



# Investigating the origins of B<sub>12</sub> biosynthesis in the most ancient roots of the tree of life

Amanda K. Petrus, Ph.D.,\* Kristen S. Swithers, Ph.D.,# Sergio Bibis, Ph.D.,+ Veronica Celone,+ Tripta Jutla\* Edil Alicea\*

\* School of Arts and Sciences, University of Bridgeport, Bridgeport, CT

# Icahn School Of Medicine at Mount Sinai, Hess Center for Science and Medicine, New York, NY

+ Communicating author (apetrus@Bridgeport.edu)

## Abstract

Vitamin B<sub>12</sub>, also known as B<sub>12</sub> or cobalamin, is a vital nutrient required across all branches of life, but the ability to synthesize this complex molecule *de novo* is limited to only a few archaea and bacteria. *De novo* synthesis begins with glutamate and utilizes over 30 gene products to produce an active cobalamin [1]. Previous studies suggest that of the available bacterial genomes, only half utilizing cobalamin can synthesize it [2]. The other half either take up complete cobalamin from the environment via an ABC transporter, or scavenge incomplete corrinoids (partial cobalamin molecules) as precursors to synthesize active cobalamin [3, 4].

The evolutionary histories and identities of multiple genes within these B<sub>12</sub> pathways are unknown, leaving gaping holes in our understanding of the only source of biosynthesis of the vitamin so essential to human survival. Genes of particular interest to this investigator are those responsible for producing reductases that act upon the central cobalt atom of B<sub>12</sub>. Three reductases with unknown gene identities are located within the B<sub>12</sub> biosynthetic pathway and it is the aim of this research to identify those genes responsible.

## Background

### Vitamin B<sub>12</sub>

- Most complex cofactor
- Essential micronutrient required by life
- Only made by certain bacteria and archaea.
- Structure contains modified tetrapyrrole ring bearing resemblance to heme, chlorophyll, siroheme and coenzyme F<sub>430</sub>.
- Each organism's B<sub>12</sub> pathway unique

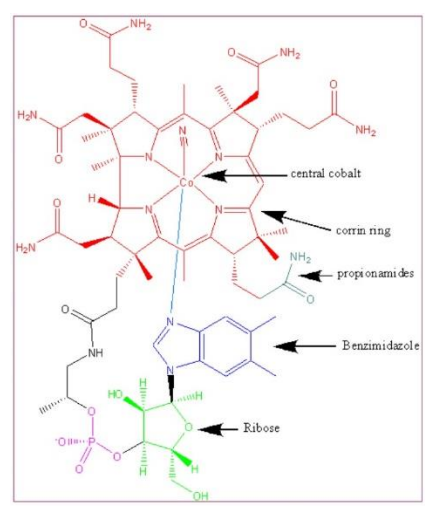


Fig. 1. Vitamin B<sub>12</sub> (cobalamin). The central cobalt atom is the key to the cofactor's biological activity and versatility.

