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## **Determination of the Content of Organochlorine Pesticides Residues in Chicken Eggs Sold in Bamako**

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### **Abstract**

Pesticides used in agriculture enter the food chain, because of their chemical stability persist and their accumulation in different media in regions where they are used regularly. Their high solubility in the fat means that they can be found in the chicken eggs through poultry feed. The present study, aimed to determine the levels of pesticide residues in eggs in 36 composite samples of chicken eggs at twelve sale points distributed between the six communes of the District of Bamako. The samples were extracted and analyzed in the laboratory by gas chromatography to determine the residues of four organochlorine pesticides. The results showed that 58% of eggs sold in Bamako are contaminated by organochlorine pesticides. The average concentrations of the pesticides detected and quantified was respectively 0.52 mg/Kg  $\pm$  0.05 for DDT, 0.15 mg/Kg  $\pm$  0.04 for endosulfan  $\alpha$ , 0.23 mg/Kg  $\pm$  0, 03 for  $\beta$  endosulfan and 0.02 mg/Kg  $\pm$  0.01 for dieldrin. DDT was the most encountered pesticide in 53% of the samples, Endosulfan A and Endosulfan B were encountered in 36% and the lowest dosed dieldrin with less than 6% of the egg samples analysed. The feed for laying hens is largely made up of the following three ingredients: maize, fishmeal and cottonseed cake which are highly exposed to contamination by pesticides. These average levels of three residues identified in the egg samples are well above the maximum residue limit (MRL) and therefore unacceptable for consumption because they are already prohibited pesticides.

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**Keywords:** Determination; Eggs; Pesticides; Organochlorines; Bamako.

## **I. Introduction**

Mali is a vast Sahelo-Saharan country with an agro-pastoral vocation, located in the heart of West Africa with a national herd estimated at 38,587,450 poultry including 2,757,313 layers placed on a forecast of 2,628,000 or 104.91 % of production targets; and 441,079,208 eggs produced out of a forecast of 467,000,000 or 94.43% of the production objectives [ 1].

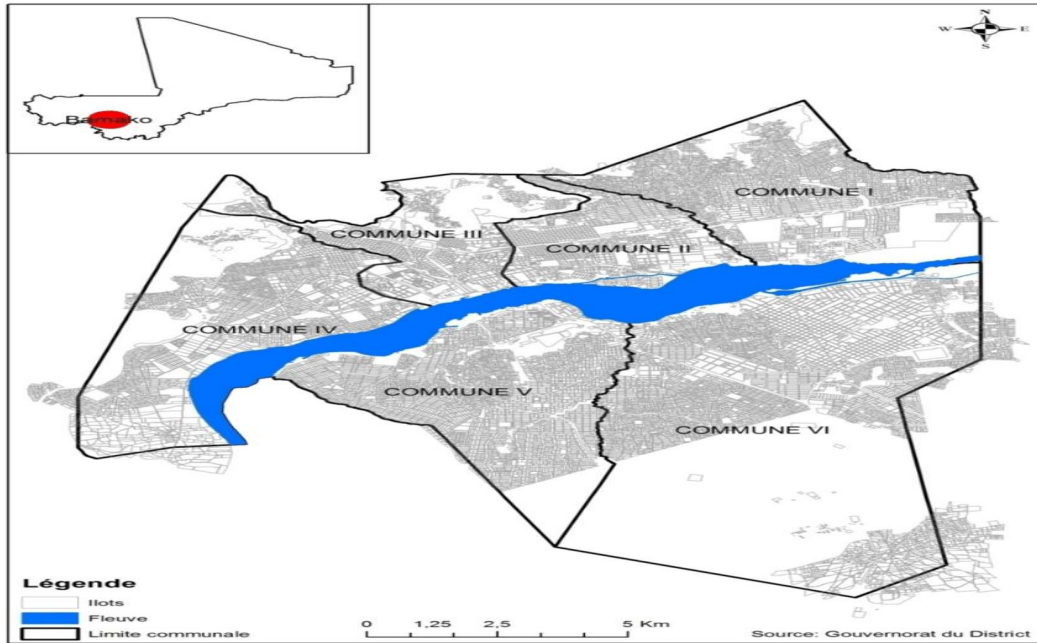
However, this sector is confronted with a series of potential contaminants such as pesticide and antibiotic residues. In West Africa, only pathogenic microbial agents, pesticide residues and aflatoxins [2,3,4] have been the subject of work or studies as part of an approach to protect the health quality of food intended for human and animal consumption. These dangers have been most reported as threatening public health [2]. Contamination of poultry and poultry by-products by pesticide and antibiotic residues is a major risk for consumers [5] but also a matter of concern, both for economic and health reasons. The consumption of poor quality animal products can cause serious diseases (cancers ([6] and [7]. (Collective food poisoning (TIAC)) and endanger the health of consumers. Since the advent of the Company Malian of Development des Textiles (CMDT), Malian agriculture has experienced extensive use of chemical inputs, including a total volume of 2 794 992 doses of insecticides, 854100 liters of herbicides and 298,500 sachets of fungicides, was used as pesticides for cotton production in 2017 [ 8].

