



Review article

Integrating physiological data with the conservation and management of fishes: a meta-analytical review using the threatened green sturgeon (Acipenser medirostris)

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Reversing global declines in the abundance and diversity of fishes is dependent on science-based conservation solutions. A wealth of data exist on the ecophysiological constraints of many fishes, but much of this information is underutilized in recovery plans due to a lack of synthesis. Here, we used the imperiled green sturgeon (Acipenser medirostris) as an example of how a quantitative synthesis of physiological data can inform conservation plans, identify knowledge gaps and direct future research actions. We reviewed and extracted metadata from peer-reviewed papers on green sturgeon. A total of 105 publications were identified, spanning multiple disciplines, with the primary focus being conservation physiology (23.8%). A meta-analytical approach was chosen to summarize the mean effects of prominent stressors (elevated temperatures, salinity, low food availability and contaminants) on several physiological traits (growth, thermal tolerance, swimming performance and heat shock protein expression). All examined stressors significantly impaired green sturgeon growth, and additional stressorspecific costs were documented. These findings were then used to suggest several management actions, such as mitigating salt intrusion in nursery habitats and maintaining water temperatures within optimal ranges during peak spawning periods. Key data gaps were also identified; research efforts have been biased towards juvenile (38.1%) and adult (35.2%) life-history stages, and less data are available for early life-history stages (embryonic, 11.4%; yolk-sac larvae, 12.4%; and post yolk-sac larvae, 16.2%). Similarly, most data were collected from single-stressor studies (91.4%) and there is an urgent need to understand interactions among stressors as anthropogenic change is multi-variate and dynamic. Collectively, these findings provide an example of how meta-analytic reviews are a powerful tool to inform management actions, with the end goal of maximizing conservation gains from research efforts.

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Introduction

The loss of biodiversity of fishes is occurring worldwide, with many fish populations in serious decline (Moyle and Leidy, 1992; Dudgeon et al., 2006). These losses are particularly acute in areas where anthropogenic stressors have led to habitat loss and degradation or changes in environmental variables (Marchetti and Moyle, 2001; Wenger et al., 2011; Hooper et al., 2012). As the need to effectively manage and conserve fishes has become more urgent, there has been a corresponding call for a greater understanding of the physiological ecology and behaviour of species in question. The profound lack of available detailed physiological and behavioural data has been underscored by fish population declines, and the data gaps often present a barrier to moving forward with new management and conservation actions. Similarly, managers are often unaware of what data currently exist and how this information could potentially be useful in mitigating or preventing further population declines. In the fields of fisheries science as well as experimental fish physiology and conservation biology, a call for greater integration of physiological information with fisheries management has been made in recent years (Wikelski and Cooke, 2006; Horodysky et al., 2015).

Physiology is extremely well suited to inform conservation efforts, including detailed information about environmental tolerances and optima (e.g. temperature, salinity, water velocity, turbidity, etc.), growth rates under different conditions, sensitivity to environmental contaminants and susceptibility to predation by invasive species (Cooke et al., 2013). This information is critical for species persistence and an awareness of the need for it has been reflected in recent status reports and assessments issued by conservation agencies such as the National Oceanographic Atmospheric Administration's National Marine Fisheries Service (NMFS). Collaborations between physiologists and fisheries managers have proven successful at more effectively managing fish populations for decades, though the emergence of a discipline to support this integration is a more recent development (Cooke et al., 2013). For example, the inclusion of water temperature as a critical abiotic factor affecting the growth, survival and distribution of fishes in management plans is long-standing (U.S. Fish and Wildlife Service, 1995). Much of our understanding of how fishes respond to both acute and gradual changes in temperature is derived from physiological

and behavioural studies conducted in the laboratory. A recent recovery plan released by NMFS included elevated temperatures as a key factor affecting populations of threatened and endangered Chinook salmon (Oncorhynchus tshawytscha) in California, citing physiological stress, increased energetic costs, reduced growth rates and higher incidence of disease as physiological mechanisms contributing to population declines (NMFS, 2014). Indeed, numerous water temperature control projects, such as regulated flow regimes or structured water releases from reservoirs, are currently in effect around the world to help create thermal habitat that will promote the survival and persistence of species of management concern. Similarly, physiology has helped mitigate direct human-induced mortality on fishes, including the regulation of fishing pressure based on predictive estimates made by integrating physiological and environmental data. Cooke et al. (2012) reviewed the numerous physiological studies that contributed to the regulation and management of Pacific salmon (Oncorhynchus) fisheries, including those that affected management of some imperiled populations of Fraser River sockeye salmon (O. nerka). As fish populations continue to decline and conservation and management plans become more detailed, the need to provide quantitative syntheses of existing data is crucial.

Although there are examples of physiological data being incorporated into biological opinions, recovery plans and assessments of conservation actions, this integration is often limited in scope or depth. As such, the need for a more systematic and quantitative approach to integrate physiological data into conservation plans is required. This includes not only a way to utilize the already existing data but also a process to identify knowledge gaps, with the overall goal of increasing the efficacy of future management and research efforts. Here, we used the imperiled green sturgeon (Acipenser medirostris, Ayres, 1854) as an example of how a quantitative synthesis of physiological data can inform conservation plans and future research actions. Green sturgeon are long-lived, anadromous fish inhabiting waters of the Pacific Northwest region of North America. Sturgeons are members of the subclass Chondrostei and family Acipenseridae, a group of evolutionarily ancient fish that diverged from the morederived teleosts over 200 million years ago (Moyle, 2002). Green sturgeon are slow to reach sexual maturity compared to most fishes (typically 15-17 years old and 1.5-1.6 m total

length, TL; Erickson and Webb, 2007) and have a spawning periodicity of 3-5 years (Moyle, 2002)—characteristics that make them particularly vulnerable to population declines. The green sturgeon population is comprised of two genetically distinct population segments (DPSs; Israel et al., 2004) that differ in spawning ground locations. The northern DPS spawns principally in the Rogue River in Oregon and the Klamath River in California, though both the Umpqua and Eel Rivers may contribute to the spawning population (Erickson et al., 2002). The southern DPS spawns primarily in mainstem Sacramento River in California (Israel et al., 2004; Israel and Kimley, 2008), though recent evidence documented spawning in the Feather River and the Yuba River as well (Seesholtz et al., 2015). Due to the restricted spawning grounds of the southern DPS, as well as habitat degradation and evidence of declining population numbers, the southern DPS was listed by the National Marine Fisheries Division of the National Oceanic and Atmospheric Administration in 2006 as 'threatened' under the federal Endangered Species Act of 1973. The northern DPS remains unlisted. although it is considered a 'species of concern' under federal designation.

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The drivers of green sturgeon population declines are unclear but key threatening processes are thought to be at play. Early life stages (embryos, larvae and young juveniles) are restricted to heavily altered riverine habitats and face multiple anthropogenic threats, such as the intrusions of contaminants and salt, the introduction of exotic species and the installation of dams and water diversion pipes (NMFS, 2010, 2018). Conversely, there are few known threats (besides fisheries capture and ocean acidification) to green sturgeon during their marine life history phase, and life-stage-specific management guidelines are required for this species. A wealth of data exists on the ecophysiological constraints of green sturgeon, but much of this information is underutilized in recovery plans due to a lack of synthesis. Therefore, this review and meta-analysis synthesized data on green sturgeon ecophysiology published in peer-reviewed journals. A metaanalytical approach was opted to summarize the effects of prominent stressors [i.e. elevated temperatures, contaminants, salinity and low food availability (food restriction)] on several physiological traits [e.g. growth, thermal tolerance, swimming performance and heat shock protein (Hsp) expression] using quantitative methods. The aim of this study was threefold: (i) to characterize the variety of research output on green sturgeon, in terms of the field of study (e.g. conservation physiology, movement ecology, etc.), type of study (experimental, field or theoretical), life-history stage examined and number of concurrent stressors examined; (ii) to quantify the overall effect of prominent stressors on key physiological traits; and (iii) to highlight areas where sufficient data exist to inform recovery plans and identify knowledge gaps to direct future research efforts. Collectively, the information derived here will provide an example of how a quantitative synthesis of physiological data can inform conservation plans and future research actions.

Materials and Methods

Review protocol

A Web of Science (Thompson Reuters) search was performed to find publications containing green sturgeon or *Acipenser medirostris* as key words. These publications were refined to only include peer-reviewed research papers. The following data were extracted from all publications: scientific journal, publication year, geographic location, type of study (experimental, field or theoretical), stressor of interest, population (southern or northern DPS), life-history stage examined (embryos, yolk-sac larvae, larvae, juveniles, sub-adults or adults) and age (days post-hatch). For studies examining the effects of stressors on physiological responses the duration of stressor exposure (acute ≤ 2 weeks < chronic) was noted, along with the number of concurrent stressors examined (i.e. single- or multi-stressor study).

Meta-analysis

Additional metadata were extracted from studies examining the effects of elevated temperatures, salinity, food restriction and contaminants on green sturgeon physiology. Only manipulative experiments with controls were included. Studies on the impact of movement barriers (e.g. dams and water diversion pipes) on green sturgeon were generally not manipulative and measured incomparable response variables (e.g. movements of free-ranging individuals versus laboratory entrainment rates in diversion pipes), and data were insufficient for quantitative analysis. However, movement barriers present a major threat to green sturgeon (NMFS, 2010, 2018), so a qualitative assessment of movement barriers was included in this review. To enable effect size calculations, studies were only included if means, sample sizes and variance (standard error, standard deviation or confidence interval) were reported. In instances where means and variance were not reported in text, data were extracted from figures using ImageJ (v. 1.51 k, National Institute of Health, USA, http://imagej.nih. gov.i)—a validated and commonly used method (Schneider et al., 2012; Schindelin et al., 2015; Foley et al., 2018). For each stressor the following definitions for control groups were used: elevated temperatures (control = 16-19°C; treatment > 19°C); salinity (control < 3 ppt; treatment > 3 ppt), food restriction (control = unrestricted, ad libitum; treatment = restricted) and contaminants (control = no contaminant; treatment = contaminant). The effects of the stressors on the following response variables were examined: growth, Hsp expression, deformity rates, hatching success, swimming performance (U_{crit}) , critical thermal maxima (CTMax), mortality rates, whole-body contaminant burden, plasma osmolality and muscle water content. A minimum of four data points for each trait were required to perform analyses, and response variables that did not meet this criterion were considered data deficient (D.D.). Effect sizes were calculated as the

log-response ratio (lnRR),

$$LnRR = ln\left(\frac{\overline{X}_T}{\overline{X}_C}\right) = ln(\overline{X}_T) - ln(\overline{X}_C),$$

where \overline{X}_T and \overline{X}_C are the mean responses from the treatment and the control groups, respectively. The variance (ν) of each ln-transformed response ratio was also calculated,

$$v = \frac{\left(SD_T\right)^2}{N_{T \,\overline{X}_T^2}} + \frac{\left(SD_C\right)^2}{N_{C \,\overline{X}_C^2}},$$

where SD and N are the standard deviation and sample size of the treatment and the control groups. Log response ratios were chosen over other methods because response ratios calculate the proportional change between the control and the treatment groups. Log response ratios are also robust to small sample sizes, less sensitive to data of different units and normalize data that are not normally distributed (Lajeunesse, 2011; Viechtbauer, 2010). Separate random effects models were used to calculate the overall effect size for each stressor (elevated temperature, salinity, food restriction and contaminants). Random effects models account for within- and between-study variation, and effect sizes were considered significant if 95% confidence intervals did not overlap with zero. Log response ratios and variances were calculated using the escalc function of the metafor package (Viechtbauer, 2010) in R Studio (version 1.0.136).

Publication bias

Contour-enhanced funnel plots of observed effect sizes were used to test for publication bias using the funnel function in *metaphor* in R Studio (version 1.0.136). Regression tests were used to test for asymmetry in the funnel plot using the regtest function (Egger et al., 1997), and publication bias was inferred if the regression slope was significantly different from zero. If asymmetry was detected, the trimfill function was used to determine the number of studies needed to obtain symmetry. The *trimfill* function assesses the sensitivity of the results to publication bias by 'filling in' the number of missing studies to obtain symmetry (Møller and Jennions, 2001). Lastly, Rosenberg's fail-safe numbers were calculated and represent the number of papers that would need to be published with non-significant results to change the mean effect size to a non-significant result (Rosenberg, 2005). Failsafe numbers are considered robust to publication bias when larger than 5n + 10 (where n is the number of studies already included in the meta-analysis; Rosenberg, 2005).

Results

A total of 105 publications were found containing green sturgeon or *Acipenser medirostris* as key words (Supplementary

material). Research on green sturgeon spans multiple disciplines (Fig. 1A), with a primary focus on conservation physiology (23.8%), movement ecology (17.1%), and basic physiology (7.6%). Mixtures of experimental, field and theoretical research approaches have been employed (Fig. 1C). Data on juvenile (38.1%) and adult (35.2%) life-history stages of green sturgeon are predominant, and less data are available for the early life-history stages (embryonic, 11.4%; yolk-sac larvae, 12.4%; and post yolk-sac larvae, 16.2%; Fig. 1B). The effects of stressors on physiological responses have included both acute (38.8%) and chronic (61.2%) responses; however, most data have been collected from single stressor studies and only 6 out of 70 experimental studies (~8.6%) examined the combined effects of multiple stressors. Thermal stress has received the majority of investigation, although research efforts (# publications) have been distributed evenly across stressors of interest (i.e. contaminants, salinity, food restriction and movement barriers) (Fig. 1D). All experimental studies were conducted on the northern DPS.

