (2018). HMDB 4.0: the human metabolome database for 2018. *Nucleic Acids Research*, 46(D1), D608–D617.
- Wold, S., Sjostrom, M., & Eriksson, L. (2001). *PLS-regression: a basic tool of chemometrics*. *Chemometrics Intelligent Laboratory Systems*, 58 (2), 109-130.
- Worley, B., & Powers, R. (2013). Multivariate Analysis in Metabolomics. *Current Metabolomics*, 1(1), 92–107. <https://doi.org/10.2174/2213235x11301010092>
- Ya Zhang, K. S. . O. . A. and A. J. (1986). Stage-Dependent Inhibition of Chloroquine on *Plasmodium falciparum* *In vitro*. *J. Paras.*, *American Society of Parasitologists Stable*, 72(6), 830–836.

Yoopan, N., Thisoda, P., Rangkadilok, N., Sahasitiwat, S., Pholphana, N., Ruchirawat, S., & Satayavivad, J. (2007). Cardiovascular effects of 14-Deoxy-11,12-didehydroAndrographolide and *Andrographis paniculata* extracts. *Planta Medica*, 73(6), 503–511. <https://doi.org/10.1055/s-2007-967181>

Yoshida, T., Prudent, M., & D'alessandro, A. (2019). Red blood cell storage lesion: causes and potential clinical consequences. *Blood Transfusion*, 17(1), 27–52. <https://doi.org/10.2450/2019.0217-18>

Zaid, O. I., Abd Majid, R., Sabariah, M. N., Hasidah, M. S., Al-Zihiry, K., Yam, M. F., & Basir, R. (2015). Andrographolide effect on both *Plasmodium falciparum* infected and non infected RBCs membranes. *Asian Pacific Journal of Tropical Medicine*, 8(7), 507–512. <https://doi.org/10.1016/j.apjtm.2015.06.007>

Zakeri, S., Afsharpad, M., Kazemzadeh, T., Mehdizadeh, K., Shabani, A., & Djadid, N. D. (2008). Association of *pfcrt* but not *pfmdr1* alleles with chloroquine resistance in Iranian isolates of *Plasmodium falciparum*. *The American Journal of Tropical Medicine and Hygiene*, 78(4), 633–640.

Zaridah, M. Z., Idid, S. Z., Wan Omar, A., & Khozirah, S. (2001). *In vitro* antifilarial effects of three plant species against adult worms of subperiodic Brugia malayi. *Journal of Ethnopharmacology*, 78(1), 79–84. [https://doi.org/10.1016/S0378-8741\(01\)00286-0](https://doi.org/10.1016/S0378-8741(01)00286-0)

Ziegler, H. L., Stærk, D., Christensen, J., Hviid, L., Hägerstrand, H., & Jaroszewski, J. W. (2002). *In vitro* *Plasmodium falciparum* drug sensitivity assay: inhibition of parasite growth by incorporation of stomatocytogenic amphiphiles into the erythrocyte membrane. *Antimicrobial Agents and Chemotherapy*, 46(5), 1441–1446.

## **BIODATA OF STUDENT**

The student, Ashraf Ahmed Issa Alapid, is a Libyan, he was born on 23rd January 1982 at Gharyan Libya. He studied the primary, intermediate and secondary schools in his hometown during the period from 1987 to 1998. he did bachelor's degree at university of Al Jabal Gharbi, department of biology science under Faculty of science in 1999. He graduated in 2003 and obtained a Degree in biology science. In 2004, he started his master study in the Libyan Academy, school basic science, department biological science. He graduated in 2008 and obtained M.Sc. Degree in medical parasitology. After that, he was worked as Assistant lecturer in Al Jabal Gharbi University as well as Gharyan University from 2009 to 2014. Through this job, he gained a thorough teaching experience that may help him to continue in his future academic life. In Jaan 2015, he decided to quit teaching, and he obtained admission in Universiti Putra Malaysia to pursue his PhD program under the supervision of the esteemed Dr Roslaini Bin Abd Majid. Then he came to Malaysia and studied English in ELS UPM school, for six months before joining a PhD program in Medical parasitology Unit/department of medical microbiology and parasitology at Faculty of Medicine and Health Sciences/ Universiti Putra Malaysia (UPM). From 2016 to 2020. On personal life, he is married, and they are blessed with three sons only.