Mali, like other developing countries, does not have reliable data on poisoning cases related to chemicals due to the absence of a surveillance or notification system, although all cases are reported. Years of cases of human poisoning sometimes followed by human deaths. The series of health safety crises have led to the conclusion of the need to strengthen health measures and consumer regulatory requirements at the national, regional and international level. With regard to the provisions of Regulation No. 07/2007/CM/UEMOA relating to the health safety of plants, animals and food in WAEMU as well as those of the national food safety policy adopted by the Government of Mali in 2002, economic operators are responsible for the sanitary quality of the foodstuffs they place on the market. They require producers and distributors to implement self-monitoring to control the sanitary and phytosanitary risks of food. The inspection services will be in charge of controlling the effectiveness of the self-checking system set up in the companies. In the specific case of contamination of eggs by pesticide residues, monitoring is required throughout the production chain. Because of their chemical stability, their persistence and their accumulation in various organs, the pesticides enter the food chain by the meat and eggs of layers which regularly receive contaminated food of residues of pesticides. The use of pesticides in the production and storage of feed for laying hens can cause their residues to end up in eggs. Then, the general objective of this study was to determine the concentration of organochlorine pesticide residues in chicken eggs sold in the District of Bamako.

## **2. Materials and methods**

### **Study area and period**

The prospecting study took place in 2020 in November. Sampling took place at the level of the six (6) communes of the District of Bamako. Twelve (12) wholesaler outlets at market level served as sampling points.



**Figure 1:** Study area and sites



**Figure 2:** Sample of fresh eggs

### Sampling

Egg samples were taken at the wholesale supply points located in the neighborhood markets of the Bamako district. For each commune, two sampling points were selected and three (03) composite samples of six (06) eggs each were taken (2 points x 6 communes x 3 samples per point). The samples (n=36) were labeled and transported to the laboratory as quickly as possible where they were kept in the refrigerator before the analyzes

**Table 1:** Summary of samples for the detection of pesticide residues

Commune	Site	Number	Total
C I	Boukassoumbougou	3	6
	Djelibougou	3	
C II	Médina Coura	3	6
	Bagadadji	3	
C III	Dibida	3	6
	Fréquence 3	3	
C IV	Lafiabougou	3	6
	Sebenikoro	3	
C V	Sabalibougou	3	6
	Bacodjicoroni ACI	3	
C VI	Magnambougou	3	6
	Yirimadjo	3	
Total		36	36

### Pesticide residue analysis method

The samples were treated and analyzed in accordance with the provisions of the standards (NF EN 12393-2: 1999) validated by the Central Veterinary Laboratory as a multi-residue method for the determination by gas chromatography of pesticide residues in non-fatty foods. It involves four (04) steps: extraction of pesticide residues, concentration, purification and data analysis.