### Ways to get Vitamin B<sub>12</sub>

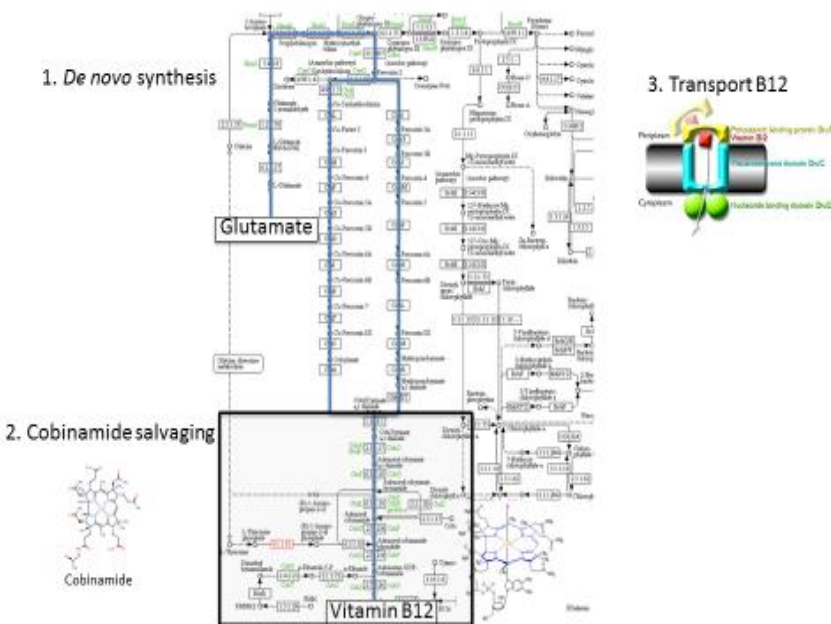


Fig. 2. Bacterial vitamin B<sub>12</sub> biosynthetic pathway. Organisms can utilize one of three major ways to get vitamin B<sub>12</sub>. 1. De novo synthesis, 2. Cobinamide salvaging, 3. Transport of external B<sub>12</sub>.

## B<sub>12</sub> synthesis in Thermotogae phylum as starting model

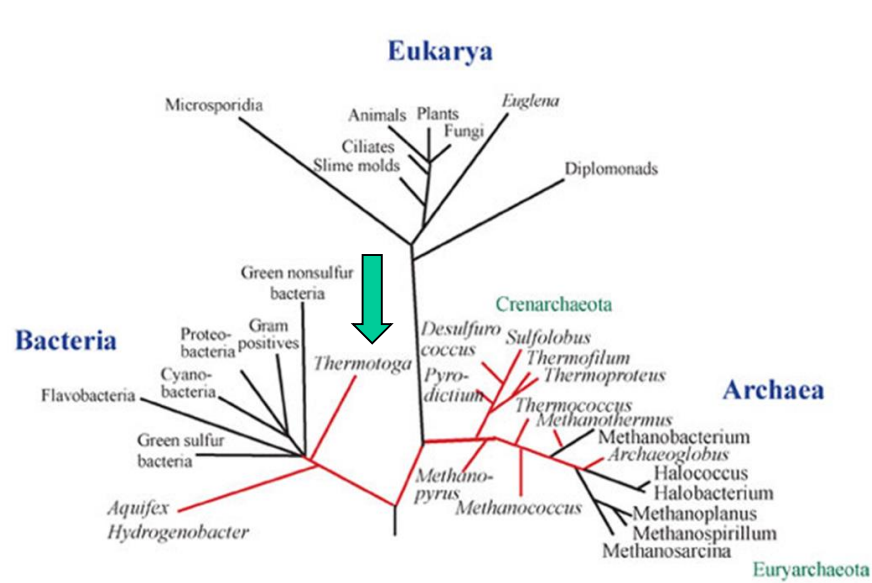


Fig. 3. Placement of Thermotogae at the base of the tree of life. The Thermotogae (green arrow) is a deep branching phylum that possess all three types of B<sub>12</sub> genes.

- We have previously shown two members of this phylum capable of the complete *de novo* synthesis of B<sub>12</sub>. [5]