## LIST OF PUBLICATIONS

### Journal Articles

**Ashraf A.Alapid**, Zaid O.Ibraheem, I.S. Ismail, Ngah Z.U, Sharif A. A , Norshariza N, Wana, M. N., Basir R. and Roslaini Abd. Majid (2020). Investigation of Andrographolide effect on non-infected Red Blood Cells using the  $^1\text{H}$ -NMR-based metabolomics approach. *Metabolites* 11 (8): 486. <https://doi.org/https://doi.org/10.3390/metabo 11080486>.

**Ashraf A.Alapid**, Zaid O.Ibraheem, I.S. Ismail, Ngah Z.U, Sharif A. A , Norshariza N, Wana, M. N., Basir R. and Roslaini Abd. Majid (2019). Determination of time dependent effect of Andrographolide and chloroquine on the different stages of the intraerythrocytic cycle of the *Plasmodium falciparum* 3D7. submitted to *Journal of Tropical Medicine*.

**Ashraf A.Alapid**, Zaid O.Ibraheem, I.S. Ismail, Ngah Z.U, Sharif A. A , Norshariza N, Wana, M. N., Basir R. and Roslaini Abd. Majid (2020). Identification of the metabolic variations of antimalaria activity of Andrographolide on *Plasmodium falciparum* 3D7 by  $^1\text{H}$  NMR-based metabolomics approach. To be submitted to *Metabolites*

Alhassan Abdullahi Sharif, Ngah Zasmy Unyah, Norshariza Nordin, Rusliza Basir, Mohammed Nasiru Wana, **Ashraf Alapid Ahmad**, Tijjani Mustapha, and Roslaini Abd. Majid, Susceptibility of *Toxoplasma gondii* to Ethanolic Extract of *Tinospora crispa* in Vero Cells. *Evidence-Based Complementary and Alternative Medicine*, vol. 2019, Article ID 2916547, 10 pages, 2019. <https://doi.org/10.1155/2019/2916547>.

Sharif Alhassan Abdullahi, Ngah Zasmy Unyah, Noshariza Nordin, Rusliza Basir Wana Mohammed Nasiru, **Ashraf, Ahmad Alapid**, Yahaya Hassan, Tijjani, Mustapha and Roslaini Abd Majid, Therapeutic Targets on *Toxoplasma gondii* Parasite in Combatting Toxoplasmosis. *Annual Research & Review in Biology* 32(2): 1-15, 2019; Article no.ARRB .49444 ISSN: 2347 - 565X, NLM ID: 101632869

## Conference Presentations

**Ashraf Ahmad Issa Alapid**, Zaid O.I, Sharif A. A, Ngah ZU, Shariza N, Wana, M. N, Rusliza B, I.S. Ismail, Roslaini AM: The Effect of Andrographolide And Chloroquine on The Erythrocytic Stages of The *Plasmodium falciparum* 3d7 *In Vitro*. Global health and Infectious Diseases. Infections 2017, 24th-25th October 2017. Faculty of Medicine and Health Sciences, Universiti Putra Malaysia.

**Ashraf Ahmad Issa Alapid**, Zaid Ibraheem, Sharif Alhassan Abdullahi, Ngah Zasmy, Shariza Nordin, Mohammed Nasiru Wana, Rusliza Basir, Intan Ismail, Roslaini Bin Abd Majid. The Morphological Effect of Andrographolide on the Different Stages of Erythrocytic Cycle of the *Plasmodium falciparum* 3D7. 54th Annual Scientific Conference of the Malaysian Society of Parasitology and tropical Medicine (MSPTM); 14-15th March 2018, Connexion Conference and events Centre, Kuala Lumpur, Malaysia.