### Extraction, concentration and purification of extracts

20g of egg yolk from each sample was taken. Then, 25ml of dichloromethane was added to the sample and the containers tightly closed, then transported in an electric shaker at medium speed for 30 minutes at room temperature. The homogeneous mixture obtained was poured into a vacuum filtration system. The filtrate obtained was poured into the micro extraction column. Then, 10ml of the first elution solution was successively added, which is the mixture of dichloromethane and hexane in the respective proportions 1/4 and 3/4, then 10ml of hexane. The various eluates obtained are evaporated to dryness on a Heidolph WB 2001 brand rotavapor at 70°C, the residues are then taken up with small quantities of hexane in a 5 mL or 10 mL volumetric flask. The extract is transferred to vials and is thus ready for chromatographic analysis.

### Instrumental analysis of the extracts

The sample extracts were analyzed on gas chromatograph (Agilent 7890A) fitted with an electron capture detector ( $\mu$ ECD) and CHEMSTATION operating software. An RTX-5 Crossbond capillary column, 30 meters x 250  $\mu$ m internal diameter x 0.50  $\mu$ m film thickness, 5% diphenyl-95% dimethyl polysiloxane stationary phase was used. An HP-5 type column (5% Phenyl methyl Siloxane) made it possible to confirm the status of the samples already analyzed (negative and positive). The injector was in splitless mode at a temperature of 280°C with nitrogen as the carrier and make-up gas at 60 mL/min and a constant flow rate of 1.6 mL/min. The programming of the oven temperature in three stages was as follows: an initial stage between 70-150°C followed by a secondary stage of 150-200°C and a final stage of 200-280°C. The detector was at a temperature of 300°C for an injected sample volume of 1  $\mu$ L.

### Standard solutions and calibration curve

The individual reference standards of the four pesticides with a purity varying between 96.3 and 98.5%, acquired from the manufacturer (Ehrenstorfer GmbH-Germany) were used. A solution of the mixture of the four standards was prepared by diluting small quantities in hexane at 5mg/L. Then an analysis range of five different levels at several concentrations (0.0125; 0.25; 0.5; 0.1 and 0.125 µg/ml) was prepared and used for GC calibration.

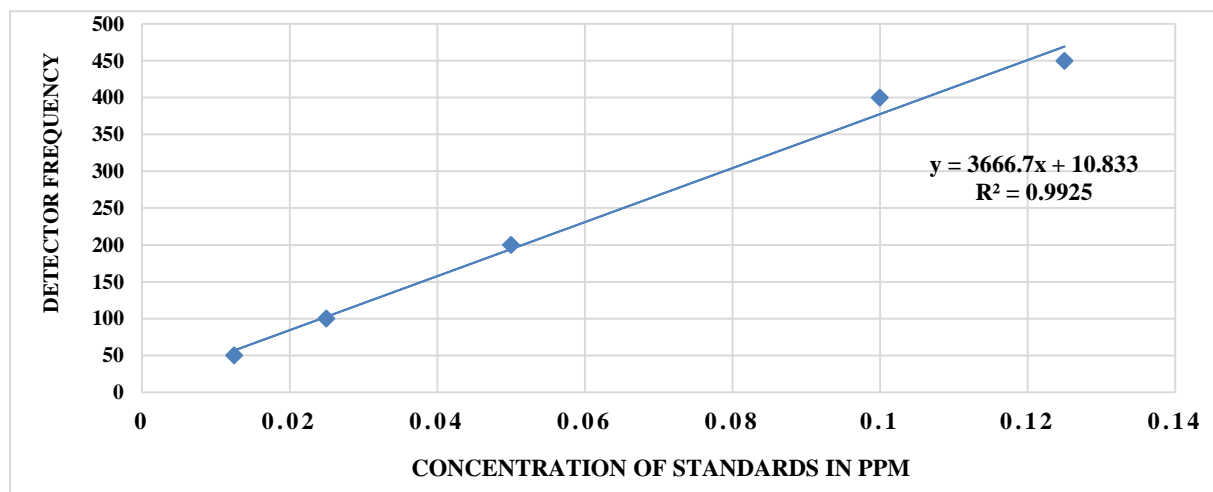


Figure 3: Calibration line of the mixture of analytical standards used for the analysis