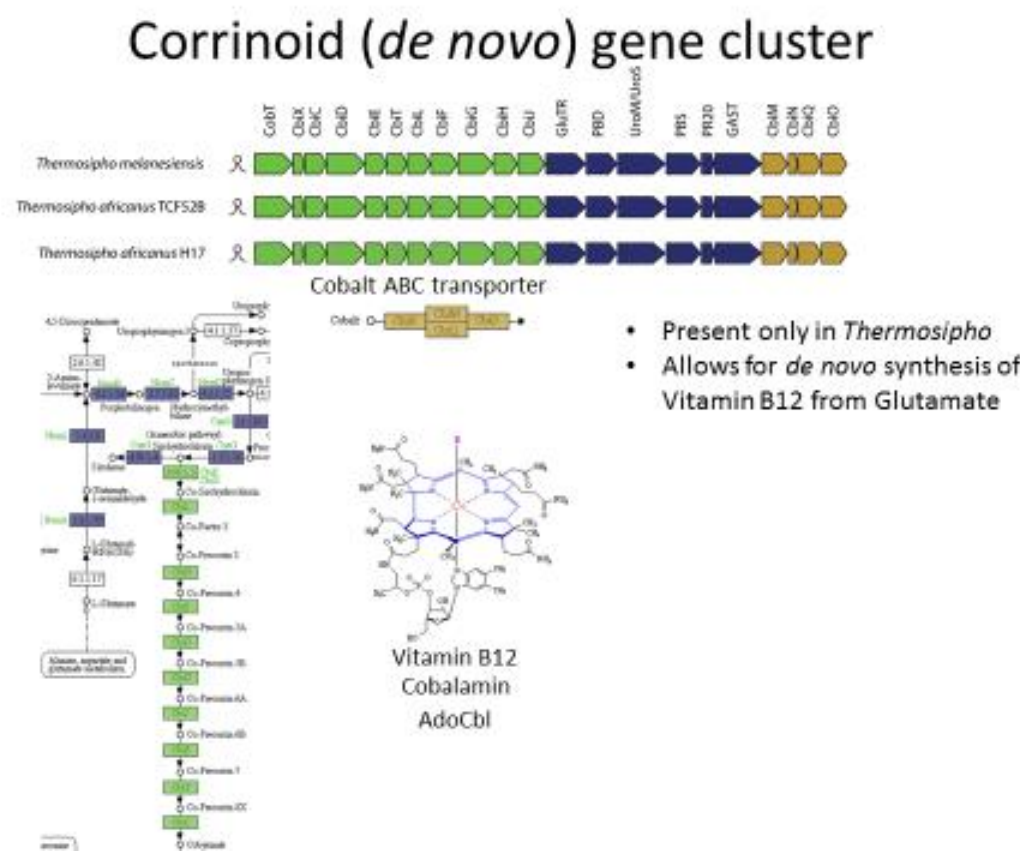


Fig. 4. B<sub>12</sub> related gene clusters in *Thermosipho*. Present as an operon in *Thermosipho* species. From presence/absence analysis 20 continuous genes (unique to *Thermosipho*) involved in vitamin B<sub>12</sub> synthesis were revealed.

### B<sub>12</sub> standard and *Ts. Africanus H17* cell extract

- B<sub>12</sub> identified in extract by UV and MS as compared to B<sub>12</sub> standard
- Detected on full scan mode

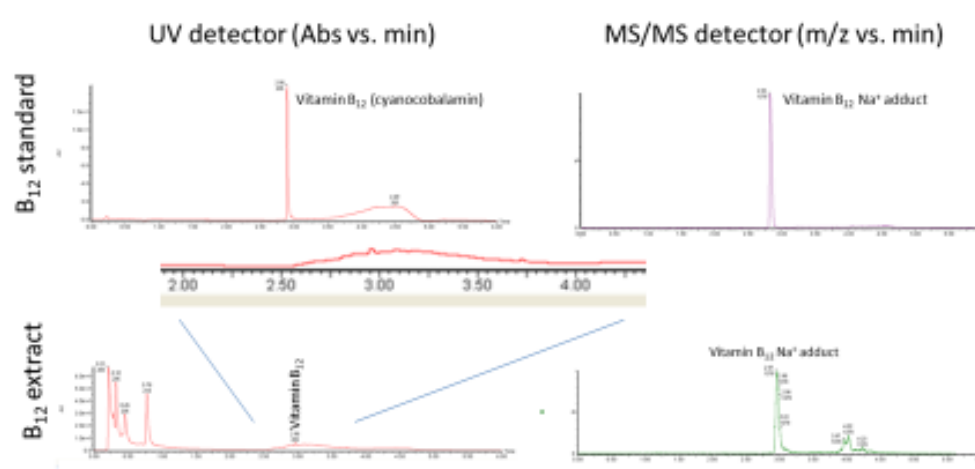


Fig. 5. B<sub>12</sub> production experimentally verified in *Thermosipho*. Cell extracts of *Ts. Africanus H17* and *TCF* tested for B<sub>12</sub> production by UPLC/MS (shown above) and *Lactobacillus* assay. This was verified by qRT-PCR analysis of gene expression