Sharif Alhassan Abdullahi, Ngah Zasmy, Norshariza Noordeen, **Ashraf Alapid**, Wana Mohammed Nasiru, Hassan Yahaya, Rusliza Basir, Roslaini Abd Majid: Susceptibility of *Toxoplasma Gondii* to Ethanolic Leaf Extract of *Andrographis Paniculata*. 54th Annual Scientific Conference of the Malaysian Society of Parasitology and tropical Medicine (MSPTM); 14-15th March 2018, Connexion Conference and events Centre, Kuala Lumpur, Malaysia.

Sharif A A, Ngah Z.U, Shariza N, Wana, MN, **Ashraf, A**, Rusliza B, Roslaini A.M. *In Vitro* Anti-Parasitic Activities of Some Malaysian Herbs on *Toxoplasma gondii*. Global health and Infectious Diseases. Infections 2017, 24th-25th October 2017. Faculty of Medicine and Health Sciences, Universiti Putra Malaysia.

Sharif A. A, Ngah ZU, Shariza N, Wana, M N, **Ashraf, A.Alapid**, Getso M.I, Dabo N.T, Yahaya H, Tijjani M, Rusliza B, Roslaini AM. Combating Toxoplasmosis: Identifying Essential Therapeutic Targets on *Toxoplasma gondii*. Malaysian Society for Microbiology Postgraduate Seminar, 9th October 2018; Faculty of Applied Sciences, University Technology Mara, Shah Alam, Selangor, Malaysia.

Sharif A A, Ngah ZU, Shariza N, Wana, M N., Tijjani, M, **Ashraf, A. Alapid**, Rusliza B, Roslaini AM. Potential of Malaysian Herbs in the Treatment of Zoonotic Infections: the *Toxoplasma gondii* Parasite, at the South East Asia One Health University Network (SEAOHUN' 2018) International Conference on 12th November, 2018, Vietnam.

Wana M. N., Mohamad A. M. M., Malaika W., Ngah Z. U., Sharif A. A., **Ashraf A. Alapid.**, Tijjani, M., Shariza N., Rusliza B., and Roslaini AM. A Preliminary Study on Molecular Characterization of *Toxoplasma gondii* Isolated from Cat Faeces in Serdang, Selangor, Malaysia. At the South East Asia One Health University Network (SEAOHUN' 2018) International Conference on 12th November 2018, Vietnam.

Sharif A. A, Ngah ZU, Shariza N, Wana, M N, **Ashraf, A.Alapid**, Yahaya H, Tijjani M, Rusliza B, Roslaini AM. *In vitro* anti-toxoplasmal activities of ethanolic extracts from *andrographis paniculata* and *Tinospora crispa* against *Toxoplasma gondii* parasite. International conference on drug discovery and translational medicine 2018 (ICDDTM, 18), held on the 4th and 5th December 2018, at the Everly hotel, Putrajaya, Malaysia.

Wana M. N, Mohamad A. M. M., Malaika W, Ngah Z. U., Sharif A. A, **Ashraf A. Alapid**, Tijjani, M, Shariza N, Rusliza B, and Roslaini AM. Biological and Molecular Characterisation of *Toxoplasma gondii* isolated from Cat Faeces in Serdang, Selangor, Malaysia. ASEAN Emerging researchers Conference, 3rd and 4th December 2018, at the Sunway University, Kuala Lumpur Malaysia.

Wana MN, Mohamad A MM, Malaika W, Ngah Z U, Sharif AA, **Ashraf A Alapid**, Tijjani M, Shariza N, Rusliza B, Roslaini AM. Effects of locally isolated Malaysian strains of *Toxoplasma gondii* on behaviors in albino waster rats. 55th Annual Scientific Conference of the Malaysian Society of Parasitology and tropical Medicine (MSPTM); 13-14th March 2019, InterContinental hotel, Kuala Lumpur, Malaysia.

#### Awards

1. The best poster award at the Global health and Infectious Diseases. Infections 2017, 24th-25th October 2017. Faculty of Medicine and Health Sciences, Universiti Putra Malaysia.



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