Meta-analysis

Elevated temperatures

The results of the random effects model show that elevated temperatures significantly reduce growth and hatching success and increase the incidence of larval deformities (Fig. 2A). Exposure to elevated temperatures also increases the expression of Hsp. There were insufficient data to assess the impact of elevated temperatures on swimming performance. Regression tests indicated that there was no significant asymmetry in the random effects models for growth, Hsp expression and deformity rate. However, significant asymmetry was detected in response to hatching success (Supplementary Table I). The trimfill method revealed that two data points were augmented in order to obtain symmetry. Rosenberg's fail-safe numbers were robust (ranging from 489 to 4439) for all response variables, indicating that a large number of non-significant results are required to alter the conclusions (Supplementary Table I).

Salinity

On average, exposure to elevated salinity levels negatively affects growth (Fig. 2B). Further, plasma osmolality and muscle moisture are significantly increased in response to salinity exposure. There were insufficient data to assess the impacts of salinity on expression of Hsp and swimming performance. Asymmetry in the regression tests for growth, plasma osmolality and muscle moisture were not detected, indicating no publication bias (Supplementary table I). Rosenberg's fail-safe numbers are robust for the plasma osmolality (1335) and muscle moisture (377) datasets, but not for growth (12).

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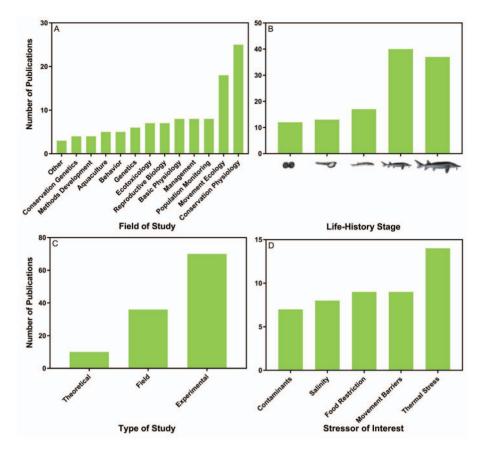


Figure 1: Summary of the research output on green sturgeon, (Acipenser medirostris) in terms of (A) field of study, (B) life-history stage examined, (C) type of study and (D) type of stressor examined

Food restriction

Growth was significantly reduced by food restriction (Fig. 2C), but food restriction does not affect the upper thermal tolerance (CTMax) or expression of Hsp. Although mortality rates tend to increase and swimming performance decrease with food restriction, there were insufficient data to assess its impact. Publication bias was not detected for the growth data set, as indicated by the regression test and Rosenberg's fail-safe number (498; Supplementary table I).

Contaminants

Exposure to environmental contaminants (carbaryl, methylmercury and selenium) significantly reduced growth and increased the body burden and mortality rates of green sturgeon (Fig. 2D). There were insufficient data to assess the impact of contaminants on the swimming performance and expression of Hsp. Regression tests indicated that there was no significant asymmetry in the random effects models for growth, mortality and body burden (Supplementary table I). Rosenberg's fail-safe numbers were robust to publication bias for the mortality (167) and body burden (244) datasets, but not for growth (5).

Discussion

Green sturgeon research has been multi-disciplinary, with a primary focus on conservation physiology. The meta-analysis revealed that all examined stressors significantly impaired green sturgeon growth, and additional stressor-specific costs were identified. These findings can be used to tailor management actions to meet the ecophysiological requirements of green sturgeon and plan future research efforts. Here we review and discuss the impact of prominent stressors (i.e. thermal stress, food restriction, elevated salinity, contaminants and movement barriers) on each green sturgeon life-history stage, identify knowledge gaps and suggest science-based management actions.

Thermal stress

As ectotherms, green sturgeon body temperature is closely tied to the external environment and thermal perturbations are particularly salient due to the thermal sensitivity of molecular, cellular and metabolic processes (Hochachka, 1967; Hochachka and Somero, 1968). The thermal sensitivity of key traits, such as swimming performance, can be

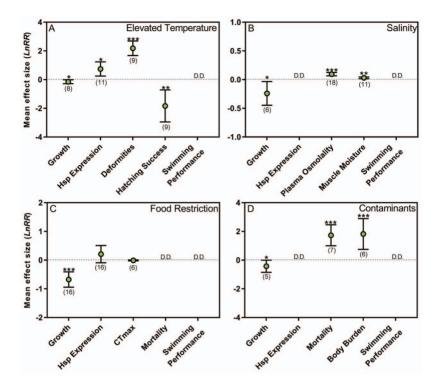


Figure 2: The impact of (A) elevated temperatures, (B) salinity, (C) food restriction, and (D) contaminants on several physiological traits in green sturgeon (*Acipenser medirostris*) from random effects meta-analyses. Data are presented as mean effect size (log ratio response, *LnRR*) and 95% confidence intervals. Sample sizes in each analysis are included in parentheses. The horizontal zero (dotted) line indicates no effect, and mean effect sizes are considered significant when 95% confidence intervals do not overlap with the zero line as denoted by an asterisks ($^*P < 0.05$, $^*P < 0.01$, $^*P < 0.001$). D.D. represents response variables with less than four data points for comparison. See main text for abbreviations.

described using thermal performance curves (TPCs), where an ectotherms' performance is modelled as a function of environmental/body temperature (Huey and Stevenson, 1979; Huev et al., 2012). Ectotherm TPCs are generally bell shaped; performance is optimized within a narrow thermal window (Topt) and performance decrements are observed as temperatures deviate towards low and high ends. Performance is entirely reduced once temperatures reach low and high critical limits-critical thermal minima (CTMin) and CTMax, respectively. However, the biological relevance of CTMin can be limited in high latitude, cold-water fishes because CTMin data are often censored at 0°C, even though eggs of some species can successfully develop at near freezing temperatures (Beitinger and Bennett, 2000; Beitinger et al., 2000). Anthropogenic manipulations of water temperatures can expose fish to stressfully low and high temperatures, where performance or survival may be compromised (Caissie, 2006). Water project operations (e.g. dams and water diversions) can alter natural thermal regimes in green sturgeon habitat (Cloern and Jassby, 2012). For example, cold water releases from thermally stratified dams can abruptly reduce water temperatures by 10-15°C (Pike et al., 2013), and temperatures are predicted to be altered even further under global climate change, with predictions of overall warmer river temperatures and intensified drought conditions (Cloern *et al.*, 2011).

Research has shown temperature to have pervasive effects on green sturgeon physiology (Mayfield and Cech, 2004), but temperature sensitivity differs depending on life-history stage, as seen in other fishes (Komoroske *et al.*, 2014). Further, fishes generally do not experience thermal stress passively and compensatory responses, such as the upregulation of Hsp and seasonal acclimatization can alleviate thermal stress (Prosser, 1991; Lindquist and Craig, 1988; Fangue *et al.*, 2011; Dalvi *et al.*, 2017). A life-history stage-specific overview of studies that examined the effects of temperature on green sturgeon is provided below.

Embryos

Temperatures experienced during fish embryogenesis can shift developmental trajectories, influence recruitment rates and have persistent effects on subsequent life-history stages (Martell and Kieffer, 2007; Schnurr *et al.*, 2014). Experimental assessments of the effects of water temperature on embryo development in green sturgeon are lacking and only a single study has been published. Van Eenennaam

et al. (2005) assessed the effects of constant incubation temperatures, ranging from 11°C to 26°C, on embryo survival and development in three progenies. Embryos were obtained using artificial tank spawning with three mature females and five males, captured in the lower Klamath River by the Yurok Tribe (Van Eenennaam et al., 2001). Hatching success was greatest between 14°C and 20°C, but the incidence of deformed embryos increased at temperatures >17.5°C. Elevated temperatures were particularly severe, and 100% mortality was observed at 23-26°C, although upper temperature tolerance differed slightly between progenies. The lower thermal tolerance limit was not identified, but hatching success was reduced at 11°C and hatched embryos were shorter. These findings suggest that 14-16°C is the optimal thermal range for successful development of green sturgeon because temperatures outside this range are associated with deleterious effects (i.e. reduced hatching success, higher deformity rates and smaller body sizes), but this needs to be replicated with more progenies and from females at different times of the spawning run, as maternal thermal history may influence progeny thermal tolerance (Le Roy et al., 2017). In field studies, embryos at early stages of development have been collected using egg mats in the upper Sacramento River at 13.9°C (Brown, 2007) and between 11.3°C and 15.8°C (Poytress et al., 2015), and in the Feather River between 16°C and 17°C (Seesholtz et al., 2015). These observations fall within the broad range indicated by laboratory studies, although they suggest that spawning sturgeon may be required to accept less than optimal temperatures based on the available environmental conditions. No studies to date have examined temperature influenced differences in development in wild embryos.

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Further research is required to understand the mechanisms underlying the emergence of embryonic abnormalities and mortality at unfavourable temperatures. Stage-specific assessments of abnormalities (e.g. at cleavage and neurulation) may provide insight into which developmental processes are most affected by suboptimal temperatures. For example, cleavage is one of the most heat-sensitive stages during embryogenesis in actinopterygian fishes, likely due to an exhaustion of maternally supplied Hsps (Motani and Wainwright, 2015). Indeed, abnormal cleavage in beluga sturgeon (Huso huso) has been observed at elevated temperatures (Dettlaff et al., 1993). Embryo thermal tolerance is also modulated by egg quality, which is underscored by several factors, including stress exposure (e.g. hypoxia and contaminants), final ovarian stage of maturity, post-ovulatory ageing and egg handling (Bobe and Labbé, 2010). Green sturgeon fecundity is lower than white sturgeon, as they have much larger egg size, and consistent low to moderate fertilization rates have been observed in green sturgeon eggs over several years of artificial spawning induction (Van Eenennaam et al., 2001, 2005, 2008, 2012). Of 23 females that ovulated in these studies, only four (17% of females) had eggs with fertility rates >80%, in comparison to most (\sim 75%) white sturgeon, which routinely have >80% fertility rates (Van Eenennaam et al., 2004). More research is required to characterize the thermal requirements of green sturgeon during embryogenesis and understand how additional stressors (e.g. hypoxia, contaminants and salinity) interact with thermal stress to affect egg quality and subsequent embryo survival.

Larvae

The impact of thermal stress on larval survival, growth and development has been the focus of several investigations. Elevated temperatures (>18°C) are particularly threatening to newly hatched larvae. Linares-Casenave et al. (2013) found survival of newly hatched larvae (hatch to yolk-sac depletion, Stage 45) to significantly decline between 22°C and 26°C and 100% mortality was observed at 28°C. An increase in notochord deformities (kyphosis) occurred at sub-lethal temperatures (20–26°C), and deformed larvae exhibited spiral swimming, which would likely compromise movements critical for predator avoidance and microhabitat selection. Thermal tolerance differed slightly between progenies, and heightened thermal tolerance was associated with an overexpression of Hsp60 and Hsp90.

Werner et al. (2007) conducted a similar study, where the capacity of newly hatched larvae to tolerate and recover from acute thermal stress was assessed. Larvae were exposed to one of three temperature treatments until yolk-sac absorption: constant 17°C (control), exposed to 26°C for 3 days and returned to 17°C or constant 26°C. Overall, deformity rates were lowest in larvae continuously maintained at 17°C and highest in larvae continuously exposed to 26°C. In the recovery treatment, ~33% of larvae exhibited deformities after 3 days of exposure to 26°C, but this rate halved once larvae were returned to cooler water, demonstrating some capacity to recover from thermal spikes. Deformed larvae also exhibited modified Hsp expression, where Hsp60 levels were lower but Hsp72 and Hsp78 levels were higher compared to healthy larvae.

Post yolk-sac larvae appear more resilient to elevated temperatures than newly hatched larvae. Allen *et al.* (2006b) assessed the effects of elevated and cycling thermal regimes (control 19°C; 19–24°C and 24°C) on growth rates in post yolk-sac larvae (15 days post-hatch, dph). Elevated and cycling thermal regimes had no adverse effects on growth rates, and growth was highest in fish maintained at 24°C. Accelerated growth rates at 24°C were facilitated by increased food consumption, but it was noted that growth of wild larvae could be limited at elevated temperatures if prey is not abundant.

There are few reported observations of larval green sturgeon in field studies; however, some yolk-sac larvae have been collected in the upper Sacramento River above Bend Bridge (Rkm 415) at temperatures of 13.2°C (Brown, 2007) and 12.9 ± 1.0 °C (Poytress *et al.*, 2015). While these temperatures are substantially lower than those at which developmental abnormalities have been observed in lab studies, it should

be noted that water temperatures in this reach are held at <13.3°C by Order 90-5 of the California State Water Resources Control Board for the benefit of winter-run Chinook salmon. It remains unclear what temperature larval green sturgeon seek out or prefer in natural field conditions.

Juveniles

More is known regarding the thermal requirements of juvenile green sturgeon compared to other life-history stages. A comprehensive study by Mayfield and Cech (2004) evaluated the thermal sensitivity of food consumption and conversion efficiency, growth rates, oxygen consumption, ventilatory frequency, volitional activity, thermal preference and swimming performance in juveniles. These data were collated to provide a temperature target for optimal bioenergetic performance. Bioenergetic performance was estimated to be optimal between 15°C and 19°C, as energetic demands increased beyond sustainable limits at higher temperatures. Swimming performance was compromised at 24°C in age-1 juveniles. suggesting that unseasonably high temperatures may impede critical movements, such as outmigration, predator avoidance and habitat selection. However, younger juveniles (155 dph) may be less vulnerable to high temperatures, as Allen et al. (2006a) found swimming performance to be greater at 24°C compared to 19°C. Young juveniles (47 dph) have also been shown to respond to daily thermal fluctuations, with fish exposed to wide temperature fluctuations (11–21°C day⁻¹) able to maintain swimming performance across a wider range of temperatures and having improved growth compared to fish maintained at stable temperatures (Rodgers et al., 2018).