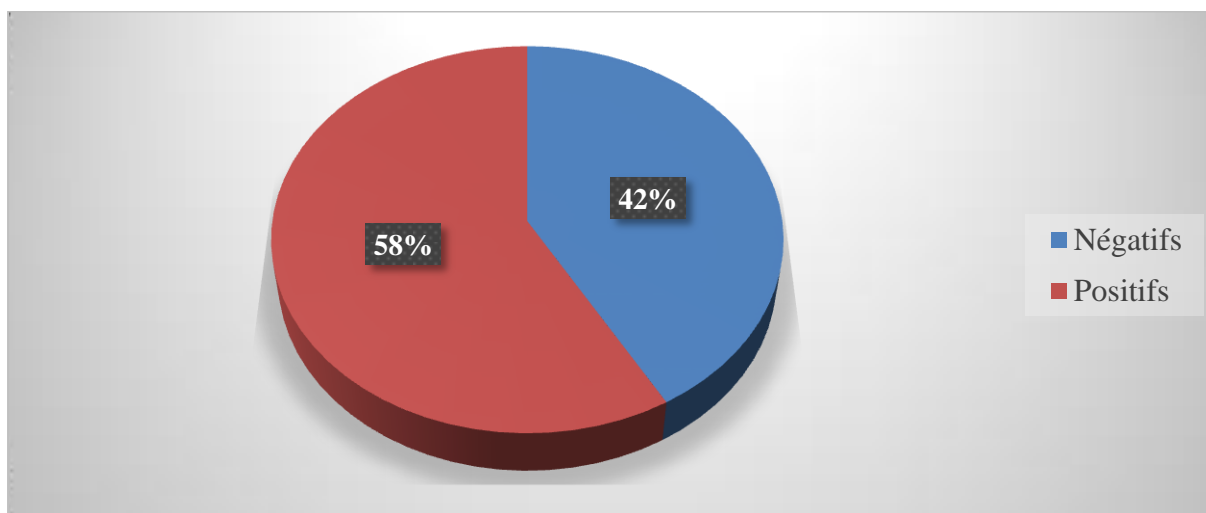
### 3. Results

The frequency of positive samples, the average concentrations of pesticide residues and the maximum and minimum are presented respectively in tables 2, 3 and 4. Table 2 presents, depending on the production site, the frequency of samples contaminated with residues the four (04) pesticides sought.

Table 2: Number of egg samples contaminated with residues of the 04 pesticides detected

Communes	Number Samples	Researched pesticides				Number contaminat ed Samples
		DDT C <sub>2</sub> H <sub>8</sub> NO <sub>2</sub> PS	Dieldrine C <sub>22</sub> H <sub>19</sub> Br <sub>2</sub> NO 3	Endosulfane α C <sub>12</sub> H <sub>21</sub> N <sub>2</sub> O <sub>3</sub> PS	Endosulfane β C <sub>12</sub> H <sub>8</sub> Cl <sub>6</sub> O	
Commune I	6	04	00	02	02	04
Commune II	6	03	00	02	02	03
Commune III	6	03	01	01	03	04
Commune IV	6	02	00	02	01	03
Commune V	6	04	01	03	02	04
Commune VI	6	03	00	03	03	03
Total	36	19	02	13	13	21

Among the 36 egg samples analyzed, 21 were contaminated (58%) with residues of organochlorine pesticides (Figure 4). The dieldrin, DDT,  $\alpha$  endosulfan,  $\beta$  endosulfan were found in most of the egg sampled at the twelve egg sale sites in the District of Bamako (Table 2).



**Figure 4:** Percentage of egg samples with or without pesticide residues

21 of the 36 egg samples submitted for analysis were contaminated with four pesticides, i.e. 58%, and 42% were found to be uncontaminated (Figure 4).

DDT was the most encountered pesticide in about 53% of the samples, Endosulfan A and Endosulfan B were encountered in 36% and the lowest dosed Dieldrin with less than 6% of the egg samples analyzes. From a statistical point of view, using the Chi-II test, no significant difference could be observed ( $p > 0.005$ ) depending on the sampling site. The average levels expressed in milligrams of pesticide per egg (mg/Kg) per municipality are given in Table 3.