## Identifying reductases in the B<sub>12</sub> pathway

- Parts of the B<sub>12</sub> pathways poorly understood
  - Specifically reductases that act upon the central cobalt atom of the B<sub>12</sub>
- Reduction of the cobalt is the driving force behind B<sub>12</sub>
- Few reductases have been identified
  - *Salmonella enterica* and *Escherichia coli* [6]
  - *Pseudomonas denitrificans* [7, 8]

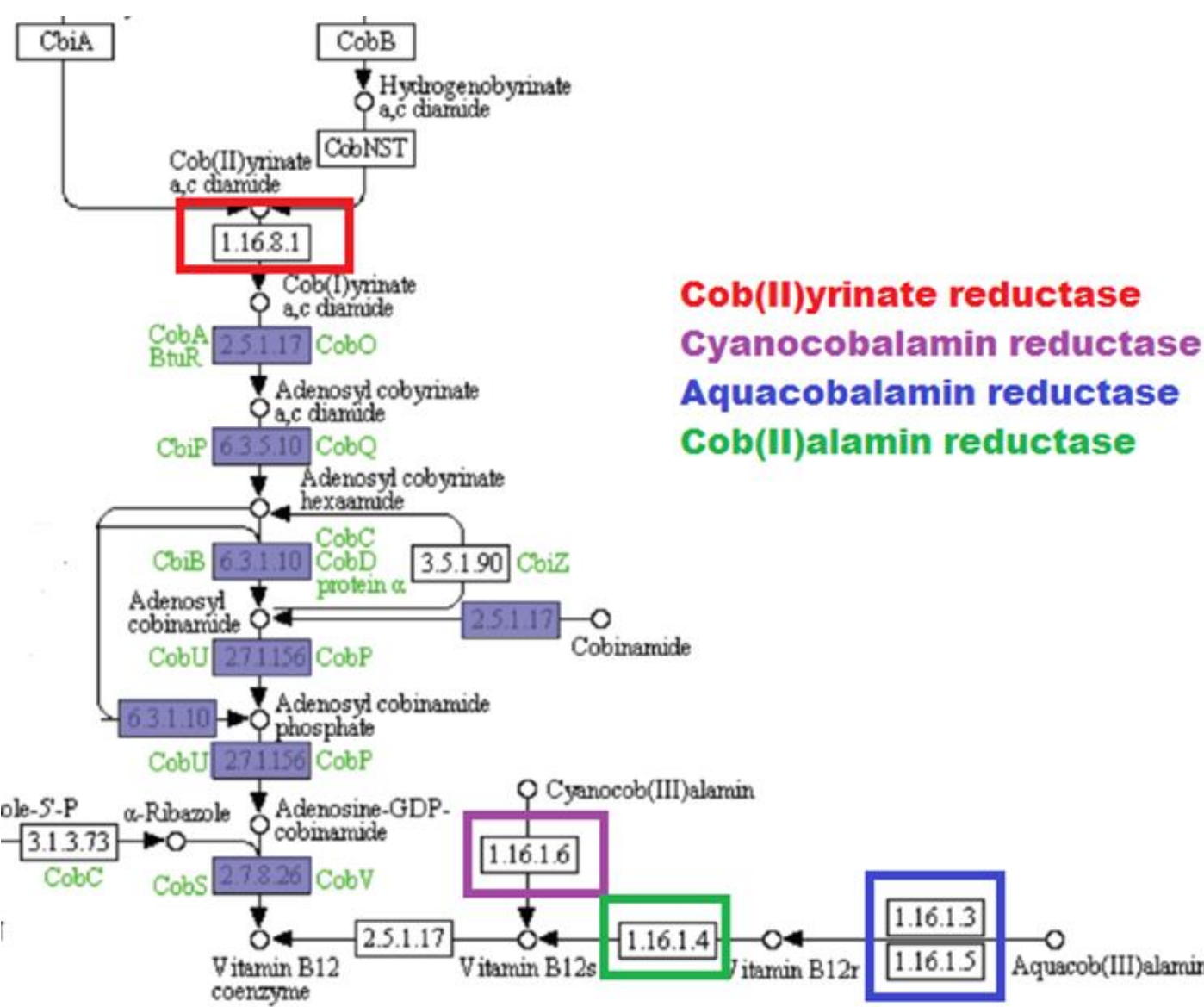


Fig. 6. B<sub>12</sub> biosynthesis and recycling in *Thermosipho melanesiensis* (KEGG). [8] The missing reductases are displayed within colored boxes. Red