Hsp responses have been elicited in juvenile green sturgeon by both acute and chronic exposures to high temperatures (Allen et al., 2006a; Wang et al., 2013). Mucus appears to be the most sensitive tissue to Hsp70 induction and white muscle is the least responsive (Wang et al., 2013). Green sturgeon exhibited significantly lower Hsp70 responses compared to white sturgeon, suggesting green sturgeon may be more vulnerable to the damaging effects of elevated temperatures. Temperature can also modulate the stress response in juvenile sturgeon. Lankford et al. (2003) assessed the physiological stress response (i.e. plasma cortisol and lactate concentrations) of age-0 fish exposed to a 1-minute air emersion at 19°C (control temperature) and 11°C. The magnitude of the stress responses did not differ between temperatures, but recovery was significantly prolonged at 11°C, providing evidence that juvenile sturgeon may therefore be more susceptible when recovering from stress at cold temperatures.

Sub-adults, adults and spawning adults

Very little is known about the thermal requirements of adult green sturgeon. During the peak Klamath River spawning migration (April–May) river temperatures generally range over 8–15°C under normal precipitation years (Van Eenennaam *et al.*, 2006). Spawning migrations begin earlier and are truncated during dry years when water temperatures

are elevated and flows are reduced (Van Eenennaam et al., 2006). Ripe northern DPS adults have also been reported in the Rogue River or when water temperatures ranged between 9.3°C and 18°C (Erickson and Webb, 2007). Similarly, spawning of the southern DPS in the Sacramento River occurred at temperatures ranging between 9.6°C and 17.6°C, with a daily mean \pm SD of $13.5 \pm 1.0^{\circ}$ C, and temperatures ranged between 16°C and 17°C during spawning in the Feather River (Poytress et al., 2015; Seesholtz et al., 2015). Additionally, when sub-adults were manually tracked in the San Francisco Estuary, they selected temperatures ranging from 14.5°C to 20.8°C as measured in the water column at the depths at which they were swimming (Kelly et al., 2007). Adult green sturgeon have been tracked along the coast of Oregon and in Willapa Bay and at temperatures ranging from 9.5°C to 21.9°C (Huff et al., 2011; Moser and Lindley, 2007).

Remote recordings of water/body temperature of free-ranging adult green sturgeon would provide valuable thermal preference data. An implantable multi-channel biotelemetry system for measuring blood flow and body temperature has been tested in sub-adult green sturgeon (age 3; Grans *et al.*, 2009). This system allowed the calculations of cardiac output as a function of heart rate and stroke volume during an acute thermal challenge. Temperatures were increased from 19°C to 26°C, and a moderate increase in cardiac output was observed as temperature increased. This methodological advance offers the possibility of collecting important physiological data on free-ranging adults, and interactions between therma l preference and sex, age, body size or reproductive status may be identified.

Management actions and data gaps

Temperature ranges where physiological performance is optimal differs between life-history stages (Fig. 3) and management actions should be geared towards the most vulnerable stage. Early developmental stages (embryos–yolk-sac larvae) are highly sensitive to thermal perturbations and only narrow thermal windows (embryos, 14-16°C; yolk-sac larvae, 16-18°C) ensure survival and growth are optimized and deformities are minimized (Fig. 3). Physiological performance is maintained across a wider breadth of temperatures in post yolk-sac larvae and juveniles, but the thermal requirements of adults remain largely uncharacterized (Fig. 3). The overall mean effect of elevated temperatures (>19°C) on green sturgeon physiology are shown in Fig. 2A. Growth and embryo hatching success are significantly reduced at elevated temperatures, likely owing to metabolic demands exceeding energy supply (Fig. 2A). The incidence of deformities increases at elevated temperatures, along with increased expression of Hsps (Fig. 2A). Insufficient data exist on the thermal sensitivity of swimming performance, despite this trait representing an important fitness correlate (Plaut, 2001).

Ensuring water temperatures are maintained within optimal ranges during peak spawning periods (April–May) in critical rearing habitat is a potential management strategy.

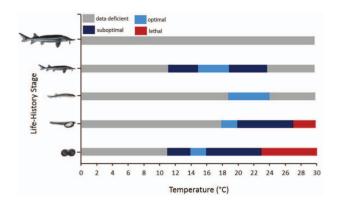


Figure 3: Life-history stage specific thermal ranges of green sturgeon (*Acipenser medirostris*) where survival and performance (growth, swimming performance and normal development) are optimal (light blue) and suboptimal (dark blue). Lethal temperatures are red and grey bars represent D.D. temperatures. Life-history stages are represented in order as embryos, yolk-sac larvae, post yolk-sac larvae, juveniles and adults. Data were extracted from peer-reviewed publications (*SI*) on the northern DPS on full feed rations.

Maintaining water temperatures below 20°C may benefit green sturgeon, particularly during early development (Fig. 2A, Fig. 3); however, more data are required before specific thermal ranges can be recommended. An understanding of how cold temperatures (<11°C) influence green sturgeon physiology is lacking across all life-history stages (Fig. 3); however, a recent study evaluating the effects of temperature on growth performance found growth to cease at 11°C suggesting that temperatures at or below this point will likely result in time limited survival (Poletto et al., 2018). Hypolimnetic water releases from dams within the Sacramento-San Joaquin (S-SJ) watershed can expose green sturgeon to unseasonably low temperatures during critical spawning periods (Pike et al., 2013). Research efforts should aim to identify lower temperature limits for survival, growth and swimming performance across all life stages.

Thermal requirements for embryos and adults are poorly characterized and represent fruitful research avenues. Specifically, additional work is required to understand transgenerational effects on thermal tolerance (Rossiter, 1996). Temperatures experienced by parental fish can influence offspring phenotype and may facilitate transgenerational plasticity (Mousseau and Fox, 1998; Burt et al., 2011). Progeny of spawners from an early spawning migration of starry sturgeon (A. stellatus) had a narrower range of temperature tolerance than progeny of spawners from the late migration whose parents experienced warmer waters during migration (Sytina and Shagaeva, 1987). These authors reported an upward shift in the temperature limits of lethal, sublethal and suboptimal effects by 2-3°C in A. stellatus embryos. Moreover, inter-progeny variation in thermal tolerance of green sturgeon reported (Van Eenennaam et al., 2005; Linares-Casenave et al., 2013) may partially reflect transgenerational plasticity (Rossiter, 1996; Mousseau and Fox, 1998). Lastly, future experiments should be designed to maximize ecological relevance by mimicking natural thermal regimes and exposing fish to multiple heterologous stressors in unison. Almost all studies (c.f. Allen *et al.*, 2006b; Rodgers *et al.*, 2018) used constant temperatures, but natural river temperatures with daily fluctuations should be mimicked.

Salinity

Green sturgeon are truly anadromous and spend a large portion of their life in the ocean (Allen and Cech, 2007). Mature adults undertake long-distance, seasonal migrations between marine and freshwater habitats to spawn (Lindley et al., 2008). Eggs and larvae develop in freshwater and juveniles may remain in freshwater habitats for up to 3 years prior to outward migration (Nakamoto et al., 1995). Juvenile sturgeon undergo physiological remodelling in preparation for seaward migrations (Allen et al., 2009, 2011). The timing of this physiological remodelling is age and size dependent and occurs at a young age relative to other sturgeons (Allen et al., 2009). Exposure to salinity prior to physiological remodelling can disrupt osmotic homeostasis (e.g. plasma osmolality and muscle moisture; Fig. 2B) and increase energetic costs through the upregulation of ionoregulatory mechanisms (Boeuf and Payan, 2001). Salinity levels within the San Francisco Bay Delta (SFBD) are forecasted to increase under climate change due to sea-level rise, seawater intrusion and altered precipitation patterns (Cayan et al., 2008). Therefore, recent research efforts have focused on understanding the timing of physiological remodelling, behavioural responses to salinity gradients and the costs associated with salinity exposure in green sturgeon. Data on the effects of elevated salinity on early life stages (embryonic and larval stages) are currently unavailable.

Juveniles

Research has been geared towards understanding the timing of physiological adjustments associated with seawater acclimation in juveniles. Findings have shown that both age and body size modulate the timing of seawater acclimation (Allen and Cech, 2007). Short-term exposure (72 h) to elevated salinity (20-33 ppt) during early life (<100 dph) causes profound declines in survival (Allen et al., 2011). However, juveniles can tolerate brackish (10 ppt) to full seawater (33 ppt) when more developed (i.e. 170 dph, ~15 g) but at the cost of reduced growth (Allen and Cech, 2007; Allen et al., 2011) resulting from the disruption of ionic homeostasis (increased plasma osmolality and muscle moisture; Fig 2B). At 1.5 years of age (1.5 kg) juveniles show complete seawater acclimation with no costs evident (Allen and Cech, 2007; Sardella and Kültz, 2009). Juvenile green sturgeon ranging in size from 19 to 58 cm TL were captured by gill net and otter trawl in the freshwater region of the Sacramento-San Joaquin Delta near Sherman Island (Radtke, 1966). It remains unclear at what point in development this species moves into saline waters in the wild, but sub-adult fish of ~1 m TL were captured and

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tracked in the San Francisco Bay Estuary in water ranging from 8.8 to 32.1 ppt (Kelly et al., 2007).

Physiological changes underlying seawater acclimation have been characterized and include increased gill and pyloric ceca sodium potassium-ATPase (NKA) activity, gill mitochondria-rich cell size, lamellar length, body condition, hematocrit and thyroid hormone levels (T₃ and T₄) (Allen *et al.*, 2009, 2011; Haller *et al.*, 2015). Behavioural responses to salinity have also been examined in juveniles, finding that at 196 dph juveniles display a preference for seawater regardless of acclimation salinity (Poletto *et al.*, 2013). These findings are consistent with the timing of the development of seawater tolerance and suggest juveniles actively seek out saline environments at this age.

Salinity tolerance can be modulated by the presence of additional stressors. Haller *et al.* (2015) found salinity tolerance to be significantly impaired when food was restricted in 240 dph juvenile sturgeon. Food-restricted fish showed altered hematological indexes, plasma parameters (osmolality, glucose, lactate and cortisol), and gill and pyloric ceca enzymatic activity. Similarly, Vaz *et al.* (2015) found combined exposure to seawater (16 and 32 ppt) and food restriction synergistically decreased body condition factor. Upper thermal tolerance (CTMax) can also change in response to environmental salinity (Sardella *et al.*, 2008). For example, CTMax increased 0.5°C in juveniles (58.4 \pm 2.5 g) acclimated to brackish water (10 ppt) compared to juveniles acclimated to freshwater or bay water (24 ppt), suggesting energy demands were lowest in brackish, near-isosmotic water.

Management actions and data gaps

Although it is appreciated that salinity in the SFBD is projected to increase under climate change, very little is known regarding the effects of salinity exposure on early life-history stages. Elevated salinity will likely impair hatching success and growth and development of larvae, but this has yet to be explored. Nonetheless, it is evident that young juveniles (<100 dph) experience heightened mortality rates in saline waters (Allen et al., 2011) and salt intrusion should be mitigated in nursery habitats. Salinity can be maintained at low levels by manipulating dam releases to increase river flows, termed 'flushing' (Liu and Liu, 2014). Moreover, climate change scenarios predict salinity levels to rise in tandem with water temperatures (Cayan et al., 2008), along with changes in other environmental variables (e.g. aquatic pH). Therefore, an understanding of the interactions among elevated temperatures, salinity, pH and other environmental stressors on the physiology of early life-history stages is essential for futurefocused management plans.

Food restriction

Low food availability is a pervasive stressor in developed waterways, and record declines in zooplankton biomass have been observed in the SFBD over the past four decades (Cloern et al., 1983; Carlton et al., 1990; Cloern et al., 2014; Winder and Jassby, 2011). Diminished food resources stem from exotic species, namely the overbite clam (Corbula amurensis), and flow regulation modifying food-web structure and lowering primary productivity (Cloern et al., 1983; Cloern and Jassby, 2012). Food scarcity can disrupt the energy budget of fish and lower stress tolerance via a divestment of energy towards protective molecular and cellular responses (Sokolova, 2013). Lower stress tolerance decreases fish resilience to additional stressors such as elevated temperatures or salinity. However, the most well-documented effect of food restriction is a decrease in somatic growth (Jobling, 1994). Slowed growth rates, particularly during early life stages, can have important consequences because smaller body sizes often confer increased mortality rates (Carlson et al., 2008) and reduced competitiveness over resources (Johnsson et al., 1999). The effect of food restriction on growth, development and susceptibility to additional stressors have been examined in green sturgeon, although studies are skewed towards juveniles and very little is known about food preferences in the wild at any life stage.