**Table 3:** Average pesticide residue concentrations per site and per Commune (mg/Kg)

Communes	Number Samples	Researched pesticides (mg/Kg)			
		DDT	Dieldrine	Endosulfane $\alpha$	Endosulfane $\beta$
Commune I	06	0,5	-	0,085	0,25
Commune II	06	0,6	-	0,18	0,04
Commune III	06	0,4	0,021	0,04	0,36
Commune IV	06	0,7	-	0,037	0,06
Commune V	06	0,4	0,019	0,23	0,42
Commune VI	06	0,5	-	0,39	0,07
<b>Average</b>		<b>0,52 <math>\pm</math> 0,05</b>	<b>0,02 <math>\pm</math> 0,01</b>	<b>0,15 <math>\pm</math> 0,04</b>	<b>0,23 <math>\pm</math> 0,03</b>
<b>Maximum Residue Limit</b>		<b>0,02mg</b>	<b>0,03mg</b>	<b>0,05 mg</b>	<b>0,05 mg</b>

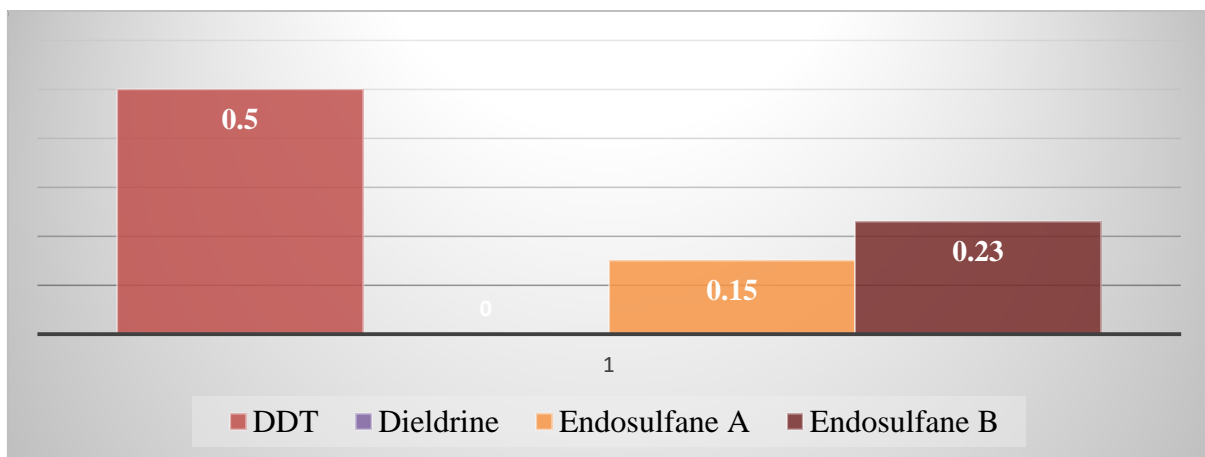
The egg samples were contaminated by DDT, Endosulfan  $\alpha$ , Endosulfan  $\beta$  were found in the egg samples in all the six sites. Dieldrin was only quantified among the samples from the two sites (Commune III and Commune V) in the area study (Table 3).

**Table 4:** Summary of data on contamination of eggs by organochlorine pesticides

Samples	DDT	Dieldrine	Endosulfane $\alpha$	Endosulfane $\beta$
Average (X)	0,52	0,02	0,15	0,23
Standard deviation (SD)	0,50	0,10	0,22	0,32
Maximum (Max)	0,7	0,021	0,39	0,42
Minimum (Min)	0,4	0,019	0,03	0,04

The average of the pesticides detected and quantified was respectively 0.52 mg/Kg  $\pm$  0.05 for DDT, 0.23 mg/Kg  $\pm$  0.03 for endosulfan  $\beta$ , 0.15 mg/Kg  $\pm$  0, 04 for  $\alpha$  endosulfan and 0.02 mg/Kg  $\pm$  0.01 for dieldrin. The maximum regulatory values are 0.01 mg of lindane, 0.05 mg of DDT and 0.02 mg aldrin + dieldrin per kg of egg (Jondreville and his colleagues 2010). These average levels of three residues identified in the egg samples are well above the Maximum Residue Limit (MRL) and therefore unacceptable for consumption.