is Cob(II)yrinate reductase (1.16.8.1), Purple is cyanocobalamin reductase (1.16.1.4), Blue is aquacobalamin reductase (1.16.1.3 and 1.16.1.1).

- We aim to identify the B<sub>12</sub> reductases
- Three candidates have been identified :
  - Tmel\_0728, Tmel\_0733 and Tmel\_1756.
- Candidates share domains with known reductases (Pfam database [10])
- Candidates were identified by BLAST searches against B<sub>12</sub> proteins in
  - *Brucella melitensis*, *Rhodobacter capsulatus* and *Pseudomonas denitrificans*

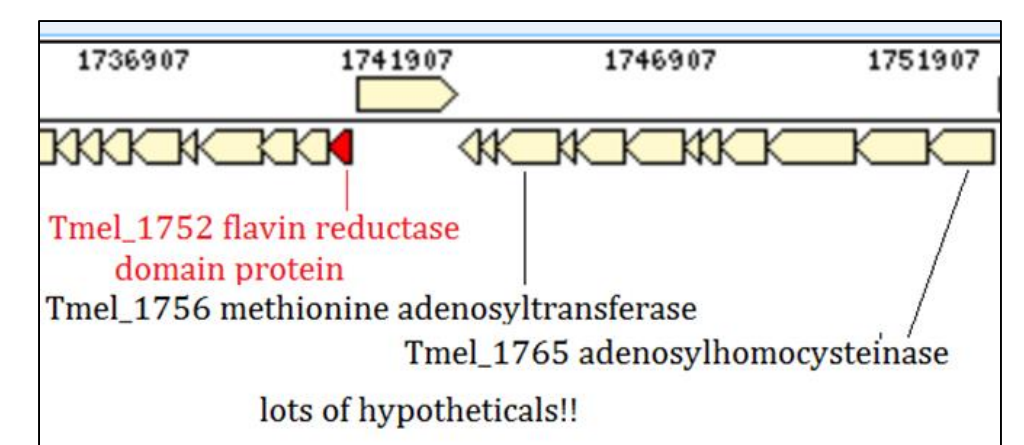
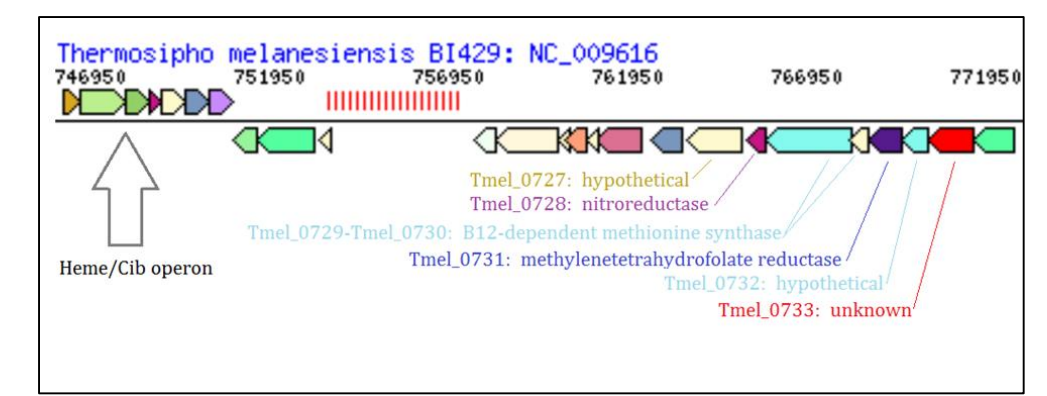