Larvae

Green sturgeon larvae typically initiate exogenous feeding at 10–16 dph, but water temperabuture modulates yolk-sac absorption rates and time to exogenous feeding (Van Eenennaam

et al., 2001). The digestive tract of sturgeon develops during the larval stage and is sensitive to changes in food availability (Gisbert and Doroshov, 2003). Gisbert and Doroshov (2003) found food deprivation in green sturgeon larvae (hatching—31 dph) to cause pathological changes after 5 days and significant deterioration of the digestive system after 10–15 days, evident as tissue degeneration, necrosis and shrinkage of digestive epithelia. These findings suggest there is a critical period of 4–5 days following hatching where larvae must feed to ensure the healthy development of their digestive system.

Juveniles and adults

Food restriction can impact growth rates, body condition, energy stores and stress tolerance in juvenile green sturgeon. Intuitively, food restriction decreases growth rates (Fig. 2C), feeding efficiency, body condition as well as whole-body lipid and protein stores (Haller et al., 2015; Zheng et al., 2015; Vaz et al., 2015; Verhille et al., 2015; Poletto et al., 2018), but effects on stress tolerance are less uniform. Haller et al. (2015) found that food restriction impaired osmoregulation capacity in juveniles (222 dph), and exposing food-restricted fish to full-strength sea water (32 ppt) resulted in high mortality rates within 72 h. Other studies have investigated the effect of food restriction on heat tolerance and Hsp expression (Wang et al., 2013; Zheng et al., 2015; Lee et al., 2016; Verhille et al., 2015). Findings have been mixed, but overall food restriction does not significantly affect upper thermal limits (CTMax) or the expression of Hsps (Fig. 2C).

Management actions and data gaps

Overall, food restriction is known to have irreversible impacts on the development of the digestive tract in young larvae and affect growth rates and energy stores in juveniles and compensatory growth following periods of food restriction were not seen in a recent study (Poletto et al., 2018). Restoration of natural food-web dynamics may alleviate the threat of food scarcity. Reversing disrupted food-web linkages in the SFBD may be achieved by restoring natural water flow regimes and controlling invasive primary consumers that disrupt the food-web from its base (Cloern et al., 1983; Strayer, 2010). Research on the effects of food restriction in green sturgeon and other fishes is still in its infancy. No empirical data exist on the effect of food restriction on green sturgeon adult body condition, but bioenergetic models suggest spawning frequency may be reduced under low prey availability scenarios (Borin et al., 2017). It is recommended that future research efforts explore the effects of food restriction across all life-history stages to further our understanding of the impacts of food restriction on a range of factors including disease susceptibility, contaminant uptake rates and reproductive investment. Dietary preferences of green sturgeon in the wild across all life stages should also be investigated in order to both identify critical habitat for management and understand the impacts of food restriction on bioenergetics.

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Contaminants

The SFBD is listed as an 'impaired waterway' under the US Clean Water Act because many contaminants exceed regulatory thresholds (SWRCB, 2010). Contaminants enter the SFBD from a variety on inputs, including municipal and industrial wastewater, urban and agricultural runoff, and direct pesticide application (Fong et al., 2016). Contaminants associated with this listing include heavy metals (e.g. mercury, cadmium, copper and zinc), pesticides (e.g. chlordane and organophosphate insecticides) and chlorinated compounds (e.g. polychlorinated biphenyls and dioxins) (Fong et al., 2016). Recent studies by Moschet et al. (2014) targeting pesticides and pharmaceuticals detected 17 common compounds across 51 samples, highlighting risks of potential synergism resulting from exposure to a multitude of co-occurring contaminants. Uptake of contaminants in fishes can occur through aqueous exposure (e.g. branchial exposure during gill ventilation) and dietary exposure (consuming contaminated prey) (Arnot and Gobas, 2006). Contaminant exposure can disrupt organismal functions across all levels of biological organization, from cellular mechanisms to whole-animal performance, impacting reproduction and population dynamics. Research investigating the effects of contaminants on green sturgeon is limited to a small number of studies examining the effects of exposure to selenium, carbaryl and methylmercury, a legacy contaminant in the SFBD. Selenium is an essential micronutrient to all vertebrates but is toxic at higher concentrations and aquatic organisms are particularly vulnerable (Janz et al., 2010). Exposure to elevated selenium levels has been shown to result in developmental abnormalities and skeletal deformities in multiple fish species (Holm et al., 2005; Shi et al., 2018; Kupsco and Schlenk, 2016). Agricultural runoff over seleniferous soils and water discharge from industrial operations are the main sources of excessive selenium inputs in the SFBD (Lemly, 2004). The SFBD also suffers from high levels of mercury (Hg) contamination, due to intense hydraulic gold mining operations from 1850 to 1960 (Dasmann, 1999). Once in the water, bacteria and fungi convert inorganic Hg into methylmercury (MeHg), a highly toxic form, owing to its high bioavailability and affinity to bind to thiol groups (Harris et al., 2007). Exposure and responses to numerous contaminant classes vary depending on life-history stage (Brooks et al., 2012), as well as with physicochemical properties such as temperature (Maulvault et al., 2016) and salinity (Smylie et al., 2016), and management actions need to be tailored accordingly. The primary studies evaluating the effect of contaminants on sturgeon address MeHg and selenomethionine (SeMet). There is a lack of information regarding the effects posed by a multitude of contaminants detected in the SFBD.

Embryos

Early life-history stages are generally most susceptible to the detrimental effects of contaminants (McKim, 1977; Foekema et al., 2012). Embryos can be exposed to contaminants through direct exposure to water and contact with sediment or via maternal transfer (Niimi, 1983). Green sturgeon embryogenesis occurs during a period of direct contact with sediment, which is potentially laden with multiple contaminants. Sediments act as pollutant sinks and release contaminants back into the water column following disruption or bacterial activity, thus the sedimentwater interface is an area of highly concentrated pollution. Despite green sturgeon embryos being at a high risk of contaminant exposure, no studies have assessed the effects of contaminant exposure in this life stage. In other fishes, exposure to contaminants during embryogenesis has led to increased mortality, slowed development, increased heart rates and dysfunctional lateral lines (Johnson et al., 2007; Vardy et al., 2013). Chorionic permeability is known to differ greatly between fish species and does not necessarily provide sufficient protection from anthropogenic contaminants. Studies conducted by Weis and Weis (1989) have demonstrated differences in adsorption of methylmercury by the chorion vs. developing embryos, correlating with population tolerance in killifish, (Fundulus heteroclitus). Green sturgeon embryos may also be exposed to maternally transferred contaminants. For example, studies have reported selenium concentrations in ovaries and ovulated eggs sampled from white sturgeon in the SFBD at levels that far exceed reproductive toxicity thresholds (Kroll and Doroshov, 1991; Lemly, 1996; Linares-Casenave et al., 2015). Several reviews and expert panels have highlighted the need to conduct studies on the effects of multiple contaminants on green sturgeon

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embryos (Johnson *et al.*, 2010; Brooks *et al.*, 2012; IEP MAST, 2015; Fong *et al.*, 2016), as exposure may contribute to recruitment failure.

Larvae

Sturgeon yolk-sac larvae take refuge in sediments prior to the onset of exogenous feedings and are likely to be chronically exposed to high concentrations of contaminants (Richmond and Kynard, 1995; McAdam, 2011). Many chemicals are lipophilic and thus will adsorb to the yolk-sac and be dispersed throughout the developing larvae. Yolk-sac larvae may be particularly vulnerable to contaminant exposure because they are energy limited (yolk-sac dependent) and energy may be redirected from growth and organogenesis towards enzymatic detoxification of contaminants, repair mechanisms and toxicant elimination (Brooks et al., 2012). Once exogenous feeding initiates, larvae spend less time in the sediment but contend with dietary intake of contaminants through prey consumption. Very little is known regarding the effects of contaminants on green sturgeon larvae, and only a single study on post yolk-sac larvae has been published. Silvestre et al. (2010) examined the effects of selenium exposure on survival, morphology and protein expression profiles in green and white sturgeon. Selenium stress (body burden of 8 µg g⁻¹) induced developmental abnormalities (i.e. peritoneal and pericardial oedemas and spinal deformities), altered behaviour (i.e. lethargy) and physiological changes (i.e. bradycardia and internal haemorrhaging). Overall, mortality rates and mechanistic responses, determined through proteome evaluations, in selenium-exposed fish were higher in green compared to white sturgeon. These findings suggest green sturgeon exhibit greater sensitivity to contaminant exposure; although it should be noted that additional research is needed as Silvestre et al. (2010) only tested one family exposed to a single contaminant.

Juveniles

More is known regarding contaminant exposure in juvenile green sturgeon compared to other life-history stages. Studies have focused on the effects of dietary exposure of l-SeMet and methylmercury (MeHg) on growth, tissue burden and histopathology (Lee et al., 2011, 2012; De Riu et al., 2014). De Riu et al. (2014) exposed juvenile green and white sturgeon to four levels of dietary SeMet (0, 50, 100 or 200 mg SeMet kg⁻¹) for 8 weeks. SeMet diets were associated with reduced body lipids, reduced growth, kidney and liver abnormalities and increased mortality rates in a dosedependent pattern. Of concern, even the low SeMet diet, which closely reflected concentrations found in prey items of juvenile sturgeon (benthic macro-invertebrates), caused detrimental effects. Overall, the adverse effects of dietary SeMet were more pronounced in green compared to white sturgeon, providing another line of evidence suggesting green sturgeon are more susceptible to contaminants. In a similar study, juvenile green sturgeon were exposed to one of four

dietary concentrations of MeHg (0, 25, 50 and 100 mg MeHg kg⁻¹) for 8 weeks (Lee et al., 2011, 2012). Methylmercury burden was detected in all assessed tissues, and highest concentrations were detected in the kidney. Green sturgeon had a higher MeHg tissue burden in the liver relative to white sturgeon, but tissue burden was higher in all other white sturgeon tissues. Despite this, dietary MeHg exposure had greater negative effects on survival, growth and development in green sturgeon compared to white sturgeon. The disparity in responses to dietary contaminants in green and white sturgeon suggest that the two species have different functional or structural capabilities in dealing with contaminant (e.g. uptake, detoxification and elimination capacities) and show that white sturgeon is an inappropriate surrogate species for green sturgeon (Lee et al., 2011, 2012). There are no additional toxicity data for green sturgeon, beyond the limited MeHg and SeMet studies detailed above.

Sub-adults and adults

Sub-adult and adult life stages of green sturgeon may be less exposed to contaminants because the marine environment is more distant from pollutant sources than freshwater systems (Vörösmarty et al., 2010). However, adults forage in polluted habitats during obligate migratory movements (Moser and Lindley, 2007) and the longevity of green sturgeon makes adults vulnerable to bioaccumulation and biomagnification of toxins. Indeed, Se tissue burden appears to increase with age in white sturgeon, suggesting significant bioaccumulation occurs prior to reaching sexual maturity (Linares-Casenave et al., 2015). Stomach content analyses show that adult green sturgeon feed on two species of burrowing shrimp (Neotrypaea californiensis and Upogebia pugettensis) in Willapa Bay and Grays Harbor (WA, USA)—major areas of commercial oyster cultivation (Dumbauld et al., 2008). Burrowing shrimp are considered a pest because they impair oyster growth and survival, and widespread applications of imidacloprid pesticides have occurred to control shrimp abundance. Concerns that green sturgeon are exposed to imidacloprid via shrimp consumption and interactions with sediment porewater prompted Frew et al. (2015) to conduct a study to characterize imidacloprid exposure. Sturgeon feeding pits (shallow depressions) were counted before, during and after imidacloprid applications at 24 field sites and compared to reference sites where no imidacloprid applications occurred. Results suggested that green sturgeon preferentially fed on imidacloprid-impaired shrimp as the number of feeding pits following a pesticide application was 78% higher compared to reference sites. Total imidacloprid uptake, from both branchial and dietary intake, was also modelled and it was estimated that porewater exposure contributes more than shrimp consumption (Frew et al., 2015). Concentrations and durations of imidacloprid exposure were suggested to be lower than toxic thresholds; however, these estimates were based on toxicity assessments in white sturgeon (Frew and Grue, 2015) and several studies have demonstrated the unsuitability of white sturgeon as a surrogate species

(Silvestre *et al.*, 2010; Lee *et al.*, 2011, 2012; De Riu *et al.*, 2014). Similarly, the effect of another pesticide used to control burrowing shrimp, carbaryl, has been studied in white sturgeon as a surrogate species (Troiano and Grue, 2016) but this work provides little information on the vulnerability of green sturgeon.

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Management actions and data gaps

Overall, certain contaminant classes appear to accumulate in green sturgeon tissues through chronic exposure, leading to significant increases in body burden and mortality (Fig. 2D). At sub-lethal levels, contaminants lower green sturgeon growth rates (Fig. 2D), potentially reflecting a disruption to energy homeostasis. Contaminant exposure may cause fish to divest energy into fitness-related activities, such as growth, predator avoidance and reproduction, to mitigate damage (Goto and Wallace, 2010). Gearing future research efforts to understand effect-based, sub-lethal physiological costs (e.g. impaired swimming performance or elevated maintenance costs) of contaminant exposure may effectively assess proximate causes of green sturgeon declines (Brooks et al., 2012). Data are lacking across all life-history stages, particularly during critical early life periods when contaminant sensitivity may be heightened (McKim, 1977). Measuring the effects of contaminant exposure along with other stressors (e.g. elevated temperatures, salinity and turbidity) or contaminant mixtures would better reflect ecologically relevant conditions. Additional stressors may influence contaminant uptake, accumulation, excretion and toxicity (Sokolova and Lannig, 2008). Once a greater understanding of green sturgeon susceptibility to a multitude of contaminant classes present in the SFBD is gained, beyond that of MeHg and SeMet, a life-history stage-specific management plan can be put into place, which explicitly considers temporal and spatial changes in habitat use and exposure to additional stressors.