The dieldrin was found in eggs at the concentration below the Maximum Residue Limit (MRL). All the samples are conform for the dieldrin. On the other hand, no egg sample was not in conformity for the DDT. The rate of conformity was respectively of 33, 33%, 16, 66%, for Endosulfan  $\alpha$ , Endosulfan  $\beta$  (Table 3).



**Figure 5:** Average concentrations by pesticides in eggs

The observation of the graph of the average concentrations shows that, DDT was the pesticide which presented the highest content in the samples of eggs marketed in the District of Bamako, followed respectively by endosulfan  $\beta$  and endosulfan  $\alpha$  while the lowest content was presented by dieldrin ( Table 3 ; Figure 5),.

#### 4. Discussion

Certain organic chemical substances ingested involuntarily by laying hens (pesticides) or voluntarily introduced into their feed (drugs, additives) are likely to be transferred to the egg [9]. The analysis of egg samples marketed in the District of Bamako show their levels of contamination by organochlorine pesticides. The presence of organochlorine compounds is undeniably the consequence of the use of these actives ingredients in agriculture for the production of corn and the conservation of fish meal. Their presence in the different samples could be explained either by fraudulent use for the conservation of fishmeal the maintenance of agricultural plots due to

the relatively low purchase price compared to less persistent and authorized pesticides; or that they have been used insidiously in the past for various reasons (public health, agriculture) and finally, because of their high remanence (persistence) in the environment. The results obtained in this study are similar to those obtained by [10] in Côte d'Ivoire. In Mali, chicken feed is largely made up of maize, shellfish, fishmeal or cottonseed cake, which is a cash crop produced using pesticides. And studies carried out by researchers have shown that some of these components of chicken feed are contaminated with pesticides [11]. In addition, the work of [10] has shown that the rate of contamination is subject to the mode of supply and manufacture of food. Thus for the manufacture of poultry feed, the two ingredients used are corn and fishmeal which receive treatment with pesticides during their storage, while these pesticides are prohibited in feed [12]. These substances are quite worrying insofar as data establishing a potential link between exposure to organochlorine pesticides and the development of cancer are available ) [ 6, 7].

Authors [13] report that so-called "first generation" organochlorine insecticides such as Aldrin, Endrin, DDT, Dieldrin, heptachlor and Lindane have been widely used in chemical control against coffee, cocoa and cotton pests. The contamination of egg samples by organochlorine pesticides can be explained by the level of use of the latter in agricultural production, which is the source of the contamination of soils and agricultural products.

The relatively high concentrations of DDT are probably related to recent uses. Indeed, once absorbed in animals (poultry), DDT and its metabolites are rapidly distributed via the blood to all the tissues of the body to be stored there and undergo partial metabolism. Because of their fat solubility, these compounds will mainly accumulate in fats, undergo reductive dechlorination and then end up in poultry products [14]. Because this metabolite is highly retained in adipose tissue [15], which in part may justify the predominance of DDT in the egg samples of this study.

## **5. conclusion**

The concentration of organochlorine pesticide residues in the chicken eggs sold in the District of Bamako was determined. The results reveal the presence of organochlorine pesticides, mainly DDT and Endosulfan, currently banned from use in Mali for several decades. The egg samples were contaminated by DDT, Endosulfan  $\alpha$ , Endosulfan  $\beta$  were found in the egg samples in all the six sites. The average of the pesticides concentration detected and quantified was respectively  $0.52 \text{ mg/Kg} \pm 0.05$  for DDT,  $0.23 \text{ mg/Kg} \pm 0.03$  for endosulfan  $\beta$ ,  $0.15 \text{ mg/Kg} \pm 0, 04$  for  $\alpha$  endosulfan and  $0.02 \text{ mg/Kg} \pm 0.01$  for dieldrin. The diedrin was found in eggs at the concentration below the Maximum Residue Limit (MRL). All the samples are in conformity for the diedrin, but, no egg sample was not in conformity for the DDT. The rate of conformity was respectively of 33, 33%, 16, 66%, for Endosulfan  $\alpha$ , Endosulfan  $\beta$ .