Fig. 6. Gene neighborhoods of reductase candidates. Genes of interest (top: Tmel\_0728 and 0733, bottom: Tmel\_1752) are located within the vicinity of genes related to B<sub>12</sub> biosynthesis or B<sub>12</sub> dependent reactions.

Reductase Protein Candidate	Domain/Family Match	# Amino acids	Mol. Mass	% α % β
Tmel_0728	Nitroreductase	182	21 kDa	79.7 58.2
Tmel_0733	4Fe-4S Binding Domain	434	48 kDa	60.6 68.9
Tmel_1752	Flavin Reductase	152	17 kDa	77.0 67.1

Table 1. Gene candidate sequence analysis as done by Pfam and CFSSP. [9, 10]

- These three candidates are currently being expressed for purification and characterization
- Candidates will be compared to known reductases from *Salmonella enterica*, *Escherichia coli* and *Pseudomonas denitrificans*

## Conclusion

The main goal of this work is to identify the genes responsible for the cobalt reductases that act within the B<sub>12</sub> pathways of the Thermotogae phylum. Three reductase candidates have been identified within *Thermosipho* and are currently being expressed and characterized for further experiments.

### References

- Martens JH, Barg H, Warren MJ, Jahn D. 2002. Microbial production of vitamin B12. Appl Microbiol Biotechnol. 58:275-285.
- Zhang Y, Rodionov DA, Gelfand MS, Gladyshev VN. 2009. Comparative genomic analyses of nickel, cobalt and vitamin B12 utilization. BMC Genomics. 10:78.
- Escalante-Semerena JC. 2007. Conversion of cobinamide into adenosylcobinamide in bacteria and archaea. J Bacteriol. 189:4555-4560.
- Woodson JD, Reynolds AA, Escalante-Semerena JC. 2005. ABC transporter for corrinoids in Halobacterium sp. strain NRC-1. J Bacteriol. 187:5901-5909.
- Swithers, KS, Petrus, AK, Gogarten, P, Noll, K., Butzin, N., 2012. Vitamin B12 synthesis and salvage pathways were acquired by horizontal gene transfer to the Thermotogales. Genome Biol. Evol., 4(8):730-9.
- Timme, RE, Pettengill, JB, Allard, MW, Strain, E, Barrangou, R, Wehnes, C, Van Kessel, JS, Musser, SM, Brown, EW, 2013. Phylogenetic Diversity of the Enteric pathogen salmonella enterica subsp enterica Inferred from Genome-wide reference-free SNP characters. Genome Biol Evol. 5 (11), 2109-2123.
- Fonseca, MV, Escalante-Semerena, JC. 2000. Reduction of cob(III)alamin to cob(II)alamin in Salmonella enterica serovar typhimurium LT2. J bacteriol. 189 (15):4304-9.
- Kanehisa, M. and Goto, S., 2000. KEGG: Kyoto Encyclopedia of Genes and Genomes. Nucleic Acids Res. 28, 27-30.
- Finn, RD, Bateman, A, Clements, P, Coggill, P, Eberhardt, RY, Eddy, SR, Heeger, A, Hetherington, K, Holm, L, Mistry, J, Sonnhammer ELL, Tate, J, Punta M, 2014. The Pfam protein families database. Nucleic Acids Research. 42: D222-D230.
- Chou, PY, Fasman, GD, 1974. Prediction of protein conformation. Biochemistry, 13 (2), 222-245.

## Acknowledgements

Current research is funded by a 2014/2015 UB Seed Grant. We would like to thank the School of Arts and Sciences for their support.