Movement barriers

Green sturgeon declines have been linked to extensive fragmentation of spawning and rearing habitat, particularly in the S-SJ watershed (Cloern and Jassby, 2012). The S-SJ watershed has been extensively altered since the mid-1800s when hydraulic gold mining operations were pervasive and exponential human population growth followed (Dasmann, 1999). The S-SJ watershed currently supplies water to ~1 million hectares of agricultural land and over 25 million people (Service, 2007). To meet this demand and provide flood protection, tens of water reservoirs and over 3300 water diversion pipes have been constructed within the river system (Herren and Kawaski, 2001; Cloern and Jassby, 2012). These alterations are part of the two largest water diversion operations worldwide (State Water Project and the Central Valley Project).

Waterway infrastructure can impede fish movements by creating physical (e.g. dams), hydraulic (e.g. excessive water

velocities), physiochemical (e.g. low dissolved oxygen) and behavioural (e.g. low light levels) barriers. The survival and reproductive success of green sturgeon and other fishes are tied to their capacity to move freely and efficiently within the environment at all life stages (Fausch et al., 2002). Small-scale movements are vital for foraging and predator avoidance, whereas large-scale movements are necessary for reaching spawning grounds, juvenile outmigration, habitat selection and maintaining population connectivity (Clapp et al., 1990; Gowan and Fausch, 2002; Yamamoto et al., 2004). Successful navigation of instream barriers is related to fish swimming capabilities and behavioural responses to water flow (Adams et al., 2007; Boysen and Hoover, 2009). To this end, research efforts have characterized green sturgeon swimming capacity and coupled this with behavioural responses to water flow and instream structures. Moreover, the efficacy of retrofitting water diversion pipes with a range of deterrents and fish-protection devices has been assessed (Poletto et al., 2014b; Poletto et al., 2015). Together these findings can guide conservation efforts aimed at providing unimpeded passage of green sturgeon.

Water diversions

Water diversion pipes are designed to maximize hydraulic capacity and water extraction efficiency. Diversion velocities far exceed the swimming capacities of many fishes, and many are consequently drawn inside (i.e. entrained; Grimaldo et al., 2009). Entrained fishes can be physically damaged by pump operations or left stranded in irrigation fields (Baumgartner et al., 2009). Juvenile green sturgeon may spend up to 1.5 y within the S-SI watershed and likely encounter multiple water diversion pipes during daily movements and outmigration. Mussen et al. (2014) were the first to assess the susceptibility of juvenile green sturgeon (~35 cm FL) to entrainment in water diversion structures. Experiments were held in a large river-scale simulation flume and diversion flows were set to replicate field conditions. Entrainment rates were high, and fish were rapidly entrained (<1 s) after passing the diversion pipe. Extrapolations of findings predicted that ~22% of outmigrating juveniles could become entrained if they passed by a single water diversion and this number increased markedly with the number of diversions passed. However, entrainment rates were reduced dramatically by lowering water extraction velocities (from 0.57 to 0.28 m³ s⁻¹), suggesting that limiting inflow velocities may be a successful management strategy.

Building on this, Verhille *et al.* (2014) characterized the swimming capacity of larval and juvenile green sturgeon by measuring critical swimming speeds ($U_{\rm crit}$). Absolute swimming speeds increased with fish size and age, likely due to greater muscle mass and enhanced energetic efficiency (Beamish, 1978; Videler, 1993). However, 50 cm-long (TL) juveniles were exceptions to this trend and exhibited highly variable and lower-than-predicted swimming speeds. Verhille *et al.* (2014) noted that juveniles of this size typically develop

salt-water tolerance in preparation for outmigration, and this decline in swimming performance likely represented an energetic trade-off between physiological remodelling required for seawater acclimation and locomotor performance. Indeed, this finding has been corroborated with another study, where a decline in $U_{\rm crit}$ corresponded with the onset of hypersaline osmoregulatory capacities in similarly sized juveniles (Allen *et al.*, 2006a).

Ontogenetic changes in swimming performance highlight the need to specify limits on water diversion inflow velocities according to season and location within the S-SJ watershed. Entrainment risk is thought to be greatest at nighttime as larval and juvenile green sturgeon perform nocturnal foraging and migration movements (Kynard et al., 2005). Therefore, Verhille et al. (2014) suggested water diversion inflow limits only be enforced at nighttime. Larval fish were identified as the weakest swimmers and it was recommended that inflow velocities, within the mid-reaches of the Sacramento River, be restricted to ≤0.29 m s⁻¹ from May through summer, to match peak rearing times (Verhille et al., 2014). Young juveniles (\sim 55 dph; TL: 7 cm) were the strongest swimmers and it was suggested that nighttime water diversion inflow velocities are limited to ≤ 0.54 m s⁻¹ in the mid-reaches of the Sacramento River from July through May. Outmigrating juveniles (TL: 8-50 cm) were also identified to be susceptible to entrainment, owing to their weak swimming ability as they prepare for seawater entry. Therefore, it was recommended that nighttime diversion inflows are limited to < 0.40 m s⁻¹ in the middle and lower river reaches during peak movements (October-November). These data offer baseline guidelines that require further testing before implementation.

Taking a different approach, installing deterrents or fishprotection devices at water diversion pipes may lower fish entrainment rates without reducing hydraulic efficiency. The efficacy of two sensory deterrents, a strobe light and acoustic vibrations, in eliciting avoidance behaviour and minimizing entrainment has been assessed in juvenile green sturgeon (Poletto et al., 2014a,b). Both sensory deterrents proved ineffective and had no effect on entrainment rates, despite reports of success in other species (Maes et al., 2004; Hamel et al., 2008). Poletto et al. (2014b, 2015) also evaluated the efficacy of several physical fish-protection devices in lowering entrainment rates. Retrofit designs included a modified upturned pipe configuration and the addition of a protective screen, louvre box, trash-rack box or perforated cylinder. All retrofit designs significantly reduced entrainment rates. For example, the addition of a protective screen lowered entrainment rates from 44% in an unmodified pipe to 13%. The upturned pipe configuration allowed water to be withdrawn from the middle of the water column rather than the bottom. This retrofit design was particularly effective at reducing entrainment rates, likely owing to the benthic behaviour of green sturgeon. However, Poletto et al. (2014a) question the feasibility of this design because it can only be installed in water bodies with sufficient depth to avoid

navigation hazards by boats and could increase entrainment risk for pelagic species. Other retrofit designs (i.e. louvre box, trash-rack box and perforated cylinder) reduced entrainment risk by 60–96%. The effectiveness of these designs was postulated to be linked to physical protection, and inflow velocities being distributed across a greater area, enabling fish to detect flow changes from a greater distance and initiate avoidance behaviour.

Behaviour of juvenile green sturgeon near water diversions with protective screens has also been investigated. Although fish exclusion screens can reduce entrainment rates, fish can become impinged on screens (Poletto et al., 2014b). Screen contacts can injure fish and cause heightened levels of physiological stress (e.g. acidosis from exhaustion; Young et al., 2010). Poletto et al. (2014b) compared impingement rates between juvenile green and white sturgeon, finding that green sturgeon were impinged twice as often and spent more time near the screen. Impingement rates also varied according to fish size and age (Poletto et al., 2018). A comparison between three size classes of green sturgeon (small juveniles, 9.2 cm FL; intermediate juveniles, 18.8 cm FL; and older juveniles, 29.5 cm FL) identified a general negative trend between fork length and the number of body contacts with the screen. Intermediate-sized juveniles (18-20 cm) were exceptions and contacted the screens more frequently than smaller fish. This size range was correlated with the time when osmoregulatory capacity is developed (Allen and Cech, 2007; Allen et al., 2009), providing yet another line of evidence to suggest juveniles of this stage are particularly susceptible to water diversions.

Water reservoirs

Water impoundments (e.g. dams and weirs) can act as large-scale movement barriers and block access to critical spawning grounds (Bednarek, 2001). Adult green sturgeon migrate upstream to freshwater habitats to spawn and perform long-distance movements within freshwater reaches during the spawning season (Thomas *et al.*, 2014). Dams can impede upstream movements by creating physical blockades (e.g. closed gates) and hydraulic barriers in the form of excessive water velocities below flow-control gates (Brown, 2007). Fish ladders (i.e. fishways) have been installed at many dams to facilitate fish passage, but these structures are designed for salmonids with superior swimming abilities compared to green sturgeon (Mallen-Cooper and Brand, 2007).

To assess the effects of dams and altered flow regimes on the geographic distribution of green sturgeon, Mora et al. (2009) characterized habitat preference by correlating environmental characteristics (i.e. river discharge, water velocity, channel gradient and temperature) with over 2000 green sturgeon sightings. The habitat preference model was then validated using tracks of acoustically tagged freeranging adults. The model predicted that $\sim\!9\%$ of historically available habitat is now blocked by dams and that much of this habitat would have been suitable for spawning. Similarly,

acoustic tracking of free-ranging green sturgeon adults has shown different migratory behaviour based on seasonal operation of Red Bluff Diversion Dam (RBDD) gates in the Sacramento River, suggesting upstream movement was hindered (Heublein et al., 2009). Further, aggregations of green sturgeon near a water pumping facility (Glen Colusa Irrigation District pumping facility) suggest these facilities may influence spawning and post-spawn holding habitat and freshwater movements (Heublein et al., 2009). Several other partial or complete barriers to migrating adult green sturgeon exist throughout the Californian Central Valley landscape. Under normal flows the Sunset Pumps Weir in the Feather River limits migration to upstream spawning habitat. Multiple reports of adult green sturgeon at the base of Daguerre Point Dam in the Yuba River infers this structure also blocks sturgeon from upstream spawning habitat (Heublein et al., 2017). Upstream migrating sturgeon are also impeded by flood control structures in the Sacramento and Feather rivers. Adult green sturgeon are stranded near Fremont and Tisdale weirs in ponded water after flood flows recede on the Yolo and Sutter bypasses (Thomas et al., 2013; Heublein et al., 2017).

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Decommissioning non-essential water reservoirs may provide improved habitat connectivity. Steel *et al.* (2018) compared migratory movements of acoustically tagged adult green sturgeon before and after the decommissioning of RBDD in the Sacramento River. Prior to decommissioning the RBDD the proportion of tagged fish migrating upstream past the dam was linked to gate closures. When gate closures were delayed by a month, significantly more green sturgeon migrated above the dam. Following dam decommissioning, the proportion of fish successfully reaching spawning habitat above the dam increased. These findings suggest the RBDD was blocking movement to spawning grounds and highlight that decommissioning of dams can be an effective management action to reconnect critical habitat.

Management actions and data gaps

The studies reviewed here present strong evidence to show that waterway infrastructure can obstruct green sturgeon movement. Water diversion entrainment likely represents a significant source of early-life mortality, particularly in outmigrating juveniles (Mussen et al., 2014). Placing limits on water diversion inflow velocities according to time of day, season and location, may be effective in lowering entrainment rates (Verhille et al., 2014), but validating these estimates under field conditions is required. Swimming performance estimates derived from non-volitional laboratory trials can sometimes underestimate true capacity, because fish can attain greater swimming speeds under open-channel, volitional conditions (Hinch and Bratty, 2000; Peake, 2004). Moreover, the swimming performance data were collected at a single test temperature (18.5°C), but the thermal sensitivity of swimming performance needs to be assessed to appropriately inform season-specific water operations. Assessing burst swimming performance, across a range of temperatures, may also provide useful information because maximal, anaerobic swimming efforts are likely employed by fish avoiding entrainment.

A suggested additional management action is to install fish-protection devices at water diversion pipes. Poletto et al. (2014b, 2015) offer several fish-protection device designs (protective screen, louvre box, trash-rack box and perforated cylinder), which significantly lowered entrainment risk in laboratory trials. Field-testing these designs under a range of conditions (e.g. manipulating turbidity, light and temperature) will provide a more detailed assessment of their effectiveness. Furthering our understanding of how fish interact with these protection devices may aid in creating a design where impingements are minimized. Constructing and maintaining fish-protection devices can be expensive, and ~98% of diversions in the S-SI watershed remain unscreened (Herren and Kawasaki, 2001). Experimental assessments of fish protection devices will allow for the identification of the most effective designs that minimize impingements that are also cost effective (Moyle and Israel, 2005).

Water reservoirs present large-scale barriers to green sturgeon and may block access to critical spawning grounds. Decommissioning non-essential dams will likely prove effective in reconnecting aquatic habitat. In instances where decommissioning is unfeasible, delaying gate closures during peak spawning migrations may allow adults to reach preferred spawning habitats (Steel et al., 2018). Additionally, the installation of fishways can greatly enhance passage prospects but designs accommodating the swimming capacity and behaviour of green sturgeon are still needed. Fish ladders designed for salmonids have been shown to be inappropriate for sturgeons (Cheong et al., 2006; Kynard et al., 2011), and green sturgeon may require a specific design to optimize passage (e.g. fishway slope, length and height). Taken together, the studies reviewed here highlight the vulnerability of green sturgeon movement to barriers, yet preliminary assessments of management strategies offer promising results.