## **References**

- [1]. DNPIA, 2015. Direction nationale des productions et des industries animales Rapport Annuel. 125 pages.
- [2]. S.E.P. Mensah , O.D. Koudandé, P. Sanders, M. Laurentie , G.A. Mensah, F.A. Abiola.,2016 Résidus



d'antibiotiques et denrées d'origine animale en Afrique : risques de santé publique.

- [3]. **Koné, A.K., 2016.** Recherche de l'aflatoxine dans le lait. Thèse de doctorat de pharmacie, Faculté de Pharmacie, USTTB, Bamako, Mali. 114p.
- [4]. Diarra, O., Babana, A. H., Maïga, K., 2017. Fungal profile and mycotoxin contamination in animal feed in urban and peri-urban zones of Bamako. *Int. J. Curr. Res. Biosci. Plant Biol.* 4(11), 50-56.
- [5]. David HW. 2004. Pesticide, Veterinary and Other Residues in Food. Woodhead Publishing Limited and CRC Press LLC, Cambridge England.
- [6]. Dorgan, J. F., Brock, J. W., Rothman, N., Needham, L. L., Miller, R., Stephenson, H. E., ... & Taylor, P. R. (1999). Serum organochlorine pesticides and PCBs and breast cancer risk: results from a prospective analysis (USA). *Cancer Causes & Control*, 10(1), 1-11.
- [7]. Høyer, A. P., Jørgensen, T., Brock, J. W., & Grandjean, P. (2000). Organochlorine exposure and breast cancer survival. *Journal of Clinical Epidemiology*, 53(3), 323-330.
- [8]. **Rapport CMDT, 2018.** Etude d'évaluation de la qualité des statistiques agricoles et propositions d'amélioration.
- [9]. Jondreville C, Fournier A, Travel A, Feidt C, Roudaut B. (2010). Contaminants chimiques organiques des œufs de poules pondeuses : aspects réglementaires, modalités et risques de transfert. *Inra Productions Animales* 23 (1), 205-214.
- [10]. Kouadio, David Léonce, Ehouman, Serge Guy Ano, Soro, Baba Donafologo, *et al.* Contamination du lait caillé et de l'œuf consommé en Côte d'Ivoire par des pesticides organochlorés. *Afrique Science: Revue Internationale des Sciences et Technologie*, 2014, vol. 10, no 4, p. 61-69.
- [11]. Traore, S. K., Mamadou, K., Dembele, A., Lafrance, P., Banton, O., & Houenou, P. (2003). Etude comparative du niveau de résidus de pesticides organochlorés chez trois espèces de poissons du lac de Buyo (sud-ouest de la Côte d'Ivoire) et estimation du potentiel de risques pour la santé humaine. *Journal de la Société ouest-africaine de chimie*, (16), 137-152.
- [12]. Savadogo PW, Traore O, Topan M, Tapsoba KH, Sedogo PM, Bonzi-Coulibaly YL. 2006. Variation de la teneur en résidus de pesticides dans les sols de la zone cotonnière du Burkina Faso. *J. Afri. Sci. l'Environ.*, 1 : 29-39.
- [13]. Adjagodo A, Tchibozo MAD, Kelome NC, Lawani R. 2016. Flux des polluants liés aux activités anthropiques, risques sur les ressources en eau de surface et la chaîne trophique à travers le monde : synthèse bibliographique. *Int. J. Biol. Chem. Sci.*, 10(3): 1459-1472. DOI: <http://dx.doi.org/10.4314/ijbcs.v10i3.43>

- [14]. Kitamura S, Shimizu Y, Shiraga Y, Yoshida M, Sugihara K, Ohta S. 2002. Reductive Metabolism of p,p'- DDT and o,p'- DDT by Rat Liver Cytochrome P450. *Drug Metabolism and Disposition*, 30(2): 113- 118. DOI: <https://doi.org/10.1124/dmd.30.2.113>.
- [15].. Tebourbi O, Driss M R, Sakly M, Rhouma K B. 2006. Metabolism of DDT in different Tissues of Young Rats. *Journal of Environmental Science and Health*, 41(2): 167-176.