Conclusion

The data synthesis provided here allowed the identification of key data gaps and priority research areas. All experimental research to date has been conducted on the less threatened northern DPS and these findings may have limited applicability to the southern DPS because stressor impacts can differ between populations in a range of fishes (Eliason *et al.*, 2011). Gaining research access to southern DPS fish could prove invaluable to the protection and long-term persistence of green sturgeon. Studying the effects of stressors on early life-history stages must also be prioritized, because research has been biased towards juveniles and adults. Less than 10% of experimental studies examined the combined effects of

stressors on green sturgeon physiology, despite the growing recognition that organisms are rarely exposed to stressors in isolation in natural settings. Future research efforts should aim to characterize interactions among both biotic (e.g. competition and predation) and abiotic stressors (e.g. elevated temperatures and hypoxia) to inform holistic management strategies which target co-occurring threats. Green sturgeon research has also centred on understanding drivers of population declines during the freshwater life history phase, and far less work has been directed towards assessing the impacts of stressors experienced during the marine life history phase, such as ocean acidification and fisheries bycatch, Lastly, several potential threats remain uninvestigated in green sturgeon, including exposure to a wide range of contaminants in their habitat (e.g. heavy metals and pesticides), predation pressure from invasive fishes and the impact of low dissolved oxygen levels (i.e. hypoxia) on their performance and survival.

Despite these data gaps, this meta-analytic review synthesized the existing research on green sturgeon and provided a quantitative assessment of the impacts of prominent stressors to identify potential management actions. Metaanalyses facilitate evidence-based conservation practices by providing a summary of relevant, empirical evidence required for management decisions (Cook et al., 2014). By combining data from numerous sources, meta-analyses also generate greater explanatory power and may reveal effects not detected by individual studies (Mulrow, 1994; Gurevitch et al., 2018). Here, we show that the use of meta-analyses can inform management decisions by synthesizing existing evidence and highlighting knowledge gaps. A wealth of literature on the ecophysiological constraints dictated by environmental stressors exists for many species of concern which can be summarized for species management guidelines. For example, several science-based management actions have been suggested here, such as, ensuring water temperatures are maintained within optimal ranges during peak spawning periods, mitigating salt intrusion in nursery habitats and placing limits on water diversion inflow velocities according to time of day, season and location.

Importantly, the meta-analytic approach implemented here can be used to synthesize ecophysiological data on a wide range of imperiled, keystone and 'umbrella' species. This approach can be extended to groups of focal species/species assemblages to reveal if stressors have generalizable effects within an ecosystem. Meta-analytical data can also determine the life-stage-specific thermal tolerance window of various species inhabiting a single waterway and management actions can target temperatures where the greatest thermal overlap occurs among species. Similarly, species-specific tolerance limits to contaminants can be determined using meta-analytics to establish lowest admissible contaminant concentrations based on the tolerance limits of the most sensitive species (O'Brien and Keough, 2014). The meta-analytic approach provided here offers an additional tool to inform conservation decisions, where a multitude of factors (e.g. economic, cultural or social value of a species, stakeholder opinion, urgency and logistics) are considered when deciding which species or habitat region to prioritize for conservation. In conclusion, quantitative syntheses can facilitate the integration of ecophysiological data into species management plans, identify knowledge gaps and direct future research actions, with the ultimate goal of maximizing conservation gains from research efforts.

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Supplementary Material

Supplementary material is available at Conservation Physiology online.

References

Adams PB, Grimes C, Hightower JE, Lindley ST, Moser ML, Parsley MJ (2007) Population status of North American green sturgeon, *Acipenser medirostris. Environ Biol Fish* 79: 339–356.

Allen PJ, Cech J Jr (2007) Age/size effects on juvenile green sturgeon, *Acipenser medirostris*, oxygen consumption, growth, and osmoregulation in saline environments. *Environ Biol Fish* 79: 211–229.

Allen PJ, Cech J Jr, Kültz D (2009) Mechanisms of seawater acclimation in a primitive, anadromous fish, the green sturgeon. *J Comp Physiol B* 179: 903–920.

Allen PJ, Hodge B, Werner I, Cech JJ Jr (2006a) Effects of ontogeny, season, and temperature on the swimming performance of juvenile green sturgeon (*Acipenser medirostris*). *Can J Fish Aquat Sci* 63: 1360–1369.

Allen PJ, McEnroe M, Forostyan T, Cole S, Nicholl MM, Hodge B, Cech J Jr (2011) Ontogeny of salinity tolerance and evidence for seawater entry preparation in juvenile green sturgeon, *Acipenser medirostris*. *J Comp Physiol B* 181: 1045–1062.

Allen PJ, Nicholl M, Cole S, Vlazny A, Cech JJ Jr (2006b) Growth of larval juvenile green sturgeon in elevated temperature regimes. *Trans Am Fish Soc* 135: 89–96.

Arnot JA, Gobas FAPC (2006) A review of bioconcentration factor (BCF) and bioaccumulation factor (BAF) assessments for organic chemicals in aquatic organisms. *Environ Rev* 14: 257–297.

Baumgartner LJ, Reynoldson NK, Cameron L, Stanger JG (2009) Effects of irrigation pumps on riverine fish. *Fish Manag Ecol* 16: 429–437.

- Beamish FWH (1978) Swimming Capacity: Fish physiology, Ed1st. Clapp DF, Clark RD, Diana JS (1990) Range, activity, and habitat of large, Academic Press, New York, p 576.
- Bednarek A (2001) Undamming rivers: a review of the ecological impacts of dam removal. Environ Manage 27: 803-814.
- Beitinger TL, Bennett WA (2000) Quantification of the role of acclimation temperature in temperature tolerance of fishes. Environ Biol Fish 58: 277-288
- Beitinger TL, Bennett WA, McCauley RW (2000) Temperature tolerances of North American freshwater fishes exposed to dynamic changes in temperature. Environ Biol Fish 58: 237-275.
- Bobe J, Labbé C (2010) Egg and sperm quality in fish. Gen Comp Endocrinol 165: 535-548.
- Boeuf G, Payan P (2001) How should salinity influence fish growth? Comp Biochem Physiol C 130: 411–423.
- Borin JM, Moser ML, Hansen AG, Beauchamp DA, Corbett SC, Dumbauld BR, Pruitt C, Ruesink JL, Donoghue C (2017) Energetic requirements of green sturgeon (Acipenser medirostris) feeding on burrowing shrimp (Neotrypaea californiensis) in estuaries: importance of temperature, reproductive investment, and residence time. Environ Biol Fish 100: 1561-1573.
- Boysen KA, Hoover JJ (2009) Swimming performance of juvenile white sturgeon (Acipenser transmontanus): training and the probability of entrainment due to dredging. J Appl Ichthyol 25: 54-59.
- Brooks ML et al. (2012) Life histories, salinity zones, and sublethal contributions of contaminants to pelagic fish declines illustrated with a case study of San Francisco Estuary, California, USA. Estuar Coasts 35:603.
- Brown K (2007) Evidence of spawning by green sturgeon, Acipenser medirostris, in the upper Sacramento River, California. Environ Biol Fish 79: 297-303.
- Burt JM, Hinch SG, Patterson DA (2011) The importance of parentage in assessing temperature effects on fish early life history: a review of the experimental literature. Rev Fish Biol and Fisher 21: 377-406.
- Caissie D (2006) The thermal regime of rivers: a review. Freshw Biol 51: 1389-1406.
- Carlson SM, Olsen EM, Asbjørn Vøllestad L (2008) Seasonal mortality and the effect of body size: a review and an empirical test using individual data on brown trout. Funct Ecol 22: 663-673.
- Carlton JT, Thompson JK, Schemel LE, Nichols FH (1990) Remarkable invasion of San Francisco Bay California USA by asian clam Potamocorbula-amurensis I. Introduction and dispersal. Introduction and dispersal. Mar Ecol Prog Ser 66: 81-95.
- Cayan DR, Bromirski PD, Hayhoe K, Tyree M, Dettinger MD, Flick RE (2008) Climate change projections of sea level extremes along the California coast. Clim Change 87: 57-73.
- Cheong TS, Kavvas ML, Anderson EK (2006) Evaluation of adult white sturgeon swimming capabilities and applications to fishway design. Environ Biol Fish 77: 197-207.

- free-ranging brown trout in a Michigan stream. Trans Am Fish Soc 119:
- Cloern JE, Alpine AE, Cole BE, Wong RLJ, Arthur JF, Ball MD (1983) River discharge controls phytoplankton dynamics in the northern San Francisco Bay estuary. Estuar Coast Shelf Sci 16: 415–429.
- Cloern JE, Foster SQ, Kleckner AE (2014) Phytoplankton primary production in the world's estuarine-coastal ecosystems. Geobioscience 11: 2477-2501.
- Cloern JE, Jassby AD (2012) Drivers of change in estuarine-coastal ecosystems: discoveries from four decades of study in San Francisco Bay. Rev Geophys 50: RG4001.
- Cloern JE et al. (2011) Projected evolution of California's San Francisco Bay-Delta River system in a century of climate change. PLoS One 6: e24465.
- Cook CN, Possingham HP, Fuller RA (2014) Contribution of systematic reviews to management decisions. Conserv Biol 27: 902-615.
- Cooke SJ et al. (2012) Conservation physiology in practice: how physiological knowledge has improved our ability to sustainably manage Pacific salmon during up-river migration. Philos Trans R Soc Lond B Biol Sci 367: 1757-1769.
- Cooke SJ, Sack L, Franklin CE, Farrell AP, Beardall J, Wikelski M, Chown SL (2013) What is conservation physiology? Perspectives on an increasingly integrated and essential science. Conserv Physiol 1: cot001.
- Dalvi RS, Das T, Debnath D, Yengkokpam S, Baruah K, Tiwari LR, Pal AK (2017) Metabolic and cellular stress responses of catfish, Horabagrus brachysoma (Günther) acclimated to increasing temperatures. J Therm Biol 65: 32-40.
- Dasmann RF (1999) Environmental changes before and after the gold rush. In JJ Rawls, RJ Orsi, eds, A Golden State: Mining and Economic Development in Gold Rush California. Berkley: University of California Press.
- De Riu N, Lee JW, Huang SS, Moniello G, Hung SS (2014) Effect of dietary selenomethionine on growth performance, tissue burden, and histopathology in green and white sturgeon. Aquat Toxicol 148: 65–73.
- Dettlaff TA, Ginsburg AS, Schmalhausen OI (1993) Sturgeon Fishes: Developmental Biology and Aquaculture. New York: Springer.
- Dudgeon D et al. (2006) Freshwater biodiversity: importance, threats, status and conservation challenges. Biol Rev 81: 163-182.
- Dumbauld BR, Holden DL, Langness OP (2008) Do sturgeon limit burrowing populations in Pacific northwest estuaries? Environ Biol Fish 83: 283-296.
- Egger M, Davey Smith G, Schneider M, Minder C (1997) Bias in metaanalysis detected by a simple, graphical test. BMJ 315: 629.
- Eliason EJ, Clark TD, Hague MJ, Hanson JM, Gallagher ZS, Jeffries KM, Gale MK, Patterson DA, Hinch SG, Farrell AP (2011) Differences in thermal tolerance among sockeye salmon populations. Science 332: 109-112.

- Erickson DL, North JA, Hightower JE, Weber J, Lauck L (2002) Movement and habitat use of green sturgeon, *Acipenser medirostris*, in the Rogue River, Oregon, USA. *J Appl Ichthyol* 18: 565–569.
- Erickson DL, Webb MAH (2007) Spawning periodicity, spawning migration, and size at maturity of green sturgeon, *Acipenser medirostris*, in the Roque River, Oregon. *Environ Biol Fish* 79: 255–268.
- Fangue NA, Osborne EJ, Todgham AE, Schulte PM (2011) The onset temperature of the heat-shock response and whole-organism thermal tolerance are tightly correlated in both laboratory-acclimated and field-acclimatized tidepool sculpins. *Physiol Biochem Zool* 84: 341–352
- Fausch KD, Torgersen CE, Baxter CV, Li HW (2002) Landscapes to riverscapes: bridging the gap between research and conservation of stream fishes. *Biosciences* 52: 483–498.
- Foekema EM, Fischer A, Lopez Parron M, Kwadijk C, de Vries P, Murk AJ (2012) Toxic concentrations in fish early life stages peak at critical moments. *Environ Toxicol Chem* 31: 1381–1390.
- Foley CJ, Feiner ZS, Malinich TD, Hook TO (2018) A meta-analysis of the effects of exposure to microplastics on fish and aquatic invertebrates. *Sci Tot Environ* 631: 550–559.
- Fong S, Louie S, Werner I, Davis J, Connon RE (2016) Contaminant effects on California Bay–Delta species and human health. *SFEWS* 14: 5.
- Frew JA, Grue CE (2015) Assessing the risk to green sturgeon from application of imidacloprid to control burrowing shrimp in Willapa Bay, Washington–part II: controlled exposure studies. *Envion Toxicol Chem* 34: 2542–2548.
- Frew JA, Sadilek M, Grue CE (2015) Assessing the risk to green sturgeon from application of imidacloprid to control burrowing shrimp in Willapa Bay, Washington—part I: exposure characterization. *Envion Toxicol Chem* 34: 2533–2541.
- Gisbert E, Doroshov SI (2003) Histology of the developing digestive system and the effect of food deprivation in larval green sturgeon (*Acipenser medirostris*). *Aquat Living Resour* 16: 77–89.
- Goto D, Wallace WG (2010) Relative importance of multiple environmental variables in structuring benthic macroinfaunal assemblages in chronically metal-polluted salt marshes. *Mar Poll Bull* 60: 363–375.
- Gowan C, Fausch KD (2002) Why do foraging stream salmonids move during summer? *Ecol* 64: 139–153.
- Grans A, Axelsson M, Pitsillides K, Olsson C, Hojesjo J, Kaufman C, Cech JJ Jr (2009) A fully implantable multi-channel biotelemetry system for measurement of blood flow and temperature: a first evaluation in the green sturgeon. *Hydrobiologia* 619: 11–25.
- Grimaldo LF, Sommer T, Van Ark N, Jones G, Holland E, Moyle PB, Herbold B, Smith P (2009) Factors affecting fish entrainment into massive water diversions in a tidal freshwater estuary: can fish losses be managed? *North Am J Fish Manag* 29: 1253–1270.
- Gurevitch J, Koricheva J, Nakagawa S, Stewart G (2018) Meta-analysis and the science of research synthesis. *Nature* 555: 175–182.

Haller LY, Hung SS, Lee S, Fadel JG, Lee JH, McEnroe M, Fangue NA (2015) Effect of nutritional status on the osmoregulation of green sturgeon (Acipenser medirostris). Physiol Biochem Zool 88: 22–42.

.....

- Hamel MJ, Brown ML, Chipps SR (2008) Behavioral responses of rainbow smelt to in situ strobe lights. *North Am J Fish Manag* 28: 394–401.
- Harris RC *et al.* (2007) Whole-ecosystem study shows rapid fish-mercury response to changes in mercury deposition. *Proc Natl Acad Sci USA* 104: 16586–16591.
- Herren JR, Kawasaki SS (2001) Inventory of water diversions in four geographic areas in California's Central Valley. Fish bulletin 179. Contributions to the biology of Central Valley Salmonids. https://nrm.dfg.ca.gov/FileHandler.ashx?DocumentID=3579.
- Heublein J et al. (2017) Life history and current monitoring inventory of San Francisco estuary sturgeon. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-SWFSC-589.
- Heublein JC, Kelly JT, Crocker CE, Klimley AP, Lindley ST (2009) Migration of green sturgeon, *Acipenser medirostris*, in the Sacramento River. *Environ Biol Fish* 84: 245–258.
- Hinch SG, Bratty J (2000) Effects of swim speed and activity pattern on success of adult sockeye salmon migration through an area of difficult passage. *Trans Am Fish Soc* 129: 598–606.
- Hochachka PW (1967) Organization of metabolism during temperature compensation. In CL Prosser, ed, Molecular Mechanisms of Temperature Adaptation. American Association for the Advancement of Science, Washington, pp. 177–203.
- Hochachka PW, Somero GN (1968) The adaptation of enzymes to temperature. *Comp Biochem Physiol* 27: 659–668.
- Holm J, Palace V, Siwik P, Sterling G, Evans R, Baron C, Werner J, Wautier K (2005) Developmental effects of bioaccumulated selenium in eggs and larvae of two salmonid species. *Environ Toxicol Chem* 24: 2373–2381.
- Hooper DU, Adair EC, Cardinale BJ, Byrnes JEK, Hungate BA, Matulich KL, Gonzalez A, Duffy JE, Gamfeldt L, O'Connor MI (2012) A global synthesis reveals biodiversity loss as a major driver of ecosystem change. *Nature* 486: 105–108.
- Horodysky AZ, Cooke SJ, Brill RW (2015) Physiology in the service of fisheries science: why thinking mechanistically matters. *Rev Fish Biol Fisheries* 25: 425–447.
- Huey RB, Stevenson RD (1979) Integrating thermal physiology and ecology of ectotherms: a discussion of approaches. *Am Zool* 19: 356–366.
- Huey RB, Kearney MR, Krockenberger A, Holtum JAM, Jess M, Williams S (2012) Predicting organismal vulnerability to climate warming: roles of behaviour, physiology and adaptation. *Philos Trans R Soc B Biol Sci* 367: 1665–1679.
- Huff DD, Lindley ST, Rankin PS, Mora EA (2011) Green sturgeon physical habitat use in the coastal Pacific Ocean. *PloS One* 6: e25156.
- IEP MAST (2015) An updated conceptual model for delta smelt: our evolving understanding of an estuarine fish h. Page 205. Technical

- Report 90, January, 2015. Interagency Ecological Program for the San Francisco Bay/Delta Estuary.
- Israel JA, Blumberg M, Cordes J, May B (2004) Geo-graphic patterns of genetic differentiation among western U.S. collections of North American green sturgeon (*Acipenser medirostris*). North Am J Fish Manage 24: 922–993.
- Israel JA, Kimley AP (2008) Life history conceptual model for North American Green sturgeon (*Acipenser medirostris*). *Delta Reg Ecosyst Restor Implement Plan Rep* 1–49.
- Janz DM et al. (2010) Selenium Toxicity to Aquatic Organisms: Ecological Assessment of Selenium in the Aquatic Environment. CRC Press, Boca Raton, FL, pp. 141–231.
- Jobling M (1994) Fish Bioenergetics. London; Chapman and Hall.
- Johnson A, Carew E, Sloman KA (2007) The effects of copper on the morphological and functional development of zebrafish embryos. *Aquat Toxicol* 84: 431–438.
- Johnson ML, Werner I, Teh SJ, Loge F (2010) Evaluation of Chemical, Toxicological, and Histopathologic Data to Determine Their Role in the Pelagic Organism Decline. Final Report to the California State Water Resources Control Board and Central Valley Regional Water Quality Control Board. Davis: University of California.
- Johnsson JI, Nöbbelin F, Bohlin T (1999) Territorial competition among wild brown trout fry: effects of ownership and body size. J Fish Biol 54: 469–472.
- Kelly JT, Klimley AP, Crocker CE (2007) Movements of green sturgeon, Acipenser medirostris, in the San Francisco Bay estuary, California. Environ Biol Fish 79: 281–295.
- Komoroske LM, Connon RE, Lindberg J, Cheng BS, Castillo G, Hasenbein M, Fangue NA (2014) Ontogeny influences sensitivity to climate change stressors in an endangered fish. *Conserv Physiol* 2: doi:10.1093/conphys/cou008.
- Kroll KJ, Doroshov SI (1991) Vitellogenin: Potential Vehicle for Selenium Bioaccumulation Ion Oocytes of the White Sturgeon: Acipenser. CEMA-GREF, Bordeaux, pp. 99–106.
- Kupsco A, Schlenk D (2016) Molecular mechanisms of selenium-induced spinal deformities in fish. *Aquat Toxicol* 179: 134–150.
- Kynard B, Parker E, Parker T (2005) Behavior of early life intervals of Klamath River green sturgeon, *Acipenser medirostris*, with a note on body color. *Environ Biol Fish* 72: 85–97.
- Kynard B, Pugh D, Parker T (2011) Passage and behaviour of cultured Lake sturgeon in a prototype side-baffle fish ladder: I. ladder hydraulics and fish ascent. *J Appl Ichthy* 27: 77–88.
- Lajeunesse MJ (2011) On the meta-analysis of response ratios for studies with correlated and multi-group designs. *Ecology* 92: 2049–2055.
- Lankford SE, Adams TE, Cech JJ Jr (2003) Time of day and water temperature modify the physiological stress response in green sturgeon *Acipenser medirostris*. *Comp Biochem Physiol A* 135: 291–302.

- Lee S, Hung SSO, Fangue NA, Haller L, Verhille CE, Zhao J, Todgham AE (2016) Effects of feed restriction on the upper temperature tolerance and heat shock response in juvenile green and white sturgeon. *Comp Biochem Physiol A* 198: 87–95.
- Lee SW, De Riu N, Lee S, Bai SC, Moniello G, Hung SS (2011) Effects of dietary methylmercury on growth performance and tissue burden in juvenile green (*Acipenser medirostris*) and white sturgeon (*A. transmontanus*). *Aquat Toxicol* 105: 227–234.
- Lee SW, Kim JW, De Riu N, Moniello G, Hung SS (2012) Histopathological alterations of juvenile green (*Acipenser medirostris*) and white sturgeon (*Acipenser transmontanus*) exposed to graded levels of dietary methylmercury. *Aquat Toxicol* 109: 90–99.
- Lemly AD (1996) Assessing the toxic threat of selenium to fish and aquatic birds. *Environ Monit Assess* 43: 19–35.
- Lemly AD (2004) Aquatic selenium pollution is a global environmental issue. *Ecotoxicol Environ Saf* 59: 44–56.
- Le Roy A, Loughland I, Seebacher F (2017) Differential effects of developmental thermal plasticity across three generations of guppies (*Poecilia reticulata*): canalization and anticipatory matching. *Sci Rep* 7: 4313.
- Linares-Casenave J, Linville R, Van Enennaam JP, Muguet JB, Doroshov SI (2015) Selenium tissue burden compartmentalization in resident white sturgeon (*Acipenser transmontanus*) of the San Francisco Bay Delta estuary. *Environ Toxicol Chem* 34: 152–160.
- Linares-Casenave J, Werner I, Van Eenennaam JP, Doroshov SI (2013) Temperature stress induces notochord abnormalities and heat shock proteins expression in larval green sturgeon (*Acipenser medirostris* Ayres 1854). *J Appl Ichthyol* 29: 958–967.
- Lindley ST, Moser ML, Erickson DL, Belchik M, Welch DW, Rechisky EL, Kelly JT, Heublein J, Klimley AP (2008) Marine migration of North American green sturgeon. *Trans Am Fish Soc* 137: 182–194.
- Lindquist S, Craig EA (1988) The heat shock proteins. *Annu Rev Genet* 22: 631–677.
- Liu WC, Liu HM (2014) Assessing the impacts of sea level rise on salinity intrusion and transport time scales in a tidal estuary, Taiwan. *Water* 6: 324–344.
- Martell DJ, Kieffer JD (2007) Persistent effects of incubation temperature on muscle development in larval haddock (*Melanogrammus aeglefinus* L.). *J Exp Biol* 210: 1170–1182.
- Maes J, Turnpenny AWH, Lambert DR, Nedwell JR, Parmentier A, Ollevier F (2004) Field evaluation of a sound system to reduce estuarine fish intake rates at a power plant cooling water inlet. *J Fish Biol* 64: 938–946.
- Mallen-Cooper M, Brand DA (2007) Non-salmonids in a salmonid fishway: what do 50 years of data tell us about past and future fish passage? *Fish Manage Ecol* 14: 319–332.
- Marchetti MP, Moyle PB (2001) Effects of flow regime on fish assemblages in a regulated California stream. *Ecol App* 11: 530–539.

- Maulvault AL, Custódio A, Anacleto P, Repolho T, Pousão P, Nunes ML, Diniz M, Rosa R, Marques A (2016) Bioaccumulation and elimination of mercury in juvenile seabass (*Dicentrarchus labrax*) in warmer environment. *Environ Res* 149: 77–85.
- Mayfield RB, Cech JJ Jr (2004) Temperature effects on green sturgeon bioenergetics. *Trans Am Fish Soc* 133: 961–970.
- McAdam SO (2011) Effects of substrate condition on habitat use and survival by white sturgeon (*Acipenser transmontanus*) larvae and potential implications for recruitment. *Can J Fish Aquat Sci* 68: 812–822.
- McKim JM (1977) Evaluation of tests with early life stages of fish for predicting long-term toxicity. *J Fish Res Board Can* 34: 1148–1154.
- Møller AP, Jennions MD (2001) Testing and adjusting for publication bias. *Trends Ecol Evol* 16: 580–586.
- Mora EA, Lindley ST, Erickson DL, Klimley AP (2009) Do impassable dams and flow regulation constrain the distribution of green sturgeon in the Sacramento River, California? *J Appl Ichthyol* 25: 39–47.
- Moschet C, Witter I, Simovic J, Junghans M, Piazzoli A, Singer H, Stamm C, Leu C, Hollender J (2014) How a complete pesticide screening changes the assessment of surface water quality. *Environ Sci Technol* 48: 5423–5432.
- Moser ML, Lindley ST (2007) Use of Washington estuaries by subadult and adult green sturgeon. *Environ Biol Fish* 79: 243–253.
- Motani R, Wainwright PC (2015) How warm is too warm for the life cycle of actinopterygian fishes? *Sci Rep* 5: 11597.
- Mousseau TA, Fox CW (1998) The adaptive significance of maternal effects. *Trends Ecol Evol* 13: 403–407.
- Moyle PB (2002) *Inland Fishes of California*. University of California Press, Berkeley, CA, p. 502.
- Moyle PB, Israel JA (2005) Untested assumptions. Fisheries 30: 20–28.
- Moyle PB, Leidy RA (1992) Loss of Biodiversity in Aquatic Ecosystems: Evidence from Fish Faunas. In PL Fiedler, SK Jain, eds, *Conservation Biology*. Springer, Boston, MA, pp. 127–169.
- Mulrow CD (1994) Systematic reviews—rationale for systematic reviews. *Br Med J* 309: 597–599.
- Mussen TD, Cocherell D, Poletto JB, Reardon JS, Hockett Z, Ercan A, Bandeh H, Kavvas ML, Cech JJr, Fangue NA (2014) Unscreened water-diversion pipes pose an entrainment risk to the threatened green sturgeon, *Acipenser medirostris*. *PloS One* 9: e86321.
- Nakamoto RJ, Kisanuki TT, Goldsmith GH (1995) Age and growth of Klamath River green sturgeon (*Acipenser medirostris*). U.S. Fish and Wildlife Service Report 93-FP-13. Yreka, CA.
- Niimi AJ (1983) Biological and toxicological effects of environmental contaminants and their eggs. *Can J Fish Aquat Sci* 40: 306–312.
- NMFS (2010) Southern Distinct Population Segment of the North American Green Sturgeon (Acipenser medirostris). Long Beach, CA, USA: West Coast Regional Office.

NMFS (2014) Recovery plan for the evolutionarily significant units of Sacramento River winter-run Chinook salmon and central Valley springrun Chinook salmon and the Distinct Population Segment of California Central Valley Steelhead. California Central Valley Area Office, CA, USA.

.....

- NMFS (2018) Recovery Plan for the Southern Distinct Population Segment of North American Green Sturgeon (Acipenser medirostris). National Marine Fisheries Service, Sacramento, CA, USA.
- O'Brien AL, Keough MJ (2014) Ecological responses to contamination: a meta-analysis of experimental marine studies. *Environ Poll* 195: 185–191.
- Peake S (2004) An evaluation of the use of critical swimming speed for determination of culvert water velocity criteria for smallmouth bass. *Trans Am Fish Soc* 133: 1472–1479.
- Pike A, Danner E, Boughton D, Melton F, Nemani R, Rajagopalan B, Lindley S (2013) Forecasting river temperatures in real time using a stochastic dynamics approach. *Water Resour Res* 49: 5168–5182.
- Plaut I (2001) Critical swimming speed: its ecological relevance. *Comp Biochem Physiol A* 131: 41–50.
- Poletto JB, Cocherell DE, Ho N, Cech J Jr, Klimley AP, Fangue NA (2014a) Juvenile green sturgeon (*Acipenser medirostris*) and white sturgeon (*Acipenser transmontanus*) behavior near water-diversion fish screens: experiments in a laboratory swimming flume. *Can J Fish Aquat Sci* 71: 1030–1038.
- Poletto JB, Cocherell DE, Ho N, Cech J Jr, Klimley AP, Fangue NA (2018) The effect of size on juvenile green sturgeon (*Acipenser medirostris*) behavior near water-diversion fish screens. *Environ Biol Fish* 101: 67–77.
- Poletto JB, Cocherell DE, Klimley AP, Cech J Jr, Fangue NA (2013) Behavioural salinity preferences of juvenile green sturgeon *Acipenser medirostris* acclimated to fresh water and full-strength salt water. *J Fish Biol* 82: 671–685.
- Poletto JB, Cocherell DE, Mussen TD, Ercan A, Bandeh H, Kavvas ML, Cech JJ Jr, Fangue NA (2014b) Efficacy of a sensory deterrent and pipe modifications in decreasing entrainment of juvenile green sturgeon (*Acipenser medirostris*) at unscreened water diversions. *Conserv Physiol* 2: doi: 10.1093/conphys/cou056.
- Poletto JB, Cocherell DE, Mussen TD, Ercan A, Bandeh H, Kavvas ML, Cech JJ, Fangue NA (2015) Fish-protection devices at unscreened water diversions can reduce entrainment: evidence from behavioural laboratory investigations. *Conserv Physiol* 3: doi:10.1093/conphys/cov040.
- Poytress WR, Gruber JJ, Van Eenennaam JP, Gard M (2015) Spatial and temporal distribution of spawning events and habitat characteristics of Sacramento River Green sturgeon. *Trans Am Fish Soc* 144: 1129–1142.
- Prosser CL (1991) Respiration and Metabolism. Environmental and Metabolic Animal Physiology: Comparative Animal Physiology. Wiley-Liss, New York, pp. 392–399.
- Radtke LD (1966) Distribution of smelt, juvenile sturgeon, and starry flounder in the Sacramento-San Joaquin Delta with observations

on food of sturgeon. In JL Turner, DW Kelly, eds, *Ecological studies of the Sacramento-San Joaquin Delta*. Part II Fishes of the Delta. pp. 115–129.

- Richmond A, Kynard B (1995) Ontogenetic behaviours of shortnose sturgeon. *Copeia* 1995: 172–182.
- Rodgers EM, Cocherell DE, Nguyen TX, Todgham AE, Fangue NA (2018) Plastic responses to diel thermal variation in juvenile green sturgeon, *Acipenser medirostris. J Therm Biol* 76: 147–155.
- Rosenberg MS (2005) The file-drawer problem revisited: a general weighted method for calculating fail-safe numbers in meta-analysis. *Evolution* 59: 464–468.
- Rossiter M (1996) Incidence and consequences of inherited environmental effects. *Annu Rev Ecol Syst* 27: 451–476.
- Sardella BA, Kültz D (2009) Osmo- and ionoregulatory responses of green sturgeon (*Acipenser medirostris*) to salinity acclimation. *J Comp Physiol B* 179: 383–390.
- Sardella BA, Sanmarti E, Kültz D (2008) The acute temperature tolerance of green sturgeon (*Acipenser medirostris*) and the effect of environmental salinity. *J Exp Zool* 309A: 477–483.
- Schindelin S, Rueden CT, Hiner MC, Eliceiri KW (2015) The ImageJ ecosystem: an open platform for biomedical image analysis. *Mol Reprod Dev* 82: 518–529.
- Schneider CA, Rasband WS, Eliceiri KW (2012) NIH image to ImageJ: 25 years of image analysis. *Nat Methods* 9: 671–675.
- Schnurr ME, Yin Y, Scott GR (2014) Temperature during embryonic development has persistent effects on metabolic enzymes in the muscle of zebrafish. *J Exp Biol* 217: 1370–1380.
- Seesholtz AM, Manuel MJ, Van Eenennaam JP (2015) First documented spawning and associated habitat conditions for green sturgeon in the Feather River, CA. *Environ Biol Fish* 98: 905–912.
- Service RF (2007) Delta blue, California Style. Science 317: 442–445.
- Shi M, Zhang C, Xia IF, Cheung ST, Wong KS, Wong KH, Au DWT, Hinton DE, Kwok KWH (2018) Maternal dietary exposure to selenium nanoparticles led to malformation in offspring. *Ecotoxicol Environ Saf* 156: 34–40.
- Silvestre F, Linares-Casenave J, Doroshov S, Kültz D (2010) A proteomic analysis of green and white sturgeon larvae exposed to heat stress and selenium. Sci Total Environ 408: 3176–3188.
- Smylie MS, McDonough CJ, Reed LA, Shervette VR (2016) Mercury bioaccumulation in an estuarine predator: biotic factors, abiotic factors, and assessments of fish health. *Environ Poll* 214: 169–176.
- Sokolova IM (2013) Energy-limited tolerance to stress as a conceptual framework to integrate the effects of multiple stressors. *Integr Comp Biol* 53: 597–608.
- Sokolova IM, Lannig G (2008) Interactive effects of metal pollution and temperature on metabolism in aquatic ectotherms: implications of global climate change. *Clim Res* 37: 181–201.

- SWRCB (2010) Transmittal of the 2010 Integrated Report [Clean Water Act Section 303(d) and Section 305(b)]. Letter to Alexis Strauss, USEPA, and four CDs of supporting materials, including the staff report, fact sheets, and responsiveness summary, dated October 11, 2010. https://www3.epa.gov/region9/water/tmdl/california.html.
- Steel AE, Thomas MJ, Klimley AP (2018) Reach specific use of spawning habitat by adult green sturgeon (*Acipenser medirostris*) under different operation schedules at Red Bluff Diversion Dam. *J Appl Ichthyol* 0: 1–8.
- Strayer DL (2010) Alien species in fresh waters: ecological effects, interactions with other stressors, and prospects for the future. *Freshw Biol* 55: 152–174.
- Sytina LA, Shagaeva VG (1987) Thermotolerance of progeny of Sevryuga, *Acipenser stellatus*, spawners from different periods of the spawning migration. *J Ichthyol* 27: 12–21.
- Thomas MJ, Peterson ML, Chapman ED, Hearn AR, Singer GP, Battleson RD, Klimley AP (2014) Behavior, movements, and habitat use of adult green sturgeon, *Acipenser medirostris*, in the upper Sacramento River. *Environ Biol Fish* 97: 133–146.
- Thomas MJ, Peterson ML, Friedenberg N, Van Eenennaam JP, Johnson JR, Hoover JJ, Klimley AP (2013) Stranding of spawning run green sturgeon in the Sacramento River: post-rescue movements and potential population-level effects. *N Am Fish Manag* 33: 287–297.
- Troiano AT, Grue CE (2016) Plasma cholinesterase activity as a biomarker for quantifying exposure of green sturgeon to carbaryl following applications to control burrowing shrimp in Washington state. *Environ Toxicol Chem* 35: 2003–2015.
- U.S. Fish and Wildlife Service (1995) Sacramento-San Joaquin Delta Native Fishes Recovery Plan. Portland, Oregon: U.S. Fish and Wildlife Service.
- Viechtbauer W (2010) Conducting meta-analyses in R with the metafor package. *J Stat Softw* 36: 1–48.
- Van Eenennaam JP, Webb MAH, Deng X, Doroshov SI, Mayfield R, Cech JJ Jr (2001) Artificial spawning and larval rearing of Klamath River green sturgeon. *Trans Am Fish Soc* 130: 159–165.
- Van Eenennaam JP, Chapman FA, Jarvis PL (2004) Aquaculture. In GTO LeBreton, FWH Beamish, RS McKinley, eds, *Sturgeons and Paddlefish of North America*. Fish and Fisheries Series 27. Kluwer Academic Publishers, Dordrecht, pp. 277–311.
- Van Eenennaam JP, Linares-Casenave J, Deng X, Doroshov SI (2005) Effect of incubation temperature on green sturgeon embryos, *Acipenser medirostris. Environ Biol Fish* 72: 145–154.
- Van Eenennaam JP, Linares-Casenave J, Doroshov SI, Hillemeier DC, Willson TE, Nova AA (2006) Reproductive conditions of the Klamath river green sturgeon (*Acipenser medirostris*). *Trans Am Fish Soc* 135: 151–163.
- Van Eenennaam JP, Linares-Casenave J, Muguet JB, Doroshov SI (2008) Induced spawning, artificial fertilization and egg incubation techniques for green sturgeon. *N Am J Aquac* 70: 434–445.

- Van Eenennaam JP, Linares-Casenave J, Doroshov SI (2012) Tank spawning of first generation domestic green sturgeon. *J Appl Ichthyol* 28: 505–511.
- Vardy DW, Oellers J, Doering JA, Hollert H, Giesy JP, Hecker M (2013) Sensitivity of early life stages of white sturgeon, rainbow trout, and fathead minnow to copper. *Ecotoxicol* 22: 139–147.
- Vaz PG, Kebreab E, Jung SSO, Fadel JG, Lee S, Fangue NA (2015) Impact of nutrition and salinity changes on biological performances of Green and white sturgeon. *Plos One* 10: e0122029.
- Verhille CE, Lee S, Todgham AE, Cocherell DE, Hung SSO, Fangue NA (2015) Effects of nutritional deprivation on juvenile green sturgeon growth and thermal tolerance. *Environ Biol Fish* 99: 145–159.
- Verhille CE, Poletto JB, Cocherell DE, DeCourten B, Baird S, Cech J Jr, Fangue NA (2014) Larval green and white sturgeon swimming performance in relation to water-diversion flows. *Conserv Physiol* 2: doi:10.1093/conphys/cou031.
- Videler JJ (1993) Fish Swimming. London: Chapman and Hall.
- Vörösmarty CJ, McIntyre PB, Gessner MO, Dudgeon D, Prusevich A, Green P Glidden S, Sullivan CA, Reidy Liermann C, Davies PM (2010) Global threats to human water security and river biodiversity. *Nature* 467: 555–561.
- Wang W, Deng D, De Riu N, Moniello G, Hung SSO (2013) Heat shock protein 70 (HSP70) responses in tissues of white sturgeon and Green sturgeon exposed to different stressors. *N Am J Aquac* 75: 164–169.

- Weis JS, Weis P (1989) Tolerance and stress in a polluted environment. *BioScience* 39: 89–95.
- Wenger SJ *et al.* (2011) Flow regime, temperature, and biotic interactions drive differential declines of trout species under climate change. *Proc Natl Acad Sci USA* 108: 14175–14180.
- Werner I, Linares-Casenave J, Van Eenennaam JP, Doroshov SI (2007) The effect of temperature stress on development and heat-shock protein expression in larval green sturgeon (*Acipenser medirostris*). *Environ Biol Fish* 79: 195–200.
- Wikelski M, Cooke SJ (2006) Conservation physiology. *Trends Ecol Evol* 21: 38–46.
- Winder M, Jassby AD (2011) Shifts in Zooplankton Community Structure: Implications for Food Web Processes in the Upper San Francisco Estuary. *Estuaries Coast* 34: 675–690.
- Yamamoto S, Morita K, Koizumi I, Maekawa K (2004) Genetic differentiation of white-spotted charr (*Salverlinus leucomaenis*) populations after habitat fragmentation: spatial–temporal changes in gene frequencies. *Conserv Genet* 5: 529–538.
- Young PS, Swanson C, Cech JJ (2010) Close encounters with a fish screen III: behavior, performance, physiological stress responses, and recovery of adult Delta smelt exposed to two-vector flows near a fish screen. *Trans Am Fish Soc* 139: 713–726.
- Zheng KK, Deng DF, De Riu N, Moniello G, Hung SSO (2015) The effect of feeding rate on the growth performance of green sturgeon (*Acipenser medirostris*) fry. *Aquacult Nutr* 21: 489–495.